

# Pathogenesis of COVID-Associated Mucormycosis

Shreya Singh, Rimjhim Kanaujia, and Shivaprakash M. Rudramurthy

# 4.1 Introduction

Mucormycosis is a serious fungal infection caused by ubiquitous fungi which belong to the order Mucorales. The common genera infecting humans include Rhizopus spp. (most commonly R. arrhizus, R. microsporus, and R. homothallicus). Rhizomucor spp., Mucor spp., Cunninghamella spp., Lichtheimia spp., and Apophysomyces spp. [1]. The major underlying health conditions predisposing to mucormycosis include diabetes mellitus (DM) and immunosuppression; trauma such as burns or road traffic accidents, and iron chelation therapy with deferoxamine [1-3]. An upsurge in the cases of mucormycosis has been noted over the last decade in the developed and developing world alike but has been especially concerning in Asia and specifically in India. Recently, a dramatic rise in the cases of mucormycosis infection has been observed in patients with Coronavirus-19 disease (COVID-19) or those recovering from it, as evidenced by several reports and institutional experiences [1, 3, 4]. COVID-19 is a viral inflammatory disease caused by SARS-CoV-2 characterized by an exaggerated proinflammatory response with cytokine surge, altered iron metabolism and iron overload, hemoglobinopathy, hypercoagulability

state causing multisystem involvement [5–8]. Several strategies to prevent and treat mucormycosis have been used over the years, and more such strategies are urgently needed in view of its raging rise [9]. It is essential to have a clear understanding of the pathogenesis of this disease to facilitate these efforts and ensure appropriate management. Therefore, in this chapter, we will provide an overview of the pathogenesis of mucormycosis, focusing on rhino-orbitalcerebral mucormycosis (ROCM) in patients with COVID-19.

## 4.2 Host Risk Factors

Mucormycosis generally occurs in immunocompromised individuals with defined risk factors. Table 4.1 shows the risk factors and underlying diseases associated with mucormycosis. Diabetes mellitus is the most common underlying disease, followed by hematological malignancies and solid organ transplants [17]. Hyperglycemia is associated with defective neutrophil migration, chemotaxis, and phagocytosis [18, 19]. Other predisposing factors associated with mucormycosis are patients receiving iron chelation therapy and steroids. In India, 3-26% of mucormycosis cases were recorded from the immunocompetent host, compared to 18-19% globally and they usually present with cutaneous or isolated renal mucormycosis [17]. Trauma is the most common

S. Singh  $\cdot$  R. Kanaujia  $\cdot$  S. M. Rudramurthy ( $\boxtimes$ ) Department of Medical Microbiology, Postgraduate Institute of Medical Institution and Research, Chandigarh, India

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2022 N. Gupta, S. G. Honavar (eds.), *Rhino-Orbito-Cerebral Mucormycosis*, https://doi.org/10.1007/978-981-16-9729-6\_4

Due dismosting and dition	The predominant site of infection	Maat aanuman anaaisa	Reference
Predisposing condition		Most common species	
Diabetes, hyperglycemia, and ketoacidosis	ROCM	Rhizopus species (R. arrhizus, R. microsporus, R. homothallicus)	[10]
Hematological and solid organ malignancy	Pulmonary, ROCM	<i>Rhizopus</i> species, <i>Cunninghamella</i> species, <i>Lichtheimia</i> species., <i>Cokeromyces</i> species	[11]
Solid-organ transplant, HSCT	Pulmonary, ROCM	Rhizopus species, Cunninghamella species, Lichtheimia species	[11]
Neutropenia	Pulmonary, disseminated	Rhizopus species, Lichtheimia species. Cunninghamella species	[11]
Iron overload and iron chelation therapy	Pulmonary, disseminated	Rhizopus species, Lichtheimia species, Cunninghamella species	[1]
Corticosteroids	ROCM, pulmonary, disseminated	<i>Rhizopus</i> species, <i>Cunninghamella</i> species, <i>Lichtheimia</i> species	[1]
Malnutrition, preterm neonates, neonates with suspected necrotizing enterocolitis	Gastrointestinal	Rhizopus species, Lichtheimia species	[12]
Breach of skin (trauma due to accidents, burns, catheterization, injection site)	Cutaneous	Apophysomyces and Saksenaea species Syncephalastrum species	[13]
New risk factors:			
SARS-CoV-2, long term steroid use for the treatment	ROCM, pulmonary	Rhizopus species (R. arrhizus, R. microsporus, R. homothallicus), Lichtheimia species	[14]
Post pulmonary tuberculosis/ COPD	Pulmonary mucormycosis	Rhizopus species	[15]
Breakthrough mucormycosis after voriconazole treatment	ROCM, pulmonary	Rhizopus species	[16]
Immunocompetent host in the Indian subcontinent	Renal mucormycosis	Rhizopus and Apophysomyces species	[1]

 Table 4.1
 Risk factors for mucormycosis

*ROCM* Rhino-orbito-cerebral mucormycosis, *SARS-CoV-2* severe acute respiratory syndrome coronavirus 2, *COPD* chronic obstructive pulmonary disease, *HSCT* Hematopoietic Stem Cell Transplantation

risk factor associated with cutaneous mucormycosis [11]. Newer risk-factor like SARS-CoV-2 is emerging. The virus itself acts as a risk factor by decreasing immunity and irrational use of steroids for the treatment, making individuals susceptible to the infection [14].

