
Potassium-Solubilizing Microbes: Diversity, Distribution, and Role in Plant Growth Promotion

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Abstract

Injudicious application of chemical fertilizers in India has a considerable negative impact on economy and environmental sustainability. There is a growing need to turn back to nature or sustainable agents that promote evergreen agriculture. Potassium (K) is an important and well-known constraint to crop production. Very low rates of potash fertilizer application in agricultural production lead to rapid depletion of K in the soil. Depletion of plant-available K in soils results in a variety of negative impacts of the crops yield and soil health. Microorganisms play important role in determining plant productivity. For successful functioning of introduced microbial bioinoculants, exhaustive efforts have been made to explore soil microbial diversity of indigenous community, their distribution, and behavior in soil habitats. Soil microorganisms are directly responsible for recycling of nutrients. K is the third major essential macronutrient for plant growth. The concentrations of soluble potassium in the soil are usually very low, and more than 90% of potassium in the soil exists in the form of insoluble rocks. Use of plant growth-promoting microorganisms (PGPMs) helps in increasing yields in addition to conventional plant protection. The most important PGPMs are

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Azotobacter, *Azospirillum*, *Acidithiobacillus ferrooxidans*, *Bacillus circulans*, *B. edaphicus*, *B. globisporus*, *B. mucilaginosus*, *B. subtilis*, *Burkholderia cepacia*, *Enterobacter hormaechei*, *Paenibacillus kribensis*, *P. mucilaginosus*, and *Pseudomonas putida* potassium solubilizes; these are eco-friendly and environmentally safe. Therefore, the efficient K-solubilizing microbes (KSM) should be applied for solubilization of a fixed form of K to an available form of K in the soils. This available K can be easily taken up by the plant for growth and development. In this chapter has been discussed isolation, characterization, diversity, and distribution of KSM from diverse stresses such as low and high temperatures, acidity, alkalinity, salinity, drought, and plant-associated applications. These studies elaborate on indigenous K-solubilizing microbes to develop efficient microbial bioinoculant for solubilization of K in different conditions of soil which enhances the plant growth and yield of crops.

Keywords

Abiotic stresses • Bioinoculant • Diversity • Distribution • K-solubilizing microbes

7.1 Introduction

Countries such as Brazil, China, and India are important food producers and consumers of high amounts of potassium-based fertilizers. In Brazil, around 90% of the potassium required for agriculture is imported (Barbosa Filho et al. 2006). Plants can uptake potassium (K) through the soil minerals, organic materials, and synthetic fertilizers. Consumption of K was exceeded 260 lakh tons for 2 consecutive years (2011 and 2012) in India, and all the K fertilizers were imported across the globe to meet the demand for agricultural productivity (Nagendran et al. 2013), indicating the injudicious application of K fertilizers. K deficiency in the rhizosphere of economically important crops has become an important limiting factor responsible for sustainable development of evergreen agriculture in India (Naidu et al. 2011).

Potassium (K) is one of the major plant macronutrients influencing plant growth, development, and grain quality; its plays a key role in the synthesis of cells, enzymes, proteins, starch, cellulose, and vitamins. Moreover, K not only participates in nutrient transportation and uptake but also confers resistance to abiotic and biotic stresses, leading to enhanced production of quality crops and providing resistance to plant diseases (Epstein 1972; Epstein and Bloom 2005; Maqsood et al. 2013; Pettigrew 2008). The K is absorbed by plants in large amount than any other mineral element except nitrogen (N) and, in some cases, calcium (Ca). Chemical or synthetic K fertilizers are the largest available sources of K rhizosphere; therefore, larger amounts of K fertilizers can be used to promote the availability of K for plant uptake (Li et al. 2007). The concentration of K in straw and grain serves as an indicator whether the K status of crop is deficient or sufficient (Rao et al. 2010).

However, K uptake by aboveground parts of plants is assimilated mainly into the straw but not into the grain (Basak and Biswas 2009).

Release of non-exchangeable K to the third exchangeable form occurs when level of exchangeable and solution K is decreased by crop removal, runoff, erosion, and/or leaching (Sparks 1987). With the introduction of high-yielding crop varieties/hybrids and the progressive intensification of agriculture, the soils are getting depleted in potassium reserve at a faster rate. Moreover, due to imbalanced fertilizer application, potassium deficiency is becoming one of the major constraints in crop production. This emphasized the search to find an alternative indigenous source of K for plant uptake and to maintain K status in soils for sustaining crop production (Sindhu et al. 2014; Supanjani et al. 2006).

Plant growth-promoting microbes are heterogeneous groups of microbes associated with plants in diverse ways. The plant-associated microbes colonize the rhizosphere (rhizospheric microbes), the phyllosphere (epiphytes), and inside of the plant tissue (endophytes). The word “endophyte” means “inside the plant” (derived from the Greek words “endon” meaning “within” and “phyton” meaning “plant”). Although there are diverse meanings for the term, endophytes are most commonly defined as those organisms whose “infections are inconspicuous, the infected host tissues are at least transiently symptomless, and the microbial colonization can be demonstrated to be internal”. While microbes are intimately involved in biogeochemical cycling of metals, anthropogenic release of metals has increased bacterial exposure to a high level of metals in some environments (Nies 1999; Suman et al. 2016a, b). A metal may be regarded as toxic if it impairs growth or metabolism of an organism above a certain threshold concentration: both essential and inessential metals may be toxic when supplied at high enough concentrations (Bowen 1966; Gadd 1992).

Many studies on microbial interactions with toxic metals have been made in the context of functions in metalloenzymes, resistance, and transport, but several aspects of metal “metabolism” remain unclear, particularly the mechanisms employed to obtain metals and associated nutrients from insoluble resources (Wakatsuki 1995). Frequently, microorganisms need to solubilize insoluble metal compounds occurring in the natural environment prior to uptake of essential metals and utilization of associated nutrients, e.g., P and S. Different bacterial species, such as species in the genera *Pseudomonas*, *Agrobacterium*, *Bacillus*, *Rhizobium*, and *Flavobacterium*, have been tested for their ability to solubilize inorganic phosphate compounds, such as tricalcium phosphate, hydroxyapatite, and rock phosphate (Goldstein 1986). Silicate bacteria were found to resolve potassium, silicon, and aluminum from insoluble minerals (Aleksandrov et al. 1967). K-solubilizing bacteria exert beneficial effects upon plant growth. Their uses as biofertilizers or control agents for agriculture improvement and environmental protection have been a focus of recent research (Deng et al. 2003; Glick 1995). Imbalanced or overdose use of chemical fertilizers have the negative environmental impacts and also increasing costs of crop production; therefore, there is an urgent need to imply eco-friendly and cost-effective agro-technologies to increase crop production. Therefore, the utilization of KSM is considered to be a sound strategy in improving the productivity of agricultural lands.

