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MRI: Quantitative Evaluation of Diseased Tissue by Viscoelastic Imaging Systems

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Abstract

Magnetic resonance elastography (MRE) and ultrasound elastography (USE) are imaging techniques that noninvasively quantify the mechanical properties of tissue by using magnetic resonance imaging and ultrasound imaging systems. In this study, we aim to develop a system that can quantitatively obtain details such as the viscoelasticity of small organs and tissues in the body by using clinical magnetic resonance imaging (MRI) and MR microscopy. By evaluating MRE and USE using a soft tissue-equivalent gel phantom with a known viscoelastic coefficient, we aim to investigate the characteristics of both devices and promote their standardization. The purpose of this study is to confrm the frequency characteristics of the developed phantom and optimize the scatterer material for ultrasound measurement. We confrmed that the phantoms are in good agreement with a physical model of the liver, and the developed phantoms are considered effective for the quantitative assessment of the MRE and USE systems.

Keywords

Magnetic resonance imaging · Magnetic resonance elastography · Ultrasound elastography · Phantom Quantitative assessment

39.1 Introduction

The mechanical property of a tissue is related to physiological and pathological states. Magnetic resonance elastography (MRE) and ultrasound elastography (USE) are imaging techniques that non-invasively quantify the mechanical properties

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of tissue by using magnetic resonance imaging and ultrasound imaging systems [[1,](#page-4-0) [2\]](#page-4-1). It is expected that measuring the mechanical properties of tissues will be useful in the diagnosis of diseases such as hepatic fbrosis and cancer. MRE visualizes shear-wave patterns within a tissue using a modifed phasecontrast MR sequence. In order to generate shear waves within the tissue, external vibration systems are used (Fig. [39.1](#page-0-0)). The local quantitative values of tissue viscoelasticity (stiffness) are calculated from the shear-wave pattern by using an inversion algorithm [\[3–](#page-4-2)[5\]](#page-4-3). We have been developing an MRE system using clinical MRI and MR microscope for measuring viscoelasticity at multi-scale and multi-frequency (Fig. [39.2](#page-1-0)) [[6,](#page-4-4) [7](#page-4-5)].

The measured viscoelasticity can be used as an imaging biomarker [[8\]](#page-4-6). A quantitative phantom is required to assess the accuracy and repeatability of elastography systems. We have been developing tough and stable polyacrylamide (PAAm) gel phantoms for this purpose [\[9](#page-4-7)[–12](#page-4-8)]. We developed a phantom with viscoelasticity close to that of living tissue by using glycerin as a solvent. For ultrasonic measurements, a scatterer is necessary for the phantom. The material and concentration of the scatterer are related to the stability of ultrasonic measurement and uniformity of the MRI image.

In this study, we compared the mechanical properties of our phantoms with those of living tissue and optimized the scatterer material and concentration for ultrasound measurement with the developed MRE system and commercial USE system.

Fig. 39.1 Schematic of the MR elastography system

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Fig. 39.2 Multi-modality equipment for measuring viscoelasticity at multi-scale and multi-frequency

mm

Scale

Modality

Rheometer

Mechanical

μm

39.1.1 Development of a Tissue-Mimicking Viscoelastic Phantom for Quantitative Assessment of MRE

We fabricated gel phantom sheets with high loss moduli (*G*″). The phantom sheets were designed to have a storage modulus (*G*^{\prime}) of 1.5 kPa and a loss tangent (tan $\delta = G''/G'$) greater than 0.2 at 60 Hz [[13\]](#page-4-9). The storage and loss moduli of the phantom and a fresh in vitro bovine liver were measured using a parallel-disc rheometer (MCR302, Anton-Parr). We repeated the measurements on these three samples with the strain amplitude set to 1% (Table [39.1](#page-1-1)).

The phantom (diameter: 120 mm, height: 150 mm) was measured with MRE using 3-T MRI (MAGNETOM Skyra, Siemens) at 30, 40, 50, and 60 Hz (Table [39.2\)](#page-1-2) by spin-echo-EPI-MRE pulse sequence (work in progress) and the vibration from a pneumatic driver system with active driver to a passive driver to create the shear wave in the phantom. The passive driver with a diameter of 18 cm was placed on the center of the phantom. The fresh in vitro bovine liver (width: 135 mm, height: 70 mm, length: 250 mm) was measured with MRE using 0.3-T MRI (Hitachi) at 31.25, 62.5, 100, and 200 Hz (Table [39.3](#page-1-3)) by spin-echo-EPI-MRE pulse sequence (a motion encoding gradient (MEG) was added to

Table 39.2 Multi-frequency MRE imaging parameters by 3-T MRI

Rheometer

(vibration freq. $<$ 30 Hz)

cm

Sequence		Spin-echo-EPI-MRE (work in progress)			
Vibrational frequency	Hz	30	40	50	60
Repetition time (TR)	S	4.5	4.5	3.0	3.0
Echo time (TE)	ms	97			
FOV	mm ²	384×384			
Matrix size	pixel	128×128			
Pixel size	mm	3			
Number of slices		15			
Slice thickness	mm	\mathcal{E}			
Temperature	$^{\circ}C$	23			

Table 39.3 Multi-frequency MRE imaging parameters by 0.3-T MRI

SE-EPI using the sequence development environment of Hitachi) and ultrasound system (ACUSON S3000, Siemens) with virtual touch IQ (VTIQ). At high-frequency measurement of the fresh in vitro bovine liver (width: 15 mm, depth: 15 mm, height: 50 mm), MRE using 1-T MR microscope (MR-MICRO, MRTecnology) was used (Table [39.4\)](#page-2-0).

