

Chapter 13

Microbial Bioremediation of Petroleum Hydrocarbon: An Overview



Debajit Borah

Abstract Increased environmental toxicity due to extensive use of petroleum-based products gradually proves itself as a major issue of global concern. The release of petroleum products to the environment may cause catastrophic effect on aquatic habitats as well as barrens of fertile soil. Petroleum oil basically contains VOCs (volatile organic compounds), paraffin, gases (methane, ethane, propane, butane, etc.), metal ions (iron, nickel, copper, vanadium, etc.), etc., out of which VOCs may cause severe health problems such as lung, liver and kidney disease. Bioremediation is a process of treatment of contaminated environment with the help of living organisms to bring back to its natural state. Treatment of hydrocarbon-contaminated sites may be accomplished with the help of indigenous microorganisms with diverse groups present in the soil by augmenting with necessary nutrients or by adding external necessary microorganisms. Further, as the petroleum hydrocarbon pollutant creates a stressful environment for growth, the bacterial species having potential to tolerate stress conditions would be an added advantage.

13.1 Introduction

Fossil fuel merchandise increases the probabilities of soil contamination that becomes one of the most important worldwide environmental issues. Statistical report released by the International Tanker Owners Pollution Federation (ITOPF) Ltd. shows incidences of oil spillage in sea since the years 1970–2015 which shows more than 700 tonnes of oil spillage which occurred in year 2015 itself (ITOPF 2016).

NAS report shows over ninetieth of oil spillage incidence is directly or indirectly as a result of human activities together with deliberate oil waste disposal (USEPA 2000; NAS 1985, 2005). Large-scale oil spills, and oil spill accidents, have received a great deal of attention worldwide, as a result of their destructive result on the

D. Borah (✉)

Centre for Biotechnology and Bioinformatics, Dibrugarh University, Dibrugarh, Assam, India

surroundings. Oil spillage on water body ends up in the transience of thousands of aquatic animals and causes major reduction in population of the many aquatic organisms and lots of long-standing environmental impacts (Spies et al. 1996; Campbell and Cary 2001; Van Hamme et al. 2003). Soil contamination primarily arises as a result of petroleum dumping or due to the rupture of underground storage tanks, etc. (Briganti et al. 1997; Butler and Mason 1997; Chang 1998). Minor oil spills and oil contamination as a result of non-point sources like urban runoff, etc. are not any less a threat to public health though they need but received nominal attention in the past. Such non-point sources of pollution remain the largest threat to water body as stated by the reports published by the *National Water Quality Inventory, USA* (Etkin 1998; USEPA 1999, 2000). It may conjointly cause severe health risks to the employees involved in treating of oil spillage areas once exposed to hydrocarbon fumes, volatile organic compounds (VOCs), polycyclic aromatic hydrocarbons (PAHs), material from controlled burns, etc. (Bach et al. 2005; Biddle et al. 2006; Campbell and Cary 2001; Chang et al. 2002, 2005; Christopher and Christopher 2004).

13.2 Composition of Petroleum Oil

Petroleum is recovered principally through oil drilling, and it's refined and fractionated into a range of commercial products.

In true sense, fossil oil includes inflammable liquid consisting of a fancy mixture of hydrocarbons of varied molecular weights and alternative liquid organic compounds; however, in common usage it includes all liquid (e.g. pentane and heavier ones), gaseous (e.g. methane, ethane, propane, butane, etc.) and solid (e.g. paraffin) hydrocarbons (Speight 1999). The hydrocarbons in fossil oil are largely alkanes, cycloalkanes and numerous aromatic hydrocarbons, whereas the other organic compounds contain varied metals like iron, nickel, copper, etc. (Speight 1999). Precisely, the molecular composition of hydrocarbons may vary from formation to formation. Chemically, fossil oil contains paraffin (15–60%), hydrocarbon (30–60%), aromatics (3–30%) and mineral (6%); however, the relative proportion of fossil oil largely varies from oil to oil (Mabro 2006). Although the constituents of fossil oil may vary, the elementary compositions may be presented as shown in Table 13.1 (Hyne 2001).

Table 13.1 Elementary compositions of fossil oil

Name of the constituents	Percentage (%)
Carbon	83–87
Hydrogen	10–14
Oxygen	0.05–1.5
Nitrogen	0.1–2
Sulphur	0.05–6
Metals	<0.1

13.3 Petroleum Oil Pollution, Environment and Health

The illegal disposal of crude oil is also one of the major causes of environmental hazard with international ramifications (Blodgett 2001; Guermouche et al. 2015). The discharge of oil into the surrounding causes environmental concern and attracts the general public attention (Roling et al. 2002). Petroleum oil and PAHs (polyaromatic hydrocarbons) have a widespread impact on the body, as prolonged exposure to crude oil might induce liver and excretory organ diseases and bone marrow injury or might cause the event of cancer (Crebelli et al. 1995; Mandri and Lin 2007; Guermouche et al. 2015). Processed engine oil contains additional heavy metals and serious PAHs and therefore contributes a lot more to chronic hazards as well as mutagenicity and carcinogenicity as compared to unused engine oil (Boonchan et al. 2000).

Unlike the claims created by the oil-exploring industries concerning the security measures taken for safe unlash of treated effluents to the surroundings, the digital and print media besides restricted scientific study claims environmental problems associated with oil contamination within the region notably within the abandoned drilling sites (Das et al. 2004; Yenn et al. 2014).

Oil contamination of soil and water isn't solely a regional issue but a worldwide issue of concern. Not only soil, the aquatic system, notably the marines, is the foremost at risk of oil spillage (Cairns and Buikema 1984). Marine oil spillage may have an effect on organisms present therein by direct toxicity or by physical stress (Perry 1980). Oil spills usually will cause varied damages to the marsh vegetation. Oil spill in water body forms a surface slick whose elements will follow several pathways. Some might pass into the mass of water, and proof suggests they will persist for an extended time before their degradation by microorganisms within the water body. The slick sometimes becomes additional viscous and forms water-in-oil emulsion. Oil in water causes depletion of dissolved gases because of transformation of the organic element into inorganic compounds, loss of biodiversity and eutrophication. Toxicity in fishes includes blood disease, dermal dysplasia and pasteurellosis (Beeby 1993). It also affects plant physiology in terms of stem height, photosynthetic rate, overall plant biomass, germination, etc. leading to their death (Krebs and Tanner 1981; Onwurah 1999). Oil spill may increase the mortality rate of a population by damaging the reproductive capacity of the respective population (Hall et al. 2006; Tiido et al. 2006).

It conjointly affects soil fertility by altering the mineral and organic matter content, ion exchange capability, salinity, pH, etc. However, the size of impact depends on the number and kind of oil spilled (Onwurah et al. 2007). As petroleum oil creates anaerobic condition in the soil, coupled to water logging and acidic metabolites, the result is high accumulation of aluminium and manganese ions, which are toxic to plant growth (Onwurah et al. 2007). Petroleum oil hydrocarbons may cause DNA damage, resulting in carcinogenesis, mutagenesis and impairment of reproductive capacity (Short and Heintz 1997). The risk of drinking water contaminated by crude oil can be extrapolated from its effect on rats that developed haemorrhagic tendencies after exposure to water-soluble components of crude oil (Onwurah 2002). Volatile components of crude oil after a spill may lead to asthma, bronchitis and other liver and kidney diseases (Kaladumo 1996; Anozie and Onwurah 2001).

13.4 Countermeasures for Oil Pollution

Conventionally physical, chemical and biological ways are used for the treatment of oil spillage.

13.4.1 *Physical Methods*

Removal of oil spill in soil and water body may involve mechanical removal methods such as skimming, manual removal (wiping), water flushing, etc. According to the recent USEPA 2015 report, current mechanical methods other than open burning of petroleum oil typically recover up to only 10–15% of oil after a massive oil spill, whereas burning of oil may reduce up to 98% but with a massive air pollution (OTA 1990; NAS 2005; USEPA 2015). Although standard ways, like physical removal of spilled oil, usually are the primary response choice, they rarely succeed in complete clean-up of oil spills.

13.4.2 *Chemical Methods*

Chemical ways, notably dispersants (synthetic surfactants), are more often utilized in several countries as a response choice (USEPA 1999). However, chemical ways haven't been extensively used due to the disagreement concerning their effectiveness (recover no over 8% of usable oil) and also the issues of their toxicity and long-run environmental effects because of their toxic formulation (USEPA 1999; <https://www.restorethegulf.gov/>; Hemmer et al. 2011). Synthetic surfactants possess both hydrophilic and hydrophobic groups which form oil-surfactant micelles by decreasing tension between water and oil interface which may lead to further degradation of droplets of the micelles in the water body towards physical and microbial degradation (Hemmer et al. 2011).

