

Chapter 5

The 100%-Complete Nuclear and Organellar Genome Sequences of the Ultrasmall Red Algal Species *Cyanidioschyzon merolae* 10D

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Abstract We were the first group to successfully sequence the 100%-complete entire eukaryotic genome in 2007, mainly using automated Sanger sequencing of the unicellular, ultrasmall red algal species *Cyanidioschyzon merolae* 10D. This world record was principally based on the ultrasmall size of the *C. merolae* genome (ca. 16 megabase pairs) as well as on three excellent previous studies: the 100%-complete mitochondrial genome in 1998, the 100%-complete plastid genome in 2003, and the first algal cell nuclear genome in 2004. The 100%-complete nuclear sequences demonstrated that this ultrasmall red alga contains unusually simple sets of genes and genetic sequences. For example, because introns are lacking in almost all of the protein-coding nuclear genes of *C. merolae*, the 100%-complete sequence can be used to directly deduce the sequences of all *C. merolae* proteins, which will be extremely valuable in further proteomics research. Thus, this small red alga represents an ideal model organism for studying the fundamental relationships among the plastid, mitochondrial, and nuclear genomes. The 100%-complete nuclear genome sequence has greatly improved the precision and value of biological analyses of *C. merolae*.

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5.1 Introduction

The eukaryotic cell has three types of nuclei that contain nuclear, mitochondrial, and plastid genomes (Kuroiwa 1982). Thus, biological attributes and/or functions in each species of photosynthetic eukaryotes are fundamentally based on consortia of these three genomic information sources, except for some nucleomorph-containing secondary phototrophs such as cryptophytes and chlorarachniophytes (Curtis et al. 2012). Therefore, complete determination of the information from all of the three genomes in a single eukaryotic species was considered tantamount to revealing the complete molecular blueprint of the eukaryote. Thanks to the skillful and patient work of Japanese researchers, the first complete plastid genome was successfully determined through manual DNA sequencing based on the radiolabeling method (Ohyama et al. 1986; Shinozaki et al. 1986) and the first cyanobacterial genome through the automated Sanger DNA sequencing method (Kaneko et al. 1996).

Even in the current era of next-generation sequencing, the 100%-complete sequencing of all three eukaryotic genomes may seem as crazy an endeavor as “eating a bicycle,” which was once recorded in the Guinness Book of World Records (although this disturbing record has since been prohibited from nomination to the book). However, we indeed succeeded in the first 100%-complete eukaryotic genome in 2007, using automated Sanger sequencing of the unicellular red algal species *Cyanidioschyzon merolae* 10D (Nozaki et al. 2007). This world record was principally based on the ultrasmall genome size (ca. 16 megabase pairs) of *C. merolae*, as well as on three excellent previous studies: the 100%-complete mitochondrial genome sequence (Ohta et al. 1998), the 100%-complete plastid genome sequence (Ohta et al. 2003), and the first algal cell nuclear genome sequence (Matsuzaki et al. 2004).

Although our original nuclear genomic sequence of *C. merolae* 10D revealed some unique features such as very few introns, only three copies of ribosomal DNA (rDNA), and a relatively small number of total genes (Matsuzaki et al. 2004), the exact features of certain important repeated elements such as histone gene clusters and telomeres remained uncertain. Due to the functional significance of these elements, it was extremely desirable to complete the sequence and resolve all ambiguities to summarize all of the unique genomic features of *C. merolae*. Thus, we painstakingly elucidated all of the nucleotide sequences remaining as gaps and telomere-lacking chromosomal ends, to construct the first 100%-complete nuclear genome sequence (Nozaki et al. 2007).

We believed that the establishment of the first 100%-complete eukaryotic genome would be of great widespread interest to the biological sciences as a whole and that demonstration of the simplest set of genomic features from the

hot spring red alga *C. merolae* would represent an important advancement in the fields of genomics and evolutionary biology. More than 10 years have passed since we determined the 100%-complete genome sequences of the mitochondrial, plastid, and nuclear genomes in *C. merolae* (Ohta et al. 1998, 2003; Nozaki et al. 2007). Recent advancements in next-generation sequencing methods have contributed much to the establishment of the organellar and nuclear whole genomes of numerous eukaryotes (e.g., Hanschen et al. 2016; Session et al. 2016). However, no other 100%-complete eukaryotic genomes have been determined. Thus, our studies (Ohta et al. 1998, 2003; Nozaki et al. 2007) remain the first and only achievement of a 100%-complete eukaryotic genome sequence.

