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New Combination/Application of Polymer-Based Nanoparticles for Biomedical Engineering

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Abstract

Polymer-based nanoparticles (PNPs) are attractive in part due to their ultra-small size, versatility and target specificity. Therefore, PNPs have been increasingly used in a variety of biomedical applications including diagnoses and therapeutic treatment. In this chapter, we focus on the recent studies (within 5 years) with some new ideas/agent's application in biomedical field and roughly divide applications of PNPs into four categories: (1) Delivery, (2) In vivo imaging, (3) Therapies, and (4) Other applications. First, we introduce how PNPs can enhance the treatment and delivery efficiency of therapeutic agent. Second, how PNPs can be used to help in vivo imaging system for disease tracking and monitor. Then, we reveal some novel PNPs which is able to function as an agent in photodynamic, photothermal, sonodynamic and neuron capture therapy. Furthermore, we also mention some

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Department of Chemistry and Biotechnology, Swinburne University of Technology, Victoria, 3122, Australia e-mail: pywang@tmu.edu.tw; pengyuanwang@swin.edu.au interesting applications of PNPs for biomedical field in individual form or cluster employment, such as immunoswitch particles, surface fabrication. Finally, the challenges and future development of PNPs are also discussed. In delivery section, we focus on how polymer "can be used" as vehicles in delivery application. But, in the section of imaging and therapies, we carried on how polymer as an "adjuvant" for functional enhancement. The biodegradable property of PNPs is the feature that they can be controllable for itself degradation and drug release as a chief actor. Besides, in imaging and therapies application, PNPs can be the support role for helping contrast agent or photo/sonosensitizer to enlarge their imaging or therapeutic effect.

Keywords

 $\begin{array}{l} Polymer-based \ nanoparticles \ (PNPs) \cdot \\ Delivery \cdot Imaging \cdot The rapeutics \cdot PNPs \\ cluster \end{array}$

14.1 Introduction

During the past decades, the development of nanotechnology makes the generation process of polymer-based nanoparticles (PNPs) more efficient. In recent years, PNPs have been increasingly used in a variety of biomedical applications including diagnoses and therapeutic treatment.

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Nanoparticles were first developed by Speiser and co-workers [1] around 1970 and are frequently defined as particles with diameters below the micron dimension (i.e., in the range 10–1000 nm) [2]. The concept of nanoscale devices has led to the development of nanoparticles. Two types of systems with different inner structures: (1) a reservoir-type system, consisting of an oily core surrounded by a polymer wall, defined here as a nanocapsule; (2) a matrix-type system composed of an entanglement of polymer units, defined here as a nanoparticle or nanosphere. The term nanoparticle can be used to refer to both systems including nanocapsules as well as nanoparticles.

Due to their ultra-small size, the PNPs have much larger surface area to volume ratio compared with bigger particles, enabling them to have unique biological behaviors. Their large functional surface area allow PNPs to interact with substances such as nucleic acids, proteins, lipids, ions, carbohydrates, probes or small molecule drugs and deliver these substances into the desired biophase via endocytosis. Furthermore, PNPs can be fabricated by different method. Their properties, such as size, shape, stability and particle composition are tunable, allowing them can be prepared to meet the requirements specific to biomedical application. For example, several pathways were involved in the endocytosis procedure of PNPs. This cellular uptake procedure can be controlled effectively by the properties of nanoparticles, such as nanoparticle size, shape, and surface chemistry [3–5]. After entering the target cell, PNPs can be made to respond to a different local stimulus, which will trigger PNPs to execute therapeutic functions, such as hyperthermia, imaging and drug release. Theses stimulus not only include biological endogenous signals like pH, glucose, enzyme, etc., but include external stimulus like alternating magnetic field (AMF), near infrared (NIR), temperature, and etc. [6, 7]. Due to their versatility and target specificity, PNPs have received much of the attention in various aspects of medicine. There are several excellent reviews on PNPs have been published, like their fabrication, characterization and strategies in the application [8–10].

In this article, we focus on the recent literature (past 5 years) and roughly divide the biomedical application of PNPs into four categories: (1) Delivery, (2) In vivo imaging, (3) Therapies, (4) Other applications, and summarized in Table 14.1.

Table 14.1 Summary of current PNPs in biomedical application

Application	Indication	References			
Delivery					
Dendrimers	Dendrimer with	Li et al. [11]			
	instantaneous size				
	switching ability for				
	tumor penetration				
Lipid	PNPs as a delivery	Yin et al. [12]			
Materials	platform for CRISPR/				
	Cas9 system				
Polymeric	Combination of PNPs	Smith et al.			
materials	with CAR T-cell therapy	[13]			
	Preventing obesity by	Xue et al. [14]			
	using PNPs				
In vivo imag	In vivo imaging				
Magnetic	pH-activatable PNPs for	Mi et al. [15]			
-	non-invasive MRI of				
	tumor				
Nuclear	Quantification of the	Keliher et al.			
	inflammation in	[16]			
	atherosclerotic plaque				
	using PET signal				
Optical	Finding a relationship	Hinde et al.			
-	between nanoparticle	[17]			
	shapes and the ability of				
	nanoparticle access into				
	the nucleus				
	Examining the radiation	Miller et al.			
	effect for tumoral	[18]			
	therapy				
	SERS for high-precision	Harmsen et al.			
	cancer imaging	[19]			
	Detection of endogenous	Zhao et al.			
	H ₂ S within living cells	[20]			
	via FRET				
	Semiconducting polymer	Pu et al. [21]			
	based PNPs as probes for				
	photoacoustic imaging				
Therapies					
PDT	Performing PDT for	Punjabi et al.			
	deep tissue penetration	[22]			
PTT	Combination of PTT and	Chen et al.			
	checkpoint-blockade	[23]			
	immunotherapy to				
	eliminate primary				
	tumors				
SDT	Combination of SDT	Huang et al.			
	and MRI for SDT	[24]			
	monitoring	_			
NCT	Incorporating Gd	Mi et al. [25]			
-	for MRI-guided				
	gadolinium NCT				
		(continued)			

Application	Indication References			
Other applications				
Individual	Separation the protein Nehilla			
application	biomarkers	et al. [101]		
	Simulates the immune	Kosmides		
	system to inhibit tumor	et al. [102]		
	growth			
	PNPs for neutralization	O'Brien		
	of venomous	et al. [104]		
	biomacromolecules			
	PNPs as a protein	Koide et al.		
	affinity reagent for	[105]		
	inhibition of			
	angiogenesis			
	Predict the therapeutic	Yaari et al.		
	efficiency of anticancer	[106]		
	drug in a personalized			
	manner			
	Identification the ideal	Dahlman		
	targeted therapeutics by	et al. [107]		
	DNA barcoded PNPs			
Cluster	Self-assembled	Wang et al.		
employment	nanoparticle monolayers	[113, 114]		
	as a cell culture tool			

Table 14.1 (continued)	Table 14.	(continued)
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Abbreviations: MRI, magnetic resonance imaging; PET, positron emission tomography; SERS, surface-enhanced Raman spectroscopy; FRET, fluorescence resonance energy transfer; PDT, photodynamic therapy; PTT, photothermal therapy; SDT, sonodynamic therapy; NCT, neutron capture therapy; Gd, gadolinium

14.2 Delivery

The ultra-small size of PNPs lead it enter cells more easily via endocytosis. Therefore, the application of PNPs in drug delivery, especially for cancer treatment, is particularly welcome. In order to grow quickly, tumor tissues tend to possess higher vascular permeability rather than normal tissues. Rapid vascularization to serve fast-growing cancerous tissue leads vessels to a leaky and defective architecture. The enhanced permeability and retention phenomenon (EPR) is based on two factors: (a) the capillary endothelium in malignant tissue is more disorderly and thus more permeable towards macromolecules than the capillary endothelium in normal tissues. This allows extravasation of circulating PNPs within the tumor interstitium, and (b) the lack of tumor lymphatic drainage in the tumor

bed results in particles' accumulation. Thus, the accumulation of PNPs in tumor tissue is much higher than normal tissues. PEGylation, coating the surface of nanocarriers with poly(ethylene glycol) (PEG), is a commonly used strategy to prolong circulation time of PNPs [26]. The EPR effect provide the PEGylation PNPs with increased opportunity to access tumors site. Due to the EPR effect, PNPs have been widely used as a vehicle to deliver the therapeutic agent, such as drug, nucleotides and protein, into tumor tissue.

Many recent studies, however, have described the limitations of EPR effect. The EPR effect is not suitable for metastatic liver cancers and less vascularized cancers, such as prostate cancer [27]. On the other hand, it has been reported that a phenomenon known as "accelerated blood clearance (ABC)" which repeated injections of PEGylated nanocarriers may cause of an unexpected immunogenic response, leading to lose their long-circulating characteristic [28]. Therefore, it is necessary to develop actively-targeted PNPs to specific recognize the specific cells.

To perform active targeting, specific ligands able to interact with the specific receptors localized on cell membranes can be coupled to the surface of PNPs. Several studies have summarized the various kind of ligand, such as small molecules, carbohydrates, peptides, enzymes or antibodies, have been used for active targeting [29–31]. In addition to ligation strategies, recently, a novel method called "cell membrane coating" has been developed for designing of PNPs. Cell membrane coating method is a technique, which the whole membrane will translocate from a cell to the surface of a nanoparticle, make the nanoparticle potentially perform some cell-specific functionalities, such as, immune evasion and targeting abilities. Zhang et al. [32, 33] have used this technique to address the issue of ABC phenomenon and improve PNPs functionality to achieve longer circulation time, higher tumor specificity and lower exocytosis of drug (Fig. 14.1) [34, 35]. The PNPs can be roughly classified nto three types; dendrimer, lipid based PNPs and biopolymeric based PNPs.



