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# Cancer Chemoprevention by Dietary Polyphenols, Flavonoids, Terpenoids, and Saponins

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#### Abstract

Cancer chemoprevention invokes the adoption of natural or man-made agents for the inhibition, delay, or reversal of carcinogenesis before an invasion. It is predicted that roughly one-third of all cancer deaths might be prevented through proper dietary alteration. Chemopreventives should be defined by low toxicity in therapeutic drugs and the possibility of an oral administration. Several epidemiological studies and preclinical evidence indicate that various nutraceuticals and dietary supplements display chemopreventive properties, which is well supported by in vitro and animal studies. Diet derived compounds widely investigated for their chemopreventive activity mostly belong to a class of polyphenols, flavonoids, terpenoids, or saponins. A well-balanced diet is an excellent source of macronutrients, micronutrients, and phytochemicals and can diminish the risk of cancer as well as provide cancer preventive activity.

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### Keywords

 $Dietary \ phytochemicals \cdot Cancer \cdot Chemoprevention \cdot Epidemiology \cdot Epigenetics \cdot Apoptosis$ 

### 5.1 Cancer Chemoprevention by the Dietary Phytochemicals

Cancer ranks as one of the top leading causes of morbidity and mortality worldwide, with roughly 18.1 million new cancer cases and 9.6 million cancer deaths in 2018. It is also expected that the number of cases will increase by about 70% over the ensuing two decades reaching 22 million annual cases (Bray et al. 2018; McGuire 2016). Carcinogenesis is a mechanism by which a normal cell is transformed into a cancer cell. This is due to the mutation and epimutation of the genetic material of normal cells, which agitates the harmony between proliferation and apoptotic cell death. This causes uncontrolled cell proliferation and the formation of cancer. Chemoprevention is action taken to thoroughly cut-off or lower the chance of getting cancer by actively intervening in the course of carcinogenesis (Zhang et al. 2017; Al Rabadi and Bergan 2017). Scientists are investigating many different ways to support the prevention of cancer, including the following:

- Avoiding or controlling cancer causing things.
- · Managing changes in diet and lifestyle.
- Early detection of precancerous conditions that may lead to cancer.
- Chemoprevention (medicine to stop or reduce cancer conditions from the initial stage).
- Risk-reducing surgery.

An abundance of data suggests that lifestyle factors, along with exposure to chemical carcinogens, diet, and lack of physical activity play a pivotal role in the advancement of common cancers. It has been predicted that appropriate lifestyle modifications could prevent more than two-thirds of human cancers.

Diet is closely linked to the incidence and prevention of different cancer types and dietary behavior has been diagnosed as one of the most significant modifiable lifestyle impetus of cancer risk. Human cancer deaths attributable to diet make up nearly 10–70% (average 35%). Therefore diet, together with a healthy lifestyle, can reduce cancer incidence by 30–40%. In fact, most authors agree that there is consistent epidemiological evidence to suggest that a diet rich in fruits or vegetables significantly rolls back the risk of certain disorders, such as cancer and cardiovascular diseases (De Stefani et al. 2000).

In consequence, several international organizations, like the World Cancer Research Fund (WCRF), American Institute for Cancer Research (AICR), and other various cancer research foundations, endorsed a boost in the ingestion of certain fruits, vegetables, and grains as their incorporation in diet is associated with a reduced risk for the spreading of certain tumors and cancers (Mosby et al. 2012; Gapstur et al.

2018). The National Cancer Institute (NCI) has picked out about 35 plant-based foods with cancer-preventive properties. Foods and herbs possessing these qualities include garlic, cabbage, soybeans, licorice root, ginger, and the umbelliferous vege-tables (including celery, carrots, coriander, parsnips, and parsley). Additional foods with anticancer activity include citrus, onions, flax, turmeric, cruciferous vegetables (Brussels sprouts, broccoli, cabbage, and cauliflower), sweet peppers and tomatoes, brown rice, oats, barley, whole wheat, various herbs, such as rosemary, mints, thyme, sage, oregano, and basil, cantaloupe, cucumber, and berries (Wiseman 2018; Surh 2003). Plenty of secondary plant products, such as polyphenols (Thomasset et al. 2007), terpenoids (Rabi and Gupta 2008), saponins (Raju and Mehta 2009), flavonoids (Galati and O'Brien 2004), carotenoids (Tanaka et al. 2012). etc., which are substantial constituents of our daily food, have thus transformed themselves from being considered as non-nutritive constituents to possibly cancer preventive ones.

# 5.2 Occurrence of Polyphenols, Flavonoids, Terpenoids, and Saponins in Diet

Dietary phytochemicals are divided into four main classes according to their chemical structures: polyphenols, terpenoids, alkaloids, and sulfur compounds. Among them, dietary polyphenols and flavonoids are the rich antioxidants in human diets (Table 5.1). They are further subdivided into groups based on the number of

Subclass	Compounds	Primary source	
Flavonols	Quercetin, myricetin, kaempferol, rutin, isorhamnetin	Vegetables: capers, chives, celery, onions, red onions, lettuce, dock leaves, broccoli, hartwort leaves, kale	
		Fruits: apricots, apples, grapes, plums, bilberries, blueberries, blackberries, cranberries, olive elderberries, black currant juice, currants, cherries, apple juice	
		Drink: red wine, tea (green and black)	
Flavanones	Hesperin, hesperidin, naringin, eriodictyol, naringenin	Citrus fruits and juices: orange, orange juice, lemon, lemon juice, lime juice, grapefruit, tangerine juice	
Flavan-3-ols	Catechin, epicatechin, galloylated derivatives	Tea, apple, plums, cranberries, berries, chocolate	
Flavones	Luteolin, apigenin	Fruits: olives, celery	
		Vegetables: hot peppers, fresh parsley, celery hearts	
		Spices and herbs: oregano, dry parsley, rosemary, thyme	
Anthocyanins	Cyanidin, delphinidin, malvidin, pelargonidin, peonidin, petunidin (mostly as glycosides)	Fruits: Cherries, blackberries, black currants, blueberries, elderberries, black grape, strawberries, plums, cranberry, raspberry, pomegranate juice	

Table 5.1 Dietary sources of polyphenols



Fig. 5.1 Chemical structure of flavones and their dietary sources



Fig. 5.2 Chemical structure of flavonols and their dietary sources

phenolic rings that link the structural elements: (1) The phenolic acids, which have subclasses derived from different hydroxybenzoic acids, such as gallic acid and hydroxycinnamic acids, containing ferulic, caffeic, and coumaric acids; (2) the giant flavonoid subclass, which consists of the flavones (Fig. 5.1), flavonols (Fig. 5.2), flavanones (Fig. 5.3), flavanols (Fig. 5.4), and anthocyanidins (Fig. 5.5); (3) the stilbenes; and (4) the lignans and their polymers (Lin et al. 2016).

