# **Chapter 4 Potentiality of Anticancer Plant-Derived Compounds of North-East India**



Mohan Lal, Nibir Ranjan Parasar, Anil Kumar Singh, and Mohd Sayeed Akhtar

### 4.1 Introduction

Cancer is a severe metabolic syndrome and the leading cause of mortality and morbidity worldwide with the number of cases increasing every year (Sharma et al. 2014; ACS 2016). In developed nations, this disease ranks second in death cases after cardiovascular disorders (Mbaveng et al. 2011; Siegel et al. 2016). The incidence of mortality and prevalence from major types of cancer as estimated by International Agency for Research on Cancer of 184 countries of the world revealed that there were 8.2 million cancer deaths, and 14.1 million new cancer cases, worldwide and it is projected that by 2030 there will be 26 million new cancer cases and 17 million cancer deaths per year (Thun and De Lancey 2010). Cancer is characterized by uncontrolled proliferation and dedifferentiation of normal cell. A typical cancer cell has marked attributes, viz. sends signals of proliferation and differentiation and is capable to sustain proliferation; they have the power of invasion and angiogenesis, and they overcome apoptosis (Sharma et al. 2014). Transformation from normal cell to malignant cell involves a sequence of alterations producing genetic instabilities which accumulate in a cell. Alterations such as mutation in DNA repair genes, oncogenes, apoptotic genes, tumour suppressor genes and gene involved in cell growth and differentiation are prominent (Sharma et al. 2014).

A. K. Singh Biotechnology Group, Biological Sciences and Technology Division, CSIR-North East Institute of Science and Technology, Jorhat, Assam, India

M. S. Akhtar Department of Botany, Gandhi Faiz-e-Aam College, Shahjahanpur, Uttar Pradesh, India

© Springer Nature Singapore Pte Ltd. 2018

M. S. Akhtar, M. K. Swamy (eds.), Anticancer Plants: Properties and Application, https://doi.org/10.1007/978-981-10-8548-2\_4

M. Lal (🖂) · N. R. Parasar

Medicinal, Aromatic and Economic Plants Group, Biological Sciences and Technology Division, CSIR-North East Institute of Science and Technology, Jorhat, Assam, India e-mail: mohan@neist.res.in

Cancer is caused by both internal (e.g. hormones, gene mutations and immune conditions) and external (e.g. smoking, radiation and pollution) factors.

North-East India is one of the nine global biodiversity hotspots lying between 22-30°N latitude and 89-97°E longitude. This region is blessed with varying flora with diversified topography and climatic conditions marked by high humidity, moderate temperature and high rainfall. There are abundant dense forests, swamps, marshes, etc. that engulf the region with vegetation ranging from tropical to the alpine forests. Different tribes of North-East India rely mostly on the ethnic traditional herbal medicine due to lack of adequate modern medical facilities (Sviem and Kharbuli 1999; Rosangkima et al. 2010; Tushara et al. 2010). The crude herbal preparations are applied by the herbal practitioners with additives, viz. milk, curd, ghee honey, etc., as adjuvant in order to enhance the effect of the respective herbal preparations (Behere et al. 2013). The aim of the present chapter is to focus on the potentiality of major plant-derived compounds from diversified medicinal plants of North-East India for their anticancer and chemopreventive activity and their mode of action. Moreover, their large-scale production, uses of structural analogs, and molecular docking studies of some of the selected plant-derived compounds are also discussed.

# 4.2 Plants as an Imperative Source of Chemopreventive Phytochemicals

Since ancient times herbal formulations are used for medicinal purposes. Herbal practitioners apply various herbal formulations which are based on different philosophies and cultural origins to heal diseases. Traditional knowledge, viz. Ayurveda, Kampo, Egyptian medicine and traditional Chinese medicine, is the science of good health and well-being (Hashimoto et al. 2000; Rosangkima et al. 2010; Sharma et al. 2014). In the recent times, hunt for novel phytochemicals for drug development based on the concepts of traditional knowledge has gained wide acceptance. Natural products derived from plants are non-toxic thereby making them ideal candidates for modern drug discovery. Only 10% of the 250,000 plant species have been investigated for therapeutic applications, and more than 50% of all the modern drugs are derived from plants. Varieties of phytochemicals and their derived metabolites are present in the bark, root, leaves, stem and flower which serve an array of pharmacological activity in human health system. Phenolics, alkaloids, flavonoids, glycosides, tannins, oils and gums are responsible compounds for various therapeutic purposes. A significant antitumour activity has been shown by altered forms of these compounds. Curcumol, betulinic acid, kaempferol, ellagic acid, dillenetin, L-borneol, taxol, tangeretin, naringin and resveratrol are some of the remarkable chemopreventive phytochemicals as lead molecules for the development of anticancer drugs (Table 4.1).