5.2 Mitochondrial Genome

The 100%-complete nucleotide sequence of the mitochondrial genome of *C. merolae* 10D is on circular DNA containing 34 or 35 genes encoding proteins (including unidentified open reading frames [ORFs]) and 28 RNA genes (3 rRNAs and 25 tRNAs) (Table 5.1, Fig. 5.1.; Ohta et al. 1998; Yang et al. 2015). The genes are encoded on both strands. This is the largest number of protein-coding genes within the current 37 sequenced red algal mitochondrial genomes (Yang et al. 2015). The G + C content of the *C. merolae* mitochondrial genome is 27.2%. The genome size is 32,211 base pairs (bp), exceeding the average size (26,866 bp) of the 37 red algal mitochondrial genomes (Yang et al. 2015). The genome size and number of protein-coding genes of another cyanidial species, *Galdieria sulphuraria*, are 21,428 bp and 19 genes, respectively. Thus, the large size and gene richness of the *C. merolae* mitochondrial genome do not represent Cyanidiales-specific characteristics.

5.3 Plastid Genome

The 100%-complete nucleotide sequence of the plastid genome of *C. merolae* 10D is on circular DNA composed of 149,987 bp lacking inverted repeats (Table 5.1, Fig. 5.2). The G + C content is 37.6% (Ohta et al. 2003). This organelle genome contains 243 genes on both strands and consists of 207 protein-coding genes (including unidentified ORFs) and 36 RNA genes (a ribonuclease P RNA component, 3 rRNAs, 31 tRNAs, and tmRNA). Approximately 40% of the protein-coding genes overlap, and none of the genes in this plastid genome have introns (Ohta et al. 2003).

Recently, the plastid genomes of various red algal species have been sequenced (Janouškovec et al. 2013; Tajima et al. 2014). The *C. merolae* plastid genome is similar to other red algal plastid genomes in that it is gene rich: 223–250 unique genes are found in plastid genomes of red algae (Janouškovec et al. 2013). The

Table 5.1 Key attributes of the 22 chromosomes constituting the 100%-complete three genomes of the ultrasmall red alga *Cyanidioschyzon merolae* 10D

Genome/ chromosome	No. of nucleotides (bp)	Shape of chromosome	No. of protein-coding genes
Cell nucleus ^a			
1	422,616	Linear	102
2	457,013	Linear	125
3	481,791	Linear	144
4	513,455	Linear	140
5	528,682	Linear	161
6	536,163	Linear	131
7	584,452	Linear	173
8	739,753	Linear	213
9	810,151	Linear	231
10	839,707	Linear	247
11	852,849	Linear	236
12	859,119	Linear	258
13	866,983	Linear	249
14	852,727	Linear	256
15	902,900	Linear	265
16	908,485	Linear	261
17	1,232,258	Linear	355
18	1,253,087	Linear	360
19	1,282,939	Linear	384
20	1,621,617	Linear	484
Total	16,546,747		4775
Unassigned	0		0
Plastid ^b	149,987	Circular	208
Mitochondrion ^c	32,211	Circular	34
Total of three genomes	16,728,945		5017

^aNozaki et al. (2007)^bOhta et al. (2003)^cOhta et al. (1998)

striking feature of the *C. merolae* genome within the red algae is the high degree of gene compaction: it has the smallest genome size and the shortest intergenic region (Janouškovec et al. 2013). The median intergenic distance of the *C. merolae* plastid genome is extremely small (only 10 bp), whereas the intergenic distances of other red algal plastid genomes (including that of another cyanidial species, *Cyanidium caldarium*) range from 61 to 85 bp (Janouškovec et al. 2013). Thus, the extraordinary compaction of the plastid genome of *C. merolae* has likely resulted from species-specific factors following the divergence of *Cyanidioschyzon*