7.2 Isolation and Identification of Potassium-Solubilizing Microorganism

Potassium-solubilizing bacteria have been isolated and purified on Aleksandrov agar plates (Hu et al. 2006). The composition of the medium (g/liter) is 5.0 g glucose, 0.5 g magnesium sulfate, 0.005 g ferric chloride, 0.1 g calcium carbonate, 2 g calcium phosphate, and 2 g potassium-bearing minerals. Aleksandrov agar medium with different pH (3–11), NaCl concentration (5–20%), temperatures (5–50 °C), and PEG 8000 (–0.5 to –1.5 MPa) were used to isolate diverse groups of K-solubilizing microbes, viz., acidophilic, alkaliphilic, halophilic, psychrophilic, thermophilic, or drought tolerant. Plates were incubated at different temperatures, and time as described earlier by Yadav et al. (2015a). Cultures were purified and maintained at 4 °C as slant and glycerol stock (20%) at –80 °C for further use. Potassium aluminosilicates and mica were used as insoluble potassium-bearing minerals. Microbes showed halo zone on plates were selected and measured the diameter of halo zone. Quantitative potassium solubilization was carried out in Aleksandrov broth. Microbial cultures showed the K solubilization qualitatively were inoculated separately in conical flasks (150 mL) containing 40 mL broth. Available potassium in culture supernatant was determined by using flame photometer.

For identification and phylogenetic profiling of K-solubilizing microbes, the genomic DNA should be extracted for the identification of microbes by the method described by Verma et al. (2016a, b). The amount of DNA was extracted and assessed by electrophoresis on a 0.8% agarose gel. The 16S rRNA/ITS gene should be amplified as described earlier (Verma et al. (2016a, b)) using the universal primers pA and pH for bacteria and ITS 1 and ITS 2 for fungus (Edwards et al. 1989). The PCR-amplified 16S rRNA/ITS gene was purified. The nucleotide sequences of purified 16S/ITS rDNA have been sequenced with fluorescent terminators (BigDye, Applied Biosystems) and run in 3130xl Applied Biosystems ABI prism automated DNA sequencer. The DNA sequence should be double-checked by sequencing both strands using primers forward and reverse reaction, respectively. The partial 16S rRNA/ITS gene sequences of the isolated strains have been compared with those available in the databases. Identification at the species level has determined using a 16S rRNA/ITS gene sequence similarity of $\geq 97\%$ with that of a prototype strain sequence in the GenBank. Sequence alignment and comparison have been performed, using the program ClustalW. One sequence from each group was selected as a representative operational taxonomic unit (OTU). The phylogenetic tree was constructed on the aligned datasets using the neighbor-joining method implemented in the program MEGA 4.0.2 (Tamura et al. 2007).

7.3 Diversity of Potassium-Solubilizing Microorganism

Microbial world unique in each ecosystem niche forms the basis of the diversity associated. Agriculture is highly on soils and climatic conditions. The ever-increasing need for food to support the growing population in the country demands

a systematic appraisal of its soil and climate resources in order to prepare effective lands. The diversity of K-solubilizing microorganisms inhabiting different environments has been extensively investigated in the past few years. The different groups of microbes have been reported such as bacteria and fungi, which included bacterial phylum *Actinobacteria*, *Bacteroidetes*, *Firmicutes*, *Proteobacteria*, α -*Proteobacteria*, β -*Proteobacteria*, and γ -*Proteobacteria* (Fig. 7.1), and only two fungal phyla were reported to solubilize potassium, namely, *Ascomycota* and *Glomeromycota* (Table 7.2). The last few decades, potassium-solubilizing bacterial genera have been recovered, that is, *Acidithiobacillus*, *Agrobacterium*, *Aminobacter*, *Arthrobacter*, *Azotobacter*, *Bacillus*, *Clostridium*, *Delftia*, *Enterobacter*, *Klebsiella*, *Methylobacterium*, *Microbacterium*, *Myroides*, *Paenibacillus*, *Pseudomonas*, *Rhizobium*, *Salmonella*, and *Sphingomonas*. Very least research has been done in K-solubilizing fungus, with only five genera reported being *Aspergillus*, *Cladosporium*, *Fusarium*, *Glomus*, and *Penicillium* (Fig. 7.2).

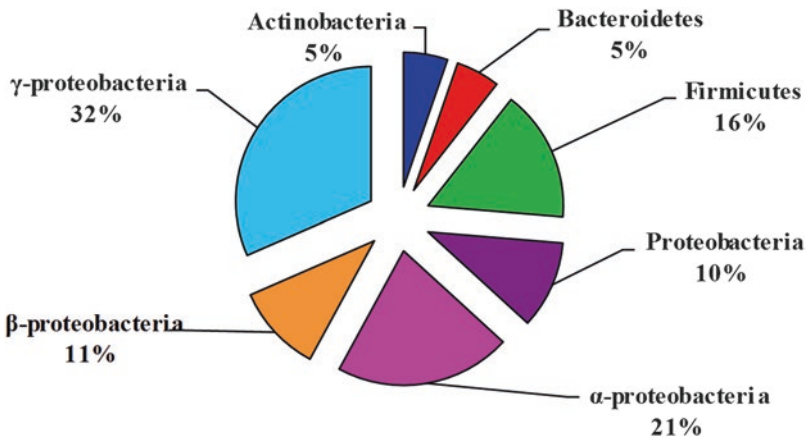


Fig. 7.1 Diversity of potassium-solubilizing bacteria

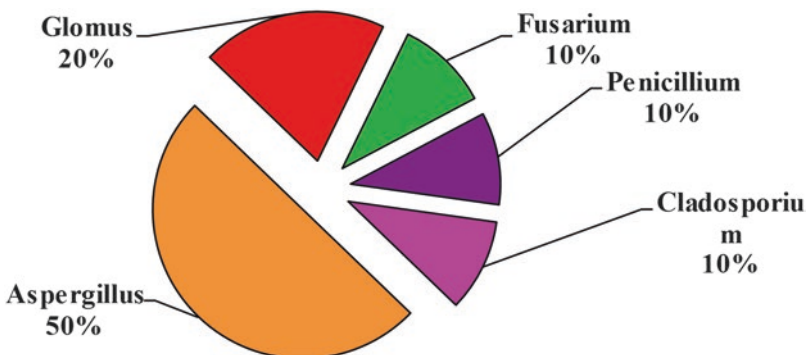


Fig. 7.2 Diversity of plant growth-promoting K-solubilizing fungi

7.3.1 Bacteria

Soil bacteria that colonize plant roots and promote growth when added to seeds, roots, or tubers have been termed plant growth-promoting rhizobacteria (PGPR). Different plant growth-promoting rhizosphere bacteria, including associative bacteria such as *Azospirillum*, *Bacillus*, *Pseudomonas*, and *Enterobacter* groups, have been used for their beneficial effects on plant growth. The mechanisms of plant growth stimulation by associative bacteria are mobilization of nutrients, stimulation of root growth by production of phytohormones, and antagonism against soil-borne plant pathogens. Several studies clearly showed the effect of plant growth-promoting bacteria on plant growth of different crops at different climates and soils. The survival of inoculated PGPR in the plant rhizosphere is in most cases a precondition for a potential plant stimulation effect during the vegetation time or at least during early plant development.

