

# Health Assessment of Loktak Lake Using Diatoms as Biological Indicators

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## 1 Introduction

Wetlands are precious freshwater resources. They are fragile ecosystems. A small change in the abiotic factors or composition of biotic can render them susceptible to damage. The large freshwater Loktak Lake is one such wetland, characterized by phumdis, the floating islands, which are the unique habitat of the endangered mammal, the brow-antlered deer popularly known as Sangai. Phumdis occur in sizes ranging from a few centimeters to about 2.5 m. This wetland is a Ramsar Site since 1990. The Keibul Lamjao National Park occupying an area of 40 sq km offers protection to the Sangai. The lake is shrinking due to soil erosion resulting from deforestation and shifting cultivation in the catchment [1]. The present study therefore proposes to determine the water quality vis-à-vis health of the Loktak Lake ecosystem. This will be useful in restoration efforts in order to maintain the benefits provided by this wetland. The results of this study will also provide a baseline data on the diatom flora and community, useful for comparing ecology of the lake in future. Diatom communities respond quickly and predictably to changes in water quality and are therefore employed as bio-indicators in aquatic monitoring programs in Europe, the USA, Canada, South America, and Australia [2–11].

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## 2 Methodology

**Study Area:** Diatom samples were collected from the Loktak Lake at 24°30'37" N to 93°47'08" E and elevation of 772 masl. Substrate comprising small boulders, pebbles and cobbles, free-floating water hyacinth, partly decomposed roots and rhizomes, small twigs and grasses were sampled to record all possible species in the ecosystem. Epilithic diatoms were obtained from hard substrate by scraping 3 × 3 cm surfaces with the help of sharp razor and brush. The epiphytic diatoms were obtained by collecting sizeable portions of root, stem and leaves. Two replicates were obtained for each substrate and mixed to form one sample. The sample was preserved in 4% formalin. Diatoms were cleaned with the acid and H<sub>2</sub>O<sub>2</sub>. Permanent mount was prepared in Naphrax. Diatom species were identified at ×1500 under bright field using NIKON 80i Trinocular Research Microscope and documented with DS-5M-LI digital camera. Diatom flora was recorded by identifying species, varieties, and forms using standard literature [12–17]. Species counts of 500 valves were made to determine relative abundance, species diversity, and evenness using Species Diversity and Richness software. The Van Dam, Hoffmann, Lange-Bertalot ecological values, and Leclercq Index were computed by OMNIDIA software.

## 3 Results and Discussion

The Loktak Lake is known as the lifeline of Manipur, owing to its socioeconomic and cultural values. A large number of fishermen depend directly on the lake resources for sustenance. Lake water is also used for generation of hydroelectric power by NHPC. A recent Union Planning Commission study stated that the habitat of the brow-antlered deer (*Cervus eldi*, locally called Sangai) has shrunk from 40 sq kms to only 6 sq kms today due to the Ithai Barrage commissioned in 1983. The bio-assessment of the water quality through diatom community to indicate the recent health of the Loktak Lake ecosystem thus becomes very important.

**Diatom flora:** The flora consisted of 115 species from 2 centric and 39 pennate genera represented by 2 and 113 taxa, respectively (Appendix 1). Relatively more species of centric diatoms are known from lakes of Jammu & Kashmir [18, 19] and lentic waters from Gujarat [20]. Among the pennates, biraphid taxa constitute the bulk of the flora (86 taxa; 77%) while the araphid, monoraphid, and raphid species are few (26 taxa; 23%) as also evident from the other studies in India [13, 20–22] and also from outside of the Indian subcontinent [23–26]. The biraphid *Cymbella* and *Navicula* are the species-rich genera in the flora as observed not only in the above-said mountain waters but also from various parts of India [22, 27–29]. *Anomoeoneis styriaca* (Grun) Hustedt and *Brachysira vitrea* Ross are notable species in the flora that occur in Central Highland but not in the Himalayan waters [30].

