

Chapter 10

Hepatitis E Virus



Vasileios Papatsiros

Abstract Hepatitis E virus (HEV) is taxonomically classified within *Hepeviridae* family and *Orthohepevirus* genus. Genotypes HEV-1 and HEV-2 infect human, while genotypes HEV-3 and HEV-4 are zoonotic viruses that infect humans, domestic pigs and other animal species (e.g. wild boar, deer). The main route of trans-species transmission is the direct contact with infected animals, as well as via the consumption of HEV-contaminated food products or via the faecal–oral route through drinking of contaminated water. HEV-3 has been detected in pigs around the world (South and North America, Europe, Africa, Asia, and Oceania). HEV-4 has mainly reported in domestic pigs and humans in Asia. Domestic pigs, wild boar, and various species of deer reported to play important role in zoonotic transmission of HEV-3 and HEV-4 from animals to humans. The most important reservoirs of the HEV genotypes are domestic pigs and the most HEV infections in humans are foodborne due mainly to consumption of undercooked meat or meat products (e.g. sausages). The main route of natural HEV transmission in pigs is via the faecal–oral. However, the HEV infection in pig is usually asymptomatic, with low impact on health status. Future studies focus on preventive measures to eliminate the appearance and persistence of HEV in pig farms (including biosecurity and vaccination) are required. Moreover, more studies are needed to investigate deeply the role of wildlife in the epidemiology of HEV infection.

Keywords Hepatitis E virus · HEV · Pig · Human · Pork · Epidemiology · Pathogenesis

V. Papatsiros (✉)

Clinic of Medicine (Farm Animal Medicine), Faculty of Veterinary Medicine, School of Health Sciences, University of Thessaly, Karditsa, Greece

e-mail: vpapatsiros@vet.uth.gr

10.1 Prologue

Hepatitis E virus (HEV) is the causative agent of hepatitis E and it is classified into family *Hepeviridae*, which is divided in two genera: *Orthohepevirus* and *Piscihepevirus* (Smith et al. 2014). The genus *Piscihepevirus* includes only *Piscihepevirus A* (cutthroat trout virus), while the genus *Orthohepevirus* is divided in four species (Khuroo et al. 2016; Purdy et al. 2017):

- Orthohepevirus A*, including isolates from such humans, domestic pigs, wild boars, deer, mongoose, rabbits, and camels—Fig. 10.1). Moreover, *Orthohepevirus A* has eight genotypes, five members of them are found to infect humans (Johne et al. 2014).
- Orthohepevirus B*, including three avian isolates (HEV-1, HEV-2, and HEV-3),
- Orthohepevirus C*, including isolates from rats, greater bandicoot, Asian musk shrews, mink, and ferrets, and
- Orthohepevirus D*, including isolates from bats

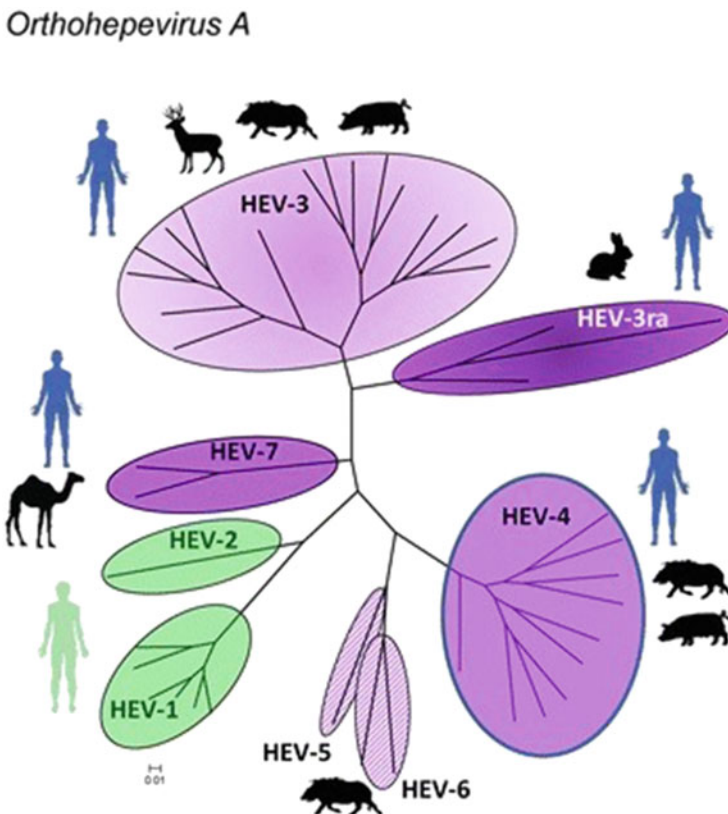


Fig. 10.1 Phylogenetic tree of HEV sequences within the species *Orthohepevirus A* (adopted from Pavio et al. 2017 and Smith et al. 2016)

HEV is a relatively stable virus, surviving in the gastrointestinal tract environment due to its resistance to gastric secretions and bile salts (Emerson and Purcell 2001). It is a small non-enveloped virus (27–33 nm in diameter) and icosahedral shaped sphere with shaped bumps visible on its surface (Balayan 1997). Its genome consists a single-stranded, positive-sense RNA molecule about 7.5 kilobases (kb) in length, which contains three open reading frames (ORF) (Tam et al. 1991). Based on ORF2 nucleotide sequence analysis, four major genotypes (HEV-1, HEV-2, HEV-3, and HEV-4) have been defined in mammals (Schlauder and Mushahwar 2001).

HEV genotypes 1 and 2 (HEV-1, HEV-2) are reported in humans (Kamar et al. 2017), while HEV genotypes 3 and 4 (HEV-3, HEV-4) are zoonotic viruses, infecting both humans and animals. The main route of trans-species transmission is the direct contact with infected animals, as well as via the consumption of HEV-contaminated food products or via the faecal–oral route through drinking of contaminated water (Colson et al. 2010; Dremsek et al. 2012; Chaussade et al. 2013; Riveiro-Barciela et al. 2015; Guillois et al. 2016). In rabbit species a separate genotype HEV-3 (HEV-3ra) was reported, which also includes a closely related human isolate (Pavio et al. 2017). Furthermore, HEV genotypes 5 and 6 (HEV-5, HEV-6) have been reported in wild boars (Takahashi et al. 2011), while HEV genotypes 7 and 8 (HEV-7, HEV-8) were found in camels (Woo et al. 2016; Lee et al. 2016).

Studies reported the isolation of HEV from various wild and domestic animals, such as domestic pigs, cattle, chickens, sheep, goats, and rodents (Favorov et al. 2000; Meng 2000).