Fig. 14.1 Schematic preparation of PNPs. poly(lacticco-glycolic acid) (PLGA) nanoparticles are enclosed entirely in plasma membrane derived from human platelets. The resulting particles possess platelet mimicking

properties for immunocompatibility, subendothelium binding, and pathogen adhesion. (Image adapted from Zhang et al. [33] and reprinted with permission from Springer Nature)

14.2.1 Dendrimers

Dendrimers, a large number of branching points, including precisely-defined molecular structure, numerous functional groups on the surface and cavities in the interior, have been widely used in the field of drug delivery in a three-dimensional spheral shape, nanometric size and well monodispersity [36]. With appropriate branching units and surface groups modification, dendrimers can provide the targeted delivery of gene or drugs. Kesharwani et al. [37, 38] reviewed the recent advancements in dendrimer-based PNPs for tumor-targeted delivery. They summerized the different kinds of ligands, including the biotin, folate, amino acids, peptides, aptamers and antibodies, which have been successfully conjugated to dendrimer [37]. Also, they have reviewd the different routes, such as parenteral, transdermal, oral, plumonary, ocular and colon, can delivery the various dendrimers [38]. In the following, several publications within these years will be discussed.

Dendrimers can be used for gene delivery. For example, Cheng et al. [39] systematically summarized the functional ligands can modifided to the dendrimers to improve the DNA- and membranebinding affinity, transfection efficacy and biocompatibility of dendrimers. Also, dendrimers can be applied in drug delivery, including doxorubicin, 5-Fluorouracil, paclitaxel and other types of chemotherapy drugs. [40]. For example, Li et al. [11] preapred the size switching PNPs construcring from assembly of platinum-prodrug conjugated polyamidoamine dendrimers. At neutral pH, this PNPs have initial size of ~80 nm. Under acidic tumor microenvironment, however, the PNPs rapidly dissociate into the dendrimer building blocks and penerate into tumor cell. Wei et al. [41] synthesize the amphiphilic dendrimer, which can form the supramolecular micelles to enclose the anticancer drug doxorubicin with high loading capacity. Not only for gene transfection and chemodrug delivery, dendrimeric nanoparticles also have advantages for ocular drug delery: nanoparticle platform of hybrid dendrimer hydrogel/poly (lactic-co-glycolic acid) with anti-glaucoma drug loading was evaluated. The hybrid dendrimer nanoparticle platform (HDNP) consists of three domains: the polyamidoamine (PAMAM) dendrimer core to encapsulate hydrophobic drugs, the poly (lactic-co-glycolic acid) (PLGA) nanoparticles to deliver either hydrophobic or hydrophilic drugs, and the PEG network to load hydrophilic drugs. The major advance of this novel dendrimer is its ability to deliver simultaneously multiple drugs in the same dosages and release them in a slow manner with sustained efficacy [42]. These platforms are very promising tools in ocular nanomedicine.

14.2.2 Lipid Based PNPs

Lipid based PNPs are delivery system mainly prepared from natural and/or synthetic phosphoand sphingo-lipids. Because they are absorbed more easily in human body and producing fewer toxic degradation products, therefore, lipid materials based PNPs are more suitable for clinical trial rather than other delivery system. In addition, some of them, particularly liposomes, due to their bilayer structure, can enclose and tranport the both hydrophobic and hydrophilic drugs. Several researchers have comprehensive review the current state of lipid based PNPs, including development process of them, strategies of them for tumor targeting, and commerical applications of them [43–45]. Zhang et al. [46] reported that upconversion nanoparticles encapsulated Azobenzene (Azo) liposome (UCNP@Azo-Lipo) could convert near infrared (NIR) light into the UV/vis region; then UV/vis light was absorbed by the Azo molecules in the liposome. When stimulated by UV/vis light, the synthesized Azo amphiphilic derivatives created continuous rotation-inversion movement for the liposome membrane then driving drug released. The results show that they can precisely control the drug release amount via tuning the intensity and duration of light irradiation. The combination of NIR liger trigger system and liposome leads an on-off switch controllable liposome was created. Recently, lipid based PNPs and clustered regularly interspaced short palindromic repeat (CRISPR)/CRISPR-associated protein 9 (Cas9) systems have been combined in a new method to increase genome editing efficiencies. Yin et al. [12] use the lipid based PNPs to deliver the sgRNAs (e-sgRNA) and mRNA encoding Cas9, which can significantly decrease the specific genes such as Pcsk9 in liver (>80% editing), and serum to undetectable levels by a single intravenous injection in mice. These lipid-Cas9 based PNPs provide a non-viral genome editing vehicles for liver genome correcting in clinical setting. The lipid based PNPs, called the liposome protamine/DNA lipoplex (LPD), was electrostatically assembled from cationic liposomes and an anionic protamine-DNA complex, it promoted efficient delivery of the retinal pigment epithelium protein 65 (Rpe65) gene in mice in a longterm expression and cell specific-mode leading to in vivo correction of blindness [47]. The efficacy of this method of restoring vision is comparable to AAV and lentiviral gene transfer of the Rpe65 gene to Rpe65 knockout mice [48].

14.2.3 Biopolymeric Based PNPs

Biopolymeric based PNPs, compare to other PNPs, have better storage stability and diversity of designs [49]. They can roughly divide into nanocapsules and nanospheres, based on their structure. Both of them have spherical structure comprising polymer, however, the cargo is dispersed within a matrix for nanoshpere and encapsulated in central cavity for nanocapsule, respectively [50]. Nanogels are common nanosphere for delivery. Li et al. [8] summarize the method to fabricate the nanogels, based on natural and synthetic polymers, for drug and nucleic acid controlled release. Gaitzsch et al. [51] systematically review papers about how to prepare the "smart" nanocapsules for delivery system based on different techniques like self-assembly, emulsion polymerization, microfluidics, or Pickering emulsion, etc. Therefore, due to their diversity of designs, various release strategies,

including heat, light, ultrasound stimulation; pH change, oxidation process, or enzyme participation, these ways have been employed to improve the efficiency of biopolymeric based PNPs to sustained drug release within the target site [52]. In the following, we introduce some newly publications within these years.

Satchi-Fainaro et al. [53] designed a biodegradable amphiphilic polyglutamate amine PNPs (APA) PNPs that can deliver the small interfering RNA (siRNA) and microRNA (miRNA) into the tumor. After arrive the tumor site, PNPs will be degraded by an enzyme highly expressed in tumor tissues leading to release the siRNA or miRNA to silence or dysregulate the gene associated with cancer, respectively. PNPs can be used to enhance the efficiency of chimeric antigen receptor T-cell immune therapy (CAR-T). Stephan et al. [13] prepare the PNPs, based on poly(beta-amino ester) (PBAE) polymer functionalized with the microtubule-associatednuclear localization (MTAS-NLS) peptide, can transport the leukaemia-targeting CAR genes into T-cell nuclei to rapidly program T cells with abilities of tumour-recognizing, resulting in simplifying the time consuming traditional method. Also, PNPs can be used to address obesity. Langer et al. [14] demonstrate the PNPs selfassembled from biodegradable triblock polymer composed of adipose vasculature targeted peptides conjugated poly(lactic-coglycolic acid)-bglycol) (PLGA-b-PEG) can poly(ethylene increase the accumulation of drugs in white adipose tissue (WAT), which accelerate the transformation of WAT into brown adipose tissue (BAT) and then cause the weight loss.

Although novel polymeric based NPs for drug/ gene delivery are designed/developed for biomedical application popularly; traditional biopolymer such as chitosan or gelatin also plays an important role in drug delivery system. For example, gelatin, the biodegradable polymer, exhibits excellent biocompatibility, plasticity, and adhesiveness [54]. Its degradation rate can be regulated by the degree of cross-linking. The functional groups on gelatin NPs, such as carboxyl, hydroxyl, and amino groups, are available for conjugation with ligands to bring about surface modifications. Variant gelatin nanoparticles (GPs) were synthesized by Tseng et al. [55–57] for different application such as inhalation delivery of cisplatin loaded GPs with EGF modification for lung cancer treatment [55]; or as an efficient and safe drug carriers for ocular drug delivery in an eye-drops formula [56]; and even be a gene delivery carriers with pEGFP-C1 loading (plasmid encoded enhanced green fluorescence protein) for transgene sic chicken manipulation [57].

Nonmatter novel one or old one, biopolymeric based PNPs is the major components as delivery system for drug/gene. Beyond this, another role such as a protector for contrast agent for imaging or other therapeutic chemical application, PNPs can also participate as a supporting role for reaching final clinical requirements. These are adders thereafter.

14.3 In vivo Imaging

Medical imaging techniques are powerful tool allowing researchers to look inside a cell or to find difference between the normal and abnormal biological processes. However, most imaging agents, such as organic molecules and inorganic nanomaterials, do not have ability to recognize and target specific cells hindering their applications to cellular studies and biological imaging. To deal with this problem, PNPs have been introduced into the biomedical imaging field as a helper.

Currently, most studies using PNPs for molecular imaging focus on designing of nanoparticles with a structure of combination of imaging agent core and polymer shell. This designing has some advantages. First, it can decrease the cytotoxicity of imaging agent. Second, various targeting ligands can be modified to the polymer shell allowing imaging agent can active bind to cellular target moieties. In addition, it will promote imaging agent accumulation in the tumor, increasing the signal-to-noise ratio, highlighting tumor tissue within the body. Third, compare to imaging agent alone, polymer shell can make it with larger size, allowing them to have longer circulation time, decreasing in the time to agent's administration. Zheng et al. [58] discuss the clearance pathways and tumor targeting of imaging nanoparticles, including ultra-small inorganic core modified with protein-adsorption-resistant zwitterionic or PEGylated surface. Lammers et al. [59] summarize the advantages and limitations of imaging modalities, such as such as magnetic resonance imaging (MRI), positron emission tomography (PET), single photon emission computed tomography (SPECT) and optical imaging. Also, they provide principles for the rational use of PNPs in noninvasive imaging. Smith et al. [60] divide the imaginable nanomaterials based on the fundamental physicochemical properties. In addition, carrier, such as micelle, liposome, dendrimer and polymeric materials, for delivery of these nanomaterials are discussed.

