

# Chapter 4

## Role of Rhizospheric Microbes in the Management of Phytopathogens



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**Abstract** Medicinal plants play very crucial role in the life of people, and they are used in official and various traditional systems of medicines throughout the world, benefitting people to prevent disease, maintain health, and cure ailments. Nearly all modern pharmaceuticals are considered to be natural products or derived from plants. Fungal diseases are the major constraints in the profitable cultivation of medicinal plants. Phytopathogenic problem of medicinal plants not only reduces the yield, but it is also responsible for the deterioration of biochemical and secondary metabolites which are of immense therapeutic value. Imprudent use of insecticides, fungicides, agrochemicals, and fertilizers poses serious threat to environment. Scientists have reported various mechanisms regarding plant rhizospheric microbes, i.e., fungi and bacteria, which colonize the roots of plant and thus help the plants in maintaining its health. In the present scenario, rhizospheric microbes (biocontrol agents) have gained popularity due to their effectiveness, safety, and eco-friendliness, and hence their demand has gradually increased. Rhizospheric microbes not only manage plant diseases but at the same time also boost plant growth by different mechanisms. Many scientists have already reported the beneficial role of rhizospheric microbes on the health of various medicinal plants. Research on medicinal plants and rhizospheric microbes is inadequate as far as biotic stresses are concerned. The mechanisms of plant disease management such as mycoparasitism, antibiosis, induced systemic resistance, plant growth promotion, root colonization, siderophore production, phosphate solubilization, etc., have been studied well in reference to medicinal plants. Still due to the distinct features of medicinal plants, future research could be a major breakthrough in the significant increase in the production of medicinal plants.

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## 4.1 Introduction

About 80% people of the globe rely on herbal medicine for their health problems according to the World Health Organization (Goto et al. 1998). Due to severe side effects of modern medicines, drugs extracted from medicinal plants are gaining popularity in many developed countries. The basics of herbal medicines mainly depend on plant diversity and past studies of their use in maintaining human health (Table 4.1). Nearly all herbal medicines are natural products or derivatives of plants; interestingly it has also been acknowledged that the discovery of artemisinin, which is an antimalarial drug extracted from medicinal plant “sweet wormwood,” has earned a 2015 Nobel Prize in medicine (George et al. 2016). Fungal diseases pose a serious threat to the profitable cultivation of crop as well as the medicinal plant. In 1845 potato late blight fungus was responsible for Irish famine which led to millions of people to migrate from Ireland. The *Plasmopara viticola* causal organism of downy mildew of grapes devastated the wine industry in France. In 1943 brown spot disease of rice was solely responsible for Bengal famine, and other catastrophic examples of phytopathogens also include apple scab; Panama wilt; wheat rust; wilt of *Cajanus cajanus*, chickpea, castor, and guava; rust; and smut of cereals. Phytopathogenic problem is not only a threat to commercial crop plants, but they are also a threat to our important medicinal plants. *Alternaria* leaf spot and *Fusarium* cause severe wilting in Ashwagandha which leads to enormous yield losses (Sharma et al. 2013; Zuhaib et al. 2016; Ansari and Mahmood 2017; Sharma and Trivedi 2010). Apart from Ashwagandha other medicinal plants were also reported to be infected by *Fusarium* wilt.

Various options of disease management such as chemical, botanical, and biological are available, and among them, chemicals are considered one of the best and reliable options, but they pose serious health and environmental risks, which have limited their use. About 0.1% of the agrochemicals used in crop protection reach to the target pest, rest 99.9% enter into the environment causes greater damage to the ecosystem (Ashraf and Zuhaib 2013). The indiscriminate use of chemical fungicides has consequently caused several health issues, such as toxicity in food, water, and soil, which ultimately leads to pollution of the ecosystem; hence it was recommended by the scientists to use nonchemical methods for the management of phytopathogenic problem of medicinal plants.

Plant rhizospheric microbes including soil fungi and bacteria colonize plant roots and, in turn, help in maintaining plant health. In the present scenario, rhizospheric microbes (biocontrol agents) have gained popularity due to their effectiveness, safety, and eco-friendliness, and hence, their demand has gradually increased. Rhizospheric microbes not only manage the plant diseases but also enhance plant growth by adhering and multiplying at the root hair surface; increase in seedling emergence, functioning of premature nodules, and nodulation; and increase in area

**Table 4.1** Medicinal plants and their uses in human health

Traditional name	Scientific name	Family	Part in use	Medicinal use
Amla	<i>Emblica officinalis</i>	Euphorbiaceae	Fruit	Rich source of ascorbic acid, cough, cold, hyperacidity, laxative, prevention of cancer
Ashok	<i>Saraca asoca</i>	Caesalpiniaceae	Bark, flower	Diabetes disorder, menstrual pain, uterine problems
Ashwagandha	<i>Withania somnifera</i>	Solanaceae	Root	Curative tonic and helps in nerves disorder
Guggul	<i>Commiphora wightii</i>	Burseraceae	Gum resin	Rheumatism, laxative, hyperlipidemia
Aloe	<i>Aloe vera</i>	Liliaceae	Leaves	Used in cosmetic industry
Bael/bilva	<i>Aegle</i>	Rutaceae	Fruit, bark	Diarrhea, dysentery, constipation
Tulsi (perennial)	<i>Ocimum sanctum</i>	Lamiaceae	Leaves/ seed	Helps in bronchitis, acts as expectorant, anticancerous
Sarpagandha (H)	<i>Rauwolfia serpentina</i>	Apocynaceae	Root	Hypertension, insomnia
Bhumiamla (H)	<i>Phyllanthus amarus</i>	Euphorbiaceae	Whole plant	Provide strength, lower the bilirubin
Shatavari	<i>Asparagus racemosus</i>	Liliaceae	Tuber, root	Pregnant women, anti-fatigue, lowers blood sugar
Brahmi	<i>Bacopa monnieri</i>	Scrophulariaceae	Whole plant	Anxiety, improve the memory enhancer,
Makoi/ Kakamachi	<i>Solanum</i>	Solanaceae	Fruit/ whole	Dropsy, general weakness, anticancerous
Isabgol	<i>Plantago ovata</i>	Plantaginaceae	Seed coat	Constipation and gastrointestinal irritations. Also used in food industry
Coleus	<i>Coleus forskohlii</i>	Lamiaceae	Tuberous root	Used in glaucoma, heart functioning, and various types of carcinoma
Henna/mehndi	<i>Lawsonia inermis</i>	Lythraceae	Leaf, seed	Burning, steam, anti-inflammatory
Pashanbheda	<i>Coleus barbatus</i>	Lamiaceae	Root	Stone problems, diabetes
Peppermint	<i>Mentha</i>	Lamiaceae	Leaves,	Digestive, painkiller
Sadabahar	<i>Vinca rosea</i>	Apocynaceae	Whole plant	Blood cancer, blood pressure muscle spasm
Vringraj	<i>Eclipta alba</i>	Compositae	Seed/ whole	Anti-allergic, digestive, hair
Neem	<i>Azadirachta</i>	Meliaceae	Whole plant	Sedative, analgesic, epilepsy

Modified from of Shahzad et al. (2015)

of leaf surface, vigor, biomass, phytohormone, and nutrient, water, and air uptake, hence stimulating the accumulation of important nutrients in plants (Shrivastava et al. 2015). This review will focus on major fungal diseases of medicinal plants and also the recent developments in the field of biological control of medicinal plant diseases by rhizospheric microbes, which will emphasize on the mechanism. In this chapter we will limit our discussion on important rhizospheric microbes, viz., species of *Trichoderma*, *Pseudomonas*, and *Bacillus*.