The richest flavonoids in the diet are flavanols (catechins plus proanthocyanidins), anthocyanins, and their oxidation analogues. The leading dietary sources of polyphenols include some common fruits and beverages (tea, coffee, fruit juice, wine, beer, and chocolate) and, to a lesser extent, dries legumes, vegetables, and cereals as shown in Table 5.1 (Bhagwat et al. 2013).

Terpenoids (Figs. 5.6 and 5.7) and Saponins (Fig. 5.8) occur in a wide range of plants but only a few of them are frequently used as food by humans. The more regularly consumed ones are spinach, soybeans, chickpeas, and peanuts. Many different saponins can exist within a sole plant species. Common dietary sources of saponins are soybeans (5.6%), lentil (4%), broad bean (3.7%), chickpeas (3.6%), garden peas (2.5%), and lucerne/alfalfa (2–3%) (Guclu-Ustundag and Mazza 2007).



Fig. 5.3 Chemical structure of flavanones and their dietary sources

# 5.3 Dietary Phytochemicals and Cancer

### 5.3.1 Epidemiological Evidence

Epidemiological studies and systematic analyses have shown the response of diet on health, and the relation between the utilization of certain foods and a marked down risk of some chronic diseases like cancer. In this view, many exercises have proven the potential of dietary phytochemicals as anticarcinogenic agents during study using different cell lines, animal models, and human epidemiological data (Scott et al. 2009; Kapinova et al. 2018). Dietary phytochemicals may inhibit numerous stages in the carcinogenesis mechanism and hence prevent or hold up tumor development. Although induction of apoptosis looks to be rather specific for the cancer cells, it may perhaps be mentioned that certain human studies have exposed no useful effects (Table 5.2). The Korean Population Cohort Study showed that a higher intake of flavonols and flavan-3-ols can slow down the risk of colorectal cancer (Cho et al. 2017). In a Netherlands Cohort Study involving 20,852 people, decreased colorectal cancer risk in normal wt. women was observed on consumption of a flavonol and catechin rich diet (Simons et al. 2009). Similarly, a decreased risk for esophageal, gastric, and prostate cancers have been reported after the consumption of anthocyanidin (Petrick et al. 2015). Also several epidemiological studies showed the pull down of different types of cancer risk after consumption of quercetin, myricetin, catechins, green tea, soy and many more dietry phytochemicals.

In summary, epidemiological and nutritional intervention studies data suggest that high consumption of dietary foods rich in polyphenols, flavonoids, terpenoids, and saponins may shorten the flourishing risk of several types of cancers (breast, colon, rectal, gastric, pancreatic, lung, ovarian, and prostatic cancer). This promising finding has motivated scientists to explore the molecular mechanisms involved in the antitumor actions in order to validate the value for cancer treatment. Various in vitro and in vivo investigations have provided broad evidence to prevent carcinogenesis and to suppress tumorigenesis over different molecular mechanisms.





Barley



Cocoa beans



Green tea



Acacia Catechu

Fig. 5.4 Chemical structure of flavan-3-ols and their dietary sources

# 5.3.2 Dietary Phytochemicals Effects in Xenograft Models

The in vivo effect of dietary phytochemicals has been studied using subcutaneous xenografts in mice. A nude mouse model of breast cancer xenografts can reduce the primary tumor growth using combined treatment with resveratrol, quercetin, and catechin (Schlachterman et al. 2008). Anthocyanidins (cyanidin, malvidin, petunidin, peonidin, and delphinidin) at 1.5 mg/mouse inevitably inhibit the development



Fig. 5.5 Chemical structure of anthocyanidins and their dietary sources



Fig. 5.6 Common dietary sources of terpenoids

of H1299 xenografts in nude mice. Cyanidin, the most active anthocyanidin, reduced the growth by  $\approx$ 60% and inhibited the growth of xenografts in nude mice (Liu et al. 2018). Significant reduction of tumor growth in a T24 bladder cancer xenograft model has been observed after treatment with the flavonol myricetin (Sun et al. 2012). Similarly, hepatocellar carcinoma xenograft growth is inhibited by flavone apigenin in nude mice (Gao et al. 2018).



Fig. 5.7 Structure of some common dietary terpenoids



Fig. 5.8 Common dietary sources of saponins

Dietary saponins, such as hederagenin, and the triterpene oleanolic acid also inhibit the tumor growth in a mouse xenograft model (Kim et al. 2017; Niu et al. 2018). Similarly, dietary diterpene carnosol (Johnson et al. 2010), monoterpene geraniol (Kim et al. 2012), and triteppene betulinic acid suppress cancer cell growth in xenograft tumor models (Cai et al. 2018).

		Sample	
	Effect	size	References
Flavonoids	Dropped off cancer risk in entire	9959 men	Knekt et al.
	sites combined		(1997)
	Deteriorated cancer risk in the	540	De Stefani et al.
	oral cavity, larynx, pharynx and	people	(1999)
	esophagus		
Quercetin, onions, white	Decreased recurrence of lung	582	Le Marchand
grapes	cancer	people	et al. (2000)
Quercetin	Decreased incidence of lung	10,054	Knekt et al.
	cancer	men	(2002)
	Decreased incidence of colon	264	Kyle et al.
	cancer	people	(2010)
Quercetin, kaempferol	Decreased risk of gastric cancer	354	Garcia-Closas
		people	et al. (1999)
Catechins	Decreased incidence of epithelial	939 men	Arts et al.
	cancer		(2001)
Tea	Decreased risk of colon cancer	12,170	Su and Arab
		people	(2002)
Green tea	Slow down risk of prostate cancer	49,920	Kurahashi et al.
		men	(2008)
	Diminished risk of recurrence	472	Inoue et al.
	breast cancer and metastasis	women	(2001)
Flavonoid intake and	Declined risk of prostate cancer	58,279	Geybels et al.
black tea		men	(2013)
Soy	Decreased risk of lung cancer	999 men	Wakai et al.
			(1999)
	Decreased risk of breast cancer	34,759	Key et al.
		women	(1999)
Anthocyanidin	Reduced risk of oesophageal and	615	Petrick et al.
	gastric cancer	people	(2015)
Flavonol and catechin	Decreased colorectal cancer risk	120, 852	Simons et al.
	in normal wt. women	women	(2009)
	Reduced risk of prostate cancer	118	Geybels et al.
		people	(2013)
Epicatechin, catechin,	Decreased lung cancer risk	558	Cui et al. (2008)
quercetin and kaempferol	among tobacco smokers	neonle	

