

# Chapter 2

## Biosurfactants in Improving Bioremediation Effectiveness in Environmental Contamination by Hydrocarbons



Paulo Renato Matos Lopes, Renato Nallin Montagnolli,  
Jaqueline Matos Cruz, Elis Marina Turini Claro, and Ederio Dino Bidoia

**Abstract** Recent biotechnological advances currently evidence new surfactant production technologies. Biocompounds produced by fermentative processes appeared as an economic and sustainable alternative to many synthetic molecules. Thereby, biosurfactants have become a promising substitute due to their synthesis potential by a wide variety of microorganisms. Biosurfactants are a highly diverse group of structures, such as glycolipids, lipopeptides, polysaccharide-protein complexes, phospholipids, fatty acids, and neutral lipids. This diversity promotes many advantages compared to synthetic surfactants, thus making biosurfactants the most natural choice for technological advances associated with sustainable development. Such advantages include fermentative production viability by using renewable resources, effectiveness in small concentrations even under extreme conditions, selective and specific potential for several applications, lower toxicity, higher biodegradability, and better stability to physicochemical variations. Despite their benefits, biosurfactants are not widely used because of the high production costs. Hence, cost-effective substrates, optimized cultivation conditions, and mutant lineage development are imperative to make these biomolecules an economically competitive product to propose a widespread replacement of synthetic surfactants.

### 2.1 Introduction

Petroleum spills, including oil-based products, can cause considerable damage to the environment, generating an enormous public concern. Oil pollution demands fast and cost-effective solutions. It is estimated that about 0.40% of global oil

---

P. R. M. Lopes (✉) · R. N. Montagnolli · J. M. Cruz · E. M. T. Claro · E. D. Bidoia  
College of Agricultural and Technological Sciences, São Paulo State University (Unesp),  
Dracena, SP, Brazil  
e-mail: [prm.lopes@unesp.br](mailto:prm.lopes@unesp.br)

production eventually reaches the oceans (Banat et al. 2000). Several accidents with oil spills in recent years have continuously shown the environmental damage from hydrocarbons (Montagnolli and Bidoia 2012).

The major oil spill sources can be traced down to all stages of petroleum processing (exploration, refining, and transport). Accidents are prone to occur in marine tanks, complex bed drilling, marine fleets, refineries, and associated leaks (El-Tarabily 2002; Mille et al. 2006). In Brazil, many accidents involving petroleum, including derivatives such as gasoline and jet fuel, have caused serious environmental problems. In 1998, 1.200 m<sup>3</sup> of diesel oil were released due to pipeline corrosion underneath a considerable area below the city of Cubatão-SP. Pipeline corrosion was also responsible for 1.300 m<sup>3</sup> of hydrocarbons spilled in Guanabara Bay in Rio de Janeiro, which had a history of being contaminated constantly by oil spills (Benincasa 2007).

The lubricating oils are an important petroleum-based product, reaching about 60% of petrochemical products. Its importance is mostly industrial but also due to the massive automobile applications (Mang and Gosalia 2017). However, this high consumption also means a serious potential for environmental problems (Rahman et al. 2002). Lubricants often leak chronically from storage tanks (Lopes et al. 2008). Thus, continuous and prolonged release sets up the scenario for a long-term and highly persistent pollution regarding lubricant oils, leading to the contamination of soils and groundwaters (Chavan and Mukherji 2008). In addition, used lubricant oil suffers structural changes caused by high pressures and temperatures inside automobile engines that affect the biodegradability (Lopes and Bidoia 2009).

The persistence of petroleum hydrocarbons in the environment depends on several factors, such as chemical structure, concentration, and dispersion (Atlas 1981; Mille et al. 2006; Prosser et al. 2016; Duan et al. 2017). Physicochemical treatments are applied in the event of the oil spill; however, these treatments are very expensive, and even more strategy might be necessary depending on the chemical agents chosen as dispersants or catalysts (Elanchezhian et al. 2016; Grote et al. 2018).

Alternatively, the conventional physical treatments can separate soil and contaminants without destroying or chemically modifying the oils. Most of the hydrocarbons are absorbed in the soil matrix, reducing the removal efficiency of any treatment. Biological processes, on the other hand, are promising clean decontamination technologies, as they combine simplicity and cost-effectiveness. Thus, among many novel strategies, bioremediation emerges as the least aggressive and the most suitable method for keeping the ecological balance (Montagnolli and Bidoia 2012).

The decrease of contaminant concentrations by biodegradation or other treatment does not always indicate a decreased toxicity. The incomplete degradation and consequent formation of toxic intermediate compounds (metabolites) can result in increased toxicity during remediation processes (Tamada et al. 2012). Therefore, the combination of chemical analysis and ecotoxicity assays is recommended to elucidate the risks associated with contamination. In this way, a detailed monitoring is crucial for better bioremediation strategies and the establishment of environmental safety standards (Al-Mutairi et al. 2008).

## 2.2 Bioremediation of Areas Impacted by Petroleum Hydrocarbon

The microbial ability to use petroleum hydrocarbons as a carbon source was first demonstrated by Zobell (1946), who also revealed that these organisms were widely distributed in nature. The author described 100 bacterial species, belonging to 30 genera capable of metabolizing petroleum. Later, Bartha and Atlas (1977) expanded this list into another 22 bacterial genera, 14 fungi, and 1 alga. Any efficient bioremediation proposal should demonstrate that the contaminant removal is primarily due to biodegradation rates. In other words, the biodegradation must be higher than natural attenuation degree of decontamination. Among the many difficulties in the development of bioremediation methods is the reliability of laboratory-scale experiments in comparison to field results (Juhasz et al. 2000; Simpanen et al. 2016).