## 4.2 Biotic Stresses on Medicinal Plants

Since there is a great demand for herbal medicinal in the international market, many biotic factors are responsible for the low productivity of medicinal plants. Biotic factors liable for the low productivity of medicinal plants include an attack of insects, arthropods, fungi, bacteria, and nematodes. Among them the fungi cause major yield losses after insects; in this chapter, we will focus on economic yield losses caused by fungi. Black leaf spot diseases of *Aloe vera* were caused by *Alternaria alternata*; two different isolates A and B of *Alternaria alternata* were isolated from the diseased leaf of *Aloe vera* (Alam et al. 2007). Similarly Shukla et al. (2008) also reported *Pythium* leaf spot of *Aloe vera* which was caused by *Pythium aphanidermatum*. An occurrence of leaf spot of Kalmegh (*Andrographis paniculata*) was also observed which causes severe yield loss of 30–45% (Alam et al. 2007). Leaf blight of *Mentha piperita* and *Ocimum sanctum* was also reported by (Alam et al. 2007) and (Ashraf and Zuhaib 2009), respectively. Same workers (Zuhaib et al. 2016) also reported leaf spot of *Withania somnifera* by *Alternaria alternata* and also screened the resistant cultivars of *Withania somnifera* for the pathogen. Twig blight of periwinkle (*Catharanthus roseus*) caused by *Sclerotinia sclerotiorum* was also studied; similarly foliar infection in the form of leaf spot of *Rauwolfia serpentina* was also studied by Alam et al. (2007). Medicinal plants are not only attacked by phyllospheric pathogens, but they are also attacked by rhizospheric pathogens. *Fusarium* causes severe wilting in Ashwagandha which leads to enormous yield losses (Bharti et al. 2014; Zuhaib et al. 2016; Sharma and Trivedi 2010). Apart from Ashwagandha other medicinal plants were also reported to be infected by *Fusarium* wilt. Important medicinal plants reported to be infected with *Fusarium* wilt include *Atractylodes lancea*, *Dioscorea zingiberensis*, *Euphorbia pekinensis*, *Ophiopogon platyphyllum*, *Pinellia ternata* (Dai et al. 2009), *Curcuma manga* (Khamna et al. 2009), *Launaea nudicaulis* (Mansoor et al. 2007), Jerusalem artichoke (sunchoke) (Jina et al. 2013), *Panax quinquefolius* (Song et al. 2014), *Coleus forskohlii* (Zheng et al. 2012), *Papaver somniferum* (Kishore et al. 1985; Sattar et al. 1995), *Calotropis gigantea* (Selvanathan et al. 2011), *Basilicum* (Elmer et al. 1994; Katan et al. 1996), and *Asparagus* (Lamondia and Elmer 1989). *Coleus forskohlii* is an important medicinal plant; wilt of *Coleus forskohlii* is a disease complex caused by *Rhizoctonia bataticola*, *Fusarium chlamydosporum*, *Sclerotium rolfsii*, and *Ralstonia solanacearum* (Bhattacharya and Jha 2012).

### 4.3 Rhizospheric Microbes as Biocontrol Agents

Plant health may be ameliorated by rhizospheric microbes (naturally present soil fungi and bacteria) by colonizing the plant roots. In the present scenario, rhizospheric microbes (biocontrol agents) have gained popularity due to their effectiveness, safety, and eco-friendliness, and hence, their demand has gradually increased. Vigor, biomass, nutrients and water absorption, yield, root hair proliferation, root hair branching, increase in seedling emergence, increase in area of leaf surface, nodulation, and promoted accumulation of carbohydrates are some of the ways in which rhizospheric microbes supplement plant growth besides providing protection to plants from diseases (Shrivastava et al. 2015). Usage of fungicides is not recommended as it is neither economical nor environmentally friendly. Moreover, its long-term use can cause the development of resistant strains of a pathogen (Ashraf and Zuhair 2014a, b; Vinale et al. 2008). However, research on biological control gained momentum in the last quarter of the tenth century, and several books (Baker and Cook 1974; Cook and Baker 1983) and review articles (Papavizas 1985) have come up stressing the potential of microorganisms in disease management. Numerous microorganisms have been reported to cause antagonism against plant pathogenic fungi in laboratory and in vivo condition. A perfect biocontrol agent/rhizospheric microbe must have the subsequent qualities (Lucy et al. 2004; Mukerji 2000).

1. Prolonged survival, either in active or passive form.
2. Greater probability of contact with the pathogen.
3. Functional under variable environments.
4. Mass multiplication should be easy, feasible, and economical.
5. Proficient and cheap.
6. Eco-friendly.

A number of rhizospheric microbes such as *Trichoderma*, *Bacillus*, and *Pseudomonas* have been found successful against a number of important fungal diseases of medicinal plants (Scher and Baker 1982; Strashnov et al. 1985; Kaur et al. 2006; Abo-Elyousr et al. 2014; Dubey et al. 2007). The most common species of *Trichoderma* which have been successfully exploited in biological control of pathogenic fungi are *T. virens*, *T. viride*, and *T. harzianum* (Benitez et al. 2004). *T. viride* has been found to significantly reduce mycelial growth, a formation of spores, and germ tube formation of *A. solani* and *A. alternate* (Latha et al. 2009). *T. harzianum* has been found active against *F. oxysporum* inciting wilt in Ashwagandha (Sharma and Trivedi 2010). Moreover, *Trichoderma* can even stimulate plant growth; reports of which have been found in the case of *T. virens* (Kumar et al. 2011) and the stimulation of plant defense mechanisms (Chet et al. 1997).

