

# Chapter 14

## Bioactivities



Kang Liu, Xue-Ying Li, Jian-Ping Luo, and Xue-Qiang Zha

**Abstract** Polysaccharides and proteins are representative natural biomacromolecules existing in animals, plants, and microorganisms. They are attracting a great attention of scholars worldwide due to their various healthy functions, such as immunomodulation, anti-tumor, anti-oxidative, hypoglycemic, and hypolipidemic activities. Besides the strong bioactivity, these natural polysaccharides and proteins are non-toxic and show no side effects. In recent decades, a large number of bioactive polysaccharides and proteins with different structure and bioactivity from natural resources have been extracted, purified, and characterized. The aim of this chapter is to summarize the bioactivities, active mechanisms, structure features, structure–activity relationships of natural polysaccharides, proteins, and their derivatives. Moreover, this chapter also presented the applications of some active natural biopolymers in foods and medicines.

**Keywords** Polysaccharide · Protein · Bioactivity · Structure–activity relationship

### 1 Introduction

The food hydrocolloid is an edible soft matter system, which determines the texture and flavor characteristics of food products (Van der Sman and Van der Goot 2009). In food processing, various food materials such as polysaccharides, proteins, lipids, emulsifiers, sugars, minerals, and water are often mixed and fabricated. Among these ingredients, polysaccharides and proteins are the most used materials, which not only acting as “building blocks” for designing food hydrocolloids, but also providing interface-stabilizing properties via the interaction with other molecules (Wijaya et al. 2017).

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K. Liu · X.-Y. Li · J.-P. Luo · X.-Q. Zha (✉)  
School of Food and Biological Engineering, Hefei University of Technology, Hefei, China  
e-mail: zhaxueqiang@hfut.edu.cn

Polysaccharide is defined as carbohydrate polymers consisting of different monosaccharide linked by glycosidic bonds (Xie et al. 2016). Protein is a macromolecular compound, which is formed by the binding of peptide chains composed of amino acids. In recent decades, many natural polysaccharides and proteins have been extracted and purified from plants, animals, and microorganisms. In addition to the properties of food hydrocolloids, these natural macromolecules also possess many bioactivities, such as anti-tumor, immunomodulation, anti-oxidative, hypoglycemic, and hypolipidemic (Cho et al. 2015). Therefore, this chapter mainly introduces the bioactivities of natural hydrocolloids, including anti-tumor, immunomodulation, anti-oxidation, antimicrobial, hypoglycemic, and hypolipidemic effects. Moreover, the applications of these natural biomacromolecules in functional foods and medicines are presented in this chapter.

## 2 Bioactivities

In recent years, natural polysaccharides and proteins extracted from different materials have attracted increasing attention because of their wide bioactivities, such as anti-oxidation, immunomodulation, anti-tumor, antimicrobial, hypoglycemic, and hypolipidemic effects. Moreover, more and more evidence indicated that most of these bioactivities of polysaccharides and proteins are related to the immune system.

### 2.1 Anti-Tumor

Cancer is a group of diseases involving abnormal cell growth with the potential to invade or spread to other parts of the body. According to the report released by the World Health Organization (WHO) 2018, cancer is one of the main causes of human death worldwide. Although there are many different types of antineoplastic drugs in clinic, these drugs not only have limited efficacy, but also have strong side effects. Since *Lentinan* was first recognized to have anti-tumor efficacy (Chihara et al. 1969), more and more studies on natural polysaccharides used in cancer treatment have been carried out in vitro and in vivo.

Up to date, a series of human carcinoma cell lines have been employed to investigate the anticancer activity of polysaccharides, such as the lung cancer cell line (A549 cell), the cervical carcinoma cell line (Hela cell), the gastric carcinoma cell line (BGC-823 cell), the breast carcinoma cell line (MCF-7 cell), the colon cancer cell line (HCT116 cell and HT29 cell), and the liver cancer cell line (HepG2 cell). In addition, some mouse-derived cancer cell lines were also used to evaluate the activity of polysaccharides. It has been suggested that the anti-tumor mechanisms of polysaccharides were possibly attributed to their inhibition of tumor cell proliferation, initiation of tumor cell apoptosis, and activation of immune system to kill tumor cells (Zong et al. 2012).

It has been reported that polysaccharides from *Dendrobium* (Yu et al. 2018), *Astragalus* (Zhai et al. 2018), *Lentinus edodes* (Ya 2017), *Ganoderma lucidum* (Mohan et al. 2017), and *Portulaca Oleracea L.* (Zhao et al. 2016) exhibited good inhibitory effects on HeLa cells proliferation. These anti-proliferation effects might be related to the increase in autophagic activity of HeLa cells via regulating the expression of some key proteins in mitochondria-mediated signaling pathway, such as beclin1, LC3, and p62 (Zhai et al. 2018). Polysaccharides extracted from *Houttuynia cordata* (Han et al. 2018), *Tremella* (Shi et al. 2018), *Sargassum integerrimum* (Liu et al. 2016), *Pleurotus nebrodensis* (Cui et al. 2016), *Auricularia polytricha* (Yu et al. 2014) exhibited strong activity to resist the proliferation of human A549 cells. Lin et al. (2018) reported that *Hedyotis diffusa* polysaccharides could induce the apoptosis of A549 cells via regulating caspase-3-dependent mitochondrial pathway. Wu et al. (2017) found that polysaccharide from *Glehnia littoralis* could inhibit A549 cell proliferation and migration via decreasing the expression of PCNA, leading to cell cycle arrested in S and G2/M phase. Luo et al. (2016) also found that *coix* polysaccharides had the function to inhibit the migration and invasion of A549 cells via down-regulating the expression of S100A4. S100A4, a member of the S100 family, is a sort of calcium binding protein with EF double helix domain. The S100A4 expresses in kinds of tumor and stem cells of human rather than normal somatocytes.

HepG2 is an immortalized cell line consisting of human liver carcinoma cells. It has been reported that polysaccharides extracted from *Phormidium versicolor* (Belhaj et al. 2018), *Ganoderma lucidum* (Yang et al. 2017), *Lentinus edodes* (Zhao et al. 2017), *Antrodia camphorata* (Li et al. 2009a, b), *Grifola frondosa* (Wang et al. 2013) showed strong ability to prevent the proliferation of HepG2 cells. Li et al. (2013a) reported that polysaccharide from *Phellinus linteus* could induce S-phase arrest in HepG2 cells via decreasing calreticulin expression and activating the P27kip1-cyclin A/D1/E-CDK2 pathway. Shen et al. (2014) found that polysaccharide from *Ganoderma lucidum mycelia* could induce HepG2 cells apoptosis via regulating the expression of miRNAs. Some algae polysaccharides have been proved to possess broad-spectrum antineoplastic effects. For instance, polysaccharides from *Sargassum plagiophyllum* and *Sargassum pallidum* showed strong inhibitory effects on the proliferation of HepG2 cells, A549 cells, and MGC-803 cells in vitro (Ye et al. 2008; Suresh et al. 2013).

$\beta$ -glucans, a type of the most abundant polysaccharides in the cell wall of bacteria and fungus, are glucose polymers linked by 1 $\rightarrow$ 3 linear  $\beta$ -glycosidic bond (Chan et al. 2009). Over the last half-century, fungi-derived  $\beta$ -glucans have received great attention because of the potential medical and edible value all over the world. Lentinan is a representative  $\beta$ -glucan. It has been widely proved to have therapeutic effect on many kinds of tumors. In clinical trials, compared to chemotherapy alone, the addition of lentinan to standard chemotherapy could relieve the pain and prolong survival in patients with gastric cancer (Oba et al. 2009), pancreatic cancer (Shimizu et al. 1999), colorectal cancer (Hazama et al. 2009), liver cancer (Ina et al. 2016), breast Cancer (Taguchi 1983). A large number of cell and animal experiments have also proved these anti-tumor effects. For instance, lentinan has ability to inhibit

proliferation and differentiation of cancer cells, such as human autologous tumor cell line (K562 cell) (Tani et al. 1993), human gastric cancer cell line (BGC823 cell) (Zhao et al. 2013), human pancreatic cancer cell line (BXPC-3 cell) (Qian et al. 2018), human cervical cancer cell line (Hela cell) (Qian et al. 2018), human breast cancer cell line (MCF-7 cell) (Yi et al. 2018), non-small cell lung cancer (Wang et al. 2020), human bladder cancer cell line (T24) (Bao et al. 2015), liver cancer cell line (H22 cell) (Yamamoto et al. 1989). Animal experiments demonstrated that lentinan could inhibit colitis-associated cancer (CAC) development via regulating TLR4/NF-kappaB signaling-mediated inflammatory responses in model mice (Liu et al. 2018).

$\beta$ -glucans have also been reported to kill cancer cells directly. The anticancer mechanisms of these polysaccharides are mainly dependent on the ability of enhancement of host immune system, increase in the antioxidant capacity of host, up-regulation of phase I and phase II enzymes in metabolic transformation, and the detoxification of mutagenic compounds (Vannucci et al. 2013). Masuda et al. (2013) reported that both oral administration and intraperitoneal injection of  $\beta$ -glucans from *Grifola frondosa* could inhibit tumor growth via regulating the systemic immune response. Moreover, the possible mechanism was revealed that the *Grifola frondosa*  $\beta$ -glucans can induce systemic tumor-antigen specific T cell response via dectin-1-dependent activation of DCs, enhance the infiltration of the activated T cells into the tumor, and decrease number of tumor-caused immunosuppressive cells such as myeloid-derived suppressor cells and regulatory T cells, thus leading to the anti-tumor activity. Yeast  $\beta$ -glucans, extracted from by-product yeast of beer production, have been known to exhibit anti-tumor activities by potentiating host immunity (Suphantharika et al. 2003).

In recent years, a large number of anticancer peptides have been identified from plant-derived proteins. For instance, corn peptides can induce the apoptosis of HepG2 cells by increasing caspase-3 expression (Díaz-Gómez et al. 2018). The lunasin peptide from soybean has the ability to resist skin cancer (Hernandez-Ledesma et al. 2009). The potato protein was found to suppress the proliferation of mouse melanoma B16 cells (Sun et al. 2013). Kannan et al. have extracted a pentapeptide (Glu-Gly-Arg-Pro-Arg) from rice bran and proved that it had the ability against the proliferation of colon cancer cells (Kannan et al. 2010). It has been reported that peptides derived from fish proteins have the inhibitory effect on MCF-7 cells in a dose-dependent manner (Hsu et al. 2011). Nongonierma and FitzGerald (2016) also found that milk protein-derived peptides exhibited anti-proliferative activity to tumor cells.

## 2.2 Immunoregulation

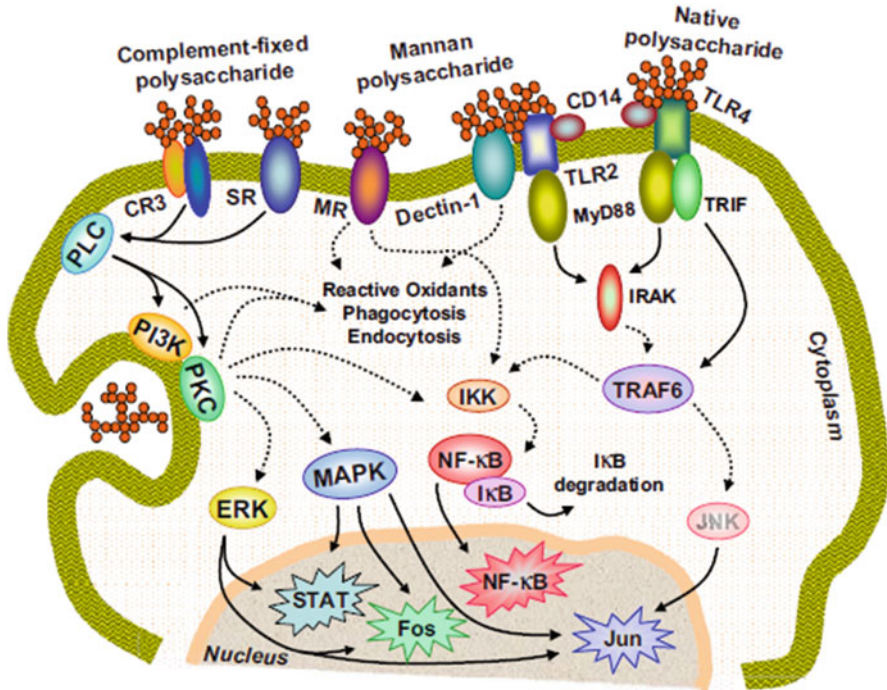
The immune system is a complex network of cells, tissues, and organs that work together to protect the body from harmful substances and organisms and defend against disease. However, when the immune system is disorder, autoimmune

**Table 14.1** Immunomodulatory effects of some polysaccharides on macrophages

Source	Mice type	Cell type	Effects on immune cells	References
<i>Ganoderma atrum</i>	–	RAW264.7	Phagocytosis ↑, NO ↑, TNF-α↑, IL-1β ↑	Yu et al. (2013)
<i>Dendrobium officinale</i>	–	RAW264.7	Phagocytosis ↑, NO ↑	Xia et al. (2012)
<i>Astragalus membranaceus</i>	H22 tumor-bearing mice	H22	IL-2↑, IL-12↑ TNF-α ↑, IL-10↓	Yang et al. (2013)
<i>Mushroom sclerotia</i>	Male BALB/c mice and athymic BALB/c nude mice	–	IL-13↑, IL-17↑, IFN-γ↑	Wong et al. (2011)
<i>Grifola frondosa</i>	–	HepG-2	NO ↑, TNF-α↑, IL-1β ↑	Mao et al. (2015)
<i>Inonotus obliquus</i>	–	SGC-7901	TNF-α↑,	Fan et al. (2012a, b)
<i>Porphyra haitanensis</i>	A BALB/c murine	RAW264.7 DC Tregs	Phagocytosis ↑, TNF-α ↑, NO↑, IL-10↑, IL-6↑	Liu et al. (2017a, b)
<i>Dictyophora indusiata</i>	–	RAW 264.7	TNF-α↑, NO↑,(IL)-6↑	Liao et al. (2015)
<i>Ganoderma lucidum</i>	–	CD	–	Lai et al. (2010)
<i>Prunella vulgaris</i>	–	RAW 264.7	NO↑, TNF-α↑, (IL)-6↑	Li et al. (2015)
<i>Longan pulp</i>	–	Splenic cells	Proliferation↑	Yi et al. (2012)
<i>Cordyceps sinensis</i>	–	RAW 264.7	TNF-α↑, (IL)-6↑, IL-10↑, IL-1α↑	Wu et al. (2014)
<i>Panax Ginseng</i>	–	DC	Proliferation↑, IL-12 ↑, TNF-α↑	Kim et al. (2010)
<i>Laminaria japonica</i>	–	RAW264.7	NO↑, TNF-α↑, IL-1β↑, IL-6↑, IL-10 ↑	Fang et al. (2015)

diseases, inflammatory diseases, and cancer will happen in the body. Many natural polysaccharides have exhibited the ability to affect the immune system via modulating the immune functions including ROS production, cytokine/chemokine production, cell proliferation, and so on (Table 14.1). Therefore, polysaccharide is considered as a potential immunomodulator with great development prospects (Schepetkin and Quinn 2006). Figure 14.1 shows the possible signaling pathways involved in macrophage activation by polysaccharides (Schepetkin and Quinn 2006). It is known that the anti-tumor activity of polysaccharide is partly related to the enhancement of immune system.