**Table 5.2** Epidemiological studies: corporation between flavonoids or foods affluent in phenolic compounds and cancer

### 5.3.3 Dietary Phytochemicals Effects on Apoptosis

Programmed cell death is defined by morphological and biochemical modifications in cells (e.g., DNA fragmentation) (Debatin 2004). Dietary phytochemicals induce apoptosis in various cancer cell lines, namely human breast cancer (Valcic et al. 1996), lung cancer (Yang et al. 1998), gastric cancer (Horie et al. 2005), colon cancer (Tan et al. 2000), and prostate cancer (Brusselmans et al. 2003). Recent investigations have shown that the dietary flavonol quercetin induces apoptotic cell death in various types of cancers, such as leukemia (Chen and Jiu-Hong 2005; Mertens-Talcott and Percival 2005), prostate cancer (Huynh et al. 2003), breast cancer

(Hakimuddin et al. 2004), lung cancer (Nguyen 2003) and hepatoma (Chi et al. 1997). Moreover, these flavonols have also been shown to promote morphological mutation and DNA cleavage in leukemia (Csokay et al. 2005) and rat pancreatic carcinoma cells (BSp73AS) (Mouria et al. 2002).

Genistein has been found to restrain the growth of various cancer cells by the modulation of genes that are related to the control of apoptosis or to another mechanism like cell growth or signal transduction pathways, after all this isoflavonoid is a magnificent inhibitor of protein tyrosine kinases (Lian et al. 1999). It can promote apoptosis in prostate cancer (Kumi-Diaka et al. 2000), breast cancer (Li et al. 1999), head and neck squamous cell carcinoma (Alhasan et al. 1999), lung cancer cells (Lian et al. 1999), and stomach cancer cells (Yanagihara et al. 1993).

Kaempferol exerts a direct effect on the apoptosis extrinsic pathway, which is based on the presence of death receptors on the cell surface able to recognize death inducing substances. These death receptors comprise tumor necrosis factor alpha (TNF- $\alpha$ ), FAS, and TRAIL (Lee et al. 2014).

Anthocyanidins have been found to activate morphological change and DNA fragmentation in hepatoma cells (Shih et al. 2005). Cell apoptosis was detected by DNA agarose gel electrophoresis when lung cancer cells NCI-H460 were treated with anthocyanidin (Zhang et al. 2005). Theaflavin digallate and epigallocatechin inhibited growth and promoted apoptosis in COLO 320DM cells (Hsu et al. 2012). Morphological observation of the tissue displayed apoptotic bodies in treated human stomach cancer KATO III cells (Hibasami et al. 1998). Viability, apoptosis, and DNA fragmentation assay indicated that the merger of EGCG and bleomycin potentiated apoptosis (Bimonte et al. 2015). Epigallocatechin-3-O-gallate provoked dosedependent cell propagation inhibition, cell cycle detention at the G0/G1 stage, and DNA cleavage in HT-1080 cells, suggesting the induction of apoptosis (Lee et al. 2011). Synergistic apoptosis of HCT 15, HCT 116, as well as Hep G-2 cells by curcumin and catechin have been observed efficiently (Alam et al. 2018). Anthocynin rich bilberry extract has been shown to induce apoptosis in HL60 cells and nucleosomal DNA fragmentation (Katsube et al. 2003). Triterpenes oleanolic acid and ursolic acid induced apoptosis in four cancer cell lines of human liver. Completion of apoptosis was proved microscopically by observing escalation in plasma membrane permeability and detecting the fragmentation of DNA (Shyu et al. 2010).

### 5.3.4 Epigenetic Markers Effects

Dietary factors play a pivotal role in many natural biological courses of action and are also convoluted in the surveillance of pathological breakthroughs. Environmental and dietary circumstances can influence diseases linked to genetic and epigenetic modifications. Recently, an increasing number of nutritional components that have an inherent epigenetic activity have been identified. These micronutrients are able to control gene expression by carrying out an inheritable DNA (or DNA-associated proteins) modification without altering the DNA sequence. The posttranslational modification of histone proteins is the most well-known epigenetic mechanism by histone deacetylases (HDACs). Synthetic HDACs cause harmful side effects like atrial fibrillation, questioning their applicability. Therefore, the discovery of new HDACs inhibitors (HDACIs) is of great interest as potential anticancer drugs (Berger et al. 2013).

Dietary polyphenols and flavonoids have a custodial role against diseases and have found an important place in cancer prevention (Yang et al. 1998; Tan et al. 2000). In fact, various mechanisms have been found that aid in demonstration of the preventive nature of polyphenols, along with their ability to amend the epigenome by chromatin remodeling or by reactivating silenced genes in cancer cells (Bag and Bag 2018). Their chemopreventive potential can be defined by their ability to restrain DNMTs and also act as histone modifiers. The epigenome of cancer cells could be changed significantly by both of these properties, and they are viewed as interesting possibilities for anticancer therapeutics (Fig. 5.9).

The DNMT inhibitory activity of a green tea catechin and epigallocatechin 3-gallate (EGCG) was described by Morris et al. (2016). More than 50% of effective compounds in green tea are EGCG. Flavones apigenin and luteolin, flavanone hesperetin, and anthocyanidin cyanidine inhibit DNMT activity when tested in vitro. The flavone apigenin shows chemopreventive properties against prostate cancer by inhibiting HDAC (Ganai 2017). The dietary flavone luteolin influenced apoptosis of HL-60 cells, is associated with c-Jun activation, and expressed the histone H3 acetylation-mediated by Fas/FasL (Wang et al. 2018). It also cut down protein levels and the enzyme actions of epigenetic modifying enzymes, such as DNA methyltransferases (DNMTs) and histone deacetylases (HDACs), in HCT116 cells (Zuo et al. 2018). Delphinidin, a dominant anthocyanidin compound found in diversified fruits, promotes p53-mediated apoptosis by abolishing HDAC activity and stimulating p53 acetylation in human prostate cancer LNCaP cells (Jeong et al. 2016).



Fig. 5.9 Epigenetic modifications by dietary phytochemicals. (Idea adapted from Szyf 2015)

### 5.3.5 Importance of Dietary Phytochemicals in Co-therapy

Used in combination, dietary phytochemicals also have great potential to enhance the therapeutic effects of antitumor drugs, a practice known as co-therapy. The combination of the flavonol kaempferol with classical chemotherapeutic agents results in greater cytotoxic effects than those achieved by each of them separately (Luo et al. 2010). It is also able to sensitize the cytotoxic effects of 5-fluorouracil to the tumor cells, cytarabine (Mohan et al. 2013) and doxorubicin (Sharma et al. 2007).