Mechanism of disease suppression by rhizospheric microbes *Trichoderma* spp. is reported to suppress plant pathogenic fungi through a combination of different mechanisms (Table 4.2) such as mycoparasitism, synthesis of antibiotics (Harman 2006; Harman et al. 2004), enzymes degrading cell wall (Jayalakshmi et al. 2009),

**Table 4.2** *Trichoderma* species, their target organism, and mechanisms involved in suppression of plant pathogens

<i>Trichoderma</i> species	Target organism	Factor responsible for biocontrol	Disease control
<i>T. harzianum</i> 1051, <i>T. harzianum</i> 39.1	<i>Crinipellis pernicioso</i>	Chitinase, <i>N</i> -acetylglucosaminidase, $\beta$ -1,3-glucanase, total cellulase, endoglucanase, aryl- $\beta$ -glucosidase, $\beta$ -glucosidase, protease, and amylase	Witches' broom disease ( <i>Crinipellis pernicioso</i> ) of cocoa
<i>T. lignorum</i> , <i>T. virens</i> , <i>T. hamatum</i> , <i>T. harzianum</i> , <i>T. pseudokoningii</i> (Rifai)	<i>Rhizoctonia solani</i>	Extracellular, metabolites or antibiotics, or lytic enzyme action	Damping-off disease of bean plants
<i>T. viride</i> , <i>T. harzianum</i>	<i>Aspergillus flavus</i> and <i>Fusarium moniliforme</i>	Lipolytic, proteolytic, pectinolytic, and cellulolytic enzymes. Unknown (mycotoxins) antibiotic compounds (e.g., peptides, cyclic polypeptides)	Fungal seed-associated
<i>T. harzianum</i> , BAFC 742	<i>Sclerotinia sclerotiorum</i> , BAFC 2232	$\beta$ -1,3-Glucanase and chitinase	Fungal-soybean plant
<i>T. harzianum</i> 25, <i>T. viride</i>	<i>Serpula lacrymans</i>	Antibiotic; anthraquinones	Fungal wood decay
<i>T. virens</i> "Q" strain	<i>Rhizopus oryzae</i> / <i>Pythium</i> sp.	Plant phytoalexin induction by antibiotic compound, gliovirin	Seedling disease of cotton
<i>T. virens</i> isolates GL3 and GL21; <i>T. harzianum</i> T-203	<i>Rhizoctonia solani</i> , <i>Pythium ultimum</i> , <i>Meloidogyne incognita</i>	Antibiotics gliovirin and gliotoxin and other inhibitory metabolites	Damping-off disease of cucumber

Source: Leng et al. (2011)

contesting for the availability of important nutrients and increase in plant health (Zimand et al. 1996), parasitism of host fungus (Komatsu 1968; Gao et al. 2001), inducing plant defense (Jayalakshmi et al. 2009), and/or induced systemic resistance (Harman et al. 2004; Sriram et al. 2009).

### 4.3.1 Mycoparasitism

The most common mechanism used by *Trichoderma* for the suppression of phytopathogens is mycoparasitism (Howell 2003; Vinale et al. 2008). Mycoparasitism is a diverse process involving recognition of the host by the mycoparasite; hyphal attachment and coiling of pathogen hyphae (Whipps 2001; Woo and Lorito 2007).

The biocontrol of *R. solani* by *T. lignorum* through mycoparasitism was very well described by Weindling (1932). Enzymes such as chitinases, proteases, and  $\beta$ -1, 3-glucanases lyse hyphal cell walls of pathogens during mycoparasitic activity (De La Cruz et al. 1993; Schirmbock et al. 1994).  $\beta$ -1, 3-Glucanases have properties for degrading the cell wall and inhibit the mycelial growth and spore germination of phytopathogenic fungi (Benítez et al. 2004; Lin et al. 2007). Degradation of pathogen hyphal membranes and cell walls was achieved by proteases produced by *T. harzianum*. Application of *T. harzianum* may inhibit the synthesis of hydrolytic enzymes such as endo-polygalacturonase and exo-polygalacturonase, produced by *Botrytis cinerea*, a causal agent of gray mold, resulting in reduced disease severity (Elad and Kapat 1999). Mustafa et al. (2009) and Kotze et al. (2011) also observed the mycoparasitic activity of *Trichoderma* species against wide range of plant pathogenic fungi.

### 4.3.2 Competition and Rhizosphere Competence

Biocontrol agents multiplication and their multiplication depends upon various factors like rhizosphere competence, successful root colonization, proliferation along the growing plant roots (Chet 1990; Irtwange 2006). Rhizospheric competence is very crucial which provides appreciable understanding pertaining to mode of action of rhizospheric microbes against wide range of plant pathogens (Whipps 2001; Bais et al. 2004; Howell 2003). *Trichoderma*, *Pseudomonas*, and *Bacillus* are considered as potent biocontrol agents and offer excellent competition in terms of food and space to the pathogens (Wells 1988). Among these three rhizospheric microbes, *Pseudomonas* was reported to be more effective comparatively *Trichoderma* followed by *Bacillus* (Weller 1988).

The mass culture of *Trichoderma* can be prepared by using different media which can be thereafter used directly either by mixing with the soil or indirectly by biopriming methods (Zhang et al. 1996; Howell et al. 2000). *T. viride* have been reported to reduce the disease severity of *Chondrostereum purpureum*, the silver leaf pathogen of plum trees (Corke and Hunter 1979).

A race for obtaining carbon in the rhizosphere was also observed in the evaluation of antagonistic activity of *Trichoderma* spp. against different plant pathogens, especially *F. oxysporum* (Sivan and Chet 1989). Competition for carbon is involved in the suppression of *F. oxysporum* f. sp. *vasinfectum* and *F. oxysporum* f. sp. *melonis* by *T. harzianum* T-35 in the rhizosphere of cotton and melon, respectively (Sivan and Chet 1989). The case of root colonization by bacteria consists of two phases, attachment to roots followed by colonization of roots (Howie et al. 1987). It was also reported that motile isolates were far more better colonizers than non-motile isolates (Toyota and Ikeda 1997). The capability of bioagents to synthesize certain antibiotics has a direct relation to being a good colonizer. Mazzola et al. (1992) sug-



gested that phenazine antibiotic production contributes to the ecological competence of *P. fluorescens* in the rhizosphere of wheat. A decrease in disease severity for take - all disease of wheat and radish wilt caused by *Fusarium* has a direct relation with the establishment of *Pseudomonas* strains (Bull et al. 1991; Raaijmakers et al. 1995). Berger et al. (1996) after thorough studies have drawn a conclusion that decrease in disease severity has a direct relation with the rhizospheric establishment by *B. subtilis*.

### 4.3.3 Antibiosis

Suppression or destruction of diseases producing propagules (spores, conidia, conidiophore) by the synthesis of antibiotics or other chemicals synthesizing the bioagents (fungi or bacteria) is known as antibiosis (Irtwange 2006; Viterbo et al. 2007; Haggag and Mohamed 2007). Most of the biocontrol agents including *Trichoderma*, *Pseudomonas* spp., and *Bacillus* species produce several types of antibiotics (Kumar et al. 2011; Handelsman and Stabb 1996). The antibiotics produced by *Trichoderma* species include gliotoxin (Anitha and Murugesan 2005), harzianic acid (Vinale et al. 2014), trichoviridin (McAlees and Taylor 1995), viridian (Zafari et al. 2008), viridiol (Phuwapraisirisan et al. 2006), alamethicins (Aidemark et al. 2010), and others (Goulard et al. 1995). Gliovirin an antibiotic isolated from *Trichoderma* (*Gliocladium*) *virens* shows a strong inhibitory effect against *Pythium ultimum* and *Phytophthora* species (Howell and Stipanovic 1983). *Thielaviopsis basicola*, *Phymatotrichum omnivorum*, *Rhizopus arrhizus*, or *Verticillium dahliae*. *B. thuringiensis* was not inhibited by gliovirin. Secretion of *T. harzianum* strain against *Gaeumannomyces graminis* var. *tritici* exhibited inhibitory effects supporting the fact that bioagent synthesizes antibiotics plays a vital role in the inhibition of the pathogen.