A wide range of rhizospheric bacteria reported as K solubilizers included *B. mucilaginosus* (Zarjani et al. 2013), *B. edaphicus* (Sheng 2005), *B. circulans* (Lin et al. 2002), *Burkholderia*, *Acidithiobacillus ferrooxidans*, *B. mucilaginosus* (Zhang and Kong 2014), *Bacillus edaphicus* (Sheng and He 2006), *Arthrobacter* spp. (Zarjani et al. 2013), *Enterobacter hormaechei* (Prajapati et al. 2013), *Paenibacillus mucilaginosus* (Liu et al. 2012; Hu et al. 2006), *P. frequentans*, *Cladosporium* (Argelis et al. 1993), *Aminobacter*, *Sphingomonas*, *Burkholderia* (Uroz et al. 2007), and *Paenibacillus glucanolyticus* (Sangeeth et al. 2012). These microbial strains have the ability to solubilize K from K-bearing minerals, but only few bacteria, such as *B. edaphicus* and *B. mucilaginosus*, have high capacity for mobilizing and solubilizing of K from minerals (Zhao et al. 2008).

Verma et al. (2016a, b) reported that most of the bacilli solubilized potassium such as *Bacillus aerophilus*, *Bacillus atrophaeus*, *Bacillus cereus*, *Bacillus circulans*, *Bacillus horikoshii*, *Bacillus licheniformis*, *Bacillus megaterium*, *Bacillus mojavensis*, *Bacillus pumilus*, *Bacillus sphaericus*, *Exiguobacterium antarcticum*, *Paenibacillus amylolyticus*, *Paenibacillus dendritiformis*, *Paenibacillus polymyxa*, *Planococcus citreus*, and *Planococcus salinarum*. The K-solubilizing bacteria may have use in the amelioration of K-deficient soil in agriculture. Diversity analysis of potassium solubilizing bacteria has been reported in different phylum such as 32% γ -*Proteobacteria*, 21% α -*Proteobacteria*, 16% *Firmicutes*, 11% β -*Proteobacteria*, 10% *Proteobacteria*, and 5% both *Actinobacteria* and *Bacteroidetes*. Maximum potassium-solubilizing bacterial genera have been report from γ -*Proteobacteria*.

7.3.2 Fungi

The alteration of rock minerals in natural environments is a well-known process mainly caused by the action of water and organic acids produced by plant roots and by microorganisms that accelerate this alteration. Molds are capable of solubilizing elements immobilized in silicates during the decomposition of organic matter, resulting in the production of organic acid. Potassium solubilization has been

obtained using molds such as *Aspergillus*, *Penicillium*, and *Fusarium*. The filamentous fungus *Aspergillus niger* is an exceptionally efficient producer of organic acids, which is one of the reasons for its relevance to industrial processes and its commercial importance. The production of organic acids by *A. niger* is dependent on the pH of the medium, since the greatest quantities of oxalic acid are produced at a pH between 5 and 8, while it is completely absent below pH 3.0.

Arbuscular mycorrhiza can increase the solubility of the mineral form of potassium by releasing protons, H⁺, or CO₂ and organic acid anions such as citrate, oxalate, and malate. This also increased the nitrogen, potassium, calcium, and iron in the plant leaves and fruits (Veresoglou et al. 2011; Yousefi et al. 2011). The inoculants of the two arbuscular mycorrhizal fungi (AMF) species *G. intraradices* and *G. mosseae* was applied in soil on a weight basis, and the increasing potassium uptake by maize crop was recorded (Wu et al. 2005). Ectomycorrhizal fungi particularly isolated UFSC-Pt22 and UFSC-Pt186 and contributed to the increase of the efficiency of alkaline breccias as a source of P and K to the plant growth of *Eucalyptus dunnii* seedlings, respectively (Alves et al. 2010).

Prajapati et al. (2012) reported that potassium-solubilizing fungi (KSF) strains such as *Aspergillus terreus* and *Aspergillus niger* were isolated from various K-rich soil samples and observed that *A. terreus* and *A. niger* could solubilize insoluble potassium and showed the highest available potassium in liquid medium by using two various insoluble sources of potassium, i.e., feldspar and potassium aluminum silicate. *Aspergillus* spp., *Aspergillus terreus* (Prajapati et al. 2013), *Aspergillus niger* (Prajapati et al. 2012), and *Penicillium* spp. (Sangeeth et al. 2012) enhanced K solubilization by mobilizing inorganic and organic K and release of structural K from rocks and minerals (Fig. 7.2). Diversity analysis of potassium solubilizing fungi has been reported in two phylum *Ascomycota* and *Glomeromycota* with five genera: *Aspergillus* 50%, *Glomus* 20%, *Cladosporium* 10%, *Fusarium* 10%, and *Penicillium* 10%. *Aspergillus* has been the more frequent potassium-solubilizing fungal genera.

7.4 Distribution of Potassium-Solubilizing Microorganisms

Microbial communities are found in most diverse conditions, including extremes of temperature, salinity, water deficiency, and pH. In order to survive under such extreme conditions, these organisms, referred to as extremophiles, have developed adaptive features that permit them to grow optimally under one or more environmental extremes, while polyextremophiles grow optimally under multiple conditions (Rothschild and Mancinelli 2001). Global work on PGPR for different crops is brief carried out on a hypothesis that PGPR can overcome the burden caused by chemical fertilizer on environment. There are diverse conditions for crops growing in different abiotic stresses of pH, salinity, temperature, and drought (Glick et al. 1999; Verma et al. 2015a).

In an efforts to understand the diversity and distribution of culturable K-solubilizing microbes associated with different crops growing in the diverse environments which included saline soil, acidic soil, water deficiency/drought stress, high temperature, and

low temperature, many researchers isolated, enumerated, and characterized potassium-solubilizing microbes for tolerances to abiotic stresses. Tolerance to stress provided by microbial inoculants become more significant with the perspectives of crop production that has losses due to the severity of abiotic stresses (Grover et al. 2011).

