

Emerging Concepts in Bacterial Taxonomy

Anusha Rai, Indu, N. Smita, G. Deepshikha, K. Gaurav, K. Dhanesh, G. Suresh, Ch. Sasikala, and Ch. V. Ramana

Abstract

Bacterial taxonomy has progressed over the years by virtue of the brisk and competent scientific developments. Ground-breaking molecular techniques have added an edge in the phylogenetic studies, resulting in the quality description of the taxa under studies. New avenues are rapidly developing whose validation has always been embraced and included, which will assist in resolution. It began with the simple application of objective procedures for classification, and now we have arrived at the genome-based taxonomy. This pedantic step has led to the meticulous examination and served to reconcile certain conflicts of the status of the taxa. This field is dynamic and is exploring more options like proteomics and metabolomics in gaining more insights into the lineal heritage. Even though there has been a significant change and addition, there is an ever-growing need for a comprehensive study, which would thread all the attributes together into one functional unit of classification. In this review, we examine the paradigm shift from traditional taxonomy to integrated taxonomy useful in the characterisation of bacteria which in addition aids in the identity of biotechnological targets.

Keywords

Bacterial taxonomy · Polyphasic · Phylogenomics · Integrated taxonomy · Average nucleotide sequence index (ANI)

A. Rai · Indu · N. Smita · G. Deepshikha · K. Gaurav · K. Dhanesh · G. Suresh · C. V. Ramana (\boxtimes) Department of Plant Sciences, School of Life Sciences, University of Hyderabad, Hyderabad, Telangana, India

C. Sasikala (🖂)

Centre for Environment, Institute of Science & Technology, JNT University Hyderabad, Hyderabad, Telangana, India

[©] Springer Nature Singapore Pte Ltd. 2019

T. Satyanarayana et al. (eds.), *Microbial Diversity in Ecosystem Sustainability and Biotechnological Applications*, https://doi.org/10.1007/978-981-13-8315-1_1

1.1 Introduction

The first description of the microorganism in the early 1670s by Robert Hooke and Antonie van Leeuwenhoek under a microscope was the beginning of the field of microbiology. Microbiology then had several dissections and bifurcations owing to novel implications, in various fields such as taxonomy, medicine, agriculture and environment. Like any other field in science, the in-depth and comprehensive knowledge requires the fundamental understanding of the subject under speculation. By implying the term 'fundamental' here, we allude to the application of 'taxonomy' in bacteriology. Bacterial taxonomy at the outset serves as a platform to figure out the basic characteristics of the species and then correlates it with its phylogenetic properties. It deals with the identification of isolates, their classification into the taxa and creating new ones (if novel) and their nomenclature which are carried out in accordance with the rules and regulations laid down in the Bacteriological Code 1990 (revised in 2008 by Parker et al. 2018). Classification and identification has been served best with polyphasic studies, whereas the nomenclature has always been advised to imply and reflect the genomic association. It is as essential as any other discipline of the biological sciences because it provides a scientific framework for the salient understanding of the bacterial species.

Taxonomy and systematics have been often used interchangeably, but there lies a thin line that governs a difference between the two. Taxonomy is based on practical classification dictated by theory, whereas systematics is the evolutionary study of the diverse group of organisms and its related taxa. Conventionally, bacterial taxonomy helps us to picturise the evolutionary history and its concordant relationship with the nearby organisms. Bacterial taxonomy is aimed at achieving authentic, reliable and reproducible knowledge ready for dissemination. As of today, the newness of the bacterial taxa is determined by the phylogenetic status of an isolate mostly using the 16S ribosomal RNA (rRNA) gene in supplementation of the phenotypic and chemotaxonomic properties of the culture. Although conscious efforts are meticulously made to describe and define taxa validly, still there are gaps which need to be filled with the help of upcoming techniques. The present system of bacterial taxonomy has progressed and developed due to inclusions in the light of various taxonomical methods. It has also led to the unearthing of new valuable taxa thus giving microbiology a new dimension.

1.2 Historical Developments

Bacterial taxonomy became the most sought-after subject in the field of microbiology after its inception in the early 1600s due to quantum surge in the discoveries and inventions, thus escalating the accumulation of knowledge. Understanding bacteria at the surface level became futile which genuinely leads to the importance in the definition of the taxa under study. This ultimately led to the development in the robust methods for the taxonomy in bacteriology. Even without the advent of molecular work, various attempts were made consciously for the classification

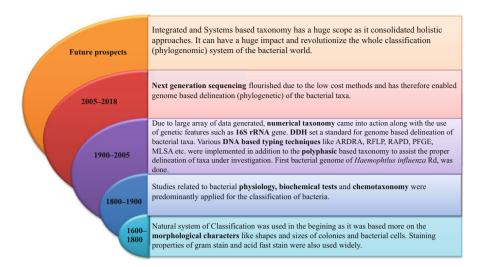


Fig 1.1 Diagrammatic representation of evolution of the classification systems extensively used in different era for appropriate delineation of bacterial taxa

although it was discreetly based on morphology, therefore always trying to systematise and correlate its phenetic with phylogenetic characteristics and its significance. Partitioning domain bacteria into various taxa levels proved beneficial and productive. Till date, various methods have been incorporated and improvised for the enhancement of the bacterial taxonomy. Direct or indirect contributions by scientists throughout the history of biological sciences made it possible, and therefore, we see that one event led to another, thereby causing the definitive birth of bacterial taxonomy (Fig. 1.1).

The development of bacterial taxonomy can be traced into different phases:

Phase I (1600–1900 AD)

Taxonomic study during this phase was based on simple biological observations and morphological descriptions. It was in this era that Antonie van Leeuwenhoek and Robert Hooke first observed 'animalcules' like structure under the single-lens microscope in the early 1670s. In the next decade of the 1800s, maximum contributions were made to aid the morphological studies. Muller and his contemporaries played a major role in promoting taxonomy as they inherently perceived the importance of assigning taxa genuinely owing to its towering application. Koch developed agar plate technique for the isolation of pure cultures, making the isolation of bacterial species convenient. In 1872, Ferdinand Cohn proposed that bacteria could also be designated as genus and species. Assorted methods like acidfast stain and Gram staining were developed for understanding the morphology by Paul Ehrlich and Christian Gram, respectively. Further, for the adept usage, petri plates were developed by R. J. Petri. These scientific landmarks improved the morphological studies and paved the ways for the future development in molecularbased taxonomy.

Phase II (1900-1980)

This phase saw the emergence of the application of biochemical and physiological properties for the taxa descriptions in accordance with the report presented by the Society of American Microbiologists (later changed into American Society for Microbiology) in 1923. This report also served as the ground for the publication of the first edition of Bergey's Manual of Determinative Bacteriology by David Hendricks Bergey. An exclusive journal was then established at the fifth International Congress for Microbiology in 1950 as the International Bulletin of Bacteriological Nomenclature and Taxonomy (IBBNT) and then later renamed the International Journal of Systematic Bacteriology (IJSB) in 1966. It was only in 2000, that it was called International Journal of Systematic and Evolutionary Microbiology solely for the description of valid taxa (Oren 2015). Numerical taxonomy was widely used owing to the large datasets arising from the characterisation. The coming years saw the emergence of semantides (or semantophoretic molecules which are biological macromolecules that carry phylogenetic information about evolutionary history) and its applications. DNA-DNA hybridisation was widely applied from the 1960s by various groups and is still considered as the golden standard for the species description. Polyphasic term was first coined by Colwell (1970), where the combined approach of phenotypic, genotypic and chemotypic characterisation was applied for the genus Vibrio. It was finally in 1977 that Carl Woese implied the use of ribosomal RNA (16S rRNA) sequence to identify Archaea and Bacteria.