		Suppressed cancerous cell lines/cancer			
Phytochemicals	Plants	models	Uses	References	
Kaempferol	Ageratum conyzoides	Lung cancer (A-549), gastric carcinoma (SGC-7901), colon carcinoma (HT-29), human glioma carcinoma (U-251), breast cancer (MDA-MB-231), prostate cancer (DU-145), hepatic carcinoma (BEL- 7402), mouse leukaemia (P-388)	In clinical use	Adebayo et al. (2010)	
Nimbolide	Azadirachta indica	Lung cancer (U937), leukaemia (HL-60, THP1), skin melanoma (B16), prostate cancer (PC-3)	In preclinical development	Baral and Chattopadhyay (2004), Giri and Lakshmi Narasu (2000) and Kumar et al. (2006)	
Taxol	Taxus baccata	Breast cancer (HER2, MDA-MB- 435), ovary (SK-OV-3 w)	In clinical use	Baselga et al. (1998) and Aggarwal and Shishodia (2005)	
$\Delta^9$ - Tetrahydrocannabinol	Cannabis sativa	Breast cancer (MCF-7, EFM-19, MDA-MB-231), skin cancer (PDV.C57, HaCa4), brain/spine tumour (U87, U373)	In preclinical development	Casanova et al. (2003), Massi et al. (2004), Cheung and Tai (2007) and Yesil-Celiktas et al. (2010)	
Curcumin	Curcuma longa	Breast cancer (BT-20,T-47D, SK-BR3 and MCF-7), leukaemia (HL60)	In preclinical use	Cui et al. (2006) and Magesh et al. (2009)	
6-Shogaol, (6)-gingerol	Zingiber officinale	Breast cancer (MCF-7 and MDA-MB-231), colon cancer (HCT 116, HT 29), ovarian cancer (SK-OV-3), lung cancer (A549), melanoma (SK-MEL-2), colorectal adenocarcinoma (HCT15)	In clinical use	De Petrocellis et al. (1998), Kim (2008) and Ligresti et al. (2006)	

 Table 4.1
 Chemopreventive phytochemicals derived from plants available in North-East India

(continued)

Phytochemicals	Plants	Suppressed cancerous cell lines/cancer models	Uses	References
Dillenetin and betulinic acid	Dillenia indica	Lung cancer (U937), promyelocytic leukaemia (HL60, K562)	In preclinical development	Gandhi and Mehta (2013)
Alexin B, emodin	Aloe vera	Liver cancer (HepG2), breast cancer (MCF-7), cervical cancer	In preclinical development	Hussain et al. (2015) and Noorolahi et al. (2016)
Taxol	T. baccata	Breast cancer (BT-474, SK-BR-3 and MCF7)		Klauber et al. (1997)
Dillenetin and betulinic acid	Dillenia pentagyna	T-cell lymphoma	In preclinical development	Mehta et al. (1997) and Rosangkima and Prasad (2007)
Catechin	Potentilla fulgens	Breast cancer (MCF-7), human glioblastoma cancer (U-87)	In clinical use	Mittal and Tripathy (2015)
Dihydroflavonol	Blumea balsamifera	Breast cancer (MCF-7), epidermal carcinoma of the mouth (KB), myeloid leukaemia (K562), lung cancer (NCI- H187), hepatocellular carcinoma (McA-RH7777)	In preclinical development	Norikura et al. (2008)
Eugenol, orientin, vicenin	Ocimum sanctum	Lung cancer (A549), human fibrosarcoma cells (HFS)	In preclinical development	Roy et al. (2007)
Oleic acid and beta-sitosterol	Mirabilis jalapa	Human laryngeal carcinoma (Hep-2), breast cancer (MCF-7)	In preclinical development	Rumzhum et al. (2008) and Gogoi and Nakhuru (2016)
Vinblastine, vincristine	Catharanthus roseus	Lung cancer (NCI-H69/P)	In clinical use	Trevor and Theodore (1993)
Epicatechin, procyanidin B <sub>2</sub> , B <sub>4</sub>	Litchi chinensis	Breast cancer (MCF-7), leukaemia (U937, K562 and HL-60), colorectal cancer (Colo320DM and SW480)	In preclinical development	Twentyman et al. (1987), Lipinsky et al. (1997) and Hsu et al. (2012)

Table 4.1 (continued)

(continued)