The host defense mechanism consists of innate immunity and adaptive immunity, where the innate immunity is the first line of defense mediated the initial protection against infections. It is known that the innate immune system mainly contains macrophages, monocytes, granulocytes, and humoral elements. Among these



**Fig. 14.1** Schematic model of potential signaling pathways involved in macrophage activation by polysaccharides. Reproduction with permission from (Schepetkin and Quinn 2006), Copyright 2006 Elsevier

components, macrophages are reported to exhibit various biological functions, such as chemotaxis, surveillance, phagocytosis, and destruction of targeted organisms, indicating the macrophages activation might be a hopeful strategy to resist diseases (José et al. 2007). It has been reported that *Juniperus scopolorum* polysaccharides could increase macrophage cytotoxic activity against tumor cells and microorganisms, activate phagocytic activity, and enhance the secretion of cytokines and chemokines, such as tumor necrosis factor (TNF- $\alpha$ ), interleukin (IL)-1, IL-6, IL-8, IL-12, interferon gamma (IFN- $\gamma$ ), and IFN-2 (Chen et al. 2010a, b; Schepetkin et al. 2005). Polysaccharide from *Lycium barbarum* could activate macrophages via regulating the transcription factors AP-1 and NF- $\kappa$ B to induce TNF- $\alpha$  production and up-regulating the expression of MHC class II costimulatory molecules, resulting in the enhancement of innate immunity (Chen et al. 2008). Those phenomena indicated that macrophage activation is required for the activation of innate immune system (Plüddemann et al. 2011). For these events, the pattern recognition receptors (PRRs) are required for these cells to recognize stimulators, triggering the activation of signaling pathways and the synthesis of pro-inflammatory cytokines (Kumar et al. 2011). Toll-like receptors (TLRs), the important PRRs, are existed on plasma membrane (Kawai and Akira 2010). It has been reported that macrophage activation

induced by polysaccharides involves TLRs mediated recognition (Li et al. 2011). Figueiredo et al. (2012) have evidenced that TLR2 and TLR4 were the receptors involved in the recognition of *fungus* polysaccharides. Ferwerda et al. (2008) reported that the *saccharomyces cerevisiae* cell wall polysaccharide (Zymosan) has function to induce macrophages to release cytokines by the recognition of TLR2, TLR4, and Dectin-1. Nuclear factor  $\kappa$ B (NF- $\kappa$ B) and mitogen-activated protein kinases (MAPK) are the key proteins in the downstream signaling pathway of TLR, which play important role in the host defenses via regulating the expression of multiple inflammatory and immune genes (DiDonato et al. 2012). With respect to MAPKs, mammalian cells expressed three representative MAPK pathways, containing C-Jun-N-terminal kinase (JNK), extracellular signal regulated kinase (ERK1/2), and p38. In recent years, the immunostimulatory activity mechanisms of polysaccharides have been widely studied. Results suggested that the regulation of intracellular signaling pathways is essential for the activation of macrophages (Diao et al. 2014; Zhang et al. 2014; Maeda et al. 2012). Extracellular polysaccharide LBP32 from *Bacillus sp.* strain was reported to inhibit LPS-induced production of pro-inflammatory cytokines via attenuating the phosphorylation of P38 and JNK, but not ERK1/2 (Diao et al. 2014). *Lycium barbarum* polysaccharide (LBPF4-OL) was found to have the ability to promote the secretion of TNF- $\alpha$  and IL-1 $\beta$  via inhibiting JNK and ERK1/2 MAPK phosphorylation and increasing the phosphorylation of p38-MAPK (Zhang et al. 2014). The sulfated polysaccharide SP1 from *Caulerpa lentillifera* had the function to activate macrophages and enhance NO production via regulating NF- $\kappa$ B and P38 MAPK signaling pathways (Maeda et al. 2012). These results demonstrated that various polysaccharides can exert their biological activities through regulating different signaling pathways. The inflammatory response has been reported to be highly dependent on MAPK signaling pathways via activating its downstream cytosolic proteins and nuclear transcriptional factors (Arthur and Ley 2013). NF- $\kappa$ B is a ubiquitous transcription factor, which plays a critical role in the host defenses via regulating the expression of multiple inflammatory and immune genes (DiDonato et al. 2012). In unstimulated cells, NF- $\kappa$ B locates in cytoplasm and combines with inhibitory proteins to form an inactive trimer (p50-p65-I $\kappa$ B). When cells are stimulated, I $\kappa$ Bs will be phosphorylated by I $\kappa$ B kinase, leading to I $\kappa$ B degradation and translocation of NF- $\kappa$ B to the nucleus for binding to its cognate DNA in the regulation region of a variety of genes (He et al. 2013). It has been reported that the ability of some polysaccharides to activate macrophages is dependent on their level to the activation of NF- $\kappa$ B pathway (Zhang et al. 2011; Yu et al. 2013).

Lymphocyte is considered as a mediator of innate and adaptive immunity. Shriner et al. (2010) reported that *pneumococcal polysaccharide* could stimulate the proliferation of IL-7-driven B lymphocytes, regulate their cytokine production, and restore impaired T cell by immune response. Among the specialized cell subsets of the innate immune system, DCs are the critical sensors via expressing various pattern recognition receptors (Steinman and Banchereau 2007). In particular, TLRs and cytosolic sensors for DNA and RNA recognition expressed by DCs use endogenous host elements carrying microbial components (such as the alarmin HMGB1),

pathogen associated molecular patterns, and/or nucleic acids to stimulate intrinsic apoptotic pathways to generate protective immune responses (Peng et al. 2005; Poeck et al. 2008; Besch et al. 2009). During this process, polysaccharide was found to regulate the immunity via inducing DC maturation. For example, *Astragalus* polysaccharides could induce the differentiation of DCs to CD11c<sup>high</sup>CD45RB<sup>low</sup> DCs by shifting of Th2 to Th1, resulting in the enhancement of T lymphocyte immune function in vitro (Liu et al. 2011a). *Achyranthes bidentata* polysaccharide was reported to enhance DC maturation and function, supplying extra IL-12 and MHC class II molecules to up-regulating antigen presentation, activating CD4+ T cell, and thus leading to an enhancement of DC-CD4+ T cell (Zou et al. 2011). Meng et al. (2011) reported that polysaccharides from *Ganoderma lucidum* could promote effective activation of murine DCs in the immune response via up-regulating the expression of CD86, CD40, and MHC II and down-regulation of acid phosphatases.

In the past decades, the structure–activity relationships of immunomodulatory polysaccharides have been studied, indicating the interaction of immunostimulatory polysaccharides with cell receptors may trigger signaling pathways and thereby result in the induction of gene transcription (Ferreira et al. 2015a, b). A *Houttuynia cordata* pectic polysaccharide (HCP-2) with a linear chain of 1,4-linked  $\alpha$ -D-galacturonic acid residues has been reported to increase the secretion of MIP-1 $\alpha$ , MIP-1 $\beta$ , TNF- $\alpha$ , IL-1 $\beta$ , and RANTES in human peripheral blood mononuclear cells via regulating TLR-4 mediated signaling (Cheng et al. 2014). Bose et al. (2014) reported that 1,3-linked  $\beta$ -D-glucans could activate innate immune functions via regulating Dectin-1 and CR3 mediated signaling pathways. SR has been shown to be the pattern recognition receptor of fucoïdan. Guo et al. (2009) found that a 1,3-linked glucan from spores could be recognized by dectin-1 on macrophages and thereby possess the biological activities. These results suggested that what is polysaccharide's pattern recognition receptor might be determined by the structure of polysaccharide. Lo et al. (2007) suggested that galactose, mannose, xylose, and arabinose played an important role in the stimulation of macrophages, but not glucose. The residues of 1,4-lined  $\beta$ -D-Rhap and 1,5-lined  $\alpha$ -L-Araf were reported to be important for lymphocytes activation (Yang et al. 2012). The 1,4-linked mannose and glucose was reported to be the important elements for macrophages activation by a purified *Laminaria japonica* polysaccharide LJP-31 (Fang et al. 2015).

In recent years, some immunomodulatory peptides have been prepared from food proteins (Agyei and Danquah 2012). Otani et al. (2003) reported that phosphopeptides from casein could stimulate gastrointestinal tracts of mice to release immunoglobulin A. Pan et al. (2013a) revealed that peptide from milk protein exhibits immunomodulatory property in ICR mice. After modification with dicarbonyl methylglyoxyl, ovalbumin has the ability to stimulate immune cells to release tumor necrosis factor (TNF) alpha (Fan et al. 2003). The immunogenic ovalbumin peptides have been employed to enhance the immune response of different cancer patients (Vidovic et al. 2002; Goldberg et al. 2003). Some fish protein-derived immune peptides have also been identified in recent years (Yang



et al. 2009; Hou et al. 2016). Sheu et al. (2004) have separated an immunomodulatory protein from the Jew's Ear mushroom *Auricularia polytricha*. Some peptides extracted from macroalgae were also reported to exhibit immunomodulatory activity via regulating the nuclear factor kappa B (NF- $\kappa$ B) pathway (Ahn et al. 2011).

### 2.3 Anti-Oxidation

Oxidative damage of biomolecules triggers not only physiological process of aging, but also causes various physiological functional disorders, leading to serious health problem ultimately (Harman 1993). In theory, antioxidants might have a positive effect on our health because they have ability to clear free radicals from human body. As is well known, free radicals can attack macromolecules such as proteins, membrane lipids, and DNA, leading to many health problems (i.e., cancer, neurodegenerative diseases, and diabetes mellitus) via damaging cells and tissues (Lim et al. 2014). Reactive nitrogen species (RNS) and reactive oxygen species (ROS) are free radicals that are formed during the normal metabolism of cells, which can be removed by cellular anti-oxidative defense systems, such as glutathione peroxidase (GSH-Px) and superoxide dismutase (SOD). Under normal physiological conditions, the generation and elimination of RNS and ROS are balanced. Once this balance is broken, either by the overproduction of ROS and RNS, or by the damage in anti-oxidative system, oxidative stress will occur (Klaus et al. 2011; Sun et al. 2010). In food industries, some synthetic commercial antioxidants such as tertbutylhydroquinone (TBHQ), butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), and propyl gallate (PG) have been extensively used to reduce the oxidation and peroxidation damage. However, these antioxidants have potential hazards to human health (Nagaoka et al. 2010). Therefore, screening antioxidants from natural resources is always a hot topic (Peña-Ramos and Xiong 2002). Natural polysaccharides have attracted extensive attention and are proposed to be the potential resource of novel antioxidants due to their low toxicity and excellent anti-oxidation. Algal polysaccharides have been demonstrated as a scavenger of free radicals for the prevention of oxidative damage in vivo (Cristina Diaz et al. 2017).