13.4.3 *Biological Methods*

Biological strategies have emerged as promising technology nowadays, notably as a secondary treatment choice for the clean-up of oil spillage. Bioremediation has been outlined as a treatment method of contaminated environment with the assistance of living organisms to bring back to its original state (OTA 1991; Ivanova et al. 2015). This technology is based on the fact that a considerable amount of oil components are mostly biodegradable in nature (Atlas 1981; Atlas 1984; Prince

1993) and hence can be augmented by adding nutrients. Bioremediation has many potential benefits over standard technologies such as it is very cost-effective and eco-friendly in terms of its finished product.

The success rate of bioremediation process is dependent on the capacity to establish favourable conditions in the contaminated site which enhances the rate of hydrocarbon degradation. The vital demand for the success of bioremediation is the presence of indigenous microorganisms with the suitable metabolic activities for degradation of specific type of fossil oil.

There are diverse group of microorganisms known to be capable of degrading variety of target constituents present in oily sludge (Eriksson et al. 2002; Barathi and Vasudevan 2001; Mishra et al. 2001). Out of so many reported microorganisms, various strains of *Pseudomonas* sp. are mostly reported by many researchers as a potential fossil oil degrader (Fall et al. 1979; Johnson et al. 1996; Guermouche et al. 2015). Some of the field trial reports claim successful bioremediation of oil-contaminated nearby areas of abandoned drill sites located particularly in Gelakey, Amguri, Lakwa and Borholla regions of Assam, India, by augmenting contaminated sites with *Pseudomonas* strains designated as N3 and N4 along with plant species *Gmelina arborea*, *Tectona grandis*, *Michelia champaca* and *Azadirachta indica* (Yenn et al. 2014).

Apart from *Pseudomonas* sp., there are some other bacteria which include *Yokenella* sp., *Alcaligenes* spp., *Roseomonas* sp., *Stenotrophomonas* sp., *Acinetobacter* spp., *Flavobacterium* sp., *Streptococcus* sp., *Providencia* sp., *Sphingobacterium* sp., *Capnocytophaga* sp., *Moraxella* sp. and *Bacillus* sp.; some fungi, viz. *Alternaria* sp., *Aspergillus* sp., *Cladosporium* sp., *Eupenicillium* sp. and *Paecilomyces* sp.; etc. are known as potential hydrocarbon degraders (Rusansky et al. 1987; Sharidah et al. 2000; Bhattacharya et al. 2002; Juwarkar 2012; Bujang et al. 2013; Ameen et al. 2015). The component of media plays a vital role in microbial growth and proliferation. Hence, optimization and improvement of media components and growth parameters are crucial factors for obtaining enhanced rate of hydrocarbon degradation and are advocated by many researchers (Xia et al. 2006; Vieira et al. 2009; Dongfeng et al. 2011; Janani et al. 2014).

13.5 Microorganisms in Petroleum Degradation and Their Sources

Hydrocarbon bioremediation of contaminated sites by indigenous microbial population allows the conversion of toxic substances into less or nontoxic forms which might represent one of the primary mechanisms by which hydrocarbon products are removed from the contaminated environment inexpensively (Atlas 1981). The capability of microorganisms to emulsify hydrocarbon by producing surface-active agents is one of the most important characteristics of hydrocarbon-degrading

bacteria. Such surface-active agents cause dispersion of hydrocarbons in water emulsions leading to the formation of micro-droplets (micelles) which may be ingested by the microbial cells for further degradation with the help of certain enzymes. One such important enzyme involved in hydrocarbon degradation is oxygenase which converts complex chain of hydrocarbons into smaller and simpler forms which finally enters into peripheral metabolic cycles as shown in Figs. 13.1 and 13.2 (Hommel 1990; Cerniglia 1992; Yakimov et al. 1995; Adebusoye et al. 2008; Ibrahim et al. 2013). A number of microorganisms, viz. *Bacillus* sp., *Corynebacterium* sp., *Edwardsiella* sp., *Staphylococcus* sp., *Pseudomonas* sp., *Citrobacter* sp., *Micrococcus* sp., *Cladosporium* sp., *Acetobacterium* sp., *Mucor* sp., *Penicillus* sp., *Monosporium* sp., *Aspergillus* sp., *Alternaria* sp., *Mucor* sp., etc., are reported to be hydrocarbon degraders by various researchers (Fall et al. 1979; Okpokwasili and Odokuma 1986; Okpokwasili and Okorie 1988; Rusansky et al. 1987; Johnson et al. 1996; Campbell and Cary 2001; Bhattacharya et al. 2002;

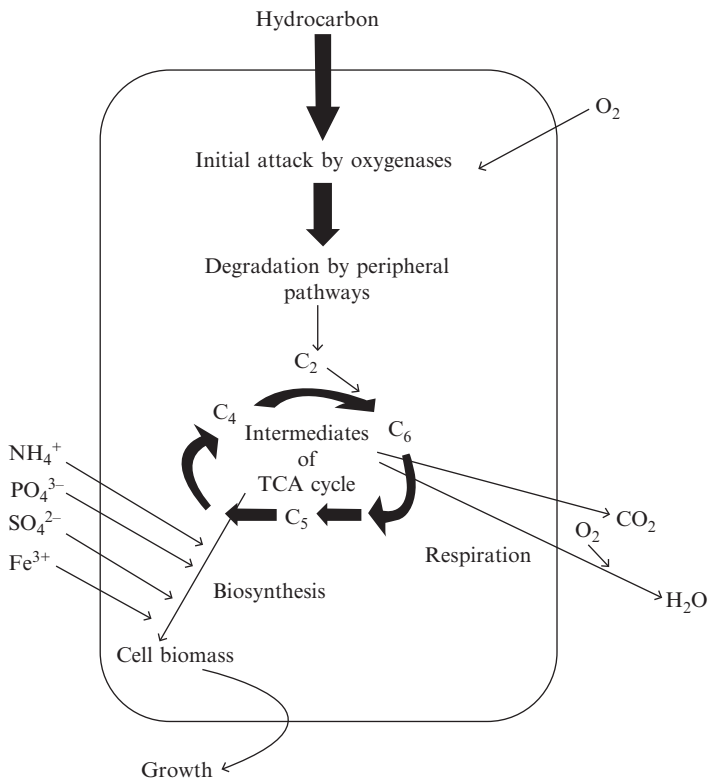


Fig. 13.1 Basic principle behind aerobic biodegradation of hydrocarbons by microorganisms. (Adopted from Das and Chandran 2011)

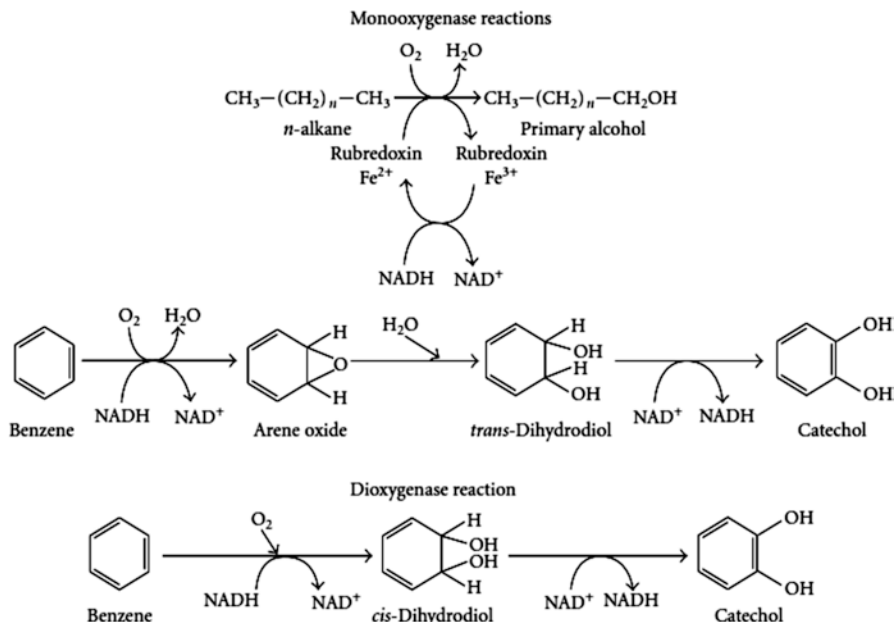


Fig. 13.2 Hydrocarbon degradation process by enzymatic action. (Adopted from Das and Chandran 2011)

Chang et al. 2002; Christopher and Christopher 2004; Chang et al. 2005; Bach et al. 2005; Biddle et al. 2006; Mandri and Lin 2007; Su et al. 2011; Luo et al. 2012; Yenn et al. 2014; Ameen et al. 2015; Jia et al. 2016).