Phytochemicals	Plants	Suppressed cancerous cell lines/cancer models	Uses	References
Etoposide, podophyllin, teniposide, podophyllotoxin	Podophyllum hexandrum	Lung cancer, testicular cancer, neuroblastoma, hepatoma	In clinical use	Uden (1989) and Abdullah and Abidin (2010)
Xanthatin, xanthinosin, 4-oxobedfordia	Xanthium strumarium	Cervical cancer	In preclinical development	Vaishnav et al. (2015)
Carnosic acid, rosmarinic acid	Rosmarinus officinalis	Breast cancer (MCF7 and MDA-MB-468), leukaemia (HL60, K-562), prostate cancer (DU-145), lung cancer (NCI-H82), liver cancer (Hep-3B), ovarian cancer (r A2780)	In preclinical development	Zhao et al. (2007), Roya et al. (2008) and Tai and Cheunga (2012)

Table 4.1 (continued)

## 4.3 Mechanism of Action and Molecular Targets of Chemopreventive Phytochemicals from North-East India

The precise mechanism of action of the bioactive molecules performing anticancer functions is an interesting area of current research. The usual targets of these molecules are the cytosolic and nuclear factors of a cancer cell. They either directly absorb the reactive oxygen species or stimulate the antioxidant enzymes, viz. catalase, glutathione and superoxide dismutase, in a transformed cell. The metabolic conversion of a procarcinogen is blocked by a phyto-molecule, or it suppresses malignant transformation of a preneoplastic cell. The cellular and signalling events involved in growth, invasion and metastasis are also regulated by these molecules. Curcumin (diferuloylmethane), a polyphenol present as a major phytochemical in the rhizome of *Curcuma longa*, is the most prominent chemopreventive bioactive molecule studied (Aggarwal et al. 2003; Fridlender et al. 2015). It has been used as a medicine for treatment of various diseases (Sharma et al. 2005; Fridlender et al. 2015; Lee and Kim 2016). Kaempferol, the major phytochemical in Potentilla fulgens, acts on proto-oncogene tyrosine protein kinase (Src), Erk1/2 and Akt pathways in pancreatic cancer cells and retards their growth and migration (Hossan et al. 2014). Ellagic acid from P. fulgens induces apoptosis in breast and prostate cancer cells and inhibits metastasis processes of various cancer types. Rosmarinic acid present in Ocimum basilicum reduces the activity of DNA methyltransferase and interferes OPG/ RANKL/RANK networks (Osakabe et al. 2004; Baliga et al. 2013). Besides, it also acts in colon cancer cells by reducing COX-2 activity and Erk phosphorylation. Moreover, it targets PKA/CREB/MITF pathway and NF-kB activation in melanoma

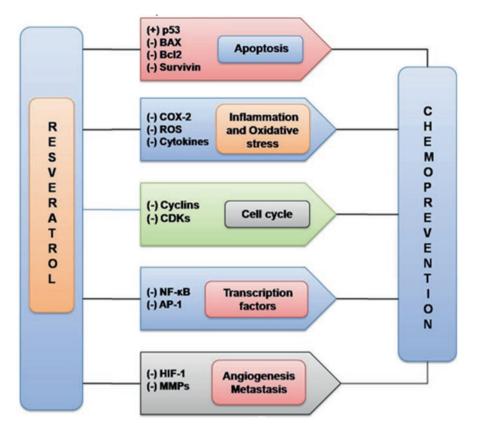


Fig. 4.1 Molecular targets of resveratrol leading to chemoprevention

and leukaemia U938 cells, respectively, thereby stimulating anti-inflammatory and antioxidant activities which consequently inhibit skin cancer (Osakabe et al. 2004; Roland et al. 2010; Baliga et al. 2013; Radhakrishnan et al. 2014). Gingerol present in Zingiber officinale induces caspase-dependent apoptosis in colon cancer cells by targeting the Erk1/2/JNK/AP-1 signalling (Fridlender et al. 2015). Tetrahydrocannabinol isolated from Cannabis sativa has been used in the past two centuries as supporting drugs for patients that receive either radiation or chemotherapies. Side effects related to these treatments such as vomiting, cachexia, nausea and loss of appetite are eased by cannabinoids (Robson 2001; Tramer et al. 2001; Ligresti et al. 2003; Massa et al. 2005; Grotenhermen and Muller-Vahl 2012). Studies imply that in the gastrointestinal system cannabinoid receptors are involved in inhibition of cell proliferation of colorectal carcinoma (Massa and Monory 2006; Varoni et al. 2016). Multiple mechanisms are performed by resveratrol in order to arrest carcinogenesis (Fig. 4.1). Paclitaxel ( $C_{47}H_{51}NO_{14}$ ) is known as taxol, probably the most well-known anticancer drug derived from the bark of Taxus brevifolia Nutt. It inhibits the microtubule disassembly by binding the polymerized microtubules (Xiao et al. 2012; Prota et al. 2013). Taxol binds to the microtubule-associated protein (MAP) microtubule complex causing further stabilization of microtubules thereby preventing mitotic spindle formation and thus inhibits mitosis as well as cell proliferation (Priyadarshini and Aparajitha 2012; Weaver 2014). Induction of multipolar divisions leads to formation of abnormal spindles bearing additional poles, and the consequence is unnatural chromosomal segregation which leads to the formation of abnormal aneuploid daughter cells that follow the apoptosis pathway (Priyadarshini and Aparajitha 2012).