Combination of autophagy inhibitors with the flavone apigenin inhibits the cell proliferation and induces autophagy by way of suppressing the PI3K/Akt/mTOR pathway (Yang et al. 2018).

A synergistic cytotoxic effect by theaflavin-3,3'-digallate, a black tea polyphenol, and cisplatin (CDDP) was shown in cisplatin resistant ovarian cancer cells A2780/CP70 and OVCAR3 (Pan et al. 2018).

Another polyphenol, epigallocatechin-3-gallate, found in tea combined with cisplatin significantly shortened the size of the tumor (Zhou et al. 2014). It also potentiated the effect of adriamycin in CaEs-17 cells (Fantini et al. 2015), fluorouracil in Hep G2 cells (Yang et al. 2012), and ponatinib in chronic myeloid leukemia cells (Goker et al. 2014). Combination of capecitabine with (-)-epigallocatechin-3-gallate inhibits tumor growth and angiogenesis with gastric cancer xenografts in nude mice (Wu et al. 2012). Green tea catechins augmented the antitumor properties of doxorubicin for chemoresistant liver cancer in a mouse model (Liang et al. 2010).

The anthocynidin delphinidin (DPN) in combination with 5-aza-2-deoxycytidine (AzaC) showed the highest inhibition of cell growth in human glioblastoma LN18 and U87MG cells (Chakrabarti and Ray 2015). Anthocyanins also potentiated the activity of trastuzumab in human epidermal growth factor receptor 2-positive breast cancer cells in vitro and in vivo (Liu et al. 2014).

Luteolin with lapatinib inhibited the growth of breast cancer cells (Zhang et al. 2017) and the doxorubicin-induced autophagy in human osteosarcoma U2OS cells (Zhang et al. 2015). The combination of the oncolytic adenovirus CD55-TRAIL with luteolin significantly decreased cytotoxicity in lung/bronchial normal epithelial cells compared with single treatment (Xiao et al. 2017). Co-treatment of tamoxifen and naringenin could inhibit cell proliferation more effectively in ER+breast cancer cells (Xu et al. 2018). Combination of myricetin with 5-fluorouracil chemotherapy can enhance tumor chemosensitivity of esophageal cancer EC9706 cells, and hence myricetin could be a potential chemosensitizer for esophageal cancer therapy (Wang et al. 2014).

Combination of afromosin with soyasaponin I enhanced their antitumor promoting activity. Consequently, many active compounds were found that might be valuable chemopreventive agents (Konoshima et al. 1992).

Limonene enhances the antitumor effect of docetaxel against prostate cancer cells without being toxic to normal prostate epithelial cells (Rabi and Bishayee 2009). Geraniol in combination with gemcitabine induced BXPC-3 cell apoptosis (Jin et al. 2013). Co-treatment 5-fluorouracil with triterpenoid lupeol induced

apoptosis by upregulating the expressions of Bax and p53 and downregulating the expressions of survivin and Bcl-2 (Liu et al. 2016).

# 5.4 Concluding Remarks

In the past few decades, several studies have been performed that support the concept of cancer chemoprevention dietary polyphenols, flavonoids, terpenoids, and saponins. Several epidemiological studies have corroborated that dietary consumptions have a huge impact on cancer prevalence. Owing to these encouraging observations, research efforts all across the globe have focused on identifying, characterizing, and providing the scientific basis behind the chemopreventive properties of dietary supplements. The results have shown that fruits and vegetables represent an untapped reservoir of various nutritive and nonnutritive phytochemicals that when incorporated into a healthy lifestyle can be a very useful step toward cancer chemopreventive.

### References

- Al Rabadi L, Bergan R (2017) A way forward for cancer chemoprevention: think local. Cancer Prev Res (Phila) 10(1):14–35. https://doi.org/10.1158/1940-6207.CAPR-16-0194
- Alam MN, Almoyad M, Huq F (2018) Polyphenols in colorectal cancer: current state of knowledge including clinical trials and molecular mechanism of action. Biomed Res Int 2018:4154185. https://doi.org/10.1155/2018/4154185
- Alhasan SA, Pietrasczkiwicz H, Alonso MD, Ensley J, Sarkar FH (1999) Genistein-induced cell cycle arrest and apoptosis in a head and neck squamous cell carcinoma cell line. Nutr Cancer 34(1):12–19
- Arts IC, Hollman PC, Bueno De Mesquita HB, Feskens EJ, Kromhout D (2001) Dietary catechins and epithelial cancer incidence: the Zutphen elderly study. Int J Cancer 92(2):298–302
- Bag A, Bag N (2018) Tea polyphenols and prevention of epigenetic aberrations in cancer. J Nat Sci Biol Med 9(1):2–5. https://doi.org/10.4103/jnsbm.JNSBM\_46\_17
- Bhagwat S, Haytowitz DB, Wasswa-Kintu SI, Holden JM (2013) USDA develops a database for flavonoids to assess dietary intakes. Procedia Food Science 2:81–86
- Berger A, Venturelli S, Kallnischkies M, Bocker A, Busch C, Weiland T, Noor S, Leischner C, Weiss TS, Lauer UM, Bischoff SC, Bitzer M (2013) Kaempferol, a new nutrition-derived pan-inhibitor of human histone deacetylases. J Nutr Biochem 24(6):977–985. https://doi. org/10.1016/j.jnutbio.2012.07.001
- Bimonte S, Leongito M, Barbieri A, Del Vecchio V, Barbieri M, Albino V, Piccirillo M, Amore A, Di Giacomo R, Nasto A, Granata V, Petrillo A, Arra C, Izzo F (2015) Inhibitory effect of (-)-epigallocatechin-3-gallate and bleomycin on human pancreatic cancer MiaPaca-2 cell growth. Infect Agent Cancer 10:22. https://doi.org/10.1186/s13027-015-0016-y
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 68(6):394–424. https://doi.org/10.3322/caac.21492
- Brusselmans K, De Schrijver E, Heyns W, Verhoeven G, Swinnen JV (2003) Epigallocatechin-3gallate is a potent natural inhibitor of fatty acid synthase in intact cells and selectively induces apoptosis in prostate cancer cells. Int J Cancer 106(6):856–862