7.4.1 Acidophiles

Acidophilus study indicated that lower pH, increase in number of cells, and the consequent increase in viscosity due to EPS are allied factors affecting K solubilization from feldspar. Fourier-transform infrared spectroscopic spectra also showed the functional groups related to them which in turn indicated the presence of EPS, organic acids, and proteins (Cao et al. 2011; Yadav et al. 2011). Numerous studies have shown that *Bacillus* sp. can promote the release of K from silicate minerals (Badar et al. 2006; Barker et al. 1998). Acidophilic microorganisms has been numerously studied from different crops, rhizospheric soil, cold deserts, etc. in which some microbes were reported as acidophiles such as *Bacillus aerophilus*, *Bacillus amyloliquefaciens*, *Bacillus atrophaeus*, *Bacillus cereus*, *Bacillus circulans*, *Bacillus licheniformis*, *Bacillus pumilus*, *Lysinibacillus fusiformis*, *Paenibacillus polymyxa*, and *Planomicrobium* sp. could grow at 3 pH (Yadav et al. 2015b). Verma et al. (2013) reported K-solubilizing microbes *Bacillus cereus*, *Bacillus pumilus*, *Bacillus thuringiensis*, *Lysinibacillus fusiformis*, *Planococcus salinarum*, *Pseudomonas rhodesiae*, and *Variovorax soli*.

7.4.2 Alkaliphiles

Alkaliphilic organisms have a pH optimum for growth above pH 9 and no growth at pH 7. Spore-forming alkaliphilic organism growing at pH 8–10 but not at pH 7 has been described which was so peculiar in its properties that the authors established a new genus for it: *Amphibacillus xylanzls* (Niimura et al. 1990). Its lack of cytochromes, quinones, or catalase and its ability to form spores under aerobic as well as anaerobic conditions clearly distinguished this organism from the genera *Bacillus*, *Clostridium*, and *Sporolactobacillus*. A wide range of rhizospheric alkaliphilic microorganisms are reported as potassium solubilizers including *Achromobacter*, *Aerobacter*, *Agrobacterium*, *Arthrobacter*, *Bacillus*, *Burkholderia*, *Duganella*, *Exiguobacterium*, *Klebsiella*, *Lysinibacillus*, *Micrococcus*, *Paenibacillus*, *Planococcus*, *Pseudomonas*, *Psychrobacter*, *Rhizobium*, *Stenotrophomonas*, and *Variovorax* (Meena and Kanwar 2015; Verma et al. 2016a, b). These bacteria have been isolated from a variety of rhizospheric and non-rhizospheric soils including sugarcane (Rosa-Magri et al. 2012), tea (Bagyalakshmi et al. 2012), tobacco (Zhang and Kong 2014), and wheat (Verma et al. 2013, 2014, 2015a, 2016a, b).

7.4.3 Halophiles

Salinity affects nearly a third of the agricultural land area worldwide. Due to the upward movement of salts in soil solution in arid and semiarid climates, it is a particular problem in irrigation agriculture under those conditions (Shabala and Cuin 2007). Salinity exerts a twofold stress on the crop (Munns and Tester 2008): it causes an osmotic stress due to decreased soil water potential and an accumulation of salts in the plant cell walls. Microbial research in saline environments has also attracted the interest of researchers due to various biotechnological applications (Sahay et al. 2012). *Bacillus alcalophilus*, *Bacillus aquimaris*, *Bacillus siamensis*, *Halobacillus*, *Paenibacillus dendritiformis*, and *Lysinibacillus xylanilyticus* were reported as halophiles (Yadav et al. 2015c).

A study of potassium transport in the haloarchaeon, *Haloferax volcanii*, has shown, however, that the intracellular concentrations of potassium observed in this organism cannot be accounted for by passive processes alone and ATP hydrolysis is required to actively transport potassium into the cell to reach the 3.6 M intracellular concentrations that are maintained by *Hfx. volcanii* (Meury and Kohiyama 1989; Oren 1999). Presently, some K-solubilizing halophilic archaea have been reported: *Haloarcula marismortui*, *Haloarcula vallismortis*, and *Haloferax volcanii* (Ouellette et al. 2015). These microbes have been used for composting of different waste products and materials.

7.4.4 Psychrophiles

Cold-adapted microbes have attracted the attention of the scientific community due to their ability to promote plant growth and produce cold-active enzymes, with potential biotechnological applications in a broad range of industrial, agricultural, and medical processes. Psychrotrophic microbes could be valuable in agriculture as bioinoculants and biocontrol agents for low-temperature habitats. Many cold-tolerant PGPBs have been reported from low-temperature environments including *Arthrobacter*, *Bacillus*, *Exiguobacterium*, *Pseudomonas*, and *Providencia* (Mishra et al. 2011; Selvakumar et al. 2011; Bisht et al. 2013; Yadav et al. 2014). Psychrophiles as biofertilizers, biocontrol agents, and bioremediators would be of great use in agriculture under cold climatic conditions.

Cold-adapted microorganisms have been reported from Antarctic subglacial, permanently ice-covered lakes, cloud droplets, ice cap cores from considerable depth, snow, and ice glaciers (Yadav et al. 2015a, b). Many K-solubilizing microbes have been sorted out from different crops growing in cold environments such as *Achromobacter piechaudii*, *Bacillus amyloliquefaciens*, *Bacillus horikoshii*, *Bacillus megaterium*, *Bacillus* sp., *Exiguobacterium antarcticum*, *Klebsiella* sp., *Stenotrophomonas maltophilia*, and *Stenotrophomonas* sp. (Verma et al. 2015c). Among the isolated microbes, four efficient lignocellulolytic psychrotrophic microbes *Eupenicillium crustaceum*, *Paecilomyces* sp., *Bacillus atrophaeus*, and *Bacillus* sp.

and commercial fungal consortia *Aspergillus awamori*, *Aspergillus nidulans*, *Trichoderma viride*, and *Phanerochaete chrysosporium* were used in the present study.

7.4.5 Thermophiles

Global warming and its associated effects are expected to impose abiotic stresses, such as extremes of temperatures, drought, and flooding, which are bound to have adverse effects on food production. Climate change affects agriculture and the food production system in many ways (Godfray et al. 2011). Crop production is affected by climatic variables such as rising temperatures, changing precipitation regimes, and increased atmospheric CO₂ levels. It is also affected by biological variables such as the lengths of the crop growth periods and the crop cycle. Over the past decades, climate change has directly affected the plant growth with different abiotic stresses and change ecosystems.

