Intraductal Neoplasms of the Pancreas

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Abstract

Here described are definitions, epidemiology, etiology, clinical features, radiology, pathology, and treatment and prognosis of intraductal neoplasms of the pancreas, namely, intraductal papillary mucinous neoplasms (IPMNs), intraductal oncocytic papillary neoplasms (IOPNs), and intraductal tubulopapillary neoplasms (ITPNs). IPMNs are grossly visible intraductal epithelial neoplasms of mucin-producing cells. IPMNs are fairly common without known etiologic factors. Imaging studies show cystically dilated ducts involving branch ducts, the main duct, or the both of ducts. Microscopically, the neoplastic cells grow in papillae with various atypical degree ranging from low-grade to high-grade. The papillae show various morphologic features with expression of characteristic mucin proteins, which are classified into gastric, intestinal, and pancreatobiliary types. Mutations in KRAS and GNAS are frequently found. IPMNs often become invasive, which show adenocarcinoma with ductal or mucinous elements. Disease-specific survivals of patients with surgically resected IPMNs are fairly good in low-grade IPMNs, modest in high-grade IPMNs, however, poor in invasive IPMNs. IOPNs show cystically dilated mucinous ducts with arborizing papillae consist of eosinophilic cells. IOPN is a rare tumor with an average age of patients <65 years. Imaging studies of IOPNs show the same feature as those of IPMNs. Pathologically, IOPNs show high-grade atypia occasionally with invasive elements. IOPNs often harbor fusion genes of ATP1B1-PRKACB, DNAJB1-PRKACA, and ATP1B1-PRKACA. Disease-specific survival rates of patients with surgically resected IOPNs are reported to be 84% for 5-year and 73% for 10-year. ITPNs are intraductal, grossly visible solid neoplasms arising in the MPD or its branches. ITPN is a rare tumor. Imaging studies show characteristic features called the two-tone duct sign and the cork-in-wine bottle sign. Pathologically, ITPNs show packed tubulo-paillary glands consist of cuboidal cells with enlarged atypical nuclei and no visible mucin in cytoplasm. ITPNs often harbor mutations in *PIK3CA, KMT2C, KMT2D*, and *BAP*. ITPNs are often with invasion, and such cases show poor prognosis.

10.1 Intraductal Papillary Mucinous Neoplasm (IPMN)

10.1.1 Definition

IPMN is a grossly visible, intraductal epithelial neoplasm of mucin-producing cells, found in the main pancreatic duct (MPD), or its branches [1, 2]. The neoplastic epithelium is usually papillary, but may include tubular glands, and the extent of mucin secretion, duct dilatation, and dysplasia can vary [3] (Fig. 10.1). Non-invasive IPMNs are classified into two categories, based on the degree of cytoarchitectural atypia: low-grade and high-grade (carcinoma *in situ*) [4]. If there is a component of invasive carcinoma, these are designated as IPMN with associated invasive carcinoma [4].

10.1.2 Epidemiology

IPMNs are fairly common, particularly in the elderly. Incidence was reported to be 1.7–2.8% in consecutive CT scans [5, 6]. The incidence doubled among the patients in their sixties, and tripled in the seventies [6].

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M. Makuuchi et al. (eds.), The IASGO Textbook of Multi-Disciplinary Management of Hepato-Pancreato-Biliary Diseases, https://doi.org/10.1007/978-981-19-0063-1_10





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Fig. 10.1 Histological images of IPMN. (a) IPMN shows a cystically dilated duct filled with mucin. Well-formed neoplastic papillae are observed inside the dilated duct. (b) The neoplastic papillae are con-

sisted of mucin containing tall columnar cells. Grade of atypia may vary from low-grade to high-grade. Hematoxylin and eosin staining. Original magnifications were \times 20 (a) and \times 100 (b)

10.1.3 Etiology

No definite etiological environmental factors associated with IPMNs are known. However, patients with Peutz-Jeghers syndrome, familial adenomatous polyposis, and McCune Albright syndrome have a greater risk of IPMNs [7–9]. Individuals with familial predisposition to pancreatic cancer often harbor cystic lesions in their pancreas that are presumably IPMNs [10].

10.1.4 Clinical Features

Clinical manifestations of the dilated main duct include epigastric pain, chronic pancreatitis, weight loss, diabetes mellitus, and jaundice [11–15], whereas the dilated branch ducts are often discovered fortuitously during clinical evaluation of some other conditions [16].