There are also several strategies for improving natural attenuation processes. Many of them are extremely cost-effective compared to physical and chemical treatments. The most common approaches in bioremediation are biostimulation, bio-aeration, bioaugmentation, land farming, composting, and phytoremediation (Wu et al. 2016; Agnello et al. 2016). The insertion of nutrients (biostimulation) and oxygen (bio-aeration) favors microbial metabolism when using pollutants as substrates (Seklemova et al. 2001). Another commonly used procedure known as bioaugmentation is the inoculation of an enriched microbial consortium in soil (Richard and Vogel 1999; Barathi and Vasudevan 2001; Agnello et al. 2016). Montagnolli et al. (2009) obtained biodegradability datasets of lubricating oils using respirometric flasks. Kinetic models have been applied to biodegradation curves, which demonstrated the rate of biodegradation of automotive lubricating oils compared to vegetable oils. It was observed that petroleum degradation tends to decrease slower and last longer. However, the influence of other factors in biodegradation had not been determined. To better establish the optimal conditions of biodegradation, it is important to know the key features of an impacted area, such as residual oil concentration, density of degrading microorganisms, and microbial biodegradation potential.

Generally, an accelerated biodegradation of hydrocarbons depends on the presence of specific microorganisms. In addition, the composition of the microbial population is directly affected by the environmental conditions and the type of hydrocarbons (Admon et al. 2001). However, these indigenous communities lack species with proper enzymatic mechanisms necessary for a rapid biodegradation, which results in long-term biodegradation processes if not bioremediated (Díaz-Ramírez et al. 2008).

In this regard, the bioremediation of soils contaminated with mixed hydrocarbons from petroleum sources is a challenge due to the poor bioavailability and complex chemical composition of these compounds (Lee et al. 2008; Sabate et al. 2004; Yu et al. 2018). The hydrocarbon concentration and the presence of oxidative metabolites with potential risks to the environment can also remain after treatment (Pagnout et al. 2006).

The chromatographic profile of lubricating oils (Lladó et al. 2012), for example, shows a considerable fraction of a nondegradable complex mixture (unresolved complex mixture, UCM). In fact, little is known about the composition of the UCM, although the major components of oils are characterized by their high resistance to biodegradation (Wang and Fingas 2003; Wang et al. 2016). Most of the petroleum-based products are composed of branched and cyclic aliphatic and aromatic compounds, including polycyclic aromatic hydrocarbons (PAH) (Nievas et al. 2008).

The PAHs are often found in the environment due to atmospheric deposition originated from natural sources such as burning biomass and volcanic activity or artificial sources including burned fuels and many environmental accidents from the petrochemical industry (Lors et al. 2010). The release of PAHs represents a great concern due to their toxic, mutagenic, and carcinogenic properties (Martins et al. 2013). Although these aromatic molecules can undergo chemical oxidation, photolysis, adsorption, and volatilization, however, the microbial degradation of PAHs is, in most cases, the main alternative of soil treatment (Yu et al. 2018, Liu et al. 2017).

Several studies on petroleum hydrocarbon biodegradation have been adopting different methodologies (Bidoia et al. 2010; Cerqueira et al. 2014; Zhang et al. 2016), but they all indicate that degradation occurs at least in some specific fractions of these substances. There is no general rule in petroleum biodegradation patterns, as most cases shown with preferential remediation of the lighter compounds were observed, whereas, in other studies, it was directed toward the heavier hydrocarbons (Huang et al. 2004).

In aquatic environments, the biodegradation of pollutants is limited and depends on the bioavailability of nutrients (such as nitrogen and phosphorus) required for the onset of microbial growth. The use of soluble salts containing these elements is an effective way to recreate and optimize the biodegradation under laboratory conditions. Field results are often not the same, as in situ treatments yield unsatisfactory stirring of the medium as well as a much reduced dissolution of salts. As a viable alternative, biosurfactants can be associated with the nutrient solution. This is importantly aimed toward petroleum pollution, because the oil is emulsified by the action of biosurfactants and thus provides a rapid microbial growth (Thavasi et al. 2011; Bezza and Chirwa 2015a, b; Mnif et al. 2017).

## 2.3 Surfactants

Surfactants are amphipathic molecules, i.e., compounds which have polar (hydrophobic) and nonpolar portions (hydrophilic), shifting solubility of other molecules in aqueous solutions based on polarity. These molecules act on the water-oil interface, thus forming micelles in various shapes and sizes (Van Hamme et al. 2006). Surfactants are an important class of chemicals widely used in modern industry (Develter and Laurysen 2010; Franzetti et al. 2010). In 2007, chemical surfactant production reached about 10 million tons (Van Bogaert et al. 2007). In this context,

the market share is led by cleaning detergents (up to 50% of surfactant production), generating a 60-billion-dollar revenue in 2004 (Scheibel 2004). It is known that almost all commercially available surfactants are now chemically synthesized from petroleum.

The conventional chemical surfactants derived from petroleum are subject to the availability of fossil fuels and pose potential threats to the environment due to their recalcitrant nature (Makkar and Rockne 2003; Aparna et al. 2012). Approximately, 0.57 tons of petrochemical intermediates are consumed for each ton of surfactant produced (Patel et al. 1999). By projecting these values to the global production of surfactants, it is estimated that 7.40 million tons of petrochemicals are destined annually for the production of surfactants (Reznik et al. 2010). Thus, there is a trend toward eco-friendly technologies mobilizing the search for novel biodegradable compounds and renewable substrates, including industrial waste (Marchant and Banat 2012; Sasayama et al. 2018).

## 2.4 Biosurfactants

The advances in surfactant technologies are within the scope of many biotechnological types of researches. There was a subtle increase in the number of patents involving biosurfactants at the beginning of this century. More than 70% of these were reported between 2000 and 2010. In contrast, most of the patent registrations for chemical surfactants were performed in the 1900s, with a sharp drop after 2000 (Müller et al. 2012).

Biosurfactants are a natural choice as substitutes to synthetic surfactants because they have several advantages, such as (1) viable fermentative production using renewable resources; (2) effectiveness in small quantities, even under extreme conditions; (3) selective and specific potential for various applications; (4) low toxicity; (5) high biodegradability; and (6) stability to pH, salinity, and temperature variations (Abdel-Mawgoud et al. 2010; Banat et al. 2000; Cameotra and Makkar 2010; Lovaglio et al. 2011; Hazra et al. 2011; Zhao et al. 2017).