## 4.3 Virulence Factors of Mucorales

Virulence factors are an essential part of Mucorales that allow these species to invade and survive within the host tissue during infection. The critical virulence traits essential to establish disease are summarized in Table 4.2.

## 4.4 Pathogen Entry

The agents of mucormycosis can gain entry to host tissues via various modes. The primary barriers against an invasion of any external pathogen include nasal mucosa, sinus cavities, skin, and endothelium layers. Typically, Mucorales are incapable of penetrating intact skin. Still, any breach in mucosal continuity by trauma or injury can promote fungal adherence to components of the basal epithelial cell layer such as laminin and type IV collagen [28]. Inhalation of Mucorales sporangiospores is the main entry portal. While the development of mucormycosis is not seen in immunocompetent hosts, those with risk factors can develop a progressive infection at various

Virulence traits	Function	Mechanism of virulence by Mucorales	Reference
Iron metabolism-rela	ated virulence traits		
Reductase/permease	systems		
Low-affinity iron	Performs in iron-rich	Patients with DKA (pH -7.3-6.88) reduce ferric ions to	[20]
reductase	environments and reduces iron	ferrous and facilitate Rhizopus spp. growth	
High-affinity ferric reductase	Functions in iron- depleted environments and reduces iron	In DKA, carrier molecules bind to iron, and free iron is available only in submicromolar concentrations. The high-affinity ferric reductase facilitates the uptake of bound iron by <i>Rhizopus</i>	[20]
Other ferric reductases	Reduce iron	Allow iron uptake in iron-depleted environments and facilitates <i>Rhizopus</i> spp. growth	[21]
Copper oxidase (Cu-oxidase)	Oxidize iron	Trans cell membrane transport of iron	[20]
High-affinity iron permease (FTR1)	Helps in the transport of iron	Trans cell membrane transport of iron	[22]
Role of siderophores	in Mucormycetes iron met	tabolism	-
Siderophore permeases	Provide iron to cells by chelating iron	This siderophore supplies iron by direct transfer across the plasma membrane for <i>Rhizopus spp.</i> growth	[23]
Rhizoferrin	Siderophore that chelates iron	This siderophore supplies iron by the energy-dependent process for <i>Rhizopus</i> spp. growth	[24]
Deferoxamine	Deferoxamine extracts ferric iron from transferrin	It acts as a xenosiderophore. It attaches on the <i>Rhizopus</i> spp. and functions by transporting iron by energy-dependent reduction of iron	[25]
Role of hemin utiliza	ution in Mucormycetes iron	metabolism	
Heme oxygenase	Iron uptake from heme	Obtain iron from host hemoglobin and allow angioinvasion of <i>Rhizopus</i> spp.	[22]
Others			
<ul><li>Proteinases</li><li>Aspartic proteinases</li><li>Subtilase protein</li></ul>	Protein lysis	Degrade host epithelium and contribute to the angioinvasive nature of the disease	[26]
Rhizoxin	Mycotoxin	Antimitotic metabolite that kills host cells	[27]
	1	1	

Table 4.2 Virulence factors for Mucorales

GRP glucose receptor protein

sites (Fig. 4.1). In patients with burns or other skin trauma/maceration, the spores can directly penetrate deeper tissues. Spores can also gain entry via application of non-sterile adhesive tapes and surgical dressings [29, 30]. Furthermore, mucormycosis spores can also gain access via contaminated tongue depressors or wooden applicators [31].

#### 4.4.1 Spore Germination

The germination of spores and hyphal formation is critical for establishing infection in the host. In immunocompetent mice, pulmonary alveolar macrophages harvested from the lungs have been found to effectively ingest and prevent the germination of *R. arrhizus* spores both *in vitro* and after intranasal infection. In contrast, those derived from immunosuppressed mice are unable to do so [32].

#### 4.4.2 Attachment and Invasion

The next step is attaching to extracellular protein matrices such as laminin and collagen IV. Any epithelial cell damage (e.g., due to COVID-19) can expose them for interaction with inhaled/ingested spores [28]. *Rhizopus* 

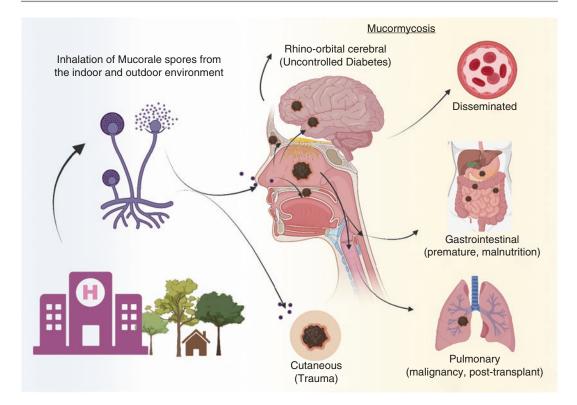


Fig. 4.1 Overview of pathogen entry and clinical presentations in mucormycosis

spp. can also attach to and invade the endothelium by recognizing a specific host receptor, the glucose-regulator protein 78 (GRP78) [33, 34]. This heat shock protein is a component of the host stress-related responses that helps in the specific binding of Mucorales germlings via the fungal ligands of the spore coat protein (CotH) family [35].