10.2 Epidemiology

10.2.1 Geographic Distribution

The geographical distribution of terrestrial animal reservoirs of HEV is summarized in Fig. 10.2. Pigs, wild boar, and various species of deer are involved in zoonotic transmission of HEV-3 and HEV-4 to humans. However, the role of mongooses, rats, and rabbits in causing human hepatitis E is unclear. Domestic pigs are the most important reservoirs of the HEV genotypes that are capable of infecting humans. In 1997, HEV-3 was first isolated from pigs in USA (Meng et al. 1997) and since then many studies reported high prevalence of HEV (seroprevalences were estimated between 5 and 100%) in pig herds worldwide (de la Caridad Montalvo Villalba et al. 2013; Owolodun et al. 2014; Aniță et al. 2014; Burri et al. 2014; Liu et al. 2015; Merino-Ramos et al. 2016; Thiry et al. 2017a, b), including countries from five continents:

- (a) Asia (China, India, Indonesia, Japan, Korea, Mongolia, Philippines, Taiwan, Thailand, and Vietnam),

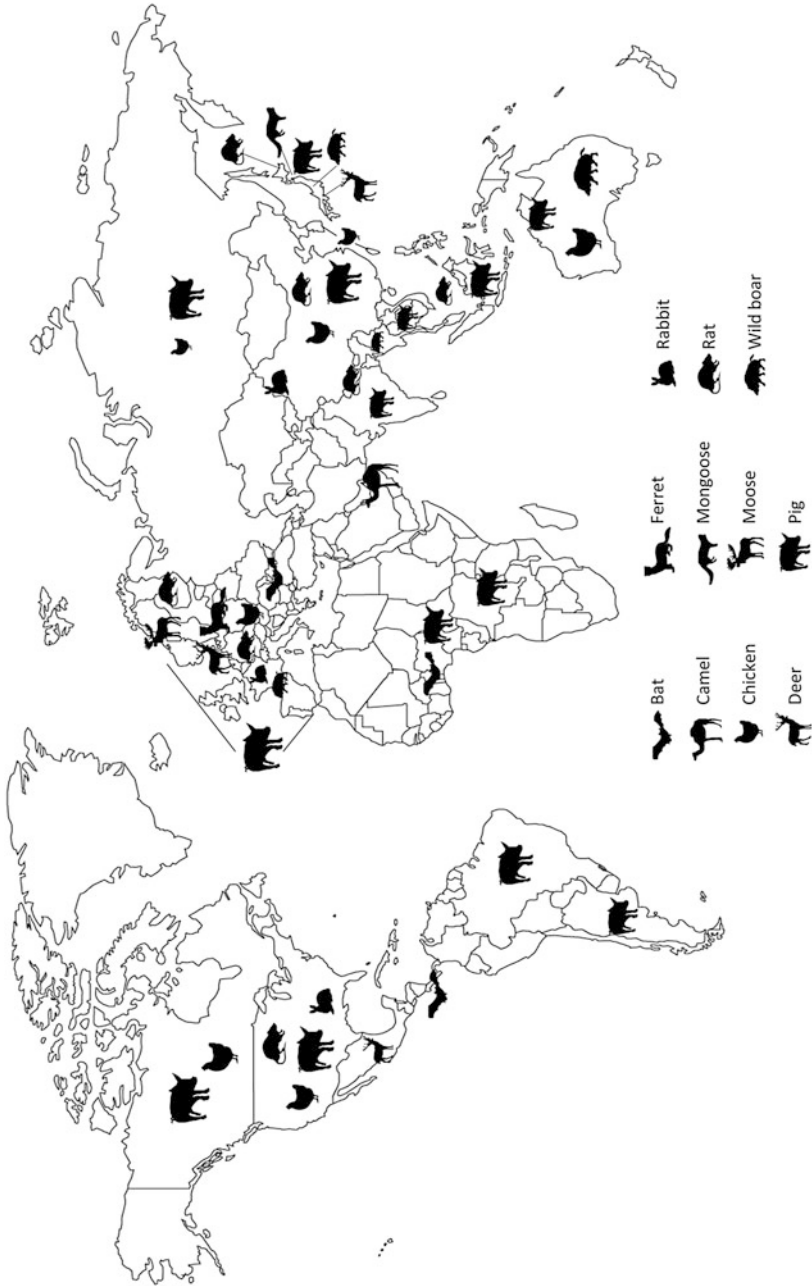


Fig. 10.2 Geographical distribution of HEV3 and HEV4, and novel HEV-like viruses in terrestrial animals (adopted from Sridhar et al. 2015)

- (b) South and North America (Argentina, Bolivia, Brazil, Cuba, and Mexico/Canada),
- (c) Africa (Cameroon, Democratic Republic of Congo, Nigeria, and Madagascar),
- (d) Europe (Belgium, Czech Republic, Finland, France, Germany, Hungary, Italy, the Netherlands, Romania, Spain, Sweden, Switzerland, and the United Kingdom), and
- (e) Oceania (Australia, New Caledonia, and New Zealand).

HEV-3 has been detected in pigs from all aforementioned continents, whereas HEV-4 has mainly been reported in pigs and humans in Asia and recently also in Europe (Hsieh et al. 1999; Schlauder and Mushahwar 2001; Cooper et al. 2005; Thiry et al. 2017a, b; Salines et al. 2017; Pavio et al. 2017).

10.2.2 Transmission of HEV to Humans

Mainly reservoirs for genotypes HEV-3 and HEV-4 are domestic pigs and wildlife (wild boars, sika deer) (Pavio et al. 2015). HEV is transmitted primarily in humans via the faecal–oral route (Purcell and Emerson 2001). Human infections are due mainly to consumption of undercooked meat or meat products (e.g. sausages), direct contact with infected animals, drinking contaminated water and environmental contamination by animal manure run-off (Khuroo et al. 2016).

10.2.2.1 Public Health

HEV is the main causative agent of hepatitis E, which is usually an asymptomatic human liver disease. However, hepatitis E is possible to induce a self-limited acute hepatitis in humans, especially in developing countries, which are characterized by problems of access to water, poor sanitation, and high population density (Purcell and Emerson 2000; Worm et al. 2002; Perez-Gracia and Rodriguez-Iglesias 2003). The genotypes HEV-1, HEV-2, HEV-3, and HEV-4 are frequently associated with clinical cases of acute hepatitis or liver failure, as well as neurological problems. Moreover, human infections from genotypes 1 and 2 (HEV-1, HEV-2) are more associated with high mortality rates in pregnant women and pancreatitis incidence (Lhomme et al. 2012, 2016). During last decade, the reported HEV infections are increasing dramatically, due to more frequent and novel application of diagnostic methods (Aspinall et al. 2015).

Generally, hepatitis E due to HEV-3 and HEV-4 infection is an important zoonosis around the world (Wu et al. 2002; Smith et al. 2014). Sporadic cases of acute and chronic hepatitis E in humans due to HEV-3 infection were reported in non-endemic regions of industrialized countries, where the pig was the major source of infection (Smith et al. 2014). Epidemic forms of hepatitis E were associated with

infection via drinking of contaminated water in developing countries with poor sanitary conditions, as the contamination of water supplies with human faeces remain a common route of HEV spread in these countries (Kamar et al. 2017). Sporadic forms of hepatitis E have been reported between epidemics of disease in these areas or in humans–patients with previous travelling to endemic areas or in humans–patients from industrialized countries, without travelling abroad (autochthonous hepatitis) (Perez-Gracia et al. 2004). Sporadic cases of hepatitis E were associated with the consumption of raw or undercooked meat products (e.g. liver, sausage) from pig or deer (Meng 2011). Large outbreaks of HEV frequently occur in many tropical and subtropical low-income regions, whereas sporadic HEV infections are seen in humans in industrialized countries. HEV sequences isolated from domestic pigs, wild boar, or deer were reported to be closely related to human HEV sequences in many countries worldwide (Meng 2011).