Phase III (From 1980 Till Now)

This phase saw the dawn of the genomic era and its wide application in the demarcation of the taxa. The following years showed marked development in the genomic techniques for the better understanding of the genome content. Walter Gilbert and Frederick Sanger in 1977 made the sequencing less tedious by initiating methods for DNA amplification (Heather and Chain 2016). Various DNA typing methods were applied for the determination of inter- and intraspecies relatedness (Stackebrandt et al. 2002). The years after 1980s saw the emergence of techniques targeting fractions of whole genome. Randomly amplified polymorphic DNA (RAPD), restriction fragment length polymorphism (RFLP), pulse-field gel electrophoresis (PFGE), ribotyping and amplified ribosomal DNA (rDNA) restriction analysis (AFLP) were widely celebrated and employed for delineating bacterial taxa. Further in 1998, multilocus sequence typing approaches (Maiden et al. 1998) and coupled in silico bar coding (Shivali et al. 2012) were used for annotation of genomic relatedness.

It was in the year 1995 that the first bacterial genome of *Haemophilus influenzae* was sequenced by Fleischmann's group (Fleischmann et al. 1995). This was a breakthrough in the genomic era owing to significant developments contributed by

the molecular biologists. Thus it began the genomic era resulting in the higher resolution in the genome analysis. Focussed genome studies are now possible owing to the marked development in molecular techniques along with advancement in the bioinformatics tools. Recently, a digital protologue database was designed in order to make a reposition of all the newly described species or the emended taxa (Rosselló-Móra et al. 2017). According to the EzBioCloud database, 63,587 16S rRNA gene sequences have been deposited, and 92,802 genomes have been sequenced. Out of which, 23.04% (14650) are valid names, 0.81% (515) are invalid names, 0.46% (292) are *Candidatus* taxa, and the rest of 75.69% (48129) are the phylotypes of the total bacterial taxa (www.ezbiocloud.net/).

1.3 Importance of Bacterial Taxonomy in Applied Sciences

Taxonomy aids scientific communication as it allows the scientists to make predictions and frame hypotheses about the organisms. Often microbiologists use informal names like purple bacteria, sulphur bacteria and spirochetes, for example. However, in the scientific classification, each organism/species is assigned to a genus using a two-part binary name written in italic (underlined when handwritten) with a majuscule first letter with the exception of sobriquets for species and subspecies, e.g. Escherichia coli. The most widely accepted prokaryotic classification by microbiologist's community appeared in the early 1990s in the Bergey's Manual of Systematic Bacteriology as 'Taxonomic outline of the Prokaryotes' aiming to aid in the identification of species. For bacterial taxonomy, valid name must be in Latin or Neo-Latin using basic Latin letters only. Many species are named after person, either discoverer or a famous person in the field, for example, Shivajiella is named after Dr. Shivaji, an eminent Indian microbiologist who has made a significant contribution to our knowledge of heterotrophic bacteria from different predominantly cold habitats worldwide (Kumar et al. 2012). Many species (the specific epithet) are named after the place they are present or found, for example, the specific epithet of *Rhodomicrobium udaipurense* is named after the place (Udaipur) from where it was isolated (Ramana et al. 2013).

Usefulness of bacterial taxonomy to its core is evident by the heterogeneity in the metabolism of strain variation. For instance, from an evolutionary point of view, the species of the genus *Shigella (S. dysenteriae, S. flexneri, S. boydii, S. sonnei)* are strains of *Escherichia coli* (polyphyletic), but due to the difference in the genetic make-up of the pathogenic strains, they cause different medical conditions. *Escherichia coli* is a poorly classified species since some strains share only 20% of their genome. Being so diverse, it should be given a higher taxonomic rank. But, due to the maladies associated with the species and to avoid confusion in medical context, it remained unchanged. According to the National Centre for Biotechnology Information (NCBI), 3180 strains of *Escherichia coli* were reported (of which 2383 strains have their genome sequenced). Merely calling *E. coli* will not assure the organism under consideration. Thus, identity of the organism to its core (up to the

strain level) is crucial before the organism is taken up either to industry or for research purpose.

There are cases where investigators misidentified the species resulting in taxonomic errors in classification. For instance, genus Agrobacterium is nested under Rhizobium based on molecular data. Thus, Agrobacterium species are transferred to the Rhizobium genus resulting in Rhizobium radiobacter (formerly Agrobacterium tumefaciens), Rhizobium rhizogenes, Rhizobium rubi, Rhizobium undicola and Rhizobium vitis. But, due to the plant pathogenic nature of Agrobacterium spp., maintaining the genus Agrobacterium was proposed and later counterargued. Similarly, in the order Pseudomonadales (Gammaproteobacteria), the genera Azotobacter and Azomonas macrocytogenes (true members of the genus *Pseudomonas*) were misclassified due to nitrogen-fixing capabilities and the large size of the genus Pseudomonas thus rendering classification problematic. Also, the Bacillus species of the phylum Firmicutes, belonging to the 'Bacillus cereus group' (Bacillus anthracis, Bacillus thuringiensis, Bacillus weihenstephanensis, Bacillus mycoides, Bacillus pseudomycoides, Bacillus cereus and Bacillus medusa), have 99-100% 16S rRNA gene sequence similarity (97% being commonly cited acceptable species cut-off) and are polyphyletic but for medical reasons retained separate. Also, there are cases where investigators rectified the errors in taxonomic classification. For instance, Deinococcus radiodurans was originally classified as Micrococcus radiodurans by Anderson et al. (1956) due to its similarity with the genus Micrococcus, but later on, it was renamed Deinococcus radiodurans based on polyphasic data (Brooks and Murray 1981). Therefore, we see that precise bacterial identification is crucial for taking an organism for any study and thus avoiding any taxonomic error.

1.4 Prevailing Methods for Classification

Bacterial taxonomy first started with a vision of resolving its physical affiliation to its phylogeny in order to correlate its genomic imprints. The phenotype-based taxonomy led to the enormous addition of bacterial species because only few morphological factors were considered for the classification. More than 90% of all the species described in *Bergey's Manual* were subsequently reduced, and only species included on the approved lists of bacterial names became validly named species (Skerman et al. 1980; Garrity 2016). Major amendments occurred due to the use of DNA-DNA hybridisation (DDH) and 16S rRNA gene applications. Hence, transitioning from simple to holistic approaches, bacterial taxonomy has come a long way. It has still left a room for numerous improvisations owing to the gradual advancement in science.