**Diatom community:** The Loktak Lake shows remarkable variations in diatom community. In any community, usually some taxa attain >10% abundance but in this study no taxa attained 10% abundance attributed to high evenness in the community. Highest abundance was recorded for *Gomphonema parvulum* Kütz. (8.0%) which is an indicator of organically enriched waters and eutrophic state, as is good count of *Nitzschia palea* (Kütz) W. Smith also [31]. Other taxa figuring >5% category are *Gomphonema neonasutum* L-B & Reichardt, *Brachysira vitrea* (Grunow) Ross, and *Cocconeis placentula* Ehrenberg which are alkaliphilous and sensitive to very sensitive for pollution [31]. *B. vitrea* has rarely been recorded to gain more than 1% abundance in the natural waters and thus is notable for >5% abundance in this wetland. *Amphora veneta* Kütz and *Navicula cryptotenella* Lange-Bertalot are also indicators of the eutrophic state. The diatom taxa *Ulnaria ulna* (Nitzsch.) Compère indicate the presence of zinc and mercury hydragyrum in water. Also, diatoms such as *Navicula capitatoradiata* Germain, *Navicula cryptocephala* Kütz., *Navicula cryptotenella*, and *Nitzschia gracillis* Hantzsch are moderately tolerant forms [31]. The community appears to be highly diverse ( $H = 5.63$ ) and even ( $E = 0.82$ ) which could happen when a wide variety of nutrients are present in limiting amounts.

**Ecological values:** The examination of ecologic values shows considerable variations in the pH conditions. While majority of the community is represented by alkaliphilous and circumneutral forms, the alkalibiontic, acidobiontic, and acidophilous forms are notable because the last two categories have not been reported even from the organically enriched waters, e.g., Khanda Gad [32]. With respect to salinity conditions, 62.6% forms in the community belong to fresh-brackish state, a condition marked by presence of <500 mg/l chloride and salinity by <0.9%. Only 6.2% are freshwater diatoms (<100 mg/l chloride and <0.2% salinity). The presence of higher ecologic values even though meagerly indicates vicious conditions due to increasing salinity in the lake; 5.6% fresh-brackish diatoms (representing 500–1000 mg/l chloride and 0.9–1.8% salinity) and 3.3% brackish water diatom forms (1000–5000 mg/l chloride with 1.8–9.0% salinity).

For  $N_2$  uptake metabolism, the lake was dominated by 28.8%  $N_2$  autotrophic diatom taxa that tolerate elevated concentration of organically bound  $N_2$ . But then all other forms are also present in low numbers which support the above view that the ecosystem has degraded from the natural or semi-natural conditions. This statement receives support from fairly even distribution of diatom community in  $O_2$  requirement categories (Table 1).

Like the above parameters, the saprobity values show presence of forms that prefer higher BOD and low  $O_2$  saturation; 28.6%  $\beta$ -mesosaprobous (2–4 mg/l BOD and 70–85%  $O_2$  saturation), 13.6% oligosaprobous (<2 mg/l BOD and >85%  $O_2$  saturated), 11.9%  $\alpha$ -mesosaprobous, and 10.6%  $\alpha$ -mesopolysaprobous representing 4–13 and 13–22 mg/l BOD and 25–70% and 10–25%  $O_2$  saturation, respectively, and 3.0% polysaprobous diatoms (>22 mg/l BOD and <10%  $O_2$  saturation). All these categories are reflected in Hoffmann and Lange-Bertalot values. The trophic