During last years, human HEV-3 infections have been dramatically increasing and the zoonotic transmission from pig to human is a common fact, based on the high sequence identity between isolated strains of human cases and contemporary isolated strains in pigs (Adlhoch et al. 2016). Nowadays, hepatitis E is an important public health concern, as about 20 million new HEV-1 and HEV-2 per year are reporting, including 3.4 million acute cases with 70,000 deaths due to acute liver disease (Rein et al. 2012). For example, studies reported a 10–40% seroprevalence rate of anti-HEV antibodies in many areas of Africa and Asia, while about 80% in Egypt (Kamar et al. 2017).

10.2.2.2 Pork and Meat Products

HEV foodborne infections in humans are caused mainly after consumption of undercooked meat or various meat products, such as sausages (Colson et al. 2010; Guillois et al. 2016). The consumption the parboiled flesh or liver from wild boar, deer, and domestic pigs is associated with autochthonous cases and outbreaks of hepatitis E (Khuroo and Khuroo 2008; Miyashita et al. 2012). Many studies reported detection of HEV-specific RNA in meat and meat products (mainly in liver as well as in sausages with and without liver) worldwide (Yazaki et al. 2003; Feagins et al. 2007; Kulkarni and Arankalle 2008; PAVIO et al. 2017). Recently, HEV-contaminated cow milk is reported as a new high risk factor for HEV foodborne infection (Huang et al. 2016) (Table 10.1).

Studies reported a seroprevalence between 2 and 15% of slaughtered pigs, while the detection of HEV in samples from sausages or meat products containing pig liver was higher (especially products prepared with raw pork liver), ranging between 16 and 47%, (Pavio et al. 2014; Di Bartolo et al. 2015; Crossan et al. 2015). For example in Europe, favourite products made from raw pig liver (e.g. fresh sausage made called Figatellu), which are traditionally eaten raw, are considered at high risk of containing HEV (Colson et al. 2010; Garbuglia et al. 2015; Matsuda et al. 2003). Except pig livers, liver from wild boar and deer are also considered at high risk of containing HEV (Tei et al. 2003). However, in a recent study reported that the

Table 10.1 Prevalence of HEV RNA-positive pork, wild boar, and deer meat products

Product	Geographic area (continent, country)	References
Pig—liver	Asia (China, India Hong Kong, Japan, and Thailand)	Li et al. (2009) Kulkarni and Arankalle (2008) Chan et al. (2017) Okano et al. (2014) Ishida et al. (2012) Intharasongkroh et al. (2017)
	North – South America (USA, Canada, Brazil, and Mexico)	Gardinali et al. (2012) Mykytczuk et al. (2017) Leblanc et al. (2010) Wilhelm et al. (2014) Cantú-Martínez et al. (2013) Feagins et al. (2007)
	Africa (Cameroon, Burkina Faso)	de Paula et al. (2013) Traoré et al. (2015)
	Western, Central and South Europe (France, the United Kingdom, The Netherlands, Germany, Czech Republic, Italy, Serbia, Spain)	Di Bartolo et al. (2010) Jori et al. (2016) Rose et al. (2011) Wenzel et al. (2011) Milojević et al. (2019) Bouwknegt et al. (2007) Berto et al. (2012a, b)
Pig—meat	Western, Central, and South Europe (The Netherlands, Czech Republic, Switzerland, Italy)	Di Bartolo et al. (2010) Boxman et al. (2019) Moor et al. (2018)
	South-East Asia (Thailand)	Intharasongkroh et al. (2017)
Sausages and other products (e.g. figatelli) containing or without liver	North – South America (Canada, Brazil)	Heldt et al. (2016) Mykytczuk et al. (2017)
	South Africa (Republic of South Africa)	Korsman et al. (2019)
	Western, Central, and South Europe (France, United Kingdom, The Netherlands Germany, Switzerland, Spain, Italy)	Colson et al. (2010) Hennechart-Collette et al. (2019) Pavio et al. (2014) Szabo et al. (2015) Garbuglia et al. (2015) Boxman et al. (2019) Martin-Latil et al. (2016) Di Bartolo et al. (2015) Giannini et al. (2018) Berto et al. (2012a, b)

(continued)

Table 10.1 (continued)

Product	Geographic area (continent, country)	References
Wild boar—liver	Western, Central, South and East Europe (Belgium, France, The Netherlands Germany, Czech Republic, Hungary, Italy, Romania)	Thiry et al. (2017a, b) Kaba et al. (2010) Anheyer-Behmenburg et al. (2017) Schielke et al. (2009) Schielke et al. (2015) Adlhoch et al. (2009) Kubankova et al. (2015) Forgách et al. (2010) Serracca et al. (2015) Montagnaro et al. (2015) Porea et al. (2018)
	East Asia (Japan)	Sato et al. (2011) Matsuda et al. (2003) Motoya et al. (2016) Sonoda et al. (2004)
Wild boar—meat	Central Europe (Germany)	Anheyer-Behmenburg et al. (2017) Schielke et al. (2015)
Wild boar—Sausages without liver	Western and Central Europe (Belgium, Germany)	Szabo et al. (2015) Thiry et al. (2017a, b)
Deer—liver	Western and Central Europe (Belgium, France, The Netherlands, Germany, Hungary)	Szabo et al. (2015) Thiry et al. (2017a, b) Lhomme et al. (2015) Anheyer-Behmenburg et al. (2017) Forgách et al. (2010) Rutjes et al. (2010)
Deer—meat	Central Europe (Germany)	Anheyer-Behmenburg et al. (2017) Schielke et al. (2015)

prevalence and the amount of HEV RNA in liver samples from deer were significantly lower in comparison to samples from domestic pigs and wild boars (Pavio et al. 2017).

10.2.2.3 Direct contact/Vocational exposure

Direct contact exposure is also reported as a possible route of HEV transmission. Moreover, studies in many countries reported that the vocational exposure of professionals in pig farms (e.g. swine veterinarians, farm workers) with pigs, manure, and sewage is an important high risk factor for HEV infections (Perez-Gracia et al. 2007; Bouwknegt et al. 2008a; Rutjes et al. 2009; Pavio et al. 2017). For example, swine veterinarians and workers in pig farms reported to be 2–5 times more under

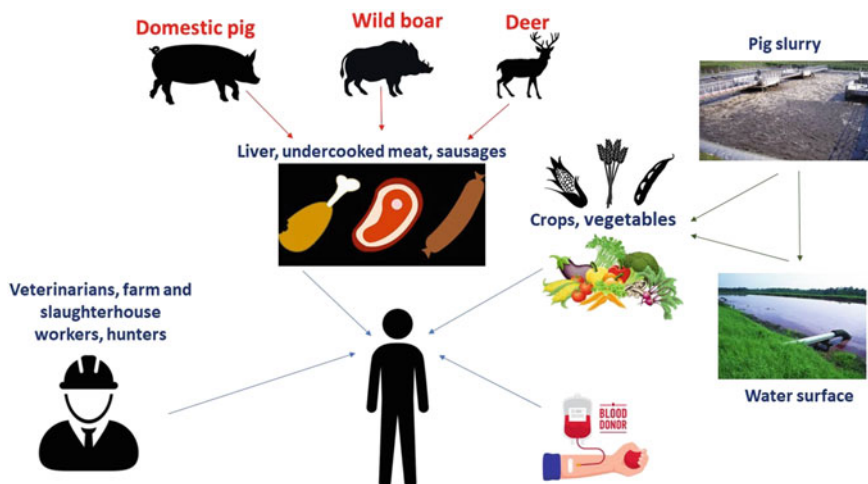


Fig. 10.3 Transmission and exposure routes of HEV infection in humans

the risk to have anti-HEV antibodies in comparison to non-swine veterinarians and the general population (Olsen et al. 2006; Vulcano et al. 2007; Galiana et al. 2008; Bouwknegt et al. 2008b). Moreover, there are reports of HEV transmission due to contact from wildlife to forest workers and hunters, as well as from frequent contact to a pet pig or to pigs at slaughterhouse (Stramer 2014; Juhl et al. 2014; Pavio et al. 2017) (Fig. 10.3).