Till date, polyphasic approach has been relevantly applied for the taxonomic purpose. It includes chemotaxonomic features (cell wall components, quinones, polar lipids, etc.), morphology, staining behaviour, culture characteristics (medium, temperature, incubation time, etc.) and genetic properties (G + C content, DDH value, 16S rRNA gene sequence identity with other closely related species) (Tindal et al. 2010). In some cases, DDH values have been strictly advised to strengthen taxa

delineation. According to the report by the Ad Hoc Committee of the International Committee for Systematic Bacteriology issued in 1897, the following parameters could be used for valid taxa description: (a) phenotypic (b) chemotaxonomic and (c) genotypic properties.

(a) Phenotypic

Phenotypic methods form the basis for formal description of taxa, from species and subspecies up to genus and family level (Garrity 2016). Traditional phenotypic tests used in classical microbiological laboratories include characteristics of organism on different growth substrates and growth range in different conditions such as pH, temperature, salinity and susceptibility towards different antibiotic stress (Prakash et al. 2007). Phenotypic data which are analysed by using computerassisted numerical comparison is known as numerical taxonomy. Phenotypic data matrices showing the degree of similarity between each pair of strains and cluster analysis resulting in dendrogram revealed a general picture of the phenotypic consistency of a particular group of strains. The advantage of phenotypic characterisation is that they can be easily observed, scored and measured without using any expensive technology. As the phenotypic characteristics depend on the conditional nature of gene expression, the same organism may show different phenotypic characteristics under various environmental conditions. Therefore, phenotypic data must be compared with a similar set of data from type strain of closely related organisms (Tindal et al. 2010).

(b) Chemotaxonomy

The term 'chemotaxonomy' refers to the application of analytical methods based on various chemical constituents of the cell to classify bacteria (Komagata and Suzuki 1987; Tindal et al. 2010; Sutcliffe 2015). It has enabled the establishment of specific chemical markers for proper classification and identification. The most commonly used chemical markers include cell wall/membrane component such as peptidoglycan, teichoic acid, polar lipid composition, relative ratios of fatty acid, sugars, lipopolysaccharide, isoprenoid quinones, carotenoids, chlorophyll composition, polyamines and fermentation products. Peptidoglycan can be an excellent marker for the taxonomic studies as it is present in most of the phyla, even including *Planctomycetes* and *Chlamydiae* according to recent studies (Pilhofer et al. 2013; Liechti et al. 2014) except for *Mycoplasma* (Rottem and Naot 1998).

Bacteria vary in their membrane lipid composition; therefore, polar lipids are considered for classification and identification of bacteria. Various chemical structures of fatty acids have been identified. The variability in chain length, double-bond position and substituent groups has proven to be very useful for the characterisation of bacterial taxa. Fatty acids are also the major constituents of lipids and lipopolysaccharides in microbial cells and have been therefore extensively used for taxonomic purposes. The process is termed the fatty acid methyl ester (FAME) analysis. Often fatty acids of variable length between 9 and 20 are considered for

classification (Sharmili and Ramasamy 2016). Hopanoids are pentacyclic triterpenoid sterol-like membrane lipids (Belin et al. 2018). Since hopanoids preserve source-specific information and can be linked with specific taxonomic group, physiological process, metabolic process or environmental condition, they can be used as a lipid biomarker (Cvejic et al. 2000; Blumenberg et al. 2012; Silipo et al. 2014). Hopanoids as chemotaxonomic markers were used in some of the recent studies (Tushar et al. 2015).

Isoprenoid quinones are components of cytoplasmic membrane of bacteria. Due to inconsistency of isoprenoid quinones along with difference in hydrogenation, saturation and side chain length, it acts as signature molecule for characterisation of bacteria at different taxonomic levels (Nowicka and Kruk 2010). Distribution of polyamines is universal in bacteria with significant quantitative and qualitative difference due to which they can be used as suitable chemotaxonomic markers. Depending on the group of organisms studied, polyamine patterning is being used to trace relatedness at and above the genus level and at the species level.

Whole-cell protein pattern can be analysed and compared for grouping of many closely related strains. Numerous studies have revealed a correlation between high similarity in whole-cell protein content and DNA-DNA hybridisation (Jarman et al. 2000). Identification is based on the comparison of the spectral database containing peptide mass fingerprints with the type strains by using the technique of MALDI-TOF (matrix assisted laser desorption/ionisation-time of flight) [Singhal et al. 2015]. It is applied to diagnose commensal bacterial species of *Enterococcus* sp. and Escherichia sp. by the determination of their unique spectra (Santos et al. 2015). Fourier transform infrared (FTIR) spectroscopy, on the other hand, uses the inherent property of the organism to produce specific metabolites to identify at the species and strain level (Naumann et al. 1991). When the whole microbial cells are excited by the absorption of the IR radiation, then it produces vibrational properties specific to the chemical bonds produced (Carlos et al. 2011). Metabolomic techniques like FTIR are often being used for the quick identification of bacteria on the basis of their particular metabolic fingerprints (Venkata Ramana et al. 2013). Biolog MicroPlates exploit the bacterial metabolism process for the utilisation of carbon sources. Species may be identified by specific colour change on the plate based on the metabolic fingerprint (Vehkala et al. 2015; Al-Dhabaan and Bakhali 2017). Lipidomes (Srinivas et al. 2016) and fermentomes (Sravanthi et al. 2016) are some of the chemomics used in recent bacterial taxonomy.

(c) Genotypic

The genotype-based methods have completely changed the scenario in the bacterial systematics world. It has finally assisted to draw lines between the various taxa levels. It mostly focusses on the retrieving of genomic information like DNA-DNA hybridisation, G + C contents, rRNA gene sequence analysis and DNA-based typing methods (DNA fingerprinting). DDH is required when a new taxon shares more than 97% 16S rRNA gene sequence similarity (Tindall et al. 2010). Value equal to or higher than 70% has been recommended for the definition of members of a species

(Wayne et al. 1987). The GC content is the calculated percentage of GC in the genome and therefore varies from one organism to another. Within prokaryotes, the G + C content varies between 20% and 80%. If the phylogenetic studies of an isolated strain reveal approximately 6% 16S rRNA gene sequence difference with its other closely related genus, then it can be recommended to represent the novel genus (Yarza et al. 2008).

It was considered that bacterial strains can be delineated with the data on 16S rRNA gene sequence analysis wherein the strains that show more than 3% sequence divergence are considered to represent different species (Rosselló-Móra and Amann 2001). However, with good quality and near full-length sequences, the value has been revised to 98.7–99% (Sackebrandt and Ebers 2006). A bacterial species can be properly defined as the group of strains sharing 70% or more DNA-DNA hybridisation with 5 °C or less ΔT_m value (T_m is the melting temperatures of the hybrid) among members of the group (Grimont 1981; Wayne et al. 1987). DDH is deemed necessary when strains share >98.7% 16S rRNA gene sequence identity. However, DDH has its own disadvantage because of which it cannot be applied to all the genera of prokaryotes. The difference in the sequences must be strongly supported by its distinctive characteristics. When a genetically close organism diverges in phenetic characteristics, then it can be ranked as a subspecies (Wayne et al. 1987).