During SARS-CoV-2 infection, the endoplasmic reticulum stress is stimulated, which drives GRP78 synthesis [36]. In a recent study, nearly five times higher serum GRP78 was observed in patients with SARS-CoV-2 infection compared to the control group [37]. The GRP78 mediated attachment and internalization of the SARS-CoV-2 virus has also been reported [38]. Thus, upregulation of GRP78 enhanced by SARS-CoV-2 spike protein for its entry and by diffuse endothelial is seen in COVID-19 may also facilitate the attachment and invasion by Mucorales [39].

#### 4.5 Immunopathogenesis

In a healthy host, the dormant/resting spores resist phagocytic killing but, swollen/germinating spores or hyphal forms are prone to degradation by immune cells. After endothelial penetration, Mucorales also encounter platelets that adhere to the spores and suppress germination, which contributes to hyphal damage, potentially hampering the fungal growth by adhering and damaging the hyphae or indirectly, by secreting inflammatory cytokines [40]. Various studies have investigated the interaction of Mucorales with host immunity (both innate and adaptive), and the details are summarized in Table 4.3.

The role of cytokines in modifying host response against the Mucorales originates from experimental studies, and its immunological interactions are depicted in Fig. 4.2.

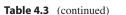
Interleukin-1 beta (IL-1 $\beta$ ) plays a pivotal role in response to pathogenic fungi with a significant role

Categories of the immune response with cellular type	Function	Immune evasion by Mucorales	Reference
Innate immune response	·	· · · · · · · · · · · · · · · · · · ·	
Bronchial alveolar macrophages (BAM)	The first line of defense and prevents germination of Mucorale spores and has a role in iron restriction	<ul> <li>Inhibits phagosome maturation by cell wall melanin</li> <li>Upregulation of iron acquisition genes</li> <li>Upregulation of genes involved in nutrient assimilation to allow the use of alternate nutrients in a hostile macrophage environment</li> </ul>	[32, 41–43]
Epithelial cells	The first line of defense and prevents the entrance of Mucorales spores	Mucorales damage the epithelial cells and upregulate epidermal growth factor receptor (EGFR) on the surface of epithelial cells, which promotes disease	[44, 45]
Polymorphonuclear leukocytes (PMNs) or neutrophil granulocytes	<ul> <li>Neutrophils produce cytokines to activate other immune cells</li> <li>They can also form tight clusters around the spores containing them in structures resembling early granulomas</li> <li>Neutrophil extracellular traps (NETs), reactive oxygen species and cationic peptides also damage both spore and hyphae</li> </ul>	<ul> <li>Sialic acids in the cell wall protect against phagocytosis</li> <li>Resting spores and the hyphae can reduce neutrophils chemotaxis</li> </ul>	[46-48]
Natural killer (NK) cells	Natural killer (NK) cells recognize Mucorales and cause hyphal damage on activation	Mucorales hyphae have an immunosuppressive effect on dendritic cells and reduce the release of immunomodulatory molecules	[49–52]
Platelets	<ul> <li>Platelet recognition of Mucorales causes inhibition of spore germination and hyphal growth by secretion of various cytokines and chemokines</li> <li>They also bind to and activate other immune cells</li> </ul>	Excessive thrombosis seen in mucormycosis causes thrombocytopenia which also makes surgical interventions difficult	[40, 53]
Endothelial cells	Endothelial cells surround the innermost layer of blood vessels. It helps in hyphae recognition	The glucose-regulated protein 78 (GRP78) receptor present on endothelial cell surface facilitate angioinvasion by Mucorales	[33, 54, 55]
Dendritic cells (D.C.s)	<ul> <li>Dendritic cells link the innate immunity with the adaptive immunity</li> <li>Production of IL23 by dendritic cells induces Th17 cells, which further promote neutrophil response</li> </ul>	The resting (dormant) spores of Mucorales stimulate the maturation of dendritic cells	[56, 57]

 Table 4.3
 Interaction of the Mucorales with the immune system

(continued)

Categories of the immune			
response with cellular type	Function	Immune evasion by Mucorales	References
Adaptive immune response			
T cells	<ul> <li>Both CD4 and CD8 cells produce cytokines that mediate hyphal damage</li> <li>Th17 cells produce IL 17, which promotes the antifungal defense of neutrophils</li> <li>Release of interleukins (IL-4, IL-10, and IL-17) and IFNγ caused damage to hyphae</li> </ul>		[58]