Thermotolerant microbes are used as plant growth promoters to protect the diverse stresses have resulted in more production and yield in many crops. Verma et al. (2016a, b) have reported thermotolerant K-solubilizing microbes represented by *Bacillus altitudinis*, *Bacillus siamensis*, *Bacillus subtilis*, *Delftia acidovorans*, *Delftia* sp., *Methylobacterium* sp., *Methylobacterium mesophilicum*, *Pseudomonas aeruginosa*, and *Salmonella bongori* from wheat crop growing in peninsular zone of India. *Bacillus altitudinis* were also reported as thermotolerant bacteria from thermal springs (Verma et al. 2015b). These bacteria produced different hydrolytic types of enzymes at high temperature.

7.4.6 Xerophiles

Being the quantitatively most important osmoticum in plants, K is a main determinant of cell turgor (White 2013). Since an adequate turgor pressure is required for cell expansion, this parameter is particularly important in growing plants (Mengel and Busch 1982). However, for a crop growing in an increasingly dry soil, the maintenance of turgidity and water uptake from the soil requires a further reduction of the plant's osmotic potential by an increase in cellular osmolyte concentration. This "osmotic adjustment" may be accomplished by the synthesis of compatible solutes, such as sugar alcohols or amino acids (Hu and Schmidhalter 2005). However, as this process is dependent on the provision of photoassimilates, it is very costly to the plant. In contrast, the uptake and storage of increased amounts of K is an energetically "cheaper" alternative. Accordingly, hyperosmotic treatments, imitating a low soil water potential, cause a sustained K uptake into roots, e.g., of barley (Chen et al. 2005). In the field, an ample K supply will thus support osmotic adjustment and sustain cell expansion at low soil water potentials (Grzebisz et al. 2013).

Microbes have solubilized potassium in water stress condition; these are some species of microbes reported: *Paenibacillus polymyxa*, *Sporosarcina* sp., *Planococcus salinarum*, *Bacillus pumilus*, *Acidithiobacillus ferrooxidans*, *Bacillus*

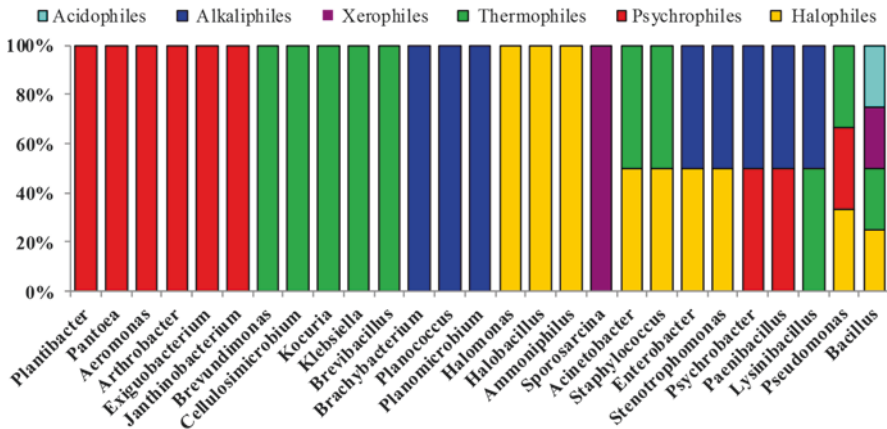


Fig. 7.3 Distribution of potassium-solubilizing microbes in diverse environments (Acidophiles: (Barker et al. 1998; Badar et al. 2006; Verma et al. 2013; Yadav et al. 2015b) Alkaliphiles: (Rosa-Magri et al. 2012; Bagyalakshmi et al. 2012; Zhang and Kong 2014; Meena and Kanwar 2015; Verma et al. 2013, 2014, 2015a, 2016a, b), Halophiles: (Meury and Kohiyama 1989; Oren 1999; Yadav et al. 2015c; Ouellette et al. 2015), Psychrophiles: (Mishra et al. 2011; Selvakumar et al. 2011; Bisht et al. 2013; Yadav et al. 2014; Verma et al. 2015c), Thermophiles: Verma et al. 2015b, 2016a, b), Xerophiles: (Sheng et al. 2002; Verma et al. 2014))

mucilaginosus, *Bacillus edaphicus*, and *Bacillus megaterium* (Sheng et al. 2002). Verma et al. (2014) have identified drought-tolerant K-solubilizing microbes such as *Bacillus megaterium*, *Duganella violaceusniger*, *Paenibacillus dendritiformis*, *Paenibacillus amylolyticus*, *Pseudomonas thivervalensis*, *Psychrobacter fozii*, *Pseudomonas monteilii*, *Pseudomonas lini*, *Stenotrophomonas maltophilia*, and *Stenotrophomonas* sp. from wheat crops growing on central zone of India. These types of microbes protect plants from water deficiency (Fig. 7.3).

7.5 Potassium Availability in the Soil and Its Relevance for Crop Production

Since the 1960s, the world population has doubled from three to seven billion, and this trend will persist in the coming decades. Because of this rapid expansion, a massive increase in crop production is required to meet the food and energy demands of future generations, while also preserving the ecological and energy-related resources of our planet. Additionally, recent climate models predict that incidences and duration of drought and heat stress periods are increasing in many regions, negatively affecting our major crops and thus our food security. Therefore, major challenges for agriculture are to enhance crop yields in more resource-efficient systems and to stabilize plant development and yield formation under biotic and abiotic stress conditions.

7.5.1 Potassium in Soils

Many soils which were initially rich in K have become deficit due to luxurious utilization by crops and inadequate application of K fertilization, soil fixation, runoff, leaching, and soil erosion by different sources (Sheng and Huang 2002; Archana et al. 2012). As mineral soils contain 0.04–3% K, the total K content of the upper 0.2 m of most agricultural soils generally ranges between 10 and 20 g kg⁻¹ (Jackson, 1964; Sparks 1987). However, in most of the soil, K (90–98%) is incorporated in the crystal lattice structure of minerals and thus not directly available for plant uptake. The availability of K differs greatly with soil type and is affected by physicochemical properties of the soil. To simplify the complex K dynamics in soil, K in soil is often classified into four groups depending on its availability to plants: water-soluble, exchangeable, non-exchangeable, and structural forms (Fig. 7.1).

Water-soluble K is directly available for plants and microbes and potentially subjected to leaching. Exchangeable K is electrostatically bound as an outer-sphere complex to the surfaces of clay minerals and humic substances (Barre et al. 2008). Both fractions are often considered to be easily available for crops. However, the size of both pools is very small. They make up only about 0.1–0.2% and 1–2% of the total K in soil, respectively (Sparks 1987). Non-exchangeable and structural forms are considered to be slowly- or non-available K sources for plants. However, these pools may also contribute significantly to the plant supply in the long term (Pal et al. 2001) (Fig. 7.4).

