# **Brassicaceae Plants Response and Tolerance to Metal/Metalloid Toxicity**



**Shyamashree Roy and Sanchita Mondal**

**Abstract** Brassicaceae is an important Family having numerous numbers of genera and species within it. Some of these species are important food crops specifically oilseed and vegetables, some are weeds also. They have significant role in accumulation of heavy metals like cadmium, arsenic, lead, caesium, uranium, zinc, etc. in their tissues and have the potential genes within them which attribute tolerance/resistance against the ill effect of these metals. These plants naturally clean and ameliorate the soil contaminated with metal/metalloids. Under Brassicaceae family, members like *Brassica juncea, Brassica oleracea, Brassica napus, Brassica carinata* and many more are known for their phytoremediation property. Like, the seedlings of *B. oleracea, B. juncea* and *Raphanus sativus* are found to uptake arsenic by their root and shoot both. Also, some of the species are known for huge accumulation of heavy metals in tissues which may be a source of potential new genes widening the future research work on phytoremediation. Therefore keeping the worlds' increasing population in mind many of them can be well adapted for growing commercially in heavy metal contaminated soil to contribute to the food basket of the world. The goal of this article is to review the use of species under Brassicaceae family in phytoremediation.

**Keywords** Heavy metals · Brassica crops · Oilseeds · Phytoextraction · Phytoremediation

## **Abbreviations**

Cd Cadmium Cr Chromium

S. Roy  $(\boxtimes)$ 

Regional Research Station, Old Alluvial Zone, Uttar Banga Krishi Viswavidyalaya, Majhian, Dakshin Dinajpur, West Bengal, India e-mail: [shree.agr@gmail.com](mailto:shree.agr@gmail.com)

S. Mondal

Department of Agronomy, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, India

© Springer Nature Singapore Pte Ltd. 2020

M. Hasanuzzaman (ed.), *The Plant Family Brassicaceae*, [https://doi.org/10.1007/978-981-15-6345-4\\_12](https://doi.org/10.1007/978-981-15-6345-4_12)



## **1 Introduction**

Brassicaceae is an important plant family contains genus Brassica. This genus is also important from agriculture point of view because it contains 39 species. It ranks among world's first ten most important plant families with high economic value of the world (Warwick et al. [2000\)](#page-13-0). There are 375 genera and 3200 species under the umbrella Brassicaceae. It is also called as mustard family. Also there are a number of cultivated and hybrids of cultivated species. The species with their high dietary fibre has possessed a valuable place in the list of food crops with their large variation of economic parts starting from oil bearing seeds, stems, buds, flowers, roots and leaves which are used as vegetables, leafy greens, salad, spices, oils, etc. (Hasanuzzaman [2008\)](#page-12-0). Within genus Brassica, *Brassica juncea, Brassica carinata, Brassica rapa, Brassica campestris* and *Brassica napus*—these six species are altogether called as Rapeseed-mustard group. Rapeseed-Mustard being the main oilseed crop is planted on more than 80% area covered under oilseeds. Their seed bear oil and contributes 35% of total oilseeds crops in India (Table [1\)](#page-6-0) (Nmoop [2018\)](#page-13-1). According to FAOSTAT [\(2013\)](#page-12-1) Brassica oilseeds occupies more than 34 million ha land annually over the world. In 1935, UN has given the relationship between these six species in a form of a triangle, designated as U's triangle (Fig. [1\)](#page-2-0).

Also some very common vegetables like cabbage, cauliflower, broccoli, Brussels sprout, kohlrabi, collards and kale come under *Brassica oleraceae* species. They are known for high nutritional value. The other vegetable Brassicaceae includes *B. napus, B. rapa*, *Raphanus sativus, Lepidium sativum* and *Nasturtium officinale*. The condiment crops include *B. juncea*, *Sinapis alba*, *Brassica nigra*, *Armoracia rusticana* and *Eutrena japonica*. Still there are numbers of crops which are being used as potherbs and salads. Among a number of weedy species, *Sinapis arvensis, Raphanus raphanistrum*, *B. rapa* and *Hirschfeldia incana* are of greatest interest with regard to cross-pollination with *B. napus*. Among wild relatives of Brassica, species like *Begonia. adpressa, Brassica fruticulosa, Brassica pinescens, Brassica oxyrrhina, Brassica barrelieri and Brassica tournefortii* comes under the group *Brassica coenospecies*. They are known for their good agronomic characteristics and used

<span id="page-2-0"></span>

widely in hybridization programme (Warwick [1993\)](#page-13-3). The genus *Brassica* is classified as follows:

Order Brassicales (= Cruciales)

Family Brassicaceae (= Cruciferae)

Tribe Brassiceae

Subtribe Brassicinae

Genus *Brassica* L.