In this section, we discuss the recent applications of PNPs in bioimaging. Based on imaging modalities, we have roughly divided imaging into three parts: magnetic, nuclear, and optical. The research we discussed here will mainly focus on how polymers are utilized to coat on the inorganic nanoparticles for *in vivo* distribution/biocompatibility. However, several studies, which utilize as polymers, like conjugated polymer, as co-component of imaging agents, will also be examined.

14.3.1 Magnetic

Magnetic resonance imaging (MRI), rather than use of damaging radiation, is based on the tiny magnetic moments produced by the spin of certain atomic nuclei within the body. MRI is a valuable technique for the clinical diagnosis that producing the non-invasive three-dimensional detailed anatomical images of soft tissues. Because soft tissue is present all over the human body, therefore, in medical diagnosis, MRI has a wide range of applications, such as detection of peripheral neuropathy, systemic cancer and cardiovascular disease. Gadolinium (Gd) and iron oxide based nanoparticles is common MRI contrast agent [61]. Recently, MRI agent made by Gd hexanedione nanoparticle has been developed to label and track the stem cell with low toxicity but higher image enhancement capacity [62].

Zhang et al. [63] describes the way to improve the biocompatibility and reflexivity of Gd-based contrast agents by conjugation of natural or synthetic polymers to it. Bakhtiary et al. [64] summarize the work of using PNPs with super paramagnetic iron oxide nanoparticles (SPION)based contrast agents to early detect and image different major cancer types, including liver, prostate, brain, breast and cervical.

In addition to Gd and SPION, manganese (Mn)-based materials have been used for MRI contrast agent in recent years. Mi et al. [15] prepared pH-activatable PNPs for non-invasive imaging of tumor. They confined Mn²⁺ within calcium phosphate (CaP) core enveloping by a PEG shell, which can avoid aggregation of core. After arrive the tumor, the acidic tumor environment make pH-sensitive CaP core to release the Mn²⁺ ion, which bind to proteins increasing the MRI contrast (Fig. 14.2).

14.3.2 Nuclear

Nuclear imaging is a technique using radioactive agents to observe metabolic processes in the body, which can be used to diagnose abnormalities in the bodily functions, especially effective in identification of cancerous tumors. The most common used nuclear imaging techniques are positron emission tomography (PET) and single photon emission computed tomography (SPECT). Both of them can provide accurate three-dimensional images, but the main difference between their imaging is the type of radioactive agents used. The decay of the radioactive agents used with PET measure positron emission, whereas the SPECT detect gamma ray emission. Radioactive agents can be used for imaging have been discussed earlier [60, 65, 66].

However, some clinical radioactive agents, such as ¹⁸F-fluorodeoxyglucose, do not have the ability to conduct themselves to target site. Moreover, due to low molecular weight, their circulation time is short. Therefore, to overcome these problems, several studies have incorporated the radioactive agents into PNPs [67]. For example, Phillips et al. [68] prepared the silica particles labeling with ¹²⁴I for PET and coating the nanoparticle with cRGDY peptide and PEG to improve the accuracy of metastatic melanoma diagnostic. Chen et al. [69] encapsulate vana-



Fig. 14.2 (a) Illustration of the PEGMnCaP structure. (b) TEM image of PEGMnCaP. (c and d) 3D MRI of C26 tumors before (c) and 1 h after (d) the intravenous injec-

dium disulfide (VS₂) nanodots, a MRI agent, labeling with 99mTc4+, a SPECT agent, inside PEG modified lipid micelle leading to fabricate the VS₂@lipid-PEG PNPs. In addition to MRI and SPECT, the high NIR absorbance of VS₂@lipid-PEG PNPs provides the PNPs with strong photoacoustic (PA) contrasts, resulting in multiple imaging functions. Keliher et al. [16] synthesize the ¹⁸F labeled polyglucose nanoparticles with size around 5 nm, can be excreted out of kidney and used to quantify the inflammation in atherosclerotic plaque using PET signal. Combination of PNPs with variant nuclear signaling agent enhances its image quality in cancer diagnosis and expands its application in other filed such as atherosclerotic detection.

tion of PEGMnCaP measured with 7T MRI. Scale bars, 50 μ m. (Image adapted from Kataoka et al. [15] and reprinted with permission from Springer Nature)

14.3.3 Optical

Optical imaging technique is typically faster and cheaper than magnetic and nuclear imaging. In addition, it can be used in high-throughput analysis in numerous cells which's interested Though the limited penetration deep of optical light hinders its clinical deployment, optical imaging can achieve highest spatial resolution in all of imaging modalities [60]. Therefore, currently, optical imaging has been widely used to observe molecular processes or detailed features of physiologic structures.

The most commonly used optical imaging agents are inorganic nanomaterials, such as semiconductors, metals and metal oxides. To provide these inorganic agents biocompatible and enhance the solubility/suspension in buffer media, several strategies have been studied to functionalize the surface of them with various amphiphilic block copolymers and to specific interact with target biomolecules [70].

In addition to inorganic nanomaterials, organic nanomaterials, such as conjugated polymer and fluorescent dye, can be also applied to bioimaging. Unlike other imaging modality, the unique opto-electronic properties of conjugated polymer can provide PNPs with intrinsic imaging ability without incorporation of imaging agent. Therefore, conjugated PNPs have widely been studied both *in vitro* and *in vivo* imaging [71].

On the other hand, current developments in the organic dyes design provide the new strategy in tuning the imaging properties of PNPs. Traditionally, the way to achieve high brightness in dye loaded PNPs depend on confining a large amount of dyes within the PNPs matrix. However, the aromatic structure of dye prefer to aggregate leading to fluorescence quenching. Recently, several studies have shown that PNPs can be used to increase the quantum yields of certain organic dyes, called aggregation-induced emission dyes (AIE dyes), by restricting the intramolecular rotation of these dyes within PNPs [72]. Klymchenko et al. [73] have reported that the brightness enhancement of AIE dyes-based PNPs are higher than quantum dots (QDs) for a comparable size, and have nearly reaching level of the brightest conjugated polymer-based nanoparticles.

Due to improve the brightness, biocompatibility, and selective targeting capability of these optical imaging agents, the development of PNPs for biomedical imaging have drawn attention [74, 75]. For example, Kim et al. [76] used the PEGylated Cornell dots (C dots) to observe the difference of propagation from cell to cell between the ferroptosis and other types of death. Hinde et al. [17] synthesize the fluorescein-labelled PNPs, based on poly(oligoethylene glycol methacrylate)- block-poly(styrene-covinylbenzaldehyde) P(OEGMA)-b-P (ST-co-PVBA) block copolymer, with different shapes but identical surface chemistries to understand how nanoparticle shapes will affect itself to access into the nucleus. Weissleder et al. [18] used the fluorescein-labelled dextran coated nanoparticles to examine the relationship between radiation effects and tumoral therapeutic nanoparticles concentration. Moreover, recently, several techniques have been used to enhance the accuracy and efficiency of imaging, including the surface-enhanced Raman spectroscopy (SERS) [19, 77], fluorescence resonance energy transfer (FRET) [20, 78], and photoacoustic effect [21, 79]. AIE dye, SERS or FRET etc. all these methodologies are developed for getting high image quality from optical signals. Combination of PNPs provides a way to enhance signal or improve biocompatibility of dye/metal/metal oxides contents to reach the goal.

14.4 Therapies

Incorporation of metal nanoparticles or organic molecule into PNPs not only can ensure a precise assessment of imaging for biomedical purposes, but also use in treating cancer, such as inhibition of tumor promotion or induction of cell death. Phototherapies based on nanotechnologies have attracted tremendous attention for their potential in biomedical application. Systematic classification of nanomaterials for phototherapies has been studied by Liu et al. [80]. Also, they review the different types of PNPs to deliver these nanomaterials to cancer cells. Zheng et al. [81] discuss about the photophysical relaxation pathways of organic molecules for phototheranostic techniques including the radiative emission, intersyscrossing/triplet-state tem relaxation and vibrational relaxation.

To monitor the treatment efficiency of therapy, most of researches will combine therapeutic PNPs with imaging agents enhancing the ability to visualize the targeted tissue. Elsabahy et al. [82] review the way to use PNPs to overcome challenges in imaging and therapy. The principle to design PNPs, such as components, type of structure and cross-linking, for delivery of diagnostic and therapeutic agents will be discussed. In the following, we will focus on the application of PNPs in photodynamic therapy (PDT) and photothermal therapy (PTT). In addition, we will also discuss the sonodynamic therapy (SDT) and neutron capture therapy (NCT), the promising strategies combining PNPs with low-intensity ultrasound or epithermal neutrons.

14.4.1 Photodynamic Therapy (PDT)

A promising biomedical application of PNPs based therapy is photodynamic therapy (PDT). PDT kill the cells via singlet oxygen or reactive oxygen species (ROS) generated from lightactivated chemical called a photosensitizer (PS). PS can transfer the absorbed light energy to either oxygen molecules to produce singlet oxygen or to surrounding molecules to form free radicals, leading to generation of different radical oxidizing agents, such as superoxide, hydrogen peroxide, and the hydroxyl radical [83]. Though, singlet oxygen only causes a destruction in nanometers, they will lead to activation of significant and complex cascade, resulting in local, regional, and systemic alteration of both cell and immune response [84]. Therefore, PDT has been extensively studied in the treatment of various disease, especially in cancers [10].

The development of PS has been comprehensive review by Zhang et al. [85]. Many photosensitizers used in PDT for cancer treatment are based on the tetrapyrrole backbone, a hydrophobic structure which is only slightly soluble and exhibits certain tendency to aggregate in aqueous resulting in a low therapeutic efficiency [86]. The combination of PS with PNPs not only decrease cytotoxicity of PS, but improve the solubility and stability of PS in water, which increase tumor accumulation of PS, which enhance the effect of PDT. Thus, many studies focused on design/synthesis different PNPs for effectively transport PS into specific tumor regions and then conduct the PDT by external light irradiation [80, 85]. Near-Infra red (NIR), due to relatively low, compared to visible light, absorption coefficient resulting the deep penetration in tissue, has been widely used in PDT treatment [87].