Microorganisms, which are capable of hydrocarbon degrading, are believed to be capable of tolerating stress conditions to some extent, as hydrocarbon-contaminated sites itself provide hostile situations for the growth and proliferation of such microorganisms. Besides, very limited numbers of scientific reports dealing with microbial hydrocarbon degradation under stressed conditions are available (Sorensen et al. 2010; Tapilatu et al. 2010; Martino et al. 2012; Abed et al. 2014; Li et al. 2016). One such literature shows the existence of hydrocarbon degradation *Pseudomonas* sp. and *Variovorax* sp. at a temperature as low as 0 °C in fuel-contaminated soils of Greenland high Arctic region (Sorensen et al. 2010). Selection of such bacteria with stress-tolerant capability to degrade mono- or polyaromatic compounds by synthesizing biosurfactants and biopolymers that may play a certain role in enhanced stress tolerance could be a good approach to find a suitable bio-augmentation agent (Martino et al. 2012; Li et al. 2016). A detailed list of hydrocarbon-degrading microorganisms and their source of isolation is given in Table 13.2.

Table 13.2 Name of the microorganisms and their source of isolation along with the reported values of percentage hydrocarbon degradation are shown

Name of the microorganisms	Source	Hydrocarbon degradation (%)	References
<i>Alcaligenes faecalis</i> , <i>Candida tropicalis</i> (*the first report on the isolation of hydrocarbon-degrading microorganisms from Amazonian soil)	Artificially augmented Amazonian rainforest soil samples	–	Bastos et al. (2000)
<i>Bacillus subtilis</i>	Contaminated soil of Kuwait	–	Sharidah et al. (2000)
<i>Alcanivorax borkumensis</i>	Marine water	–	Golyshin et al. (2003)
<i>Gordonia</i> sp., <i>Aeromicrobium</i> sp., <i>Brevibacterium</i> sp., <i>Dietzia</i> sp., <i>Paecilomyces</i> sp., <i>Burkholderia</i> sp., <i>Yarrowia</i> sp., <i>Aspergillus</i> sp., <i>Fusarium</i> sp., <i>Penicillium</i> sp., <i>Neosartorya</i> sp., <i>Talaromyces</i> sp., <i>Graphium</i> sp., <i>Pichia</i> sp. and <i>Amorphoteca</i> sp.,	Contaminated soils of Indonesia	4.9–22%	Chaillan et al. (2004)
Microcosm of six microbes (names not mentioned)	Contaminated oil refinery plant in China	63.2 ± 20.1%	Ma et al. (2015)
<i>Alkanindiges</i> sp., <i>Arthrobacter</i> sp., <i>Pseudomonas</i> sp., <i>Mycobacterium</i> sp. and <i>Rhodococcus</i> sp.	Contaminated soils of Northern China	–	Sun et al. (2014)
<i>Alternaria alternata</i> , <i>Cladosporium</i> sp., <i>Aspergillus terreus</i> , <i>Eupenicillium hirayamae</i> , <i>Sphaerospermum</i> sp., <i>Paecilomyces variotii</i>	Contaminated mangrove sediments from Red Sea coast of Saudi Arabia	28–56%	Ameen et al. (2015)
<i>Stenotrophomonas</i> sp., <i>Pseudomonas</i> sp.	Dredged sediments of a river estuary in Italy	43–95%	Gregorio et al. (2016)
<i>Polaromonas</i> sp., <i>Sphingomonas</i> sp., <i>Alcaligenes</i> sp., <i>Caulobacter</i> sp. and <i>Variovorax</i> sp.	Uncontaminated Arctic soil	–	Eriksson et al. (2002)
<i>Nocardia otitidiscaviarum</i>	Contaminated desert soil of Iran	–	Zeinali et al. (2007)
<i>Mycobacterium</i> sp.	Soil	–	Miller et al. (2004)
<i>Mycobacterium</i> sp.	Uncontaminated Natural Park Soil of Schwa of Germany	–	Kim et al. (2005)
<i>Acinetobacter</i> sp., <i>Aeromonas</i> sp., <i>Alcaligenes</i> sp., <i>Bacillus</i> sp., <i>Kocuria</i> sp. (<i>Micrococcus</i>), <i>Ochrobactrum</i> sp., <i>Pseudomonas</i> sp. and <i>Xanthomonas</i> sp.	Contaminated Patagonian soil	0.028–100%	Peressutti et al. (2003)

(continued)

Table 13.2 (continued)

Name of the microorganisms	Source	Hydrocarbon degradation (%)	References
<i>Rhodococcus</i> sp. and <i>Pseudomonas</i> sp.	Contaminated soils of Kaluga, Kirov, Moscow	0–95%	Baryshnikova et al. (2001)
Not defined	Uncontaminated soils of Western Siberia (Arctic region)	3.8–51.2%	Belousova et al. (2002)
<i>Alcanivorax borkumensis</i>	North Sea, Atlantic Ocean, Mediterranean Sea, Sea of Japan, South China Sea and the Antarctic	80–90%	Golyshin et al. (2003)
<i>Nocardia otitidiscaviarum</i>	Soil contaminated with wastewater of a petroindustrial site in Iran	10–55%	Zeinali et al. (2007)
<i>Afipia</i> sp., <i>Janthinobacterium</i> sp., <i>Leptothrix</i> sp., <i>Massilia</i> sp., <i>Methylobacterium</i> sp., <i>Rhizobium</i> sp., <i>Sinorhizobium</i> sp. and <i>Thiobacillus</i> sp.	Uncontaminated soil of Arizona	88%	Bodour et al. (2003)
<i>Mucor mucedo</i>	Uncontaminated soils of Shenfu irrigation area, China	87%	Jia et al. (2016)
<i>Haloarcula</i> sp., <i>Haloferax</i> sp. (*the first report on the potential role of halophilic archaea belonging to the genera <i>Haloarcula</i> and <i>Haloferax</i>)	Uncontaminated pond water of Camargue, France	32–95%	Tapilatu et al. (2010)
<i>Marinobacter</i> sp., <i>Pseudomonas</i> sp., <i>Halomonas</i> sp., <i>Hahella</i> sp. and <i>Alcanivorax</i> sp.	Contaminated sediments from coastal region in Oman	67%	Abed et al. (2014)
<i>Pseudomonas</i> sp. and <i>Variovorax</i> sp.	Contaminated soils from Station Nord (St. Nord) in Greenland high Arctic region	70%	Sorensen et al. (2010)

13.6 Microbial Degradation of Petroleum Hydrocarbon

13.6.1 Enzymatic Pathways

Short-chain hydrocarbons are considered as the most biodegradable petroleum hydrocarbons. Hydrocarbons with C₅–C₁₀ carbon numbers are homologues of solvents that tend to disrupt lipid membranes of hydrocarbon-degrading microorganisms, and hence, they are reported to have inhibitory action. Waxes, with alkane

of C_{20} – C_{40} , are solids with hydrophobic nature at room temperature and hence are less biodegradable in nature (Atlas and Bartha 1973). Literature suggests that the majority of organic pollutants completely or optimally degrade under aerobic condition. Enzymes such as oxygenases and peroxidases play a vital role in hydrocarbon degradation, and these are responsible for initial intracellular oxidation of organic pollutants. The organic pollutants are then converted into intermediates of central intermediary metabolism such as the Krebs cycle in a stepwise manner. Cell biomass are then synthesized by central precursor metabolites such as acetyl CoA, succinate, pyruvate, etc. Sugars are the essential source of carbon, which is required for different types of biosynthesis and growth of the microbes, and are synthesized by gluconeogenesis (Fritsche and Hofrichter 2000). Figures 13.1 and 13.2 show the initial action on hydrocarbon by oxygenases (Das and Chandran 2011).