**Table 1** Ecological characteristics of Loktak Lake based on analysis using OMNIDIA software

N° PREP	Date	Basin		Imphal						
1	02/28/2008	River/Site		Loktak/KLNIP						
<b>Van Dam 1994</b>										
	1	2	3	4	5	6	7	Dominant		
pH	0.3	1.4	26.5	46.1	6.0	0		4	Alkaliphilous	
Salinity	6.2	62.6	5.6	3.3				2	Fresh brackish	
N <sub>2</sub> uptake	21.4	28.8	9.4	3.5				2	N <sub>2</sub> autotrophic taxa tolerating elevated concentrations of organically bound nitrogen	
O <sub>2</sub> requirements	14.9	24.0	14.1	11.3	1.4			2	Fairly high	
Saprobity	13.6	28.5	11.9	10.6	3.0			2	β-mésosaprobic	
Trophic state	3.0	12.3	3.1	6.8	39.1	3.1	3.9	5	Eutraphentic	
Moisture	5.0	31.2	27.8	2.6	0			2	Mainly occurring in water sometimes on wet places	
<b>Lange-Bertalot 1979</b>										
	11.8	6.6	8.0	0.3	8.0	3.7		1	Most pollution tolerant	
<b>Hofmann 1994</b>										
	0	1	2	3	4	5	6	7	8	9
Trophic state	34.3	1.6	9.2	0.3	5.7	12.1	32.3	0.9	3.5	6
Saprobity	35.0	7.2	9.1	22.8	1.6	6.5	0	6.0	0.11	3
									11.7	Mesosaprobe

status clearly indicates high share of eutrathentic forms (39.1% nearly double of natural waters). There is a reasonable presence of oligo-mesotrathentic forms. Other categories though low in share were consistent in presence; oligo-eutrathentic, mesotrathentic, and hyper-eutrathentic and oligotrathentic as observed in Hoffmann values. Moisture preferences showed that 31.2% taxa were mainly occurring in water bodies, sometimes on wet place, 27.8% taxa were occurring in water bodies, also rather regularly on wet and moist places, 5.0% taxa were never or very rarely occurring outside water bodies, and 2.6% taxa were occurring on wet and moist or temporarily dry places. Leclercq Index shows very high degradation in the lake. Probably the barrage prevents flushing of nutrients accumulated from decaying phumdis causing perturbations in the bio-geochemical cycles. There were 16.5% diatom taxa that indicated organic pollution while 15.33% indicated anthropogenic eutrophication, indicating low levels as indicated by green color (Fig. 1).

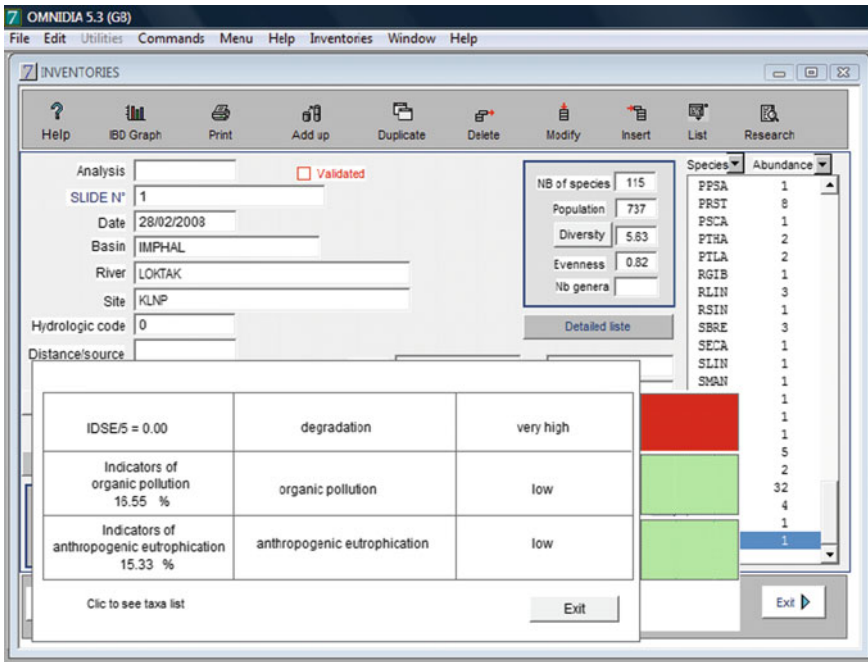


Fig. 1 OMNIDIA software showing levels of degradation, organic pollution, and anthropogenic eutrophication in Loktak Lake