Under Brassicaceae, there are 25 tribes with a supplementary 5 currently under study (Al-Shehbaz et al. [2006\)](#page-11-0). Brassiceae contains the genus *Brassica* and wild relatives has 48 genera and near about 240 species (Table [2\)](#page-7-0) (Warwick and Hall [2009\)](#page-13-4). As the population is increasing, it is providing pace to the industrialization also; and the whole, leads to the exposure of the toxic matters as waste materials directly or indirectly to the environment. All the major parts of the environment air, soil and water are contaminated highly by these toxics to such extent that it has become a life threat to humankind (Zhuang et al. [2007\)](#page-14-0), as well as for animal and plant kingdom (Singh et al. [2011;](#page-13-5) Hasanuzzaman et al. [2012;](#page-12-2) Hasanuzzaman and Fujita [2012\)](#page-12-3). These include metal, metalloids, radio nucleotides and different kinds of environmental contaminants. Human activities related to industries and agricultural practices expose the environment to various heavy metals and metalloids like Molybdenum, Cadmium, Arsenic, Nickel, Cobalt, etc. (Pilon-Smits [2005\)](#page-13-6). There is very less or narrow chance to find out a remedy (Cunningham et al. [1997;](#page-11-1) Prasad

and Freitas [2003\)](#page-13-7). Though there are policies and routine precautions and a big list of Do's and DON'Ts, which is not enough to control the heavy metal pollution of the earth (Zayad and Tery [2003\)](#page-13-8). There is a huge pressure of providing enough food to supply a square meal to this burgeoning population of the world. Horizontal expansion of cultivable land is not possible until the degraded, polluted and waste fallows are being converted to cultivated one. But these contaminated and marginal lands are always a threat to food production with a huge chance of food contamination when used for cultivation (He et al. [2005\)](#page-12-4). But nature has always an answer to every situation. Thangahu et al. [\(2011\)](#page-13-9) reported about development of different techniques to judge and observe the status and movement of metals inside the soil, water and wastewater. Again some plants/adopted a technique to extract, sequester and detoxify the toxic elements. The plant is known as accumulator plant. The technique is termed as Phytoremediation (Fig. [2\)](#page-3-0) where the Greek originated prefix *Phyto* means plant and latin word remedium means to correct (Erakhrumen and Agbontalor [2007;](#page-11-2) Tangahu et al. [2011\)](#page-13-9). From the observation of (USEPA [2000;](#page-13-10) Sarma [2011](#page-13-11) and Tangahu et al. [2011\)](#page-13-9), it can be stated that phytoremediation is comparatively cheap and ecofriendly method to get rid of the toxic materials from soil, water or air. Some specific plants are known to successfully engross contaminants include metals and metalloids responsible for environmental contamination. Phytoextraction being the significant process, used to eliminate heavy metals from soil utilizing its ability to take up the essential nutrients which are contaminants too under specific conditions (Fe, Mn, Zn, Cu, Mg, Mo, and Ni). Besides, other metals with unidentified function (Cd, Cr, Pb, Co, Ag, Se, Hg) can also be stored (Cho-Ruk et al. [2006\)](#page-11-3). Among angiosperms, Brassicaceae is the most important family having the huge number

<span id="page-3-0"></span>

of (25%) metal accumulating species (90) (Prasad and Freitas [2003;](#page-13-7) Palmer et al. [2001a,](#page-13-12) [b;](#page-13-13) Krämer [2010;](#page-12-5) Sarma [2011\)](#page-13-11). Some member of the family Brassicaceae can accumulate metals even the toxic ones in relatively high amount (Kumar et al. [1995](#page-12-6) and Anjumet al. [2012\)](#page-11-4). They are potential phytoextraction plants (Van Ginneken et al. [2007;](#page-13-14) Gall and Rajkaruna [2013\)](#page-12-7) who show visible symptoms of metal accumulation and being the food crops they contaminate the food chain (Gall et al. [2015\)](#page-12-8). Brassicas accumulate heavy metals in their stem due to the inherent tolerance to metals present in excess. Tangahu et al. [\(2011\)](#page-13-9) observed that some of the member of Brassica genus (*B. campestris* L.*, B. carinata* A. Br.*, B. juncea* (L.) *Czern*) can accumulate lead in their tissue more than 50 mg g−<sup>1</sup> dry weight of plant whereas *B. nigra* Koch. can gather more than 100 mg g<sup>-1</sup> dry weight. From the same study we came to know that in case of mercury, *B. juncea* accumulated more than 1 mg  $g^{-1}$ dry weight compared to other species (<0.2 mg/g dry weight). Therefore they may be used as tool for phytoextraction (Neison and Rajkaruna [2012\)](#page-12-9). Indian mustard or *B. juncea*, an important oil contributor in India has been reported to have a reduced oil percentage in seed due to accumulation of Cadmium (Ahmad et al. [2015;](#page-11-5) Mourato et al[.2015\)](#page-12-10). Not only Cadmium, according to Alkorta et al. [\(2004\)](#page-11-6) and Szczyglowska et al. [\(2011\)](#page-13-15), *B. juncea* has a superior capacity of different toxic metal accumulation. *Brassica napus* also known as rapeseed is the chief source of eatable oil in the world can accumulate Zn in its plant biomass (Grispenet al. [2006;](#page-12-11) Carrier et al. [2003\)](#page-11-7). *Brassica campestris* also known as turnip rape, also an oilseed can also accumulate metals in its below ground parts (Gleba et al. [1999\)](#page-12-12). It is considered for the phytoremediation of Chromium which can be accumulated in huge quantity by this species (Dheri et al. [2007\)](#page-11-8). Kabata-Pendias [\(2001\)](#page-12-13), from a study confirmed that *B. oleracea* accumulated lead (Pb) in good quantity. Kumar et al. [\(1995\)](#page-12-6) observed that roots are more efficient in Pb accumulation than above ground parts. Gisbert found that not only Pb, it also accumulate Zn and Cu in its shoot. Addae [\(2010\)](#page-11-9) observed in a study that *B. oleracea* species can extract significant amount of Cd and Pb. All these research work has been conducted with a focus in the matter to find out a plant which will be pretty good to be used for phytoremediation with the characters like, more accumulation efficiency in shoot, tolerance capacity in soils with high amount of metals, fast growing and easy to grow and harvest (Table [3\)](#page-8-0) (Marchiol et al. [2004\)](#page-12-14). Some of the best studies citing the role of Brassicaceae in tolerance to metals and metalloids are presented hereunder for better understanding.