*Bacillus* and *Pseudomonas* species are also effective microbes in managing plant diseases by the production of antibiotics (Weller 1988; Kumar et al. 2011). Plant disease suppression due to *P. fluorescens* may be due to synthesis of pyoluteorin, phenazine, oomycin A, IAA, siderophores, phenazine, siderophore (Whistler et al. 2000; Schoonbeek et al. 2002; Suzuki et al. 2003; Johri et al. 2003; Rachid and Ahmed 2005), extracellular hydrolytic enzymes (Siddiqui 2006), alginate, HCN (Bagnasco et al. 1998), and pseudomonic acid. The antimicrobial compounds discussed above are responsible to cause fungistasis, inhibition of spore germination, and degradation of a mycelial wall and also induce other fungicidal effect (Thomashow and Weller 1990). Production of iturin and surfactin by *B. subtilis* RB 14 played important role in the protection of tomato plant against *R. solani* (Asaka and Shoda 1996). *B. subtilis* synthesize about five antibiotics, namely, subtilin, bacitracin, bacillin, subtenolin, and bacillomycin (Young et al. 1974). Pukall et al. (2005) isolated four toxin-producing strains of *Bacillus* spp., such as *B. pumilus*, *B. fusiformis*, *B. subtilis*, and *B. mojavensis* apart from *B. cereus*.



#### 4.3.4 Plant Growth Promotion

PGPR helps in improving the plant health by the producing of different metabolites such as siderophore and hydrocyanic acid (HCN) (Bhatia et al. 2008); other metabolites also include phytohormones like indole acetic acid, gibberellins, cytokinins, and ethylene (Patten and Glick 2002). Another mechanism is the breaking of ethylene molecules which inhibits the growth of roots by certain rhizobacteria and also improves the plant health (Glick et al. 1999). Great number of rhizospheric microbes produces the enzyme 1-aminocyclopropane-1-carboxylate (ACC) deaminase, which breaks down the ACC molecule, the direct originator of the plant hormone ethylene (Belimov et al. 2001; Glick 1995). They kindle the root propagation of different crop plants (Belimov et al. 2001). The abovementioned mode of action (breakdown of ACC) is most efficient in plants which undergo stresses like flooding, drought, and phytopathogens (Grichko and Glick 2001; Wang et al. 2000). The plant health improvement by rhizospheric microbes lies in the fact of initiation of phosphorous plant nutrition (Bertrand et al. 2001). (The increase in yield of groundnut by *Pseudomonas* strains is the best example of initiation of phosphorous plant nutrition which leads to easy uptake of soil phosphorus by plants) (Dey et al. 2004). Effect of rhizospheric microbes on plant growth is because of synthesis of siderophores; synthesis of phytohormones which leads to increase in plant growth (Garcia de Salamone et al. 2001); and initiation of phosphorous plant nutrition leading to readily available phosphorous (Richardson 2001). Sen et al. (2012) reported that *Stevia rebaudiana* Bertoni, a natural sweetener, is composed of two main sweetest compounds which make it 300 and 450 times sweeter than sucrose. *Pseudomonas* BRL-1 isolated from the rhizosphere showed both in vitro and in vivo antagonistic activity against the pathogen *Alternaria alternata* inciting leaf blight disease in *Stevia rebaudiana*. Siderophore produced by fluorescent *Pseudomonas* has very high affinity for ferric ion and is secreted during growth under low iron conditions (Johri et al. 2003) which is then converted to ferrous ions and thus reduces iron availability to pathogens. However, the producing strain can utilize this via a very specific receptor in its outer cell membrane (Buyer and Leong 1986). In this way the bacteria may restrict the growth of deleterious bacteria and fungi at the plant root surface (Loper and Buyer 1991). Consequent iron starvation condition prevents the germination of fungal spores. Elad and Baker (1985) have demonstrated the direct relationship between synthesis of siderophores and their tendency to control the germination of chlamydospores of *Fusarium oxysporum*. Johri et al. (2003) have also reported that fluorescent pseudomonas during low iron concentration secrete siderophores which reduces ferric into ferrous ions, and thus no more iron is available to pathogens. However, the synthesizing strain has a tendency to use this with the help of specific receptors in its outer cell membrane (Buyer and Leong 1986). In this is how the bacteria can check the growth of harmful bacteria and fungi at the surface of roots (Loper and Buyer 1991) and hence promote the plant growth.

## 4.4 Conclusions

Now it is very much clear that rhizospheric microbes have a positive trend in increasing the growth and yield of medicinal plants under biotic stresses. Although understanding the mechanism of rhizospheric microbes as a plant growth promoter is still an interesting field of qualitative research, therefore, it is the right time to think about the potential candidate of microbes which can improve the plant health even under biotic stresses. Application of suitable strain of microorganisms in the field infested with the soil borne pathogens may exert some reliable results. Consortium of microorganism of different origin can enhance the potentiality of the bioagents which may be very useful in the disease management. However, mechanisms behind the control of diseases are still the matter of research as this will unravel various important facts related to disease management. Due to the distinct features of medicinal plants, future research could also pave a new platform in understanding the subject. Adequate research in this thrust area could be a major breakthrough in the improvement of health of various economically important medicinal plants.