10.1.5 Radiology

Radiological imaging reveals three distinct types of IPMN, including branch duct-type, main duct-type, and mixed type [16–18]. Branch duct IPMNs show dilated secondary pancreatic ducts of >5 mm size, without the dilatation of the main duct, while the main duct IPMNs show segmental or diffused dilation of the MPD, without any other causes of obstruction. Mixed type IPMNs show the characteristics of both these types of IPMNs [16, 18]. Mural nodules and/or irregular ductal wall thickening can be the sign of high-grade or invasive neoplasms [19, 20].

10.1.6 Pathology

10.1.6.1 Macroscopic Appearance

IPMNs show dilated ducts containing mucin. Dilated branch ducts are seen as cysts anywhere in the pancreas. IPMNs involving the main duct can be identified as segmental/fusiform or diffuse/tortuous dilatation of the duct, often accompanied by dilated secondary branch ducts [3], containing mural nodules or polypoid tumors.

10.1.6.2 Microscopic Appearance and Variations

Papillary proliferation of mucin-containing tall columnar cells is a characteristic histopathological feature of IPMNs [1] (Fig. 10.1). Shapes of papillae are diverse, and cellular atypia also vary. According to the shapes of papillae, IPMNs are subdivided into gastric, intestinal, and pancreatobiliary types [21–23]. Gastric-type IPMNs show thick, finger-like papillae, resembling the gastric foveolar cells, or tubular structures of the pyloric glands. Intestinal-type IPMNs show villous papillae mimicking a villous colonic tumor, whereas pancreatobiliary-type show complex, arborizing papillae.

10.1.6.3 Immunohistochemistry

Ductal markers, including cytokeratins 7 and 19, CA19-9, and CEA, are strongly expressed in most of the IPMNs [24, 25]. Mucin glycoproteins MUC1, MUC2, MUC5AC, and MUC6, show subtype-specific expression patterns among IPMNs [21, 26–28]. Gastric-type of IPMNs express MUC5AC and MUC6, while those of the intestinal-type IPMNs contain MUC2 and MUC5AC. Pancretobiliary-type IPMNs express MUC1, MUC5AC, and MUC6 [22, 29].

10.1.6.4 Grading

IPMNs can be low- or high-grade, based on the degree of cytoarchitectural atypia [4]. A high-grade lesion corresponds to carcinoma in situ. Existence of invasive carcinoma with IPMN leads to designation of IPMN with associated invasive carcinoma [4].

10.1.6.5 Differential Diagnosis

Mucinous cystic neoplasm (MCN), oligocystic serous cystic neoplasm (OSC), intraductal tubulopapillary neoplasm (ITPN), lymphoepithelial cyst (LEC), and chronic pancreatitis (CP) with a retention cyst or pseudocyst should be differentially diagnosed from IPMN. MCNs have characteristic ovarian-type stroma in the cyst wall [30]. OSCs are lined by distinct glycogen-rich cuboidal cells [31]. ITPN is a solid intraductal tumor clogging the duct, composed of high-grade cuboidal cells that form tubulopapillae [32]. LECs are lined by keratinized squamous epithelium with lymphoid stroma [33]. When the cysts in CP have flat epithelial lining, they are identified as retention cysts, and when no lining cells are present, they are called pseudocysts [34].

10.1.6.6 Molecular Pathology

Sixty to eighty percent of the IPMNs harbor somatic mutations in *KRAS*, while 50–70% have mutations in *GNAS* [35, 36]. Although *KRAS* mutations are prevalent in the pancreatic ductal adenocarcinomas (PDAC) as well, *GNAS* mutations are rarely found here, which makes them a specific characteristic of IPMN [36–38]. In 14% of the IPMNs, *RNF43* also shows somatic mutations [36, 39, 40]. Overexression of p53, which presumably indicates missense mutations of *TP53*, is found in 10–40% of the high-grade IPMNs and in 40–60% of the associated invasive carcinomas [29, 38, 41, 42]. While loss of SMAD4 is rare [43, 44], nuclear expression of β-catenin is seen in 18–39% of the IPMNs [38, 45].

10.1.6.7 Treatment and Prognosis

IPMNs with high-grade dysplasia or invasive carcinoma should be surgically resected. According to the international consensus guidelines for management of patients with IPMN [16], surgery is indicated by high-risk stigmata and worrisome features, such as cysts \geq 3 cm, enhancing mural nodules <5 mm, thickened enhanced cyst walls, MPD with a size of 5–9 mm, abrupt change in the MPD caliber with distal pancreatic atrophy, lymphadenopathy, elevated serum level of CA19–9, and a rapidly growing cyst at the rate of >5 mm/2 years. High-risk stigmata are usually associated with obstructive jaundice, an enhanced solid component, and MPD with a size \geq 10 mm. IPMNs with high-risk stigmata should be resected immediately, while those with worrisome features should be evaluated by endoscopic ultrasound, to further risk-stratify the lesions, whether they have mural nodules or involvement of the MPD. High-grade lesions can be evaluated with cyst fluid or pancreatic juice cytology [16].