## **2 Amelioration of Cadmium (Cd) Toxicity by** *Coronopus didymus*

*Coronopus didymus,* a native of South America is an annual herb of family Brassicaceae and widely distributed across the world (Yannitsaros [1986\)](#page-13-16). In Northern India, it is being grown along the road sides and gardens specifically during October to February. Sidhu et al. [\(2017\)](#page-13-17) worked with *C. didymus* to assess the development

and survival of *C. didymus* under varied Cd treatments and also on the tolerance and extraction efficiency of C*. didymus* under various levels of Cd concentrations. They reported that under 400 mg kg<sup>-1</sup> Cd, there was an increase in Cd content in roots and shoots (867.2 and 864.5 mg kg−<sup>1</sup> DW), reflecting the potential of this herb *C. didymus* in ameliorating the toxicity effect of Cd in soils. The possible explanation for this tolerance is the compartmentalization of metals in vacuoles that restricted the excess Cd transport in the plants. Besides, Phytochelatins has a significant role in accumulating Cd within roots (Zeng et al. [2009\)](#page-14-1). Toxicity of Cd induces stress stimulates the generation and build-up of Reactive Oxygen Species (Devi et al. [2007\)](#page-11-10). Enzymes like super oxide dismutase, catalase and peroxidase play a crucial role in quenching Reactive Oxygen Species (ROS) produced as because of heavy metal stress (Zhang et al. [2013\)](#page-14-2). Due to the antioxidative responses, *C. didymus* could be one of the possible options to be used in Cd contaminated soils with no risk to human or animals as it is unpalatable and cannot enter the food chain.

#### **3 Response of** *Hirschfeldia incana* **to Lead (Pb)**

Lead (Pb) is one of the toxic elements having ill effect on the quality of environment as well as human health (Lee et al. [2005\)](#page-12-15). Although it is not essential for plants, but still imposes a toxicity threat to plants and animals. Auguy et al. [\(2013\)](#page-11-11) identified *Hirschfeldia incana*, a member of the Brassicaceae family, with a high potential to accumulate lead in their biomass. *Hirschfeldia incana* was formerly known as *Brassica geniculata* is a species of flowering plant in the mustard family with common names as shortpod mustard, buchan weed, hoary mustard and Mediterranean mustard. In their study, chosen area was heavily affected by lead mining contamination with a concentration range of 26–9479 mg kg−<sup>1</sup> (Smouni et al. [2010\)](#page-13-18). The Brassicaceae *H. incana* was found to be important for its ability to accumulate toxic heavy metals in leaves. *H. incana* was able to accumulate 106 mg Pb  $g^{-1}$  DW in roots at their vegetative growth and 77 mg Pb  $g^{-1}$  DW in roots at the floral stage after 60 days of lead exposure. In one of the study, Kaur [\(2018\)](#page-12-16) identified the ability of Indian mustard with and without chelants like EDTA and salicylic acid. Pb concentrations increased in root and shoot organs of the plant with EDTA treatment, but it decreased with salicylic acid treatment. The results also revealed that *B. juncea var. arawali* accumulated high amount of Pb in roots with low translocation in the parts of shoots; nevertheless it could be used for phytoremediation of Pb (Tables [1](#page-6-0) and [2\)](#page-7-0).