## References

- Abo-Elyousr, K. A. M., Sobhy, A.-H., & Abdel-Rahim, I. R. (2014). Isolation of *Trichoderma* and evaluation of their antagonistic potential against *Alternaria porri*. *Journal of Phytopathology*, *162*, 567–574.
- Aidemark, M., Tjellström, H., Sandelius, A. S., Stålbrand, H., Andreasson, E., Rasmusson, A. G., & Widell, S. (2010). *Trichoderma viride* cellulase induces resistance to the antibiotic pore-forming peptide alamethicin associated with changes in the plasma membrane lipid composition of tobacco BY-2 cells. *BMC Plant Biology*, *10*(1), 274.
- Alam, M., Khaliq, A., Shukla, R. S., Sattar, A., Singh, H. N., Samad, A., Gupta, M. L., Pandey, R., Ajayakumar, P. V., Sharma, A., & Khanuja, S. P. S. (2007). *Healthy plants for health, a complete treatise on major diseases of medicinal and aromatic plants & their management*. Lucknow, U.P. (Central Institute of Medicinal and Aromatic Plant) CIMAP, India.
- Anitha, R. I., & Murugesan, K. (2005). Production of gliotoxin on natural substrates by *Trichoderma virens*. *Journal of Basic Microbiology*, *45*(1), 12–19.
- Ansari, R. A., & Mahmood, I. (2017). Optimization of organic and bio-organic fertilizers on soil properties and growth of pigeon pea. *Scientia Horticulturae*, *226*, 1–9.
- Asaka, O., & Shoda, M. (1996). Biocontrol of *Rhizoctonia solani* damping-off of tomato with *Bacillus subtilis* RB14. *Applied and Environmental Microbiology*, *62*, 4081–4085.
- Ashraf, S., & Zuhaib, M. (2009). Studies on the development of powdery mildew on *Ocimum sanctum* (Linn) using growth model. *Journal Trends in Biosciences*, *2*(1), 70–72.
- Ashraf, S., & Zuhaib, M. (2013). Fungal biodiversity: A potential tool in plant disease management. In *Management of microbial resources in the environment* (pp. 69–90). Dordrecht: Springer.
- Ashraf, S., & Zuhaib, M. (2014a). Fungal biodiversity a potential tool in plant disease management. In A. Malik, M. Alves, & E. Grohmann (Eds.), *Management of microbial resources in the environment* (pp. 1–530). Dordrecht: Springer.
- Ashraf, S., & Zuhaib, M. (2014b). Efficacy of rhizospheric microorganism against wilt of Ashwagandha (*Withania somnifera* DUNAL) and their influence on its growth. *Trends in Biosciences*, *7*(16), 2165–2167.

- Bagnasco, P., De La Fuente, L., Gaultieri, G., Noya, F., & Arias, A. (1998). Fluorescent *Pseudomonas* spp. as biocontrol agents against forage legume root pathogenic fungi. *Soil Biology and Biochemistry*, 30, 1317–1322.
- Bais, H. P., Fall, R., & Vivanco, J. M. (2004). Biocontrol of *Bacillus subtilis* against infection of *Arabidopsis* roots by *Pseudomonas syringae* is facilitated by biofilm formation and surfactin production. *Plant Physiology*, 134(1), 307–319.
- Baker, K. F., & Cook, R. J. (1974). *Biological control of plant pathogens*. San Francisco: WH Freeman and Co, 433 pp. (Book, reprinted in 1982, Am Phytopathol Soc, St Paul, Minnesota).
- Belimov, A. A., Safronova, V. I., Sergeyeve, T. A., Egorova, T. N., Matveyeva, V. A., Stepanok, V. V., Tsyganov, V. E., Borisov, A. Y., Kluge, C., Preisfeld, A., Dietz, K. J., & Tikhonovich, I. A. (2001). Characterization of plant growth promoting rhizobacteria isolated from polluted soils and containing 1-aminocyclopropane-1-carboxylate deaminase. *Canadian Journal of Microbiology*, 47, 642–652.
- Benítez, T., Rincón, A. M., Limón, M. C., & Codón, A. C. (2004). Mecanismos de biocontrol de cepas de *Trichoderma*. *International Microbiology*, 7(4), 249–260.
- Berger, F., Li, H., White, D., Frazer, R., & Leifert, C. (1996). Effect of pathogen inoculum, antagonist density, and plant species on biological control of *Phytophthora* and *Pythium* damping-off by *Bacillus subtilis* Cot1 in high-humidity fogging glasshouses. *Phytopathology*, 86, 428–433.
- Bertrand, H., Nalin, R., Bally, R., & Cleyet-Marel, J. C. (2001). Isolation and identification of the most efficient plant growth promoting bacteria associated with canola (*Brassica napus*). *Biology and Fertility of Soils*, 33, 152–156.
- Bharti, N., Barnawal, D., Awasthi, A., Yadav, A., & Kalra, A. (2014). Plant growth promoting rhizobacteria alleviate salinity induced negative effects on growth, oil content and physiological status in *Mentha arvensis*. *Acta Physiologiae Plantarum*, 36, 45–60.
- Bhatia, S., Maheshwari, D. K., Dubey, R. C., Arora, D. S., Bajpai, V. K., & Kang, S. C. (2008). Beneficial effects of fluorescent Pseudomonads on seed germination, growth promotion, and suppression of charcoal rot in groundnut (*Arachis hypogea* L). *Journal of Microbiology and Biotechnology*, 18, 1578–1583.
- Bhattacharya, P. N., & Jha, D. K. (2012). Plant growth-promoting rhizobacteria (PGPR): Emergence in agriculture. *World Journal of Microbiology and Biotechnology*, 28, 1327–1350.
- Bull, C. T., Weller, D. M., & Thomashow, L. S. (1991). Relationship between root colonization and suppression of *Gaeumannomyces graminis* var. *tritici* by *Pseudomonas fluorescens* strain 2-79. *Phytopathology*, 81(9), 954–959.
- Buyer, J. S., & Leong, J. (1986). Iron transport-mediated antagonism between plant growth-promoting and plant-deleterious *Pseudomonas* strains. *Journal of Biological Chemistry*, 261(2), 791–794.
- Chet, I. (1990). Mycoparasitism – Recognition, physiology and ecology. In R. Baker & P. Dunn (Eds.), *New directions in biological control: Alternatives for suppressing agricultural pests and diseases* (pp. 725–783). New York: Alan R Liss.
- Chet, I., Inbar, J., & Hadar, I. (1997). *Fungal antagonists and mycoparasites. The mycota IV: Environmental and microbial relationships* (pp. 165–184). Berlin: Springer-Verlag.
- Cook, R. J., & Baker, K. F. (1983). *The nature and practices of biological control of plant pathogens* (p. 539). St. Paul: American Phytopathology Society.
- Corke, A. T. K., & Hunter, T. (1979). Biocontrol of *Nectria galligena* infection of pruning wounds on apple shoots. *Journal of Horticultural Science*, 54(1), 47–55.
- Dai, C. C., Xie, H., Wang, X. X., Li, P. D., Zhang, T. L., Li, Y. L., & Tan, X. (2009). Intercropping peanut with traditional Chinese medicinal plants improves soil microcosm environment and peanut production in subtropical. *China African Journal Biotechnology*, 8, 3739–3746.
- De La Cruz, J., Rey, M., Lora, J. M., Hidalgo-Gallego, A., Domínguez, F., Pintor-Toro, J. A., et al. (1993). Carbon source control on  $\beta$ -glucanases, chitinase and chitinase from *Trichoderma harzianum*. *Archives of Microbiology*, 159(4), 316–322.