Recently, HEV transmission from ruminants to farmers (Tritz et al. 2018) and rabbits to humans (slaughterhouse workers) were reported (Geng et al. 2019).

10.2.2.4 Water/Pig Slurry

Hepatitis E is primarily transmitted through the faecal–oral route (Khuroo 1991). Gross faecal contamination of community water supplies has been associated with several outbreaks in developing countries (Khuroo 1980; Naik et al. 1992; Kamar et al. 2017).

The presence of pig manure indicates the potential spread to humans through contact with contaminated crops or in personnel that handle swine manure and spread this waste on agricultural fields (Fernandez-Barredo et al. 2006). Use of pig slurry as pasture can infect agricultural products, such as raspberries, strawberries, and many vegetables used in the salad (Ward et al. 2008; Brassard et al. 2012). Run-off from outdoor pig farms causes contamination of surface water as well as produce receiving surface water (Steyer et al. 2011; Tyrrel and Quinton 2003).

10.2.2.5 Iatrogenic

HEV transmission from blood HEV-infected donors to human by blood transfusion is reported in many studies (Baylis et al. 2012; Hewitt et al. 2014; Gallian et al. 2014; Sauleda et al. 2015; Hogema et al. 2016). Moreover, a case of HEV-7 transmission to human is reported for a liver transplant recipient (Lee et al. 2016).

10.2.3 *Transmission of HEV to Pigs*

Wild boars are recognized as a potential reservoir of HEV, while HEV is transmitted from them to domestic pigs (Thiry et al. 2016; Schlosser et al. 2015; Jori et al. 2016).

The primary route of natural HEV transmission in pigs is the faecal–oral route, but it may require repeated exposure and high doses of virus (Kasorndorkbua et al. 2004). It is remarkable that the duration of detection of HEV in pig faeces is considerably longer than the duration of HEV viremia (Kasorndorkbua et al. 2004).

Previous studies reported a seroprevalence of HEV in pig between 5% and 100% (Pavio et al. 2017). The prevalence of the virus is depended on the animals age, the kind of tested sample, and the diagnostic method. Usually, HEV infection is detected at an early age after the loss of maternal antibodies. The virus load is high in all ages (weaners, growers, and fatteners), but is reported to be the highest in fatteners. Moreover, the seroprevalence is depended on the production system, as a slightly higher seroprevalence was reported in organic farms compared with conventional and free-range pig farms (Berto et al. 2012a, b). A comprehensive review (Salines et al. 2017) reported that the detection of HEV RNA in pig faeces and serum depends on the pig's age, while the shedding period ranges from 1.5 to 5 months of age (Salines et al. 2017). However, the peak of shedding in faeces happens around 3–4 months of age, whereas the shedding prevalence at slaughter age (around 185 days of age) is possible to be around 6% (Fig. 10.4).

10.3 Pathogenesis

HEV replication occurs mainly in the liver, but the virus can also be detected in other organs, such as small intestine, lymph nodes, and colon (Williams et al. 2001; Ha and Chae 2004). Viraemia is transient (duration of 1–2 weeks), while the peak of viral shedding in faeces occurs 3–8 weeks after weaning (Kantala et al. 2015). The viral shedding in faeces of infected pigs may persist for up to 7 weeks (Pavio et al. 2010). Then it is decreased around 15–18 weeks of age (McCreary et al. 2008), with the appearance of antibodies IgM followed by IgG (seroconversion) (Pavio et al. 2010).

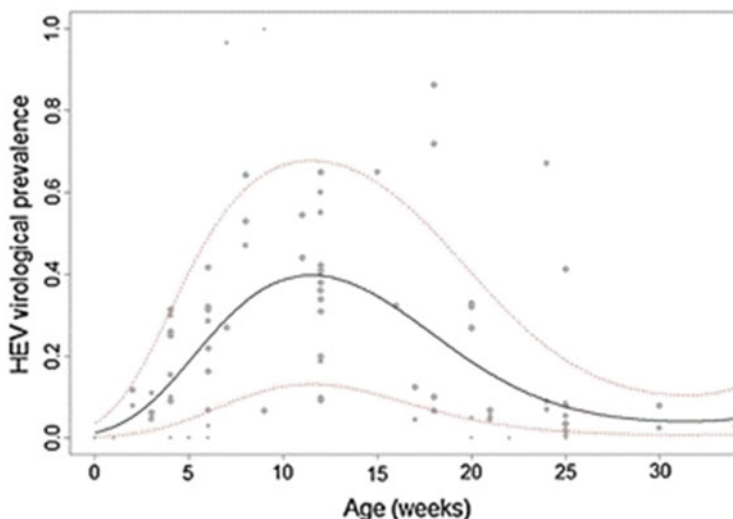


Fig. 10.4 Predicted HEV prevalence in faeces according to animal age (adapted from Salines et al. 2017)

The duration of the immunity acquired after HEV infection is not clear, but re-infection in case of transient decrease of immunity could be possible. A decrease of protection (antibodies or cellular response) over time may happen in older animals and especially in sows (Casas et al. 2011). Many studies reported that the majority of pigs are infected at 8–15 weeks of age, but some of them could remain positive at slaughter age (de Deus et al. 2008; Meng et al. 1997; Casas et al. 2011). The infection happens at an early age after the loss of maternal antibodies (MAbs), which can be transferred from HEV-Ab positive sows to offspring (Feng et al. 2011). High levels of MAbs are very important for the reduction of prevalence of HEV positive animals (Krog et al. 2019).

10.4 Clinical Signs in Domestic Pig and Wild Boar

HEV infection in pig is usually asymptomatic, without important impact on their health status. HEV replication occurs in the liver and the intestine (Ha and Chae 2004), while it may enhance the clinical performance of disease caused by other porcine viruses, such as porcine reproductive and respiratory syndrome virus (PRRSV) (Salines et al. 2015) or porcine circovirus 2 (PCV2) (Yang et al. 2015; Jäckel et al. 2018). Immune modulatory effects have been reported in cases of PCV2 and HEV co-infection (Jäckel et al. 2018). The aforementioned enhancing activity of the HEV may be due to immunosuppressive properties (Cao et al. 2017).

Genotypes HEV-3 and HEV-4 were also reported in wild boars, but without characteristic clinical symptoms in most cases. However, the prevalence of HEV in wild boars is lower than in domestic pigs (Pavio et al. 2017). Furthermore, HEV was detected in species of deer (Neumann et al. 2016; Anheyer-Behmenburg et al. 2017), while other species of animals (e.g. ruminants) are reported to be susceptible to HEV infection (Spahr et al. 2018).

10.5 HEV Monitoring/Prevention

Future studies focus on preventive measures to eliminate the appearance and persistence of HEV in pig farms (including biosecurity and vaccination) are required. Moreover, more studies are needed to investigate deeply the role of wildlife (wild boars, deer, etc.) in the epidemiology of HEV infection.