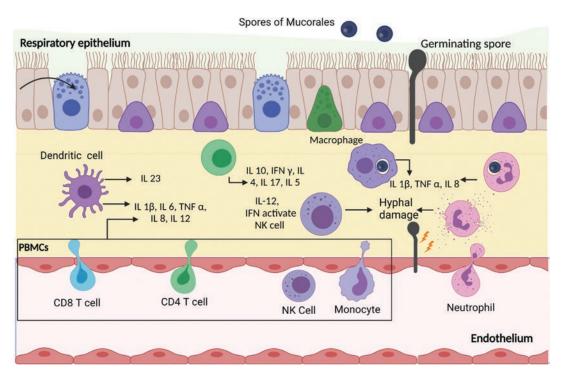


Fig. 4.2 Schematic representation of immune cells involved in the pathogenesis of mucormycosis

in the induction of other proinflammatory responses, hematopoiesis, Th17 cell differentiation, etc. [59]. Tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) has a dual role as an activator of inflammatory response and immunosuppression by mediating host apoptosis. At the same time, Interleukin-6 mediates leukocyte trafficking and production of acute-phase reactants while promoting T-cell proliferation and B cell responses [60]. The IL-12 promotes Th1 cell response, natural killer (N.K.) cell activation, Dendritic cell maturation, and production of interferon- $\gamma$  (IFN- $\gamma$ ) and chemoattractant proteins like IL-8 also recruit immune cells, thereby mediating an effective immune response [60].

# 4.6 Factors Affecting the Pathogenesis of Mucormycosis in COVID-19

- Hyperglycemia: The SARS-CoV-2 impairs pancreatic beta-cell function and precipitates acute diabetic ketoacidosis (DKA) [61], and DKA has been was observed even with T2DM cases [62]. In the past, infections with SARS-CoV-1 have been seen to be associated with hyperglycemia persisting for as long as 3 years indicating long-term damage to pancreatic β-cells by these viruses [63]. Thus, SARS-CoV-2 mediated DKA could present even weeks or months after apparent recovery from the disease substantiating the late onset of CAM [14].
- Corticosteroids: Worsening of glycemic con-٠ trol can also occur secondary to the use of systemic steroids and antiviral agents in the management of COVID-19. A higher incidence of mucormycosis infections has been observed in patients who received corticosteroids during COVID-19 treatment, often in terms of inappropriate doses ( $\geq 6 \text{ mg/kg dexa-}$ methasone), duration (more than 10 days) or even when not indicated [14, 64]. Impaired macrophage and neutrophil function due to steroids explains suppressed antifungal immunity.
- Iron metabolism: Dysregulated iron metabolism is known to occur during COVID-19 [65]. The interaction between viral particles and hemoglobin perpetuates a cascade of dysfunctional hemoglobin synthesis, hemolysis, and heme accumulation with increased serum ferritin also seen in response to the inflammation [66–68]. The activation of macrophages and high IL-6 secretion also accentuates hyperferritinemia. This excessive intracellular free iron generates free radicals that cause endothelial destruction, leading to endothelitis, promoting fungal invasion [69].
- *Immune dysfunction*: The mucociliary clearance by the nasal epithelium is the primary

innate immune defense against inhaled microorganisms. This clearance is delayed in COVID-19, allowing Mucorales spores to attach to the nasal epithelium [70]. Among innate immune cells, lymphopenia has been observed among COVID-19 patients, but its role in escalating the host susceptibility to mucormycosis is not clear due to insufficient data [71] (Fig. 4.3).

## 4.7 Common Misconceptions

The unprecedented epidemic of mucormycosis in the background of the COVID-19 pandemic has given rise to various misconceptions regarding CAM. The term "black fungus" has been used incorrectly and indiscriminately to indicate mucormycosis infection, which should be discouraged. Although the clinical presentation of mucormycosis comprises eschar and black necrotic lesions, the term "black fungus" actually denotes a completely different group of fungi that produce the pigment melanin, resulting in black hyphae, which are not seen in Mucorales. Another misconception is regarding the source of infection. The use of respirators, oxygenation during COVID-19 management, reuse of masks, etc. have been implicated as potential sources of infection, but this lacks scientific backing. There is also no evidence of human-to-human transmission, and this disease is acquired via Mucorales spores present ubiquitously in the environment, both indoors and outdoors. The prophylactic use of antifungals to prevent this infection in COVID-19 is also strongly discouraged since the incidence of infection is <10% in any COVID-19 cohort [8].

#### 4.8 Future Perspectives

Although CAM is a new entity, various reports describing this infection have emerged over the past few months. Unfortunately, there is still a

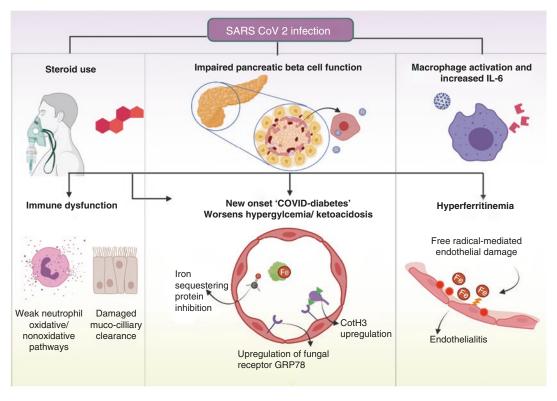


Fig. 4.3 Factors affecting pathogenesis of mucormycosis in patients with SARS CoV-2 infection

shortage of systematic prospective studies evaluating the various factors involved in disease pathogenesis and outcome, particularly in comparison to non-COVID mucormycosis cases. Studying the nature and extent of immunological dysfunction in COVID-19 and its impact on the pathogenesis of mucormycosis is a promising area of research.