Five-year survival rate for patients with surgical resection of the low-grade IPMNs is 100%, and 95–85% with high-grade IPMNs [22, 46, 47]. Survival rate varies between 36–90% when the IPMNs are associated with invasive carcinoma, depending on the stage [22, 46–48]. The morphological subtypes of IPMNs can be a prognostic indicator; 5-year survival rate is 94% for the gastric-type, 90% for the intestinal-type, and 50% for the pancreatobiliary-type [22, 48].

10.2 Intraductal Oncocytic Papillary Neoplasms (IOPN)

10.2.1 Definition

IOPN is an intraductal neoplasm of eosinophilic epithelial cells that form arborizing papillae [49] (Fig. 10.2). It is categorized as a variant of IPMNs, as it shows a grossly visible intraductal neoplasm with mucin production [21], similar to IPMN. However, a number of studies have reported that IOPNs have distinct molecular features that distinguish them from IPMNs in the current fifth edition of the World Health Organization classification of tumors of the digestive system [50].

10.2.2 Epidemiology

IOPNs are fairly infrequent, making up only 4.5-8.4% of the cystic neoplasms of the pancreas [22, 51]. They occur more frequently in men than in women, between 20–80 years of age. With an average age of <65 years, patients with IOPNs are younger than those with IPMNs (> 65 years) [49, 51–53].

10.2.3 Etiology

No etiological factor associated with IOPN is known.

10.2.4 Clinical Features

Clinical manifestations of IOPN are the same as those of IPMNs, including abdominal pain, weight loss, diabetes mellitus, and jaundice [49, 51].



Fig. 10.2 Histological images of IOPN. (**a**) IOPN shows mucinous dilated ducts. Arborising papillae are seen inside them. (**b**) The neoplastic papillae of IOPN consist of eosinophilic cells with enlarged nuclei

and prominent nucleoli, which show high-grade atypia. Intraepithelial lumina are often seen in the cells. Hematoxylin and eosin staining. Original magnifications were \times 20 (a) and \times 100 (b)

10.2.5 Radiology

Like IPMNs, IOPNs show cystic dilatation of the pancreatic duct, and can be of branch duct type, main duct type, and mixed type. Mural nodules are often seen inside the dilated duct [53].

10.2.6 Pathology

10.2.6.1 Macroscopic Appearance

IOPNs show cystic dilation of ducts filled with mucin [49, 53], which is indistinguishable from IPMNs, and may involve branch ducts or the main duct. Polypoid tortuous tumors are often seen inside them.

10.2.6.2 Microscopic Appearance

In IOPNs, the dilated ducts are lined by arborizing papillae, consisting of eosinophilic cells with enlarged nuclei and prominent nucleoli [21, 49], and show high-grade atypia. Intraepithelial lumina are often seen in the neoplastic papillae (Fig. 10.2). Occasionally, the neoplasm invades the parenchymal stroma, resulting in a diagnosis of IOPN with associated invasive carcinoma. The invading mass usually shows clusters of eosinophilic cells in the mucin pools [54].

10.2.6.3 Immunohistochemistry

Neoplastic cells frequently express HepPar1 and mesothelin [54]. The expression of mucin proteins MUC5AC and MUC6 are consistently positive, while MUC1 and MUC2 are infrequently and focally positive [21, 54].

10.2.6.4 Differential Diagnosis

IOPN should be distinguished from cystic neoplasms, including IPMN, mucinous cystic neoplasm, oligocystic

serous cystic neoplasm, lymphoepithelial cyst, and solid pseudopapillary neoplasm.

10.2.6.5 Molecular Pathology

IOPNs infrequently show somatic mutations in *KRAS* and *GNAS* unlike IPMNs [36, 40]. Instead, they carry fusion genes of *ATP1B1-PRKACB*, *DNAJB1-PRKACA*, and *ATP1B1-PRKACA* [55, 56]. Interestingly, these fusion genes upregulate the protein kinase A pathway, which is the main target of *GNAS* that shows frequent gain-of-function mutations in IPMNs, indicating that both IOPNs and IPMNs, are driven by activation of the protein kinase A pathway. Aberrant expression of p53 or SMAD4 is observed in up to 10% of IOPNs [52], and about 30% of these show nuclear accumulation of *B*-catenin [48],whereas *RNF43* mutations are not reported [40, 57]. A patient with a germline *SMAD4* mutation was reported to have developed IOPN [58].