13.6.2 Role of Biosurfactant in Petroleum Degradation

As discussed earlier, the ability to produce biosurfactants by microbes is the most important feature of a potential hydrocarbon degrader. Biosurfactants are a complex mixture of biomolecules such as proteins, exopolysaccharides, fatty acids, amino acids, glycolipids, etc. with hydrophobic and hydrophilic components that emulsify hydrocarbon by minimizing the interstitial surface tension which makes them suitable for bioremediation and microbial enhanced oil recovery process (Ibrahim et al. 2013). Thus, surfactant subsequently increases the aqueous solubility of oily contaminants and makes them available for microorganisms for further degradation (Karanth et al. 1999). On the other hand, synthetic surfactants such as SDS (sodium dodecyl sulphate), LAS (linear alkylbenzene sulphonate), Brij 30, Tween 80, etc. used for the treatment of oil contaminants are often toxic, which also act as an additional source of contamination (Bognolo 1998). Microbial surfactants have analogous properties to that of synthetic surfactants but are biodegradable in nature and can be produced at the contaminated site (in situ) (Cha 2000). Isolation of microorganisms with desired capabilities to emulsify and solubilize hydrophobic contaminants both ex situ and in situ is a major benefit over competitors in contaminated areas (Reddy and Singh 1982; Vecchioli et al. 1990; Chattré et al. 1996; Cassidy and Hudak 2001). Nowadays, these are not only important for bioremediation purpose but also area of research for microbial enhance oil recovery from wells. Such process involves the implementation of microbes directly or microbial surfactants in the well which helps in reducing the viscosity of oil to increase the free flow through the pipelines and also stabilizes fuel water–oil emulsions (Ghurye and Vipulanandan 1994; Makkar and Cameotra 1997; Bognolo 1998).

Contact angle of a fluid with a solid surface is an index used to examine the action of biosurfactant on the rock surface wettability (i.e. the interaction between fluid and solid surface) (Ibrahim et al. 2013). It is defined as the angle between the tangent to the periphery of the point of fluid contact with the solid and the surface of the solid in the direction where the droplet exists as shown in Fig. 13.3. Contact

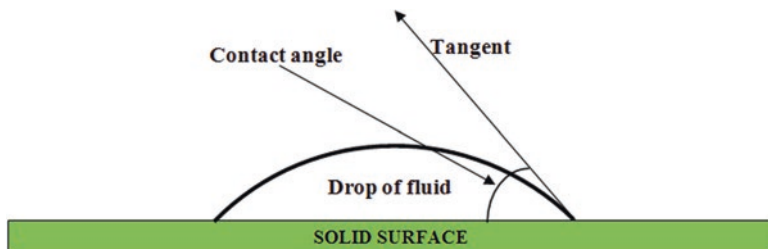


Fig. 13.3 The contact angle formed by a drop of fluid on a solid surface

angle less than 90° for surfactant-containing liquid is considered to be wet on solid surface, and a contact angle greater than 90° is said to be non-wetting (i.e. the liquid which does not wet the solid surface). The alteration in wettability has been projected as one of the mechanisms of MEOR (Alvarez et al. 2015).

Some of the field trials carried out by researchers in Poland, Holland, the Czech Republic, the United States, Romania and Hungary reported significant enhancement in oil recovery by MEOR process (Karanth et al. 1999). Apart from the cost-effective nature of biosurfactants, they are also widely used in food, pharmaceutical and cosmetic industry which makes it an important microbial product of commercial importance (Batista et al. 2006).

A review work carried out by Satpute et al. (2010) represents marine biosphere as a wealthy natural source of flora and fauna with functional commercial-grade bioactive compounds, biosurfactants/bioemulsifiers, etc. which may be used for the purpose of bioremediation. Marine microorganisms such as *Bacillus*, *Acinetobacter*, *Halomonas*, *Myroides*, *Pseudomonas*, *Corynebacterium*, *Arthrobacter* and *Alteromonas* sp. were reported for their potential application for the production of biosurfactants and exopolysaccharides. (Filonov et al. 2004; Coral and Karagoz 2005; Das and Mukherjee 2007; Simpson et al. 2011).

Various species of the genus *Pseudomonas* are probably the most widely studied hydrocarbon-degrading bacteria. Existing literature shows the role of various species of *Pseudomonas* isolated from contaminated sites in hydrocarbon degradation (Baryshnikova et al. 2001; Filonov et al. 2004; Shin et al. 2006; Niepceron et al. 2010; Aresta et al. 2010; Nie et al. 2010; Singh and Malik 2013; Li et al. 2016). Their source of isolation even ranges from hot springs (Perfumo et al. 2006), marine water (Satpute et al. 2010) and cow dung (Singh and Fulekar 2010) up to the Arctic region (Sorensen et al. 2010).

Some of the researchers studied the capability of various *Pseudomonas* strains in BTEX (benzene, toluene, ethylene and xylene), naphthalene and TPH degradation, surface tension reduction of petroleum oil, formation of biofilm on hydrocarbon-supplemented medium, formation of rhamnolipid, glycolipid-based surfactants, etc. (Baryshnikova et al. 2001; Perfumo et al. 2006; Kim and Jaffé 2008; Zhang et al. 2011; Cébron et al. 2011; John and Okpokwasili 2012; Martino et al. 2012; Li et al. 2013; Oyetibo et al. 2013; Pedetta et al. 2013; Dudášová et al. 2014;

Pacwa-Płociniczak et al. 2014; Ma et al. 2015). This report advocates their capability to achieve 20–100% of hydrocarbon degradation within 7–30 days of incubation. But physicochemical parameters such as pH, temperature, salinity, type of culture medium used, carbon source, etc. may influence the yield of biosurfactant production in industrial scale (Lang 2002; Batista et al. 2006).

13.7 Genes Involved in Petroleum Degradation

Molecular techniques not only play a vital role for the identification of petroleum degraders but are also largely used for the characterization and identification of different genes involved in hydrocarbon degradation. Genes responsible for hydrocarbon degradation may be present in either genomic or plasmid DNA or may be in both at the same time in a certain microorganism. Plasmid curing is a convenient tool for the determination of the involvement of plasmid-encoded genes in hydrocarbon degradation (Karpagam and Lalithakumari 1999; Liu et al. 2004; Vasudevan et al. 2007; Kumar and Gopal 2015; John and Okpokwasili 2012). Reports also show presence of certain genes in the genomic DNA of some microbes that are involved in hydrocarbon degradation (Kim et al. 2007; Quatrini et al. 2008; Weelink et al. 2009; Li et al. 2011; Sun et al. 2014).

In contrary to the above, there are also reports which show contribution of both genomic and plasmid DNA-encoded genes in the same organism in hydrocarbon degradation (Fondi et al. 2013). A brief list of important genes responsible for microbial hydrocarbon degradation is shown in Table 13.3.

13.8 Conclusions

Even though widespread research has been conducted on petroleum bioremediation during the last decade, the usefulness of these tools has only rarely been convincingly established, and on the commercially available bioremediation products, the literatures are nearly lacking with supportive evidence of success. Existing literatures chiefly demonstrated the assessment of factors affecting bioremediation under laboratory condition. Out of these, very few numbers of literature reports their implementation in field trials at pilot-scale convincingly demonstrated this technology. Only 27 commercially available bioremediation agents such as Inipol EAP22, BIOREN 1 and 2, Oil Spill Eater II® (OSE II), ENVIROZYME BR, BioCATalystIOS-500, Petro-Clean, IOS-500, Micro-Blaze, WMI-2000, etc. are listed on the NCP schedule of USEPA till March 2015 (USEPA 2015). But the scientific community is still trying to screen out more effective microorganisms of the same type or sometimes a consortium of different types of microbes for bioremediation of a hydrocarbon-contaminated site considering their advantages over commercially available synthetic bioremediation agents. In conclusion, as the petroleum

Table 13.3 A brief list of hydrocarbon-degrading genes and their source organisms

Name of the microbes	Genes involved	Encodes for	References
<i>Rhodococcus</i> sp., <i>Gordonia</i> sp.	alkB	Non-haem iron-containing alkane monooxygenases and hydroxylases	Kim et al. (2007), Quatrini et al. (2008)
<i>Acinetobacter</i> sp., <i>Staphylococcus haemolyticus</i>	C23O	Cytochrome C230	Onur et al. (2015)
<i>Betaproteobacteria</i> sp., <i>Sterolibacterium denitrificans</i>	bssA	α -Subunit of Bss enzyme	Weelink et al. (2009)
<i>Proteobacteria</i> sp., <i>Firmicutes</i> sp., <i>Acidobacteria</i> sp., <i>Actinobacteria</i> sp., <i>Deferribacteres</i> sp., <i>Bacteroidetes</i> sp., <i>Thauera chlorobenzoica</i>	bamA	6-oxocyclohex-1-ene-1-carbonyl-CoA hydrolase	Sun et al. (2014), Li et al. (2011)
<i>Gammaproteobacteria</i> sp.		Catechol	Sei and Fathepure (2009)
<i>Acinetobacter</i> sp., <i>Alcanivorax</i> sp.	almA	Flavin-binding monooxygenase	Wang and Shao (2012)
	arfA	Arthofactin	Roongsawang et al. (2003), Das et al. 2008
<i>B. tequilensis</i> , <i>E. coli</i> , <i>B. subtilis</i> , <i>Aeromonadaceae</i> sp., <i>Bacillaceae</i> sp., <i>Enterobacteriaceae</i> sp., <i>Gordoniaceae</i> sp., <i>Pseudomonadaceae</i> sp.	sfp	4'-phosphopantetheinyl transferase protein	Porob et al. (2013), Anburajan et al. (2015), Ndlovu et al. (2016)
	sfpO	Phosphopantetheinyl transferase	Anburajan et al. (2015)
<i>Aeromonadaceae</i> sp., <i>Bacillaceae</i> sp., <i>Enterobacteriaceae</i> sp., <i>Gordoniaceae</i> sp. and <i>Pseudomonadaceae</i> sp.	rhlB	Rhamnosyltransferase subunit B	Ndlovu et al. (2016)
<i>Aeromonadaceae</i> sp., <i>Bacillaceae</i> sp., <i>Enterobacteriaceae</i> sp., <i>Gordoniaceae</i> sp. and <i>Pseudomonadaceae</i> sp.	bamC	Bacillomycin C	Ndlovu et al. (2016)
<i>Pseudomonas</i> sp.	rhl	Rhamnolipid	Pacwa-Płociniczak et al. (2014)
<i>A. xylosoxidans</i>	bphA		Pacwa-Płociniczak et al. (2014)
<i>Pseudomonas putida</i>	ndoB	Iron-sulphur protein	Hamann et al. (1999)
<i>Rhodococcus</i> sp.	thmA		Kim et al. (2007)
<i>Rhodococcus</i> sp.	PrmA		Kim et al. (2007)