The biosurfactants are produced by microorganisms to increase cellular access to hydrophobic substrates. This facilitates the metabolism and promotes the development of biomass, hence increasing biodegradation (Bordoloi and Konwar 2009; Singh et al. 2007). The major advantage of biosurfactants compared to synthetic surfactants lies in their structural diversity and environmental acceptability. Biosurfactants are biodegradable, biocompatible, and less toxic with higher specificity, the possibility of in situ production. They can be produced from renewable substrates and organic residues (Mulligan 2009). A wide range of biosurfactants are potential to apply in various industrial approaches: food, petrochemical, mining, agriculture, cosmetics, pharmaceutical, textile, leather, construction, dyes, and chemicals (Araújo et al. 2016; Ferreira et al. 2017). In addition, these molecules have the ability to decrease surface and interfacial tension. These properties can promote microbial growth, aid microbial enhanced oil recovery (MEOR) procedures

in drilling oil wells, and facilitate bioremediation of pollutants (Zhao et al. 2017). Thus, biosurfactants are a multifunctional material and an important alternative to replace compounds and chemical processes (Silva et al. 2017).

Typically, biosurfactants have hydrophilic structures (amino acids, peptides, mono-/disaccharides, and/or polysaccharides) and hydrophobic structures (saturated and/or unsaturated fatty acids) in their molecules (Smyth et al. 2010; Shao et al. 2015). Biosurfactants are classified by their chemical structure, and this composition depends on the producing microorganisms, the substrate, and the conditions of the fermentation process (Cameotra and Makkar 2004; Nitschke et al. 2005a; Singh et al. 2007; Makkar and Cameotra 1999; Nitschke and Pastore 2006).

Among the various microorganisms able to produce biosurfactants, bacteria belonging to the genus *Pseudomonas* are often the most promising group. They are able to synthesize biosurfactants known as rhamnolipids. These molecules are glycolipids containing fatty acid groups linked to a rhamnose. The lipid portion is composed of  $\beta$ -hydroxydecanoic acid (Benincasa et al. 2004; Mulligan 2009; Abdel-Mawgoud et al. 2010). These different types of rhamnolipids slightly differ on their chemical structures and surface activities. The production of each homolog depends on the nutrient availability, the environmental conditions, and the biosynthesis capabilities of the specific *P. aeruginosa* strain (Oluwaseun et al. 2017; Mondal et al. 2017).

Rhamnolipids are considered as the most promising biosurfactant class in terms of industrial production, due to their physicochemical characteristics, outstandingly high productivity, and deep understanding of the rhamnolipid production (Müller et al. 2012).

## 2.5 Biological Function of Biosurfactants

Biosurfactants reduce surface tension or emulsify hydrophobic substrates (Diaz de Rienzo et al. 2016). However, the biological function of the biosurfactants into the cell is beyond just solubilizing substrates. From the ecological point of view, biosurfactants provide advantages to surfactant-producing microorganisms in relation to other organisms that do not have such capacity. Biosurfactants also have different chemical structures and are produced by different microorganisms (Mulligan 2009; Abdel-Mawgoud et al. 2010; Banat et al. 2010). These biosurfactants are described in the literature as molecules with antimicrobial and antiviral activities (Plaza et al. 2013, Remichkova et al. 2014). This ability benefits ecological interactions (e.g., competition).

There is a quorum sensing mechanism that controls the genes *rhl* and *pqs* responsible to produce rhamnolipids (Pearson et al. 1997; Dusane et al. 2010). Quorum sensing (“sufficient amount” in Latin) is a mechanism for assessing population density through molecular signals to activate a response that requires a certain population density (Madigan et al. 2015). Rhamnolipids are also important for the formation of water channels in the biofilm. These channels provide the homogenous

flow of nutrients and oxygen in the biofilm, in addition to allowing the release of metabolites. Davey et al. (2003) silenced the *rhl* gene of *Pseudomonas aeruginosa*, which caused the blockade of rhamnolipid production and prevented the formation of water channels in the biofilm.

Due to the multiple functions of these biosurfactants, their applicability was observed in activities such as the recovery of areas contaminated with hydrophobic compounds (Amani et al. 2013), emulsifiers in cosmetics and the pharmaceutical industry (Ferreira et al. 2017; Bhadoriya et al. 2013), and phytopathogen control (Ongena and Jacques 2008; D'aes et al. 2010, Falardeau et al. 2013). However, the cost of producing these biomolecules today is high, resulting in unfeasible applications. In comparison with synthetic surfactants with an average cost around \$ 1 to \$ 3 per kg, rhamnolipids cost \$ 20 to \$ 25 per kg (Chong and Li 2017). To overcome these challenges, many strategies are being adopted to increase productivity and reduce costs. These costs are mainly related to the costly carbon source required for the biosurfactant production and also to extraction and purification processes (Chong and Li 2017).

## 2.6 Production of Biosurfactants from Alternative Substrates

At the beginning of scientific interest in biosurfactants around 1980 (Syldatk and Wagner 1987), only pure hydrocarbons were used as carbon sources for their production (Fish et al. 1982; Hisatsuka et al. 1971; Itoh and Suzuki 1972; Syldatk et al. 1985). This consequently raised the biosurfactant market value to an unfeasible acceptance scenario. Despite its many advantages over synthetic chemical surfactants, the biosurfactants still have economic obstacles to overcome in the large-scale process, including a drastic reduction in production costs. In fact, there are efforts in the recent decades that focused on minimizing biosurfactant production costs to promote commercial acceptance (Mukherjee et al. 2006; Costa et al. 2008; Nitschke et al. 2011; Heryani and Putra 2017).

Currently, biosurfactants commercialized in the USA are more expensive than synthetic surfactants (Rosenberg and Ron 1997; Maier and Soberon-Chavez 2000; Makkar et al. 2011). In this context, alternative strategies have been adopted to establish a competitive price. Among the strategies are the development of genetically modified microorganisms toward better yields during the fermentation process (Dobler et al. 2016; Du et al. 2017), use of more cost-effective raw materials for biosurfactant production, and the development of economically viable production processes on a large scale (Mukherjee et al. 2006; Hazra et al. 2011; Makkar et al. 2011).