# 4.9 Conclusion

Although host-immune response and inflammatory signalling against Mucorales have been established, the underlying molecular mechanism responsible for producing proinflammatory cytokines and activating inflammatory response against Mucorales is unknown. In the context of CAM, this field is even more unexplored. An understanding of the pathogenesis of CAM can provide new perspectives for developing novel diagnostic and treatment modalities.

## References

- Hariprasath P, Arunaloke C. Global epidemiology of mucormycosis. J Fungi. 2019;5(1). https://doi. org/10.3390/jof5010026.
- Skiada A, Pagano L, Groll A, Zimmerli S, Dupont B, Lagrou K, et al. Zygomycosis in Europe: analysis of 230 cases accrued by the registry of the European Confederation of Medical Mycology (ECMM) Working Group on Zygomycosis between 2005 and 2007. Clin Microbiol Infect. 2011;17(12):1859–67. https://doi.org/10.1111/j.1469-0691.2010.03456.x.
- Arunaloke C. Fungal infections in Asia: eastern frontier of mycology. ECAB Clinical update infectious Disease. 2014. Publisher: Elsevier India
- Arunaloke C, Harsimran K, Jayanthi S, Rudramurthy Shivaprakash M, Atul P, Prakash S, et al. Epidemiology and clinical outcomes of invasive mould infections in Indian intensive care units (FISF study). J Crit Care. 2019;51:64–70. https://doi.org/10.1016/j. jcrc.2019.02.005.
- Antonella F, Zan M, Alexander RR, Assas Bakri M. Cytokine storm and COVID-19: a chronicle of proinflammatory cytokines: cytokine storm: the elements of rage! Open Biol. 2020; https://doi.org/10.1098/ rsob.200160.

- Nilam M, Hunter Christopher A. Cytokine storms: understanding COVID-19. Immunity. 2020:19–25. https://doi.org/10.1016/j.immuni.2020.06.017.
- Attilio C, Emidio T, Salvatore C. COVID-19: hemoglobin, iron, and hypoxia beyond inflammation. A narrative review. Clin Pract. 2020;10(2):24–30. https://doi.org/10.4081/CP.2020.1271.
- Rudramurthy Shivaprakash M, Martin H, Meis Jacques F, Cornely Oliver A, Valliappan M, Pierre GJ, et al. ECMM/ISHAM recommendations for clinical management of COVID-19 associated mucormycosis in low-and middle-income countries. https://doi. org/10.1111/MYC.13335.
- Arunaloke C, Shreya S. Management of mucormycosis. Curr Fungal Infect Rep. 2020;14(4):348–60. https://doi.org/10.1007/S12281-020-00406-2.
- Patel A, Kaur H, Xess I, Michael JS, Savio J, Rudramurthy S, et al. A multicentre observational study on the epidemiology, risk factors, management and outcomes of mucormycosis in India. Clin Microbiol Infect. 2020;26(7):944.e9–944.e15. https:// doi.org/10.1016/j.cmi.2019.11.021.
- Jeong W, Keighley C, Wolfe R, Lee WL, Slavin MA, Kong DCM, et al. The epidemiology and clinical manifestations of mucormycosis: a systematic review and meta-analysis of case reports. Clin Microbiol Infect. 2019;25(1):26–34. https://doi.org/10.1016/j. cmi.2018.07.011.
- Sushma P, Chirla DK, Narendar K, Mukul V, Samal SC. Unsuspected invasive neonatal gastrointestinal mucormycosis: a clinicopathological study of six cases from a tertiary care hospital. J Indian Assoc Pediatr Surg. 2012;17(4):153. https://doi. org/10.4103/0971-9261.102329.
- Robin K. Primary cutaneous Zygomycosis in India. Indian J Surg. 2012;74(6):468–75. https://doi. org/10.1007/s12262-012-0429-4.
- Atul P, Ritesh A, Rudramurthy Shivaprakash M, Manoj S, Immaculata X, Ratna S, et al. Multicenter epidemiologic study of coronavirus disease-associated mucormycosis, India. Emerg Infect Dis. 2021;27(9). https://doi.org/10.3201/eid2709.210934.
- Hariprasath P, Kumar GA, Mandya RS, Pankaj S, Immaculata X, Jayanthi S, et al. A prospective multicenter study on mucormycosis in India: epidemiology, diagnosis, and treatment. Med Mycol. 2019;57(4):395–402. https://doi.org/10.1093/mmy/ myy060.
- Sushil M, Chetanya S, Sanjay T, Sreenivas V, Kabra SK, Immaculata X, et al. Oral voriconazole versus intravenous low dose amphotericin B for primary antifungal prophylaxis in pediatric acute leukemia induction. J Pediatr Hematol Oncol. 2011;33(8):e333–41. https://doi.org/10.1097/MPH.0b013e3182331bc7.
- Hariprasath P, Arunaloke C. Epidemiology of mucormycosis in India. Microorganisms. 2021;9(3):1–12. https://doi.org/10.3390/ MICROORGANISMS9030523.
- Jagdish C, Mandeep K, Nidhi S, Punia RPS, Singhal Surinder K, Attri Ashok K, et al. Mucormycosis:

battle with the deadly enemy over a five-year period in India. J Fungi (Basel, Switzerland). 2018;4(2). https://doi.org/10.3390/jof4020046.