hydrocarbon pollutant creates a stressed environment for the growth and proliferation of indigenous microbial species, hence research on finding out novel hydrocarbon-degrading microorganisms with stress-tolerant potential would be an added advantage.

Acknowledgements The author acknowledges DBT Delcon facility for providing access to e-journals at the Centre for Biotechnology and Bioinformatics, Dibrugarh University, India.

References

- Abed RMM, Al-Sabahi J, Al-Maqrashi F, Al-Habsi A, Al-Hinai M (2014) Characterization of hydrocarbon-degrading bacteria isolated from oil-contaminated sediments in the Sultanate of Oman and evaluation of bioaugmentation and biostimulation approaches in microcosm experiments. *Int Biodeterioration Biodegrad* 89:58–66
- Adebusoye SA, Amund OO, Ilori MO, Domeih DO, Okpuzor J (2008) Growth and biosurfactant synthesis by Nigerian hydrocarbon-degrading estuarine bacteria. *Int J Tropical Biol* 56:1603–1611
- Alvarez VM, Jurelevicius D, Marques JM, de Souza PM, de Araújo LV, Barros TG, Alves de Souza ROM, Freire DMG, Seldin L (2015) *Bacillus amyloliquefaciens* TSBSO 3.8, a biosurfactant-producing strain with biotechnological potential for microbial enhanced oil recovery. *Colloids Surf B: Biointerfaces* 136:14–21
- Ameen F, Moslem M, Hadi S, Al-Sabri AE (2015) Biodegradation of diesel fuel hydrocarbons by mangrove fungi from Red Sea Coast of Saudi Arabia. *Saudi J Biol Sci* 23(2):211–218
- Anburajan L, Meena B, Raghavan RV, Shridhar D, Joseph TC, Vinithkumar NV, Dharani G, Dheenan PS, Kirubakaran AR (2015) Heterologous expression, purification, and phylogenetic analysis of oil-degrading biosurfactant biosynthesis genes from the marine sponge-associated *Bacillus licheniformis* NIOT-06. *Bioprocess Biosyst Eng* 38:1009–1018
- Anozie O, Onwurah INE (2001) Toxic effects of Bonny light crude oil in rats after ingestion of contaminated diet. *Nigerian J Biochem Mol Biol(Proceedings Supplement)* 16(3):1035–1085
- Aresta M, Acquaviva MI, Baruzzi F, Lo Noce RM, Matarante A, Narracci M, Stabili L, Cavallo RA (2010) Isolation and characterization of polyphenols-degrading bacteria from olive-mill wastewaters polluted soil. *World J Microbiol Biotechnol* 26:639–647
- Atlas RM (1981) Microbial degradation of petroleum hydrocarbons: an environmental perspective. *Microbiol Rev* 45:180–209
- Atlas RM (1984) Petroleum microbiology. Macmillan Publishing Company, New York
- Atlas RM, Bartha R (1973) Simulated biodegradation of oil slicks using oleophilic fertilizers. *Environ Sci Technol* 7:538–541
- Bach QD, Kim SJ, Choi SC, Oh YS (2005) Enhancing the intrinsic bioremediation of PAH-contaminated anoxic estuarine sediments with biostimulating agents. *J Microbiol* 43:319–324
- Barathi S, Vasudevan N (2001) Utilization of petroleum hydrocarbons by *Pseudomonas fluorescens* isolated from a petroleum-contaminated soil. *Environ Int* 26:413–416
- Baryshnikova LM, Grishchenkov VG, Arinbasarov MU, Shkidchenko AN, Boronin LM (2001) Biodegradation of oil products by individual degrading strains and their associations in liquid media. *Appl Biochem Microbiol* 37(5):463–468
- Bastos AEB, Moon DH, Rossi A, Trevors JT, Tsai SM (2000) Salt tolerant phenol degrading microorganisms isolated from Amazonian soil samples. *Arch Microbiol* 174:346–352
- Batista SB, Mounteer AH, Amorim FR, Totola MR (2006) Isolation and characterization of biosurfactant/bioemulsifier-producing bacteria from petroleum contaminated sites. *Bioresour Technol* 97:868–875

- Beeby A (1993) Applying ecology, 7th edn. Chapman and Hall Publishers, New York
- Belousova NI, Baryshnikova LM, Shkidchenko AN (2002) Selection of microorganisms capable of degrading petroleum and its products at low temperatures. *Appl Biochem Microbiol* 38(5):437–440
- Bhattacharya D, Sarma PM, Krishnan S, Mishra S, Lal B (2002) Evaluation of genetic diversity among *Pseudomonas citronellolis* strains isolated from oily sludge-contaminated sites. *Appl Environ Microbiol* 69(3):1435–1441
- Biddle JF, Lipp JS, Lever MA, Lloyd KG, Sørensen KB, Andersonc R, Fredricks HF, Elvert M, Kelly TJ, Schrag DP, Sogin ML, Brenchley JE, Teske A, House CH, Hinrichs KU (2006) Heterotrophic Archaea dominate sedimentary subsurface ecosystems off Peru. *Proc Natl Acad Sci U S A* 103:3846–3851
- Blodgett WC (2001) Water-soluble mutagen production during the bioremediation of oil-contaminated soil. *Florida Scientist* 60(1):28–36
- Bodour AA, Wang JM, Brusseau ML, Maier RM (2003) Temporal change in culturable phenanthrene degraders in response to long-term exposure to phenanthrene in a soil column system. *Environ Microbiol* 5(10):888–895
- Bognolo G (1998) Biosurfactants as emulsifying agents for hydrocarbons. *Colloid Surf A Physicochem Eng Asp* 152:41–52
- Boonchan S, Britz ML, Stanley GA (2000) Degradation and mineralization of high-molecular weight polycyclic aromatic hydrocarbons by defined fungal-bacterial cocultures. *Appl Environ Microbiol* 66(3):1007–1019
- Briganti F, Pessione E, Giunta C, Scozzafava A (1997) Purification, biochemical properties and substrate specificity of a catechol 1, 2-dioxygenase from a phenol degrading *Acinetobacter radioresistens*. *FEBS Lett* 416:61–64
- Bujang M, Ibrahim NA, EA R (2013) Biodegradation of oily wastewater by pure culture of *Bacillus cereus*. *ARPN J Agric Biol Sci* 8:108–115
- Butler CS, Mason JR (1997) Structure-function analysis of the bacterial aromatic ring hydroxylating dioxygenases. *Adv Microb Physiol* 38:47–84
- Cairns J, Buikema AL (1984) Restoration of habitats impacted by oil spills. *Ann Arbor Science Publishers/Butterworth, Boston*
- Campbell BJ, Cary SC (2001) Characterization of a novel Spirochete associated with the hydrothermal vent polychaete annelid, *Alvinella pompejana*. *Appl Environ Microbiol* 67:110–117
- Cassidy DP, Hudak AJ (2001) Microorganism selection and biosurfactant production in a continuously and periodically operated bioslurry reactor. *J Hazard Mater* 84:253–264
- Cébron A, Louvel B, Faure P, Lanord CF, Chen Y, Murrell JC, Leyval C (2011) Root exudates modify bacterial diversity of phenanthrene degraders in PAH-polluted soil but not phenanthrene degradation rates. *Environ Microbiol* 13(3):722–736
- Cerniglia CE (1992) Biodegradation of polycyclic aromatic hydrocarbons. *Int Biodegradation* 3:351e368
- Cha DK (2000) The effect of biosurfactants on the fate and transport of nonpolar organic contaminants in porous media. *Environ Eng* 20:1–17
- Chaillan F, Le Flèche A, Bury E, Phantavong YH, Grimont P, Saliot A, Oudot J (2004) Identification and biodegradation potential of tropical aerobic hydrocarbon-degrading microorganisms. *Res Microbiol* 155(7):587–595
- Chang R (1998) Chemistry, 6th edn. McGraw-Hill Companies, Inc, New York
- Chang BV, Shiung LC, Yuan SY (2002) Anaerobic biodegradation of polycyclic aromatic hydrocarbon in soil. *Chemosphere* 48:717–724
- Chang W, Um Y, Hoffman B, Holoman TRP (2005) Molecular characterization of polycyclic aromatic hydrocarbon (PAH)-degrading methanogenic communities. *Biotechnol Prog* 21:682–688
- Chattre S, Purohit H, Shanker R, Khanna P (1996) Bacterial consortia for crude oil spill remediation. *Water Sci Technol* 34:187–193
- Christopher WK, Christopher LK (2004) Bacterial succession in a petroleum land treatment unit. *Appl Environ Microbiol* 70(3):1777–1785