In general, the polysaccharide eliminates free radicals through four aspect, including: (1) Hydrogen atoms on the structure of polysaccharides react with free radicals to form water, and the single electrons generated by the reaction can be further reduced. (2) Polysaccharides capture free radicals produced in lipid reactions or chelate with metal ions, which are important factors for the formation of free radicals. (3) Polysaccharides enhance the activity of some antioxidant enzymes. (4) Polysaccharides indirectly achieve antioxidant effect by regulating immunity. As shown in Table 14.2,  $\beta$ -glucan extracted from mushrooms and yeast have been reported to be the potential antioxidants. Three polysaccharides isolated from *Ganoderma lucidum* (GLP-H, GLP-V, and GLP-F) were found to possess the stronger radical scavenging activities (Fan et al. 2012a, b). *Astragalus*

**Table 14.2** Antioxidants activity of  $\beta$ -glucan

Source	Antioxidant activity	References
<i>Jinqian mushroom</i>	ABT radical scavenging activity was 63.96% at 5 mg/mL DPPH scavenging ratio was 89.84% at 5 mg/mL Iron chelating effect was 14.06% at 5 mg/mL Hydroxyl radical scavenging activity was 24.30% at 5 mg/mL	Liu et al. (2014a, b)
<i>Polyporus dermatopus</i>	Hydroxyl radicals inhibition was 96% at 267 $\mu$ g/mL Lipid peroxidation inhibition was 42.9% at 67 $\mu$ g/mL Superoxide inhibition was 83.3% at 67 $\mu$ g/mL	Dore et al. (2014)
<i>Saccharomyces cerevisiae</i>	Decreasing the formation of RBARS in LPS stimulated human blood platelets Decreasing the formation of O <sub>2</sub> in LPS stimulated human blood platelets	Saluk et al. (2013)
<i>Geastrum saccatum mushroom</i>	Inhibition of the formation of hydroxyl radicals in a dose-dependent manner	Guerra Dore et al. (2007)
<i>Pleurotus ostreatus</i>	The antioxidant enzymes activity, ferric reducing activity, and ascorbate concentration in human red blood cells hemolysates were markedly increased	Pietrzycka et al. (2006)
<i>Lentinus edodes</i>	Inhibition of lipid peroxidation, as well as a strong hydroxyl radical scavenging activity and superoxide radical scavenging activity	Feng et al. (2010)
<i>Yeast</i>	The level of glutathione was replenished and myeloperoxidase activity was suppressed in a rat model of sepsis	Sener et al. (2005)

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polysaccharides were reported to inhibit the generation of ROS via suppressing the NF- $\kappa$ B signal pathway (Xue et al. 2015). When the mice were orally administrated with extracellular polysaccharides of *Morchella esculenta*, the activity of SOD and GSH-Px were elevated in the blood, heart, liver, spleen, and kidney of mice (Meng 2010).

It has been reported that some compounds such as proteins, peptides, pigments, polyphenols, and flavones could bind to polysaccharides. Compared to the polysaccharides, these complexes have stronger antioxidant activity. For instance, the antioxidant effects of tea polysaccharides (TPS)-protein conjugates are dose-dependent on the protein content (Nie et al. 2008). In addition, the in vivo and in vitro antioxidant activities of crude tea polysaccharides were found to be better than that of tea polysaccharide fraction, which can be interpreted by the relatively higher proportion of tea pigments, vitamins, tea polyphenols, and other antioxidant components in the cruder fractions (Zhou et al. 2007). Moreover, Zhang et al. (2016a, b, c) suggested that the antioxidant activity of polysaccharides from *Ganoderma atrum* (PSG) was depended on the content of phenolic compounds/proteins.

It is a fact that the antioxidant activity of polysaccharides is closely correlated with their structural parameters, such as solubility, degree of substitution, degree of

branching, molecular weight, monosaccharide composition, solution conformation, and functional groups. Jing et al. (2009) reported that phosphorylated modification could enhance the ability of fucoidan to scavenge hydroxyl and superoxide radicals. The possible mechanism might be that the phosphate is a polyelectrolyte group and could activate the hydrogen atom of the anomeric carbon.

The acetylated modification was reported to enhance the ability of mushroom *Inonotus obliquus* polysaccharides to inhibit lipid peroxidation via affecting the conformation of polysaccharides (Ma et al. 2012). The carboxymethylated modification can increase the water solubility and antioxidant activity of  $\beta$ -(1,3)-glucan from the sclerotium of *Poria coco* via changing the flexibility of polysaccharides chain (Wang and Zhang 2006). Molecular weight was an important factor that can influence the antioxidant ability of polysaccharides. Yan et al. (2009) used an acidic solution to hydrolyze the exopolysaccharide from *Cordyceps sinensis*, giving a degraded exopolysaccharide. Results showed that the degraded products had much higher antioxidant activity than that of original polysaccharide.

## 2.4 Antimicrobial

$\beta$ -glucans are ubiquitously found in both bacterial and fungal cell walls and have been implicated in the initiation of antimicrobial immune response. It has been proved that the (1 $\rightarrow$ 3)- $\beta$ -D glucan with (1 $\rightarrow$ 6)- $\beta$ -D branches could act as antimicrobial agent in vivo (Ferreira et al. 2015a, b). Hetland et al. (2000) obtained a soluble branched  $\beta$ -(1,3)-glucan (SSG) from the culture broth of the fungus *Sclerotinia sclerotiorum* IFO9395. Results exhibited that the oral administration of SSG could help the mice to resist the infection of *Streptococcus pneumoniae* sero type 4 and 6B. Faccin et al. (2007) reported that  $\beta$ -glucan from the fruiting body of *Agaricus brasiliensis* mushrooms exhibited an antiviral activity against poliovirus typ1 in HEp-2 cells. Peter et al. found that oral administration of  $\beta$ -glucan (glucan phosphate, scleroglucan, and laminarin) at a dose of 1 mg/kg/day could enhance the survival of mice infected with *S. aureus* or *Candida albicans*. In Sharma's study (Sharma et al. 2015), it was found that both extracellular polysaccharide (EPS) and intracellular polysaccharides (IPS) extracted from *Cordyceps* species exhibited obvious antimicrobial activities against all pathogenic microorganism tested. Moreover, IPS showed stronger antimicrobial activity than that of EPS. The polysaccharides extracted from *Ganoderma* species have also been proved to exhibit spectral antimicrobial activity (Table 14.3). It has been found that the acidic polysaccharide CS-F2 from green tea had a selective anti-adhesive activity against some pathogenic bacteria and strongly inhibited the growth of gastric and skin pathogenic bacteria.

More and more evidences exhibited that some proteins and their derived peptides from plants, mammals, insects, and bacteria also have antimicrobial activity against eukaryotes, fungi, bacteria, and viruses (Zhu et al. 2019). The first antimicrobial peptide was found from the moth *Hyalophora cecropia* in 1981 (Steiner et al. 1981). A lectin-like peptide from red lentil seeds exhibited antimicrobial activity against

**Table 14.3** The antimicrobial activity of polysaccharides

Specie	Microorganisms used	Type of assay	References
<i>Ganoderma lucidum</i>	<i>Bacillus subtilis</i> , <i>Bacillus cereus</i> , <i>Erwinia carotovora</i> , <i>Escherichia coli</i> , <i>Penicillium digitatum</i> , <i>Botrytis cinerea</i>	In vitro	Bai et al. (2008)
<i>Ganoderma applanatum</i>	<i>Acrobacter aerogenes</i> , <i>Acitenobacter aerogenes</i> , <i>Arthrobacter citreus</i> , <i>Bacillus brevis</i> , <i>B. subtilis</i> , <i>Corynebacterium insidiosum</i> , <i>Clostridium pasteurianum</i> , <i>Escherichia coli</i> , <i>Micrococcus roseus</i> , <i>Mycobacterium phlei</i> , <i>Proteus vulgaris</i> , <i>Sarcina lutea</i> St	In vitro	Bhattacharyya et al. (2006)
<i>Ganoderma formosanum</i>	<i>Listeria monocytogenes</i>	In vivo	Wang et al. (2011)
Glucans	<i>Staphylococcus aureus</i> or <i>Candida albicans</i>	In vitro	Rice et al. (2005)
<i>Schizophyllum</i>	<i>Salmonella enterica serovar</i>	In vivo	Chen et al. (2008)
Oat	Herpes simplex virus 1	In vivo	Murphy et al. (2009)

*Mycosphaerella arachidicola* (Wang and Ng 2007). The defensin PDC1 peptide, a fermentation product of corn using *Pichia pastoris* and *Escherichia coli* as the mixed strain, has an ability to inhibit the growth of *Fusarium graminearum* (Kant et al. 2009). Using *Bacillus subtilis* sck-2 to ferment soybean paste, a peptide was obtained and exhibited antibacterial activity against *Bacillus cereus* (Yeo et al. 2012). These antimicrobial peptides can be considered as the good candidates for food antiseptis.

## 2.5 Hypoglycemic

Diabetes mellitus (DM) is a group of metabolic disorders characterized by high blood sugar levels over a prolonged period. Symptoms of high blood sugar mainly include frequent urination, increased thirst, and increased hunger. If not treated in time, diabetes will cause many complications. Acute complications include hyperosmolar hyperglycemic state, diabetic ketoacidosis, or death. Serious long-term complications include cardiovascular disease, chronic kidney disease, foot ulcers, stroke, and damage to the eyes. According to the International Diabetes Federation (IDF) in 2017, an estimated 425 million individuals are living with DM. At present, the main treatment of diabetes is oral hypoglycemic drugs and insulin injection. However, long-term use of these drugs could lead to insulin resistance and other side effects. Thus, it is quite necessary to develop helpful, innocuous, and inexpensive drugs for DM patients. As the non-toxic biological

**Table 14.4** The anti-diabetic effects of polysaccharides

Source	Model	Effects and mechanisms	References
<i>Opuntia dillenii</i>	STZ mice	Protecting the liver from peroxidation damage, maintaining tissue function, improving the sensitivity and response of target cells in diabetic mice to insulin	Zhao et al. (2011)
<i>Tremella aurantia</i>	KK-Ay mice	Reducing levels of insulin, total cholesterol and triglyceride in the mice blood, decreasing the level of plasma lipoperoxide	Kiho et al. (2001)
<i>Phellinus linteus</i>	NOD mice	Inhibiting the development of autoimmune diabetes by regulating cytokine expression	Kim et al. (2010)
<i>Morus alba fruit</i>	T2DM mice	Repairing of damaged pancreatic tissues of diabetic rats.	Jiao et al. (2017)
<i>Lycium barbarum</i>	T2DM mice	Increasing insulinogenic index	Cai et al. (2011)
<i>Pleurotus ostreatus</i>	STZ mice	Reducing the risk of oxidative damage by increasing catalase (CAT), glutathione peroxidase (GSH-Px) and superoxide dismutase (SOD) activities and decreasing malonaldehyde (MDA) level	Zhang et al. (2016a, b, c)
<i>Portulaca oleracea L.</i>	Alloxan-induced diabetic mice	Controlling blood glucose, and modulating the metabolism of glucose and blood lipid in diabetes mellitus mice	Li et al. (2009a, b)
<i>Corn silk</i>	T2DM	Regulating the levels of serum lipid profile, decreasing the levels of glycated serum protein, non-esterified fatty acid	Pan et al. (2017)
<i>Hedysarum polybotrys</i>	Alloxan-induced diabetic mice	Increasing insulin secretion, inhibiting lipid peroxidation, promoting the sensitivity to insulin, suppressing gluconeogenesis, and reducing the biosynthesis fatty acid, cholesterol, and cell cytokines related to insulin resistance	Hu et al. (2010)
<i>Taxus cuspidata</i>	STZ mice	Increasing the body weight of diabetic mice, and reversing the decrease of SOD and the increase of thiobarbituric acid reactive substances (TBARS) in kidney and liver of diabetic mice	Zhang et al. (2012)
<i>Ganoderma lucidum</i>	T2DM	Down-regulation of the hepatic glucose regulated enzyme mRNA levels via AMPK activation, improvement of insulin resistance and decrease of epididymal fat/BW ratio	Xiao et al. (2016)

macromolecules, natural polysaccharides were observed to have positive effects to treat DM (Huie and Di 2004; Zhou et al. 2007). Table 14.4 presents the anti-diabetic effects of different polysaccharides.

A polysaccharide (CSP-1) from *Cordyceps sinensis* consists of glucose, mannose, and galactose in a molar ratio of 1:0.6:0.75, and exhibited a significant drop in blood glucose level in both normal and streptozotocin (STZ)-diabetic animals. These results could be related to the increase in blood insulin level via the release of insulin from the residual pancreatic cells and/or CSP-1-induced the reduction of insulin

metabolism in body (Li et al. 2006a, b). Another two purified polysaccharides (CS-F30 and CS-F10) from *Cordyceps sinensis mycelia* was also proved to have hypoglycemic effects on mice (Kiho et al. 1996, 1999). Compared to CSP-1, these two polysaccharides have different monosaccharide composition. In recent years, the hypoglycemic effect of tea polysaccharides (TPS) has also attracted much attention. Chen et al. (2010a, b) reported that the oral administration of TPS at 150 mg/kg/day can significantly reduce the blood glucose level in non-obese diabetic (NOD) mice. From TPS, two water-soluble polysaccharide fractions of TFP-1 and TFP-2 were obtained. The average molecular weight was determined to be  $15.9 \times 10^4$  and  $1.12 \times 10^4$  Da, respectively. Results showed that continuous administration of TFP-2 could dose-dependently decrease the blood glucose level in alloxan-induced diabetic mice by inhibiting  $\alpha$ -amylase and  $\alpha$ -glucosidase (Han et al. 2011a, b). Zhou et al. (2007) isolated a crude tea polysaccharides (CTP) and a tea polysaccharide fraction (TPF) from green tea. It was found that CTP and TPF have hypoglycemic effects on alloxan-induced diabetic mice. The hypoglycemic action of polysaccharides from oolong tea and black tea has also been investigated. Results exhibited that these tea polysaccharides could alleviate the diabetic mice and improve diabetes symptoms, indicating tea polysaccharides have good effects on the prevention of hyperglycemia (Nie et al. 2011).