#### 4.4 Purification of Anticancer Phytochemicals

The curative efficacy of medicinal plants is determined by the quality and quantity of bioactive molecule(s) which varies with altitude, latitude, climactic conditions and seasons. Varieties of chemopreventive bioactive molecules are distributed across different parts of a plant accounting for varying levels of pharmacological activity. Development of the phytochemicals as antitumor entities becomes a daunting task owing to the synergistic effects of such bioactive phyto-constituents rather than the purified one. Purification of bioactive phytochemicals includes isolation and assay, combinatorial chemistry and bioassay-guided fractionation. Prior to fractionation of the crude plant extract, the bioactivity of the extract is confirmed by subjecting it to bioassays. Various analytical platforms are used for examination of the eluted fractions, viz. FT-IR, mass spectroscopy, HPLC and thin-layer chromatography (Fig. 4.2). Solvents should be used in an increasing polarity order of silica, Sephadex, Superdex or any other suitable matrix that can be used for fractionation. Purification is followed by in vivo examination of extracts for evaluation of anticancer activity. The killing activity of tumour and other parameters like pharmacokinetics, safety and adverse effects, dose concentration, drug interactions, etc. must be explored before the development of novel anticancer drugs. However, the major bottleneck for rapid manufacturing of medicines using natural products is the poor solubility and bioavailability of plant secondary metabolites (Guo et al. 2006). In order to meet market demands, the development and use of synthetic or semisynthetic analogs to plant-derived substances are adopted. Morphine is a wellknown example that has been modified to morphine-6-glucuronide in order to enhance its therapeutic efficacy (Parc et al. 2002). Taxol, an important plant-derived (Taxus sp.) anticancer drug, is present in low amounts in all Taxus species along with its insolubility in water. These limitations in taxol manufacturing are overcome by combining the use of 10-deacetylbaccatin III and a semi-synthetic process for production of the drug. Docetaxel is the semi-synthetic soluble analog of taxol which is widely used, and additional strategies for improvement are needed to enhance its features and also to meet future market demands of this important drug (Aggarwal et al. 2003; Sharma et al. 2005; Malik et al. 2011; Fridlender et al. 2015). Curcumin analogs have been prepared owing to its insolubility in water. Besides due to reduced absorption in the liver and in intestinal walls and systemic

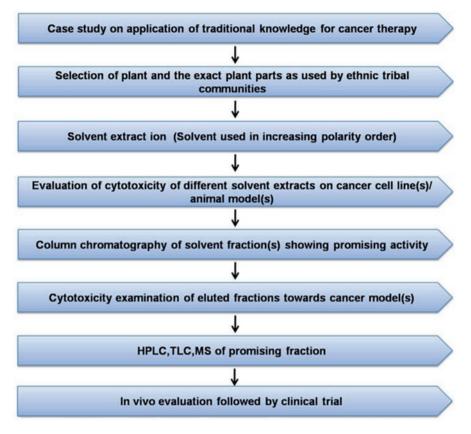


Fig. 4.2 Development of novel anticancer drugs derived from bioactive phytochemical

elimination, curcumin has poor bioavailability (Gordaliza 2007). However, the modified analog (beta-diketone monocarbonyl dienone) of curcumin has a good bioavailability and therapeutic effect as demonstrated in rodents (Twentyman et al. 1987; Mosley et al. 2007).

#### 4.5 In Vitro Propagation of Anticancer Plants

The anticancer plants are seems to be endangered. Therefore, there is a need to raise these plants for drugs and other therapeutic applications through various standardized protocols under in vitro conditions (Kingston 2000; Malik et al. 2011). The induction of callus and proliferation of gametophytes of *T. baccata*, found abundantly in north-eastern states of Nagaland and Arunachal Pradesh, was reported by past investigators (Rohr 1973; David and Plasitra 1974). However, David and Plastira (1976) studied the mineral and phytohormone composition of culture

		Sucrose			Root	
Explants	Medium	(%)	Auxin	Cytokinin	proliferation	Reference
Leaf	MS	3	IAA (1 mg/l)	BAP (1 mg/l)	Axillary	Wilken et al. (2011)
Leaf	MS	3	NAA (1 mg/l)	Kinetin (1 mg/l)	Adventitious	Klauber et al. (1997)

Table 4.2 Standardized in vitro shoot proliferation conditions of P. fulgens

medium to improve callus proliferation using mature stems as explants. Moreover, callus induction studies have been also performed using different explants, viz. hypocotyls, cotyledons, young or mature stems, and the roots from young seedlings, by past investigators (Brunakova et al. 2004, 2005). These studies concluded that the young tissues are more responsive and prone to callus induction than mature plants or adult trees (Brunakova et al. 2004). However, in vitro conditions for auxiliary and adventitious shoots of *P. fulgens*, an endangered anticancer herb of higher Himalayas, using leaf as explants (Wilken et al. 2011) were tabulated in Table 4.2.