To evaluate the temporal changes, the phantom designed to have a storage modulus of 3 kPa was examined by MRE for the duration of 1 year.

 \times 2 mm

Sequence		Spin-echo-MRE
Vibrational frequency	Hz	200
Repetition time (TR)	S	0.5
Echo time (TE)	ms	21
FOV	mm ²	25×25
Matrix size	pixel	128×128
Pixel size	mm	0.2
Number of slices		1
Slice thickness	mm	1.8
Average		\mathcal{E}
Temperature	\circ C	23

Table 39.4 MRE imaging parameters by 1-T MR microscope

39.1.2 Ultrasound-Based Shear-Wave Speed Measurement on a Highly Viscous Embedded Phantom

The PAAm gel is composed of a three-dimensional network polymer and a large amount of liquid. The storage modulus (stiffness) of the PAAm gel depends mainly on the quantity of acrylamide. Additionally, the density of the threedimensional network polymer depends mainly on the quantity of cross-linker. The loss modulus (viscosity) depends mainly on the ratio of water and glycerin. To make compatible phantoms for MRE and USE, the aluminum oxide powder was added to the PAAm gel for the scatterer.

A highly viscoelastic embedded phantom was measured with US-based shear-wave elastography (SWE). The phantom composed square soft part (background part; width: 130 mm, depth: 130 mm, height: 160 mm) and embed two cylindrical hard parts (embedded part; length: 130 mm, diameter: 10 mm and 20 mm). The weight percent of acrylamide in the embedded part is 1.5 times higher than the background part. In addition, we have created a homogeneous phantom that is content the same as the embedded part.

The SWS was measured with virtual touch quantifcation (VTQ) and VTIQ (Table [39.5\)](#page-2-1). VTQ provides only singlepoint SWS measurement, and VTIQ provides twodimensional color-coded SWE (2D SWE), which displays 2D color velocity maps and allows for multiple measurements to be obtained. The stiffness is proportional to squaring of the SWS.

We measured the embedded parts, and the background parts were 5 mm apart from the outline of the embedded ones. VTIQ measurements were repeated 4 points on the same depth and three times at each part, VTQ measurements were repeated fve times at each part, and the mean value and SD of the SWS were calculated. Reference values of the embedded part were measured in a homogenous phantom

Table 39.5 SWE measurement parameters

made with the same material component. The reference value of the background part was measured at a deeper area of the embedded phantom.

39.2 Results

39.2.1 Development of a Tissue-Mimicking Viscoelastic Phantom for Quantitative Assessment of MRE

Figure [39.3](#page-3-0) shows the stiffness (square root of the sum of squares of the storage and loss modulus) obtained with MRE, the rheometer, and USE. The stiffness of the phantom and bovine liver increased with frequency. Figure [39.4](#page-3-1) shows the change in the mechanical properties of the phantom over a year. The change in the storage and loss modulus during the 1-year period was within ±3%.

39.2.2 Ultrasound-Based Shear-Wave Speed Measurement on a Highly Viscous Embedded Phantom

Table [39.6](#page-3-2) shows the SWS in the embedded part and background part. In the background part, the SWS was equivalent to the reference value. The SWS of the embedded part with a diameter of 20 mm in a highly viscous phantom was measured accurately with the SWE; however, with a diameter of 10 mm was lower than the reference value. Figure [39.5](#page-3-3) shows the B-mode image and the VTIQ image around the embedded part with a diameter of 10 mm. On the VTIQ image, the embedded part was demarcated from the background; however, the border was not sharp. In addition, the embedded part on the VTIQ was visualized to be larger than the part on the B-mode image. This phantom has the potential to be used as a quality control phantom to mimic living tissues.

Table 39.6 Results of SWS measurement

Fig. 39.4 Changes in the mechanical properties of high-viscosity phantom over time

Fig. 39.5 Image of the phantom containing an embedded part with a diameter of 10 mm. (**a**) B mode image. (**b**) VTIQ image

39.3 Conclusion

The purpose of this study was to confrm the frequency characteristics of the developed phantom with commercial USE system and the developed MRE system at multi-scale and multi-frequency. We confrmed that the phantoms are in good agreement with a physical model of the liver, and the developed phantoms are considered effective for the quantitative assessment of the MRE and USE system.

Furthermore, an MRE system using an MR microscope and clinical MRI was developed. We succeeded in performing measurements at multiple frequencies. These developed systems allow quantitative multi-scale and multi-parameter imaging of diseased tissues.

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