The prevention of zoonotic HEV infection demands a monitoring system to investigate and prevent the contamination of pork-derived meat products. HEV monitoring activities in the pork production chain are important to be implemented for the following targets:

- (a) to maintain a database for the prevalence of HEV and follow-up the prevalence of the different HEV strains;
- (b) to investigate in detail the dynamics of HEV infection, as well as their risk factors;
- (c) to remove contaminated livers and other high-risk meat products from the food chain;
- (d) to inform consumers regarding handling and cooking of high-risk pork-derived meat products (Salines et al. 2017; ANSES 2013).

Acknowledgements The author of the manuscript thanks and acknowledges their Institute.

Conflict of Interest There is no conflict of interest.

References

- Adlhoch C, Wolf A, Meisel H et al (2009) High HEV presence in four different wild boar populations in East and West Germany. *Vet Microbiol* 139(3–4):270–278
- Adlhoch C, Avellon A, Baylis SA et al (2016) Hepatitis E virus: assessment of the epidemiological situation in humans in Europe, 2014/15. *J Clin Virol* 82:9–16
- Agence nationale de sécurité sanitaire de l'alimentation, de l'environnement et du travail (ANSES) (2013) Opinion of the French Agency for Food, Environmental and Occupational Health & Safety concerning the "Request to assess the risks related to contamination of delicatessen meats products derived from raw pork liver with hepatitis E virus (HEV)." ANSES opinion request 2012-SA-0012. <https://www.anses.fr/fr/system/files/BIORISK2012sa0012EN.pdf>. Accessed 17 Aug 2019

- Anheyer-Behmenburg HE, Szabo K, Schotte U et al (2017) Hepatitis E Virus in Wild Boars and Spillover Infection in Red and Roe Deer, Germany, 2013-2015. *Emerg Infect Dis* 23 (1):130-133
- Aniță A, Gorgan L, Aniță D et al (2014) Evidence of hepatitis E infection in swine and humans in the East Region of Romania. *Int J Infect Dis* 29:232-237
- Aspinall EJ, Couturier E, Faber M et al (2017) On behalf of the country experts. Hepatitis E virus infection in Europe: Surveillance and descriptive epidemiology of confirmed cases, 2005 to 2015. *Eurosurveillance* 22:30561
- Balayan MS (1997) Type E hepatitis: state of the art. *Int J Infect Dis* 2:113-120
- Baylis SA, Gärtner T, Nick S et al (2012) Occurrence of hepatitis E virus RNA in plasma donations from Sweden, Germany and the United States. *Vox Sang* 103:89-90
- Berto A, Martelli F, Grierson S et al (2012a) Hepatitis E virus in pork food chain, United Kingdom, 2009-2010. *Emerg Infect Dis* 18:1358-1360
- Berto A, Backer JA, Mesquita JR et al (2012b) Prevalence and transmission of hepatitis E virus in domestic swine populations in different European countries. *BMC Res Notes* 5:190
- Bouwknegt M, Lodder-Verschoor F, van der Poel WHM et al (2007) Hepatitis E virus RNA in commercial porcine livers in The Netherlands. *J Food Prot* 70:2889-2895
- Bouwknegt M, Frankena K, Rutjes SA et al (2008a) Estimation of hepatitis E virus transmission among pigs due to contact-exposure. *Vet Res* 39:40
- Bouwknegt M, Engel B, Herremans MM et al (2008b) Bayesian estimation of hepatitis E virus seroprevalence for populations with different exposure levels to swine in The Netherlands. *Epidemiol Infect* 136:567-576
- Boxman ILA, Jansen CCC, Hägele G et al (2019) Monitoring of pork liver and meat products on the Dutch market for the presence of HEV RNA. *Int J Food Microbiol* 296:58-64
- Brassard J, Gagne MJ, Genreux M et al (2012) Detection of human food-borne and zoonotic viruses on irrigated, field-grown strawberries. *Appl Environ Microbiol* 78:3763-3766
- Burri C, Vial F, Ryser-Degiorgis M-P et al (2014) Seroprevalence of hepatitis E virus in domestic pigs and wild boars in Switzerland. *Zoonoses Public Health* 61:537-544
- Cantú-Martínez MA, Roig-Sagués AX, Cedillo-Rosales S et al (2013) Molecular detection of hepatitis E virus in pig livers destined for human consumption in the state of Nuevo Leon, Mexico. *Salud Publica Mex* 55:193-195
- Cao D, Cao QM, Subramaniam S et al (2017) Pig model mimicking chronic hepatitis E virus infection in immunocompromised patients to assess immune correlates during chronicity. *Proc Natl Acad Sci USA* 114:6914-6923
- Casas M, Cortés R, Pina S et al (2011) Longitudinal study of hepatitis E virus infection in Spanish farrow-to-finish swine herds. *Vet Microbiol* 148:27-34
- Chan MCW, Kwok K, Hung T-N et al (2017) Molecular epidemiology and strain comparison between hepatitis E virus in human sera and pig livers during 2014-2016 in Hong Kong. *J Clin Microbiol* 55:1408-1415
- Chaussade H, Rigaud E, Allix A et al (2013) Hepatitis E virus seroprevalence and risk factors for individuals in working contact with animals. *J Clin Virol* 58:504-508
- Colson P, Borentain P, Queyriaux B et al (2010) Pig liver sausage as a source of hepatitis E virus transmission to humans. *J Infect Dis* 202(6):825-834
- Cooper K, Huang FF, Batiste L et al (2005) Identification of genotype 3 hepatitis E virus (HEV) in serum and fecal samples from pigs in Thailand and Mexico, where genotype 1 and 2 HEV strains are prevalent in the respective human population. *J Clin Microbiol* 43:1684-1688
- Crossan C, Grierson S, Thomson J et al (2015) Prevalence of hepatitis E virus in slaughter-age pigs in Scotland. *Epidemiol Infect* 143:2237-2240
- de Deus N, Casas M, Peralta B et al (2008) Hepatitis E virus infection dynamics and organic distribution in naturally infected pigs in a farrow-to-finish farm. *Vet Microbiol* 132:19-28
- de la Caridad Montalvo Villalba MD, Owot JC, Benedito EC et al (2013) Hepatitis E virus genotype 3 in humans and swine, Cuba. *Infect Genet Evol* 14:335-339
- de Paula VS, Wiele M, Mbunkah AH et al (2013) Hepatitis E virus genotype 3 strains in domestic pigs, Cameroon. *Emerg Infect Dis* 19:666-668