10.2.6.6 Treatment and Prognosis

IOPN should be surgically resected. Disease-specific survival rates of patients with surgically resected IOPNs are reported to be 84% for 5-year and 73% for 10-year [22].

10.3 Intraductal Tubulopapillary Neoplasms (ITPN)

10.3.1 Definition

ITPN is an intraductal, grossly visible solid neoplasm arising in the MPD or its branches [32]. Cystic lesions are infrequent and are only focally and peripherally observed. The neoplastic epithelium has the mixture of tubular and papillary configurations, and the neoplastic cells show uniformly high-grade atypia [32] (Fig. 10.3). The neoplasm is often



Fig. 10.3 Histological images of ITPN. (a) ITPNs show a solid tumor clogged in dilated pancreatic ducts, without visible mucin. Cystic ducts are occasionally seen in the periphery of a neoplasm-obstructed duct. (b) ITPN consist of cuboidal to columnar cells with enlarged nuclei,

with little cytoplasmic mucin, exhibiting solid tubulopapillary growth. Necrotic foci are often seen. Hematoxylin and eosin staining. Original magnifications were $\times 20$ (a) and $\times 100$ (b)

invasive, and is designated as intraductal tubulopapillary neoplasm with associated invasive carcinoma.

10.3.2 Epidemiology

ITPN is rare, barely accounting for 0.9% of the exocrine neoplasms and 3% of the intraductal neoplasms of the pancreas [32].

10.3.3 Etiology

No definite etiological factors of ITPNs are known.

10.3.4 Clinical Features

Clinical manifestations include abdominal pain, nausea, vomiting, jaundice, weight loss, and exacerbation of diabetes mellitus [32, 59]. Some patients have a history of acute pancreatitis [32].

10.3.5 Radiology

In CT scan, patients with ITPN show a solid enhanced mass, packed in a dilated duct. Magnetic resonance cholangiopancreatography indicates a filling defect in a dilated duct or an abrupt disruption of the dilated duct. Images of the dilated duct packed with tumor, are known as the two-tone duct sign and the cork-in-wine bottle sign [60]. ITPNs that involve the MPD show a sausage-like image in magnetic resonance imaging [61].

10.3.6 Pathology

10.3.6.1 Macroscopic Appearance

ITPNs show a solid tumor packed in dilated pancreatic ducts, without visible mucin. Cystic ducts are occasionally seen in the periphery of a neoplasm-obstructed duct [32, 59].

10.3.6.2 Microscopic Appearance and Variations

Cuboidal to columnar cells with enlarged nuclei, with little cytoplasmic mucin, exhibit solid tubulopapillary growth [32, 59]. Some may consist of tubular glands. The neoplastic cells show uniformly high-grade atypia (Fig. 10.3). Intraductal comedo-like necrosis is often present. Stromal invasion can be seen in the periductal parenchyma, which shows clusters of tubulopapillary glands. Tumor emboli are occasionally evident in the veins [32].

10.3.6.3 Immunohistochemistry

Cytokeratin (CK) 7 and CK19 are consistently positive. Mucin glycoproteins, MUC1 and MUC6 are positive, while MUC2 and MUC5AC are negative [32, 59]. Trypsin, an acinar marker, is negative.

10.3.6.4 Differential Diagnosis

A solid intraductal neoplasm can be observed in the intraductal variant of acinar cell carcinoma and intraductal neuroendocrine tumors [62, 63]. Histologically, neuroendocrine tumors can be easily differentiated; however, the intraductal variant of acinar cell carcinoma may show solid tubular growth with necrosis, mimicking ITPNs. Acinar cell carcinomas usually express trypsin, an immunohistochemical marker of differentiation [32]. IPMNs occasionally show solid nodules inside the dilated duct. IPMN of the pyloric gland variant may particularly show a polypoid tumor in a dilated duct, mainly consisting of tubular glands. However, it can be distinguished from ITPNs by its well-formed tubular glands with low-grade atypia and the expression pattern of mucin which is negative for MUC1 and positive for MUC5AC [64].

10.3.6.5 Molecular Pathology

ITPNs often show somatic mutations in *PIK3CA, KMT2C, KMT2D*, and *BAP1* [65, 66]. Some tumors also harbor *FGFR2* fusion genes [66]. *KRAS* or *GNAS* mutations that are common in IPMNs, are not seen in ITPNs. Aberrant expression of p16 and p53 is reported in up to 70% of the cases [59], while the atypical expression of SMAD4 is rare [32, 59].

10.3.7 Treatment and Prognosis

Surgery is the treatment of choice for patients with ITPN, but some of them may experience a recurrence in the remnant tissue after pancreatectomy [32, 59]. ITPNs diffusely involving the pancreatic duct, need total pancreatectomy [32, 59, 67]. Efficacy of the adjuvant therapy is not known. About 10% of the patients with ITPN die of the disease [32, 59], and all of them are reported to have invasive tumors.