Food-derived peptides have been reported to have antagonistic effects against glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1) in type 2 diabetes via inhibiting the activity of Dipeptidyl peptidase IV (DPP-IV) (Matteucci and Giampietro 2009). So far, this active peptide has been isolated from corn, milk, soybean, rice, and other foods (Zhu et al. 2019; Mochida et al. 2010; Hira et al. 2009; Sanjukta and Rai 2016). The dipeptides and tripeptides isolated from whey proteins have been used as competitive inhibitors to interact with DPP-IV substrates (Gunnarsson et al. 2006). Recently, more attention have been paid on the application of marine bioactive peptides to treat type 2 diabetes. The peptides from *Porphyra dioica* were observed to have the inhibitory effect on DPP-IV (Stack et al. 2017). Three polypeptides (ILAP, LLAP, and MAGVDHI) with similar bioactivity were also isolated from macroalga *Palmaria palmata* (Harnedy et al. 2015).

## 2.6 Hypolipidemic

Hyperlipidemia is a lipid metabolic disorder characterized by the enhancement of blood total cholesterol (TC), low-density lipoprotein (LDL), very low-density lipoprotein-cholesterol (VLDL-C) and triglyceride (TG), with a concomitant decrease in the level of high-density lipoprotein-cholesterol (HDL-C) in the plasma. With the improvement of people's living standard and the change of dietary structure, the incidence of hyperlipidemia is increasing year by year (Chan et al. 2009). Hyperlipidemia is one of the main risk factors for inducing cardiovascular diseases, such as hypertension, atherosclerosis, and coronary heart disease (Rosenson et al. 2002).

In a meanwhile, lipid accumulation in liver (steatosis) can result in oxidative stress and inflammation, leading to the damage of liver (Esposito et al. 2002). Thus, how to reduce lipid level is one of most important clinical problems. At the present time, statins, nicotinic acid, and its derivatives are the most common lipid-lowering prescribed drugs. Although these drugs help in lowering lipids, their side effects are also enormous, such as headache, muscle pain, and nausea. Moreover, the long-term use of these drugs can increase the risk of type 2 diabetes (Hsu et al. 2015). In recent decades, more and more evidences exhibited that natural polysaccharides had the function to low lipids. For instance, *Enteromorpha prolifera* polysaccharides exhibited a high hypolipidemic action in high fat rats via decreasing the plasma LDL-C, TC, and TG levels and increasing HDL-C level (Teng et al. 2013). *Ganoderma lucidum*  $\beta$ -glucan was also reported to decrease the TC, TG, and LDL-C levels in the serum of diabetic mice, whereas the HDL-C level was increased (Li et al. 2011). Recently, significant in vivo studies testing, the efficacy of cereal  $\beta$ -glucans using animal and human subjects has led to health claims for this material in many industrialized countries. The soluble dietary fibers (i.e.,  $\beta$ -glucan from cereal grains) are well accepted to be the polysaccharides with the ability to lower lipids (Pomeroy et al. 2001). Therefore, adding  $\beta$ -glucan-rich fiber products in the daily diet is now considered as an effective approach to lower lipids. Extensive efforts have been made to establish the relationship between the structure of cereal  $\beta$ -glucans and reduction of LDL-C levels (Lazaridou and Biliaderis 2004; Li et al. 2006a, b). Cereal  $\beta$ -glucans are linear homopolysaccharides formed by the linkage of  $\beta$ -D-glucopyranosyl units via (1 $\rightarrow$ 4)- $\beta$ -linkage and separated by single (1 $\rightarrow$ 3)- $\beta$ -linkages. They are the main component of water-soluble dietary fibers from cereals. The physiological activity of soluble  $\beta$ -glucan from cereal is closely related to its unique structure of (1 $\rightarrow$ 3)(1 $\rightarrow$ 4)- $\beta$ -D-glucan. It was concluded that the LDL-C lowering effect and the ability to control blood glucose of cereal  $\beta$ -glucan may depend on its viscosity in solution, which is controlled by the  $M_w$ , structure, and concentration in the intestine (Wood 2007).

It has been reported that  $\beta$ -glucan can bind to the bile acid in intestinal lumen. This combination would reduce the circulation of bile acid liver and further stimulate the production of more bile acids from cholesterol. Moreover,  $\beta$ -glucan could be fermented in the large bowel by colonic bacteria, and producing the short-chain fatty acids. The short-chain fatty acids could be absorbed by portal vein, and inhibit hepatic cholesterol synthesis via regulating the activity of HMG-CoA reductase (a rate-limiting enzyme required for cholesterol biosynthesis), or increasing catabolism of LDL-cholesterol. Another phenomenon was observed that  $\beta$ -glucan reduced the concentration of postprandial serum insulin by delaying gastric emptying, leading to the inhibition of hepatic cholesterol production. It was also reported that  $\beta$ -glucan could interfere with the absorption of dietary fat via increasing intestinal viscosity (Bell et al. 1999).

## 2.7 Other Activities

Besides the healthy benefits mentioned above, some other bioactivities of polysaccharides and protein-related compounds have been studied in recent years, including antidiarrheal, anti-fatigue, anti-tussive, anti-analgesia, anti-allergic, and anti-dyszoospermia activities. Baek et al. (2010) evaluated the anti-diarrhea effect of *ginseng* polysaccharides in vitro using the model of rotavirus infection. Results showed that two pectic ginseng polysaccharides dose-dependently rescued the cell viability from rotavirus infection. Moreover, the prevention of polysaccharide on rotavirus was attributed to the inhibitory effects of rotaviral attachment to cells. In recent years, the anti-fatigue effect of polysaccharides extracted from different materials is gradually accepted by people (Jing et al. 2009). Pimentel et al. (2019) demonstrated that macroalgae-derived peptides and enzymes have protective effects on skin via eliminating free radicals and promoting moisture.

## 3 Structure Features

Chemical structure is the basis of polysaccharides and to exert their biological activity, including monosaccharide composition, monosaccharide arranging order, anomeric carbon configuration, glycoside bond types, branches, substituted groups, and the spatial conformation. It has been reported that most bioactive polysaccharides are mainly composed of glucose, fucose, galactose, arabinose, mannose, xylose, ribose, glucuronic acid, and galacturonic acid. According to the published literatures, the fungal derived polysaccharides have been found to be  $\alpha$ -mannan,  $\beta$ -glucans and hetero- $\beta$ -glucans,  $\alpha$ -mannan- $\beta$ -glucan complexes, heteroglycans, glycopeptides or glycoprotein and proteoglycan. The polysaccharides extracted from plants mainly include glucans, glucomannans, heteroglycans, arabinans, arabinogalactan, pectins, rhamnogalacturonan, and their sulfated or acetylated forms. Animal polysaccharides are mainly composed of glycosaminoglycan and sulfated glycosaminoglycan.

Although it is quite difficult to determine the structural variability of polysaccharides from different resources, a series of analytical methods have been established to achieve this. The high performance liquid chromatography (HPLC) is often performed to determine the molecular weight of polysaccharides. The infrared spectroscopy (IR), ultraviolet spectroscopy (UV), gas chromatography-mass spectrometry, NMR spectroscopy, periodate oxidation, partial acid hydrolysis, methylation, and periodate oxidation-Smith degradation are the common methods to analyze structural features of polysaccharides (Yang et al. 2009).

The chemical structure of natural polysaccharides is very complex. Table 14.5 showed the structure features of some polysaccharides extracted from different species. In most cases, one material could contain a variety of polysaccharides with different structures. Moreover, the same species growing in different places



**Table 14.5** Structure features of polysaccharides from various sources. Reproduction with permission from (Nie et al. 2018), Copyright 2018 Elsevier

Source	Mw (Da)	Monosaccharide composition	Backbone	References
<i>Pleurotus florida</i>	180,000	Glucose	(1→6)-linked-β-D-Glcp (1→3,6)-linked-β-D-Glcp	Maji et al. (2012)
<i>Auricularia polytricha</i>	120,000	Glucose	1,3-β-glucan 1,3-α-glucan 1,4-α-glucan	Song and Du (2012)
<i>Cistanche Deserticola Y. Ma</i>	10,000	Glucose	1,4-linked-α-D-glucan	Dong et al. (2007)
<i>Cordyceps sinensis</i>	–	Glucose, Mannose, Galactose	(1→4)-linked-α-D-Glcp	Nie et al. (2011)
<i>Ganoderma lucidum</i>	83,000	Rhamnose, Galactose, Glucose	1,4-linked-α-D-Glcp 1,6-linked-β-D-Glcp	Bao et al. (2002a, b)
<i>Ganoderma lucidum</i>	200,000	Glucose, Mannose	1,3-, 1,4-, 1,6-linked-β-D-Glcp 1,6-linked-β-D-Manp	Bao et al. (2002a, b)
<i>Ophiopogon japonicus</i>	35,200	Arabinose, Glucose, Galactose	1,4-linked-Glcp 1,6-linked-Glcp 1,4,6-linked-Glcp	Chen et al. (2011)
<i>Lentinus squarrosulus (Mont.) Singer</i>	196,000	Galactose, Glucose, Fucose	(1→4)-linked-α-D-Glcp (1→6)-linked-β-D-Glcp (1→4,6)-linked-D-Glcp (1→3,6)-linked-D-Glcp	Bhunia et al. (2010)
<i>Dendrobium huoshanense</i>	73,000	Glucose, Galactose, Xylose	1,4-linked-β-D-Glcp 1,6-linked-β-D-Glcp 1,4,6-linked-β-D-Glcp	Pan et al. (2013b)
<i>Radix Astragali</i>	1334,000	Rhamnose, Glucose, Arabinose, Galactose, Galactose acid	1,4-linked-α-Glcp 1,4-linked-α-GalAp6Me 1,2,4-linked-Rhap 1,3,6-linked-β-Galp	Yin et al. (2012)

could contain different polysaccharides. It is also reported that the polysaccharides in different organs of the same species is also different in the structures. It has been observed that polysaccharides extracted from the spores of *Ganoderma lucidum* possessed a backbone of (1–3)-β-linked glucans (Bao et al. 2001). However, the backbone of fruit bodies polysaccharides was composed of 1,3-linked glucose, 1,6-linked glucose, 1,6-linked mannose, 1,4-linked glucose, and 1,6-linked galactose (Bao et al. 2002a, b).

For the preparation of natural polysaccharides, the extraction methods and extraction parameters are also critical factors affecting polysaccharide's structures and properties. For instance, Palacios et al. compared the effects of different extraction methods on the structure of polysaccharides from *Pleurotus ostreatus* mushroom. Results showed that the polysaccharides extracted by cold-water mainly

consisted of 1–3- $\alpha$ -linked and 1–6- $\alpha$ -linked galactose. The polysaccharide extracted by hot-water was mainly composed of 1–4- $\alpha$ -linked glucose. However, the fraction extracted by hot-aqueous NaOH was mainly composed of 1–3- $\beta$ -linked and 1–6- $\beta$ -linked glucose (Palacios et al. 2012).

In addition to the natural factors, chemical modification can also change the polysaccharide's structure, leading to the variation of physicochemical properties and bioactivities (Fiorito et al. 2018). According to the published literatures, the acetylation, carboxymethylation, sulfation, phosphorylation, alkylation, and selenization have been employed to modify the structure of polysaccharides (Prashanth and Tharanathan 2007). Moreover, some modified polysaccharides have been developed into drug delivery systems (Shah et al. 2011).

In addition to the natural factors, polysaccharide derivatives also contribute to the structural diversity, which can also be classified as a semisynthetic polysaccharide. It has been reported that the effective chemical modification of this natural structure could improve the bioactivities and some key parameters, including solubility, bioavailability, and pharmacokinetics (Fiorito et al. 2018). Chemical modification can control the final structure of polysaccharides, and thus determining the specific biological functions. In addition, the chemical modification of polysaccharide structure mainly utilizes the polysaccharide's reactive groups, such as hydroxyl, carboxyl, and amino groups, to chemically introduce new functional groups. Chemical modification of polysaccharide includes sulfation, carboxymethylation, acetylation, alkylation, phosphorylation, and selenization. Some semisynthetic polysaccharides have been developed into various drug delivery systems.

In recent years, a large number of studies have been carried out on the structure–activity relationship of polysaccharides. Results exhibited that the activity of polysaccharides was mainly related to the molecular weight, chemical structure, and physical properties (Jin et al. 2012). The (1,3)- $\beta$ -D-glucan from *Poria cocos* sclerotium is a water-insoluble polysaccharide and exhibited low bioactivity. After carboxymethylation, both the water solubility and bioactivity were enhanced (Wang et al. 2009). Di et al. (2017) reported that sulfated polysaccharides from *Gracilaria rubra* exerted immunologic activity by promoting the proliferation of RAW264.7 cells. Moreover, the activity was improved with the decrease of polysaccharide molecular weight. Chen et al. (2015) demonstrated that sulfated polysaccharide from *Ganoderma atrum* has the strongest immunological activity when the molecular weight was intermediate ( $4.0 \times 10^{-6}$  Da). Zhang et al. (2005) found that the physicochemical property and steric conformation of polysaccharides from *Poria cocos* mycelia were changed by the introduction of sulfate groups, leading to the changes in the bioactivity of polysaccharides. Similarly, sulfated modification changed the structure and conformation of polysaccharides from *Hypsizygus marmoreus*, enhancing the ability of anticancer and immunity (Bao et al. 2010). The substitution degree of substituent groups also has a great influence on the biological activity of polysaccharides. For instance, chitosan with different substitution degree of sulfuric acid group exhibited different strength of immunoregulation ability (Yang et al. 2018). Liu et al. (2017a, b) investigated the structure–activity relationship of selenium-containing polysaccharide. Results showed that the

anti-diabetic activity was improved with the increase of selenium content of *Catathelasma ventricosum* polysaccharide with triple helical structure. However, when the tri-helical structure was damaged, the anti-diabetic effect was decreased. With respect to the structure–activity relationship of proteins, it has been reported that the bioactivity of proteins was highly related to its structure characteristics, such as amino acid composition, amino acid sequence, molecular weight, and hydrophobicity (Silva et al. 2017).