- Coral G, Karagoz S (2005) Isolation and characterization of phenanthrene degrading bacteria from a petroleum refinery soil. *Ann Microbiol* 55:255–259
- Crebelli R, Conti L, Crochi B, Carere A, Bertoli C, Giacomo ND (1995) The effect of fuel composition on the mutagenicity of diesel engine exhaust. *Mutation Res* 346:167–172
- Das N, Chandran P (2011) Microbial degradation of petroleum hydrocarbon contaminants: an overview. *Biotechnol Res Int* 1:1–13. <https://doi.org/10.4061/2011/941810>
- Das K, Mukherjee AK (2007) Crude petroleum-oil biodegradation efficiency of *Bacillus subtilis* and *Pseudomonas aeruginosa* strains isolated from a petroleum-oil contaminated soil from North-East India. *Bioresour Technol* 98:1339–1345
- Das SN, Swamy YV, Rao KK, Misra VN (2004) Pollution in urban environment. Proceedings National Seminar on Pollution in Urban Environment (NSPUIE 2004): Regional Research Laboratory, Bhubaneswar. ISBN:8177648578
- Das P, Mukherjee S, Sen R (2008) Genetic regulations of the biosynthesis of microbial surfactants: an overview. *Biotechnol Genet Eng Rev* 25(1):165–186
- Dongfeng Z, Weilin W, Yunbo Z, Qiyu L, Haibin Y, Chaocheng Z (2011) Study on isolation, identification of a petroleum hydrocarbon degrading bacterium *Bacillus fusiformis* sp. and influence of environmental factors on degradation efficiency. *Chin Pet Process Pe Technol (Environ Prot)* 13(4):74–82
- Dudášová H, Lukáčová L, Murínová L, Puškárová A, Pangallo D, Dercová K (2014) Bacterial strains isolated from PCB-contaminated sediments and their use for bioaugmentation strategy in microcosms. *J Basic Microbiol* 54:253–260
- Eriksson M, Dalhammar G, Mohn WW (2002) Bacterial growth and biofilm production on pyrene. *FEMS Microbiol Ecol* 40:21–27
- Etkin DS (1998, October) Oil spills from production and exploration activities. Oil spill intelligence report, white paper series Vol. II, no. 8, Publication of Cutter Information Corp
- Fall RR, Brown JL, Schaeffer TL (1979) Enzyme recruitment allows the biodegradation of recalcitrant-branched hydrocarbons by *Pseudomonas citronellolis*. *Appl Environ Microbiol* 38:715–722
- Filonov AE, Puntus IF, Karpov AV, Kosheleva IA, Kashparov KI, Slepkin AV, Boronin AM (2004) Efficiency of naphthalene biodegradation by *Pseudomonas putida* G7 in soil. *J Chem Technol Biotechnol* 79:562–569
- Fondi M, Rizzi E, Emiliani G, Orlandini V, Berna L, Papaleo MC, Perrin E, Maida I, Corti G, Bellis GD, Baldi F, Dijkshoorn L, Vaneeschoutte M, Fani R (2013) The genome sequence of the hydrocarbon-degrading *Acinetobacter venetianus* VE-C3. *Res Microbiol* 164(5):439–449
- Fritsche W, Hofrichter M (2000) In: Klein J (ed) *Aerobic degradation by microorganisms in environmental processes- soil decontamination*. Wiley-VCH, Weinheim
- Ghurye GL, Vipulanandan C (1994) A practical approach to biosurfactant production using non-aerobic fermentation of mixed cultures. *Biotechnol Bioeng* 44:661–666
- Golyshin PN, Santos VAPMD, Kaiser O, Ferrer M, Sabirova YS, Lünsdorf H, Chernikova TN, Golyshina OV, Yakimov MM, Pühler A, Timmis KN (2003) Genome sequence completed of *Alcanivorax borkumensis*, a hydrocarbon-degrading bacterium that plays a global role in oil removal from marine systems. *J Biotechnol* 106:215–220
- Gregorio SD, Siracusa G, Becarelli S, Mariotti L, Gentini A, Lorenzi R (2016) Isolation and characterization of a hydrocarbonoclastic bacterial enrichment from total petroleum hydrocarbon contaminated sediments: potential candidates for bioaugmentation in bio-based processes. *Environ Sci Pollut Res* 23:10587. <https://doi.org/10.1007/s11356-015-5944-y>
- Guermouche MA, Bensalah F, Gury J, Duran R (2015) Isolation and characterization of different bacterial strains for bioremediation of n-alkanes and polycyclic aromatic hydrocarbons. *Environ Sci Pollut Res* 22(20):15332–16346
- Hall AJ, Hugunin K, Deaville R, Law RJ, Allchin CR, Jepson PD (2006) The risk of infection from polychlorinated biphenyl exposure in the Harbor Porpoise (*Phocoena phocoena*): a case-control approach. *Environ Health Perspect* 114:704–711

- Hamann C, Hegemann J, Hildebrandt A (1999) Detection of polycyclic aromatic hydrocarbon degradation genes in different soil bacteria by polymerase chain reaction and DNA hybridization. *FEMS Microbiol Lett* 173(1):255–263
- Hemmer MJ, Barron MG, Greene RM (2011) Comparative toxicity of eight oil dispersants, Louisiana sweet crude oil (LSC), and chemically dispersed LSC to two aquatic test species. *Environ Toxicol Chem* 30(10):2244–2252
- Hommel RK (1990) Formation and physiological role of biosurfactants produced by hydrocarbon utilizing microorganisms. *Biodegradation* 1:107–119. <https://www.restorethegulf.gov/>. Accessed 8 Aug 2016
- Hyne NJ (2001) Nontechnical guide to petroleum geology, exploration, drilling and production, 2nd edn. PennWell Books, USA. ISBN: 978-0878148233
- Ibrahim ML, Ijah UJJ, Manga SB, Bilbis LS, Umar S (2013) Production and partial characterization of biosurfactant produced by crude oil degrading bacteria. *Int Biodeterioration Biodegrad* 81:28–34
- ITOPF (2016) <http://www.itopf.com/knowledge-resources/data-statistics/statistics/>. Accessed 17 Feb 2016
- Ivanova AA, Vetrova AA, Filonov AE, Boronin AM (2015) Oil biodegradation by microbial–plant associations. *Appl Biochem Microbiol* 51(2):191–197
- Janani PG, Keerthi K, Deshpande A, Bhattacharya S, Indira RP (2014) Molecular identification of the isolated diesel degrading bacteria and optimization studies. *J Biochem Technol* 5(3):727–730
- Jia C, Li X, Allinson G, Liu C, Gong Z (2016) Composition and morphology characterization of exopolymeric substances produced by the PAH-degrading fungus of *Mucor mucedo*. *Environ Sci Pollut Res* 23(9):8421–8440
- John RC, Okpokwasili GC (2012) Crude oil degradation and plasmid profile of nitrifying bacteria isolated from oil impacted mangrove sediment in the Niger Delta of Nigeria. *Bull Environ Contam Toxicol* 88:1020–1026
- Johnson K, Anderson S, Jacobson CS (1996) Phenotypic and genotypic characterization of phenanthrene-degrading fluorescent *Pseudomonas biovars*. *Appl Environ Microbiol* 62:3818–3825
- Juwarkar AA (2012) Microbe-assisted phytoremediation for restoration of biodiversity of degraded lands: a sustainable solution. *Proc Natl Acad Sci India* 82:313–318
- Kaladumo COK (1996) The implications of gas flaring in the Niger Delta environment. Proceedings of the 8th biennial international NNPC seminar. In: *The Petroleum Industry and the Nigerian Environment*, Port Harcourt, Nigeria, pp 277–290
- Karanth NGK, Deo PG, Veenanadig NK (1999) Microbial production of biosurfactant and their importance. *Ferment Sci Technol* 77:116–126
- Karpagam S, Lalithakumari D (1999) Plasmid mediated degradation of *o*- and *p*-phthalate by *Pseudomonas fluorescens*. *World J Microbiol Biotechnol* 15:565–569
- Kim H, Jaffé PR (2008) Degradation of toluene by a mixed population of archetypal aerobes, microaerophiles, and denitrifiers: laboratory sand column experiment and multispecies biofilm model formulation. *Biotechnol Bioeng* 99(2):290–301
- Kim YH, Engesser KH, Cerniglia CE (2005) Numerical and genetic analysis of polycyclic aromatic hydrocarbon-degrading mycobacteria. *Microbiol Ecol* 50:110–119
- Kim YH, Engesser KH, Kim SJ (2007) Physiological, numerical and molecular characterization of alkyl ether-utilizing rhodococci. *Environ Microbiol* 9(6):1497–1510
- Krebs CT, Tanner CE (1981) Restoration of oiled marshes through sediment stripping and *Spartina* propagation. Proceeding of the 1981 oil spill conference, American Petroleum Institute, Washington, DC, pp 375–385
- Kumar BL, Gopal DVRS (2015) Effective role of indigenous microorganisms for sustainable environment. *3 Biotech* 5(6):867–876
- Lang S (2002) *Biological amphiphiles* (microbial biosurfactants). *Curr Opin Colloid Interface Sci* 7:12–20