- Corzo-León DE, Chora-Hernández LD, Rodríguez-Zulueta AP, Walsh TJ. Diabetes mellitus as the major risk factor for mucormycosis in Mexico: epidemiology, diagnosis, and outcomes of reported cases. Med Mycol. 2018;56(1):29–43. https://doi.org/10.1093/ MMY/MYX017.
- Howard DH. Acquisition, transport, and storage of iron by pathogenic fungi. Clin Microbiol Rev. 1999;12(3):394–404. https://doi.org/10.1128/ CMR.12.3.394.
- Artis WM, Fountain JA, Delcher HK, Jones HE. A mechanism of susceptibility to mucormycosis in diabetic ketoacidosis transferrin and iron availability. Diabetes. 1982;31(12):1109–14. https://doi. org/10.2337/diacare.31.12.1109.
- 22. Ibrahim AS, Teclegiorgis G, Lin L, Guanpingsheng L, Husseiny Mohamed I, Skory Christopher D, et al. The high affinity iron permease is a key virulence factor required for Rhizopus oryzae pathogenesis. Mol Microbiol. 2010;77(3):587–604. https://doi.org/10.1111/j.1365-2958.2010.07234.x.
- Ibrahim AS. Spellberg Brad, Edwards John. Iron acquisition: a novel perspective on mucormycosis pathogenesis and treatment. Curr Opin Infect Dis. 2008:620–5. https://doi.org/10.1097/ QCO.0b013e3283165fd1.
- Thieken A. Rhizoferrin: a complexone type siderophore of the mocorales and entomophthorales (zygomycetes). FEMS Microbiol Lett. 1992;94(1–2):37–41. https://doi.org/10.1016/0378-1097(92)90579-D.
- Boelaert JR, de Locht M, Van Cutsem J, Kerrels V, Cantinieaux B, Verdonck A, et al. Mucormycosis during deferoxamine therapy is a siderophore-mediated infection. In vitro and in vivo animal studies. J Clin Invest. 1993;91(5):1979–86. https://doi.org/10.1172/ JCI116419.
- Farley PC, Sullivan PA. The Rhizopus oryzae secreted aspartic proteinase gene family: an analysis of gene expression. Microbiology. 1998;144(8):2355–66. https://doi.org/10.1099/00221287-144-8-2355.
- White JD, Blakemore PR, Green NJ, Bryan HE, Holoboski Mark A, Keown Linda E, et al. Total synthesis of rhizoxin D, a potent antimitotic agent from the fungus Rhizopus chinensis. J Org Chem. 2002;67(22):7750–60. https://doi.org/10.1021/ jo020537q.
- Bouchara JP, Oumeziane NA, Lissitzky JC, Larcher G, Tronchin G, Chabasse D. Attachment of spores of the human pathogenic fungus Rhizopus oryzae to extracellular matrix components. Eur J Cell Biol. 1996;70(1):76–83.
- Rammaert B, Lanternier F, Zahar JR, Dannaoui E, Bougnoux ME, Lecuit M, et al. Healthcare-associated mucormycosis. Clin Infect Dis. 2012;54(Suppl 1). https://doi.org/10.1093/CID/CIR867.
- 30. Alsuwaida K. Primary cutaneous mucormycosis complicating the use of adhesive tape to secure the

endotracheal tube. Can J Anaesth. 2002;49(8):880–2. https://doi.org/10.1007/BF03017426.