The ribosomal locus such as the internal transcribed spacer region which is located between the 16S and the 23S rRNA genes has been scrutinised for the phylogenetic properties. Although this technique can outline the species/strain level but still at the lower level, it remains incongruous (Valera and Garcia-Martinez 2000). Multilocus sequence typing has been mostly used in epidemiology and pathological purposes but still generated its place in bacterial systematics. It has out sided the traditional procedure for determination of the genomic relatedness at inter- and intraspecific levels by sequence profiling of housekeeping genes (Maiden et al. 1998). The advantage of it not only lies in the application of the cultivable species, but also to those which are difficult to cultivate (Martens et al. 2008). Here, 6–11 housekeeping genes of the microbial species are profiled, which are around 470 bp long and stably selected. Amplified ribosomal DNA (rDNA) restriction analysis (ARDRA) can be used for the characterisation of bacterial isolates and has potential for analysing mixed bacteria communities. It is based on the principle of conserved restriction sites on the 16S rRNA which forms particular phylogenetic patterns specific to certain taxa (Abed 2008). The obtained banding pattern serves as a fingerprint for identification of respective bacteria. BOX-A1R-based repetitive extragenic palindromic-PCR (BOX-PCR) techniques play a vital role in the studies of microbial isolates from various environments.

The sequencing of the *Haemophilus influenzae* genome was a landmark in modern biology, as it marked the beginning of the genomic era. Next-generation sequencing (NGS) technologies introduced from 2005 provided a new platform resulting in a rapid increase in the prokaryotic genomes getting sequenced (Deurenberg et al. 2017; Besser et al. 2018). Genomic taxonomy is the newest

addition to the bacterial systematics world. Genome microbial taxonomy is paving a new path for the dynamic system-based classification.

1.5 Advanced Genome-Based Bacterial Taxonomy

Polyphasic taxonomy complemented along with the molecular fingerprinting techniques (AFLP, RFLP and others) served for the delineation of the taxa for a long time (Rademaker et al. 2000; Gurtler and Mayall 2001; Van Belkum et al. 2001). There are certain gaps generated in the definition of species which remain to be duly filled. Genome-based taxonomy is the missing link and can bridge the gap between the genome and phenotype-based classification. It has been well proven that genomic signatures can be tapped for the definition of bacterial species. A uniform definition of the bacterial species on the establishment of genomics would be to consider the strains from the same species.

With all the advantages that each technique has added in this field, the wholegenome sequencing will be the ultimate step for the resolution of taxonomic position of prokaryotes. The fine advancement in technology and the reduction in the cost of the whole-genome sequencing have led to the monumental shift in the sequencing of the genomes. There are thousands of whole-genome sequences of prokaryotes available, but still only a few hundred are of the type strains, therefore greatly restricting the use of genomic data for the comparative use in taxonomy (Chun and Rainey 2014). Therefore, the governing body has made mandatory in some cases for the whole-genomic sequencing where it would be used to break the incertitude situation in outlining its taxonomic rank (Konstantinidis and Tiedje 2005a). Although not mandatory for publication, the inclusion of this genome sequence data is highly recommended and will be expected to include new taxa descriptions submitted to *International Journal of Systematic and Evolutionary Microbiology*.

In the recent years, whole-genome sequencing has assisted in solving the complex taxonomical positions of certain species of *Vibrio*, *Mycoplasma*, *Xanthomonas* and *Prochlorococcus* (Jones et al. 2004; Thompson et al. 2013; Barak et al.2016). On the basis of genomic parameters like average amino acid sequence identity (AAI) and average nucleotide sequence identity (ANI), data has been applied for the bacterial species definition and classification (Qin et al. 2014). The ANI of common genes between strains being compared is especially closely correlated with the level of DDH, and a 95–96% ANI value can serve as a genomic measure for prokaryotic species delineation (Konstantinidis and Tiedje 2005b). At the genus level, the percentage of conserved proteins (POCPs) was used for the robust indexing of the genus boundary for the prokaryotic group. If all the pairwise POCP values are higher than 50%, then it could be defined as prokaryotic genus (Qin et al. 2014).

Vibrios belonging to the *Gammaproteobacteria* are found in the surrounding environment and also in close association as pathogen with the plant and animal. There are around 152 species of the genus, most of which are in specific hostpathogenic relationship whether it be human or animals and some in mutualistic relationship (http://www.bacterio.cict.fr/index.html). Often techniques like MLSA, DDH and $\Delta T_{\rm m}$ are often conventionally used for the delineation of the Vibrio species (Thompson et al. 2004). They are usually difficult to segregate into taxa owing to similar genomic and phenotypic characteristics. In a recent study, Thompson et al. (2009) restructured the Vibrio genus by a study comprising 43 genomes and observed that vibrios were distributed into three major groups or genera of Vibrio, *Photobacterium* and *Aliivibrio*. Critical genome analysis of vibrios has evidently revealed the description of two novel species which were closely related to Vibrio cholerae. Similarly, species closely related to *Mycoplasma* are very difficult to delineate on the basis of their 16S rRNA similarity and DDH. For example, Mycoplasma pneumoniae and Mycoplasma genitalium have a 16S rRNA similarity of about 98%. When critical analyses of 46 different genomes of Mycoplasmas were done, it was observed that Mycoplasma pneumonia and Mycoplasma genitalium had only 73% MLSA similarity, 67% AAI and 88 Karlin genomic signatures. With many more observations based on genome, Mycoplasma was seen to be paraphyletic (Thompson et al. 2013).

More evidences need to be generated using core genome for structuring the flexible positions of few species. Another interesting case is that of *Mycobacterium*. Till date, there are 193 species and 13 subspecies validly described (http://www. bacterio.net). Mycobacteriaceae consist of pathogenic as well as non-pathogenic species. Mycobacterium tuberculosis and Mycobacterium leprae and Mycobacterium abscessus are considered as pathogenic, whereas Mycobacterium smegmatis and Mycobacterium thermoresistibile are non-pathogenic (Brosch et al. 2000; Prasanna and Mehra 2013). Miscellaneous software for bioinformatics especially designed for the analysis of genomes makes the annotation of several mycobacterial species feasible. In the present scenario, the advancement has led to the collective information in relation to the evolutionary traits, sequence homology, conserved regions and gene ontology content (Malhotra et al. 2017). The study of comparative genomic analysis of 21 mycobacteria conducted by Zakham et al. (2012) revealed that 1250 Mycobacterium gene families were conserved across all species. The Mycobacterium pan-genome showed a total of 20,000 gene families (Zakham et al. 2012). Moreover, it was seen that the pathogenic ones had undergone genome reduction and gained defined group of genes for repair and protection (Wassenaa et al. 2009; Zakham et al. 2012). Mycobacterium leprae is the pathogenic one with the diminutive genome with 1600 genes and approximately 1300 pseudogenes (Singh and Cole 2011). Functional orthologs of these pseudogenes (>75%) were present in other mycobacterial species belonging to various protein groups (Malhotra et al. 2017; Muro et al. 2011). Therefore, tapping these variations for the genomic identity can be an excellent tool for the taxonomic purposes.