#### 4.6 Conclusions and Future Prospects

The plant of North-East India or their derived compounds have the potential to be used as anticancer agents. The use of single drug to treat a single disease is questionable, and is a matter of debate since several decades. The recent trends in genomics, which is concerned with the genetic diversity or polymorphisms, clearly indicated that different human population need different drugs to cure the diseases. In this regard, the use of herbal medicines is gaining popularity because of its cost-effectiveness and potentials in the conventional medicinal practices. However, lacks of consistency in terms of their composition, efficacy, quality, safety, consistent manufacturing practices, regulations and approval processes lead the idea to combine the traditional and modern medicine practices. Therefore, the exploration of the plants found in North-East India is highly desired for the search of novel bioactive anticancer compounds and their mechanisms of action involved in the treatment of various types of malignant cells.

#### References

- Abdullah S, Abidin SAZ (2010) Ginger extract (*Zingiber officinale*) triggers apoptosis and G0/ G1 cells arrest in HCT 116 and HT 29 colon cancer cell lines. Afr J Biochem Res 4:134–142
- ACS (2016) Cancer facts and figures 2016 special section: cancer in Asian Americans, Native Hawaiians, and Pacific Islanders. American Cancer Society, Atlanta, pp 35–42
- Adebayo AH, Tan NH, Akindahunsi AA, Zeng GZ, Zhang YM (2010) Anticancer and antiradical scavenging activity of *Ageratum conyzoides* L. (Asteraceae). Pharmacogn Mag 6:62–66

- Aggarwal BB, Shishodia S (2005) Curcumin suppresses the Paclitaxel induced nuclear factor-KB pathway in breast cancer cells and inhibits lung metastasis of human breast. Clin Cancer Res 11:7490–7498
- Aggarwal BB, Kumar A, Bharti AC (2003) Anticancer potential of curcumin: preclinical and clinical studies. Anticancer Res 23:363–398
- Baliga MS, Jimmy R, Thilakchand KR, Sunitha V, Bhat NR, Saldanha E, Rao S, Rao P, Arora R, Palatty PL (2013) *Ocimum sanctum* L. (Holy Basil or Tulsi) and its phytochemicals in the prevention and treatment of cancer. Nutr Cancer 65:S26–S35
- Baral R, Chattopadhyay U (2004) Neem (*Azadirachta indica*) leaf mediated immune activation causes prophylactic growth inhibition of murine ehrlich carcinoma and B16 melanoma. Int Immunopharmacol 4:355–366
- Baselga J, Norton L, Albanell J, Kim YM, Mendelsohn J (1998) Recombinant humanized anti-HER2 antibody enhances the antitumor activity of Paclitaxel and Doxorubicin against HER2/ neu overexpressing human breast cancer xenografts. Cancer Res 58:2825–2831
- Behere PB, Das A, Yadav R, Behere AP (2013) Ayurvedic concepts related to psychotherapy. Indian J Psychiatry 55:S310–S314
- Brunakova K, Babincova Z, Takac M, Cellarova E (2004) Selection of callus cultures of *Taxus baccata* L. as a potential source of Paclitaxel production. Eng Life Sci 4:465–469
- Brunakova K, Babincova Z, Cellarova E (2005) Production of taxanes in callus and suspension cultures of *Taxus baccata* L. In: Hvoslef-Eide AK, Preil W (eds) Liquid culture systems for in vitro plant propagation. Springer, The Netherlands, pp 567–574
- Casanova ML, Blazquez C, Martinez-Palacio J, Villanueva C, Fernandez-Acenero MJ, Huffman JW, Jorcano JL, Manuel Guzman M (2003) Inhibition of skin tumor growth and angiogenesis in vivo by activation of cannabinoid receptors. J Clin Invest 111:43–50
- Cheung S, Tai J (2007) Anti-proliferative and antioxidant properties of rosemary Rosmarinus officinalis. Oncol Rep 17:1525–1531
- Cui SX, Qu XJ, Xie YY, Zhou L, Nakata M, Makuuchi M, Tang W (2006) Curcumin inhibits telomerase activity in human cancer cell lines. Int J Mol Med 18:227–231
- David A, Plasitra V (1974) Histophysiologie vegetale realisation de cultures de tissues d'If (*Taxus baccata* L.) a partir de fragments de pousses ages d'un an; obtention d'une culture indefinie. CR Acad Sci Paris 279:1757–1759
- David A, Plastira V (1976) Histophysiologie vegetale. Comportement en culture in vitro de cellules isolees de deux gymnospermes: *Taxus baccata* L. et Pinus pinaster Sol. CR Acad Sci Paris 110:5917–5919
- De Petrocellis L, Melck D, Palmisano A, Bisogno T, Laezza C, Bifulco M, Di Marzo V (1998) The endogenous cannabinoid an and amide inhibits human breast cancer cell proliferation. Proc Natl Acad Sci USA 95:8375–8380
- Fridlender M, Kapulnik Y, Koltai H (2015) Plant derived substances with anticancer activity: from folklore to practice. Front Plant Sci 6:799
- Gandhi D, Mehta P (2013) *Dillenia indica* Linn. and *Dillenia pentagyna* Roxb.: pharmacognostic, phytochemical and therapeutic aspects. J Appl Pharma Sci 3:134–142
- Giri A, Lakshmi Narasu M (2000) Production of podophyllotoxin from *Podophyllum hexandrum*: a potential natural product for clinically useful anticancer drugs. Cytotechnology 34:17–26
- Gogoi J, Nakhuru KS (2016) Isolation and characterization of bioactive components from *Mirabilis* jalapa L. radix. J Tradit Complement Med 6:41–47
- Gordaliza M (2007) Natural products as leads to anticancer drugs. Clin Transl Oncol 9:767–776
- Grotenhermen F, Muller-Vahl K (2012) The therapeutic potential of cannabis and cannabinoids. Dtsch Arztebl Int 109:495–501
- Guo BH, Kai GY, Jin HB, Tang KX (2006) Taxol synthesis. Afr J Biotechnol 5:15-20
- Hashimoto T, Ashida H, Sano T, Furuyashiki T, Shiotani B, Kanazawa K, Danno G (2000) 3-Amino-1,4-dimethyl-5H-pyrido[4,3-b]indole(Trp-P-1) induces apoptosis in rat splenocytes and thymocytes by different mechanisms. Mutat Res 457:57–67