## 4 Application

Because natural polysaccharides have wide biological activities and low toxicity, some of them have been successfully applied in fields of drugs and foods (Table 14.6). In the field of drugs, some polysaccharides such as Lentinan, *Astragalus* polysaccharide, Ginseng polysaccharide, Poria polysaccharide, and Chondroitin have been developed into injections, tablets, and capsules. In the field of foods, some polysaccharides have been used as the additives to endow new nutritional and healthy functions of foods. For instance, *Lycium barbarum* polysaccharides (LBPs) have been processed into different forms of functional foods, such as Goji beverage, Goji wine, Goji tea, Goji oral liquid, Goji tablet, Goji capsule, and Goji granules. Among these healthy foods, the Goji capsule and Goji oral liquid were popular in the public. Long-term use of Goji capsule and Goji oral liquid can enhance the immunity, improve sleep, protect liver, and reduce fatigue (Wu et al. 2018).

Proteins are the important components in foods. Since soybean protein isolate was produced in large-scale in 1958, the application of food protein becomes more and more pluralistic. In the field of traditional foods, the soybean protein, milk protein, egg protein, meat protein, and nut protein have been applied in beverage food, baby food, baked food, pastry, and meat products. In the field of non-traditional foods, the protein has been used as a main component to prepare reproduced foods and simulated foods, such as soybean protein beef, plant protein chicken, vegetarian ham, vegetarian sausage, and imitation meat hamburger. Moreover, food protein widely applied in cosmetics and biomedicine.

## 5 Conclusions and Future Prospects

In recent decades, the research of bioactive polysaccharides has made some important progress and has been widely used in pharmaceutical, biochemical cosmetic, and functional food industries. However, due to the limitations of existing experimental methods and the complexity of their structure, research about polysaccharides is still far behind than that of proteins and nucleic acids. Although the functional properties of polysaccharides have been widely studied, the mechanism of their actions is still unknown. One of the main reasons is that the structural

**Table 14.6** Practical Applications of Polysaccharides

Applications areas	Polysaccharides	Practical applications	Bioactivities	References
Clinical drugs and medicines	<i>Astragalus polysaccharide</i>	Astragalus polysaccharide injection	Immunomodulation; Antioxidant; Anti-hypertensive	Xu (2012) Zhang et al. (2016a, b, c)
	<i>Ginseng polysaccharide</i>	Ginseng polysaccharide injection	Anti-tumor immunity	Xu (2015)
	<i>Lentinan polysaccharide</i>	Lentinan injection; Lentinan capsules	Immunomodulation; Anti-tumor	Wang (2012) Wang et al. (2013)
	<i>Poria polysaccharide</i>	Poria polysaccharide oral solution	Anti-gastric cancer	Yang et al. (2017) Hou and Luo (2017)
	<i>Chondroitin sulfate</i>	Chondroitin sulfate tablets; chondroitin sulfate capsules	Anti-arthritis; Anti-angiocardopathy	Gacci et al. (2015) Liu et al. (2014a, b)
	<i>Fucoidan</i>	Active pharmaceutical ingredient; Antivirus drugs	Hypolipidemic; Antivirus	Wu and Yang (2010) Mandal et al. (2007)
	<i>Ganoderma atrum polysaccharide</i>	Hypoglycemic drugs	Hypoglycemic	Zhu et al. (2013)
	<i>Heparin</i>	Anticoagulant drugs	Anticoagulant	Bai and Ahsan (2009) Dong and Fang (2001)
Food industry	<i>Exopolysaccharides</i>	Fermented dairy products	Anti-ulcer; Immunomodulation; Anti-tumor	Yang et al. (2010)
	<i>Soybean soluble polysaccharide</i>	Yogurt; Milk beverage	Anti-hypertensive; Reduce weight; Hypoglycemic	Ron et al. (2010) Nakamura et al. (2006)
	<i>Red ginseng polysaccharide</i>	Noodles, bread, or cake making	Immunomodulation; Anti-tumor	Yu et al. (2012)
	<i>Carrageenan</i>	Desserts (ice cream and puddings)	Antivirus	Prajapati et al. (2014)
Cosmetics industry	<i>Hyaluronic acid</i>	Sodium hyaluronate injection	Moisture absorption and moisture retention	Zhang et al. (2008)
	<i>Aloe polysaccharides</i>	Aloe Vera gel	Anti-aging; Versatile skin care	Takahashi et al. (2009)
	<i>Marine algae polysaccharides</i>	Marine algae deep moisturizing cream	Anti-radiation; Whitening and moisturizing	Moore (2002)

diversity and heterogeneity of natural polysaccharides hamper the research and product development. Moreover, the polysaccharides extracted from the same raw material by different preparation methods often have different compositions. Therefore, the preparation process of polysaccharides needs to be standardized. To reveal the structure–activity relationship is always the focus of polysaccharides research, which will disclose the structural basis for polysaccharides to perform their healthy functions. This is not only necessary to screen and design high-active polysaccharides, but also have important theoretical guiding significance to study the medicinal mechanism of polysaccharides. In addition, because polysaccharides have a wide range of biological activities, they will have broad application prospects in functional foods in the future. Therefore, it is also necessary to strengthen the research on the activity development and mechanism of polysaccharides.

## References

- Agyei D, Danquah MK (2012) Rethinking food-derived bioactive peptides for antimicrobial and immunomodulatory activities. *Trends Food Sci Technol* 23:62–69
- Ahn G, Park E, Lee WW et al (2011) Enzymatic extract from *Ecklonia cava* induces the activation of lymphocytes by IL-2 production through the classical NF- $\kappa$ B pathway. *Mar Biotechnol* 13:66–73
- Arthur JSC, Ley SC (2013) Mitogen-activated protein kinases in innate immunity. *Nat Rev Immunol* 13(9):679–692
- Baek SH, Lee JG, Park SY et al (2010) Pectic polysaccharides from *Panax ginseng* as the antirotavirus principals in ginseng. *Biomacromolecules* 11(8):2044–2052
- Bai D, Chang NT, Li DH et al (2008) Antitubercular activity of *Ganoderma lucidum* polysaccharides. *Acta Agric Bor Sin* 23(S1):282–285
- Bai S, Ahsan F (2009) Synthesis and evaluation of pegylated dendrimeric nanocarrier for pulmonary delivery of low molecular weight heparin. *Pharm Res* 26:539–548
- Bao HH, Wonseok C, You SG (2010) Effect of sulfated modification on the molecular characteristics and biological activities of polysaccharides from *Hypsizigus marmoreus*. *Biosci Biotech Bioch* 74:1408–1414
- Bao L, Wang Y, Ma R et al (2015) Apoptosis-inducing effects of *lentinan* on the proliferation of human bladder cancer T24 cells. *Pak J Pharm Sci* 28(5):1595–1600
- Bao X, Liu C, Fang J et al (2001) Structural and immunological studies of a major polysaccharide from spores of *Ganoderma lucidum* (fr.) karst. *Carbohydr Res* 332(1):67–74
- Bao XF, Wang XS, Dong Q et al (2002a) Structural features of immunologically active polysaccharides from *Ganoderma lucidum*. *Phytochemistry* 59(2):175–181
- Bao XF, Zhen Y, Ruan L et al (2002b) Purification, characterization, and modification of T lymphocyte-stimulating polysaccharide from spores of *Ganoderma lucidum*. *Chem Pharm Bull* 50(5):623–629
- Belhaj D, Athmouni K, Ahmed MB et al (2018) Polysaccharides from *Phormidium versicolor* (NCC466) protecting HepG2 human hepatocellular carcinoma cells and rat liver tissues from cadmium toxicity: evidence from in vitro and in vivo tests. *Int J Biol Macromol* 113:813–820
- Bell S, Goldman VM, Bistrain BR et al (1999) Effect of  $\beta$ -glucan from oats and yeast on serum lipids. *Crit Rev Food Sci* 39(2):189–202
- Besch R, Poeck H, Hohenauer T et al (2009) Proapoptotic signaling induced by RIG-I and MDA-5 results in type I interferon-independent apoptosis in human melanoma cells. *J Clin Invest* 119(8):2399–2411

- Bhattacharyya C, De S, Basak A et al (2006) Antimicrobial activities of some *Basidiomycetous fungi*. *J Mycopathol Res* 44:129–135
- Bhunia SK, Dey B, Maity KK et al (2010) Structural characterization of an immunoenhancing heteroglycan isolated from an aqueous extract of an edible mushroom, *Lentinus squarrosulus* (mont.) singer. *Carbohydr Res* 345(17):2542–2549
- Bose N, Wurst LR, Chan AS et al (2014) Differential regulation of oxidative burst by distinct beta-glucan-binding receptors and signaling pathways in human peripheral blood mononuclear cells. *Glycobiology* 24:379–391
- Cai X, Wang YF, Mao FF et al (2011) Hypoglycemic and hyperglycemia-prevention effects of crude tea flower polysaccharide. *Mod Food Sci Technol* 27:262–266
- Chan G, Chan W, Sze D (2009) The effects of beta-glucan on human immune and cancer cells. *J Hematol Oncol* 2(1):25
- Chen JR, Yang ZQ, Hu TJ et al (2010a) Immunomodulatory activity in vitro and in vivo of polysaccharide from *Potentilla anserina*. *Fitoterapia* 81(8):1117–1124
- Chen KL, Weng BC, Chang MT et al (2008) Direct enhancement of the phagocytic and bactericidal capability of abdominal macrophage of chicks by beta-1, 3-1, 6-glucan. *Poult Sci* 87:2242–2249
- Chen X, Jin J, Tang J et al (2011) Extraction, purification, characterization and hypoglycemic activity of a polysaccharide isolated from the root of *Ophiopogon japonicus*. *Carbohydr Polym* 83(2):749–754
- Chen XQ, Lin Z, Ye Y et al (2010b) Suppression of diabetes in non-obese diabetic (NOD) mice by oral administration of water-soluble and alkali-soluble polysaccharide conjugates prepared from green tea. *Carbohydr Polym* 82:28–33
- Chen Y, Zhang H, Wang Y et al (2015) Sulfated modification of the polysaccharides from *Ganoderma atrum* and their antioxidant and immunomodulating activities. *Food Chem* 186:231–238
- Cheng BH, Chan JYW, Chan BCL et al (2014) Structural characterization and immunomodulatory effect of a polysaccharide HCP-2 from *Houttuynia cordata*. *Carbohydr Polym* 103:244–249
- Chihara G, Maeda Y, Hamuro J et al (1969) Inhibition of mouse sarcoma 180 by polysaccharides from *Lentinus edodes* (Berk.) sing. *Nature* 222:687–688
- Cho CW, Han CJ, Rhee YK et al (2015) *Cheonggukjang* polysaccharides enhance immune activities and prevent cyclophosphamide-induced immunosuppression. *Int J Biol Macromol* 72:519–525
- Cristina Diaz A, Laura Espino M, Arzoz NS et al (2017) Free radical scavenging activity of extracts from seaweeds *Macrocystis pyrifera* and *Undaria pinnatifida*: applications as functional food in the diet of prawn *Artemesia longinaris*. *Lat Am J Aquat Res* 45(1):104–112
- Cui H, Wu S, Shang Y et al (2016) *Pleurotus nebrodensis* polysaccharide (PN50G) evokes A549 cell apoptosis by the ROS/AMPK/PI3K/AKT/mTOR pathway to suppress tumor growth. *Food Funct* 7(3):1616–1627
- Di T, Chen G, Sun Y et al (2017) Antioxidant and immunostimulating activities in vitro of sulfated polysaccharides isolated from *Gracilaria rubra*. *J Funct Foods* 28:64–75
- Diao Y, Xin Y, Zhou Y et al (2014) Extracellular polysaccharide from *Bacillus sp. strain* LBP32 prevents LPS-induced inflammation in RAW 264.7 macrophages by inhibiting NF- $\kappa$ B and MAPKs activation and ROS production. *Int Immunopharmacol* 18(1):12–19
- Díaz-Gómez J, Ortiz-Martínez M, Aguilar O et al (2018) Antioxidant activity of zein hydrolysates from zein species and their cytotoxic effects in a hepatic cell culture. *Molecules* 23(2):312–326
- DiDonato JA, Mercurio F, Karin M (2012) NF- $\kappa$ B and the link between inflammation and cancer. *Immunol Rev* 246(1):379–400
- Dong Q, Fang J (2001) Structural elucidation of a new arabinogalactan from the leaves of *Nerium indicum*. *Carbohydr Res* 332(1):109–114
- Dong Q, Yao J, Fang JN et al (2007) Structural characterization and immunological activity of two cold-water extractable polysaccharides from *Cistanche deserticola* Y. C. Ma. *Carbohydr Res* 342(10):1343–1349