Coleman and Spain (2003) first described *Mycobacterium* strain JS623 from the environmental sample based on the identity value of 96.7% (421 bp) 16S rRNA being more similar to *Mycobacterium smegmatis*. It was still studied as the strain under the *Mycobacterium smegmatis* species until Ramasamy et al. (2014) increased the species limit of delimitation to 98.7%. Subsequently undertaken methodical gene and genome analyses showed that strain JS623 is a mycobacterium more related to

M. moriokaense than to *M. smegmatis* and indicate that it is not a member of this last species, as was previously believed (Garcia and Gola 2016). Therefore, JS623 was probably not a member of *M. smegmatis*.

We see that standard methods like 16S rRNA gene sequence analysis and DDH might not be superior in terms of establishing phylogenetic relationships. It has to be corroborated with other references generated from whole-genome sequencing. Genome-based taxonomy becomes very essential for the delineation of the closely related species in order to disclose the species-specific patterns. In terms of genomics, a collate prokaryotic species can be defined as the strains from the same species which share <10 in Karlin signatures (Karlin et al. 1997; Coenye and Vandamme 2004), > 95% AAI and ANI (Goris et al. 2007; Konstantinidis and Tiedje 2005a, b; Rohwer and Edwards 2002), > 95% identity based on multiple alignment genes (Thompson et al. 2008) and > 70% in silico genome-to-genome distance (Auch et al. 2010).

1.6 Future Prospectives

We have seen the expansion of bacterial taxonomy, mainly due to the contributions of more of accessible and unambiguous techniques. Further advent of molecular techniques added to their improved taxonomic studies. Now, we are in the transitional stages of omics wherein a lot of data generated shall be subdivided and specific markers shall be applied for the taxa studies. Already, huge genome data have been submitted in various databases. The Genomic Encyclopaedia of Bacteria and Archaea (GEBA) project is started by the DOE Joint Genome Institute in 2007 to pilot genome sequences of the type strains (Wu et al. 2009). This effort will definitely bridge in the gap which has arisen from the biased sequencing of only the physiologically advantageous ones. It has been suggested that time and again that the type strains of genomes can be used for comparative studies for taxonomical purposes (Chun and Rainey 2014). Whitman also emphasised on the future complementation of DNA and genome sequences to some extent to substitute the pure cultures as type materials to be deposited in culture collections (Whitman 2015). Moreover, the genomes reflect the biology of the organism and its evolutionary lineage pattern, therefore avoiding fabrication of redundant species. The expansion of the next-generation sequencing and its subsequent decrease in the price have resulted in the better understanding and scanning of gene families for fine delineation of the taxa. Dynamic approaches are being embraced for the accelerated taxonomical purposes of bacteria. Innovative technologies and study systems are underway for effective identification. The following systems are highly ambitious and visionary, thus enabling the filling of gaps and errors in the taxonomic studies. Advanced taxonomy can be summarised as follows:

(i) Integrated Taxonomy: Although polyphasic taxonomy has solved the problem of ambiguous ranking of the taxa by its methodical application of phylogeny, chemotaxonomy and phenotype-based studies, there remains a generous gap for upgradation. Polyphasic studies have added some dimension to taxonomy, but it has remained confined to certain aspects as it does not reflect the genomic content of the organism. Incorporation of various metabolomic and physiological affixes in polyphasic taxonomy has its own impediment owing to the variability caused by the environmental differences. All the more, the attributes considered for classification remain boxed and not interlinked inherently with the genomic content. Therefore, there is a certain need for an immediate classification system that would encompass genome data for the formulation of new taxa. The unparallel system for such kind of taxonomy would be that of 'integrated taxonomy'.

In the present context, scientists are exploring the aspects of genomics, transcriptomics and metabolomics to elucidate various processes and functions. Whole-genome sequencing plays a vital role in describing bacterial phylogeny through systems biology approach by their mechanistic genome annotation. Application of the same genomics for the translational-based studies would lead to the legitimate discerning of the relative species taxa based on the data generated. This type of integrated studies will help us assemble diverse information and put forth re-analysed phylogenetic history and novel biological proteins (Wu et al. 2009). Genome annotation of the taxa under study can ravel huge information owing to large generation of dataset. Translational and prediction-based inspection of the genome sequences can help us ascertain the production of possible novel metabolites and proteins specific to the taxa. Thus, all these compendious works shall complement 16S rRNA gene base for describing bacterial phylogeny with added values.

In the next-generation bacterial identification (NGBI), taxonomist would rely on both genomics and metabolomics for determining the microbial phylogeny. Postgenomic developments are, therefore useful in describing the phylogeny in a more determined way. Taxonomy, especially the bacterial one, has lawfully accepted the new advancing technologies. Therefore, polyphasic taxonomy can be clearly replaced by the integrated taxonomy.

(ii) Systems Taxonomy: Even with the integrated taxonomy providing the genome framework for classification, yet system-based taxonomy remains the advisable and prudent system for classification. It is an ultimate and ambitious goal to have inclusive taxonomic studies embracing genomics, proteomics and metabolomics along with various other components. It would in fact consider all the factors which influence the survival of the bacteria under study. It is not too far owing to the paramount advancement in the technologies for the development of system-based taxonomy wherein compendious interdisciplinary subjects are included for the comprehensive yet precise functional taxonomy.

Generally, it would be a holistic approach to identify and assign taxa to the bacteria under studies. Microbial metabolomics, proteomics and transcriptomics are variable and dynamic in nature under different environmental conditions, but still they play a vital role in bacterial physiological processes. So far, all these factors are

not considered for taxa delineation because of which we are failing to understand certain evolutionary divergences and convergences leading to speciation. Systembased taxonomy can be the missing link for critical studies for taxa delineation. Moreover, systems-based taxonomy constitutes all the consolidated component of systems biology that affects the organism's existence. It would as such consider all the parameters encompassing the taxa citing from a single cell to that of its complex interaction in the environments. Starting with the cell architecture (shape and size). its membrane components, biochemical activities, metataxon information (polar lipids, quinones, etc.) and physiological processes (growth mode, respiration, reproduction and energy metabolism) to its genomic fingerprints all would be considered. At another higher level, all the proteomic and metabolomic networking along with its ecological habitats and environmental factors (such as pH, temperature and salinity) shall be connected. Basically, these types of studies would network all the possible factors together into functional units for the proper understanding of the taxa under study and subsequently describe them into novel ones. Therefore, we see that this type of system-based taxonomy can solve major issues arising out of biased studies.