References

- Hruban RH, Takaori K, Klimstra DS, Adsay NV, Albores-Saavedra J, Biankin AV, et al. An illustrated consensus on the classification of pancreatic intraepithelial neoplasia and intraductal papillary mucinous neoplasms. Am J Surg Pathol. 2004;28(8):977–87.
- Tanaka M, Chari S, Adsay V, Fernandez-del Castillo C, Falconi M, Shimizu M, et al. International consensus guidelines for management of intraductal papillary mucinous neoplasms and mucinous cystic neoplasms of the pancreas. Pancreatology. 2006;6(1–2):17–32.
- Furukawa T, Takahashi T, Kobari M, Matsuno S. The mucushypersecreting tumor of the pancreas. Development and extension visualized by three-dimensional computerized mapping. Cancer. 1992;70(6):1505–13.
- Basturk O, Hong SM, Wood LD, Adsay NV, Albores-Saavedra J, Biankin AV, et al. A revised classification system and recommendations from the baltimore consensus meeting for neoplastic precursor lesions in the pancreas. Am J Surg Pathol. 2015;39(12):1730–41.
- Laffan TA, Horton KM, Klein AP, Berlanstein B, Siegelman SS, Kawamoto S, et al. Prevalence of unsuspected pancreatic cysts on MDCT. AJR Am J Roentgenol. 2008;191(3):802–7.

- Su GH, Hruban RH, Bansal RK, Bova GS, Tang DJ, Shekher MC, et al. Germline and somatic mutations of the STK11/LKB1 Peutz-Jeghers gene in pancreatic and biliary cancers. Am J Pathol. 1999;154(6):1835–40.
- Maire F, Hammel P, Terris B, Olschwang S, O'Toole D, Sauvanet A, et al. Intraductal papillary and mucinous pancreatic tumour: a new extracolonic tumour in familial adenomatous polyposis. Gut. 2002;51(3):446–9.
- Wood LD, Noe M, Hackeng W, Brosens LA, Bhaijee F, Debeljak M, et al. Patients with McCune-Albright syndrome have a broad spectrum of abnormalities in the gastrointestinal tract and pancreas. Virchows Arch. 2017;470(4):391–400.
- Canto MI, Hruban RH, Fishman EK, Kamel IR, Schulick R, Zhang Z, et al. Frequent detection of pancreatic lesions in asymptomatic high-risk individuals. Gastroenterology. 2012;142(4):796–804; quiz e14–5.
- Kloppel G. Clinicopathologic view of intraductal papillarymucinous tumor of the pancreas. Hepato-Gastroenterology. 1998;45(24):1981–5.
- Traverso LW, Peralta EA, Ryan JA Jr, Kozarek RA. Intraductal neoplasms of the pancreas. Am J Surg. 1998;175(5):426–32.
- Yasuda H, Takada T, Amano H, Yoshida M. Surgery for mucin-producing pancreatic tumor. Hepato-Gastroenterology. 1998;45(24):2009–15.
- Salvia R, Fernandez-del Castillo C, Bassi C, Thayer SP, Falconi M, Mantovani W, et al. Main-duct intraductal papillary mucinous neoplasms of the pancreas: clinical predictors of malignancy and longterm survival following resection. Ann Surg. 2004;239(5):678–685; discussion 85–7.
- Sohn TA, Yeo CJ, Cameron JL, Hruban RH, Fukushima N, Campbell KA, et al. Intraductal papillary mucinous neoplasms of the pancreas: an updated experience. Ann Surg. 2004;239(6):788– 97; discussion 97–9.
- Tanaka M, Fernandez-Del Castillo C, Kamisawa T, Jang JY, Levy P, Ohtsuka T, et al. Revisions of international consensus Fukuoka guidelines for the management of IPMN of the pancreas. Pancreatology. 2017;17(5):738–53.
- Kobari M, Egawa S, Shibuya K, Shimamura H, Sunamura M, Takeda K, et al. Intraductal papillary mucinous tumors of the pancreas comprise 2 clinical subtypes: differences in clinical characteristics and surgical management. Arch Surg. 1999;134(10):1131–6.
- Tanaka M, Fernandez-del Castillo C, Adsay V, Chari S, Falconi M, Jang JY, et al. International consensus guidelines 2012 for the management of IPMN and MCN of the pancreas. Pancreatology. 2012;12(3):183–97.
- Furukawa T, Oohashi K, Yamao K, Naitoh Y, Hirooka Y, Taki T, et al. Intraductal ultrasonography of the pancreas: development and clinical potential. Endoscopy. 1997;29(6):561–9.
- Koito K, Namieno T, Nagakawa T, Hirokawa N, Ichimura T, Syonai T, et al. Pancreas: imaging diagnosis with color/power Doppler ultrasonography, endoscopic ultrasonography, and intraductal ultrasonography. Eur J Radiol. 2001;38(2):94–104.
- Furukawa T, Kloppel G, Adsay NV, Albores-Saavedra J, Fukushima N, Horii A, et al. Classification of types of intraductal papillarymucinous neoplasm of the pancreas: a consensus study. Virchows Arch. 2005;447(5):794–9.
- Furukawa T, Hatori T, Fujita I, Yamamoto M, Kobayashi M, Ohike N, et al. Prognostic relevance of morphological types of intraductal papillary mucinous neoplasms of the pancreas. Gut. 2011;60(4):509–16.
- 23. Basturk O, Tan M, Bhanot U, Allen P, Adsay V, Scott SN, et al. The oncocytic subtype is genetically distinct from other pancreatic