Most of the K in soil is in the structural form, mainly comprised of K-bearing primary minerals such as muscovite, biotite, and feldspars. K-feldspars may directly release K to the soil solution, whereas interlayer K of micas is held tightly by electrostatic forces. Weathering of K-feldspars and micas inherited from soil parent materials produces secondary soil minerals which represent the potential sources of plant-available K in soils (Singh and Goulding 1997). K in trioctahedral micas (such as biotite and phlogopite) is reported to be more readily released by weathering, and to stabilize plant development and yield formation under biotic and abiotic stress conditions (Reynolds et al. 2011). In this context, among the many plant nutrients,

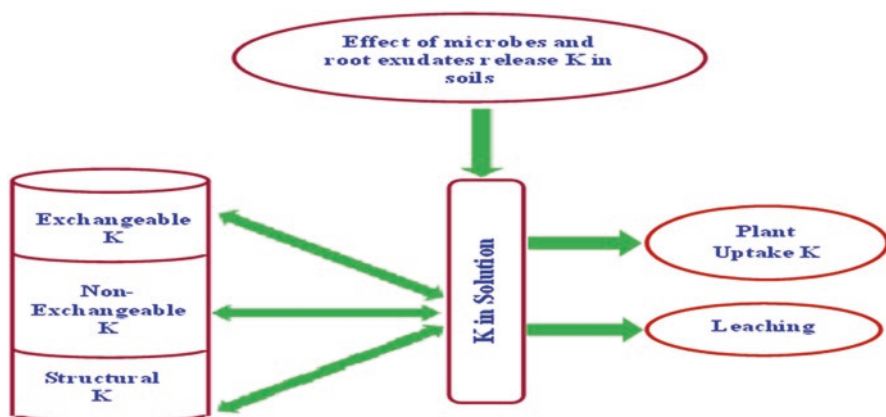


Fig. 7.4 Effects of microbes and root exudates release different forms of K in the soil

potassium (K) plays a particularly crucial role in a number of physiological processes vital to growth, yield, quality, and stress resistance of all crops.

7.5.2 Mechanisms of Potassium Solubilization

Mechanism of potassium solubilization means by which the insoluble potassium and structurally unavailable form of potassium compounds are mobilized and solubilized due to the production of various types of organic acids which are accompanied by acidolysis and complexolysis exchange reactions and these are key processes attributed to the conversion in a soluble form. The efficiency of the K solubilization by various microorganisms was found to vary according to the nature of potassium-bearing minerals and aerobic conditions (Uroz et al. 2009). The release of various types of organic acids were reflected by microorganisms to solubilized the insoluble K to an available form of K. Solubilization of feldspar and illite via rhizospheric microorganisms is due to the production of organic acids like citric acid, tartaric acids, 2-ketogluconic acid, oxalic acid, gluconic acid, malic acid, propionic acid, and fumaric acid, which is easily taken up by the plant. Glycolic and succinic acid seems to be the most frequent agent of K solubilization of mineral (Prajapati and Modi 2012; Zarjani et al. 2013).

Potassium solubilizing microbe solubilized K is done by lowering the pH or by enhancing chelation of the cations bound to K and acidolysis of the surrounding area of microorganism. Such acidolysis by organic acids produced by the rhizospheric microorganisms can either directly dissolve the mineral K as a result of slow releases of exchangeable K, readily available exchangeable K, or can chelate both Si and Al ions associated with K minerals (Romheld and Kirkby 2010). Thus, the synthesis and discharge of organic acids by the microorganisms into the surrounding environment acidify the microbe's cells and their surrounding environment that ultimately leads to the release of K ions from the mineral by protonation and acidification (Goldstein 1994).

7.5.3 Role of Potassium in Plant Growth Promotion

Plant species are known to differ in their K requirement and in their ability to take up K. The differences in absorption of K among different plant species are attributed to variations in root structure, such as root density, rooting depth, and root hair length for more details on the mechanisms of K uptake in plant roots (Nieves-Cordones et al. 2014) and for aspects on the distribution of K throughout the plant (Ahmad and Maathuis 2014; Wigoda et al. 2014). Positive correlations between K uptake efficiency and root hair length or density in K-depleted soils have been reported for maize, oilseed rape, tomato (Jungk 2001), pea, red clover, barley, rye, and perennial ryegrass (Hogh-Jensen and Pedersen 2003). Mengel and Steffens (1985) hypothesized that rye grass competes for K more effectively than red clover due to its longer root hairs and denser root system. Both morphological parameters may also deplete the K in larger volumes of soil solution, and this depletion of K can initiate the release of non-exchangeable K.

In intact plants, K uptake by leaves is stimulated by light (Blum et al. 1992). However, literature reports comparing the effect of K on photosynthesis in different plant species tend to be inconsistent. Tsonev et al. (2011) showed positive effects of K nutrition on the rate of photosynthesis only in crops subjected to some drought treatment. Similarly, Sen Gupta et al. (1989) also reported that plants supplied with elevated K levels showed similar levels of photosynthetic rates. However, when similar plants were exposed to drought, rates of photosynthesis were positively correlated with application rates of K. There are no clear explanations for how K starvation or suboptimal K nutrition downregulate photosynthesis, e.g., under drought conditions. Therefore, further research is needed to explain these findings, especially for crops. In this context, techniques such as chlorophyll fluorescence imaging may be used for noncontact detection of key physiological parameters regulating photosynthesis (e.g., quantum yield, electron transport rate) and stress defense mechanisms (heat dissipation, chlorophyll fluorescence) from the microscopic to the remote sensing scale (Chaerle et al. 2007). Capturing critical threshold values for potassium deficiencies (and quantifying optimum or slight super-optimum potassium nutrition) under high light, drought stress, or heat stress may be possible at early stages of the vegetation period. In the agronomic literature, high K concentrations in crops have often been termed “luxury consumption” which may be considered as an “insurance strategy” to enable the plant to better survive a sudden environmental stress (Kafkafi 1990).