- De Salamone, I. E. G., Hynes, R. K., & Nelson, L. M. (2001). Cytokinin production by plant growth promoting rhizobacteria and selected mutants. *Canadian Journal of Microbiology*, *47*, 404–411.
- Dey, R., Pal, K. K., Bhatt, D. M., & Chauhan, S. M. (2004). Growth promotion and yield enhancement of peanut (*Arachis hypogaea* L) by application of plant growth promoting rhizobacteria. *Microbiological Research*, *159*, 371–394.
- Dubey, S. C., Suresh, M., & Singh, B. (2007). Evaluation of *Trichoderma* species against *Fusarium oxysporum* f. sp. ciceris for integrated management of chickpea wilt. *Journal of Biological Control*, *40*, 118–127.
- Elad, Y., & Baker, R. (1985). The role of competition for iron and carbon in suppression of chlamyospore germination of *Fusarium* spp. by *Pseudomonas* spp. *Phytopathology*, *75*(9), 1053–1059.
- Elad, Y., & Kapat, A. (1999). The role of *Trichoderma harzianum* protease in the biocontrol of *Botrytis cinerea*. *European Journal of Plant Pathology*, *105*, 177–189.
- Elmer, W. H., Wick, R. L., & Haviland, P. (1994). Vegetative compatibility among *Fusarium oxysporum* f. sp. *basilicum* isolates recovered from basil seeds and infected plants. *Plant Diseases*, *78*, 789–791.
- Gao, K., Liu, X., Guo, R., Huai, W., & Zhang, M. (2001). Study on the antagonism of *Trichoderma* species on canker pathogen fungi of poplar. *Scientia Silvae Sinicae*, *37*(5), 82–86.
- George, D. R., Edris, W., Hanson, R., & Gilman, F. (2016). Medicinal plants – The next generation. *The Lancet*, *387*(10015), 220–221.
- Glick, B. R. (1995). The enhancement of plant growth by free living bacteria. *Canadian Journal of Microbiology*, *41*, 1376–1381.
- Glick, B. R., Patten, C. L., Holguin, G., & Penrose, D. M. (1999). *Biochemical and genetic mechanisms used by plant growth promoting bacteria*. London: Imperial College Press.
- Goto, S., Nishioka, T., & Kanehisa, M. (1998). LIGAND: Chemical database for enzyme reactions. *Bioinformatics*, *14*, 591–599.
- Goulard, C., Hlimi, S., Rebuffat, S., & Bodo, B. (1995). Trichorzins HA and MA, antibiotic peptides from *Trichoderma harzianum*, I: Fermentation, isolation and biological properties. *Journal of Antibiotics*, *48*, 1248–1253.
- Grichko, V. P., & Glick, B. R. (2001). Amelioration of flooding stress by ACC deaminase-containing plant growth-promoting bacteria. *Plant Physiology and Biochemistry*, *39*, 11–17.
- Haggag, W. M., & Mohamed, H. A. A. (2007). Biotechnological aspects of microorganisms used in plant biological control. *American-Eurasian Journal of Sustainable Agriculture*, *1*, 7–12.
- Handelsman, J., & Stabb, E. V. (1996). Biocontrol of soilborne plant pathogens. *The Plant Cell*, *8*(10), 1855.
- Harman, G. E. (2006). Overview of mechanisms and uses of *Trichoderma* spp. *Phytopathology*, *96*(2), 190–194.
- Harman, G. E., Howell, C. R., Viterbo, A., Chet, I., & Lorito, M. (2004). *Trichoderma* species-opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, *2*, 43–56.
- Howell, C. R. (2003). Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: The history and evolution of current concepts. *Plant Disease*, *87*(1), 4–10.
- Howell, C. R., & Stipanovic, R. D. (1983). Gliovirin, a new antibiotic from *Gliocladium virens*, and its role in the biological control of *Pythium ultimum*. *Canadian Journal of Microbiology*, *29*(3), 321–324.
- Howell, C. R., Hanson, L. E., Stipanovic, R. D., & Puckhaber, L. S. (2000). Induction of terpenoid synthesis in cotton roots and control of *Rhizoctonia solani* by seed treatment with *Trichoderma virens*. *Phytopathology*, *90*(3), 248–252.
- Howie, W. J., Cook, R. J., & Weller, D. M. (1987). Effects of soil matric potential and cell motility on wheat root colonization by fluorescent pseudomonads suppressive to take-all. *Phytopathology*, *77*(2), 286–292.
- Irtwange, S. (2006). Application of biological control agents in pre- and post-harvest operations. *Agricultural Engineering International* 8, Invited Overview 3, A & M University Press, Texas.