- Di Bartolo I, Diez-Valcarce M, Vasickova P et al (2010) Hepatitis E virus in pork production chain in Czech Republic, Italy, and Spain, 2010. *Emerg Infect Dis* 18(8):1282–1289
- Di Bartolo I, Angeloni G, Ponterio E et al (2015) Detection of hepatitis E virus in pork liver sausages. *Int J Food Microbiol* 193:29–33
- Dremsek P, Wenzel JJ, Johne R et al (2012) Seroprevalence study in forestry workers from eastern Germany using novel genotype 3- and rat hepatitis E virus-specific immunoglobulin G ELISAs. *Med Microbiol Immunol* 201:189–200
- Emerson SU, Purcell RH (2001) Recombinant vaccines for hepatitis E. *Trends Mol Med* 7:462–466
- Favorov MO, Kosoy MY, Tsarev SA (2000) Prevalence of antibody to hepatitis E virus among rodents in the United States. *J Infect Dis* 181:449–455
- Feagins AR, Opriessnig T, Guenette DK et al (2007) Detection and characterization of infectious Hepatitis E virus from commercial pig livers sold in local grocery stores in the USA. *J Gen Virol* 88:912–917
- Feng R, Zhao C, Li M et al (2011) Infection dynamics of hepatitis E virus in naturally infected pigs in a Chinese farrow-to-finish farm. *Infect Genet Evol* 11:1727–1731
- Galiana C, Fernandez-Barredo S, Galiana C, Garcia A et al (2006) Detection of hepatitis E virus shedding in feces of pigs at different stages of production using reverse transcription- polymerase chain reaction. *Vet Diagn Invest* 18:462–465
- Forgách P, Nowotny N, Erdélyi K et al (2010) Detection of hepatitis E virus in samples of animal origin collected in Hungary. *Vet Microbiol* 143(2-4):106–116
- Galiana C, Fernandez-Barredo S, Garcia A et al (2008) Occupational exposure to hepatitis E virus (HEV) in swine workers. *Am J Trop Med Hyg* 78:1012–1015
- Gallian P, Lhomme S, Piquet Y et al (2014) Hepatitis E virus infections in blood donors, France. *Emerg Infect Dis* 20(11):1914–1917
- Garbuglia AR, Alessandrini AI, Pavo N et al (2015) Male patient with acute hepatitis E in Genoa, Italy: figatelli (pork liver sausage) as probable source of the infection. *Clin Microbiol Infect* 21(1):e4–e6
- Gardinali NR, Barry AF, Otonel RAA et al (2012) Hepatitis E virus in liver and bile samples from slaughtered pigs of Brazil. *Mem Inst Oswaldo Cruz* 107:935–939
- Geng Y, Zhao C, Geng K, et al (2019) High seroprevalence of hepatitis E virus in rabbit slaughterhouse workers. *Transbound Emerg Dis* 66(2):1085–1089
- Giannini P, Jermini M, Leggeri L et al (2018) Detection of Hepatitis E virus RNA in raw cured sausages and raw cured sausages containing pig liver at retail stores in Switzerland. *J Food Prot* 81(1):43–45
- Guillois Y, Abravanel F, Miura T et al (2016) High proportion of asymptomatic infections in an outbreak of hepatitis E associated with a spit-roasted piglet, France, 2013. *Clin Infect Dis* 62:351–357
- Ha SK, Chae C (2004) Immunohistochemistry for the detection of swine hepatitis E virus in the liver. *J Viral Hepat* 11:263–267
- Heldt FH, Staggmeier R, Gularte JS (2016) Hepatitis E virus in surface water, sediments, and pork products marketed in Southern Brazil. *Food Environ Virol* 8(3):200–205
- Hennechart-Collette C, Fraisse A, Guillier L et al (2019) Evaluation of methods for elution of HEV particles in naturally contaminated sausage, figatelli and pig liver. *Food Microbiol* 84:103235
- Hewitt PE, Ijaz S, Brailsford SR et al (2014) Hepatitis E virus in blood components: a prevalence and transmission study in southeast England. *Lancet* 384:1766–1773
- Hogema BM, Molier M, Sjerps M et al (2016) Incidence and duration of hepatitis E virus infection in Dutch blood donors. *Transfusion* 56:722–728
- Hsieh SY, Meng XJ, Wu YH et al (1999) Identity of a novel swine hepatitis E virus in Taiwan forming a monophyletic group with Taiwan isolates of human hepatitis E virus. *J Clin Microbiol* 37(12):3828–3834
- Huang F, Li Y, Yu W et al (2016) Excretion of infectious hepatitis E virus into milk in cows imposes high risks of zoonosis. *Hepatology* 64(2):350–359

- Intharasongkroh D, Sa-Nguanmoo P, Tuanthap S et al (2017) Hepatitis E virus in pork and variety meats sold in fresh markets. *Food Environ Virol* 9:45–53
- Ishida S, Yoshizumi S, Ikeda T et al (2012) Detection and molecular characterization of hepatitis E virus in clinical, environmental and putative animal sources. *Arch Virol* 157:2363–2368
- Jäckel S, Muluneh A, Pöhle D et al (2018) Co-infection of pigs with Hepatitis E and porcine circovirus 2, Saxony 2016. *Res Vet Sci* 123:35–38
- Johne R, Dremsek P, Reetz J et al (2014) Hepeviridae: an expanding family of vertebrate viruses. *Infect Genet Evol* 27:212–229
- Jori F, Laval M, Maestrini O et al (2016) Assessment of domestic pigs, wild boars and feral hybrid pigs as reservoirs of hepatitis E virus in Corsica, France. *Viruses* 8(8):E236
- Juhl D, Baylis SA, Blümel J et al (2014) Seroprevalence and incidence of hepatitis E virus infection in German blood donors. *Transfusion* 54:49–56
- Kaba M, Davoust B, Marié J-L et al (2010) Detection of hepatitis E virus in wild boar (*Sus scrofa*) livers. *Vet J* 186:259–261
- Kamar N, Izopet J, Pavo N et al (2017) Hepatitis E virus infection. *Nat Rev Dis Primers* 3:17086
- Kantala T, Heinonen M, Oristo S et al (2015) Hepatitis E virus in young pigs in Finland and characterization of the isolated partial genomic sequences of genotype 3 HEV. *Foodborne Pathog Dis* 12:253–260
- Kasorndorkbua C, Guenette DK, Huang FF et al (2004) Routes of transmission of swine hepatitis E virus in pigs. *J Clin Microbiol* 42(11):5047–5052
- Khuroo MS (1980) Study of an epidemic of non-a, non-b hepatitis. Possibility of another human hepatitis virus distinct from post-transfusion non-a, non-b type. *Am J Med* 68:818–824
- Khuroo MS (1991) Hepatitis E: the enterically transmitted non-a, non-b hepatitis. *Indian J Gastroenterol* 10:96–100
- Khuroo MS, Khuroo MS (2008) Hepatitis E virus. *Curr Opin Infect Dis* 21:539–543
- Khuroo MS, Khuroo MS, Khuroo NS (2016) Transmission of Hepatitis E virus in developing countries. *Viruses* 8(9):253
- Korsman S, Bloemberg J, Brombacher M (2019) Hepatitis E in pig-derived food products in Cape Town, South Africa, 2014. *S Afr Med J* 109(8):584–586
- Krog JS, Larsen LE, Breum SØ (2019) Tracing hepatitis E virus in pigs from birth to slaughter. *Front Vet Sci* 6:50
- Kubankova M, Kralik P, Lamka J et al (2015) Prevalence of hepatitis E virus in populations of wild animals in comparison with animals bred in game enclosures. *Food Environ Virol* 7:159–163
- Kulkarni MA, Arankalle VA (2008) The detection and characterization of hepatitis E virus in pig livers from retail markets of India. *J Med Virol* 80:1387–1390
- Leblanc D, Poitras E, Gagné MJ (2010) Hepatitis E virus load in swine organs and tissues at slaughterhouse determined by real-time RT-PCR. *Int J Food Microbiol* 139(3):206–209
- Lee GH, Tan BH, Teo EC et al (2016) Chronic infection with Camelid hepatitis E virus in a liver transplant recipient who regularly consumes Camel Meat and Milk. *Gastroenterology* 150:55–57
- Lhomme S, Abravanel F, Dubois M et al (2012) Hepatitis E virus quasispecies and the outcome of acute hepatitis E in solid-organ transplant patients. *J Virol* 86:10006–10014
- Lhomme S, Top S, Bertagnoli S et al (2015) Wildlife reservoir for hepatitis E virus, Southwestern France. *Emerg Infect Dis* 21:1224–1226
- Lhomme S, Marion O, Abravanel F et al (2016) Hepatitis E. *Viruses* 8:212
- Li W, She R, Wei H et al (2009) Prevalence of hepatitis E virus in swine under different breeding environment and abattoir in Beijing, China. *Vet Microbiol* 133:75–83
- Liu X, Saito M, Sayama Y et al (2015) Seroprevalence and molecular characteristics of hepatitis E virus in household-raised pig population in the Philippines. *BMC Vet Res* 11:11
- Martin-Latil S, Hennechart-Collette C, Delannoy S et al (2016) Quantification of hepatitis E virus in naturally contaminated pig liver products. *Front Microbiol* 7:1183
- Matsuda H, Okada K, Takahashi K et al (2003) Severe hepatitis E virus infection after ingestion of uncooked liver from a wild boar. *J Infect Dis* 188:944