intraductal papillary mucinous neoplasm subtypes. Mod Pathol. 2016;29(9):1058-69.

- Terada T, Ohta T, Nakanuma Y. Expression of oncogene products, anti-oncogene products and oncofetal antigens in intraductal papillary-mucinous neoplasm of the pancreas. Histopathology. 1996;29(4):355–61.
- 25. Terada T, Ohta T, Kitamura Y, Ashida K, Matsunaga Y. Cell proliferative activity in intraductal papillary-mucinous neoplasms and invasive ductal adenocarcinomas of the pancreas: an immunohistochemical study. Arch Pathol Lab Med. 1998;122(1):42–6.
- 26. Adsay NV, Merati K, Basturk O, Iacobuzio-Donahue C, Levi E, Cheng JD, et al. Pathologically and biologically distinct types of epithelium in intraductal papillary mucinous neoplasms: delineation of an "intestinal" pathway of carcinogenesis in the pancreas. Am J Surg Pathol. 2004;28(7):839–48.
- Nakamura A, Horinouchi M, Goto M, Nagata K, Sakoda K, Takao S, et al. New classification of pancreatic intraductal papillarymucinous tumour by mucin expression: its relationship with potential for malignancy. J Pathol. 2002;197(2):201–10.
- Adsay NV, Merati K, Andea A, Sarkar F, Hruban RH, Wilentz RE, et al. The dichotomy in the preinvasive neoplasia to invasive carcinoma sequence in the pancreas: differential expression of MUC1 and MUC2 supports the existence of two separate pathways of carcinogenesis. Mod Pathol. 2002;15(10):1087–95.
- Furukawa T, Fujisaki R, Yoshida Y, Kanai N, Sunamura M, Abe T, et al. Distinct progression pathways involving the dysfunction of DUSP6/MKP-3 in pancreatic intraepithelial neoplasia and intraductal papillary-mucinous neoplasms of the pancreas. Mod Pathol. 2005;18(8):1034–42.
- Compagno J, Oertel JE. Mucinous cystic neoplasms of the pancreas with overt and latent malignancy (cystadenocarcinoma and cystadenoma). A clinicopathologic study of 41 cases. Am J Clin Pathol. 1978;69(6):573–80.
- Lewandrowski K, Warshaw A, Compton C. Macrocystic serous cystadenoma of the pancreas: a morphologic variant differing from microcystic adenoma. Hum Pathol. 1992;23(8):871–5.
- 32. Yamaguchi H, Shimizu M, Ban S, Koyama I, Hatori T, Fujita I, et al. Intraductal tubulopapillary neoplasms of the pancreas distinct from pancreatic intraepithelial neoplasia and intraductal papillary mucinous neoplasms. Am J Surg Pathol. 2009;33(8):1164–72.
- Truong LD, Rangdaeng S, Jordan PH Jr. Lymphoepithelial cyst of the pancreas. Am J Surg Pathol. 1987;11(11):899–903.
- Kloppel G. Pseudocysts and other non-neoplastic cysts of the pancreas. Semin Diagn Pathol. 2000;17(1):7–15.
- Wu J, Matthaei H, Maitra A, Dal Molin M, Wood LD, Eshleman JR, et al. Recurrent *GNAS* mutations define an unexpected pathway for pancreatic cyst development. Sci Tansl Med. 2011;3(92):92ra66.
- 36. Furukawa T, Kuboki Y, Tanji E, Yoshida S, Hatori T, Yamamoto M, et al. Whole-exome sequencing uncovers frequent *GNAS* mutations in intraductal papillary mucinous neoplasms of the pancreas. Sci Rep. 2011;1:161.
- 37. Wu J, Jiao Y, Dal Molin M, Maitra A, de Wilde RF, Wood LD, et al. Whole-exome sequencing of neoplastic cysts of the pancreas reveals recurrent mutations in components of ubiquitin-dependent pathways. Proc Natl Acad Sci USA. 2011;108(52):21188–93.
- Kuboki Y, Shimizu K, Hatori T, Yamamoto M, Shibata N, Shiratori K, et al. Molecular biomarkers for progression of intraductal papillary mucinous neoplasm of the pancreas. Pancreas. 2015;44(2):227–35.
- 39. Amato E, Molin MD, Mafficini A, Yu J, Malleo G, Rusev B, et al. Targeted next-generation sequencing of cancer genes dissects the molecular profiles of intraductal papillary neoplasms of the pancreas. J Pathol. 2014;233(3):217–27.
- 40. Sakamoto H, Kuboki Y, Hatori T, Yamamoto M, Sugiyama M, Shibata N, et al. Clinicopathological significance of somatic RNF43 mutation and aberrant expression of ring finger protein 43 in intra-