To increase the depth of PDT, recently, PS combined with upconversion nanoparticles (UCNPs) has attracted great attention. UCPNs is a materials converting low energy NIR light into high energy UV/visible light, which can activate the PS creating a photodynamic reaction [88]. Punjabi et al. [22] prepared the PNPs containing the PS (aminolevulinic acid) to convert it into protoporphyrin IX in the UCNPs treated by a biocompatible laser, which perform PDT for deep tissue penetration (>1.2 cm) than others with <1.0 cm depth).

To further penetrate into the deeper layer, scintillating nanoparticles (SCNP) have emerged as promising candidates for PDT application. After exposure to ionizing radiation, SCNP absorb the radiation and emit energy as visible light, which can trigger the PDT more efficiency. Cai et al. [89] summarized recent developments of SCNP. They also discuss about the strategies to loading SCNP, including the way to wrap it with polymer. Although the incorporation of SCNP within PNPs can overcome the penetration limit, the application of scintillating nanoparticles is still in infant stage. The relevant techniques for this strategy need to find ways to improve further.

14.4.2 Photothermal Therapy (PTT)

Another type for phototherapy is photothermal therapy (PTT). Different from PDT killing the cell by ROS, PTT use the heat to treat tumors. After light irradiation, the PTT agents will produce the photothermal effects, which can transform the energy of light into local heat. The heat not only can be used to cause thermal ablation of tumor cell, leading to cell membrane disruption and protein denaturation, but also can be helpful to address some of the limitations of nanodelivery systems, such as endosomal escape or cargo release. Therefore, to improve the therapeutic efficiency, PTT therapy has been combined with other therapeutic approaches. Kim et al. [90] revealed synergistic therapeutic systems combining gene and PTT ablation, they focus on how PTT effect enhanced cellular uptake, facilitate endosomal escape and induce

gene release for transfection. Zou et al. [91] showed when PTT combined with PNPs, it can be used alone or synergize with the imaging, radiotherapy, chemotherapy and immunotherapy to improve the efficiency of cancer treatment. For example, El-Sayed et al. [92] prepared the gold nanoparticles containing PNPs that can induce PPT cell death and molecular changes of single cell through real-time surface enhanced Raman spectroscopy (SERS). Chen et al. [23] design the PNPs, which combine the PTT with checkpoint-blockade immunotherapy, could eliminate primary tumors and inhibit metastases.

Different types of agents for PTT, including inorganic and organic nanomaterials, have been reported in various works [80, 93]. Inorganic nonmaterial, such as metallic nanomaterials, carbon nanostructures, quantum dots, and heavy metal nanocrystals and porous silicon nanomaterials, have shown the ability to efficiently convert light into heat. Shao et al. produced the biodegradable poly (lactic-co-glycolic acid) (PLGA) nanoparticle loaded with black phosphorus quantum dots (BPQDs), these hydrophobic PLGA polymer shell not only increased the photothermal stability via separation the BPQDs from oxygen and water, but mediated the BPQDs degradation [94]. On the other hand, organic nanomaterials, including organic compound and conjugated polymer based PNPs, have considered as potential agents in PTT due to their biodegradability. Among organic agents, conjugated polymer based PNPs have drawn great attention arising for their photothermal efficiencies, which is similar to gold nanoparticles [93]. Zhou et al. [95] prepare the conjugated polymer based PNPs, which can release the heat shock protein inhibitor. This inhibitor could reduce the cellular tolerance to heat resulted in better PTT effect.

14.4.3 Sonodynamic Therapy (SDT)

Different to the phototherapies conducting by light, sonodynamic therapy (SDT) executes the remedy by "sound". After activated by lowintensity ultrasound stimulation, specialized chemical agents, sonosensitizers, can produce ROS leading to cell damage. Due to ultrasound can non-invasive penetrate deeper to internal organs, compared to traditional phototherapies, SDT has attracted more attention in recent years.

The sonosensitizer, however, suffer the same problems as photosensitizer, such as low biological stability, tumor-accumulation. Therefore, to increase the SDT efficiency, several researchers have explored ways to combine PNPs with sonosensitizer, such as inorganic nanoparticles or organic compound. Xu et al. [96] confirmed the design of PNPs as carrier for sonosensitizer nanoparticles, including gold, Fe₃O₄, silver, porous silicon, and carbon fluoroxide, they introduce different methods to encapsulate the sonosensitizer into PNPs. Qian et al. [97] provides systematic description about the development of amplified SDT performance assisted by PNPs. A recent study demonstrated that, with proper design, MRI imaging can be displayed by sonosensitizers. Huang et al. [24] chelate MRI agents, Mn ion, to the sonosensitizers, protoporphyrinc (PpIx), and anchored the Mn chelateing sonosensitizers, Mn-protoporphyrin (MNPpIx), in to the inner mesoporous organosilica nanoparticles (HMONs-MnPpIX); then, the surface of HMONs-MnPpIX was also covalently modified with PEG, these PNPs made by HMONs-MnPpIX not only increased the SDT efficiency, but also augmented the Mn²⁺ ions chelating on protoporphyrin, these make this composite sonosensitizers with good MRI performance combined with SDT monitoring.

14.4.4 Neutron Capture Therapy (NCT)

Neutron capture therapy (NCT) is a treatment based on the nuclear reaction. Unlike aforementioned therapies which destroy the cells by ROS or heat, NCT kill the cell by gamma ray. After penetrate into the tissue, the epithermal neutrons would slow down and be captured by NCT agents, causing lethal radiation to injury in tumor cells. Boron (B) has been extensively



Fig. 14.3 Scheme of Gd-DTPA/CaP hybrid micelles targeting tumors for gadolinium neutron capture therapy (GdNCT). (a) The accumulation of Gd-DTPA delivered by Gd-DTPA/CaP in tumors through the EPR effect. (b) Low energy thermal neutron irradiation does not kill normal cells

studied as NCT agent. Subsequent to the capture of neutron, the nuclear fission of B could produce high energy alpha particles and lithium-7 nuclei [98].

To achieve sufficient deposition of NCT agents in a tumor site, NCT agents could be delivered by PNPs. For example, Huang et al. [99] synthesize the amphiphilic carboraneconjugated polycarbonates, which can selfassembly into different sizes in water, after irradiation of thermal neutron, the boron content of the carboranes conducts the NCT and suppresses tumor growth. In Kataoka's study [25], they replace the B with Gd, and loaded Gd into calcium phosphate core with hybridizing of PEG-polyanion block copolymers. There's two reasons to use Gd in their study: first, Gd has the largest capture crosssection, among NCT agents for thermal neutron absorption (larger than B) resulting in the emission of high energy gamma rays; the other reason is Gd can helpful for guiding the NCT under MRI (Fig. 14.3).

without NCT agents. (c) Thermal neutron irradiation could kill or cause hazardous damage to cancer cells by the γ -rays emitted from the Gd nuclides after nuclear reaction with captured thermal neutrons. (Image adapted from Mi et al. [25] and reprinted with permission from ACS publish)

14.5 Other Applications

The development of nanotechnology has been successfully selected as a alternative way to design, synthesize, and apply to present materials in biomedical filed. In addition to the abovementioned application, such as diagnosis and therapy, PNPs have been addressed in various applications. In the following, we roughly divide the application of PNPs into two categories by its condition; individual application and cluster employment.

14.5.1 PNPs Individual Application

In the preceding sections, we described how metal nanoparticles can be incorporated into PNPs to adhere to cells for imaging or treatment of them. However, the use of PNPs is not only limited to these. Recent studies have shown excellent performance of PNPs in mediation of cell behavior, biomolecular isolation, immunotherapy, protein affinity reagents and personalized medicine. In the following, we will introduce these applications.

PNPs with magnetic nanoparticle can control the mechanical stimulation of single-cell behavior. Tseng et al. [100] manipulate the dextranmagnetic nanoparticles, via magnetic fields, within HeLa cells. They apply localized nanoparticle-mediated forces producing the tension on the cortex of cells and observe the responses in cellular behavior. This technique offer a tool to analysis of molecular, such as proteins or nucleic acids, localization and its functional role in cell.

In addition to mediation of cell behavior, magnetic PNPs can be employed for separation the protein biomarkers. Nehilla et al. [101] use the classical temperature-responsive polymer poly (N-isopropylacrylamide), or pNIPAAM, to create the stimuli- responsive binary reagent system, including polymer coating magnetic nanoparticles (PNPs) and polymer-antibody conjugates (Ab), to capture the antigen. When the stimulus is applied, the PNPs and Ab with captured antigen will aggregate to form magnetically separable species. After removal of a stimulus, the captured antigen will release from PNPs. This system can customize for types of affinity reagents, providing a potential platform for biomarker discovery and diagnostics.

By combining immunotherapy with PNPs, nanoparticle platform can simulates the immune system to inhibit tumor growth is developed Kosmides et al. [102] prepare the "immunoswitch particle" by coating dextran-magnetic nanoparticles with two different antibodies that switch off the inhibitory checkpoint PD-L1 pathway on tumor cells and switch on CD8+T cells via the 4-1BB co-stimulatory pathway. These immunoswitch PNPs can increase effectiveness of immunotherapy over soluble antibody, resulting in reduction in cost and complexity of therapeutic.

PNPs are used for protein affinity reagent in biomedical application. Because the biological affinity reagents, such as antibody, have some disadvantage, including high cost, difficulties in production and storage. Several researchers have tried to try to find out alternatives. Hoshino et al. [103] prepare the PNPs, which interact strongly and weakly with denatured and native lysozyme respectively, to refold the aggregated lysozyme. Shea et al. [104] develop a PNPs can neutralize venomous biomacromolecules, phospholipase A₂ (PLA₂), inhibiting the hemolytic reaction. Koide et al. [105] synthesis the PNPs with affinity to a key vascular endothelial growth factor, VEGF₁₆₅, leading to inhibit VEGF-mediated angiogenesis without exhibition off-target activity.