(iii) Virtual Taxa: It is evident from numerous studies that less than 1% of the actual microbial wealth is known, whereas the remaining part lies undiscovered. Metagenomics has also strengthened the same scientific belief of yet to be uncultivated microbial wealth. Therefore, forming ranks with the help of virtual taxa would be the most appropriate need of the hour. Particularly, it would further strengthen the taxonomy of the uncultured or the *Candidatus* species status; therefore, virtual taxa can be well correlated. It is also at par with the *Candidatus* species status as it is strenuous to cultivate them. Virtual taxonomy can be defined as the identification and classification of single-cell bacteria based on its genomic DNA and other cellular parameters under consideration. Single cell, screened using fluorescence-activated cell sorting (FACS), can be genome sequenced to frame the virtual taxonomy of the organism using bioinformatics tools. This tendency can be targeted to make use of *Candidatus* species for studying the community analysis.

As such genome sequences would serve as the main source of information in postulating and circumscribing species that are not available as pure cultures (Konstantinidis and Rosselló-Móra 2015). Based on the DNA sequences retrieved from the metagenomic studies, virtual taxonomy could be functional and practical in which prediction and conclusive studies based on the genome information about its cell structure, physiology and biological roles could be reported (Fig. 1.2). Combining and analysing the results thus obtained will fetch 'consensus taxonomy' of a microorganism. As a consequence, there is an immediate need of non-culture-based rapid method of identification utilising molecular approaches mainly centring DNA-based methods selecting taxa-specific loci. This propensity will foster a rapid and cost-effective method for bacterial identification in the coming years.

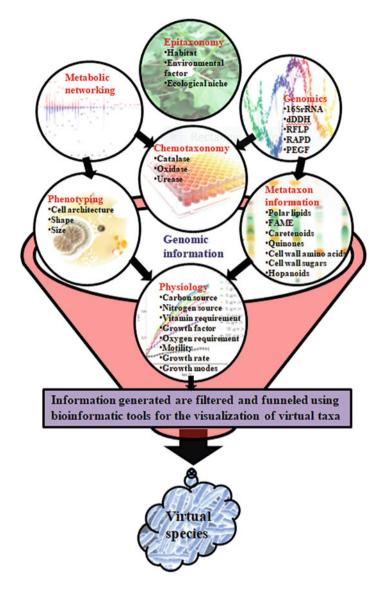


Fig. 1.2 Identification and classification of finely delineated virtual species by filtration of the epitaxonomic information and genomic information such as genomics, phenotyping, chemataxonomy, metataxon information, physiology and metabolic networking, using bioinformatic tools

1.7 Concluding Remarks

Since all the information applied in the taxonomic studies are based on assets and not just liability, integrated and system-based taxonomy has a good probability of expansive implementation in the future owing to its functionality. It may be a gradual change, but it surely helps in obtaining the bigger picture basically concatenating its genome to its state of being (phenotype) and also various environmental factors. Formulating virtual taxa related to taxonomy is on the other side a liability used only for aiding taxonomists to understand its inherent property of uncultivated state. It is a visualisation aid. Despite the fact that many methods have been realised and still being developed, there is still an undying need for the quick and rapid method of identification of bacteria.

Acknowledgements Anusha, Indu, Gaurav and Suresh thank DST, CSIR, DBT and UGC, Government of India, for the award of INSPIRE and SRF fellowships, respectively. Dhanesh is grateful to UGC for Dr. D.S. Kothari postdoctoral fellowship. C.V.R. thanks DBT, Government of India, for the award of TATA-Innovative Fellowship. Infrastructural facilities created under DST-FIST and UGC-SAP are also acknowledged.

References

- Abed RMM (2008) Nucleic acid-based techniques for studying diversity and activity of bacterial communities in oil-contaminated sediments. In: Handbook of environmental chemistry. Springer, Berlin/Heidelberg, pp 97–160
- Al-Dhabaan FAM, Bakhali AH (2017) Analysis of the bacterial strains using biolog plates in the contaminated soil from Riyadh community. Saudi J Biol Sci 24:901–906
- Anderson AW, Nordan HC, Cain RF, Parish G, Duggan D (1956) Studies on radio resistant micrococcus. I. The isolation, morphology, cultural characteristics and resistance to gamma radiation. Food Technol 10:575–577
- Auch AF, von Jan M, Klenk HP, Göker M (2010) Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison. Stand Genomic Sci 2:117–134
- Barak JD, Vancheva T, Lefeuvre P, Jones JB, Timilsina S, Minsavage GV, Vallad GE, Koebnik R (2016) Whole-genome sequences of *Xanthomonas euvesicatoria* strains clarify taxonomy and reveal a stepwise erosion of type 3 effectors. Front Plant Sci 7:1805
- Belin BJ, Busset N, Giraud E, Molinaro A, Silipo A, Newman DK (2018) Hopanoid lipids: from membranes to plant–bacteria interactions. Nat Rev Microbiol 16:304–315
- Besser J, Carleton HA, Gerner-Smidt P, Lindsey RL, Trees E (2018) Next-generation sequencing technologies and their application to the study and control of bacterial infections. Clin Microbiol Infect 24:335–341
- Blumenberg M, Thiel V, Riegel W, Kah LC, Reitner J (2012) Biomarkers of black shales formed by microbial mats, late Mesoproterozoic (1.1 Ga) Taoudeni Basin, Mauritania. Precambrian Res 196:113–127
- Brooks BW, Murray RGE (1981) Nomenclature for *Micrococcus radiodurans* and other radiationresistant Cocci: Deinococcaceae fam. nov. and *Deinococcus* gen. nov., including five species. Int J Syst Bacteriol 31:353–360
- Brosch R, Gordon SV, Eiglmeier K, Garnier T, Cole ST (2000) Comparative genomics of the leprosy and tubercle bacilli. Res Microbiol 151:135–142