- McCreary C, Martelli F, Grierson S et al (2008) Excretion of hepatitis E virus by pigs of different ages and its presence in slurry stores in the United Kingdom. *Vet Rec* 163:261–265
- Meng XJ (2000) Novel strains of hepatitis E virus identified from humans and other animal species: is hepatitis E a zoonosis? *J Hepatol* 33:842–845
- Meng XJ (2010) Hepatitis E virus: animal reservoirs and zoonotic risk. *Vet Microbiol* 140:256–265
- Meng XJ (2011) From barnyard to food table: the omnipresence of hepatitis E virus and risk for zoonotic infection and food safety. *Virus Res* 161:23–30
- Meng XJ, Purcell RH, Halbur PG et al (1997) A novel virus in swine is closely related to the human hepatitis E virus. *Proc Natl Acad Sci USA* 94:9860–9865
- Merino-Ramos T, Martín-Acebes MA, Casal J et al (2016) Prevalence of hepatitis E virus (HEV) antibodies in Mexican pigs. *Food Environ Virol* 8:156–159
- Milojević L, Velebit B, Teodorović V et al (2019) Screening and molecular characterization of hepatitis E virus in slaughter pigs in Serbia. *Food Environ Virol* 11:410–419. <https://doi.org/10.1007/s12560-019-09393-1>
- Miyashita K, Kang JH, Saga A et al (2012) Three cases of acute or fulminant hepatitis E caused by ingestion of pork meat and entrails in Hokkaido, Japan: zoonotic food-borne transmission of hepatitis E virus and public health concerns. *Hepatol Res* 42:870–878
- Montagnaro S, De Martinis C, Sasso S et al (2015) Viral and antibody prevalence of hepatitis E in European wild boars (*Sus scrofa*) and hunters at zoonotic risk in the Latium region. *J Comp Pathol* 153(1):1–8
- Moor D, Liniger M, Baumgartner A et al (2018) Screening of ready-to-eat meat products for hepatitis E virus in Switzerland. *Food Environ Virol* 10(3):263–271
- Motoya T, Nagata N, Komori H et al (2016) The high prevalence of hepatitis E virus infection in wild boars in Ibaraki Prefecture, Japan. *J Vet Med Sci* 77:1705–1709
- Mykytczuk O, Harlow J, Bidawid S et al (2017) Prevalence and molecular characterization of the hepatitis E virus in retail pork products marketed in Canada. *Food Environ Virol* 9:208–218
- Naik SR, Aggarwal R, Salunke PN et al (1992) A large waterborne viral hepatitis E epidemic in Kanpur, India. *Bull World Health Organ* 70:597–604
- Neumann S, Hackl SS, Piepenschneider M et al (2016) Serologic and molecular survey of hepatitis E virus in German deer populations. *J Wildl Dis* 52:106–113
- Okano H, Takahashi M, Isono Y et al (2014) Characterization of sporadic acute hepatitis E and comparison of hepatitis E virus genomes in acute hepatitis patients and pig liver sold as food in Mie, Japan. *Hepatol Res* 44:E63–E76
- Olsen B, Axelsson-Olsson D, Thelin A et al (2006) Unexpected high prevalence of IgG-antibodies to hepatitis E virus in Swedish pig farmers and controls. *Scand J Infect Dis* 38:55–58
- Owolodun OA, Gerber PF, Giménez-Lirola LG et al (2014) First report of hepatitis E virus circulation in domestic pigs in Nigeria. *Am J Trop Med Hyg* 91:699–704
- Pavio N, Meng XJ, Renou C (2010) Zoonotic hepatitis E: animal reservoirs and emerging risks. *Vet Res* 41:46
- Pavio N, Merbah T, Thébault A (2014) Frequent hepatitis E virus contamination in food containing raw pork liver, France. *Emerg Infect Dis* 20:1925–1927
- Pavio N, Meng XJ, Doceul V (2015) Zoonotic origin of hepatitis E. *Curr Opin Virol* 10:34–41
- Pavio N, Doceul V, Bagdassarian E et al (2017) Recent knowledge on hepatitis E virus in Suidae reservoirs and transmission routes to human. *Vet Res* 48:78
- Perez-Gracia MT, Rodriguez-Iglesias M (2003) Aspectos actuales del virus de la hepatitis E [Hepatitis E virus: current status]. *Med Clin (Barc)* 121:787–792
- Perez-Gracia MT, Garcia-Valdivia MS, Galan F et al (2004) Detection of hepatitis E virus in patients' sera in southern Spain. *Acta Virol* 48:197–200
- Perez-Gracia MT, Mateos ML, Galiana C et al (2007) Autochthonous hepatitis E infection in a slaughterhouse worker. *Am J Trop Med Hyg* 77:893–896
- Porea D, Anita A, Demange A et al (2018) Molecular detection of hepatitis E virus in wild boar population in eastern Romania. *Transbound Emerg Dis* 65(2):527–533
- Purcell RH, Emerson SU (2000) Hepatitis E virus infection. *Lancet* 355:578