PNPs can apply to personalized medicine to match specific patient with the most effective treatment. Schroeder et al. [106] load the different cancer drug and corresponding DNA barcodes into PNPs. Through the analysis of DNA barcodes, the correlation between the drug and cell viability is revealed finding out the most efficient drug. Similar to Schroeder et al., Dahlman et al. [107] use DNA barcoding to analyze the biodistribution of PNPs with varying PEG characteristics. The DNA barcoding can facilitate the researcher to understand the relationships between treatment and disease.

14.5.2 PNPs Cluster Employment

PNPs, especially spherical particles, have been used as building blocks to form close-packed two-dimensional (2D) or 3D structures so called self-assembled colloidal crystals [108, 109]. Various methods have been proposed such as evaporation induced colloidal self-assembly or self-assembly of colloids at air-water interfaces [109, 110]. The chosen method is often dependent on the particle material and size. The method and particle used also often determines the ultimate quality of crystal structure (i.e. the size of defect). High quality of 2D or 3D crystals is a common goal in various applications.

Colloidal crystal (CC) monolayers provide unique property which can be utilized in different fields such as photonics, sensors, and biology [111]. Recently, CC monolayers or CC-derived substrates have been utilized as a cell culture tool for modulating cell behavior [112, 113]. CC monolayers can be composed of single, binary, and even more types of particles. For example, binary colloidal crystal (BCC) monolayers have been fabricated on glass substrate in large area using large silicon particles and small polymer particles. Both 2D single and binary CCs have been used in cell culture. BCCs provide higher complexity on the surface than single CC such as heterogeneous chemistry. The ability of present more chemical or physical cues on cell adhering surface such as ternary CCs may elicit useful bioinformation.

Wang et al. [109] first used BCCs as a substrate for stem cell culture. BCCs were fabricated using evaporation-induced colloidal self-assembly. The detailed mechanism was described using this method. Depending on the BCC combinations, long ranged BCC can form on a 2D surface. Mammalian cells including MG63 osteoblasts, L929 fibroblasts, and human adipose stem cells (hASCs) were cultured on those long ranged BCCs, and cell spreading area was found to be inhibited on BCCs compared to flat controls. However, once cell adhesive protein, fibronectin, was pre-adsorbed onto BCCs, a synergic effect of BCCs and cell adhesive protein was found that cell spreading was significantly increased. Based on this result, hASCs were expanded on BCCs for longer term [113]. Interestingly, osteo- (BSP, RUNX2, and OPN) and chondro-genes (AGG, SOX9, and COL2) were upregulated on BCCs, but not adipogenes (PPAR γ and adiponetin). Thus, it has been demonstrated that BCCs can induce early osteochondral differentiation during stem cell expansion.

In another study, BCCs have been demonstrated that can replace the cell adhesive protein, vitronectin, during cell reprogramming of fibroblasts into induced pluripotent stem cells (iPSCs) (Fig. 14.4) [114]. Currently, the standard protocol for *in vitro* cell reprogramming needs pre-coating of extracellular proteins which support not only cell attachment but also provides abundant biosignals during cell growth. The result showed that human iPSC colonies formed on BCCs without vitronectin coating with high percentage while the colony was difficult to form on flat sur-



Fig. 14.4 PNPs self-assembly into monolayers as cell culture substrates. (a) Binary colloidal mixture self-assembled into crystal monolayer. (b) Scanning electron micrograph of one example, 2 micron silica and 0.1 micron polystyrene (2SiPS). (c) Atomic force microscopy

of 2SiPS. (d) Human adipose stem cells (hASCs) grew on 2SiPS for 4 days and formed clumps on the surface. (Image adapted from Wang et al. [114] and reprinted with permission from Springer Nature)

face without vitronectin. This result implies that BCCs regulates the cytoskeleton of fibroblasts and/or promotes extracellular matrix synthesis by fibroblasts which in turns facilitate cell reprogramming process. Overall, PNPs can be selfassembled into monolayer as a mask to producing biochemical patterns or substrate for *in vitro* cell expansion.

14.6 Conclusions

Although negative impacts of nanoparticle on patient tissue, such as liver [115], lung [116] and cardiovascular [117] were reported, due to some PNPs have capacity to cause inflammatory condition of lung, or accumulated lung to weaken the immune system, then activate a portion of latent virus in body, therefore, the proper design of PNPs can help decrease the risk of problems we mention.

Due to their good biocompatibility, large surface area to volume ratio, and versatile design, PNPs are considered competitive potential system for biomedical application. As a delivery system, PNPs not only can protect and transport of cargo, such as drug, plasmid or protein, more specific into the desired target site but can "smart" control of cargo release rate from PNPs. Also, via entrapment of metallic nanoparticles or fluorescent organic compounds, PNPs could serve as imaging or therapy platform to observe or destroy the specific tissue. Moreover, PNPs could be used to control of cell behavior, biomolecule isolation, protein affinity reagents and surface fabrication. Recently, by associating specific PNPs with photoreactvie agents as therapeutics, PNPs become a powerful tool for the application of personalized medicine to treat and improve cancer care.

In this chapter, we introduce some recent publications, including original and review articles with new agent or nanoparticles combination. All of them show that PNPs offer the new possibilities to develop both new diagnostic and therapeutic ways for biomedical application. We believe that PNPs could create more medical breakthroughs in the future.

References

- Kreuter J, Speiser PP (1976) *In vitro* studies of poly(methyl methacrylate) adjuvants. J Pharm Sci 65(11):1624–1627. https://doi.org/10.1002/ jps.2600651115
- Rao JP, Geckeler KE (2011) Polymer nanoparticles: preparation techniques and size-control parameters. Prog Polym Sci 36(7):887–913. https://doi. org/10.1016/j.progpolymsci.2011.01.001
- Albanese A, Tang PS, Chan WCW (2012) The effect of nanoparticle size, shape, and surface chemistry on biological systems. Annu Rev Biomed Eng 14(1):1–16. https://doi.org/10.1146/ annurev-bioeng-071811-150124
- Canton I, Battaglia G (2012) Endocytosis at the nanoscale. Chem Soc Rev 41(7):2718–2739. https:// doi.org/10.1039/C2CS15309B
- Verma A, Stellacci F (2010) Effect of surface properties on nanoparticle–cell interactions. Small 6(1):12–21. https://doi.org/10.1002/ smll.200901158
- Liu X, Yang Y, Urban MW (2017) Stimuli-responsive polymeric nanoparticles. Macromol Rapid Commun 38(13). https://doi.org/10.1002/marc.201700030
- Molina M, Asadian-Birjand M, Balach J, Bergueiro J, Miceli E, Calderon M (2015) Stimuli-responsive nanogel composites and their application in nanomedicine. Chem Soc Rev 44(17):6161–6186. https:// doi.org/10.1039/C5CS00199D
- Li Y, Maciel D, Rodrigues J, Shi X, Tomás H (2015) Biodegradable polymer nanogels for drug/nucleic acid delivery. Chem Rev 115(16):8564–8608. https://doi.org/10.1021/cr500131f
- Tang Z, He C, Tian H, Ding J, Hsiao BS, Chu B, Chen X (2016) Polymeric nanostructured materials for biomedical applications. Prog Polym Sci 60:86–128. https://doi.org/10.1016/j. progpolymsci.2016.05.005
- Thakor AS, Gambhir SS (2013) Nanooncology: the future of cancer diagnosis and therapy. CA Cancer J Clin 63(6):395–418. https://doi.org/10.3322/caac.21199
- 11. Li HJ, Du JZ, Liu J, Du XJ, Shen S, Zhu YH, Wang XY, Ye XD, Nie SM, Wang J (2016) Smart superstructures with ultrahigh pH-sensitivity for targeting acidic tumor microenvironment: instantaneous size switching and improved tumor penetration. ACS Nano 10(7):6753–6761. https://doi.org/10.1021/ acsnano.6b02326
- 12. Yin H, Song C-Q, Suresh S, Wu Q, Walsh S, Rhym LH, Mintzer E, Bolukbasi MF, Zhu LJ, Kauffman K, Mou H, Oberholzer A, Ding J, Kwan S-Y, Bogorad RL, Zatsepin T, Koteliansky V, Wolfe SA, Xue W, Langer R, Anderson DG (2017) Structure-guided chemical modification of guide RNA enables potent non-viral *in vivo* genome editing. Nat Biotechnol 35:1179. https://doi.org/10.1038/nbt.4005
- Smith TT, Stephan SB, Moffett HF, McKnight LE, Ji W, Reiman D, Bonagofski E, Wohlfahrt ME, Pillai SPS, Stephan MT (2017) In situ programming

of leukaemia-specific T cells using synthetic DNA nanocarriers. Nat Nanotechnol 12:813. https://doi. org/10.1038/nnano.2017.57