- Dore C, Alves M, Santos M et al (2014) Antioxidant and anti-inflammatory properties of an extract rich in polysaccharides of the mushroom *Polyporus dermatopus*. *Antioxidants (Basel)* 3 (4):730–744
- Esposito K, Nappo F, Marfella R et al (2002) Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation* 106(16):2067
- Faccin LC, Benati F, Rincão VP et al (2007) Antiviral activity of aqueous and ethanol extracts and of an isolated polysaccharide from *Agaricus brasiliensis* against poliovirus type I. *Lett Appl Microbiol* 45:24–28
- Fan L, Ding S, Ai L et al (2012a) Antitumor and immunomodulatory activity of water-soluble polysaccharide from *inonotus obliquus*. *Carbohydr Polym* 90(2):870–874
- Fan L, Li J, Deng K et al (2012b) Effects of drying methods on the antioxidant activities of polysaccharides extracted from *Ganoderma lucidum*. *Carbohydr Polym* 87(2):1849–1854
- Fan X, Subramaniam R, Weiss MF et al (2003) Methylglyoxal-bovine serum albumin stimulates tumor necrosis factor alpha secretion in RAW 264.7 cells through activation of mitogen-activating protein kinase, nuclear factor kappaB and intracellular reactive oxygen species formation. *Arch Biochem Biophys* 409:274–286
- Fang Q, Wang JF, Zha XQ et al (2015) Immunomodulatory activity on macrophage of a purified polysaccharide extracted from *Laminaria japonica*. *Carbohydr Polym* 134:66–73
- Feng Y, Li W, Wu X et al (2010) Rapid and efficient microwave-assisted sulfate modification of *lentinan* and its antioxidant and antiproliferative activities in vitro. *Carbohydr Polym* 82 (3):605–612
- Ferreira IC, Heleno SA, Reis FS et al (2015a) Chemical features of *ganoderma* polysaccharides with antioxidant, antitumor and antimicrobial activities. *Phytochemistry* 114:38–55
- Ferreira SS, Passos CP, Madureira P et al (2015b) Structure-function relationships of immunostimulatory polysaccharides: a review. *Carbohydr Polym* 132:378–396
- Ferwerda G, Meyer-Wentrup F, Kullberg BJ et al (2008) Dectin-1 synergizes with TLR2 and TLR4 for cytokine production in human primary monocytes and macrophages. *Cell Microbiol* 10 (10):2058–2066
- Figueiredo RT, Bittencourt VCB, Lopes LCL et al (2012) Toll-like receptors (TLR2 and TLR4) recognize polysaccharides of *Pseudallescheria boydii* cell wall. *Carbohydr Res* 356:260–264
- Fiorito S, Epifano F, Prezioso F et al (2018) Selenylated plant polysaccharides: a survey of their chemical and pharmacological properties. *Phytochemistry* 153:1–10
- Gacci M, Saleh O, Giannessi C et al (2015) Sodium hyaluronate and chondroitin sulfate replenishment therapy can improve nocturia in men with post-radiation cystitis: results of a prospective pilot study. *BMC Urol* 15(1):1–6
- Goldberg J, Shrikant P, Mescher MF (2003) In vivo augmentation of tumor-specific CTL responses by class I/peptide antigen complexes on microspheres (large multivalent immunogen). *J Immunol* 170:228–235
- Guerra Dore CMP, Azevedo TCG, De Souza MCR et al (2007) Antiinflammatory, antioxidant and cytotoxic actions of  $\beta$ -glucan-rich extract from *Geastrum saccatum* mushroom. *Int Immunopharmacol* 7:1160–1169
- Gunnarsson PT, Winzell MS, Deacon CF et al (2006) Glucose-induced incretin hormone release and inactivation are differently modulated by oral fat and protein in mice. *Endocrinology* 147 (7):3173–3180
- Guo L, Xie J, Ruan Y et al (2009) Characterization and immunostimulatory activity of a polysaccharide from the spores of *Ganoderma lucidum*. *Int Immunopharmacol* 9:1175–1182
- Han K, Jin C, Chen H et al (2018) Structural characterization and anti-A549 lung cancer cells bioactivity of a polysaccharide from *Houttuynia cordata*. *Int J Biol Macromol* 120:288–296
- Han Q, Yu QY, Shi J et al (2011a) Molecular characterization and hypoglycemic activity of a novel water soluble polysaccharide from tea (*Camellia sinensis*) flower. *Carbohydr Polym* 86:797–805

- Han QA, Yu QY, Shi JA et al (2011b) Structural characterization and antioxidant activities of 2 water soluble polysaccharide fractions purified from tea (*Camellia sinensis*) flower. *J Food Sci* 76:462–471
- Harman D (1993) Free radical involvement in aging pathophysiology and therapeutic implications. *Drug Aging* 3(1):60–80
- Harnedy PA, O’Keeffe M, FitzGerald R (2015) Purification and identification of dipeptidyl peptidase (DPP) IV inhibitory peptides from the macroalga *Palmaria palmata*. *Food Chem* 172:400–406
- Hazama S, Watanabe S, Ohashi M et al (2009) Efficacy of orally administered superfine dispersed lentinan ( $\beta$ -1,3-glucan) for the treatment of advanced colorectal cancer. *Anticancer Res* 29 (7):2611–2617
- He L, Ji P, Cheng J et al (2013) Structural characterization and immunostimulatory activity of a novel protein-bound polysaccharide produced by *Hirsutella sinensis* Liu, Guo, Yu & Zeng. *Food chem* 141(2):946–953
- Hernandez-Ledesma B, Hsieh CC, De Lumen BO (2009) Lunasin, a novel seed peptide for cancer prevention. *Peptides* 30(2):426–430
- Hetland G, Ohno N, Aaberge IS et al (2000) Protective effect of  $\beta$ -glucan against systemic *Streptococcus pneumoniae* infection in mice. *FEMS Immuno Med Mic* 27:111–116
- Hira T, Mochida T, Miyashita K et al (2009) GLP-1 secretion is enhanced directly in the ileum but indirectly in the duodenum by a newly identified potent stimulator, zein hydrolysate, in rats. *Am J Physiol Gastroint Liver Physiol* 297(4):G663–G671
- Hou H, Fan Y, Wang S et al (2016) Immunomodulatory activity of Alaska Pollock hydrolysates obtained by glutamic acid biosensor-artificial neural network and the identification of its active central fragment. *J Funct Foods* 24:37–47
- Hou WT, Luo JB (2017) The study of the compound *poria* polysaccharide oral liquid on the antitumor activity and immune regulation function. *Pharm Clin Chin Mat Med* 33(2):78–81
- Hsu K, Li-Chan E, Jao C (2011) Antiproliferative activity of peptides prepared from enzymatic hydrolysates of tuna dark muscle on human breast cancer cell lines MCF-7. *Food Chem* 126:617–622
- Hsu SY, Lee WJ, Chong K et al (2015) Laparoscopic bariatric surgery for the treatment of severe hypertriglyceridemia. *Asian J Surg* 38(2):96–101
- Hu F, Li X, Zhao L et al (2010) Antidiabetic properties of purified polysaccharide from *Hedysarum polybotrys*. *Can J Physiol Pharm* 88(1):64–72
- Huie C, Di X (2004) Chromatographic and electrophoretic methods for lingzhi pharmacologically active components. *J Chromatogr B* 812(1–2):241–257
- Ina K, Furuta R, Kataoka T et al (2016) P1-035 chemo-immunotherapy using *lentinan* for the treatment of inoperable gastric cancer with multiple liver metastases. *Ann Oncol* 27(suppl 7): vii94
- Jiao Y, Wang X, Jiang X et al (2017) Antidiabetic effects of *Morus alba* fruit polysaccharides on high-fat diet- and streptozotocin-induced type 2 diabetes in rats. *J Ethnopharmacol* 199:119–127
- Jin ML, Zhao K, Huang QS et al (2012) Isolation, structure and bioactivities of the polysaccharides from *Angelica sinensis* (Oliv.) Diels: a review. *Carbohydr Polym* 89(3):713–722
- Jing L, Cui G, Feng Q et al (2009) Orthogonal test design for optimization of the extraction of polysaccharides from *Lycium barbarum* and evaluation of its anti-athletic fatigue activity. *J Med Plants Res* 3(5):433–437
- José ML, Castro R, Arranz JA et al (2007) Immunomodulating activities of acidic sulphated polysaccharides obtained from the seaweed *Ulva rigida* C. Agardh. *Int Immunopharmacol* 7 (7):879–888
- Kannan A, Hettiarachchy NS, Lay JO et al (2010) Human cancer cell proliferation inhibition by a pentapeptide isolated and characterized from rice bran. *Peptides* 31(9):1629–1634
- Kant P, Liu WZ, Pauls KP (2009) PDC1, a corn defensin peptide expressed in *Escherichia coli* and *Pichia pastoris* inhibits growth of *Fusarium graminearum*. *Peptides* 30(9):1593–1599



- Kawai T, Akira S (2010) The role of pattern-recognition receptors in innate immunity: update on toll-like receptors. *Nat Immunol* 11(5):373–384
- Kiho T, Kochi M, Usui S et al (2001) Antidiabetic effect of an acidic polysaccharide (TAP) from *tremella aurantia* and its degradation product (TAP-H). *Biol Pharm Bull* 24(12):1400–1403
- Kiho T, Ookubo K, Usui S et al (1999) Structural features and hypoglycemic activity of a polysaccharide (CS-F10) from the cultured mycelium of *Cordyceps sinensis*. *Biol Pharm Bull* 22(9):966–970
- Kiho T, Yamane A, Hui J et al (1996) Polysaccharides in fungi. xxxvi. hypoglycemic activity of a polysaccharide (CS-F30) from the cultural mycelium of *Cordyceps sinensis* and its effect on glucose metabolism in mouse liver. *Biol Pharm Bull* 19(2):294–296
- Kim HM, Kang JS, Kim JY et al (2010) Evaluation of antidiabetic activity of polysaccharide isolated from *Phellinus linteus* in non-obese diabetic mouse. *Int Immunopharmacol* 10(1):72–78
- Klaus A, Kozarski M, Niksic M et al (2011) Antioxidative activities and chemical characterization of polysaccharides extracted from the *basidiomycete* schizophyllum commune. *LWT- Food Sci Technol* 44(10):2005–2011
- Kumar H, Kawai T, Akira S (2011) Pathogen recognition by the innate immune system. *Int Rev Immunol* 30(1):16–34
- Lai CY, Hung JT, Lin HH et al (2010) Immunomodulatory and adjuvant activities of a polysaccharide extract of *Ganoderma lucidum* in vivo and in vitro. *Vaccine* 28(31):4945–4954
- Lazaridou A, Biliaderis CG (2004) Cryogelation of cereal  $\beta$ -glucans: structure and molecular size effects. *Food Hydrocolloid* 18(6):933–947
- Li C, Huang Q, Fu X et al (2015) Characterization, antioxidant and immunomodulatory activities of polysaccharides from *Prunella vulgaris* linn. *Int J Biol Macromol* 75:298–305
- Li F, Li Q, Gao D et al (2009a) Preparation and antidiabetic activity of polysaccharide from *Portulaca oleracea* L. *Afr J Biotechnol* 8(4):569–573
- Li F, Zhang Y, Zhong Z et al (2011) Antihyperglycemic effect of *Ganoderma lucidum* polysaccharides on streptozotocin-induced diabetic mice. *Int J Mol Sci* 12(12):6135–6145
- Li SL, Huang ZN, Hsieh HH et al (2009b) The augmented anti-tumor effects of *Anrotdia camphorata* co-fermented with Chinese medicinal herb in human hepatoma cells. *Am J Chin Med* 37(4):771–783
- Li SP, Zhang GH, Zeng Q et al (2006a) Hypoglycemic activity of polysaccharide with antioxidation isolated from cultured *Cordyceps mycelia*. *Phytomedicine* 13(6):428–433
- Li W, Wang Q, Cui SW et al (2006b) Elimination of aggregates of (1 $\rightarrow$ 3) (1 $\rightarrow$ 4)- $\beta$ -d-glucan in dilute solutions for light scattering and size exclusion chromatography study. *Food Hydrocolloid* 20:361–368
- Li YG, Ji DF, Zhong S et al (2013a) Polysaccharide from *Pellinus linteus* induces S-phase arrest in HepG2 cells by decreasing calreticulin expression and activating the P27kip1-cyclin A/D1/E-CDK2 pathway. *J Ethnopharmacol* 150(1):187–195
- Li YG, Ji DF, Zhong S et al (2013b) Polysaccharide from *Pellinus linteus* induces S-phase arrest in HepG2 cells by decreasing calreticulin expression and activating the P27kip1-cyclin A/D1/E-CDK2 pathway. *J Ethnopharmacol* 150(1):187–195
- Liao W, Luo Z, Liu D et al (2015) Structure characterization of a novel polysaccharide from *Dictyophora indusiata* and its macrophage immunomodulatory activities. *J Agric Food Chem* 63(2):535–544
- Lim JY, Kim OK, Lee J et al (2014) Protective effect of the standardized green tea seed extract on uvb-induced skin photoaging in hairless mice. *Nutr Res Pract* 8(4):398–403
- Lin L, Cheng K, Xie Z et al (2018) Purification and characterization a polysaccharide from *Hedyotis diffusa* and its apoptosis inducing activity toward human lung cancer cell line A549. *Int J Biol Macromol* 122:64–71
- Liu G, Kuang S, Wu S et al (2016) A novel polysaccharide from *Sargassum integerrimum* induces apoptosis in A549 cells and prevents angiogenesis in vitro and in vivo. *Sci Rep* 6:26722
- Liu QM, Xu SS, Li L et al (2017a) In vitro and in vivo immunomodulatory activity of sulfated polysaccharide from *Porphyra haitanensis*. *Carbohydr Polym* 165:189–196