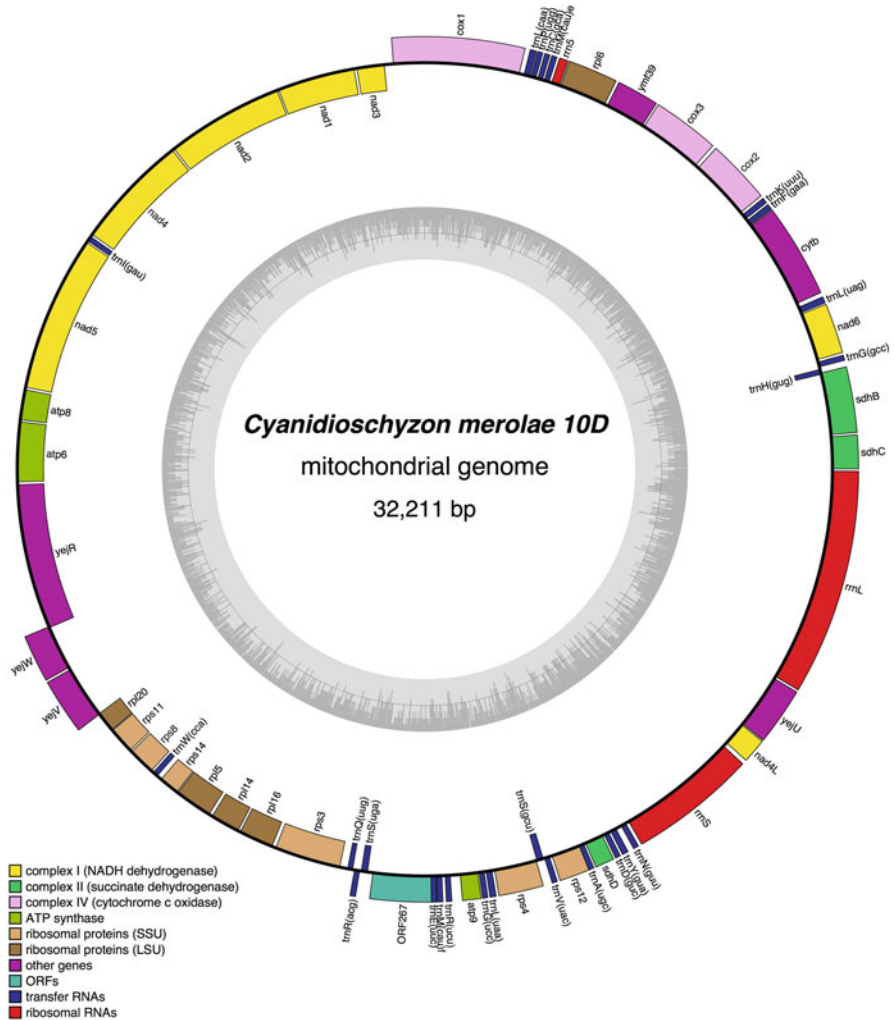


Fig. 5.1 Genetic map of the *C. merolae* mitochondrial genome. Note the *C. merolae* mtDNA is a circular-mapping molecule. The map was drawn based on the 100%-complete sequence (NC_000887; Ohta et al. 1998) using OrganellarGenomeDRAW <http://ogdraw.mpimp-golm.mpg.de/index.shtml>

from other cyanidiallean algae, such as *Cyanidium* (Janoušek et al. 2013). The plastid genome of *C. merolae* encodes several genes that are rarely present in other plastid genomes (Ohta et al. 2003). Recently, 16 cyanobacterial genes were resolved to be present in the plastid genomes of only two Cyanidiales (*C. merolae* and *Cyanidium caldarium*) out of all known red algal plastid genomes (Janoušek et al. 2013). Because plastid gene loss (endosymbiotic gene transfer) is slow in red algal plastid genomes (Nozaki et al. 2003; Janoušek et al. 2013)