The use of agro-industrial waste becomes an economically interesting strategy since the raw material accounts for about 10–30% of the total cost in this biotechnological process (Makkar and Cameotra 1999; Mukherjee et al. 2006; Makkar



et al. 2011). There are many alternative substrates currently proposed for the production of biosurfactants: residues generated by the vegetable oil manufacturing (peanut, coconut, corn, olive, soybean), cooking oils (sunflower, olive, soy), vegetable processing waste (potato, barley, cashew, cassava, wheat), sugar cane molasses, whey, peat, oily waste from oil refineries, lignocellulosic waste (fruit peels, corn-cobs), and glycerol from biodiesel production (Desai and Banat 1997; Nitschke et al. 2004; Benincasa 2007; Moldes et al. 2007; Barros et al. 2008; Thavasi et al. 2008; Monteiro et al. 2009; Rocha et al. 2009; Thavasi et al. 2011; Aparna et al. 2012; Cruz et al. 2017; Meneses et al. 2017; Rane et al. 2017). These compounds are known to exhibit high levels of carbohydrates and lipid, both required for the growth of microorganisms and the biosynthesis of biosurfactants (Nitschke et al. 2005b; Nee' Nigam and Pandey 2009; Benincasa 2007; Diaz et al. 2018).

Many microbial genera proved to be able to produce biosurfactants from these residues – *Azotobacter*, *Bacillus*, *Brevibacterium*, *Burkholderia*, *Corynebacterium*, *Flavobacterium*, *Lactobacillus*, *Micrococcus*, *Nocardiosis*, *Pseudomonas*, *Pseudoxanthomonas*, *Rhodococcus*, *Tsukamurella*, *Candida*, *Pseudozyma*, and *Trichosporon* (Boudour et al. 2004; Thavasi et al. 2008; Monteiro et al. 2009; Thavasi et al. 2011).

Agricultural residues result in lower production costs and a much smaller volume of compounds released into the environment. By successfully developing effective ways of producing surfactants, the environmental impact of surfactant industry may become smaller. Moreover, there is a sustainable gain by recycling waste (Mulligan 2009; Accorsini et al. 2012).

## 2.7 Conclusion

Microbial ability to use petroleum hydrocarbons as a carbon source enables an alternative treatment based on bioremediation. Moreover, biotechnological strategies are able to improve natural attenuation processes, and they present cost-effectiveness compared to physicochemical treatments. Generally, an accelerated hydrocarbon biodegradation depends on the presence of specific microorganisms and/or bioavailability of pollutant compounds. Thus, environmental contamination by petroleum derivatives presents many hydrophobic molecules, and microbial metabolism can be enhanced by using tensioactive compounds. In this context, biosurfactants are demonstrated to act on water-oil interface thus forming micelles that raise bioavailability of hydrocarbons to biological treatment. This facilitates the metabolism and promotes the development of biomass, hence increasing biodegradation. Therefore, these biocompounds produced by microorganisms increase cellular access to hydrophobic substrates, and they are a natural choice as substitutes for synthetic surfactants. The advantages of biosurfactant application are currently based on technological advancement associated with sustainable development. Biosurfactants still have economic obstacles to overcome in the large-scale process, including a drastic reduction in production costs. In fact, there are efforts in the recent decades



that focused on minimizing biosurfactant production costs to promote commercial acceptance. Hence, the use of agro-industrial waste as a substrate for fermentative processes becomes an economically interesting strategy. There are many alternative substrates currently proposed for the production of biosurfactants, and also many microbial genera proved to be able to produce biosurfactants from these residues.

## References

- Abdel-Mawgoud AM, Lépine F, Déziel E (2010) Rhamnolipids: diversity of structures, microbial origins and roles. *Appl Microbiol Biotechnol* 86:1323–1336
- Accorsini FR, Mutton MJR, Lemos EGM, Benincasa M (2012) Biosurfactants production by yeasts using soybean oil and glycerol as low-cost substrate. *Braz J Microbiol* 43:116–125
- Admon S, Green M, Avnimelech Y (2001) Biodegradation kinetics of hydrocarbons in soil during land treatment of oily sludge. *Bioremediat J* 5:193–209
- Agnello AC, Bagard M, Van Hullebusch ED, Esposito G, Huguenot D (2016) Comparative bioremediation of heavy metals and petroleum hydrocarbons co-contaminated soil by natural attenuation, phytoremediation, bioaugmentation and bioaugmentation-assisted phytoremediation. *Sci Total Environ* 563–564:693–703
- Al-Mutairi N, Bufarsan A, Al-Rukaibi F (2008) Ecorisk evaluation and treatability potential of soils contaminated with petroleum hydrocarbon-based fuels. *Chemosphere* 74:142–148
- Amani H, Müller MM, Sylatk C, Hausmann R (2013) Production of microbial rhamnolipid by *Pseudomonas aeruginosa* MM1011 for ex situ enhanced oil recovery. *Appl Biochem Biotechnol* 170:1080
- Aparna A, Srinikethan G, Smitha H (2012) Production and characterization of biosurfactant produced by a novel *Pseudomonas* sp. 2B. *Colloids Surf B: Biointerfaces* 95:23–29
- Araujo LV, Guimarães CR, Marquita RLS, Santiago VMJ, De Souza MP, Nitschke M, Freire DMG (2016) Rhamnolipid and surfactin: Anti-adhesion/antibiofilm and antimicrobial effects. *Food Control* 63:171–178
- Atlas MR (1981) Microbial degradation of petroleum hydrocarbons: an environmental perspective. *Microbiol Rev* 45:180–209
- Banat IM, Makkar RS, Cameotra SS (2000) Potential commercial application of microbial surfactants. *Appl Microbiol Biotechnol* 53:495–508
- Banat IM, Franzetti A, Gandolfi I, Bestetti G, Martinotti MG, Fracchia L, Smyth TJ, Marchant R (2010) Microbial biosurfactants production, applications and future potential. *Appl Microbiol Biotechnol* 87:427–444
- Barathi S, Vasudevan N (2001) Utilization of petroleum hydrocarbons by *Pseudomonas fluorescens* isolated from a petroleum contaminated soil. *Environ Int* 26:413–416
- Barros FFC, Ponezi AN, Pastore GM (2008) Production of biosurfactant by *Bacillus subtilis* LB5a on a pilot scale using cassava wastewater as substrate. *J Ind Microbiol Biotechnol* 35:1071–1078
- Bartha R, Atlas RM (1977) The Microbiology of Aquatic oil Spills. *Adv Appl Microbiol* 22:225–226
- Benincasa M (2007) Rhamnolipid produced from agroindustrial wastes enhances hydrocarbon biodegradation in contaminated soil. *Curr Microbiol* 54:445–449
- Benincasa M, Abalos A, Oliveira I, Manresa A (2004) Chemical structure, surface properties and biological activities of the biosurfactant produced by *Pseudomonas aeruginosa* LBI from soap-stock. *Antonie van Leeuwenhoek* 85:1–8
- Bezza FA, Chirwa EMN (2015a) Biosurfactant from *Paenibacillus dendritiformis* and its application in assisting polycyclic aromatic hydrocarbon (PAH) and motor oil sludge removal from contaminated soil and sand media. *Process Saf Environ Prot* 98:354–364