- Cai Y, Zheng Y, Gu J, Wang S, Wang N, Yang B, Zhang F, Wang D, Fu W, Wang Z (2018) Betulinic acid chemosensitizes breast cancer by triggering ER stress-mediated apoptosis by directly targeting GRP78. Cell Death Dis 9(6):636. https://doi.org/10.1038/s41419-018-0669-8
- Chakrabarti M, Ray SK (2015) Direct transfection of miR-137 mimics is more effective than DNA demethylation of miR-137 promoter to augment anti-tumor mechanisms of delphinidin in human glioblastoma U87MG and LN18 cells. Gene 573(1):141–152. https://doi.org/10.1016/j. gene.2015.07.034
- Chen J, Jiu-Hong K (2005) Quercetin and trichostatin a cooperatively kill human leukemia cells. Pharmazie 60(11):856–860
- Chi C, Chang Y, Ou Y, Hsieh C, Lui W, PEng F, Liu T (1997) Effect of quercetin on the in vitro and in vivo growth of mouse hepatoma cells. Oncol Rep 4(5):1021–1024
- Cho YA, Lee J, Oh JH, Chang HJ, Sohn DK, Shin A, Kim J (2017) Dietary flavonoids, CYP1A1 genetic variants, and the risk of colorectal cancer in a Korean population. Sci Rep 7(1):128. https://doi.org/10.1038/s41598-017-00117-8
- Csokay B, Prajda N, Weber G, Olah E (2005) Molecular mechanisms in the antiproliferative action of quercetin. Life Sci 60(24):2157–2163
- Cui Y, Morgenstern H, Greenland S, Tashkin DP, Mao JT, Cai L, Cozen W, Mack TM, Lu QY, Zhang ZF (2008) Dietary flavonoid intake and lung cancer – a population-based case-control study. Cancer 112(10):2241–2248. https://doi.org/10.1002/cncr.23398
- De Stefani E, Deneo-Pellegrini H, Mendilaharsu M, Ronco A (1999) Diet and risk of cancer of the upper aerodigestive tract I. Foods. Oral Oncol 35(1):17–21
- De Stefani E, Brennan P, Boffetta P, Ronco AL, Mendilaharsu M, Deneo-Pellegrini H (2000) Vegetables, fruits, related dietary antioxidants, and risk of squamous cell carcinoma of the esophagus: a case-control study in Uruguay. Nutr Cancer 38(1):23–29. https://doi.org/10.1207/ S15327914NC381\_4
- Debatin KM (2004) Apoptosis pathways in cancer and cancer therapy. Cancer Immunol Immunother 53(3):153–159. https://doi.org/10.1007/s00262-003-0474-8
- Fantini M, Benvenuto M, Masuelli L, Frajese GV, Tresoldi I, Modesti A, Bei R (2015) In vitro and in vivo antitumoral effects of combinations of polyphenols, or polyphenols and anticancer drugs: perspectives on cancer treatment. Int J Mol Sci 16(5):9236–9282. https://doi. org/10.3390/ijms16059236
- Galati G, O'Brien PJ (2004) Potential toxicity of flavonoids and other dietary phenolics: significance for their chemopreventive and anticancer properties. Free Radic Biol Med 37(3):287– 303. https://doi.org/10.1016/j.freeradbiomed.2004.04.034
- Ganai SA (2017) Plant-derived flavone Apigenin: the small-molecule with promising activity against therapeutically resistant prostate cancer. Biomed Pharmacother 85:47–56. https://doi.org/10.1016/j.biopha.2016.11.130
- Gao AM, Zhang XY, Hu JN, Ke ZP (2018) Apigenin sensitizes hepatocellular carcinoma cells to doxorubic through regulating miR-520b/ATG7 axis. Chem Biol Interact 280:45–50. https:// doi.org/10.1016/j.cbi.2017.11.020
- Gapstur SM, Drope JM, Jacobs EJ, Teras LR, McCullough ML, Douglas CE, Patel AV, Wender RC, Brawley OW (2018) A blueprint for the primary prevention of cancer: targeting established, modifiable risk factors. CA Cancer J Clin 68(6):446–470. https://doi.org/10.3322/caac.21496
- Garcia-Closas R, Gonzalez CA, Agudo A, Riboli E (1999) Intake of specific carotenoids and flavonoids and the risk of gastric cancer in Spain. Cancer Causes Control 10(1):71–75
- Geybels MS, Verhage BA, Arts IC, van Schooten FJ, Goldbohm RA, van den Brandt PA (2013) Dietary flavonoid intake, black tea consumption, and risk of overall and advanced stage prostate cancer. Am J Epidemiol 177(12):1388–1398. https://doi.org/10.1093/aje/kws419
- Goker B, Caliskan C, Onur Caglar H, Kayabasi C, Balci T, Erbaykent Tepedelen B, Aygunes D, Yilmaz Susluer S, Mutlu Z, Selvi Gunel N, Korkmaz M, Saydam G, Gunduz C, Biray Avci C (2014) Synergistic effect of ponatinib and epigallocatechin-3-gallate induces apoptosis in chronic myeloid leukemia cells through altering expressions of cell cycle regulatory genes. J BUON 19(4):992–998