- Hossan MS, Rahman S, Bashar ABMA, Jahan R, Al-Nahain A, Rahmatullah M (2014) Rosmarinic acid: a review of its anticancer action. World J Pharm Pharm Sci 3:57–70
- Hsu CP, Lin CC, Huang CC, Lin YH, Chou JC, Tsia YT, Su JR, Chung YC (2012) Induction of apoptosis and cell cycle arrest in human colorectal carcinoma by litchi seed extract. J Biomed Biotechnol 2012:341479. https://doi.org/10.1155/2012/341479
- Hussain A, Sharma C, Khan S, Shah K, Haque S (2015) Aloe vera inhibits proliferation of human breast and cervical cancer cells and acts synergistically with cisplatin. Asian Pac J Cancer Prev 16:2939–2946
- Kim JS (2008) Cytotoxic components from the dried rhizomes of *Zingiber officinale* Roscoe. Arch Pharm Res 31:415
- Kingston DG (2000) Recent advances in the chemistry of taxol. J Nat Prod 63:726-734
- Klauber N, Parangi S, Flynn E, Hamel E, D'Amato RJ (1997) Inhibition of angiogenesis and breast cancer in mice by the microtubule inhibitors 2-methoxyestradiol and taxol. Cancer Res 57:81–86
- Kumar S, Suresh PK, Vijayababu MR, Arunkumar A, Arunakaran J (2006) Anticancer effects of ethanolic neem leaf extract on prostate cancer cell line (PC-3). J Ethnopharmacol 105:246–250
- Lee J, Kim JH (2016) Kaempferol inhibits pancreatic cancer cell growth and migration through the blockade of EGFR-related pathway in vitro. PLoS One 11:e0155264
- Ligresti A, Bisogno T, Matias I, De Petrocellis L, Cascio MG, Cosenza V, D'argenio G, Scaglione G, Bifulco M, Sorrentini I, Di Marzo V (2003) Possible endocannabinoid control of colorectal cancer growth. Gastroenterology 125:677–687
- Ligresti A, Moriello AS, Starowicz K, Matias I, Pisanti S, De Petrocellis L, Laezza C, Portella G, Bifulco M, Di Marzo V (2006) Antitumor activity of plant cannabinoids with emphasis on the effect of cannabidiol on human breast carcinoma. J Pharma Exp Therap 318:1375–1387
- Lipinsky CA, Lombardo F, Dominy BW, Feeney PJ (1997) Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. Adv Drug Deliv Rev 46:3–26
- Magesh V, Lee JC, Ahn KS, Lee HJ, Lee HJ, Lee EO, Shim BS, Jung HJ, Kim JS, Kim DK, Choi SH, Ahn KS, Kim SH (2009) Ocimum sanctum induces apoptosis in A549 lung cancer cells and suppresses the in vivo growth of lewis lung carcinoma cells. Phytother Res 23:1385–1391
- Malik S, Cusido RM, Mirjalili MM, Moyano E, Palazon J, Bonfill M (2011) Production of the anticancer drug taxol in *Taxus baccata* suspension cultures: a review. Process Biochem 46:23–34
- Massa F, Monory K (2006) Endocannabinoids and the gastrointestinal tract. J Endocrinol Investig 29:47–57
- Massa F, Storr M, Lutz B (2005) The endocannabinoid system in the physiology and pathophysiology of the gastrointestinal tract. J Mol Med 83:944–954
- Massi P, Vaccani A, Ceruti S, Colombo A, Abbracchio MP, Parolaro D (2004) Antitumor effects of cannabidiol, a nonpsychoactive cannabinoid, on human glioma cell lines. J Pharma Exp Therap 3:838–845
- Mbaveng AT, Kuete V, Mapunya BM, Beng VP, Nkengfack AE, Meyer JJM, Lall N (2011) Evaluation of four Cameroonian medicinal plants for anticancer, antigonorrheal and antireverse transcriptase activities. Environ Toxicol Pharmacol 32:162–167
- Mittal AK, Tripathy D (2015) Biosynthesis of silver nanoparticles using *Potentilla fulgens* ex wall. hook and its therapeutic evaluation as anticancer and antimicrobial agent. Mater Sci Eng 53:120–127
- Mosley CA, Liotta DC, Snyder JP (2007) Highly active anticancer curcumin analogues. Adv Exp Med Biol 595:77–103
- Noorolahi SM, Sadeghi S, Mohammadi M, Azadi M, Rahimi NA, Vahabi F, Arjmand M, Hosseini H, Mosallatpur S, Zamani Z (2016) Metabolomic profiling of cancer cells to *Aloe vera* extract by <sup>1</sup>HNMR spectroscopy. J Metabol 2:1–7. https://doi.org/10.7243/2059-0008-2-1
- Norikura T, Kojima-Yuasa A, Shimizu M, Huang X, Xu S, Kametani S, Rho SN, Kennedy DO, Matsui-Yuasa I (2008) Mechanism of growth inhibitory effect of *Blumea balsamifera* extract in *Hepatocellular carcinoma*. Biosci Biotechnol Biochem 72:1183–1189