- Jayalakshmi, S. K., Raju, S., Rani, S. U., Benagi, V. I., & Sreeramulu, K. (2009). *Trichoderma harzianum* L<sup>^</sup> sub 1<sup>^</sup> as a potential source for lytic enzymes and elicitor of defense responses in chickpea (*Cicer arietinum* L.) against wilt disease caused by *Fusarium oxysporum* f. sp. *ciceri*. *Australian Journal of Crop Science*, 3(1), 44–52.
- Jina, S., Liua, L., Liua, Z., Longa, X., Shaoa, H., & Chenc, J. (2013). Characterization of marine *Pseudomonas* spp. antagonist towards three tuber-rotting fungi from Jerusalem artichoke, a new industrial crop. *Industrial Crops and Products*, 43, 556–561.
- Johri, B. N., Sharma, A., & Viridi, J. S. (2003). Rhizobacterial diversity in India and its influence on soil and plant health. In *Biotechnology in India I* (pp. 49–89). Berlin/Heidelberg: Springer.
- Katan, T., Gamliel, A., & Katan, J. (1996). Vegetative compatibility of *Fusarium oxysporum* from sweet basil in Israel. *Plant Pathology*, 45(4), 656–661.
- Kaur, R., Macleod, J., Foley, W., & Nayudu, M. (2006). Gluconic acid: An antifungal agent produced by *Pseudomonas* species in biological control of take-all. *Phytochemistry*, 67, 595–604.
- Khamna, S., Yokota, A., & Lumyong, S. (2009). Actinomycetes isolated from medicinal plant rhizosphere soils: Diversity and screening of antifungal compounds, indole-3-acetic acid and siderophore production. *World Journal of Microbiology and Biotechnology*, 25, 649–655.
- Kishore, R. A. J., Tripathi, R. D., Johrf, J. K., et al. (1985). Some new fungal diseases of opium poppy (*Papaver somniferum* L.). *Indian Journal of Plant Pathology*, 3, 213–217.
- Komatsu, M. (1968). *Trichoderma viride* as an antagonist of wood inhabiting Hymenomycetes, VIII. The antibiotic activity against the Mycelial growth of *Lentinus edodes* (Berk) sig. of three genera *T. pachybasium*, *Gliocladium* and other sterile forms. Japan: Tottori Mycological Institute.
- Kotze, C., Van Niekerk, J. M., Halleen, F., & Fourie, P. H. (2011). Evaluation of biocontrol agents for grapevine pruning wound protection against trunk pathogen infection. *Phytopathologia Mediterranea*, 50(Supplement), S247–S263.
- Kumar, S., Gupta, P., Sharma, S., & Kumar, D. (2011). A review on immunostimulatory plants. *Journal of Chinese Integrative Medicine*, 9, 117–128.
- LaMondia, J. A., & Elmer, W. H. (1989). Pathogenicity and vegetative compatibility of isolates of *Fusarium oxysporum* and *Fusarium moniliforme* colonizing *Asparagus* tissue. *Canadian Journal of Botany*, 67, 2420–2424.
- Latha, P., Anand, T., Ragupathi, N., Prakasam, V., & Samiyappan, R. (2009). Antimicrobial activity of plant extracts and induction of systemic resistance in tomato plants by mixtures of PGPR strains and Zimmu leaf extract against *Alternaria solani*. *Biological Control*, 50, 85–93.
- Leng, P., Zhang, Z., Pan, G., & Zhao, M. (2011). Applications and development trends in biopesticides. *African Journal of Biotechnology*, 10, 19864–19873.
- Lin, C., Yang, J., Sun, H., Huang, X., Wang, R., & Zhang, K. Q. (2007). Purification and characterization of a  $\beta$ -1, 3-glucanase from the novel mycoparasite *Periconia byssoides*. *Biotechnology Letters*, 29, 617–622.
- Loper, J. E., & Buyer, J. S. (1991). Siderophores in microbial interactions on plant surfaces. *Molecular Plant-Microbe Interactions*, 4, 5–13.
- Lucy, M., Reed, E., & Glick, B. R. (2004). Application of free living plant growth promoting rhizobacteria. *Antonie Van Leeuwenhoek*, 86, 1–25.
- Mansoor, F., Sultana, V., & Ehteshamul-Haque, S. (2007). Enhancement of biocontrol potential of *Pseudomonas aeruginosa* and *Paecilomyces lilacinus* against root rot of mungbean by a medicinal plant *Launaea nudicaulis*. *Pakistan Journal of Botany*, 39(6), 2113–2119.
- Mazzola, M., Cook, R. J., Thomashow, L. S., Weller, D. M., & Pierson, L. S. (1992). Contribution of phenazine antibiotic biosynthesis to the ecological competence of fluorescent pseudomonads in soil habitats. *Applied and Environmental Microbiology*, 58(8), 2616–2624.
- McAlees, A. J., & Taylor, A. (1995). The biodegradation of L-tyrosine by *Trichoderma hamatum* to trichoviridin and related compounds. *Proceedings of the Nova Scotian Institute of Science*, 40(2), 61–65.

- Mukerji, K. G. (2000). Exploitation of protoplast fusion technology in improving biocontrol potential. In *Biocontrol potential and its exploitation in Sustainable agriculture* (pp. 39–48). Boston: Springer.
- Mustafa, A., Aslam, M., Khan, M., Inam-ul-Haq, M., Pervez, A., & Ummad-ud-Din Umar. (2009). Usefulness of different culture media for in-vitro evaluation of *Trichoderma* sp. against seed-borne fungi of economic importance. *Pakistan Journal of Phytopathology*, 21(1), 83–88.
- Papavizas, G. C. (1985). *Trichoderma* and *Gliocladium* their biology, ecology and potential of biocontrol. *Annual Review of Phytopathology*, 23, 23–54.
- Patten, C. L., & Glick, B. R. (2002). Role of *Pseudomonas putida* indole-acetic acid in development of host plant root system. *Applied and Environmental Microbiology*, 68, 3795–3801.
- Phuwapraisrisan, P., Rangsan, J., Siripong, P., & Tin-Pyang, S. (2006). 9-epiViridiol, a novel cytotoxic furanosteroid from soil fungus *Trichoderma virens*. *Natural Product Research*, 20(14), 1321–1325.
- Pukall, C. R., Schumann, P., Hormazabal, V., & Granum, P. (2005). Toxin producing ability among *Bacillus* spp. outside *Bacillus cereus* group. *Applied and Environmental Microbiology*, 71, 1178–1183.
- Raaijmakers, J. M., van der Sluis, I., Koster, M., Bakker, P. A. H. M., Weisbeek, P. J., & Schippers, B. (1995). Utilization of heterologous siderophores and rhizosphere competence of fluorescent *Pseudomonas* spp. *Canadian Journal of Microbiology*, 41, 126–135.
- Rachid, D., & Ahmed, B. (2005). Effect of iron and growth inhibitors on siderophores production by *Pseudomonas fluorescens*. *African Journal of Biotechnology*, 4, 697–702.
- Richardson, A. E. (2001). Prospects for using soil microorganisms to improve the acquisition of phosphorus by plants. *Australian Journal of Plant Physiology*, 28, 897–906.
- Sattar, A., Samad, A., Alam, M., et al. (1995). Screening of opium poppy (*Papaver somniferum*) germplasm for disease resistance. *Current Research on Medicinal and Aromatic Plants*, 17, 315–320.
- Scher, F. M., & Baker, R. (1982). Effect of *Pseudomonas putida* and a synthetic iron chelator on induction of soil suppressiveness to Fusarium wilt pathogen. *Phytopathology*, 72, 1567–1573.
- Schirmbock, M., Lorito, M., Wang, Y. L., Hayes, C. K., Arsian-Atac, I., Scala, F., Harman, G. E., & Kubicek, C. P. (1994). Parallel formation and synergism of hydrolytic enzymes and peptaibol antibiotics, molecular mechanisms involved in the antagonistic action of *Trichoderma harzianum* against phytopathogenic fungi. *Applied and Environmental Microbiology*, 60, 4364–4370.
- Schoonbeek, H., Raaijmakers, J. M., & De Waard, M. A. (2002). Fungal ABC transporters and microbial interactions in natural environments. *Molecular Plant-Microbe Interactions*, 15, 1165–1172.
- Selvanathan, S., Indrakumar, I., & Johnpaul, M. (2011). Biodiversity of the endophytic fungi isolated from *Calotropis gigantea* (L). *Recent Research in Science and Technology*, 3, 94–100.
- Sen, S., Biswas, G., Basu, S. K., & Acharya, K. (2012). Management of leaf spot disease of *Stevia rebaudiana* Bertoni with antagonistic bacteria. *Australian Journal of Crop Science*, 6, 350–356.
- Shahzad, S. M., Ashraf, M., Arif, M. S., Riaz, M., Yasmeen, T., Abid, M., Ghazanfar, M. U., & Zahid, M. A. (2015). *Plant-Growth-Promoting Rhizobacteria (PGPR) and medicinal plants* (Soil Biology 42, Ed. D. Egamberdieva, et al.). Cham: Springer. <https://doi.org/10.1007/978-3-319-13401-7>.
- Sharma, P., & Trivedi, P. C. (2010). Evaluation of different fungal antagonists against *Fusarium oxysporum* infecting *Withania somnifera* (L) Dunal. *Biology and Environmental Sciences*, 6, 37–41.
- Sharma, I., Kumari, N., & Sharma, V. (2013). Defense gene expression in Sorghum defense gene expression in Sorghum bicolor against *Macrophomina phaseolina* in leaves and roots of susceptible and resistant cultivars. *Journal of Plant Interactions*, 9(1), 315–323.
- Shrivastava, S., Egamberdieva, D., & Varma, A. (2015). Plant growth-promoting rhizobacteria (PGPR) and medicinal plants: The state of the art. In *Plant-growth-promoting Rhizobacteria (PGPR) and medicinal plants* (pp. 1–16). Cham: Springer.