- Bezza FA, Chirwa EMN (2015b) Production and applications of lipopeptide biosurfactant for bioremediation and oil recovery by *Bacillus subtilis* CN2. *Biochem Eng J* 101:168–178
- Bhadoriya SS, Madoriya N, Shukla K, Parihar MS (2013) Biosurfactants: a new pharmaceutical additive for solubility enhancement and pharmaceutical development. *Biochem Pharmacol* 2:1–5
- Bidoia ED, Montagnoli RN, Lopes PRM (2010) Microbial biodegradation potential of hydrocarbons evaluated by colorimetric technique: a case study. In: Méndez-Vilas A (ed) *Current research, technology and education topics in applied microbiology and microbial biotechnology*, Formatex Research Center: Espanha, vol 2, pp 1277–1288
- Bordoloi NK, Konwar BK (2009) Bacterial biosurfactant in enhancing solubility and metabolism of petroleum hydrocarbons. *J Hazard Mater* 170:495–505
- Boudour AA, Guerrero-Baraja C, Jiorle BV, Malcomson ME, Paull AK, Somogyi A, Trinh LN, Bater RB, Maier RM (2004) Structure and characterization of flavolipids, a novel class of biosurfactants produced by *Flavobacterium* sp. strain MTN11. *Appl Environ Microbiol* 70:1114–1120
- Cameotra SS, Makkar RS (2004) Recent applications of biosurfactants as biological and immunological molecules. *Curr Opin Microbiol* 7:262–266
- Cameotra SS, Makkar RS (2010) Biosurfactant-enhanced bioremediation of hydrophobic pollutants. *Pure Appl Chem* 82:97–116
- Cerqueira VS, Peralba MCR, Camargo FAO, Bento FM (2014) Comparison of bioremediation strategies for soil impacted with petrochemical oily sludge. *Int Biodeterior Biodegrad* 95:338–345
- Chavan A, Mukherji S (2008) Treatment of hydrocarbon-rich wastewater using oil degrading bacteria and phototrophic microorganisms in rotating biological contactor: effect of N:P ratio. *J Hazard Mater* 154:63–72
- Chong H, Li Q (2017) Microbial production of rhamnolipids: opportunities, challenges and strategies. *Microb Cell Fact* 16:137
- Costa SGVAO, Nitschke M, Contiero J (2008) Produção de biotenssoativos a partir de resíduos de óleos e gorduras. *Ciênc Tecnol Aliment* 28:34–38
- Cruz JM, Hughes C, Quilty B, Montagnoli RN, Bidoia ED (2017) Agricultural Feedstock Supplemented with Manganese for Biosurfactant Production by *Bacillus subtilis*. *Waste Biomass Valorization*:1–6
- D'Aes J, Maeyn K, Pauwelyn E, Höfte M (2010) Biosurfactants in plant – *Pseudomonas* interactions and their importance to biocontrol. *Environment Microbiol Rep* 2:359–372
- Davey ME, Caiazza NC, O'Toole GA (2003) Rhamnolipid surfactant production affects biofilm architecture in *Pseudomonas aeruginosa* PAO1. *J Bacteriol* 185:1027–1036
- Desai JD, Banat IM (1997) Microbial production of surfactants and their commercial potential. *Microbiol Mol Biol Rev* 61:47–64
- Develter DWG, Laurysen LML (2010) Properties and industrial applications of sophorolipids. *Eur J Lipid Sci Technol* 112:628–638
- Díaz de Rienzo MA, Kamalanathan ID, Martin PJ (2016) Comparative study of the production of rhamnolipid biosurfactants by *B. thailandensis* E264 and *P. aeruginosa* ATCC 9027 using foam fractionation. *Process Biochem* 51:820–827
- Díaz AB, Blandino A, Caro I (2018) Value added products from fermentation of sugars derived from agro-food residues. *Trends Food Sci Technol* 71:52–64 *in press*
- Díaz-Ramírez II, Escalante-Espinosa E, Favela-Torres E, Gutiérrez-Rojas M, Ramírez-Saad H (2008) Design of bacterial defined mixed cultures for biodegradation of specific crude oil fractions, using population dynamics analysis by DGGE. *Int Biodeterior Biodegradation* 62:21–30
- Dobler L, Vilela LF, Almeida RV, Neves BC (2016) Rhamnolipids in perspective: Gene regulatory pathways, metabolic engineering, production and technological forecasting. *N Biotechnol* 33:123–135
- Du J, Zhang A, Hao J, Wang J (2017) Biosynthesis of di-rhamnolipids and variations of congeners composition in genetically-engineered *Escherichia coli*. *Biotechnol Lett* 39:1041–1048