#### **4 Arsenic Tolerance by Brassicaceae Seedlings**

Arsenic is one of the potential toxic elements with a high risk of entering food chains and one report by Bhattacharya et al. [\(2010\)](#page-11-12) emphasize that 93% of the total arsenic are being consumed by humans. This consumption is always accompanied

| Year        | Groundnut | Rapeseed-mustard | Soybean | Sunflower |
|-------------|-----------|------------------|---------|-----------|
| 1950-51     | 3.48      | 0.76             | -       | -         |
| 1960-61     | 4.81      | 1.35             |         |           |
| 1970-71     | 6.11      | 1.98             | 0.01    | 0.08      |
| 1980-81     | 5.01      | 2.30             | 0.44    | 0.07      |
| 1990-91     | 7.51      | 5.23             | 2.60    | 0.87      |
| $2000 - 01$ | 6.41      | 4.19             | 5.28    | 0.65      |
| $2010 - 11$ | 8.26      | 8.18             | 12.74   | 0.65      |
| $2011 - 12$ | 6.96      | 6.60             | 12.21   | 0.52      |
| $2012 - 13$ | 4.70      | 8.03             | 14.67   | 0.54      |
| $2013 - 14$ | 9.71      | 7.88             | 11.86   | 0.43      |
| $2014 - 15$ | 7.40      | 6.28             | 10.37   | 0.43      |
| $2015 - 16$ | 6.73      | 6.80             | 8.57    | 0.30      |
| $2016 - 17$ | 7.46      | 7.92             | 13.16   | 0.25      |
| $2017 - 18$ | 9.25      | 8.43             | 10.98   | 0.22      |
| $2018 - 19$ | 6.52      | 8.78             | 13.74   | 0.20      |

<span id="page-6-0"></span>**Table 1** Year-wise production of major oilseeds in India (in Million tonnes)

*Source* Modified from Directorate of Economics and Statistics

Third Advance Estimates of Production of Commercial Crops for 2018–19

with a health hazard to human population as it is considered one of the most unsafe ingredients to humans (Khan et al. [2009\)](#page-12-17). Numerous species of Brassicaceae are designated as arsenic tolerant to metals (Srivastava et al. [2009\)](#page-13-19). Freitas-Silva et al. [\(2016\)](#page-12-18) showed that even Brassicaceae seedlings at initial growing conditions can accumulate arsenic in their biomass (Table [4\)](#page-9-0). Roots accumulated high arsenic when compared to shoots suggesting little movement of these metalloids to the shoots.

#### **5 Hyperaccumulation of Selenium by Brassicaceae Crops**

Seleniferous soils are those with high levels of selenium and there are some Brassicaceae species that can accumulate high levels of Se within their biomass (Brown and Shrift [1981\)](#page-11-13). Plants can normally accumulate around 0.05–1 mg kg<sup>-1</sup> Se dry weight, but there are hyperaccumulators too which can engross higher concentrations and occasionally hundreds of times more than the usual range of Se accumulated by normal plants. Hladun et al.  $(2011)$  studied the accumulation of Se in the plant parts of *Stanleya pinnata* and *B. Juncea*. *Stanleya pinnata was* found to be the efficient accumulator of selenium which has a potential to accumulate 22% (Nectar) and 85% (pollen) excess over *B. juncea*. *Stanleya pinnata* is a species under mustard family and also known as desert prince's plume and is a native to North America. The mechanism behind the accumulation of Se in *S. pinnata* is the mobilization of



(continued)

<span id="page-7-0"></span>**Table 2** Cultivated *Brassica* species and related genera

#### Table 2 (continued)



Modified from OECD [\(2016\)](#page-13-20)

<span id="page-8-0"></span>**Table 3** Heavy metal accumulation by *Brassica* and related genera



Modified from Dar et al. [\(2015\)](#page-11-14) and Pantola and Alam [\(2014\)](#page-13-21)



<span id="page-9-0"></span>**Table 4**  $(\mu g g^{-1})$ dry matt  $12$ -days (Freitas-

the element in the leaves into the reproductive organs of the plants; some reports are also there relating the volatilization of Se from leaves directly into the atmosphere. Accumulation of selenium by *S. pinnata* in the reproductive parts (flowers and seeds) is also being confirmed in a number of field studies with corresponding decrease in the leaf system (Galeas et al. [2007\)](#page-12-20). Although *B. juncea* can also accumulate selenium but they basically prefer soils with moderate levels of Se; on the other hand, *Stanleya pinnata* can take up high amount of selenium even under soils with low amount of selenium (Terry et al. [2000\)](#page-13-22).