- 31. Enrique M-P, Rodríguez-Tudela JL, García DJJ, Alfonso M-L, Luis T, Jesús U, et al. Outbreak of gastric mucormycosis associated with the use of wooden tongue depressors in critically ill patients. Intensive Care Med. 2004;30(4):724–8. https://doi. org/10.1007/S00134-003-2132-1.
- Waldorf AR, Ruderman N, Diamond RD. Specific susceptibility to mucormycosis in murine diabetes and bronchoalveolar macrophage defense against Rhizopus. J Clin Invest. 1984;74(1):150–60. https:// doi.org/10.1172/JCI111395.
- 33. Mingfu L, Brad S, Phan QT, Yue F, Yong F, Lee AS, et al. The endothelial cell receptor GRP78 is required for mucormycosis pathogenesis in diabetic mice. J Clin Invest. 2010;120(6):1914–24. https://doi. org/10.1172/JCI42164.
- Ibrahim Ashraf S, Brad S, Walsh Thomas J, Kontoyiannis Dimitrios P. Pathogenesis of mucormycosis. Clin Infect Dis. 2012;54(SUPPL 1). https://doi. org/10.1093/cid/cir865.
- 35. Teclegiorgis G, Sondus A, Soliman Sameh SM, Yiyou G, Jeon HH, Lina Z, et al. Anti-CotH3 antibodies protect mice from mucormycosis by prevention of invasion and augmenting opsonophagocytosis. Sci Adv. 2019;5(6). https://doi.org/10.1126/sciadv.aaw1327.
- 36. Köseler A, Sabirli R, Gören T, Türkçüer I, Kurt Ö. Endoplasmic reticulum stress markers in SARS-COV-2 infection and pneumonia: case-control study. In Vivo. 2020;34(3 Suppl):1645–50. https://doi. org/10.21873/INVIVO.11956.
- Sabirli R, Koseler A, Goren T, Turkcuer I, Kurt O. High GRP78 levels in Covid-19 infection: a case-control study. Life Sci. 2021;265. https://doi. org/10.1016/J.LFS.2020.118781.
- Ibrahim IM, Abdelmalek DH, Elshahat ME, Elfiky AA. COVID-19 spike-host cell receptor GRP78 binding site prediction. J Infect. 2020;80(5):554–62. https://doi.org/10.1016/J.JINF.2020.02.026.
- Baldin C, Ibrahim AS. Molecular mechanisms of mucormycosis-the bitter and the sweet. PLoS Pathog. 2017;13(8). https://doi.org/10.1371/JOURNAL. PPAT.1006408.
- Susanne P, Barbara K, Kehrel BE, Dierich MP, Walter N, Cornelia L-F. Potential antifungal effects of human platelets against zygomycetes in vitro. J Infect Dis. 2009;200(7):1176–9. https://doi.org/10.1086/605607.
- 41. Kaswara K, Hea-Reung P, Hans-Martin D, Christine S, Kerstin V, Thilo FM. Virulent strain of Lichtheimia corymbifera shows increased phagocytosis by macrophages as revealed by automated microscopy image analysis. Mycoses. 2014;57:56–66. https://doi. org/10.1111/myc.12237.
- 42. Andrianaki AM, Irene K, Kalliopi T, Clara B, Elias D, Soliman Sameh SM, et al. Iron restriction inside macrophages regulates pulmonary host defense against Rhizopus species. Nat Commun. 2018;9(1):3333. https://doi.org/10.1038/s41467-018-05820-2.

- 43. Calo S, Nicolás FE, Lee SC, Vila A, Cervantes M, Torres-Martinez S, et al. A non-canonical RNA degradation pathway suppresses RNAi-dependent epimutations in the human fungal pathogen Mucor circinelloides. PLoS Genet. 2017;13(3). https://doi. org/10.1371/JOURNAL.PGEN.1006686.
- Ghuman H, Voelz K. Innate and adaptive immunity to mucorales. J Fungi. 2017;3(3):48. https://doi. org/10.3390/jof3030048.
- 45. Watkins TN, Teclegiorgis G, Marc S, Shetty AC, Graf KT, Abdullah A, et al. Inhibition of EGFR signaling protects from mucormycosis. MBio. 2018;9(4). https://doi.org/10.1128/mBio.01384-18.
- Venizelos P. Neutrophil extracellular traps in immunity and disease. Nat Rev Immunol. 2018;18(2):134–47. https://doi.org/10.1038/nri.2017.105.
- 47. Chinn RY, Diamond RD. Generation of chemotactic factors by Rhizopus oryzae in the presence and absence of serum: relationship to hyphal damage mediated by human neutrophils and effects of hyperglycemia and ketoacidosis. Infect Immun. 1982;38(3):1123–9. https://doi.org/10.1128/iai.38.3.1123-1129.1982.
- Waldorf AR, Diamond RD. Neutrophil chemotactic responses induced by fresh and swollen Rhizopus oryzae spores and Aspergillus fumigatus conidia. Infect Immun. 1985;48(2):458–63. https://doi.org/10.1128/ iai.48.2.458-463.1985.
- Eric V, Elena T, Myriam B, Thierry W, Sophie U. Functions of natural killer cells. Nat Immunol. 2008;9(5):503–10. https://doi.org/10.1038/ni1582.
- Yokoyama WM. Natural killer cell immune responses. Immunol Res. 2005;32(1–3):317–26. https://doi. org/10.1385/IR:32:1-3:317.
- 51. Stanislaw S, Lars T, Susanne P, Cornelia L-F, Mitra H, Frauke R, et al. Rhizopus oryzae hyphae are damaged by human natural killer (NK) cells, but suppress NK cell mediated immunity. Immunobiology. 2013;218(7):939–44. https://doi.org/10.1016/j. imbio.2012.10.013.
- Stanislaw S, Andreas S, Asuman D, Cornelia L-F, Thomas L. Natural killer cell-mediated damage of clinical isolates of mucormycetes. Mycoses. 2016;59(1):34–8. https://doi.org/10.1111/myc.12431.
- Jenne CN, Paul K. Platelets in inflammation and infection. Platelets. 2015;26(4):286–92. https://doi. org/10.3109/09537104.2015.1010441.
- Aird WC. Endothelial cell heterogeneity. Cold Spring Harb Perspect Med. 2012;2(1):a006429. https://doi. org/10.1101/cshperspect.a006429.
- 55. Evgenii S, Ulrike B, Christina P, Alexander M, Helmut E, Justin H, et al. In situ validation of the endothelial cell receptor GRP78 in a case of rhinocerebral mucormycosis. Antimicrob Agents Chemother. 2018;62(5). https://doi.org/10.1128/AAC.00172-18.
- Lambrecht BN, Prins JB, Hoogsteden HC. Lung dendritic cells and host immunity to infection. Eur Respir J. 2001;18(4):692–704.
- 57. Sebastian W, Vanessa T, Philipp W, Paul W, Johannes E, Maria W-GA, et al. Mucorales spores induce a pro-