- Guclu-Ustundag O, Mazza G (2007) Saponins: properties, applications and processing. Crit Rev Food Sci Nutr 47(3):231–258. https://doi.org/10.1080/10408390600698197
- Hakimuddin F, Paliyath G, Meckling K (2004) Selective cytotoxicity of a red grape wine flavonoid fraction against MCF-7 cells. Breast Cancer Res Treat 85(1):65–79
- Hibasami H, Komiya T, Achiwa Y, Ohnishi K, Kojima T, Nakanishi K, Akashi K, Hara Y (1998) Induction of apoptosis in human stomach cancer cells by green tea catechins. Oncol Rep 5(2):527–529
- Horie N, Hirabayashi N, Takahashi Y, Miyauchi Y, Taguchi H, Takeishi K (2005) Synergistic effect of green tea Catechins on cell growth and apoptosis induction in gastric carcinoma cells. Biol Pharm Bull 28(4):574–579
- Hsu CP, Shih YT, Lin BR, Chiu CF, Lin CC (2012) Inhibitory effect and mechanisms of an anthocyanins- and anthocyanidins-rich extract from purple-shoot tea on colorectal carcinoma cell proliferation. J Agric Food Chem 60(14):3686–3692. https://doi.org/10.1021/jf204619n
- Huynh H, Nguyen T, Chan E, Tran E (2003) Inhibition of ErbB-2 and ErbB-3 expression by quercetin prevents transforming growth factor alpha (TGF-alpha)- and epidermal growth factor (EGF)-induced human PC-3 prostate cancer cell proliferation. Int J Oncol 23(3):821–829
- Inoue M, Tajima K, Mizutani M, Iwata H, Iwase T, Miura S, Hirose K, Hamajima N, Tominaga S (2001) Regular consumption of green tea and the risk of breast cancer recurrence: follow-up study from the Hospital-based Epidemiologic Research Program at Aichi Cancer Center (HERPACC), Japan. Cancer Lett 167(2):175–182
- Jeong MH, Ko H, Jeon H, Sung GJ, Park SY, Jun WJ, Lee YH, Lee J, Lee SW, Yoon HG, Choi KC (2016) Delphinidin induces apoptosis via cleaved HDAC3-mediated p53 acetylation and oligomerization in prostate cancer cells. Oncotarget 7(35):56767–56780. https://doi.org/10.18632/ oncotarget.10790
- Jin X, Sun J, Miao X, Liu G, Zhong D (2013) Inhibitory effect of geraniol in combination with gemcitabine on proliferation of BXPC-3 human pancreatic cancer cells. J Int Med Res 41(4):993–1001. https://doi.org/10.1177/0300060513480919
- Johnson JJ, Syed DN, Suh Y, Heren CR, Saleem M, Siddiqui IA, Mukhtar H (2010) Disruption of androgen and estrogen receptor activity in prostate cancer by a novel dietary diterpene carnosol: implications for chemoprevention. Cancer Prev Res (Phila) 3(9):1112–1123. https://doi. org/10.1158/1940-6207.CAPR-10-0168
- Kapinova A, Kubatka P, Golubnitschaja O, Kello M, Zubor P, Solar P, Pec M (2018) Dietary phytochemicals in breast cancer research: anticancer effects and potential utility for effective chemoprevention. Environ Health Prev Med 23(1):36. https://doi.org/10.1186/s12199-018-0724-1
- Katsube N, Iwashita K, Tsushida T, Yamaki K, Kobori M (2003) Induction of apoptosis in cancer cells by Bilberry (*Vaccinium myrtillus*) and the anthocyanins. J Agric Food Chem 51(1):68–75. https://doi.org/10.1021/jf025781x
- Key TJ, Sharp GB, Appleby PN, Beral V, Goodman MT, Soda M, Mabuchi K (1999) Soya foods and breast cancer risk: a prospective study in Hiroshima and Nagasaki, Japan. Br J Cancer 81(7):1248–1256. https://doi.org/10.1038/sj.bjc.6690837
- Kim SH, Park EJ, Lee CR, Chun JN, Cho NH, Kim IG, Lee S, Kim TW, Park HH, So I, Jeon JH (2012) Geraniol induces cooperative interaction of apoptosis and autophagy to elicit cell death in PC-3 prostate cancer cells. Int J Oncol 40(5):1683–1690. https://doi.org/10.3892/ ijo.2011.1318
- Kim EH, Baek S, Shin D, Lee J, Roh JL (2017) Hederagenin induces apoptosis in cisplatin-resistant head and neck cancer cells by inhibiting the Nrf2-ARE antioxidant pathway. Oxidative Med Cell Longev 2017:5498908. https://doi.org/10.1155/2017/5498908
- Knekt P, Jarvinen R, Seppanen R, Hellovaara M, Teppo L, Pukkala E, Aromaa A (1997) Dietary flavonoids and the risk of lung cancer and other malignant neoplasms. Am J Epidemiol 146(3):223–230
- Knekt P, Kumpulainen J, Jarvinen R, Rissanen H, Heliovaara M, Reunanen A, Hakulinen T, Aromaa A (2002) Flavonoid intake and risk of chronic diseases. Am J Clin Nutr 76(3):560– 568. https://doi.org/10.1093/ajcn/76.3.560

- Konoshima T, Kokumai M, Kozuka M, Tokuda H, Nishino H, Iwahima A (1992) Anti-tumorpromoting activities of Afromosin and Soyasaponin I isolated from Wistaria brachybotrys. J Nat Prod 55(12):1776–1778
- Kumi-Diaka J, Sanderson N-A, Hall A (2000) The mediating role of caspase-3 protease in the intracellular mechanism of genistein-induced apoptosis in human prostatic carcinoma cell lines, DU145 and LNCaP. Biol Cell 92(8–9):595–604
- Kurahashi N, Sasazuki S, Iwasaki M, Inoue M, Tsugane S, Group JS (2008) Green tea consumption and prostate cancer risk in Japanese men: a prospective study. Am J Epidemiol 167(1):71– 77. https://doi.org/10.1093/aje/kwm249
- Kyle JA, Sharp L, Little J, Duthie GG, McNeill G (2010) Dietary flavonoid intake and colorectal cancer: a case-control study. Br J Nutr 103(3):429–436. https://doi.org/10.1017/ S0007114509991784
- Le Marchand L, Murphy SP, Hankin JH, Wilkens LR, Kolonel LN (2000) Intake of flavonoids and lung cancer. J Natl Cancer Inst 92(2):154–160
- Lee H, Cho H, Yu R, Lee K, Chun H, Park J (2014) Mechanisms underlying apoptosis-inducing effects of Kaempferol in HT-29 human Colon Cancer cells. Int J Mol Sci 15(2):2722–2737
- Lee MH, Han DW, Hyon SH, Park JC (2011) Apoptosis of human fibrosarcoma HT-1080 cells by epigallocatechin-3-O-gallate via induction of p53 and caspases as well as suppression of Bcl-2 and phosphorylated nuclear factor-kappaB. Apoptosis 16(1):75–85. https://doi.org/10.1007/s10495-010-0548-y
- Li Y, Bhuiyan M, Sarkar FH (1999) Induction of apoptosis and inhibition of c-erbB-2 in MDA-MB-435 cells by genistein. Int J Oncol 15(3):525–533
- Lian F, Li Y, Bhuiyan M, Sarkar FH (1999) p53-independent apoptosis induced by genistein in lung cancer cells. Nutr Cancer 33(2):125–131
- Liang G, Tang A, Lin X, Li L, Zhang S, Huang Z, Tang H, Li QQ (2010) Green tea catechins augment the antitumor activity of doxorubicin in an in vivo mouse model for chemoresistant liver cancer. Int J Oncol 37(1):111–123
- Lin D, Xiao M, Zhao J, Li Z, Xing B, Li X, Kong M, Li L, Zhang Q, Liu Y, Chen H, Qin W, Wu H, Chen S (2016) An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. Molecules 21(10):1374. https://doi.org/10.3390/molecules21101374
- Lingyan Z, Fan Y, Li H, Aixue L, Jiren Z (2017) Luteolin enhances the antitumor activity of lapatinib in human breast cancer cells. Biomed Res 28(11):4902–4907
- Liu W, Xu J, Liu Y, Yu X, Tang X, Wang Z, Li X (2014) Anthocyanins potentiate the activity of trastuzumab in human epidermal growth factor receptor 2-positive breast cancer cells in vitro and in vivo. Mol Med Rep 10(4):1921–1926. https://doi.org/10.3892/mmr.2014.2414
- Liu Y, Bi T, Dai W, Wang G, Qian L, Shen G, Gao Q (2016) Lupeol enhances inhibitory effect of 5-fluorouracil on human gastric carcinoma cells. Naunyn Schmiedeberg's Arch Pharmacol 389(5):477–484. https://doi.org/10.1007/s00210-016-1221-y
- Liu X, Zhang D, Hao Y, Liu Q, Wu Y, Liu X, Luo J, Zhou T, Sun B, Luo X, Xu J, Wang Q, Yang Z, Li L (2018) Cyanidin curtails renal cell carcinoma tumorigenesis. Cell Physiol Biochem 46(6):2517–2531. https://doi.org/10.1159/000489658
- Luo H, Daddysman MK, Rankin GO, Jiang BH, Chen YC (2010) Kaempferol enhances cisplatin's effect on ovarian cancer cells through promoting apoptosis caused by down regulation of cMyc. Cancer Cell Int 10:16. https://doi.org/10.1186/1475-2867-10-16
- McGuire S (2016) World Cancer Report 2014. Geneva, Switzerland: World Health Organization, International Agency for Research on Cancer, WHO Press, 2015. Adv Nutr 7(2):418–419. https://doi.org/10.3945/an.116.012211
- Mertens-Talcott SU, Percival SS (2005) Ellagic acid and quercetin interact synergistically with resveratrol in the induction of apoptosis and cause transient cell cycle arrest in human leukemia cells. Cancer Lett 218(2):141–151
- Mohan A, Narayanan S, Sethuraman S, Krishnan UM (2013) Combinations of plant polyphenols & anti-cancer molecules: a novel treatment strategy for cancer chemotherapy. Anticancer Agents Med Chem 13(2):281–295