- Liu QY, Yao YM, Zhang SW et al (2011a) *Astragalus* polysaccharides regulate T cell-mediated immunity via CD11chighcd45RBlow DCs in vitro. *J Ethnopharmacol* 136(3):457–464
- Liu QY, Yao YM, Zhang SW et al (2011b) *Astragalus* polysaccharides regulate T cell-mediated immunity via CD11chighcd45RBlow DCs in vitro. *J Ethnopharmacol* 136(3):457–464
- Liu Y, Du YQ, Wang JH et al (2014a) Structural analysis and antioxidant activities of polysaccharide isolated from *jinqian mushroom*. *Int J Biol Macromol* 64:63–68
- Liu Y, Liu Y, Jiang H et al (2014b) Preparation, antiangiogenic and antitumoral activities of the chemically sulfated glucan from *Pellinus ribis*. *Carbohydr Polym* 106:42–48
- Liu Y, You Y, Li Y et al (2017b) The characterization, selenylation and antidiabetic activity of mycelial polysaccharides from *Catathelasma ventricosum*. *Carbohydr Polym* 174:72–81
- Liu Y, Zhao J, Zhao Y et al (2018) Therapeutic effects of lentinan on inflammatory bowel disease and colitis-associated cancer. *J Cell Mol Med* 23(2):1–11
- Lo TCT, Jiang YH, Chao AL et al (2007) Use of statistical methods to find the polysaccharide structural characteristics and the relationships between monosaccharide composition ratio and macrophage stimulatory activity of regionally different strains of *Lentinula edodes*. *Anal Chim Acta* 584:50–56
- Luo C, Luo C, Wang X et al (2016) Molecular inhibition mechanisms of cell migration and invasion by *coix* polysaccharides in A549 NSCLC cells via targeting S100A4. *Mol Med Rep* 15(1):309–316
- Ma L, Chen H, Zhang Y et al (2012) Chemical modification and antioxidant activities of polysaccharide from mushroom *inonotus obliquus*. *Carbohydr Polym* 89(2):371–378
- Maeda R, Ida T, IHARA H et al (2012) Immunostimulatory activity of polysaccharides isolated from *Caulerpa lentillifera* on macrophage cells. *Biosci Biotech Bioch* 76(3):501–505
- Maji PK, Sen IK, Behera B et al (2012) Structural characterization and study of immunoenhancing properties of a glucan isolated from a hybrid mushroom of *pleurotus florida* and *lentinula edodes*. *Carbohydr Res* 358:110–115
- Mandal P, Mateu CG, Chattopadhyay K et al (2007) Structural features and antiviral activity of sulphated fucans from the *brown seaweed cystoseira indica*. *Antivir Chem Chemother* 18(3):153–162
- Mao GH, Ren Y, Feng WW et al (2015) Antitumor and immunomodulatory activity of a water-soluble polysaccharide from *grifola frondosa*. *Carbohydr Polym* 134:406–412
- Masuda Y, Inoue H, Ohta H et al (2013) Oral administration of soluble  $\beta$ -glucans extracted from, *grifola frondosa*, induces systemic antitumor immune response and decreases immunosuppression in tumor-bearing mice. *Int J Cancer* 133(1):108–119
- Matteucci E, Giampietro O (2009) Dipeptidyl peptidase-4 (CD26): knowing the function before inhibiting the enzyme. *Curr Med Chem* 16:2943–2951
- Meng FY (2010) Extraction optimization and in vivo antioxidant activities of exopolysaccharide by *morchella esculenta* SO-01. *Bioresour Technol* 101(12):4564–4569
- Meng J, Hu X, Shan F et al (2011) Analysis of maturation of murine dendritic cells (DCs) induced by purified *ganoderma lucidum* polysaccharides (GLPs). *Int J Biol Macromol* 49(4):693–699
- Mochida T, Hira T, Hara H (2010) The corn protein, zein hydrolysate, administered into the ileum attenuates hyperglycemia via its dual action on glucagon-like peptide-1 secretion and dipeptidyl peptidase-IV activity in rats. *Endocrinology* 151(7):3095–3104
- Mohan K, Padmanaban AM, Uthayakumar V et al (2017) Anti-cancer effect of the polysaccharide extract from the *ganoderma lucidum* against hela cell lines. *Bangladesh J Pharmacol* 12(1):56–57
- Moore A (2002) The biochemistry of beauty. *EMBO Rep* 3(8):714–717
- Murphy EA, Davis JM, Carmichael MD et al (2009) Benefits of oat  $\beta$ -glucan and sucrose feedings on infection and macrophage antiviral resistance following exercise stress. *Am J Phys* 297:1188–1194
- Nagaoka M, Shibata H, Kimura-Takagi I et al (2010) Anti-ulcer effects and biological activities of polysaccharides from *marine algae*. *Biofactors* 12:267–274

- Nakamura A, Yoshida R, Maeda H et al (2006) The stabilizing behaviour of *soybean* soluble polysaccharide and pectin in acidified milk beverages. *Int Dairy J* 16(4):361–369
- Nie S, Xie M, Fu Z et al (2008) Study on the purification and chemical compositions of tea glycoprotein. *Carbohydr Polym* 71(4):626–633
- Nie SP, Cui SW, Phillips AO et al (2011) Elucidation of the structure of a bioactive hydrophilic polysaccharide from *cordyceps sinensis* by methylation analysis and nmr spectroscopy. *Carbohydr Polym* 84(3):894–899
- Nie SP, Cui SW, Xie MY (2018) Bioactive polysaccharides. Elsevier, Amsterdam, pp 1–141
- Nongonierma AB, FitzGerald RJ (2016) Strategies for the discovery, identification and validation of milk protein-derived bioactive peptides. *Trends Food Sci Technol* 50:26–43
- Oba K, Kobayashi M, Matsui T et al (2009) Individual patient based meta-analysis of lentinan for unresectable/recurrent gastric cancer. *Anticancer Res* 29(7):2739–2745
- Otani H, Nakano K, Kawahara T (2003) Stimulatory effect of a dietary casein phosphopeptide preparation on the mucosal IgA response of mice to orally ingested lipopolysaccharide from *Salmonella typhimurium*. *Biosci Biotech Bioch* 67(4):729–735
- Palacios I, García-Lafuente A, Guillamón E et al (2012) Novel isolation of water-soluble polysaccharides from the fruiting bodies of *pleurotus ostreatus* mushrooms. *Carbohydr Res* 358:72–77
- Pan DD, Wu Z, Liu J et al (2013a) Immunomodulatory and hypoallergenic properties of milk protein hydrolysates in ICR mice. *J Dairy Sci* 96(8):4958–4964
- Pan LH, Feng BJ, Wang JH et al (2013b) Structural characterization and anti-glycation activity in vitro of a water-soluble polysaccharide from *dendrobium huoshanense*. *J Food Biochem* 37(3):313–321
- Pan Y, Wang C, Chen Z et al (2017) Physicochemical properties and antidiabetic effects of a polysaccharide from corn silk in high-fat diet and streptozotocin-induced diabetic mice. *Carbohydr Polym* 164:370–378
- Peña-Ramos EA, Xiong YL (2002) Antioxidative activity of whey protein hydrolysates in a liposomal system. *J Dairy Sci* 84(12):2577–2583
- Peng GY, Guo Z, Kuniwa Y, Voo KS et al (2005) Toll-like receptor 8-mediated reversal of CD4+ regulatory T cell function. *Science* 309(5739):1380–1384
- Pietrzycka A, Stepniewski M, Waszkielewicz AM et al (2006) Effect of vita glucan on some antioxidant parameters of the human blood in vitro study. *Acta Pol Pharm* 63(6):547
- Pimentel FB, Alves RC, Harnedy PA et al (2019) Macroalgal-derived protein hydrolysates and bioactive peptides: enzymatic release and potential health enhancing properties. *Trends Food Sci Technol* 93:106–124
- Plüddemann A, Mukhopadhyay S, Gordon S (2011) Innate immunity to intracellular pathogens: macrophage receptors and responses to microbial entry. *Immunol Rev* 240(1):11–24
- Poeck H, Besch R, Maihoefer C et al (2008) 5'-triphosphate-sirna: turning gene silencing and rig-i activation against melanoma. *Nat Med* 14(11):1256–1263
- Pomeroy S, Tupper R, Cehun-Aders M et al (2001) Oat  $\beta$ -glucan lowers total and LDL-cholesterol. *Aust J Nutr Diet* 58:51–55
- Prajapati VD, Maheriya PM, Jani GK et al (2014) Carrageenan: a natural seaweed polysaccharide and its applications. *Carbohydr Polym* 105:97–112
- Prashanth KVH, Tharanathan RN (2007) Chitin/chitosan: modifications and their unlimited application potential—an overview. *Trends Food Sci Technol* 18(3):117–131
- Qian Y, Wang D, Fan M et al (2018) Effects of intrinsic metal ions of lentinan with different molecular weights from *Lentinus edodes* on the antioxidant capacity and activity against proliferation of cancer cells. *Int J Biol Macromol* 120:73–81
- Rice PJ, Adams EL, Ozment-Skelton T et al (2005) Oral delivery and gastrointestinal absorption of soluble glucans stimulate increased resistance to infectious challenge. *J Pharmacol Exp Ther* 314:1079–1086

- Ron N, Zimet P, Bargarum J et al (2010) Beta-lactoglobulin-polysaccharide complexes as nanovehicles for hydrophobic nutraceuticals in non-fat foods and clear beverages. *Int Dairy J* 20:686–693
- Rosenson RS, Otvos JD, Freedman DS (2002) Relations of lipoprotein subclass levels and low-density lipoprotein size to progression of coronary artery disease in the pravastatin limitation of atherosclerosis in the coronary arteries (plac-i) trial. *Am J Cardiol* 90(2):89–94
- Saluk J, Bijak M, Ponczek MB et al (2013) (1→3)- $\beta$ -d-glucan reduces the damages caused by reactive oxygen species induced in human platelets by lipopolysaccharides. *Carbohydr Polym* 97(2):716–724
- Sanjukta S, Rai AK (2016) Production of bioactive peptides during soybean fermentation and their potential health benefits. *Trends Food Sci Technol* 50:1–10
- Schepetkin IA, Faulkner CL, Nelson-Overton LK et al (2005) Macrophage immunomodulatory activity of polysaccharides isolated from *Juniperus scopolorum*. *Int Immunopharmacol* 5 (13–14):1783–1799
- Schepetkin IA, Quinn MT (2006) Botanical polysaccharides: macrophage immunomodulation and therapeutic potential. *Int Immunopharmacol* 6(3):317–333
- Sener G, Toklu H, Ercan F et al (2005) Protective effect of  $\beta$ -glucan against oxidative organ injury in a rat model of sepsis. *Int Immunopharmacol* 5(9):1387–1396
- Shah N, Shah T, Amin A (2011) Polysaccharides: a targeting strategy for colonic drug delivery. *Expert Opin Drug Deliv* 8:779–796
- Sharma SK, Gautam N, Atri NS (2015) Optimized extraction, composition, antioxidant and antimicrobial activities of exo and intracellular polysaccharides from submerged culture of *cordyceps cicadae*. *BMC Complement Altern Med* 15(1):446
- Shen J, Park HS, Xia YM et al (2014) The polysaccharides from fermented *ganoderma lucidum* mycelia induced miRNAs regulation in suppressed HepG2 cells. *Carbohydr Polym* 103:319–324
- Sheu F, Chien PJ, Chien AL et al (2004) Isolation and characterization of an immunomodulatory protein (APP) from the Jew's Ear mushroom *Auricularia polytricha*. *Food Chem* 87(4):593–600
- Shi X, Wei W, Wang N (2018) *Tremella* polysaccharides inhibit cellular apoptosis and autophagy induced by *Pseudomonas aeruginosa* lipopolysaccharide in A549 cells through sirtuin 1 activation. *Oncol Lett* 15:9609–9616
- Shimizu K, Watanabe S, Watanabe S et al (1999) Efficacy of oral administered superfine dispersed lentinan for advanced pancreatic cancer. *Hepato-Gastroenterology* 56(89):240–244
- Shriner AK, Liu H, Sun G et al (2010) IL-7-dependent B lymphocytes are essential for the anti-polysaccharide response and protective immunity to *streptococcus pneumoniae*. *J Immunol* 185 (1):525–531
- Silva FGDE, Hernández-Ledesma B, Amigo L et al (2017) Identification of peptides released from flaxseed (*Linum usitatissimum*) protein by Alcalase®; hydrolysis: antioxidant activity. *LWT Food Sci Technol* 76:140–146
- Song G, Du Q (2012) Structure characterization and antitumor activity of an  $\alpha\beta$ -glucan polysaccharide from *auricularia polytricha*. *Food Res Int* 45(1):381–387
- Stack J, Tobin PR, Gietl A et al (2017) Seasonal variation in nitrogenous components and bioactivity of protein hydrolysates from *Porphyra dioica*. *J Appl Phycol* 29:2439–2450
- Steiner H, Hultmark D, Engstrom A et al (1981) Sequence and specificity of two antibacterial proteins involved in insect immunity. *Nature* 292(5820):246–248
- Steinman RM, Banchereau J (2007) Taking dendritic cells into medicine. *Nature* 449:419–426
- Sun Y, Jiang L, Wei D (2013) Partial characterization, in vitro antioxidant and antiproliferative activities of patatin purified from potato fruit juice. *Food Funct* 4:1502–1511
- Sun Y, Li X, Yang J et al (2010) Water-soluble polysaccharide from the fruiting bodies of *chroogomphus rutilus* (Schaeff.: Fr.) o. k. miller: isolation, structural features and its scavenging effect on hydroxyl radical. *Carbohydr Polym* 80(3):720–724