ductal papillary mucinous neoplasms of the pancreas. Mod Pathol. 2015;28(2):261–7.

- 41. Biankin AV, Biankin SA, Kench JG, Morey AL, Lee CS, Head DR, et al. Aberrant p16(INK4A) and DPC4/Smad4 expression in intraductal papillary mucinous tumours of the pancreas is associated with invasive ductal adenocarcinoma. Gut. 2002;50(6):861–8.
- 42. Abe K, Suda K, Arakawa A, Yamasaki S, Sonoue H, Mitani K, et al. Different patterns of p16INK4A and p53 protein expressions in intraductal papillary-mucinous neoplasms and pancreatic intraepithelial neoplasia. Pancreas. 2007;34(1):85–91.
- 43. Iacobuzio-Donahue CA, Klimstra DS, Adsay NV, Wilentz RE, Argani P, Sohn TA, et al. Dpc-4 protein is expressed in virtually all human intraductal papillary mucinous neoplasms of the pancreas: comparison with conventional ductal adenocarcinomas. Am J Pathol. 2000;157(3):755–61.
- 44. Inoue H, Furukawa T, Sunamura M, Takeda K, Matsuno S, Horii A. Exclusion of SMAD4 mutation as an early genetic change in human pancreatic ductal tumorigenesis. Genes Chromosomes Cancer. 2001;31(3):295–9.
- 45. Chetty R, Serra S, Salahshor S, Alsaad K, Shih W, Blaszyk H, et al. Expression of Wnt-signaling pathway proteins in intraductal papillary mucinous neoplasms of the pancreas: a tissue microarray analysis. Hum Pathol. 2006;37(2):212–7.
- 46. Chari ST, Yadav D, Smyrk TC, DiMagno EP, Miller LJ, Raimondo M, et al. Study of recurrence after surgical resection of intraductal papillary mucinous neoplasm of the pancreas. Gastroenterology. 2002;123(5):1500–7.
- 47. Maire F, Hammel P, Terris B, Paye F, Scoazec JY, Cellier C, et al. Prognosis of malignant intraductal papillary mucinous tumours of the pancreas after surgical resection. Comparison with pancreatic ductal adenocarcinoma. Gut. 2002;51(5):717–22.
- Mino-Kenudson M, Fernandez-del Castillo C, Baba Y, Valsangkar NP, Liss AS, Hsu M, et al. Prognosis of invasive intraductal papillary mucinous neoplasm depends on histological and precursor epithelial subtypes. Gut. 2011;60(12):1712–20.
- Adsay NV, Adair CF, Heffess CS, Klimstra DS. Intraductal oncocytic papillary neoplasms of the pancreas. Am J Surg Pathol. 1996;20(8):980–94.
- 50. Basturk O, Esposito I, Fukushima N, Furukawa T, Hong SM, Klöppel G, et al. Pancreatic intraductal oncocytic papillary neoplasm. In: Gill AJ, Klimstra DS, Lam AK, Washington MK, editors. WHO Classification of Digestive System Tumours. WHO Classification of Tumours 1. Lyon: International Agency for Research on Cancer; 2019. p. 315–6.
- Marchegiani G, Mino-Kenudson M, Ferrone CR, Warshaw AL, Lillemoe KD, Fernandez-del CC. Oncocytic-type intraductal papillary mucinous neoplasms: a unique malignant pancreatic tumor with good long-term prognosis. J Am Coll Surg. 2015;220(5):839–44.
- 52. Xiao HD, Yamaguchi H, Dias-Santagata D, Kuboki Y, Akhavanfard S, Hatori T, et al. Molecular characteristics and biological behaviours of the oncocytic and pancreatobiliary subtypes of intraductal papillary mucinous neoplasms. J Pathol. 2011;224(4):508–16.
- 53. D'Onofrio M, De Robertis R, Tinazzi Martini P, Capelli P, Gobbo S, Morana G, et al. Oncocytic intraductal papillary mucinous neoplasms of the pancreas: imaging and histopathological findings. Pancreas. 2016;45(9):1233–42.
- 54. Basturk O, Chung SM, Hruban RH, Adsay NV, Askan G, Iacobuzio-Donahue C, et al. Distinct pathways of pathogenesis of intraductal oncocytic papillary neoplasms and intraductal papillary mucinous neoplasms of the pancreas. Virchows Arch. 2016;469(5):523–32.
- 55. Singhi AD, Wood LD, Parks E, Torbenson MS, Felsenstein M, Hruban RH, et al. Recurrent rearrangements in *PRKACA* and *PRKACB* in intraductal oncocytic papillary neoplasms of the pancreas and bile duct. Gastroenterology. 2020;158(3):573–82 e2.
- Vyas M, Hechtman JF, Zhang Y, Benayed R, Yavas A, Askan G, et al. DNAJB1-PRKACA fusions occur in oncocytic pancreatic and