## 4 Conclusion

The freshwater Loktak Lake ecosystem exhibits perturbations in the nutrient regimes and hence the water chemistry. This is reflected in the flora (presence of salinity loving forms *A. styriaca* and *B. vitrea*) and the community (higher abundance of *G. parvulum* that prefers organically enriched waters). The Van Dam ecologic values also support this observation, as evident by the higher share of nitrogen autotrophic diatom taxa that tolerate elevated concentration of organically bound N<sub>2</sub>, forms that prefer higher BOD, low O<sub>2</sub> and eutraphentic conditions. The Leclercq index shows low organic pollution, anthropogenic eutrophication and very high degradation. Lake restoration is the only solution to current state of the Loktak Lake ecosystem.

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## Appendix 1

Diatom flora of Loktak Lake

Pennales	C
<i>Diatoma vulgare</i> Bory morpho <i>producta</i> Gr. in VH	4
<i>Fragilaria rumpens</i> (Kützing) G.W.F. Carlson	26
<i>Tabellaria fenestrata</i> (Lyngbye) Kützing	5
<i>Ulnaria ulna</i> (Nitzsch) Compère	1
<i>U. u.</i> var. <i>aequalis</i> (Kützing) Aboal	4
<i>U. u.</i> var. <i>acus</i> (Kützing) Abol	32
<i>U. amphirhynchus</i> (Ehrenberg) Compere et Bukhtiyarova	1
<i>U. danica</i> (Kützing) Compere et Bukhtiyarova	2
<i>Achnanthes brevipes</i> Agardh	1
<i>Achnantheidium catenatum</i> (Bily& Mar.) Lange-Bertalot	14
<i>A. exigum</i> (Grunow) var. <i>heterovalvum</i> (Krasske) Czarnecki	2
<i>Achnanthes microcephala</i> (Kützing) Grunow	1
<i>A. minutissimum</i> (Kützing) Czarnecki	1
<i>Cocconeis euglypta</i> Ehrenberg	31
<i>C. pediculus</i> Ehrenberg	7
<i>C. placentula</i> Ehrenberg	40
<i>Lemnicola hungarica</i> (Gr) Round & Basson	1
<i>Planothidium hauckianum</i> (Gr) Round & Bukhtiyarova	2
<i>P.lanceolatum</i> (Brebisson ex Kützing) Lange-Bertalot	2
<i>P. rostratum</i> (Öestrup) Round & Bukhtiyarova	8

(continued)

(continued)

<b>Pennales</b>	<b>C</b>
<i>Rossithidium linearis</i> (W. Smith) Round & Bukhtiyarova	3
<i>Actinella guianensoides</i> Metzeltin & Lange-Bertalot	1
<i>Eunotia alpina</i> (Naegeli) Hustedt	1
<i>E. flexuosa</i> (Brebisson) Kützing	1
<i>E. pectinalis</i> (Dyllumyn) Rabenhorst	1
<i>E. monodon</i> Ehrenberg	4
<i>E. m. var. bidens</i> (Gregory) Hustedt	1
<i>Amphora libyca</i> Ehrenberg	1
<i>A. montana</i> Krasske	2
<i>A. pediculus</i> (Kützing) Grunow	3
<i>A. veneta</i> Kützing	2
<i>Amphipleura pellucida</i> Kützing	1
<i>Anomooneis styriaca</i> (Grunow) Hustedt	1
<i>Brachysira vitrea</i> (Grunow) Ross in Hartley	40
<i>Caloneis bacillum</i> (Grunow) Cleve	1
<i>Craticula accomoda</i> (Hustedt) D.G. Mann	2
<i>Craticula cuspidata</i> (Kützing) D.G. Mann	1
<i>Cymbella aspera</i> (Ehrenberg) H. Peragallo	1
<i>C. austriaca</i> Grunow	1
<i>C. excisa</i> Kützing	10
<i>C. gracilis</i> (Ehrenberg) Kützing	1
<i>C. hantzschiana</i> Krammer	5
<i>C. hustedtii</i> Krasske	13
<i>C. h. var. crassipunctata</i> Lange-Bertalot & Krammer	7
<i>C. mesiana</i> Cholnoky	7
<i>C. orientalis</i> Lee in Lee Gotoh & Chung	1
<i>C. perparva</i> Krammer	1
<i>C. parva</i> (W. Smith) Kirchner in Cohn	1
<i>C. pervarians</i> Krammer	1
<i>C. subleptoceros</i> Krammer	11
<i>C. subhelvetica</i> Krammer	4
<i>C. stigmaphora</i> Østrup	5
<i>C. tumida</i> (Brebisson) Van Heurck	6
<i>C. turgidula</i> Grunow	1
<i>C. t. var. venezolana</i> Krammer	1
<i>Diadesmis confervacea</i> Kützing	1
<i>Diploneis elliptica</i> (Kützing) Cleve	1
<i>D. modica</i> Hustedt	1
<i>Encyonema minutum</i> (Hilse in Rabenhorst) D.G. Mann	27
<i>Epithemia sorex</i> Kützing	3