- Purcell RH, Emerson SU (2001) Hepatitis E virus. In: Knipe DM, Howley PM (eds) *Fields virology*, vol 2, 4th edn. Lippincott Williams & Wilkins, Philadelphia, pp 3051–3061
- Purdy MA, Harrison TJ, Jameel S et al (2017) ICTV virus taxonomy profile: Hepeviridae. *J Gen Virol* 98:2645–2646
- Rein DB, Stevens GA, Theaker J et al (2012) The global burden of hepatitis E virus genotypes 1 and 2 in 2005. *Hepatology* 55:988–997
- Riveiro-Barciela M, Mínguez B, Gironés R et al (2015) Phylogenetic demonstration of hepatitis E infection transmitted by pork meat ingestion. *J Clin Gastroenterol* 49:165–168
- Rose N, Lunazzi A, Dorenlor V et al (2011) High prevalence of Hepatitis E virus in French domestic pigs. *Comp Immunol Microbiol Infect Dis* 34(5):419–427
- Rutjes SA, Lodder WJ, el L-VF e (2009) Sources of hepatitis E virus genotype 3 in The Netherlands. *Emerg Infect Dis* 15:381–387
- Rutjes SA, Lodder-Verschoor F, Lodder WJ et al (2010) Seroprevalence and molecular detection of hepatitis E virus in wild boar and red deer in The Netherlands. *J Virol Methods* 168 (1–2):197–206
- Salines M, Barnaud E, Andraud M et al (2015) Hepatitis E virus chronic infection of swine co-infected with porcine reproductive and respiratory syndrome virus. *Vet Res* 46:55
- Salines M, Andraud M, Rose N (2017) From the epidemiology of hepatitis E virus (HEV) within the swine reservoir to public health risk mitigation strategies: a comprehensive review. *Vet Res* 48:31
- Sato Y, Sato H, Naka K et al (2011) A nationwide survey of hepatitis E virus (HEV) infection in wild boars in Japan: identification of boar HEV strains of genotypes 3 and 4 and unrecognized genotypes. *Arch Virol* 156(8):1345–1358
- Sauleda S, Ong E, Bes M et al (2015) Seroprevalence of hepatitis E virus (HEV) and detection of HEV RNA with a transcription-mediated amplification assay in blood donors from Catalonia (Spain). *Transfusion* 55:972–979
- Schielke A, Sachs K, Lierz M et al (2009) Detection of hepatitis E virus in wild boars of rural and urban regions in Germany and whole genome characterization of an endemic strain. *Virol J* 6:58
- Schielke A, Ibrahim V, Czogiel I et al (2015) Hepatitis E virus antibody prevalence in hunters from a district in Central Germany, 2013: a cross-sectional study providing evidence for the benefit of protective gloves during disembowelling of wild boars. *BMC Infect Dis* 15:440
- Schlauder GG, Mushahwar IK (2001) Genetic heterogeneity of hepatitis E virus. *J Med Virol* 65:282–292
- Schlosser J, Vina-Rodriguez A, Fast C et al (2015) Chronically infected wild boar can transmit genotype 3 hepatitis E virus to domestic pigs. *Vet Microbiol* 180:15–21
- Serracca L, Battistini R, Rossini I et al (2015) Molecular investigation on the presence of hepatitis e virus (HEV) in wild game in North-Western Italy. *Food Environ Virol* 7:206–212
- Smith DB, Simmonds P, Izopet J et al (2016) Proposed reference sequences for hepatitis E virus subtypes. *J Gen Virol* 97:537–542
- Smith DB, Simmonds P, International Committee on Taxonomy of Viruses Hepeviridae Study Group et al (2014) Consensus proposals for classification of the family Hepeviridae. *J Gen Virol* 95:2223–2232
- Sonoda H, Abe M, Sugimoto T et al (2004) Prevalence of hepatitis E virus (HEV) Infection in wild boars and deer and genetic identification of a genotype 3 HEV from a boar in Japan. *J Clin Microbiol* 42(11):5371–5374
- Spahr C, Knauf-Witzens T, Vahlenkamp T et al (2018) Hepatitis E virus and related viruses in wild, domestic and zoo animals: a review. *Zoonoses Public Health* 65:11–29
- Sridhar S, Lau SK, Woo PC (2015) Hepatitis E: a disease of reemerging importance. *J Formos Med Assoc* 114(8):681–690
- Steyer A, Naglic T, Mocilnik T et al (2011) Hepatitis E virus in domestic pigs and surface waters in Slovenia: prevalence and molecular characterization of a novel genotype 3 lineage. *Infect Genet Evol* 11:1732–1737

- Stramer SL (2014) Current perspectives in transfusion-transmitted infectious diseases: emerging and re-emerging infections. *ISBT Sci Ser* 9:30–36
- Szabo K, Trojnar E, Anheyer-Behmenburg H et al (2015) Detection of hepatitis E virus RNA in raw sausages and liver sausages from retail in Germany using an optimized method. *Int J Food Microbiol* 215:149–156
- Takahashi M, Nishizawa T, Sato H et al (2011) Analysis of the full-length genome of a hepatitis E virus isolate obtained from a wild boar in Japan that is classifiable into a novel genotype. *J Gen Virol* 92(4):902–908
- Tam AW, Smith MM, Guerra ME et al (1991) Hepatitis E virus (HEV): molecular cloning and sequencing of the full-length viral genome. *Virology* 185:120–131
- Tei S, Kitajima N, Takahashi K et al (2003) Zoonotic transmission of hepatitis E virus from deer to human beings. *Lancet* 362(9381):371–373
- Thiry D, Rose N, Mauroy A et al (2016) Susceptibility of pigs to zoonotic hepatitis E virus genotype 3 isolated from a wild boar. *Transbound Emerg Dis* 64(5):1589–1597
- Thiry D, Mauroy A, Pavo N et al (2017a) Hepatitis E virus and related viruses in animals. *Transbound Emerg Dis* 64:37–52
- Thiry D, Mauroy A, Saegerman C (2017b) Belgian wildlife as potential zoonotic reservoir of hepatitis E virus. *Transbound Emerg Dis* 64(3):764–773
- Traoré KA, Ouoba JB, Huot N et al (2015) Hepatitis E virus exposure is increased in pork butchers from Burkina Faso. *Am J Trop Med Hyg* 93:1356–1359
- Tritz SE, Khounvisith V, Pommassichan S et al (2018) Evidence of increased hepatitis E virus exposure in Lao villagers with contact to ruminants. *Zoonoses Public Health* 65(6):690–701
- Tyrril SF, Quinton JN (2003) Overland flow transport of pathogens from agricultural land receiving faecal wastes. *J Appl Microbiol* 94:87S–93S
- Vulcano A, Angelucci M, Candelori E et al (2007) HEV prevalence in the general population and among workers at zoonotic risk in Latium region. *Ann Ig* 19:181–186
- Ward P, Muller P, Letellier A et al (2008) Molecular characterization of hepatitis E virus detected in swine farms in the province of Quebec. *Can J Vet Res* 72:27–31
- Wenzel JJ, Preiss J, Schemmerer M et al (2011) Detection of hepatitis E virus (HEV) from porcine livers in Southeastern Germany and high sequence homology to human HEV isolates. *J Clin Virol* 52:50–54
- Wilhelm B, Leblanc D, Houde A et al (2014) Survey of Canadian retail pork chops and pork livers for detection of hepatitis E virus, norovirus, and rotavirus using real time RT-PCR. *Int J Food Microbiol* 185:33–40
- Williams TP, Kasorndorkbua C, Halbur PG et al (2001) Evidence of extrahepatic sites of replication of the hepatitis E virus in a swine model. *J Clin Microbiol* 39:3040–3046
- Woo PC, Lau SK, Teng JL et al (2016) New hepatitis E virus. Genotype in Bactrian Camels, Xinjiang, China, 2013. *Emerg Infect Dis* 22:2219–2221
- Worm HC, Van der Poel WH, Brandstatter G (2002) Hepatitis E: an overview. *Microbes Infect* 4:657–666
- Wu JC, Chen CM, Chiang TY et al (2002) Spread of hepatitis E virus among different-aged pigs: two-year survey in Taiwan. *J Med Virol* 66:488–492
- Yang Y, Shi R, She R et al (2015) Fatal disease associated with Swine Hepatitis E virus and Porcine circovirus 2 co-infection in four weaned pigs in China. *BMC Vet Res* 11:77
- Yazaki Y, Mizuo H, Takahashi M et al (2003) Sporadic acute or fulminant hepatitis E in Hokkaido, Japan, may be food-borne, as suggested by the presence of hepatitis E virus in pig liver as food. *J Gen Virol* 84(9):2351–2357