7.5.4 Effects of Potassium-Solubilizing Microorganisms on Different Crops

The application of organo-minerals with a combination of silicate bacteria for enhancing plant growth and yield of maize and wheat was first reported by Aleksandrov et al. (1967). More importantly, research investigation conducted under field level test crops such as wheat, forage crop, maize, and Sudan grass crops has revealed that KSMs could drastically reduce the usage of chemical or organic fertilizers (Xie 1998). KSMs have been isolated from rhizospheric soil of various plants and from K-bearing mineral (Parmar and Sindhu 2013; Zhang et al. 2013); feldspar (Sheng et al. 2008); potato-soybean-cropping sequence (Biswas 2011); Iranian soils (Zarjani et al. 2013); ceramic industry soil (Prajapati and Modi 2012); mica core of Andhra Pradesh (Gundala et al. 2013); common bean (Kumar et al. 2012); biofertilizers (Zakaria 2009); sorghum, maize, bajra, and chili (Archana et al. 2013); cotton, tomato, soybean, groundnut, and banana (Archana et al. 2012); soil of Tianmu Mountain, Zhejiang Province (China) (Hu et al. 2006); rice (Muralikannan 1996); tea (Bagyalakshmi et al. 2012); Valencia orange (Shaaban et al. 2012); black pepper (Sangeeth et al. 2012); potato (Abdel-Salam and Shams 2012); thyme (Yadegari et al. 2012); eggplant (Han and Lee 2006); peanut and sesame (Youssef et al. 2010); and tobacco (Subhashini and Kumar 2014). Better crop performance was reported to be achieved from several horticultural plants, vegetables, and cereals, which were successfully inoculated with KSMs (Singh

et al. 2010; Basak and Biswas 2012; Prajapati et al. 2013). Inoculation with KSMs has been reported to exert beneficial effects on growth of cotton and rape (Sheng 2005), pepper and cucumber (Han et al. 2006), khella (Hassan et al. 2010), sorghum (Badr 2006), wheat (Sheng and He 2006), tomato (Lin et al. 2002), chili (Ramarethinam and Chandra 2005), Sudan grass (Basak and Biswas 2010), and tobacco (Zhang and Kong 2014) Table 7.1.

Prajapati et al. (2012) isolated four different potassium-solubilizing fungi from soils nearby ceramic industries and found that *Aspergillus niger* and *A. terreus* possess a greater potassium-solubilizing activity. *Aspergillus*, *Penicillium*, and *Fusarium* were reported for their remarkable activity to solubilize different kinds of insoluble mineral salts in rocks including phosphates, zinc, and potassium salts (Gour 1990; Simine et al. 1998). Lopes-Assad et al. (2010a, b) reported that *Aspergillus niger* has a better ability to solubilize silicates of potassium and aluminum. Rock powder has been solubilized by *Aspergillus niger* as a source of potassium for agroecological systems Table 7.2.

According to Archana et al. (2012), the efficient K-solubilizing bacteria *Bacillus* spp. showed increase in growth and yield of maize. It indicates that the KSMs significantly increased yield, plant growth, and nutrient uptake component over absolute fertilizer control. Supanjani et al. (2006) reported that integration of P and K rocks with inoculation of K- and P-solubilizing bacteria increased K availability from 13 to 15% and P availability from 12% to 21%, respectively. Soil application of KSMs on plant has ~16% photosynthesis and 35% higher leaf area to control. The overall result of this experiment is the treatment of P and K rocks with P- and K-solubilizing bacterial strains that were sustainable and alternative of chemical fertilizer for crop production. Bagyalakshmi et al. (2012) reported that K-solubilizing strains were isolated from rhizosphere of tea and used as biofertilizers of K in tea that have a solubilizing capacity of muriate of potash (MOP) was increased as compared to mineral K sources. Supplementation of glucose and ammonium nitrate was found to be highly effective in solubilization of MOP as compared to the other sources which should be considered prior to the application of these strains in tea soils as bioinoculants.

K-solubilizing microbes as biofertilizers for agriculture improvement can reduce the use of agrochemicals and support eco-friendly crop production (Archana et al. 2012, 2013; Kloepper et al. 1989; Requena et al. 1997; Sheng et al. 2003; Sindhu et al. 2010; Prajapati et al. 2012, 2013). Therefore, it is imperative to isolate more species of mineral-solubilizing bacteria to enrich the pool of microbial species and genes as microbial fertilizers, which will be of great benefit to the ecological development of agriculture (Liu et al. 2012). Plant growth promoting bioinoculants were assumed to have greater importance in sustainable crop protection which could increase the shelf life providing tolerance to increase adverse conditions (Suman et al. 2016a, b). K-solubilizing microorganisms develop efficient indigenous microbial consortia which are required for enhancing plant growth and yield of various crops as well as improving the soil fertility. This type of microbial consortium is cost-effective and environmentally friendly for enhancing the sustainable agriculture.

Table 7.1 Beneficial effect of potassium-solubilizing bacteria in different plants

KSM	Phylum	Source	References
<i>Acidithiobacillus ferrooxidans</i>	<i>Proteobacteria</i>	Tobacco	Zhang and Kong (2014)
<i>Agrobacterium tumefaciens</i>	α - <i>Proteobacteria</i>	Tobacco	Zhang and Kong (2014)
<i>Aminobacter</i>	<i>Proteobacteria</i>	Rhizosphere	Uroz et al. (2007)
<i>Arthrobacter</i> sp.	<i>Actinobacteria</i>	Iranian soils	Zarjani et al. (2013)
<i>Azotobacter chroococcum</i>	γ - <i>Proteobacteria</i>	Wheat and maize	Singh et al. (2010) and Sheng and He (2006)
<i>Bacillus</i>	<i>Firmicutes</i>	Cotton, tomato, soybean, groundnut, banana	Archana et al. (2012)
<i>Bacillus altitudinis</i>	<i>Firmicutes</i>	Wheat	Verma et al. (2015a, b, c)
<i>Bacillus amyloliquefaciens</i>	<i>Firmicutes</i>	Wheat	Verma et al. (2015a, b, c)
<i>Bacillus amyloliquefaciens</i>	<i>Firmicutes</i>	Mica core of Andhra Pradesh	Gundala et al. (2013)
<i>Bacillus circulans</i>	<i>Firmicutes</i>	Potato	Abdel-Salam and Shams (2012)
<i>Bacillus edaphicus</i>	<i>Firmicutes</i>	Rhizosphere	Sheng (2002)
<i>Bacillus globisporus</i>	<i>Firmicutes</i>	Weathered feldspar	Sheng et al. (2008)
<i>Bacillus licheniformis</i>	<i>Firmicutes</i>	<i>Oryza sativa</i> , <i>Zea mays</i> , <i>Sorghum bicolor</i> , and wheat	Sheng et al. (2008), Singh et al. (2010) and Basak and Biswas (2012)
<i>Bacillus megaterium</i>	<i>Firmicutes</i>	Valencia orange	Shaaban et al. (2012)
<i>Bacillus mucilaginosus</i>	<i>Firmicutes</i>	Eggplant, black pepper, maize, wheat	Han and Lee 2006, Sangeeth et al. (2012) and Prajapati et al. (2013)
<i>Bacillus</i> sp.	<i>Firmicutes</i>	Rice	Muralikannan (1996)
<i>Bacillus</i> sp. BPR7	<i>Firmicutes</i>	Common bean	Kumar et al. (2012)
<i>Bacillus subtilis</i>	<i>Firmicutes</i>	Wheat	Verma et al. (2016a, b)
<i>Bacillus thuringiensis</i>	<i>Firmicutes</i>	Cold desert	Yadav et al. (2016)
<i>Burkholderia</i>	β - <i>Proteobacteria</i>	Tobacco	Uroz et al. (2007) and Zhang and Kong (2014)
<i>Burkholderia cepacia</i>	β - <i>Proteobacteria</i>	Tobacco	Zhang and Kong (2014)
<i>Clostridium pasteurianum</i>	<i>Firmicutes</i>	Rhizosphere	Reitmeir (1951)
<i>Delftia acidovorans</i>	β - <i>Proteobacteria</i>	Wheat	Verma et al. (2016a, b)
<i>Delftia</i> sp.	β - <i>Proteobacteria</i>	Wheat	Verma et al. (2016a, b)
<i>Enterobacter aerogenes</i>	γ - <i>Proteobacteria</i>	Tobacco	Zhang and Kong (2014)
<i>Enterobacter asburiae</i>	γ - <i>Proteobacteria</i>	Tobacco	Zhang and Kong (2014)
<i>Enterobacter cloacae</i>	γ - <i>Proteobacteria</i>	Tobacco	Zhang and Kong (2014)
<i>Enterobacter hormaechei</i>	γ - <i>Proteobacteria</i>	Ceramic industry soil	Prajapati and Modi (2012)