- 14. Xue Y, Xu X, Zhang X-Q, Farokhzad OC, Langer R (2016) Preventing diet-induced obesity in mice by adipose tissue transformation and angiogenesis using targeted nanoparticles. Proc Natl Acad Sci 113(20):5552–5557. https://doi.org/10.1073/ pnas.1603840113
- 15. Mi P, Kokuryo D, Cabral H, Wu H, Terada Y, Saga T, Aoki I, Nishiyama N, Kataoka K (2016) A pHactivatable nanoparticle with signal-amplification capabilities for non-invasive imaging of tumour malignancy. Nat Nanotechnol 11:724. https://doi. org/10.1038/nnano.2016.72
- 16. Keliher EJ, Ye Y-X, Wojtkiewicz GR, Aguirre AD, Tricot B, Senders ML, Groenen H, Fay F, Perez-Medina C, Calcagno C, Carlucci G, Reiner T, Sun Y, Courties G, Iwamoto Y, Kim H-Y, Wang C, Chen JW, Swirski FK, Wey H-Y, Hooker J, Fayad ZA, Mulder WJM, Weissleder R, Nahrendorf M (2017) Polyglucose nanoparticles with renal elimination and macrophage avidity facilitate PET imaging in ischaemic heart disease. Nat Commun 8:14064. https://doi.org/10.1038/ncomms14064
- Hinde E, Thammasiraphop K, Duong HTT, Yeow J, Karagoz B, Boyer C, Gooding JJ, Gaus K (2016) Pair correlation microscopy reveals the role of nanoparticle shape in intracellular transport and site of drug release. Nat Nanotechnol 12:81. https://doi. org/10.1038/nnano.2016.160
- Miller MA, Chandra R, Cuccarese MF, Pfirschke C, Engblom C, Stapleton S, Adhikary U, Kohler RH, Mohan JF, Pittet MJ, Weissleder R (2017) Radiation therapy primes tumors for nanotherapeutic delivery via macrophage-mediated vascular bursts. Sci Transl Med 9(392):eaal0225. https://doi.org/10.1126/scitranslmed.aal0225
- Harmsen S, Huang R, Wall MA, Karabeber H, Samii JM, Spaliviero M, White JR, Monette S, O'Connor R, Pitter KL, Sastra SA, Saborowski M, Holland EC, Singer S, Olive KP, Lowe SW, Blasberg RG, Kircher MF (2015) Surfaceenhanced resonance Raman scattering nanostars for high-precision cancer imaging. Sci Transl Med 7(271):271ra277–271ra277. https://doi. org/10.1126/scitranslmed.3010633
- 20. Zhao CC, Zhang XL, Li KB, Zhu SJ, Guo ZQ, Zhang LL, Wang FY, Fei Q, Luo SH, Shi P, Tian H, Zhu WH (2015) Forster resonance energy transfer switchable self-assembled micellar nanoprobe: ratiometric fluorescent trapping of endogenous H₂S generation via Fluvastatin-stimulated upregulation. J Am Chem Soc 137(26):8490–8498. https://doi. org/10.1021/jacs.5b03248
- Pu KY, Shuhendler AJ, Jokerst JV, Mei JG, Gambhir SS, Bao ZN, Rao JH (2014) Semiconducting polymer nanoparticles as photoacoustic molecular imaging probes in living mice. Nat Nanotechnol 9(3):233– 239. https://doi.org/10.1038/nnano.2013.302

- 22. Punjabi A, Wu X, Tokatli-Apollon A, El-Rifai M, Lee H, Zhang Y, Wang C, Liu Z, Chan EM, Duan C, Han G (2014) Amplifying the red-emission of upconverting nanoparticles for biocompatible clinically used prodrug-induced photodynamic therapy. ACS Nano 8(10):10621–10630. https://doi. org/10.1021/nn505051d
- 23. Chen Q, Xu LG, Liang C, Wang C, Peng R, Liu Z (2016) Photothermal therapy with immune-adjuvant nanoparticles together with checkpoint blockade for effective cancer immunotherapy. Nat Commun 7. https://doi.org/10.1038/ncomms13193
- 24. Huang P, Qian X, Chen Y, Yu L, Lin H, Wang L, Zhu Y, Shi J (2017) Metalloporphyrin-encapsulated biodegradable nanosystems for highly efficient magnetic resonance imaging-guided sonodynamic cancer therapy. J Am Chem Soc 139(3):1275–1284. https://doi.org/10.1021/jacs.6b11846
- 25. Mi P, Dewi N, Yanagie H, Kokuryo D, Suzuki M, Sakurai Y, Li YM, Aoki I, Ono K, Takahashi H, Cabral H, Nishiyama N, Kataoka K (2015) Hybrid calcium phosphate-polymeric micelles incorporating gadolinium chelates for imaging-guided gadolinium neutron capture tumor therapy. ACS Nano 9(6):5913–5921. https://doi.org/10.1021/ acsnano.5b00532
- 26. Suk JS, Xu QG, Kim N, Hanes J, Ensign LM (2016) PEGylation as a strategy for improving nanoparticle-based drug and gene delivery. Adv Drug Deliv Rev 99:28–51. https://doi.org/10.1016/j. addr.2015.09.012
- 27. Maeda H (2012) Vascular permeability in cancer and infection as related to macromolecular drug delivery, with emphasis on the EPR effect for tumor-selective drug targeting. Proc Jpn Acad Ser B Phys Biol Sci 88(3):53–71. https://doi.org/10.2183/pjab.88.53
- Abu Lila AS, Kiwada H, Ishida T (2013) The accelerated blood clearance (ABC) phenomenon: clinical challenge and approaches to manage. J Control Release 172(1):38–47. https://doi.org/10.1016/j.jconrel.2013.07.026
- Bertrand N, Wu J, Xu XY, Kamaly N, Farokhzad OC (2014) Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. Adv Drug Deliv Rev 66:2–25. https://doi. org/10.1016/j.addr.2013.11.009
- 30. Nicolas J, Mura S, Brambilla D, Mackiewicz N, Couvreur P (2013) Design, functionalization strategies and biomedical applications of targeted biodegradable/biocompatible polymer-based nanocarriers for drug delivery. Chem Soc Rev 42(3):1147–1235. https://doi.org/10.1039/C2CS35265F
- 31. Ulbrich K, Hola K, Subr V, Bakandritsos A, Tucek J, Zboril R (2016) Targeted drug delivery with polymers and magnetic nanoparticles: covalent and non-covalent approaches, release control, and clinical studies. Chem Rev 116(9):5338–5431. https://doi.org/10.1021/acs.chemrev.5b00589
- 32. Hu C-MJ, Fang RH, Wang K-C, Luk BT, Thamphiwatana S, Dehaini D, Nguyen P,

Angsantikul P, Wen CH, Kroll AV, Carpenter C, Ramesh M, Qu V, Patel SH, Zhu J, Shi W, Hofman FM, Chen TC, Gao W, Zhang K, Chien S, Zhang L (2015) Nanoparticle biointerfacing by platelet membrane cloaking. Nature 526:118. https://doi. org/10.1038/nature15373

- 33. Dehaini D, Wei X, Fang RH, Masson S, Angsantikul P, Luk BT, Zhang Y, Ying M, Jiang Y, Kroll AV, Gao W, Zhang L (2017) Erythrocyte–platelet hybrid membrane coating for enhanced nanoparticle functionalization. Adv Mater 29(16). https://doi. org/10.1002/adma.201606209
- 34. Rao L, Bu L-L, Xu J-H, Cai B, Yu G-T, Yu X, He Z, Huang Q, Li A, Guo S-S, Zhang W-F, Liu W, Sun Z-J, Wang H, Wang T-H, Zhao X-Z (2015) Red blood cell membrane as a biomimetic nanocoating for prolonged circulation time and reduced accelerated blood clearance. Small 11(46):6225–6236. https://doi.org/10.1002/ smll.201502388
- 35. Tian H, Luo Z, Liu L, Zheng M, Chen Z, Ma A, Liang R, Han Z, Lu C, Cai L (2017) Cancer cell membrane-biomimetic oxygen nanocarrier for breaking hypoxia-induced chemoresistance. Adv Funct Mater 27(38):1703197-n/a. https://doi. org/10.1002/adfm.201703197
- 36. Kalomiraki M, Thermos K, Chaniotakis NA (2016) Dendrimers as tunable vectors of drug delivery systems and biomedical and ocular applications. Int J Nanomed 11:1–12
- 37. Kesharwani P, Lyer AK (2015) Recent advances in dendrimer-based nanovectors for tumortargeted drug and gene delivery. Drug Discov Today 20(5):536–547. https://doi.org/10.1016/j. drudis.2014.12.012
- Kesharwani P, Jain K, Jain NK (2014) Dendrimer as nanocarrier for drug delivery. Prog Polym Sci 39(2):268–307. https://doi.org/10.1016/j. progpolymsci.2013.07.005
- 39. Yang JP, Zhang Q, Chang H, Cheng YY (2015) Surface-engineered dendrimers in gene delivery. Chem Rev 115(11):5274–5300. https://doi. org/10.1021/cr500542t
- 40. Sharma AK, Gothwal A, Kesharwani P, Alsaab H, Iyer AK, Gupta U (2017) Dendrimer nanoarchitectures for cancer diagnosis and anticancer drug delivery. Drug Discov Today 22(2):314–326. https:// doi.org/10.1016/j.drudis.2016.09.013
- 41. Wei T, Chen C, Liu J, Liu C, Posocco P, Liu XX, Cheng Q, Huo SD, Liang ZC, Fermeglia M, Pricl S, Liang XJ, Rocchi P, Peng L (2015) Anticancer drug nanomicelles formed by self-assembling amphiphilic dendrimer to combat cancer drug resistance. Proc Natl Acad Sci U S A 112(10):2978–2983. https:// doi.org/10.1073/pnas.1418494112
- 42. Yang H, Leffler CT (2013) Hybrid dendrimer hydrogel/poly(lactic-co-glycolic acid) nanoparticle platform: an advanced vehicle for topical delivery of antiglaucoma drugs and a likely solution to improving compliance and adherence in glaucoma man-

agement. J Ocul Pharmacol Ther 29(2):166–172. https://doi.org/10.1089/jop.2012.0197