- Osakabe N, Yasuda A, Natsume M, Yoshikawa T (2004) Rosmarinic acid inhibits epidermal inflammatory responses: anti-carcinogenic effect of *Perilla frutescens* extract in the murine two-stage skin model. Carcinogenesis 25:549–557
- Parc G, Canaguier A, Landre P, Hocquemiller H, Chriqui D, Meyera M (2002) Production of taxoids with biological activity by plants and callus culture from selected *Taxus* genotypes. Phytochemistry 59:725–730
- Priyadarshini K, Aparajitha UK (2012) Paclitaxel against cancer : a short review. Med Chem 2:139–141
- Prota AE, Bargsten K, Zurwerra D, Field JJ, Díaz JF, Altmann KH, Steinmetz MO (2013) Molecular mechanism of action of microtubule-stabilizing anticancer agents. Science 339:587–590
- Radhakrishnan E, Bava SV, Narayanan SS, Nath LR, Thulasidasan AKT, Soniya EV, Anto RJ (2014) [6]-Gingerol induces Caspase-dependent apoptosis and prevents PMA-induced proliferation in colon cancer cells by inhibiting MAPK/AP-1 signalling. PLoS One 9:e104401
- Robson P (2001) Therapeutics aspects of cannabis and cannabinoids. Br J Psychiatry 178:107-115
- Rohr R (1973) Cytologie vegetale. Ultrastructure des spermatozoides de *Taxus baccata* L. obtenus a partir de cultures aseptiques de microspores sur un milieu artificiel. CR Acad Sci Paris 277:1869–1871
- Roland CL, Dineen SP, Toombs JE, Carbon JG, Smith CW, Brekken RA, Barnett CC Jr (2010) Tumour-derived intercellular adhesion molecule-1 mediates tumour-associated leukocyte infiltration in orthotopic pancreatic xenografts. Exp Biol Med 235:263–270
- Rosangkima G, Prasad SB (2007) Changes in endogenous glutathione level associated with the antitumor activity of the stem bark extract of *Dillenia pentagyna* against Murine Ascites Dalton's. Pharmacology 2:11–19
- Rosangkima G, Rongpi T, Prasad SB (2010) Ethno-medicinal value of some anticancer medicinal plants from north-east India. Sci Vis 10:123–132
- Roy MK, Kobori M, Takenaka M (2007) Antiproliferative effect on human cancer cell lines after treatment with nimbolide extracted from an edible part of the neem tree (*Azadirachta indica*) Phytother Res 21:245–250
- Roya S, Bersa SE, De T, Banerjee B, Mukherjee J, Vedasiromoni JR (2008) Induction of apoptosis in human leukemic cell lines U937, K562 and HL-60 by *Litchi chinensis* leaf extract via activation of mitochondria mediated caspase cascades. Open Leuk J 1:1–14. https://doi. org/10.2174/1876816400901010001
- Rumzhum NN, Rahman MM, Islam MS, Chowdhury SA, Sultana R (2008) Cytotoxicity and antioxidant activity of extractives from *Mirabilis jalapa*. St J Pharm Sci 1:85–88
- Sharma RA, Gescher AJ, Steward WP (2005) Curcumin: the story so far. Eur J Cancer 41:1955-1968
- Sharma B, Singh S, Kanwar SS (2014) L-methionase: a therapeutic enzyme to treat malignancies. Bio Med Res Int 2014:13. https://doi.org/10.1155/2014/506287
- Siegel RL, Miller KD, Jemal A (2016) Cancer statistics, 2016. CA Cancer J Clin 66:7-30
- Syiem D, Kharbuli B (1999) Medicinal plants and herbal medicine: a case study in Meghalaya. In: Kharbuli B, Syiem D, Kayang H (eds), Biodiversity. North-East India Perspectives Shillong, India, pp 1–8
- Tai J, Cheunga S (2012) Antiproliferation effect of Rosemary (*Rosmarinus officinalis*) on human ovarian cancer cells in vitro. Phytomedicine 19:436–443
- Thun MJ, De Lancey JO (2010) The global burden of cancer: priorities for prevention. Carcinogenesis 31:100–110
- Tramer MR, Carroll D, Campbell FA, Reynolds DJM, Moore RA, Mcquay HJ (2001) Cannabinoids for control of chemotherapy induced nausea and vomiting quantitative systematic review. BMJ 323:1–8
- Trevor P, Theodore A (1993) Determination of Taxol in *Taxus* media needles in the presence of interfering components. J Liq Chromatogr 16:723–731
- Tushara BS, Sharma SC, Rangan L (2010) Ethnomedical uses of Zingiberaceous plants of Northeast. India J Ethnopharmacol 132:286–296