(continued)

(continued)

<b>Pennales</b>	<b>C</b>
<i>E. zebra</i> (Ehrenberg) Kützing	10
<i>Frustulia vulgaris</i> (Thwaite) De Toni	2
<i>Gomphonema affine</i> Kützing	4
<i>G. acuminatum</i> Ehrenberg	18
<i>G. neonasutum</i> Lange-Bertalot & Reichardt	51
<i>G. angustum</i> Agardh	1
<i>G. augur</i> Ehrenberg	6
<i>G. clevei</i> Fricke	7
<i>G. gracile</i> Ehrenberg	2
<i>G. olivaceum</i> (Hornemann) Brébisson	1
<i>G. parvulum</i> Kützing	59
<i>G. subtile</i> Ehrenberg	1
<i>G. truncatum</i> Ehrenberg	1
<i>Gyrosigma obtusatum</i> (Sullivan & Wormley) Boyer	29
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	5
<i>Luticola goeppertiana</i> (Bleish in Rbenhorst) D.G. Mann	2
<i>Navicula capitatoradiata</i> Germain	1
<i>N. cryptocephala</i> Kützing	10
<i>N. cryptotenella</i> Lange-Bertalot	9
<i>N. krammerae</i> Lange-Bertalot	1
<i>N. upsaliensis</i> (Grunow) Peragallo	1
<i>N. phyllepta</i> Kützing	20
<i>N. radiosa</i> Kützing	16
<i>N. schroeteri</i> Meister	16
<i>N. rostellata</i> Kützing	3
<i>N. veneta</i> Kützing	1
<i>N. v. var. viridula</i> (Kützing) Ehrenberg	4
<i>N. v. var. v. forma linearis</i> (Hustedt) Kobayasi	1
<i>Neidium affine</i> (Ehrenberg) Piftzer	1
<i>N. ampliatum</i> (Ehrenberg) Krammer	1
<i>N. binodeforme</i> Krammer	1
<i>Nitzschia clausii</i> Hantzsch	23
<i>N. dissipata</i> (Kützing) Grunow	1
<i>N. dravellensis</i> Coste & Ricard	1
<i>N. frustulum</i> (Kützing) Grunow	4
<i>N. gracillis</i> Hantzsch	2
<i>N. hantzschiana</i> Rabenhorst	12
<i>N. obtusa</i> var. <i>scalpelliformis</i> Grunow	1
<i>N. palea</i> (Kützing) W. Smith	20
<i>Placoneis elliptica</i> (Hustedt) Ohtsuka	1

(continued)



(continued)