- Bose RJC, Ravikumar R, Karuppagounder V, Bennet D, Rangasamy S, Thandavarayan RA (2017) Lipid– polymer hybrid nanoparticle-mediated therapeutics delivery: advances and challenges. Drug Discov Today 22(8):1258–1265. https://doi.org/10.1016/j. drudis.2017.05.015
- 44. Teixeira MC, Carbone C, Souto EB (2017) Beyond liposomes: recent advances on lipid based nanostructures for poorly soluble/poorly permeable drug delivery. Prog Lipid Res 68:1–11. https://doi. org/10.1016/j.plipres.2017.07.001
- 45. Yingchoncharoen P, Kalinowski DS, Richardson DR (2016) Lipid-based drug delivery systems in cancer therapy: what is available and what is yet to come. Pharmacol Rev 68(3):701–787. https://doi.org/10.1124/pr.115.012070
- 46. Yao C, Wang P, Li X, Hu X, Hou J, Wang L, Zhang F (2016) Near-infrared-triggered azobenzeneliposome/upconversion nanoparticle hybrid vesicles for remotely controlled drug delivery to overcome cancer multidrug resistance. Adv Mater 28(42):9341–9348. https://doi.org/10.1002/ adma.201503799
- 47. Rajala A, Wang YH, Zhu Y, Ranjo-Bishop M, Ma JX, Mao CB, Rajala RV (2014) Nanoparticle-assisted targeted delivery of eye-specific genes to eyes significantly improves the vision of blind mice in vivo. Nano Lett 14:5257–5263. https://doi.org/10.1021/ nl502275s
- Wang YH, Rajala A, Rajala RVS (2015) Lipid nanoparticles for ocular gene delivery. J Funct Biomater 6:379–394. https://doi.org/10.3390/ jfb6020379
- 49. Muthu MS, Leong DT, Mei L, Feng S-S (2014) Nanotheranostics - application and further development of nanomedicine strategies for advanced theranostics. Theranostics 4(6):660–677. https://doi. org/10.7150/thno.8698
- Suffredini G, East JE, Levy LM (2014) New applications of nanotechnology for neuroimaging. Am J Neuroradiol 35(7):1246–1253. https://doi. org/10.3174/ajnr.A3543
- 51. Gaitzsch J, Huang X, Voit B (2016) Engineering functional polymer capsules toward smart nanoreactors. Chem Rev 116(3):1053–1093. https:// doi.org/10.1021/acs.chemrev.5b00241
- Kamaly N, Yameen B, Wu J, Farokhzad OC (2016) Degradable controlled-release polymers and polymeric nanoparticles: mechanisms of controlling drug release. Chem Rev 116(4):2602–2663. https://doi. org/10.1021/acs.chemrev.5b00346
- 53. Gibori H, Eliyahu S, Krivitsky A, Ben-Shushan D, Epshtein Y, Tiram G, Blau R, Ofek P, Lee JS, Ruppin E, Landsman L, Barshack I, Golan T, Merquiol E, Blum G, Satchi-Fainaro R (2018) Amphiphilic nanocarrier-induced modulation of PLK1 and miR-34a leads to improved therapeutic response in pan-

creatic cancer. Nat Commun 9(1):16. https://doi. org/10.1038/s41467-017-02283-9

- Peppas NA (1996) An introduction to materials in medicine. In: Ratner BD, Hoffman AS, Schoen FJ, Lemons JE (eds) Biomaterials science. Academic, New York, pp 60–64
- 55. Tseng C, Su W, Yen K, Yang K, Lin F (2009) The use of biotinylated-EGF-modified gelatin nanoparticle carrier to enhance cisplatin accumulation in cancerous lungs via inhalation. Biomaterials 30:3476–3485. https://doi.org/10.1016/j. biomaterials.2009.03.010
- 56. Tseng C, Chen K, Su W, Lee Y, Wu C, Fang H, Lin F (2013) Cationic gelatin nanoparticles for drug delivery to the ocular surface: in vitro and in vivo evaluation. J Nanomater 2013:1–11. https://doi.org/10.1155/2013/238351
- 57. Tseng CL, Peng CL, Huang JY, Chen JC, Lin FH (2012) Gelatin nanoparticles as gene carriers for transgenic chicken applications. J Biomater Appl 27(8):1055–1065. https://doi.org/10.1177/0885328211434089
- Yu M, Zheng J (2015) Clearance pathways and tumor targeting of imaging nanoparticles. ACS Nano 9(7):6655–6674. https://doi.org/10.1021/ acsnano.5b01320
- Kunjachan S, Ehling J, Storm G, Kiessling F, Lammers T (2015) Noninvasive imaging of nanomedicines and nanotheranostics: principles, progress, and prospects. Chem Rev 115(19):10907– 10937. https://doi.org/10.1021/cr500314d
- Smith BR, Gambhir SS (2017) Nanomaterials for in vivo imaging. Chem Rev 117(3):901–986. https:// doi.org/10.1021/acs.chemrev.6b00073
- Estelrich J, Sanchez-Martin MJ, Busquets MA (2015) Nanoparticles in magnetic resonance imaging: from simple to dual contrast agents. Int J Nanomed 10:1727–1741. https://doi.org/10.2147/ijn.s76501
- Tseng C-L, Shih IL, Stobinski L, Lin F-H (2010) Gadolinium hexanedione nanoparticles for stem cell labeling and tracking via magnetic resonance imaging. Biomaterials 31(20):5427–5435. https://doi. org/10.1016/j.biomaterials.2010.03.049
- 63. Zhang L, Liu R, Peng H, Li P, Xu Z, Whittaker AK (2016) The evolution of gadolinium based contrast agents: from single-modality to multimodality. Nanoscale 8(20):10491–10510. https:// doi.org/10.1039/C6NR00267F
- 64. Bakhtiary Z, Saei AA, Hajipour MJ, Raoufi M, Vermesh O, Mahmoudi M (2016) Targeted superparamagnetic iron oxide nanoparticles for early detection of cancer: possibilities and challenges. Nanomedicine 12(2):287–307. https://doi. org/10.1016/j.nano.2015.10.019
- 65. Goel S, England CG, Chen F, Cai W (2017) Positron emission tomography and nanotechnology: a dynamic duo for cancer theranostics. Adv Drug Deliv Rev 113:157–176. https://doi.org/10.1016/j. addr.2016.08.001

- 66. Morais GR, Paulo A, Santos I (2012) Organometallic complexes for SPECT imaging and/or radionuclide therapy. Organometallics 31(16):5693–5714. https:// doi.org/10.1021/om300501d
- Pant K, Sedláček O, Nadar RA, Hrubý M, Stephan H (2017) Radiolabelled polymeric materials for imaging and treatment of cancer: quo vadis? Adv Healthc Mater 6(6):1601115-n/a. https://doi.org/10.1002/ adhm.201601115
- 68. Phillips E, Penate-Medina O, Zanzonico PB, Carvajal RD, Mohan P, Ye Y, Humm J, Gönen M, Kalaigian H, Schöder H, Strauss HW, Larson SM, Wiesner U, Bradbury MS (2014) Clinical translation of an ultrasmall inorganic optical-PET imaging nanoparticle probe. Sci Transl Med 6(260):260ra149. https:// doi.org/10.1126/scitranslmed.3009524
- 69. Chen Y, Cheng L, Dong Z, Chao Y, Lei H, Zhao H, Wang J, Liu Z (2017) Degradable vanadium disulfide nanostructures with unique optical and magnetic functions for cancer theranostics. Angew Chem Int Ed 56(42):12991–12996. https://doi.org/10.1002/ anie.201707128
- 70. Palui G, Aldeek F, Wang W, Mattoussi H (2015) Strategies for interfacing inorganic nanocrystals with biological systems based on polymer-coating. Chem Soc Rev 44(1):193–227. https://doi.org/10.1039/ C4CS00124A
- Feng L, Zhu C, Yuan H, Liu L, Lv F, Wang S (2013) Conjugated polymer nanoparticles: preparation, properties, functionalization and biological applications. Chem Soc Rev 42(16):6620–6633. https://doi. org/10.1039/C3CS60036J
- Zhang X, Wang K, Liu M, Zhang X, Tao L, Chen Y, Wei Y (2015) Polymeric AIE-based nanoprobes for biomedical applications: recent advances and perspectives. Nanoscale 7(27):11486–11508. https:// doi.org/10.1039/C5NR01444A
- Reisch A, Klymchenko AS (2016) Fluorescent polymer nanoparticles based on dyes: seeking brighter tools for bioimaging. Small 12(15):1968–1992. https://doi.org/10.1002/smll.201503396
- Chen M, Yin M (2014) Design and development of fluorescent nanostructures for bioimaging. Prog Polym Sci 39(2):365–395. https://doi.org/10.1016/j. progpolymsci.2013.11.001
- Peng H-S, Chiu DT (2015) Soft fluorescent nanomaterials for biological and biomedical imaging. Chem Soc Rev 44(14):4699–4722. https://doi.org/10.1039/ C4CS00294F
- 76. Kim SE, Zhang L, Ma K, Riegman M, Chen F, Ingold I, Conrad M, Turker MZ, Gao M, Jiang X, Monette S, Pauliah M, Gonen M, Zanzonico P, Quinn T, Wiesner U, Bradbury MS, Overholtzer M (2016) Ultrasmall nanoparticles induce ferroptosis in nutrient-deprived cancer cells and suppress tumour growth. Nat Nanotechnol 11:977. https://doi. org/10.1038/nnano.2016.164
- 77. Gao X, Yue Q, Liu Z, Ke M, Zhou X, Li S, Zhang J, Zhang R, Chen L, Mao Y, Li C (2017) Guiding braintumor surgery via blood–brain-barrier-permeable

gold nanoprobes with acid-triggered MRI/SERRS signals. Adv Mater 29(21). https://doi.org/10.1002/adma.201603917