- Duan J, Liu W, Zhao X, Han Y, O'Reilly SE, Zhao D (2017) Study of residual oil in Bay Jimmy sediment 5 years after the Deepwater Horizon oil spill: Persistence of sediment retained oil hydrocarbons and effect of dispersants on desorption. *Sci Total Environ.* 618:1244–1253 *In press*
- Dusane DH, Zinjarde SS, Venugopalan VP, MClean RJC, Weber MM, Rahman PKSM (2010) Quorum sensing: implications on rhamnolipid biosurfactant production. *Biotechnol Genet Eng Rev* 27:159–184
- Elanchezhyan SS, Sivasurian N, Meenakshi S (2016) Enhancement of oil recovery using zirconium-chitosan hybrid composite by adsorptive method. *Carbohydr Polym* 145:103–113
- El-Tarabily KA (2002) Total microbial activity and microbial composition of a mangrove sediment are reduced by oil pollution at a site in the Arabian Gulf. *Can J Microbiol* 48:176–182
- Falardeau J, Wise C, Novitsky L, Avis TJ (2013) Ecological and mechanistic insights into the direct and indirect antimicrobial properties of *Bacillus subtilis* lipopeptides on plant pathogens. *J Chem Ecol* 39:869–878
- Ferreira A, Vecino X, Ferreira D, Cruz JM, Moldes AB, Rodrigues LR (2017) Novel cosmetic formulations containing a biosurfactant from *Lactobacillus paracasei*. *Colloids Surf B Biointerfaces* 155:522–529
- Fish NM, Allenby DJ, Lilly MD (1982) Oxidation of n-alkanes: growth of *Pseudomonas putida*. *Eur J Appl Microbiol Biotechnol* 14:259–262
- Franzetti A, Gandolfi I, Bestett IG, Smyth TJP, Banat IM (2010) Production and applications of trehalose lipid biosurfactants. *Eur J Lipid Sci Technol* 112:617–627
- Grote M, Van Bernem C, Böhme B, Callies U, Calvez I, Christie B, Colcomb K, Damian HP, Farke H, Gräbsch C, Hunt A, Höfer T, Knaack J, Kraus U, Le Floch S, Le Lann G, Leuchs H, Nagel A, Nies H, Nordhausen W, Rauterberg J, Reichenbach D, Scheiffarth G, Schwichtenberg F, Theobald N, Voß J, Wahrendorf DS (2018) The potential for dispersant use as a maritime oil spill response measure in German waters. *Mar. Pollut. Bull.* 129(2):623–632
- Hazra C, Kundu D, Ghosh P, Joshi S, Dandia N, Chaudharia A (2011) Screening and identification of *Pseudomonas aeruginosa* AB4 for improved production, characterization and application of a glycolipid biosurfactant using low-cost agro-based raw materials. *J Chem Technol Biotechnol* 86:185–198
- Heryani H, Putra MD (2017) Dataset on potential large scale production of biosurfactant using *Bacillus* sp. *Data Br* 13:196–201
- Hisatsuka K, Nakahara T, Sano N, Yamada K (1971) Formation of rhamnolipid by *Pseudomonas aeruginosa* and its function in hydrocarbon fermentation. *Agric Biol Chem* 35:686–692
- Huang XD, El-Alawi Y, Penrose DM, Glick B, Greenberg BM (2004) A multiprocess phytoremediation system for removal of polycyclic aromatic hydrocarbons from contaminated soils. *Environ Pollut* 130:465–476
- Itoh S, Suzuki T (1972) Effect of rhamnolipids on growth of *Pseudomonas aeruginosa* mutant deficient in n-paraffin-utilizing ability. *Agric Biol Chem* 36:2233–2235
- Juhász A, Stanley GA, Britz ML (2000) Degradation of high molecular weight PAHs in contaminated soil by a bacterial consortium: effects on Microtox and mutagenicity bioassays. *Bioremediation J* 4:271–283
- Lee SH, Oh BI, Kim JG (2008) Effect of various amendments on heavy mineral oil bioremediation and soil microbial activity. *Biores Technol* 99:2578–2587
- Liu SH, Zeng GM, Niu QY, Liu Y, Zhou L, Jiang LH, Tan XF, Xu P, Zhang C, Cheng M (2017) Bioremediation mechanisms of combined pollution of PAHs and heavy metals by bacteria and fungi: A mini review. *Bioresour Technol* 224:25–33
- Lladó S, Solanas AM, De Lapuente J, Borràs M, Viñas M (2012) A diversified approach to evaluate biostimulation and bioaugmentation strategies for heavy-oil-contaminated soil. *Sci Total Environ* 435-436:262–269
- Lopes PRM, Bidoia ED (2009) Evaluation of the biodegradation of different types of lubricant oils in liquid medium. *Braz Arch Biol Technol* 52:1285–1290