- Li YN, Porter AW, Mumford A, Zhao XH, LY Y (2011) Bacterial community structure and bama gene diversity in anaerobic degradation of toluene and benzoate under denitrifying conditions. *J Appl Microbiol* 112:269–279
- Li J, Toledo RA, Chung J, Shim H (2013) Removal of mixture of cis-1, 2 dichloroethylene/trichloroethylene/benzene, toluene, ethylbenzene, and xylenes from contaminated soil by *Pseudomonas plecoglossicida*. *J Chem Technol Biotechnol* 89(12):1934–1940
- Li F, Guo S, Hartog N, Yuan Y, Yang X (2016) Isolation and characterization of heavy polycyclic aromatic hydrocarbon-degrading bacteria adapted to electrokinetic conditions. *Biodegradation* 27:1–13
- Liu Y, Zhang J, Zhang Z (2004) Isolation and characterization of polycyclic aromatic hydrocarbons-degrading *Sphingomonas sp.* strain ZL5. *Biodegradation* 15:205–212
- Luo Q, Zhang JG, Shen XR, Fan ZQ, He Y, Hou DY (2012) Isolation and characterization of marine diesel oil-degrading *Acinetobacter sp.* strain Y2. *Ann Microbiol* 63:633–640
- Ma J, Yan G, Ma W, Cheng C, Wang Q, Guo S (2015) Isolation and characterization of oil-degrading microorganisms for bench-scale evaluations of autochthonous bioaugmentation for oil remediation. *Water Air Soil Pollut* 226:272–280
- Mabro RE (2006) *Oil in the twenty-first century: issues, challenges and opportunities*. Oxford University Press, Oxford ISBN-13: 9780199207381
- Makkar RS, Cameotra SS (1997) Utilization of molasses for biosurfactant production by two *Bacillus strains* at thermophilic conditions. *J Am Oil Chem Soc* 74:887–889
- Mandri T, Lin J (2007) Isolation and characterization of engine oil degrading indigenous microorganisms in Kwazulu-Natal, South Africa. *Afr J Biotechnol* 6:23–27
- Martino CD, Lopez NI, Iustman LJR (2012) Isolation and characterization of benzene, toluene, and xylene degrading *Pseudomonas sp.*, selected as candidate for bioremediation. *Int Biodeterioration Biodegrad* 67:15–20
- Miller CD, Hall K, Liang YN, Nieman K, Sorensen D, Issa B, Anderson AJ, Sims RC (2004) Isolation and characterization of polycyclic aromatic hydrocarbon-degrading mycobacterium isolates from soil. *Microbial Ecol* 48:230–238
- Mishra S, Jyot J, Kuhad RC, Lal B (2001) Evaluation of inoculum addition to stimulate in situ bioremediation of oily-sludge-contaminated soil. *Appl Environ Microbiol* 67:1675–1681
- National Academy of Science (NAS) (2005) *Oil spill dispersants: efficacy and effects*; Ocean Studies Board. <http://dels.nas.edu/Report/Spill-Dispersants-Efficacy-Effects/11283>
- National Academy of Sciences (1985) *Oil in the sea: inputs, fates and effects*. National Academy Press, Washington DC
- Ndlovu T, Khan S, Khan W (2016) Distribution and diversity of biosurfactant producing bacteria in a wastewater treatment plant. *Environ Sci Pollut Res* 23(10):9993–10004
- Nie M, Yin X, Ren C, Wang Y, Xu F, Shen Q (2010) Novel rhamnolipid biosurfactants produced by a polycyclic aromatic hydrocarbon-degrading bacterium *Pseudomonas aeruginosa* strain NY3. *Biotechnol Adv* 28(5):635–643
- Niepceron M, Koltalo FP, Merlin C, Massei AM, Barray S, Bodilis J (2010) Both *Cycloclasticus spp.* and *Pseudomonas spp.* as PAH-degrading bacteria in the Seine estuary (France). *FEMS Microbiol Ecol* 71:137–147
- Office of Technology Assessment (1990) *Coping with an oiled sea: an analysis of oil spill response technologies*, OTA-BP-O-63, Washington, DC
- Office of Technology Assessment (1991) *Bioremediation of marine oil spills: an analysis of oil spill response technologies*, OTA-BP-O-70, Washington, DC
- Okpokwasili GC, Odokuma LO (1986) Tolerance of Nitrobacter to toxicity of some Nigerian crude oils. *Bull Environ Contam Toxicol* 52:388–395
- Okpokwasili GC, Okorie BB (1988) Biodeterioration potentials of microorganisms isolated from car-engine lubricating oil. *Tribol Znt* 21:215–220
- Onur G, Yilmaz F, and Içgen B (2015) Diesel Oil Degradation Potential of a Bacterium Inhabiting Petroleum Hydrocarbon Contaminated Surface Waters and Characterization of Its Emulsification Ability. *J Surfactant Deterg* 18:707–717.