## **6 Zinc (Zinc) Tolerance of** *Arabidopsis halleri*

*Arabidopsis halleri* (syn: *Cardaminopsis halleri*) can tolerate higher concentrations of heavy metals (Brooks [1998\)](#page-11-15). Zinc tolerance was investigated in five populations of *A. halleri* (syn.: *C. halleri*). All these were raised from the seeds collected from the even sites, i.e. both contaminated and uncontaminated. Irrespective of the origin, *A. halleri* seedlings showed a continuous growth of roots at 100  $\mu$ M Zn, with no toxic symptoms (dry or chlorotic leaves) in the plants. In contrast to *A. halleri*, roots of other species like *Arabidopsis thaliana* and *Arabidopsis lyrata* subsp. *petraea* seedlings were completely repressed at 100  $\mu$ MZn concentration (Bert et al. [2000\)](#page-11-16). This study was the first to demonstrate the ability of *A. halleri* to tolerate high level of zinc in the soil. Even, uncontaminated site seedlings showed a tolerance suggested a constitutive trait in *A. halleri*.

<span id="page-10-0"></span>

Adopted from Drozdova et al. [\(2017\)](#page-11-17)

#### **7 Ni Tolerance of Brassicaceae Species**

With an idea to identify potential accumulators, Drozdova et al. [\(2017\)](#page-11-17) conducted a comparative study and reported the outcome of higher doses of Nickel on few wild growing plant species of Brassicaceae. Results of the experiments revealed distinct differences in the species in terms of tolerance to Ni. There were clear symptoms like chlorosis and necrosis on the leaves of species which are non-hyperaccumulators. After 11 days of Ni exposure, about 80% of plants *Alyssum alyssoides* and *Alyssum campestre* and 50% *Erysimum ibericum* started showing the toxicity symptoms. Surprisingly, *Alyssum murale* developed normally even in the same concentrations of Ni applied for *A. alyssoides*, *A. campestre*a nd *E. ibericum*. A comparative study of the accumulation of Ni is shown in Table [5.](#page-10-0) The accumulation of Ni was highest in the roots of *A. murale* suggesting its high efficiency of extracting Ni ions from the soil solution, even in areas with low Ni content. In terms of tolerance to high concentrations of Ni, the species could be arranged in the order: A. murale  $\geq E$ . *ibericum* > *A. alyssoides* > *A. campestre*.

### **8 Conclusion**

Across many studies, it has been confirmed that the family Brassicaceae is remarkably vital for remediation of heavy metals and metalloids across the world. Brassicaceae comprises a number of species with better accumulation efficiency signifying a great tolerance level to metals. Although a number of species are identified with proper scientific theories on the mechanisms leading to the tolerance or hyperaccumulation; but their genetic level study linking with their metal tolerance ability needs to be explored for better understanding.