<b>Pennales</b>	<b>C</b>
<i>Pinnularia acrospheria</i> W. Smith	1
<i>P. braunii</i> (Grunow) Cleve	1
<i>P. subcapitata</i> Gregory	1
<i>Pleurosigma angulatum</i> (Quekett) W. Smith	4
<i>Reimeria sinuata</i> (Gregory) Kociolek & Stoermer	1
<i>Rhopalodia gibba</i> (Ehrenberg) O. Müller	1
<i>Sellaphora mantasoana</i> Metzeltin et Lange-Bertalot	1
<i>S. parapupula</i> Lange-Bertalot	1
<i>S. pupula</i> (Kützing) Mereschkowsky	1
<i>Stauroneis anceps</i> Ehrenberg	1
<i>S. phoenicenteron</i> (Nitzsch) Ehrenberg	1
<i>urirella capronii</i> Brebisson in Kitton	3
<i>S. linearis</i> W. Smith	1
<b>Centrales</b>	
<i>Cyclotella meneghiniana</i> Kützing	8
<i>Discostella stelligera</i> (Cleve at Grunow) Houk & Klee	1

Acronym: C = Count

## References

1. LRIS: Catchment characterization of Loktak lake using remote sensing and GIS. Report published in Map Asia (2003)
2. Round, F.: Use of diatoms for monitoring rivers. In: Whitton, B., Rott, E., Friedrich, G. (eds.) *Use of Algae for Monitoring Rivers*, pp. 25–32. Proceedings of International Symposium, Institut für Botanik, Universität of Innsbruck, Innsbruck Austria (1991)
3. Whitton, B., Rott, E., Friedrich, G. (eds.): *Use of Algae for Monitoring Rivers*, 193 pp. Institut für Botanik, Universität of Innsbruck, Innsbruck Austria (1991)
4. Kelly, M.G., Whitton, B.A.: The Trophic diatom index: a new index for monitoring Eutrophication in Rivers. *J. Appl. Phycol.* 7, 433 (1995)
5. Whitton, B., Rott, E. (eds.): *Use of algae for monitoring rivers II*. In: Proceedings of the 2nd European Workshop, Innsbruck, 1995, 196 pp. Institut für Botanik, University of Innsbruck, Innsbruck (1996)
6. Lowe, R.L., Pan, Y.: Benthic algal communities and biological monitors. In: Stevenson, R.J., Bothwell, M., Lowe, R.L. (eds.) *Algal Ecology: Freshwater Benthic Ecosystems*, pp. 705–39. Academic Press, San Diego, California, USA (1996)
7. Lobo, E.A., Callegaro, V.L.M., Hermany, G., Go'mez, N., Ector, L.: Review of the use of microalgae in South America for monitoring rivers, with special reference to diatoms. *Vie Milieu-Life Environ.* 54, 105–114 (2004)
8. Poulíčková, A., Dokulil, M., Duchoslav, M.: Littoral diatom assemblages as bioindicators of lake trophic status: a case study from perialpine lakes in Austria. *European J. Phycol.* 39, 143–152 (2004)