- Morris J, Moseley VR, Cabang AB, Coleman K, Wei W, Garrett-Mayer E, Wargovich MJ (2016) Reduction in promotor methylation utilizing EGCG (epigallocatechin-3-gallate) restores RXRalpha expression in human colon cancer cells. Oncotarget 7(23):35313–35326. https:// doi.org/10.18632/oncotarget.9204
- Mosby TT, Cosgrove M, Sarkardei S, Platt KL, Kaina B (2012) Nutrition in adult and childhood cancer: role of carcinogens and anti-carcinogens. Anticancer Res 32(10):4171–4192
- Mouria M, Gukovskaya AS, Jung Y, Buechler P, Hines OJ, Reber HA, Pandol SJ (2002) Foodderived polyphenols inhibit pancreatic cancer growth through mitochondrial cytochrome C release and apoptosis. Int J Cancer 98(5):761–769
- Nguyen TTT (2003) The role of activated MEK-ERK pathway in quercetin-induced growth inhibition and apoptosis in A549 lung cancer cells. Carcinogenesis 25(5):647–659
- Niu G, Sun L, Pei Y, Wang D (2018) Oleanolic acid inhibits colorectal cancer angiogenesis by blocking the VEGFR2 signaling pathway. Anticancer Agents Med Chem 18(4):583–590. https://doi.org/10.2174/1871520617666171020124916
- Pan H, Li J, Rankin GO, Rojanasakul Y, Tu Y, Chen YC (2018) Synergistic effect of black tea polyphenol, theaflavin-3,3'-digallate with cisplatin against cisplatin resistant human ovarian cancer cells. J Funct Foods 46:1–11. https://doi.org/10.1016/j.jff.2018.04.037
- Petrick JL, Steck SE, Bradshaw PT, Trivers KF, Abrahamson PE, Engel LS, He K, Chow WH, Mayne ST, Risch HA, Vaughan TL, Gammon MD (2015) Dietary intake of flavonoids and oesophageal and gastric cancer: incidence and survival in the United States of America (USA). Br J Cancer 112(7):1291–1300. https://doi.org/10.1038/bjc.2015.25
- Rabi T, Bishayee A (2009) d -Limonene sensitizes docetaxel-induced cytotoxicity in human prostate cancer cells: generation of reactive oxygen species and induction of apoptosis. J Carcinog 8:9. https://doi.org/10.4103/1477-3163.51368
- Rabi T, Gupta S (2008) Dietary terpenoids and prostate cancer chemoprevention. Front Biosci 13:3457–3469
- Raju J, Mehta R (2009) Cancer chemopreventive and therapeutic effects of diosgenin, a food saponin. Nutr Cancer 61(1):27–35. https://doi.org/10.1080/01635580802357352
- Schlachterman A, Valle F, Wall KM, Azios NG, Castillo L, Morell L, Washington AV, Cubano LA, Dharmawardhane SF (2008) Combined resveratrol, quercetin, and catechin treatment reduces breast tumor growth in a nude mouse model. Transl Oncol 1(1):19–27
- Scott EN, Gescher AJ, Steward WP, Brown K (2009) Development of dietary phytochemical chemopreventive agents: biomarkers and choice of dose for early clinical trials. Cancer Prev Res (Phila) 2(6):525–530. https://doi.org/10.1158/1940-6207.CAPR-08-0223
- Sharma V, Joseph C, Ghosh S, Agarwal A, Mishra MK, Sen E (2007) Kaempferol induces apoptosis in glioblastoma cells through oxidative stress. Mol Cancer Ther 6(9):2544–2553. https:// doi.org/10.1158/1535-7163.MCT-06-0788
- Shih P-H, Yeh C-T, Yen G-C (2005) Effects of anthocyanidin on the inhibition of proliferation and induction of apoptosis in human gastric adenocarcinoma cells. Food Chem Toxicol 43(10):1557–1566
- Shyu MH, Kao TC, Yen GC (2010) Oleanolic acid and ursolic acid induce apoptosis in HuH7 human hepatocellular carcinoma cells through a mitochondrial-dependent pathway and downregulation of XIAP. J Agric Food Chem 58(10):6110–6118. https://doi.org/10.1021/jf100574j
- Simons CC, Hughes LA, Arts IC, Goldbohm RA, van den Brandt PA, Weijenberg MP (2009) Dietary flavonol, flavone and catechin intake and risk of colorectal cancer in the Netherlands Cohort study. Int J Cancer 125(12):2945–2952. https://doi.org/10.1002/ijc.24645
- Su LJ, Arab L (2002) Tea consumption and the reduced risk of colon cancer results from a national prospective cohort study. Public Health Nutr 5(3):419–425. https://doi.org/10.1079/ PHNPHN2001314
- Sun F, Zheng XY, Ye J, Wu TT, Wang J, Chen W (2012) Potential anticancer activity of myricetin in human T24 bladder cancer cells both in vitro and in vivo. Nutr Cancer 64(4):599–606. https://doi.org/10.1080/01635581.2012.665564
- Surh YJ (2003) Cancer chemoprevention with dietary phytochemicals. Nat Rev Cancer 3(10):768– 780. https://doi.org/10.1038/nrc1189