## **References**

- <span id="page-11-9"></span>Addae C, Piva M, Bednar AJ, Zaman MS (2010) Cadmium and lead bioaccumulation in cabbage plants grown in metal contaminated soils. AdvSciTechnol 4:79–82
- <span id="page-11-5"></span>Ahmad P, Sarwat M, Bhat NA, Wani MR, Kazi AG, Tran LSP (2015) Alleviation of cadmium toxicity in *Brassica juncea*L. (czern. & coss.) by calcium application involves various physiological and biochemical strategies. PLoS ONE10e0114571
- <span id="page-11-6"></span>Alkorta I, Hernandez-Allica J, Becerril JM, Amezaga I, Albizu I, Garbisu C (2004) Recent findings on the phytoremediation of soils contaminated with environmentally toxic heavy metals and metalloids such as zinc, cadmium, lead, and arsenic. Rev Environ Sci Biotechnol 3:71–90
- <span id="page-11-4"></span>Anjum NA, Ahmad I, Pereira ME, Duarte AC, Umar S, Khan NA (2012) The plant family Brassicaceae: an introduction. In: Anjum NA, Ahmad I, Pereira ME, Duarte AC, Umar S, Khan NA (eds) The plant family Brassicaceae: contribution towards phytoremediation, Environmental pollution series no. 21. Springer, Dordrecht
- <span id="page-11-11"></span>Auguy F, Fahr M, Moulin P, Brugel A, Laplaze L, Mzibri ME, Filali-Maltouf A, Doumas P, Smouni A (2013) Lead tolerance and accumulation in Hirschfeldia incana, a Mediterranean Brassicaceae from metalliferous mine spoils. PLoS ONE 8(5):e61932
- <span id="page-11-0"></span>Al-Shehbaz LA, Beilstein, MA, Kellogg EA (2006) Systematics and phylogeny of the Brassicaceae (Cruciferae): an overview. Plant Syst Evol 259(2):89–120
- <span id="page-11-16"></span>Bert V, Macnair MR, De-Laguerie P, Saumitou-Laprade P, Petit D (2000) Zinc tolerance and accumulation in metallicolous and nonmetallicolous populations of *Arabidopsis helleri* (Brassicaceae). New Phytologists 146:225–233
- <span id="page-11-12"></span>Bhattacharya P, Samal AC, Majumdar J (2010) Arsenic contamination in rice, wheat, pulses, and vegetables: a study in an arsenic affected area of West Bengal, India. Water Air Soil Pollut 21:3–13
- <span id="page-11-15"></span>Brooks RR (1998) Plants that hyperaccumulate heavy metals. Wallingford, UK: CAB International
- <span id="page-11-13"></span>Brown TA, Shrift A (1981) Exclusion of selenium from proteins of selenium tolerant *Astragalus* species. Plant Physiol 67:1051–1053
- <span id="page-11-7"></span>Carrier P, Baryla A, Havaux M (2003) Cadmium distribution and microlocalization in oilseed rape (*Brassica napus*) after long-term growth on cadmium contaminated soil. Planta 216:939–950
- <span id="page-11-3"></span>Cho-Ruk K, Kurukote J, Supprung P, Vetayasuporn S (2006) Perennial plants in the phytoremediation of lead contaminated soils. Biotechnology 5(1):1–4
- <span id="page-11-1"></span>Cunningham SD, Shann JR, Crowley DE, Anderson TA (1997) Phytoremediation of contaminated water and soil. In: Kruger EL, Anderson TA, Coats JR (eds) Phytoremediation of soil and water contaminants, ACS symposium series 664. American Chemical Society, Washington, DC
- <span id="page-11-14"></span>Dar MI, Khan FA, Rehman F, Masoodi A, Ansari AA, Varshney D, Naushin F, Naikoo MI (2015) Roles of Brassicaceae in phytoremediation of metals and metalloids. In: Ansari A, Gill S, Gill R, Lanza G, Newman L (eds) Phytoremediation. management of environmental contaminants, vol 1. [https://doi.org/10.1007/978-3-319-10395-2\\_14,](https://doi.org/10.1007/978-3-319-10395-2_14) © Springer International Publishing Switzerland 2015. pp 201–215
- <span id="page-11-10"></span>Devi R, Munjral N, Gupta AK, Kaur N (2007) Cadmium induced changes in carbohydrate status and enzymes of carbohydrate metabolism, glycolysis and pentose phosphate pathway in pea. Environ Exp Bot 61:167–174
- <span id="page-11-8"></span>Dheri GS, Brar MS, Malhi SS (2007) Comparative phytoremediation of chromium-contaminated soils by Fenugreek, Spinach, and Raya. Commun Soil Sci Plant Anal 38:1655–1672
- <span id="page-11-17"></span>Drozdova IV, Alekseeva-Popova NV, Kalimova IB, Belyaeva AI, Smirnova NA (2017) The accumulating ability and nickel tolerance of Brassicaceae species of the North Caucasus in connection with the problem of phytoremediation. J Geo chem. Expln 182:235–241
- <span id="page-11-2"></span>Erakhrumen A, Agbontalor A (2007) Phytoremediation: an environmentally sound technology for pollution prevention, control and remediation in developing countries. Educ Res Rev 2:151–156