- Carlos C, Maretto DA, Poppi RJ, Sato MI, Ottoboni LM (2011) Fourier transform infrared microspectroscopy as a bacterial source tracking tool to discriminate fecal *E. coli* strains. Microchem J 99:15–19
- Chun J, Rainey FA (2014) Integrating genomics into the taxonomy and systematics of the Bacteria and Archaea. Int J Syst Evol Microbiol 64:316–324
- Coenye T, Vandamme P (2004) Use of the genomic signature in bacterial classification and identification. Syst Appl Microbiol 27:175–185
- Coleman NV, Spain JC (2003) Distribution of the coenzyme M pathway of epoxide metabolism among ethene- and vinyl chloride-degrading *Mycobacterium* strains. Appl Environ Microbiol 69:6041–6046
- Colwell RR (1970) Polyphasic taxonomy of the genus Vibrio: numerical taxonomy of Vibrio cholerae, Vibrio parahaemolyticus and related Vibrio species. J Bacteriol 104:410–433
- Cvejic JH, Putra SR, El-Beltagy A, Hattori R, Hattori T, Rohmer M (2000) Bacterial triterpenoids of the hopane series as biomarkers for the chemotaxonomy of *Burkholderia*, *Pseudomonas* and *Ralstonia* spp. FEMS Microbiol Lett 183:295–299
- Deurenberg RH, Bathoorn E, Chlebowicz MA, Couto N, Ferdous M, García-Cobos S, Kooistra-Smid AM, Raangs EC, Rosema S, Veloo AC, Zhou K (2017) Application of next generation sequencing in clinical microbiology and infection prevention. J Biotechnol 243:16–24
- Fleischmann RD, Adams MD, White O, Clayton RA, Kirkness EF, Kerlavage AR, Bult CJ, Tomb JF, Dougherty BA, others (1995) Whole-genome random sequencing and assembly of *Haemophilus influenza* RD. Science 269:496–512
- Garcia MJ, Gola S (2016) Gene and whole genome analyses reveal that the mycobacterial strain JS623 is not a member of the species *Mycobacterium smegmatis*. Microbiol Biotechnol 9:269–274
- Garrity GM (2016) A genomics driven taxonomy of Bacteria and Archaea: are we there, yet? J Clin Microbiol 54:1956–1963
- Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM (2007) DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. Int J Syst Evol Microbiol 57:81–91
- Grimont PAD (1981) Use of DNA reassociation in bacterial classification. Can J Microbiol 34:541–546
- Gürtler V, Mayall BC (2001) Genomic approaches to typing, taxonomy and evolution of bacterial isolates. Int J Syst Evol Microbiol 51:3–16
- Heather J, Chain B (2016) The sequence of sequencers: the history of sequencing DNA. Genomics 107:1–8
- Jarman KH, Cebula ST, Saenz AJ, Petersen CE, Valentine NB, Kingsley MT, Wahl KL (2000) An algorithm for automated bacterial identification using matrix-assisted laser desorption/ionization mass spectrometry. Anal Chem 72:1217–1223
- Jones JB, Lacy GH, Bouzar H, Stall RE, Schaad NW (2004) Reclassification of the xanthomonads associated with bacterial spot disease of tomato and pepper. Syst Appl Microbiol 27:755–762
- Karlin S, Mrázek J, Campbell AM (1997) Compositional biases of bacterial genomes and evolutionary implications. J Bacteriol 179:3899–3913
- Komagata K, Suzuki KI (1987) Lipid and cell-wall analysis in bacterial systematics. Methods Microbiol 19:161–207
- Konstantinidis KT, Rosselló-Móra R (2015) Classifying uncultivated microbial majority: a place for metagenomics data in the Candidatus proposal. Syst Appl Microbiol 38:223–230
- Konstantinidis KT, Tiedje JM (2005a) Towards a genome-based taxonomy for prokaryotes. J Bacteriol 187:6258–6264
- Konstantinidis KT, Tiedje JM (2005b) Genomic insights that advance the species definition for prokaryotes. Proc Natl Acad Sci U S A 102:2567–2572
- Kumar A, Aravind R, Francis K, Bhumika V, Ritika C, Priyashanth P (2012) Shivajiella indica gen. nov., sp. nov., a marine bacterium of the family "Cyclobacteriaceae" with nitrate reducing activity. Syst Appl Microbiol 35:320–325

- Liechti GW, Kuru E, Hall E, Kalinda A, Brun YV, van Nieuwenhze M, Maurelli AT (2014) A new metabolic cell-wall labelling method reveals peptidoglycan in *Chlamydia trachomatis*. Nature 506:507
- Maiden MC, Bygraves JA, Feil E, Morelli G, Russell JE, Urwin R, Zhang Q, Zhou J, Zurth K, Caugant DA, Feavers IM (1998) Multilocus sequence typing: a portable approach to the identification of clones within populations of pathogenic organisms. Proc Natl Acad Sci U S A 95:3140–3145
- Malhotra S, Vedithi SC, Blundell TL (2017) Decoding the similarities and differences among mycobacterial species. PLoS Negl Trop Dis 11:e0005883
- Martens M, Dawyndt P, Coopman R, Gillis M, De Vos P, Willems A (2008) Advantages of multilocus sequence analysis for taxonomic studies: a case study using 10 housekeeping genes in the genus *Ensifer* (including former *Sinorhizobium*). Int J Syst Evol Microbiol 58:200–214
- Muro EM, Mah N, Moreno-Hagelsieb G, Andrade-Navarro MA (2011) The pseudogenes of *Mycobacterium leprae* reveal the functional relevance of gene order within operons. Nucleic Acids Res 39:1732–1738
- Naumann D, Helm D, Labischinski H (1991) Microbiological characterizations by FT-IR spectroscopy. Nature 351:81–82
- Nowicka B, Kruk J (2010) Occurrence, biosynthesis and function of isoprenoidquinones. Biochim Biophys Acta Bioenerg 1797:1587–1605
- Oren A (2015) 70th anniversary collection for the microbiology society: international journal of systematic and evolutionary microbiology. Int J Syst Bacteriol 65:4291–4293
- Parker CT, Tindall BT, Garrity GM (2018) International code of nomenclature of prokaryotes. Int J Syst Evol Microbiol. In Press
- Pilhofer M, Aistleitner K, Biboy J, Gray J, Kuru E, Hall E, Brun YV, van Nieuwenhze MS, Vollmer W, Horn M, Jensen GJ (2013) Discovery of chlamydial peptidoglycan reveals bacteria with murein sacculi but without Fts Z. Nat Commun 4:2856
- Prakash O, Verma M, Sharma P, Kumar M, Kumari K, Singh A, Kumari H, Jit S, Gupta SK, Khanna M, Lal R (2007) Polyphasic approach of bacterial classification-an overview of recent advances. Ind J Microbiol 47:98–108
- Prasanna AN, Mehra S (2013) Comparative phylogenomics of pathogenic and non-pathogenic *Mycobacterium*. PLoS One 8:e71248
- Qin QL, Xie BB, Zhang XY, Chen XL, Zhou BC, Zhou J, Oren A, Zhang YZ (2014) A proposed genus boundary for the prokaryotes based on genomic insights. J Bacteriol 196:2210–2215
- Rademaker JL, Hoste B, Louws FJ, Kersters K, Swings J, Vauterin L, Vauterin P, de Bruijn FJ (2000) Comparison of AFLP and rep-PCR genomic fingerprinting with DNA-DNA homology studies: *Xanthomonas* as a model system. Int J Syst Evol Microbiol 50:665–677
- Ramana VV, Raj PS, Tushar L, Sasikala C, Ramana CV (2013) *Rhodomicrobium udaipurense* sp. nov., a psychrotolerant, phototrophic alphaproteobacterium isolated from a freshwater stream. Int J Syst Evol Microbiol 63:2684–2689
- Ramasamy D, Mishra AK, Lagier JC, Padhmanabhan R, Rossi M, Sentausa E, Raoult D, Fournier PE (2014) A polyphasic strategy incorporating genomic data for the taxonomic description of novel bacterial species. Int J Syst Evol Microbiol 64:384–391
- Rohwer F, Edwards R (2002) The phage proteomic tree: a genome-based taxonomy for phage. J Bacteriol 184:4529–4535
- Rosselló-Móra R, Amann R (2001) The species concept for prokaryotes. FEMS Microbiol Rev 25:39–67
- Rosselló-Móra R, Trujillo ME, Sutcliffe IC (2017) Introducing a digital protologue: a timely move towards a database-driven systematics of archaea and bacteria. Antonie Van Leeuwenhoek 110:455–456
- Rottem S, Naot Y (1998) Subversion and exploitation of host cells by mycoplasmas. Trends Microbiol 6:436–440