- Hildebrandt N, Spillmann CM, Algar WR, Pons T, Stewart MH, Oh E, Susumu K, Díaz SA, Delehanty JB, Medintz IL (2017) Energy transfer with semiconductor quantum dot bioconjugates: a versatile platform for biosensing, energy harvesting, and other developing applications. Chem Rev 117(2):536– 711. https://doi.org/10.1021/acs.chemrev.6b00030
- 79. Weber J, Beard PC, Bohndiek SE (2016) Contrast agents for molecular photoacoustic imaging. Nat Methods 13:639. https://doi.org/10.1038/ nmeth.3929
- Cheng L, Wang C, Feng L, Yang K, Liu Z (2014) Functional nanomaterials for phototherapies of cancer. Chem Rev 114(21):10869–10939. https:// doi.org/10.1021/cr400532z
- Ng KK, Zheng G (2015) Molecular interactions in organic nanoparticles for phototheranostic applications. Chem Rev 115(19):11012–11042. https://doi. org/10.1021/acs.chemrev.5b00140
- Elsabahy M, Heo GS, Lim S-M, Sun G, Wooley KL (2015) Polymeric nanostructures for imaging and therapy. Chem Rev 115(19):10967–11011. https:// doi.org/10.1021/acs.chemrev.5b00135
- Henderson BW, Dougherty TJ (1992) How does photodynamic therapy work ? Photochem Photobiol 55(1):145–157. https://doi.org/10.1111/ j.1751-1097.1992.tb04222.x
- Allison RR, Moghissi K (2013) Photodynamic therapy (PDT): PDT mechanisms. ClinEndosc 46(1):24– 29. https://doi.org/10.5946/ce.2013.46.1.24
- Lucky SS, Soo KC, Zhang Y (2015) Nanoparticles in photodynamic therapy. Chem Rev 115(4):1990– 2042. https://doi.org/10.1021/cr5004198
- Abrahamse H, Hamblin MR (2016) New photosensitizers for photodynamic therapy. Biochem J 473:347–364. https://doi.org/10.1042/bj20150942
- Shanmugam V, Selvakumar S, Yeh CS (2014) Nearinfrared light-responsive nanomaterials in cancer therapeutics. Chem Soc Rev 43(17):6254–6287. https://doi.org/10.1039/c4cs00011k
- Zhou B, Shi BY, Jin DY, Liu XG (2015) Controlling upconversion nanocrystals for emerging applications. Nat Nanotechnol 10(11):924–936. https://doi. org/10.1038/nnano.2015.251
- Kamkaew A, Chen F, Zhan Y, Majewski RL, Cai W (2016) Scintillating nanoparticles as energy mediators for enhanced photodynamic therapy. ACS Nano 10(4):3918–3935. https://doi.org/10.1021/ acsnano.6b01401
- Kim J, Kim J, Jeong C, Kim WJ (2016) Synergistic nanomedicine by combined gene and photothermal therapy. Adv Drug Deliv Rev 98:99–112. https://doi. org/10.1016/j.addr.2015.12.018
- 91. Zou LL, Wang H, He B, Zeng LJ, Tan T, Cao HQ, He XY, Zhang ZW, Guo SR, Li YP (2016) Current approaches of photothermal therapy in treating can-

cer metastasis with nanotherapeutics. Theranostics 6(6):762–772. https://doi.org/10.7150/thno.14988

- Aioub M, El-Sayed MA (2016) A real-time surface enhanced raman spectroscopy study of plasmonic photothermal cell death using targeted gold nanoparticles. J Am Chem Soc 138(4):1258–1264. https:// doi.org/10.1021/jacs.5b10997
- 93. Jaque D, Martinez Maestro L, del Rosal B, Haro-Gonzalez P, Benayas A, Plaza JL, Martin Rodriguez E, Garcia Sole J (2014) Nanoparticles for photothermal therapies. Nanoscale 6(16):9494–9530. https:// doi.org/10.1039/C4NR00708E
- 94. Shao J, Xie H, Huang H, Li Z, Sun Z, Xu Y, Xiao Q, Yu X-F, Zhao Y, Zhang H, Wang H, Chu PK (2016) Biodegradable black phosphorus-based nanospheres for in vivo photothermal cancer therapy. Nat Commun 7:12967. https://doi.org/10.1038/ncomms12967
- 95. Liu D, Ma L, An Y, Li Y, Liu Y, Wang L, Guo J, Wang J, Zhou J (2016) Thermoresponsive nanogelencapsulated PEDOT and HSP70 inhibitor for improving the depth of the photothermal therapeutic effect. Adv Funct Mater 26(26):4749–4759. https:// doi.org/10.1002/adfm.201600031
- 96. Xu H, Zhang X, Han R, Yang P, Ma H, Song Y, Lu Z, Yin W, Wu XX, Wang H (2016) Nanoparticles in sonodynamic therapy: state of the art review. RSC Adv 6(56):50697–50705.
- 97. Qian XQ, Zheng YY, Chen Y (2016) Micro/ nanoparticle-augmented sonodynamic therapy (SDT): breaking the depth shallow of photoactivation. Adv Mater 28(37):8097–8129. https://doi. org/10.1002/adma.201602012
- 98. Barth RF, Coderre JA, Vicente MGH, Blue TE (2005) Boron neutron capture therapy of cancer: current status and future prospects. Clin Cancer Res 11(11):3987–4002. https://doi.org/10.1158/1078-0432.Ccr-05-0035
- 99. Xiong H, Wei X, Zhou D, Qi Y, Xie Z, Chen X, Jing X, Huang Y (2016) Amphiphilic polycarbonates from carborane-installed cyclic carbonates as potential agents for boron neutron capture therapy. Bioconjug Chem 27(9):2214–2223. https://doi. org/10.1021/acs.bioconjchem.6b00454
- 100. Tseng P, Judy JW, Di Carlo D (2012) Magnetic nanoparticle-mediated massively parallel mechanical modulation of single-cell behavior. Nat Methods 9:1113. https://doi.org/10.1038/ nmeth.2210
- 101. Nehilla BJ, Hill JJ, Srinivasan S, Chen Y-C, Schulte TH, Stayton PS, Lai JJ (2016) A stimuli-responsive, binary reagent system for rapid isolation of protein biomarkers. Anal Chem 88(21):10404–10410. https://doi.org/10.1021/acs.analchem.6b01961
- 102. Kosmides AK, Sidhom J-W, Fraser A, Bessell CA, Schneck JP (2017) Dual targeting nanoparticle stimulates the immune system to inhibit tumor growth. ACS Nano 11(6):5417–5429. https://doi. org/10.1021/acsnano.6b08152

- 103. Nakamoto M, Nonaka T, Shea KJ, Miura Y, Hoshino Y (2016) Design of synthetic polymer nanoparticles that facilitate resolubilization and refolding of aggregated positively charged lysozyme. J Am Chem Soc 138(13):4282–4285. https://doi.org/10.1021/ jacs.5b12600
- 104. O'Brien J, Lee SH, Onogi S, Shea KJ (2016) Engineering the protein corona of a synthetic polymer nanoparticle for broad-spectrum sequestration and neutralization of venomous biomacromolecules. J Am Chem Soc 138(51):16604–16607. https://doi. org/10.1021/jacs.6b10950
- 105. Koide H, Yoshimatsu K, Hoshino Y, Lee S-H, Okajima A, Ariizumi S, Narita Y, Yonamine Y, Weisman AC, Nishimura Y, Oku N, Miura Y, Shea KJ (2017) A polymer nanoparticle with engineered affinity for a vascular endothelial growth factor (VEGF165). Nat Chem 9:715. https://doi. org/10.1038/nchem.2749
- 106. Yaari Z, da Silva D, Zinger A, Goldman E, Kajal A, Tshuva R, Barak E, Dahan N, Hershkovitz D, Goldfeder M, Roitman JS, Schroeder A (2016) Theranostic barcoded nanoparticles for personalized cancer medicine. Nat Commun 7:13325. https://doi. org/10.1038/ncomms13325
- 107. Dahlman JE, Kauffman KJ, Xing Y, Shaw TE, Mir FF, Dlott CC, Langer R, Anderson DG, Wang ET (2017) Barcoded nanoparticles for high throughput in vivo discovery of targeted therapeutics. Proc Natl Acad Sci 114(8):2060–2065. https://doi. org/10.1073/pnas.1620874114
- 108. Parchine M, McGrath J, Bardosova M, Pemble ME (2016) Large area 2D and 3D colloidal photonic crystals fabricated by a roll-to-roll langmuir-blodgett method. Langmuir 32(23):5862–5869. https://doi. org/10.1021/acs.langmuir.6b01242
- 109. Wang P-Y, Pingle H, Koegler P, Thissen H, Kingshott P (2015) Self-assembled binary colloidal crystal monolayers as cell culture substrates. J Mater Chem B 3(12):2545–2552. https://doi.org/10.1039/ C4TB02006E
- 110. Wang P-Y, Bennetsen DT, Foss M, Ameringer T, Thissen H, Kingshott P (2015) Modulation of human mesenchymal stem cell behavior on ordered tantalum nanotopographies fabricated using colloi-

dal lithography and glancing angle deposition. ACS Appl Mater Interfaces 7(8):4979–4989. https://doi. org/10.1021/acsami.5b00107

- 111. Li F, Josephson DP, Stein A (2011) Colloidal assembly: the road from particles to colloidal molecules and crystals. Angew Chem Int Ed 50(2):360–388. https://doi.org/10.1002/anie.201001451
- 112. Ji LJ, LaPointe VLS, Evans ND, Stevens MM (2012) Changes in embroynic stem cell colony mophology and early differentiation markers driven by colloidal crystal topographical cues. Eur Cell Mater 23:135–146. https://doi.org/10.22203/ eCM.v023a10
- 113. Wang P-Y, Thissen H, Kingshott P (2016) Stimulation of early osteochondral differentiation of human mesenchymal stem cells using binary colloidal crystals (BCCs). ACS Appl Mater Interfaces 8(7):4477–4488. https://doi.org/10.1021/ acsami.5b12660
- 114. Wang P-Y, Hung SS-C, Thissen H, Kingshott P, Wong RC-B (2016) Binary colloidal crystals (BCCs) as a feeder-free system to generate human induced pluripotent stem cells (hiPSCs). Sci Rep 6:36845. https://doi.org/10.1038/srep36845
- 115. Esch MB, Mahler GJ, Stokol T, Shuler ML (2014) Body-on-a-chip simulation with gastrointestinal tract and liver tissues suggests that ingested nanoparticles have the potential to cause liver injury. Lab Chip 14(16):3081–3092. https://doi.org/10.1039/ C4LC00371C
- 116. Sattler C, Moritz F, Chen S, Steer B, Kutschke D, Irmler M, Beckers J, Eickelberg O, Schmitt-Kopplin P, Adler H, Stoeger T (2017) Nanoparticle exposure reactivates latent herpesvirus and restores a signature of acute infection. Part Fibre Toxicol 14:2. https://doi.org/10.1186/ s12989-016-0181-1
- 117. Miller MR, Raftis JB, Langrish JP, McLean SG, Samutrtai P, Connell SP, Wilson S, Vesey AT, Fokkens PHB, Boere AJF, Krystek P, Campbell CJ, Hadoke PWF, Donaldson K, Cassee FR, Newby DE, Duffin R, Mills NL (2017) Inhaled nanoparticles accumulate at sites of vascular disease. ACS Nano 11(5):4542–4552. https://doi.org/10.1021/ acsnano.6b08551