- Sun Y, Li X, Yang J et al (2010) Water-soluble polysaccharide from the fruiting bodies of *Chroogomphus rutilus* (Schaeff.: Fr.) O. K. Miller: isolation, structural features and its scavenging effect on hydroxyl radical. *Carbohydr Polym* 80(3):720–724
- Suphantharika M, Khunrae P, Thanardkit P et al (2003) Preparation of spent brewer's yeast  $\beta$ -glucans with a potential application as an immunostimulant for black tiger shrimp, *Penaeus monodon*. *Bioresour Technol* 88(1):55–60
- Suresh V, Senthilkumar N, Thangam R et al (2013) Separation, purification and preliminary characterization of sulfated polysaccharides from *Sargassum plagiophyllum* and its in vitro anticancer and antioxidant activity. *Process Biochem* 48(2):364–373
- Taguchi T (1983) Effects of lentinan in advanced or recurrent cases of gastric, colorectal, and breast cancer. *Gan To Kagaku Ryoho* 10(2):387–393
- Takahashi M, Kitamoto D, Asikin Y et al (2009) Liposomes encapsulating *aloe vera* leaf gel extract significantly enhance proliferation and collagen synthesis in human skin cell lines. *J Oleo Sci* 58(12):643–650
- Tani M, Tanimura H, Yamaue H et al (1993) Augmentation of lymphokine-activated killer cell activity by lentinan. *Anticancer Res* 13(5):1773–1776
- Teng Z, Qian L, Zhou Y (2013) Hypolipidemic activity of the polysaccharides from *enteromorpha prolifera*. *Int J Biol Macromol* 62:254–256
- Van der Sman R, Van der Goot A (2009) The science of food structuring. *Soft Matter* 5(3):501–510
- Vannucci L, Krizan J, Sima P et al (2013) Immunostimulatory properties and antitumor activities of glucans (review). *Int J Oncol* 43(2):357–364
- Vidovic D, Graddis T, Chen F et al (2002) Antitumor vaccination with HER-2-derived recombinant antigens. *Int J Cancer* 102:660–664
- Wang CL, Meng M, Liu SB et al (2013) A chemically sulfated polysaccharide from *grifola frondosa* induces HepG2 cell apoptosis by notch1-NF- $\kappa$ B pathway. *Carbohydr Polym* 95(1):282–287
- Wang CL, Pi CC, Kuo CW et al (2011) Polysaccharides purified from the submerged culture of *Ganoderma formosanum* stimulate macrophage activation and protect mice against *Listeria monocytogenes* infection. *Biotechnol Lett* 33:2271–2278
- Wang GJ (2012) A systematic research on lentinan injection preparation quality standard. The Second Military Medical University, Shanghai
- Wang HX, Ng TB (2007) An antifungal peptide from red lentil seeds. *Peptides* 28(3):547–552
- Wang X, Wang Y, Zhou Q et al (2020) Immunomodulatory effect of Lentinan on aberrant T subsets and cytokines profile in non-small cell lung cancer patients. *Pathol Oncol Res* 26(1):499–505
- Wang Y, Zhang L (2006) Chain conformation of carboxymethylated derivatives of (1  $\rightarrow$ 3)- $\beta$ -D-glucan from *poria cocos sclerotium*. *Carbohydr Polym* 65(4):504–509
- Wang YJ, Yu YZ, Mao JW (2009) Carboxymethylated D-glucan derived from *Poria cocos* with biological activities. *J Agric Food Chem* 57(22):10913–10915
- Wijaya W, Patel AR, Setiowati AD et al (2017) Functional colloids from proteins and polysaccharides for food applications. *Trends Food Sci Technol* 68:56–69
- Wood PJ (2007) Cereal  $\beta$ -glucans in diet and health. *J Cereal Sci* 46:230–238
- Wong KH, Lai CKM, Cheung PCK (2011) Immunomodulatory activities of mushroom sclerotial polysaccharides. *Food Hydrocolloid* 25(2):150–158
- Wu DT, Guo H, Lin S et al (2018) Review of the structural characterization, quality evaluation, and industrial application of, *lycium barbarum*, polysaccharides. *Trends Food Sci Technol* 79:171–183
- Wu DT, Meng LZ, Wang LY et al (2014) Chain conformation and immunomodulatory activity of a hyperbranched polysaccharide from *cordyceps sinensis*. *Carbohydr Polym* 110:405–414
- Wu J, Chen J, Song Z et al (2017) Anticancer activity of polysaccharide from *glehnia littoralis* on human lung cancer cell line A549. *Int J Biol Macromol* 106:464–472
- Wu Q, Yang BX (2010) Observation on effect of combined use of fucoidan polysaccharide sulfate and benazepril for treatment of early diabetic nephropathy. *Clin J Med Off* 38(5):743–745

- Xia L, Liu X, Guo H et al (2012) Partial characterization and immunomodulatory activity of polysaccharides from the stem of *dendrobium officinale* (Tiepishihu) in vitro. *J Funct Foods* 4 (1):294–301
- Xiao C, Wu Q, Zhang J et al (2016) Antidiabetic activity of *ganoderma lucidum* polysaccharides F31 down-regulated hepatic glucose regulatory enzymes in diabetic mice. *J Ethnopharmacol* 196:47–57
- Xie JH, Jin ML, Morris GA et al (2016) Advances on bioactive polysaccharides from medicinal plants. *Crit Rev Food Sci* 60:S60–S84
- Xu J (2012) Effects of *astragalus* polysaccharide combined with Megestrol on life quality improvement of patients with advanced malignant tumor. *Chin Trad Herb Drug* 43 (7):1385–1386
- Xu JD (2015) Effect of *ginseng* polysaccharide injection combined with chemotherapy on immune function and therapeutic of the patients with advanced gastric cancer. *Cent South Pharm* 13 (3):316–321
- Xue H, Gan F, Zhang Z et al (2015) *Astragalus* polysaccharides inhibits pcv2 replication by inhibiting oxidative stress and blocking NF- $\kappa$ B pathway. *Int J Biol Macromol* 81:22–30
- Ya GW (2017) A *Lentinus edodes* polysaccharide induces mitochondrial-mediated apoptosis in human cervical carcinoma HeLa cells. *Int J Biol Macromol* 103:676–682
- Yamamoto S, Takatori K, Ohmoto K et al (1989) NK activity and T cell subsets in percutaneous ethanol injection therapy of liver cancer-effect of lentinan with combined use. *Gan To Kagaku Ryoho* 16(9):3291–3294
- Yan JK, Li L, Wang ZM et al (2009) Acidic degradation and enhanced antioxidant activities of exopolysaccharides from *cordyceps sinensis* mycelial culture. *Food Chem* 117(4):641–646
- Yang B, Xiao B, Sun T (2013) Antitumor and immunomodulatory activity of *astragalus membranaceus* polysaccharides in H22 tumor-bearing mice. *Int J Biol Macromol* 62 (11):287–290
- Yang R, Zhang Z, Pei X et al (2009) Immunomodulatory effects of marine oligopeptide preparation from chum Salmon (*Oncorhynchus keta*) in mice. *Food Chem* 113(2):464–470
- Yang Y, Xing R, Liu S et al (2018) Immunostimulatory effects of sulfated chitosans on RAW 264.7 mouse macrophages via the activation of PI3K/Akt signaling pathway. *Int J Biol Macromol* 108:1310–1321
- Yang Y, Zhang MW, Liao ST et al (2012) Structural features and immunomodulatory activities of polysaccharides of *longan* pulp. *Carbohydr Polym* 87:636–643
- Yang Z, Li S, Zhang X et al (2010) Capsular and slime-polysaccharide production by *lactobacillus rhamnosus* jaas8 isolated from chinese sauerkraut: potential application in fermented milk products. *J Biosci Bioeng* 110(1):53–57
- Yang Z, Wu F, Yang H et al (2017) Endocytosis mechanism of a novel proteoglycan, extracted from *ganoderma lucidum*, in HepG2 cells. *RSC Adv* 7(66):41779–41786
- Ye H, Wang K, Zhou C et al (2008) Purification, antitumor and antioxidant activities in vitro of polysaccharides from the brown seaweed *sargassum pallidum*. *Food Chem* 111(2):428–432
- Yeo IC, Lee NK, Hahm YT (2012) Genome sequencing of *Bacillus subtilis* SC-8, antagonistic to the *Bacillus cereus* group, isolated from traditional Korean fermented-soybean. *Food J Bacteriol* 194:536–537
- Yi W, Zhang P, Hou J et al (2018) Enhanced response of tamoxifen toward the cancer cells using a combination of chemotherapy and photothermal ablation induced by lentinan-functionalized multi-walled carbon nanotubes. *Int J Biol Macromol* 120:1525–1532
- Yi Y, Zhang MW, Liao ST et al (2012) Structural features and immunomodulatory activities of polysaccharides of *longan* pulp. *Carbohydr Polym* 87(1):636–643
- Yin JY, Lin HX, Nie SP et al (2012) Methylation and 2d nmr analysis of arabinoxylan from the seeds of *plantago asiatica*. *Carbohydr Polym* 88(4):1395–1401
- Yu J, Sun R, Zhao Z et al (2014) *Auricularia polytricha* polysaccharides induce cell cycle arrest and apoptosis in human lung cancer A549 cells. *Int J Biol Macromol* 68:67–71

- Yu L, Yue H, Liu Y et al (2012) Effect of red *ginseng* polysaccharide on bread quality. *Sci Technol Food Ind* 24:332–338
- Yu Q, Nie SP, Li WJ et al (2013) Macrophage immunomodulatory activity of a purified polysaccharide isolated from *ganoderma atrum*. *Phytother Res* 27(2):186–191
- Yu W, Ren Z, Zhang X et al (2018) Structural characterization of polysaccharides from *dendrobium officinale* and their effects on apoptosis of HeLa cell line. *Molecules* 23(10):2484
- Zhai QL, Hu XD, Xiao J et al (2018) *Astragalus* polysaccharide may increase sensitivity of cervical cancer HeLa cells to cisplatin by regulating cell autophagy. *China J Chin Mat Med* 43(4):805–812
- Zhang D, Meng H, Yang HS (2012) Antidiabetic activity of *taxus cuspidata* polysaccharides in streptozotocin-induced diabetic mice. *Int J Biol Macromol* 50(3):720–724
- Zhang H, Cui SW, Nie SP et al (2016a) Identification of pivotal components on the antioxidant activity of polysaccharide extract from *ganoderma atrum*. *Bioact Carbohydr Diet Fib* 7(2):9–18
- Zhang L, Chen L, Xu X et al (2005) Comparison on chain stiffness of a water-insoluble (1→3)- $\alpha$ -d-glucan isolated from *Poria cocos* mycelia and its sulfated derivative. *Carbohydr Polym* 59:257–263
- Zhang WQ, Huang YS, Zhi XX (2008) Application of hyaluronic acid to clinical medicine. *J Clin Rehabil Tissue Eng Res* 12(23):4515–4518
- Zhang XR, Zhou WX, Zhang YX et al (2011) Macrophages, rather than t and b cells are principal immunostimulatory target cells of *lycium barbarum L.* polysaccharide LBPF4-OL. *J Ethnopharmacol* 136(3):465–472
- Zhang XR, Qi CH, Cheng JP et al (2014) *Lycium barbarum* polysaccharide LBPF4-OL may be a new toll-like receptor 4/MD2-MAPK signaling pathway activator and inducer. *Int Immunopharmacol* 19(1):132–141
- Zhang Y, Hu T, Zhou HL et al (2016b) Antidiabetic effect of polysaccharides from *pleurotus ostreatus* in streptozotocin-induced diabetic rats. *Int J Biol Macromol* 83:126–132
- Zhang Y, Wang L, Du MN (2016c) Research progress on treatment of tumor with *Astragalus* polysaccharides for injection. *Drug Eval Res* 39(6):1092–1094
- Zhao L, Xiao Y, Xiao N (2013) Effect of lentinan combined with docetaxel and cisplatin on the proliferation and apoptosis of BGC823 cells. *Tumor Biol* 34(3):1531–1536
- Zhao LY, Lan QJ, Huang ZC et al (2011) Antidiabetic effect of a newly identified component of *opuntia dillenii* polysaccharides. *Phytomedicine* 18(8–9):661–668
- Zhao R, Zhang T, Ma B et al (2016) Antitumor activity of *Portulaca Oleracea L.* polysaccharide on HeLa cells through inducing TLR4/NF- $\kappa$ B signaling. *Nutr Cancer* 69(1):131–139
- Zhao YM, Yang JM, Liu YH et al (2017) Ultrasound assisted extraction of polysaccharides from *lentinus edodes* and its anti-hepatitis B activity in vitro. *Int J Biol Macromol* 107:2217–2223
- Zhou X, Wang D, Sun P et al (2007) Effects of soluble tea polysaccharides on hyperglycemia in alloxan-diabetic mice. *J Agric Food Chem* 55(14):5523–5528
- Zhu BY, He H, Hou T (2019) A comprehensive review of corn protein-derived bioactive peptides: production, characterization, bioactivities, and transport pathways. *Compr Rev Food Sci Food Saf* 18:329–345
- Zhu K, Nie S, Li C et al (2013) A newly identified polysaccharide from *ganoderma atrum* attenuates hyperglycemia and hyperlipidemia. *Int J Biol Macromol* 57:142–150
- Zong A, Cao H, Wang F (2012) Anticancer polysaccharides from natural resources: a review of recent research. *Carbohydr Polym* 90:1395–1410
- Zou Y, Meng J, Chen W et al (2011) Modulation of phenotypic and functional maturation of murine dendritic cells (DCS) by purified *achyranthes bidentata* polysaccharide (ABP). *Int Immunopharmacol* 11(8):1103–1108