- Twentyman PR, Fox NE, White DJ (1987) Cyclosporin A and its analogues as modifiers of adriamycin and vincristine resistance in a multi-drug resistant human lung cancer cell line. Br J Cancer 56:55–57
- Uden WN (1989) Detection and identification of *Podophyllotoxin* produced by cell cultures derived from *Podophyllum hexandrum* royle. Plant Cell Rep 8:165–168
- Vaishnav K, George LB, Highland HN (2015) Antitumor activity of Xanthium strumarium L. on human cervical cancer HeLa cells. J Cancer Tum Int 2:1–13
- Varoni EM, Lo Faro AF, Sharifi-Rad J, Iriti M (2016) Anticancer molecular mechanisms of resveratrol. Front Nutr 3:8. https://doi.org/10.3389/fnut.2016.00008
- Weaver BA (2014) How taxol/paclitaxel kills cancer cells. Mol Biol Cell 25:2677-2268
- Wilken R, Veena MS, Wang MB, Srivatsan ES (2011) Curcumin: a review of anti-cancer properties and therapeutic activity in head and neck squamous cell carcinoma. Mol Cancer 10:12. https:// doi.org/10.1186/1476-4598-10-12
- Xiao H, Wang H, Zhang X, Tu Z, Bulinski C, Khrapunovich-Baine M, Angeletti RH, Horwitz SB (2012) Structural evidence for cooperative microtubule stabilization by Taxol and the endogenous dynamics regulator MAP 4. ACS Chem Biol 7:744–752
- Yesil-Celiktas O, Sevimli C, Bedir E, Vardar-Sukan F (2010) Inhibitory effects of Rosemary extracts, carnosic acid and rosmarinic acid on the growth of various human cancer cell lines plant foods. Human Nutr 65:158–163
- Zhao M, Yang B, Wang J, Liu Y, Yu L, Jiang Y (2007) Immunomodulatory and anticancer activities of flavonoids extracted from litchi (*Litchi chinensis Sonn.*) pericarp. Int Immunopharmacol 7:162–166