(continued)

Table 7.1 (continued)

KSM	Phylum	Source	References
<i>Klebsiella variicola</i>	γ -Proteobacteria	Tobacco	Zhang and Kong (2014)
<i>Methylobacterium mesophilicum</i>	α -Proteobacteria	Wheat peninsular zone	Verma et al. (2016a, b)
<i>Methylobacterium</i> sp.	α -Proteobacteria	Wheat peninsular zone	Verma et al. (2016a, b)
<i>Microbacterium foliorum</i>	α -Proteobacteria	Tobacco	Zhang and Kong (2014)
<i>Myroides odoratimimus</i>	Bacteroidetes	Tobacco	Zhang and Kong (2014)
<i>Paenibacillus frequentans</i>	Firmicutes	Rhizosphere	Argelis et al. (1993)
<i>Paenibacillus glucanolyticus</i>	Firmicutes	Rhizosphere	Sangeeth et al. (2012)
<i>Paenibacillus kribensis</i> CX-7	Firmicutes	Rhizosphere soil, wheat soil of Chang'an, Shanxi Province	Parmar and Sindhu (2013) and Zhang et al. (2013)
<i>Paenibacillus mucilaginosus</i>	Firmicutes	Soil of Tianmu Mountain, Zhejiang Province (China)	Hu et al. (2006)
<i>Paenibacillus</i> spp.	Firmicutes	Rhizosphere	Sheng et al. (2008), Singh et al. (2010) and Basak and Biswas (2012)
<i>Pantoea agglomerans</i>	γ -Proteobacteria	Tobacco	Zhang and Kong (2014)
<i>Pseudomonas azotoformans</i>	γ -Proteobacteria	<i>Oryza sativa</i> , <i>Zea mays</i> , <i>Sorghum bicolor</i> , and <i>Triticum aestivum</i>	Sheng et al. (2008), Singh et al. (2010) and Basak and Biswas (2012)
<i>Pseudomonas</i>	γ -Proteobacteria	Sorghum, maize, bajra, chili	Archana et al. (2013)
<i>Pseudomonas putida</i>	γ -Proteobacteria	Tea	Bagyalakshmi et al. (2012)
<i>Rhizobium</i>	α -Proteobacteria	Wheat and maize	Sheng and He (2006)
<i>Salmonella bongori</i>	γ -Proteobacteria	Wheat	Verma et al. (2016a, b)
<i>Sphingomonas</i>	α -Proteobacteria	Rhizosphere	Uroz et al. (2007)

Table 7.2 Beneficial effect of potassium-solubilizing fungi in different sources

KSM	Phylum	Source	References
<i>Aspergillus fumigatus</i>	Ascomycota	Waste disposal	Lopes et al. (2010)
<i>Aspergillus awamori</i>	Ascomycota	Compost	Biswas DR (2011) and Shukla et al. (2016)
<i>Aspergillus niger</i>	Ascomycota	Rock powder, tea	Prajapati et al. (2012) and Nath et al. (2015)
<i>Aspergillus</i> spp.	Ascomycota	K-rich soil	Prajapati et al. (2013)
<i>Aspergillus terreus</i>	Ascomycota	K-rich soil	Prajapati et al. (2012)
<i>Cladosporium</i>	Ascomycota	Tobacco	Zhang and Kong (2014)
<i>Fusarium solani</i>	Ascomycota	Cutinase enzymes	Sebastiao et al. (1993)
<i>Glomus intraradices</i>	Glomeromycota	Maize	Wu et al. (2005)
<i>Glomus mosseae</i>	Glomeromycota	Maize	Wu et al. (2005)
<i>Penicillium</i> spp.	Ascomycota	K-rich soil	Sangeetha et al. (2012)

7.6 Conclusions

In this book chapter, we summarize current knowledge regarding the importance of K in plant growth and quality in changing climate and discuss also the factors controlling K availability in soil. Potassium solubilizing microorganisms play an important role in plant nutrition that enhances the K acquisition of plants through soil which increase plant growth promotion activities; these KSM contributions play an important role to bio-fertilization of agricultural crops. Accordingly, further investigation is required to improve the performance and use of potassium-solubilizing microorganism as efficient microbial bioinoculants. The greater attention is needed for studies and application of new efficient combinations of potassium-solubilizing microorganisms and other plant growth-promoting microorganisms for improved results. The mechanisms explaining the synergistic interaction among KSM required further research to elucidate the molecular basis of these interactions. On the other hand, the application of biotechnological tools for genetic manipulation of potassium solubilizing microorganism increases their potassium-solubilizing efficiency/ability/capabilities and/or the insertion of this trait into other strains of plant growth-promoting effects.

7.7 Future Prospects

The K fertilizer supply is often inadequate due to economic reasons, unavailability of fertilizers, or limited knowledge. Fertilizer application techniques may be still better adjusted to the prevailing crop and growth conditions, e.g. as foliar sprays. An increased utilization of the large plant-non-available pool of soil K could decrease the fertilization requirements and improve crop performance, in particular in low-input systems. Ways to tap this resource would be the introduction of competitive K-mobilizing bacterial strains and the design of more K-efficient crop genotypes by

conventional breeding or targeted biotechnological strategies. Promising targets for an improvement of K uptake are root morphology and anatomy, transporter kinetic and regulation, as well as the release of root exudates. There is considerable variation among species and cultivars in those traits. Optimized K fertilizer application in K limited soils is crucial in order to enhance plant response especially to drought stress via enhancing adaptive/resistance mechanisms of crop plants. Especially, because the demand of K is expected to increase significantly, in particular in developing regions of the world.

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