- Lopes PRM, Domingues RF, Bidoia ED (2008) Descarte de embalagens e quantificação do volume de óleo lubrificante residual no município de Rio Claro-SP. *HOLOS Environ* 8:166–178
- Lors C, Rynngaert A, Périé F, Diels L, Damidot D (2010) Evolution of bacterial community during bioremediation of PAHs in a coal tar contaminated soil. *Chemosphere* 81:1263–1271
- Lovaglio RB, Santos FJ, Jafelicci M Jr, Contiero J (2011) Rhamnolipid emulsifying activity and emulsion stability: pH rules. *Colloids Surf B Biointerfaces* 85:301–305
- Madigan MT, Martinko JM, Bender KS, Buckley DH, Stahl DA (2015) *Brock biology of microorganisms*, 14<sup>th</sup> ed. Pearson Education, New York
- Maier RM, Soberón-Chávez G (2000) *Pseudomonas aeruginosa* rhamnolipids: biosynthesis and potential applications. *Appl Microbiol Biotechnol* 54:625–633
- Makkar RS, Cameotra SS (1999) Biosurfactant production by microorganisms on unconventional carbon sources – a review. *J Surfactants Deterg* 2:237–241
- Makkar RS, Rockne KJ (2003) Comparison of synthetic surfactants and biosurfactants in enhancing biodegradation of polycyclic aromatic hydrocarbons. *Environ Toxicol Chem* 22:2280–2292
- Makkar RS, Cameotra SS, Banat IM (2011) Advances in utilization of renewable substrates for biosurfactant production. *AMB Express* 1:5
- Mang T, Gosalia A (2017) Lubricants and their market. In: Mang T, Dresel W (eds) *Lubricants and lubrication*, 3rd edn. Wiley-VCH, Weinheim, pp 1–9
- Marchant R, Banat IM (2012) Microbial biosurfactants: challenges and opportunities for future exploitation. *Trends Biotechnol* 30:558–565
- Martins M, Costa PM, Ferreira AM, Costa MH (2013) Comparative DNA damage and oxidative effects of carcinogenic and non-carcinogenic sediment-bound PAHs in the gills of a bivalve. *Aquat Toxicol* 142–143:85–95
- Meneses DP, Gudiña EJ, Fernandes F, Gonçalves LRB, Rodrigues LR, Rodrigues S (2017) The yeast-like fungus *Aureobasidium thalaidense* LB01 produces a new biosurfactant using olive oil mill wastewater as an inducer. *Microbiol Res* 204:40–47
- Mille G, Guiliano M, Asia L, Malleret L, Jalaluddin N (2006) Sources of hydrocarbons in sediments of the Bay of Fort France (Martinique). *Chemosphere* 64:1062–1073
- Mnif I, Sahnoun R, Ellouz-Chaabouni S, Ghribi D (2017) Application of bacterial biosurfactants for enhanced removal and biodegradation of diesel oil in soil using a newly isolated consortium. *Process Saf Environ Prot* 109:72–81
- Moldes AB, Torrado AM, Barral MT, Domínguez JM (2007) Evaluation of biosurfactant production from various agricultural residues by *Lactobacillus pentosus*. *J Agric Food Chem* 55:4481–4486
- Mondal MH, Sarkar A, Maiti TK, Saha B (2017) Microbial assisted (*Pseudomonas* sp.) production of novel bio-surfactant rhamnolipids and its characterisation by different spectral studies. *J Mol Liq* 242:873–878
- Montagnolli RN, Bidoia ED (2012) Petroleum derivatives biodegradation: environmental impact and bioremediation strategies. Amazon
- Montagnolli RN, Lopes PRM, Bidoia ED (2009) Applied models to biodegradation kinetics of lubricant and vegetable oils in wastewater. *Int Biodeterior Biodegradation* 63:297–305
- Monteiro AS, Coutinho JOPA, Júnior AC, Rosa CA, Siqueira EP, Santos VL (2009) Characterization of new biosurfactant produced by *Trichosporon montevidense* CLOA 72 isolated from dairy industry effluents. *J Basic Microbiol* 49:553–563
- Mukherjee S, Das P, Sen R (2006) Towards commercial production of microbial surfactants. *Trends Biotechnol* 24:09–515
- Müller MM, Kügler JH, Henkel M, Gerlitzki M, Hörmann B, Pöhnlein M, Syldatk C, Hausmann R (2012) Rhamnolipids-next generation surfactants? *J Biotechnol* 162:366–380
- Mulligan CN (2009) Recent advances in the environmental applications of biosurfactants. *Curr Opin Colloid Interface Sci* 14:372–378
- Nee’Nigam PS, Pandey (2009) *A Biotechnology for agro-industrial residues utilisation: utilisation of agro-residues*. Springer, p 466

- Nievas ML, Commendatore MG, Esteves JL, Bucala V (2008) Biodegradation pattern of hydrocarbons from a fuel oil-type complex residue by an emulsifier-producing microbial consortium. *J Hazard Mater* 154:96–104
- Nitschke M, Pastore GM (2006) Production and properties of a surfactant obtained from *Bacillus subtilis* grown on cassava wastewater. *Biores Technol* 97:336–341
- Nitschke M, Ferraz C, Pastore GM (2004) Selection of microorganisms for biosurfactant production using agro industrial wastes. *Braz J Microbiol* 35:81–85
- Nitschke M, Costa SGVAO, Contiero J (2005a) Rhamnolipid surfactants: an update on the general aspects of these remarkable biomolecules. *Biotechnol Prog* 21:1593–1600
- Nitschke M, Costa SGVAO, Hadad R, Gonçalves LA, Eberlin MN, Contiero J (2005b) Oil wastes as unconventional substrates for rhamnolipid biosurfactant production by *Pseudomonas aeruginosa* LBI. *Biotechnol Progress* 21:1562–1566
- Nitschke M, Costa SGVAO, Contiero J (2011) Rhamnolipids and PHAs: recent reports on *Pseudomonas*-derived molecules of increasing industrial interest. *Process Biochem* 46:621–630
- Oluwaseun AC, Kola OJ, Mishra P, Singh JR, Singh AK, Cameotra SS, Micheal BO (2017) Characterization and optimization of a rhamnolipid from *Pseudomonas aeruginosa* C1501 with novel biosurfactant activities. *Sustain Chem Pharm* 6:26–36
- Ongena M, Jacques P (2008) *Bacillus lipopeptides*: versatile weapons for plant disease biocontrol. *Trends Microbiol* 16:115–125
- Pagnout C, Rast C, Veber AM, Poupin P, Féraud JF (2006) Ecotoxicological assessment of PAHs and their dead-end metabolites after degradation by *Mycobacterium* sp. strain SNP11. *Ecotoxicol Environ Saf* 65:151–158
- Patel MK, Theiss A, Worrell E (1999) Surfactant production and use in Germany: Resource requirements and CO<sub>2</sub> emissions. *Resour Conserv Recyc* 25:61–78
- Pearson JP, Pesci EC, Iglewski BH (1997) Roles of *Pseudomonas aeruginosa* las and rhl quorum-sensing systems in control of elastase and rhamnolipid biosynthesis genes. *J Bacteriol* 179:5756–5767
- Plaza GA, Turek A, Król E, Szczygłowska R (2013) Antifungal and antibacterial properties of surfactin isolated from *Bacillus subtilis* growing on molasses. *Afr J Microbiol Res* 7:3165–3170
- Prosser CM, Redman AD, Prince RC, Paumen ML, Letinski DJ, Butler JD (2016) Evaluating persistence of petroleum hydrocarbons in aerobic aqueous media. *Chemosphere* 155:542–549
- Rahman KS, Banat IM, Thahira J, Thayumanavan T, Lakshmanaperumalsamy P (2002) Bioremediation of gasoline contaminated soil by a bacterial consortium amended with poultry litter, coir pith and rhamnolipid biosurfactant. *Biores Technol* 81:25–32
- Rane AN, Baikar VV, Ravi Kumar DV, Deopurkar RL (2017) Agro-industrial wastes for production of biosurfactant by *Bacillus subtilis* ANR 88 and its application in synthesis of silver and gold nanoparticles. *Front Microbiol* 8:492
- Remichkova M, Danka G, Ivana R, Karpenko E, Shulga A, Galabov AS (2014) Anti-Herpesvirus activities of *Pseudomonas* sp. S-17 rhamnolipid and its complex with alginate. *Zeitschrift für Naturforschung C* 63:75–81
- Reznik GO, Vishwanath P, Pynn MA, Sitnik JM, Todd JJ, WU J, Jiang Y, Keenan BG, Castle AB, Haskell RF, Smith TF, Somasundaran P, Jarrell KA (2010) Use of sustainable chemistry to produce and acyl amino acid surfactant. *Appl Microbiol Biotechnol* 86:1387–1397
- Richard JY, Vogel TM (1999) Characterization of a soil bacterial consortium capable of degrading diesel fuel. *Int Biodeterior Biodegradation* 44:93–100
- Rocha MVP, Barreto RVG, Melo VMM, Gonçalves LRB (2009) Evaluation of cashew apple juice for surfactin production by *Bacillus subtilis* LAMI008. *Appl Biochem Biotechnol* 155:366–378
- Rosenberg E, Ron EZ (1997) Bioemulsans: Microbial polymeric emulsifiers. *Curr Opin Biotechnol* 8:313–316
- Sabaté J, Viñas M, Solanas A (2004) Laboratory-scale bioremediation experiments on hydrocarbon-contaminated soils. *Int Biodeterior Biodegradation* 54:19–25
- Sasayama T, Kamikanda Y, Shibasaki-Kitakawa N (2018) Process design for green and selective production of bio-based surfactant with heterogeneous resin catalyst. *Chem Eng J* 334:2231–2237