- Onwurah INE (1999) Restoring the crop sustaining potential of crude oil polluted soil by means of *Azotobacter* inoculation. *Plant Prod Res J* 4:6–16
- Onwurah INE (2002) Anticoagulant potency of water-soluble fractions of Bonny light oil and enzyme induction in rats. *Biomed Res* 13(1):33–37
- Onwurah INE, Ogueva VN, Onyike NB, Ochonogor AE, Otitoju OF (2007) Crude oil spills in the environment, effects and some innovative clean-up biotechnologies. *Int J Environ Res* 1(4):307–320
- Oyetibo GO, Ilori MO, Obayori OS, Amund OO (2013) Biodegradation of petroleum hydrocarbons in the presence of nickel and cobalt. *J Basic Microbiol* 53:917–927
- Pacwa-Plociniczak M, Płaza GA, Poliwoła A, Piotrowska-Seget Z (2014) Characterization of hydrocarbon-degrading and biosurfactant-producing *Pseudomonas* sp. P-1 strain as a potential tool for bioremediation of petroleum-contaminated soil. *Environ Sci Pollut Res* 21:9385–9395
- Pedetta A, Pouyte K, Seitz MKH, Babay PA, Espinosa M, Costagliola M, Studdert CA, Peressutti SR (2013) Phenanthrene degradation and strategies to improve its bioavailability to microorganisms isolated from brackish sediments. *Int Biodeterior Biodegrad* 84:161–167
- Peressutti SR, Alvarez HM, Pucci OH (2003) Dynamics of hydrocarbon-degrading bacteriocenosis of an experimental oil pollution in Patagonian soil. *Int Biodeterior Biodegrad* 52:21–30
- Perfumo A, Banat IM, Canganella F, Marchant R (2006) Rhamnolipid production by a novel thermophilic hydrocarbon-degrading *Pseudomonas aeruginosa* AP02-1. *Appl Microbiol Biotechnol* 72:132–138
- Perry JJ (1980) Oil in the biosphere. In: Guthrie FE, Perry JJ (eds) *Introduction to environmental toxicology*. Elsevier, New York
- Porob S, Nayak S, Fernandes A, Padmanabhan P, Patil BA, Meena RM, Ramaiah N (2013) PCR screening for the surfactin (*sfp*) gene in marine *Bacillus* strains and its molecular characterization from *Bacillus tequilensis* NIOS11. *Turkish J Biol* 37:212–221
- Prince RC (1993) Petroleum spill bioremediation in marine environments. *Crit Rev Microbiol* 19:217–242
- Quatrini P, Scaglione G, De Pasquale C, RIELA S, Puglia AM (2008) Isolation of gram-positive n-alkane degraders from a hydrocarbon-contaminated Mediterranean shoreline. *J Appl Microbiol* 104:251–259
- Reddy PG, Singh HD (1982) Bacterial degradation of emulsified crude oil and the effect of various surfactants. *J Microbiol* 43:17–22
- Roling WFM, Milner MG, Jones DM, Lee K, Daniel F, Swannell RJP (2002) Robust hydrocarbon degradation and dynamics of bacterial communities during nutrient-enhanced oil spill bioremediation. *Appl Environ Microbiol* 68(11):5537–5548
- Roongsawang N, Hase K, Haruki M, Imanaka T, Morikawa M, Kanaya S (2003) Cloning and characterization of the gene cluster encoding arthrofactin synthetase from *Pseudomonas* sp. MIS38. *Chem Biol* 10:869–880
- Rusansky S, Avigad R, Michaeli S, Gutnick DL (1987) Involvement of a plasmid in growth on and dispersion of crude oil by *Acinetobacter calcoaceticus* RA57. *Appl Environ Microbiol* 53:1918–1923
- Satpute SK, Banat IM, Dhakephalkar PK, Banpurkar AG, Chopade BA (2010) Biosurfactants, bioemulsifiers and exopolysaccharides from marine microorganisms. *Biotechnol Adv* 28:436–450
- Sei A, Fathepure BZ (2009) Biodegradation of BTEX at high salinity by an enrichment culture from hypersaline sediments of Rozel Point at Great Salt Lake. *J Appl Microbiol* 107:2001–2008
- Sharidah AA, Richardt A, Goleckil JR, Diersteinl R, Tadros MH (2000) Isolation and characterization of two hydrocarbon-degrading *Bacillus subtilis* strains from oil contaminated soil of Kuwait. *Microbiol Res* 155:157–164
- Shin KH, Kim KW, Ahn Y (2006) Use of biosurfactant to remediate phenanthrene-contaminated soil by the combined solubilisation biodegradation process. *J Hazard Mater* 137(3):1831–1837
- Short JW, Heintz RA (1997) Identification of Exxon Valdez oil in sediments and tissue from Prince William sound and the North Western Gulf of William based in a PAH weathering model. *Environ Sci Technol* 31:2375–2384

- Simpson DR, Natraj NR, McInerney MJ, Duncan KE (2011) Biosurfactant-producing *Bacillus* are present in produced brines from Oklahoma oil reservoirs with a wide range of salinities. *Appl Microbiol Biotechnol* 91:1083–1093
- Singh D, Fulekar MH (2010) Biodegradation of petroleum hydrocarbons by *Pseudomonas putida* strain MHF 7109. *Clean Soil Air Water* 38(8):781–786
- Singh G, Malik DK (2013) Utilization of 2T engine oil by *Pseudomonas* sp. isolated from automobile workshop contaminated soil. *Int J Chem Anal Sci* 4(2):80–84
- Sorensen SR, Johnsen AR, Jensen A, Jacobsen CS (2010) Presence of psychrotolerant phenanthrene-mineralizing bacterial populations in contaminated soils from the Greenland High Arctic. *FEMS Microbiol Lett* 305:148–154
- Speight JG (1999) *The chemistry and technology of petroleum*, Marcel Dekker, ISBN 0-8247-0217-4
- Spies RB, Rice SD, Wolfe DA, Wright BA (1996) The effect of the Exxon Valdez oil spill on Alaskan coastal environment. Proceedings of the 1993 Exxon Valdez oil spill symposium, American Fisheries Society, Bethesda, MD
- Sun W, Sun X, Cupples AM (2014) Identification of Desulfosporosinus as toluene-assimilating microorganisms from a methanogenic consortium. *International Biodeterioration and Biodegradation*. 88:13–19
- Su WT, Wu BS, Chen WJ (2011) Characterization and biodegradation of motor oil by indigenous *Pseudomonas aeruginosa* and optimizing medium constituents. *J Taiwan Inst Chem Eng* 42:689–695
- Tapilatu YH, Grossi V, Acquaviva M, Milton C, Bertrand J, Cuny P (2010) Isolation of hydrocarbon-degrading extremely halophilic archaea from an uncontaminated hypersaline pond (Camargue, France). *Extremophiles* 14:225–231
- Tiido T, Rignell-Hydbom A, Jönsson BAG, Giwercman YL, Pederson HS, Wojtyniak B, Ludwicki JK, Lesovoy V, Zvyezday V, Spano M, Manicardi GC, Bizzaro D, Bonefeld-Jørgensen EC, Toft G, Bonde JP, Rylander L, Hagmar L, Giwercman A (2006) Impact of PCB and p,p'-DDE contaminants on human sperm Y:X chromosome ratio: studies on three European populations and the Inuit population in Greenland. *Environ Health Perspect* 114:718–724
- U.S. EPA (1999) A series of fact sheets on Nonpoint Source (NPS) pollution. EPA841-F-96-004, Office of Water, U.S. Environmental Protection Agency
- U.S. EPA (2000) The quality of our nation's waters: a summary of the national water quality USEPA (2015) National contingency plan product schedule (March 2015). http://epa.gov/ncert/rfa/2015/2015_star_gradfellow.html. Accessed 17 March 2015
- Van Hamme JD, Singh A, Ward OP (2003) Recent advances in petroleum microbiology. *Microbiol Mol Biol Rev* 67(4):503–549
- Vasudevan N, Bharathi S, Arulazhagan P (2007) Role of plasmid in the degradation of petroleum hydrocarbon by *Pseudomonas fluorescens* NS1. *J Environ Sci Health Part A: Tox* 42(8):1141–1146
- Vecchioli GI, Panno MTD, Paineira MT (1990) Use of selected autochthonous soil bacteria to enhance degradation of hydrocarbons in soil. *Environ Pollut* 67:249–258
- Vieira PA, Faria S, Vieira RB, De Franc FP, Cardoso VL (2009) Statistical analysis and optimization of nitrogen, phosphorus, and inoculum concentrations for the biodegradation of petroleum hydrocarbons by response surface methodology. *World J M Biotechnol* 25:427–438
- Wang W, Shao Z (2012) Diversity of flavin-binding monooxygenase genes (*almA*) in marine bacteria capable of degradation long-chain alkanes. *FEMS Microbiol Ecol* 80:523–533
- Weelink SAB, Doesburg WV, Talarico FS, Rijkstra WIC, Smidt H, Stams AJM (2009) A strictly anaerobic betaproteobacterium *Georgfuchsia toluolica* gen. nov., sp. nov. degrades aromatic compounds with Fe (III), Mn (IV) or nitrate as an electron acceptor. *FEMS Microbiol Ecol* 70:575–585
- Xia WX, Li JC, Zheng XL, Bi XJ, Shao JL (2006) Enhanced biodegradation of diesel oil in seawater supplemented with nutrients. *Eng Life Sci* 6(1):80–85

- Yakimov MM, Timmis KN, Wray V, Fredrickson HL (1995) Characterization of a new lipopeptide surfactant produced by thermotolerant and halotolerant subsurface *Bacillus licheniformis* BAS50. *Appl Environ Microbiol* 61:1706–1713
- Yenn R, Borah M, Boruah HP, Roy AS, Baruah R, Saikia N, Sahu OP, Tamuli AK (2014) Phytobioremediation of abandoned crude oil contaminated drill sites of Assam with the aid of a hydrocarbon-degrading bacterial formulation. *Int J Phytoremediation* 16(7–12):909–925
- Zeinali M, Vossoughi M, Ardestani SK, Babanezhad E, Masoumian M (2007) Hydrocarbon degradation by thermophilic *Nocardia oititidiscaviarum* strain TSH1: physiological aspects. *J Basic Microbiol* 47:534–539
- Zhang Z, Hou Z, Yang C, Ma C, Tao F, Xu P (2011) Degradation of *n*-alkanes and polycyclic aromatic hydrocarbons in petroleum by a newly isolated *Pseudomonas aeruginosa* DQ8. *Bioresour Technol* 102(5):4111–4116