- <span id="page-12-1"></span>FAOSTAT (2013) FAO Statistics online database, Production/Crops,– rapeseed, Year 2013. Food and Agriculture Organization of the United Nations. <http://faostat3.fao.org/home/E>
- <span id="page-12-18"></span>Freitas-Silva LD, Araujo TOD, Silva LCD, Oliveira JAD, Araujo JMD (2016) Arsenic accumulation in Brassicaceae seedlings and its effect on plant growth and plant anatomy. Eco Toxicol Environ Saf 124:1–9
- <span id="page-12-20"></span>Galeas ML, Zhang LH, Freeman JL, Wegner M, Pilon-Smits EAH (2007) Seasonal fluctuations of selenium and sulfur accumulation in selenium hyperaccumulators and related nonaccumulators. New Phytol 173:517–525
- <span id="page-12-7"></span>Gall JE, Rajakaruna N (2013) The physiology, functional genomics, and applied ecology of heavy metal-tolerant brassicaceae. In: Lang M (ed) Brassicaceae: characterization, functional genomics and health benefits. Nova Science Publishers: Hauppauge, NY, USA, pp 121–148
- <span id="page-12-8"></span>Gall JE, Boyd RS, Rajakaruna N (2015) Transfer of heavy metals through terrestrial food webs: a review. Environ Monit Assess 187:201
- <span id="page-12-12"></span>Gleba D, Borisjuk MV, Borisjuk LG, Kneer R, Poulev A, Skarzhinskaya M (1999) Use of plant roots for phytoremediation and molecular farming. Proc Natl Acad Sci USA 96:5973–5977
- <span id="page-12-11"></span>Grispen VMJ, Nelissen HJM, Verkleij JAC (2006) Phytoextraction with *Brassica napus* L.: a tool for sustainable management of heavy metal contaminated soils. Environ Pollut 144:77–83
- <span id="page-12-0"></span>Hasanuzzaman M (2008) Siliqua and seed development in rapeseed (*Brassica campestris* L.) as affected by different irrigation levels and row spacings. Agric Conspectus Scientificus 73(4):221– 226
- <span id="page-12-3"></span>Hasanuzzaman M, Fujita M (2012) Heavy metals in the environment: current status, toxic effects on plants and possible phytoremediation. In: Anjum NA, Pereira MA, Ahmad I, Duarte AC, Umar S, Khan NA (eds) Phytotechnologies: remediation of environmental contaminants. CRC Press, Boca Raton, pp 7–73
- <span id="page-12-2"></span>Hasanuzzaman M, Hossain MA, Teixeira da Silva JA, Fujita M (2012) Plant Responses and tolerance to abiotic oxidative stress: antioxidant defense is a key factor. In: Bandi V, Shanker AK, Shanker C, Mandapaka M (eds) Crop stress and its management: perspectives and strategies. Springer, New York, pp 261–316
- <span id="page-12-4"></span>He ZL, Yang XE, Stoffella PJ (2005) Trace elements in agroecosystems and impacts on the environment. J Trace Elem Med Biol 19:125–140
- <span id="page-12-19"></span>Hladun KR, Parker DR, Trumble JT (2011) Selenium, accumulation in floral tissues of two Brassicaceae species and its impacts on floral traits and plant performance. Environ Exp Bot 74:90–97
- <span id="page-12-13"></span>Kabata-Pendias A (2001) Trace elements in soils and plants, 3rd edn.CRC, Boca Raton, FL
- <span id="page-12-16"></span>Kaur L (2018) Accumulation potential of Indian mustard (*Brassica juncea* var. *arawali*) and fenugreek (*Trigonellafoenum-graecum* L.) planted on Lead and Nickel contaminated soil. Trop Plant Res 5(2):217–223
- <span id="page-12-17"></span>Khan I, Ahmad A, Iqbal M (2009) Modulation of antioxidant defense system for arsenic detoxification in Indian mustard. Ecotoxicol Environ Saf 72:626–634
- <span id="page-12-5"></span>Krämer U (2010) Metal hyperaccumulation in plants. Annu Rev Plant Biol 61:517–534
- <span id="page-12-6"></span>Kumar PBAN, Dushenkov V, Motto H, Raskin I (1995) Phytoextraction: the use of plants to remove heavy metals from soils. Environ Sci Technol 29:1232–1238
- <span id="page-12-15"></span>Lee M, Lee K, Lee J, Noh EW, Lee Y (2005) AtPDR12 contributes to lead resistance in *Arabidopsis*. Plant Physiol 138:827–836
- <span id="page-12-14"></span>Marchiol L, Assolari S, Sacco P, Zerbi G (2004) Phytoextraction of heavy metals by canola (*Brassica napus*) and radish (*Raphanus sativus*) grown on multi contaminated soil. Environ Pollut 132:21– 27
- <span id="page-12-10"></span>Mourato MP, Moreira IN, Leitão I, Pinto FR, Sales JR, Martins LL (2015) Effect of heavy metals in plants of the *Genus Brassica*. Int J Mol Sci 16:17975–17998
- <span id="page-12-9"></span>Neilson S, Rajakaruna N (2012) Roles of rhizospheric processes and plant physiology in applied phytoremediation of contaminated soils using brassica oilseeds. In: Anjum NA, Ahmad I, Pereira ME, Duarte AC, Umar S, Khan NA (eds) The Plant Family Brassicaceae, vol. 21. Springer Netherlands: Dordrecht, The Netherland, pp 313–330