- Szyf M (2015) Prospects for the development of epigenetic drugs for CNS conditions. Nat Rev Drug Discov 14(7):461–474
- Tan X, Hu D, Li S, Han Y, Zhang Y, Zhou D (2000) Differences of four catechins in cell cycle arrest and induction of apoptosis in LoVo cells. Cancer Lett 158(1):1–6
- Tanaka T, Shnimizu M, Moriwaki H (2012) Cancer chemoprevention by carotenoids. Molecules 17(3):3202–3242. https://doi.org/10.3390/molecules17033202
- Thomasset SC, Berry DP, Garcea G, Marczylo T, Steward WP, Gescher AJ (2007) Dietary polyphenolic phytochemicals – promising cancer chemopreventive agents in humans? A review of their clinical properties. Int J Cancer 120(3):451–458. https://doi.org/10.1002/ijc.22419
- Valcic S, Timmermann BN, Alberts DS, Wachter GA, Krutzsch M, Wymer J, Guillen JM (1996) Inhibitory effect of six green tea catechins and caffeine on the growth of four selected human tumor cell lines. Anticancer Drugs 7(4):461–468
- Wakai K, Ohno Y, Genka K, Ohmine K, Kawamura T, Tamakoshi A, Lin Y, Nakayama T, Aoki K, Fukuma S (1999) Risk modification in lung cancer by a dietary intake of preserved foods and soyfoods: findings from a case-control study in Okinawa, Japan. Lung Cancer 25(3):147–159
- Wang L, Feng J, Chen X, Guo W, Du Y, Wang Y, Zang W, Zhang S, Zhao G (2014) Myricetin enhance chemosensitivity of 5-fluorouracil on esophageal carcinoma in vitro and in vivo. Cancer Cell Int 14:71. https://doi.org/10.1186/s12935-014-0071-2
- Wang SW, Chen YR, Chow JM, Chien MH, Yang SF, Wen YC, Lee WJ, Tseng TH (2018) Stimulation of Fas/FasL-mediated apoptosis by luteolin through enhancement of histone H3 acetylation and c-Jun activation in HL-60 leukemia cells. Mol Carcinog 57(7):866–877. https:// doi.org/10.1002/mc.22807
- Wiseman MJ (2018) Nutrition and cancer: prevention and survival. Br J Nutr 1–7. doi:https://doi. org/10.1017/S0007114518002222
- Wu H, Xin Y, Xu C, Xiao Y (2012) Capecitabine combined with (-)-epigallocatechin-3-gallate inhibits angiogenesis and tumor growth in nude mice with gastric cancer xenografts. Exp Ther Med 3(4):650–654. https://doi.org/10.3892/etm.2012.448
- Xiao B, Qin Y, Ying C, Ma B, Wang B, Long F, Wang R, Fang L, Wang Y (2017) Combination of oncolytic adenovirus and luteolin exerts synergistic antitumor effects in colorectal cancer cells and a mouse model. Mol Med Rep 16(6):9375–9382. https://doi.org/10.3892/mmr.2017.7784
- Xu Z, Huang B, Liu J, Wu X, Luo N, Wang X, Zheng X, Pan X (2018) Combinatorial antiproliferative effects of tamoxifen and naringenin: the role of four estrogen receptor subtypes. Toxicology 410:231–246. https://doi.org/10.1016/j.tox.2018.08.013
- Yanagihara K, Akihiro I, Tetsuya T, Michitaka N (1993) Antiproliferative effects of isoflavones on human cancer cell lines established from the gastrointestinal tract. Cancer Res 53(23):5815–5821
- Yang GY, Liao J, Kim K, Yurkow EJ, Yang CS (1998) Inhibition of growth and induction of apoptosis in human cancer cell lines by tea polyphenols. Carcinogenesis 19(4):611–616
- Yang XW, Wang XL, Cao LQ, Jiang XF, Peng HP, Lin SM, Xue P, Chen D (2012) Green tea polyphenol epigallocatechin-3-gallate enhances 5-fluorouracil-induced cell growth inhibition of hepatocellular carcinoma cells. Hepatol Res 42(5):494–501. https://doi. org/10.1111/j.1872-034X.2011.00947.x
- Yang J, Pi C, Wang G (2018) Inhibition of PI3K/Akt/mTOR pathway by apigenin induces apoptosis and autophagy in hepatocellular carcinoma cells. Biomed Pharmacother 103:699–707. https://doi.org/10.1016/j.biopha.2018.04.072
- Zhang B, Xin Y, Hong X (2015) The flavonoid luteolin enhances doxorubicin-induced autophagy in human osteosarcoma U2OS cells. Int J Clin Exp Med 8(9):15190–15197
- Zhang Y, Vareed SK, Nair MG (2005) Human tumor cell growth inhibition by nontoxic anthocyanidins, the pigments in fruits and vegetables. Life Sci 76(13):1465–1472. https://doi. org/10.1016/j.lfs.2004.08.025
- Zhang KJ, Gu QL, Yang K, Ming XJ, Wang JX (2017) Anticarcinogenic effects of alpha-mangostin: a review. Planta Med 83(3–04):188–202. https://doi.org/10.1055/s-0042-119651

- Zhou DH, Wang X, Feng Q (2014) EGCG enhances the efficacy of cisplatin by downregulating hsa-miR-98-5p in NSCLC A549 cells. Nutr Cancer 66(4):636–644. https://doi.org/10.1080/0 1635581.2014.894101
- Zuo Q, Wu R, Xiao X, Yang C, Yang Y, Wang C, Lin L, Kong AN (2018) The dietary flavone luteolin epigenetically activates the Nrf2 pathway and blocks cell transformation in human colorectal cancer HCT116 cells. J Cell Biochem 119(11):9573–9582. https://doi.org/10.1002/ jcb.27275