- Santos T, Capelo JL, Santos HM, Oliveira I, Marinho C, Gonçalves A, Araújo JE, Poeta P, Igrejas G (2015) Use of MALDI-TOF mass spectrometry fingerprinting to characterize *Enterococcus* spp. and *Escherichia coli* isolates. J Proteome 127:321–331
- Sharmili AS, Ramasamy P (2016) Fatty Acid Methyl Ester (FAME) analysis of moderately thermophilic bacteria isolated from the coramandal coast, Chennai, Tamilnadu. Eur J Exp Biol 6:1–7
- Shivali K, Sasikala C, Ramana CV (2012) MLSA barcoding of Marichromatium spp. and reclassification of Marichromatium fluminis (Sucharita et al., 2010) as Phaeochromatium fluminis gen. nov.comb. nov. Syst Appl Microbiol 35:221–225
- Silipo A, Vitiello G, Gully D, Sturiale L, Chaintreuil C, Fardoux J, Gargani D, Lee HI, Kulkarni G, Busset N, Marchetti R (2014) Covalently linked hopanoid-lipid A improves outer-membrane resistance of a *Bradyrhizobium* symbiont of legumes. Nat Commun 5:5106
- Singh P, Cole ST (2011) Mycobacterium leprae: genes, pseudogenes and genetic diversity. Future Microbiol 6:57–71
- Singhal N, Kumar M, Kanaujia PK, Virdi JS (2015) MALDI-TOF mass spectrometry: an emerging technology for microbial identification and diagnosis. Front Microbiol 6:791
- Skerman VBD, McGowan V, Sneath PHA (1980) Approved lists of bacterial names. Int J Syst Bacteriol 30:225–420
- Sravanthi T, Tushar L, Sasikala C, Ramana CV (2016) Alkalispirochaeta cellulosivorans gen. nov., sp. nov., a cellulose-hydrolysing, alkaliphilic, halotolerant bacterium isolated from the gut of a wood-eating cockroach (Cryptocercus punctulatus), and reclassification of four species of Spirochaeta as a new combinations within Alkalispirochaeta gen. nov. Int J Syst Evol Microbiol 66:1612–1619
- Srinivas A, Divyasree B, Sasikala C, Tushar L, Dave B, Ramana CV (2016) Description of Jeotagalibacillus alkaliphilus sp. nov., isolated from a solar salt pan, and Jeotagalibacillus terrae sp. nov., a name to replace 'Jeotgalibacillus soli' Chen et al. 2010. Int J Syst Evol Microbiol 66:1–6
- Stackebrandt E, Ebers J (2006) Taxonomic parameters revisited: tarnished gold standards. Microbiol Today 33:152–155
- Stackebrandt E, Frederiksen W, Garrity GM, Grimont PA, Kämpfer P, Maiden MC, Nesme X, Rosselló-Móra R, Swings J, Trüper HG, Vauterin L (2002) Report of the ad hoc committee for the re-evaluation of the species definition in bacteriology. Int J Sys Evol Microbiol 52:1043–1047
- Sutcliffe IC (2015) Challenging the anthropocentric emphasis on phenotypic testing in prokaryotic species descriptions: rip it up and start again. Front Genet 6:218
- Thompson FL, Iida T, Swings J (2004) Biodiversity of vibrios. Microbiol Mol Biol Rev 68:403-431
- Thompson CC, Thompson FL, Vicente ACP (2008) Identification of vibrio cholerae and Vibrio mimicus by multilocus sequence analysis (MLSA). Int J Syst Evol Microbiol 58:617–621
- Thompson CC, Vicente ACP, Souza RC, Vasconcelos ATR, Vesth T, Alves N Jr, Ussery DW, Iida T, Thompson FL (2009) Genomic taxonomy of vibrios. BMC Evol Biol 9:258
- Thompson CC, Chimetto L, Edwards RA, Swings J, Stackebrandt E, Thompson FL (2013) Microbial genomic taxonomy. BMC Genomics 14(1):913
- Tindall BJ, Rossello-Mora R, Busse HJ, Ludwig W, Kämpfer P (2010) Notes on the characterization of prokaryote strains for taxonomic purposes. Int J Syst Evol Microbiol 60:249–266
- Tushar DL, Srinivas A, Sasikala C, Ramana CV (2015) Hopanoid inventory of *Rhodoplanes* spp. Arch Microbiol 197:861–867
- Valera RF, Garcia-Martinez J (2000) Spacers online. ASM News 66:712-713
- Van Belkum A, Struelens M, de Visser A, Verbrugh H, Tibayrenc M (2001) Role of genomic typing in taxonomy, evolutionary genetics, and microbial epidemiology. Clin Rev Microbiol 14:547–560
- Vehkala M, Shubin M, Connor TR, Thomson NR, Corander J (2015) Novel R pipeline for analyzing biolog phenotypic microarray data. PLoS One 10(3):e0118392

- Venkata Ramana V, Shalem Raj P, Tushar L, Sasikala C, Ramana CV (2013) *Rhodomicrobium udaipurense* sp. nov. a psychrotolerant, phototrophic alphaproteobacterium isolated from a freshwater stream. Int J Syst Evol Microbiol 63:2684–2689
- Wassenaar T, Bohlin J, Binnewies T, Ussery D (2009) Genome comparison of bacterial pathogens. Microb Pathog 6:1–20
- Wayne LG, Brenner DJ, Colwell RR, Grimont PAD, Kandler O, Krichevsky MI, Moore LH, Moore WEC, Murray RGE, Stackebrandt E, Starr MP, Trüper HG (1987) Report of the *ad hoc* committee on reconciliation of approaches to bacterial systematics. Int J Syst Bacteriol 37:463–464
- Whitman WB (2015) Genome sequences as the type material for taxonomic descriptions. Syst Appl Microbiol 38:217–222
- Wu D, Hugenholtz P, Mavromatis K, Pukall R, Dalin E, Ivanova NN, Kunin V, Goodwin L, Wu M, Tindall BJ, Hooper SD, Pati A, Lykidis A, Spring S, Anderson IJ, D'haeseleer P, Zemla A, Singer M, Lapidus A, Nolan M, Copeland A, Han C, Chen F, Cheng JF, Lucas S, Kerfeld C, Lang E, Gronow S, Chain P, Bruce D, Rubin EM, Kyrpides NC, Klenk HP, Eisen JA (2009) A phylogeny-driven genomic encyclopaedia of Bacteria and Archaea. Nature 462:1056–1060
- Yarza P, Richter M, Peplies J, Euzeby J, Amann R, Schleifer KH, Ludwig W, Glockner FO, Rosselló-Móra R (2008) The all-species living tree project: a 16S rRNA-based phylogenetic tree of all sequenced type strains. Syst Appl Microbiol 31:241–250
- Zakham F, Aouane O, Ussery D, Benjouad A, Ennaji MM (2012) Computational genomicsproteomics and phylogeny analysis of twenty one mycobacterial genomes (tuberculosis & non-tuberculosis strains). Microb Inf Exp 2:7