<span id="page-13-1"></span>NMOOP (2018) Present status of oilseed crops and vegetable oils in India

- <span id="page-13-20"></span>OECD (2016) Brassica crops (*Brassica species*). In: Safety assessment of transgenic organisms in [the environment, vol 5, OECD Consensus Documents. OECD Publishing, Paris.](https://doi.org/10.1787/9789264253018-6-en) https://doi.org/ 10.1787/9789264253018-6-en
- <span id="page-13-12"></span>Palmer CE, Warwick S, Keller W (2001a) Brassicaceae (Cruciferae) family, plant biotechnology, and phytoremediation. Int J Phytorem 3(3):245–287
- <span id="page-13-13"></span>Palmer CE, Warwick S, Keller W (2001b) Brassicaceae (Cruciferae) family, plant biotechnology, and phytoremediation. Int J Phytoremediation 3:245–287
- <span id="page-13-21"></span>Pantola RC, Alam A (2014) Potential of Brassicaceae burnett (Mustard family; Angiosperms) in phytoremediation of heavy metals. Int J Sci Res Environ Sci 2(4):120–138
- <span id="page-13-6"></span>Pilon-Smits, EAH (2005) Phytoremediation. Ann. Rev. Plant Biol. 56:15–39
- <span id="page-13-7"></span>Prasad MNV, Freitas HMO (2003) Metal hyperaccumulation in plants—biodiversity prospecting for phytoremediation technology. Electron J Biotechnol 6:285–321
- <span id="page-13-11"></span>Sarma H (2011) Metal hyperaccumulation in plants: a review focusing on phytoremediation technology. Environ Sci Technol 4:118–138
- <span id="page-13-17"></span>Sidhu GPS, Singh HP, Batish DR, Kohli RK (2017) Tolerance and hyperaccumulation of cadmium by a wild, unpalatable herb *Coronopus didymus* (L.) Sm. (Brassicaceae). Eco toxicol Environ Saf 135:209–215
- <span id="page-13-5"></span>Singh J, Kalamdhad Ajay S (2011) Effects of heavy metals on soil, plants, human health and aquatic life. Int J Res Chem Environ 1(2):15–21
- <span id="page-13-18"></span>Smouni A, Ater M, Auguy F, Laplaze L, El Mzibri M (2010) Assessment of contamination by metallic trace elements in a mining area of eastern Morocco. Cah Agric 19:26–76
- <span id="page-13-19"></span>Srivastava S, Srivastava AK, Suprasanna P, D´Souza SF (2009) Comparative biochemical and transcriptional profiling of two contrasting varieties of *Brassica juncea* L. in response to arsenic exposure reveals mechanisms of stress perception and tolerance. J Exp Bot 60:3419–3431
- <span id="page-13-15"></span>Szczyglowska M, Piekarska A, Konieczka P, Namiesnik J (2011) Use of Brassica plants in the phytoremediation and biofumigation processes. Int J Mol Sci 12:7760–7771
- <span id="page-13-9"></span>Tangahu BV, Abdullah SRS, Basri H, Idris M, Anuar N, Mukhlisin M (2011) A review on heavy [metals \(As, Pb, and Hg\) uptake by plants through phytoremediation. Int J Chem Eng.](https://doi.org/10.1155/2011/939161) https://doi. org/10.1155/2011/939161
- <span id="page-13-22"></span>Terry N, Zayed AM, de Souza MP, Tarun AS (2000) Selenium in higher plants. Annu Rev Plant Physiol Plant MolBiol 51:401–432
- <span id="page-13-2"></span>UN (1935) Genomic analysis in Brassica with special reference to the experimental formation of Brassica napus and peculiar mode of fertilization. Jpn J Bot 7:389–452
- <span id="page-13-10"></span>USEPA (2000) Introduction to phytoremediation. EPA, Washington, DC
- <span id="page-13-14"></span>Van Ginneken L, Meers E, Guisson R, Ruttens A, Elst K, Tack FMG, Vangronsveld J, Diels L, Dejonghe W (2007) Phytoremediation for heavy metal-contaminated soils combined with bioenergy production. J Environ Eng Land Sc Manag 15:227–236
- <span id="page-13-3"></span>Warwick SI (1993) Guide to wild germplasm of Brassica and allied crops Part IV: wild species in the tribe Brassicaceae (cruciferae) as sources of agronomic trait. Technical Bulletin 17E-1993. Centre for Land and Biological Resources Research, Agriculture Canada, Ottawa, Ontaria
- <span id="page-13-4"></span>Warwick SI, Hall JC (2009) Phylogeny of *Brassica* and wild relatives. In: Gupta SK (ed) Biology and breeding of Crucifers. CRC Press, Boca Raton, Florida, pp 19–36
- <span id="page-13-0"></span>Warwick SI, Francis A, Lafleche J (2000) Guide to Wild Germplasm of Brassica and allied crops (*Tribe Brassiceae*[, Brassicaceae\), 2nd edn. Agriculture and Agri-food Canada.](http://www.brassica.info/resources/crucifer_genetics/guidewild/htm) http://www.bra ssica.info/resources/crucifer\_genetics/guidewild/htm
- <span id="page-13-16"></span>Yannitsaros A (1986) New data on the naturalization and distribution of *Coronopusdidymus* (Cruciferae) in Greece. Willdenowia 61–64
- <span id="page-13-8"></span>Zayed AM, Terry N (2003) Chromium in the environment: factors affecting biological remediation. Plant Soil 249:139–156
- <span id="page-14-1"></span>Zeng X, Ma LQ, Qiu R, Tang Y (2009) Responses of non-protein thiols to Cd exposure in Cd hyperaccumulator *Arabispaniculata* Franch. Environ Exp Bot 66:242–248
- <span id="page-14-2"></span>Zhang S, Lin H, Deng L, Gong G, Jia Y, Xu X, Li T, Li Y, Chen H (2013) Cadmium tolerance and accumulation characteristics of *Siegesbeckiaorientalis* L. Ecol Eng 51:133–139
- <span id="page-14-0"></span>Zhuang X, Chen J, Shim H, Bai Z (2007) New advances in plant growth-promoting rhizobacteria for bioremediation. Environ Int 33:406–413