9. Csilla S.K., Buczko K., Hajnal E., Judit, P.: Epiphytic, littoral diatoms as bioindicators of shallow lake trophic status: trophic Diatom Index for Lakes (TDIL) developed in Hungary. *Hydrobiologia*, **589**, 141–154 (2007)
10. Juttner I., Chimonides P.J., Ormerod S.J.: Using diatoms as quality indicators for a newly-formed urban lake and its catchment. *Environ. Monit. Assess.* **162**, 47–65 (2009)
11. Szczepocka, S., Szczepocka E., Szulc B.: The use of benthic diatoms in estimating water quality of variously polluted rivers. *Oceanol. Hydrobiol. Stud.* **38**(1), 17–26 (2009)
12. Hustedt F., Jensen N.G.: The pennate diatom (a translation of Hustedt's *Die kieslagen 2 Teil*' with supplement by N G Jensen). Koeltz Scientific Books, Koenigstein (1985)
13. Sarode, P.T., Kamat N.D.: *Freshwater diatoms of Maharashtra*, 338 pp. Siakripa Prakashan Aurangabad, India (1984)
14. Krammer, K., Lange-Bertalot H.: *Susswasser Flora von Mitteleurope*, 2/1. Gutsav Fuscher Verlag, Stuttgart, New York (1986)
15. Lange Bertalot, H.: *Iconographia Diatomologica*, vol. 8. A. R. G. Gantner and Verlag K. G, FAL 94191 Ruggell. Koeltz Scientific Books, Koenigstein (1999)
16. Krammer, K.: *Diatoms of Europe: Diatoms of European Inland waters and comparable habitats*. In: Lange-Bertalot, H. (ed.). *The genus Cymbella*, vol. 3, 584 pp., 194 pl. A. R. G. Gantner and Verlag K. G, FAL 94191 Ruggell. Koeltz Scientific Books, Koenigstein (2002)
17. Lange Bertalot, H.: *Iconographia Diatomologica*, vol. 14. In: Gantner, A.R.G., Verlag, K.G. (eds.) *Diatoms of North America* (2005)
18. Kant, S., Vohra, S.: Algal flora of J & K State. *J. Indian Bot. Soc.* **78**, 51–64 (1999)
19. Khan, M.A.: *Phycological studies in Kashmir I*. In: Kumar, A. (ed.) *Algal Biodiversity*, pp. 69–93. APH Publishing Corporation, New Delhi (2000)
20. Gandhi, H.P.: *Fresh water diatoms of central Gujarat with a review and some others*, 313 pp. Bishen Pal Singh Mahendra Pal Singh, Dehradun (1998)
21. Verma, J. Nautiyal, P.: Floristic composition of the epilithic diatoms central highland region of Indian Subcontinent: Thalassiosiraceae, Fragilariaceae, Eunotiaceae and Achnantheaceae. *J. Indian Bot. Soc.* **89**(3, 4), 397–400 (2010)
22. Karthick, B., Hamilton, P.B., Kociolek, J.P.: *An illustrated guide to common diatoms of Peninsular India*, 206 pp. Gubbi Labs, Gubbi (2013)
23. Kawecka, B.: Zonal distribution of algal communities in streams of the polish high Tatra Mts. *Acta Hydrobiol.* **13**, 393–414 (1971)
24. Kawecka, B.: Vertical distribution of algae communities in Maljovica stream (Rila-Bulgaria). *Polskie Arch Hydrobiol.* **22**, 371–420 (1974)
25. Ohtsuka, T.: Check list and illustration of diatoms in the Hii river. *Diatom* **18**, 23–56 (2002)
26. Iijima, T.: The diatoms in the River of Tenryu-gawa of Nagano Prefecture, Japan. *Bull. Bot. Soc. Nagano* **35**, 1–15 (2002)
27. Venkataramam, G.: A systematic account of some South Indian diatoms. *Proc. Indian Acad. Sci.* **10**, 293–368 (1939)
28. Krishna Murthy, V.: A contribution to the diatom flora of South India. *J. Indian Bot. Soc.* **33**, 354–381 (1954)
29. Suxena, M.R.: *Algae from Kodaikanal Hill, South India*. *Bibliotheca Phycol.* **66**, 43–99 (1983)
30. Verma, J., Nautiyal, P.: *Lotic diatom diversity patterns in mountain chains (North India)*, 270 pp. Lambert Academic Publishing, Amazon Distribution GmbH, Leipzig (2011)
31. Xing, W., Binghui, Z., Lusan, L., Li, L.: Use of diatoms in River health assessment. *Ann. Res. Rev. Biol.* **4**(24), 4054–4074 (2014)
32. Nautiyal, P., Mishra, A.S.: Epilithic diatom assemblages in a mountain stream of the lesser Himalaya (India): longitudinal patterns. *Int. J. Ecol. Environ. Sci.* **39**(3), 171–185 (2013)