- Shukla, R. S., Abdul-Khaliq, Singh H. N., & Alam, M. (2008). Phytotoxin production by *Alternaria alternata* and its role in black leaf spot disease of Aloe vera. In *4th National Interactive Meet Souvenir (NIM-08)*. CIMAP (CSIR), Lucknow.
- Siddiqui, Z. A. (2006). PGPR: Prospective biocontrol agents of plant pathogens. In Z. A. Siddiqui (Ed.), *PGPR: Biocontrol and biofertilization* (pp. 111–142). Dordrecht: Springer.
- Sivan, A., & Chet, T. (1989). Biological control effects of a new isolate of *Trichoderma harzianum* on *Pythium aphanidermatum*. *Phytopathology*, *74*, 498–498.
- Song, M., Yun, H. Y., & Kim, Y. H. (2014). Antagonistic *Bacillus* species as a biological control of ginseng root rot caused by *Fusarium cf. incarnatum*. *Journal of Ginseng Research*, *38*, 136–145.
- Sriram, S., Manasa, S. B., & Savitha, M. J. (2009). Potential use of elicitors from *Trichoderma* in induced systemic resistance for the management of *Phytophthora capsici* in red pepper. *Journal of Biological Control*, *23*, 449–456.
- Strashnov, Y., Elad, Y., Sivan, A., Rerdick, Y., & Chet, I. (1985). Control of *Rhizoctonia solani* fruit rot of tomatoes by *Trichoderma harzianum* Rifai. *Crop Protection*, *4*, 359–336.
- Suzuki, S., He, Y., & Oyaizu, H. (2003). Indole-3-Acetic acid production in *Pseudomonas fluorescens* HP72 and its association with suppression of creeping bentgrass brown patch. *Current Microbiology*, *47*(2), 138–143.
- Thomashow, S. L., & Weller, M. D. (1990). Role of antibiotics and siderophore in biocontrol of take-all disease of wheat. *Plant and Soil*, *129*, 95–99.
- Toyota, K., & Ikeda, K. (1997). Relative importance of motility and antibiosis in the rhizoplane competence of a biocontrol agent *Pseudomonas fluorescens* MelRC2Rif. *Biology and Fertility of Soils*, *25*(4), 416–420.
- Vinale, F., Sivasithamparam, K., Ghisalberti, E. L., Marra, R., Woo, S. L., & Lorito, M. (2008). *Trichoderma* – Plant pathogens interactions. *Soil Biology and Biochemistry*, *40*, 1–10.
- Vinale, F., Sivasithamparam, K., Emilio, L., Wool, L., Nigro, M., Marra, R., Lombardi, N., Pascale, A., Ruocco, M., Lanzuise, S., Manganiello, G., & Lorito, M. (2014). *Trichoderma* secondary metabolites active on plants and fungal pathogens. *The Open Mycology Journal*, *8*(Suppl-1, M5), 127–139.
- Viterbo, A., Inbar, J., Hadar, Y., & Chet, I. (2007). Plant disease biocontrol and induced resistance via fungal mycoparasites. In C. P. Kubicek & I. S. Druzhinina (Eds.), *Environmental and microbial relationships (The Mycota IV)* (2nd ed., pp. 127–146). Berlin/Heidelberg: Springer.
- Wang, C., Knill, E., Glick, B. R., & Defago, G. (2000). Effect of transferring 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase genes into *Pseudomonas fluorescens* strain CHA0 and its *gacA* derivative CHA96 on their growth promoting and disease-suppressive capacities. *Canadian Journal of Microbiology*, *46*, 898–907.
- Weindling, R. (1932). *Trichoderma lignorum* as a parasite of other soil fungi. *Phytopathology*, *22*, 837–845.
- Weller, D. M. (1988). Biological control of soilborne plant pathogens in the Rhizosphere with bacteria. *Annual Review of Phytopathology*, *26*(1), 379–407.
- Wells, D. H. (1988). *Trichoderma* as a biocontrol agent. In K. G. Mukerji & K. L. Garg (Eds.), *Biocontrol and plant diseases* (p. 73). Boca Raton: CRC Press.
- Whipps, J. M. (2001). Microbial interactions and biocontrol in the rhizosphere. *Journal of Experimental Botany*, *52*, 487–511.
- Whistler, C. A., Stockwell, V. O., & Loper, J. E. (2000). Lon protease influences antibiotic production and UV tolerance of *Pseudomonas fluorescens* Pf-5. *Applied and Environmental Microbiology*, *66*, 2718–2725.
- Woo, S. L., & Lorito, M. (2007). Exploiting the interactions between fungal antagonists, pathogens and the plant for biocontrol. In *Novel biotechnologies for biocontrol agent enhancement and management* (pp. 107–130). Dordrecht: Springer.
- Young, F. E., Tupper, J., & Strominger, J. L. (1974). Autolysis of cell walls of *Bacillus subtilis* mechanism and possible relationship to competence. *The Journal of Biological Chemistry*, *249*, 3600–3602.



- Zafari, D., Koushki, M. M., & Bazgir, E. (2008). Biocontrol evaluation of wheat take-all disease by *Trichoderma* screened isolates. *African Journal of Biotechnology*, 7(20), 3653–3659.
- Zhang, J., Howell, C. R., & Starr, J. L. (1996). Suppression of *Fusarium* colonization of cotton roots and *Fusarium* wilt by seed treatments with *Gliocladium virens* and *Bacillus subtilis*. *Biocontrol Science and Technology*, 6(2), 175–188.
- Zheng, L., Liu, J., Liu, T., Zhu, Z., Jiang, D., & Huang, J. (2012). Fusarium wilt of *Coleus forskohlii* caused by *Fusarium oxysporum* in China. *Canadian Journal of Plant Pathology*, 34, 310–314.
- Zimand, G., Elad, Y., & Chet, I. (1996). Effect of *Trichoderma harzianum* on *Botrytis cinerea* pathogenicity. *Phytopathology*, 86(11), 1255–1260.
- Zuhaib, M., Ashraf, S., & Ali, M. (2016). Screening of *Withania somnifera* L. Germplasm for resistance against leaf spot caused by *Alternaria alternata* (Fr.) Keissler. *Journal of Functional And Environmental Botany*, 6(1), 54–57.