biliary neoplasms and are not specific for fibrolamellar hepatocellular carcinoma. Mod Pathol. 2020;33(4):648–56.

- Chang XY, Wu Y, Jiang Y, Wang PY, Chen J. RNF43 mutations in IPMN cases: a potential prognostic factor. Gastroenterol Res Pract. 2020;2020:1457452.
- 58. Takai E, Nakamura H, Chiku S, Kubo E, Ohmoto A, Totoki Y, et al. Whole-exome sequencing reveals new potential susceptibility genes for japanese familial pancreatic cancer. Ann Surg. 2020.
- Basturk O, Adsay V, Askan G, Dhall D, Zamboni G, Shimizu M, et al. Intraductal tubulopapillary neoplasm of the pancreas: a clinicopathologic and immunohistochemical analysis of 33 cases. Am J Surg Pathol. 2017;41(3):313–25.
- 60. Motosugi U, Yamaguchi H, Furukawa T, Ichikawa T, Hatori T, Fujita I, et al. Imaging studies of intraductal tubulopapillary neoplasms of the pancreas: 2-tone duct sign and cork-of-wine-bottle sign as indicators of intraductal tumor growth. J Comput Assist Tomogr. 2012;36(6):710–7.
- Lu ZF, Kang B, Li JM, Sun C. Intraductal tubulopapillary neoplasm of the pancreas presenting as sausage. Am J Gastroenterol. 2020.
- 62. Basturk O, Zamboni G, Klimstra DS, Capelli P, Andea A, Kamel NS, et al. Intraductal and papillary variants of acinar cell carcinomas: a new addition to the challenging differential diagnosis of intraductal neoplasms. Am J Surg Pathol. 2007;31(3):363–70.

- 63. Fabre A, Sauvanet A, Flejou JF, Belghiti J, Palazzo L, Ruzniewski P, et al. Intraductal acinar cell carcinoma of the pancreas. Virchows Arch. 2001;438(3):312–5.
- 64. Yamaguchi H, Kuboki Y, Hatori T, Yamamoto M, Shimizu K, Shiratori K, et al. The discrete nature and distinguishing molecular features of pancreatic intraductal tubulopapillary neoplasms and intraductal papillary mucinous neoplasms of the gastric type, pyloric gland variant. J Pathol. 2013;231(3):335–41.
- 65. Yamaguchi H, Kuboki Y, Hatori T, Yamamoto M, Shiratori K, Kawamura S, et al. Somatic mutations in PIK3CA and activation of AKT in intraductal tubulopapillary neoplasms of the pancreas. Am J Surg Pathol. 2011;35(12):1812–7.
- 66. Basturk O, Berger MF, Yamaguchi H, Adsay V, Askan G, Bhanot UK, et al. Pancreatic intraductal tubulopapillary neoplasm is genetically distinct from intraductal papillary mucinous neoplasm and ductal adenocarcinoma. Mod Pathol. 2017;30(12):1760–72.
- 67. Kosmidis C, Varsamis N, Atmatzidis S, Koimtzis G, Mantalovas S, Anthimidis G, et al. Total pancreatectomy with splenectomy for multifocal Intraductal Tubulopapillary Neoplasm (ITPN) of the pancreas associated with invasive component: report of a rare case. Am J Case Rep. 2020;21:e924760.