- Scheibel JJ (2004) The evolution of anionic surfactant technology to meet the requirements of the laundry detergent industry. *J Surfactants Deterg* 7:319–328
- Seklemova E, Pavlova A, Kovacheva K (2001) Biostimulation based bioremediation of diesel fuel: field demonstration. *Biodegradation* 12:311–316
- Shao C, Liu L, Gang H, Yang S, Mu B (2015) Structural diversity of the microbial surfactin derivatives from selective esterification approach. *Int J Mol Sci* 16:1855–1872
- Silva RCFS, Almeida DG, Meira HM, Silva EJ, Farias CBB, Rufino RD, Luna JM, Sarubbo LA (2017) Production and characterization of a new biosurfactant from *Pseudomonas cepacia* grown in low-cost fermentative medium and its application in the oil industry. *Biocatal Agric Biotechnol* 12:206–215
- Simpanen S, Mäkelä R, Mikola J, Silvennoinen H, Romantschuk M (2016) Bioremediation of creosote contaminated soil in both laboratory and field scale: Investigating the ability of methyl- $\beta$ -cyclodextrin to enhance biostimulation. *Int Biodeterior Biodegrad* 106:117–126
- Singh A, Van Hamme JD, Ward OP (2007) Surfactants in microbiology and biotechnology: Part 2. Application aspects. *Biotechnol Adv* 25:99–121
- Smyth TJP, Perfumo A, Marchant R, Banat IM (2010) Isolation and analysis of low molecular weight microbial glycolipids. In: *Handbook of hydrocarbon and lipid microbiology*. Springer, Berlin, pp 3705–3723
- Syldatk C, Wagner F (1987) Production of biosurfactants. *Biosurf Biotechnol* 25:89–120
- Syldatk C, Lang S, Matulovic U, Wagner F (1985) Production of four interfacial active rhamnolipids from n-alkanes or glycerol by resting cells of *Pseudomonas* species DSM 2874. *Z fur Naturforsch Sect C: Biosci* 40:61–67
- Tamada IS, Montagnoli RN, Lopes PRM, Bidoia ED (2012) Toxicological evaluation of vegetable oils and biodiesel in soil during the biodegradation process. *Braz J Microbiol* 43:1576–1581
- Thavasi R, Jayalakshmi S, Balasubramanian T, Banat IM (2008) Production and characterization of a glycolipid biosurfactant from *Bacillus megaterium* using economically cheaper sources. *World J Microbiol Biotechnol* 24:917–925
- Thavasi R, Jayalakshmi IS, Banat IM (2011) Application of biosurfactant produced from peanut oil cake by *Lactobacillus delbrueckii* in biodegradation of crude oil. *Biores Technol* 102:3366–3372
- Van Bogaert I, Saerens K, De Muynck C, Develter D, Soetaert W, Vandamme E (2007) Microbial production and application of sophorolipids. *Appl Microbiol Biotechnol* 76:23–34
- Van Hamme JD, Singh A, Ward OP (2006) Physiological aspects: Part 1 in a series of papers devoted to surfactants in microbiology and biotechnology. *Biotechnol Adv* 24:604–620
- Wang Z, Fingas MF (2003) Development of oil hydrocarbon fingerprinting and identification techniques. *Mar Pollut Bull* 47:423–452
- Wang SY, Kuo YC, Hong A, Chang YM, Kao CM (2016) Bioremediation of diesel and lubricant oil-contaminated soils using enhanced landfarming system. *Chemosphere* 164:558–567
- Wu M, Dick WA, Li W, Wang X, Yang Q, Wang T, Xu L, Zhang M, Chen L (2016) Bioaugmentation and biostimulation of hydrocarbon degradation and the microbial community in a petroleum-contaminated soil. *Int Biodeterior Biodegrad* 107:158–164
- Yu L, Duan L, Naidu R, Semple KT (2018) Abiotic factors controlling bioavailability and bioaccessibility of polycyclic aromatic hydrocarbons in soil: Putting together a bigger picture. *Sci Total Environ* 613–614:1140–1153 *In press*
- Zhang H, Tang J, Wang L, Liu J, Gurav RG, Sun K (2016) A novel bioremediation strategy for petroleum hydrocarbon pollutants using salt tolerant *Corynebacterium variabile* HRJ4 and biochar. *J Environ Sci (China)* 47:7–13
- Zhao F, Shi R, Cui Q, Han S, Dong H, Zhang Y (2017) Biosurfactant production under diverse conditions by two kinds of biosurfactant-producing bacteria for microbial enhanced oil recovery. *J Pet Sci Eng* 157:124–130
- Zobell CE (1946) Action of microorganisms on hydrocarbons. *Bacteriol Rev* 10:1–49