inflammatory cytokine response in human mononuclear phagocytes and harbor no rodlet hydrophobins. Virulence. 2017;8(8):1708–18. https://doi.org/10.108 0/21505594.2017.1342920.

- 58. Leonardo P, Daniela V, Patrizia B, Giovanni R, Fabio F, Eleonora Z, et al. Mucorales-specific T cells emerge in the course of invasive mucormycosis and may be used as a surrogate diagnostic marker in highrisk patients. Blood. 2011;118(20):5416–9. https:// doi.org/10.1182/blood-2011-07-366526.
- Griffiths JS, Giorgio C, Kotowicz NK, Jemima H, Richardson JP, Naglik Julian R. Role for IL-1 family cytokines in fungal infections. Front Microbiol. 2021;12:633047. https://doi.org/10.3389/ FMICB.2021.633047.
- Harlene G, Kerstin V. Innate and adaptive immunity to mucorales. J Fungi. 2017;3(3):48. https://doi. org/10.3390/JOF3030048.
- 61. Smith SM, Boppana A, Traupman JA, Unson E, Maddock DA, Chao K, et al. Impaired glucose metabolism in patients with diabetes, prediabetes, and obesity is associated with severe COVID-19. J Med Virol. 2021;93(1):409–15. https://doi.org/10.1002/ JMV.26227.
- 62. Pal R, Banerjee M, Yadav U, Bhattacharjee S. Clinical profile and outcomes in COVID-19 patients with diabetic ketoacidosis: a systematic review of literature. Diabetes Metab Syndr. 2020;14(6):1563–9. https:// doi.org/10.1016/J.DSX.2020.08.015.
- 63. Yang JK, Lin SS, Ji XJ, Guo LM. Binding of SARS coronavirus to its receptor damages islets and causes acute diabetes. Acta Diabetol. 2010;47(3):193–9. https://doi.org/10.1007/S00592-009-0109-4.
- 64. Mrittika S, Honavar Santosh G, Rolika B, Sabyasachi S, Raksha R, Usha K, et al. Epidemiology, clinical profile, management, and outcome of COVID-19-associated rhino-orbital-cerebral mucormycosis in 2826 patients in India collaborative OPAI-IJO study

on mucormycosis in COVID-19 (COSMIC), report 1. Indian J Ophthalmol. 2021;69(7):1670–92. https:// doi.org/10.4103/IJO.IJO\_1565\_21.

- 65. Anson J, Shagun S, Ajoy R, Yathin K, Saurabh A, Sunanda R. Current understanding in the pathophysiology of SARS-CoV-2-associated rhino-Orbitocerebral mucormycosis: a comprehensive review. J Maxillofac Oral Surg. 2021;1:1–8. https://doi. org/10.1007/S12663-021-01604-2.
- 66. Amer M, Ashraf SM. Is hemoglobin the missing link in the pathogenesis of COVID-19? Anaesth Pain Intensive Care. 2020;24(1):9–12. https://doi. org/10.35975/APIC.V24I1.1216.
- 67. Carlo P, Elena B, Roberto B, Giacomo C, Maria GG, Yehuda S, et al. COVID-19 as part of the hyperferritinemic syndromes: the role of iron depletion therapy. Immunol Res. 2020;68(4):213. https://doi. org/10.1007/S12026-020-09145-5.
- Jenifer G-P, Mitchell W, James K, Xian W, Jacob S, Palmer Andre F, et al. Hyperferritinemia in critically ill COVID-19 patients – is ferritin the product of inflammation or a pathogenic mediator? Clin Chim Acta. 2020;509:249. https://doi.org/10.1016/J. CCA.2020.06.033.
- Vlahakos VD, Marathias KP, Nikolaos A, Vlahakos Demetrios V. Hyperferritinemia in patients with COVID-19: an opportunity for iron chelation? Artif Organs. 2021;45(2):163–7. https://doi.org/10.1111/ AOR.13812.
- Koparal M, Kurt E, Altuntas EE, Dogan F. Assessment of mucociliary clearance as an indicator of nasal function in patients with COVID-19: a cross-sectional study. Eur Arch Otorhinolaryngol. 2021;278(6):1863– 8. https://doi.org/10.1007/S00405-020-06457-Y.
- Soheil T, Taha R, Wei EX, Mohammad R. Lymphopenia during the COVID-19 infection: what it shows and what can be learned. Immunol Lett. 2020;225: 31–2. https://doi.org/10.1016/j.imlet.2020.06.013.