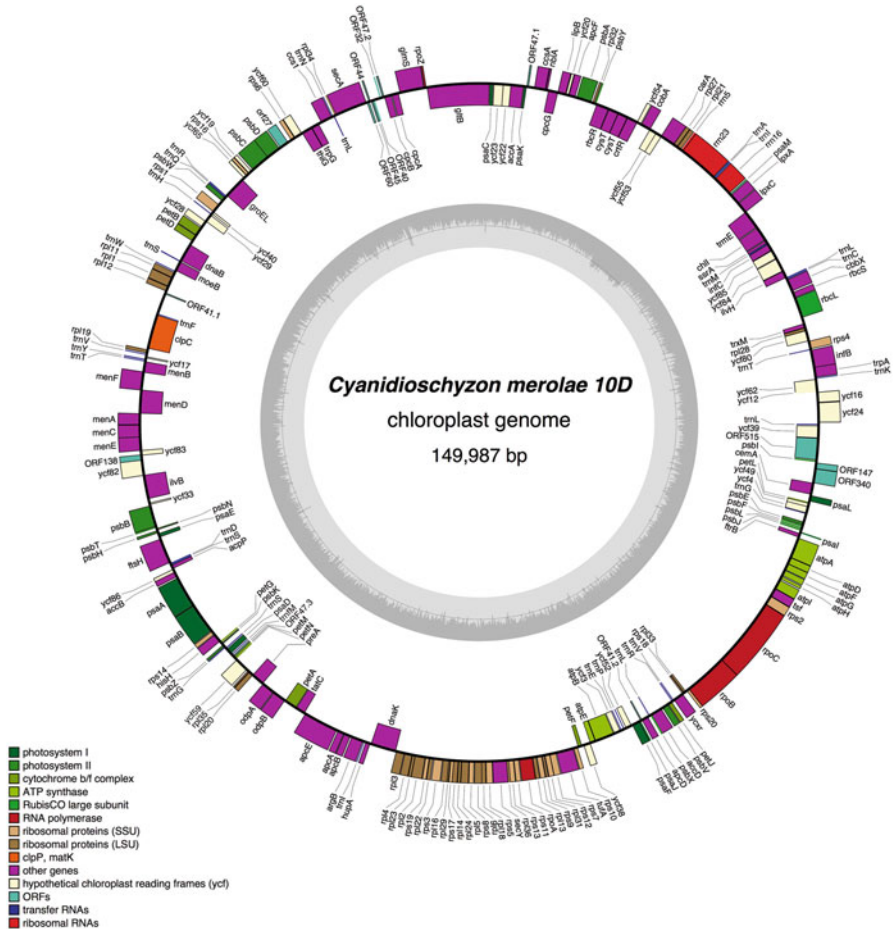


Fig. 5.2 Genetic map of the *C. merolae* chloroplast genome. The map was drawn based on the 100%-complete sequence (NC_004799; Ohta et al. 2003) using OrganellarGenomeDRAW <http://ogdraw.mpimp-golm.mpg.de/index.shtml>

and these two cyanidiallean species are distantly related to other red algae with published plastid genomes, the 16 genes may be ancestral cyanobacterial genes that were lost in the latter red algae.

5.4 Nuclear Genome

To construct a 100%-complete nuclear genome sequence of *C. merolae* 10D, unresolved gaps between contigs and undetermined chromosome ends were filled using our previously constructed *C. merolae* bacterial artificial chromosome (BAC)

clones (Matsuzaki et al. 2004). Polymerase chain reactions (PCRs) of BAC clones containing these gaps were performed using specific primers complementary to sequences flanking the gaps (Nozaki et al. 2007). We used DNA walking annealing control primer technology in order to directly amplify unknown sequences adjacent to known sequences within a contig. PCR products were sequenced by using the cycle sequencing methodology, except for a single gap with extremely high G + C content on chromosome 10, which was filled by an in vitro transcription sequencing reaction (Nozaki et al. 2007). Chromosomal ends were sequenced by the inverse PCR method, polyC-tailing and the anchor primer method, and the asymmetric PCR method (Nozaki et al. 2007). To completely determine the sequences of the histone cluster area in this red algal genome, we performed *NotI* and *ApaI* subcloning of the BAC clone GESZ2-b20, which includes possible histone clusters on chromosome 14 (Matsuzaki et al. 2004). We performed Southern blot analysis with histone-related probes, restriction enzyme analysis, and end sequencing of the subclones to reveal the relative positions of the subclones on this chromosome. We completely filled six gaps between contigs/fragments in the area of histone cluster (Matsuzaki et al. 2004) using primer walking of the subclones (Nozaki et al. 2007). Our complete nuclear genome sequence of *C. merolae* consisting of 16,546,747 nucleotides covers 100% of the 20 linear chromosomes from telomere to telomere (Table 5.1, Fig. 5.3). These 20 unambiguous DNA molecules represent the simple and unique structures of eukaryotic chromosomes (Nozaki et al. 2007): a histone gene cluster of the smallest known size, all chromosomal ends with a unique telomeric repeat, and an extremely low number of transposons. Based on these genomic features and others that were discovered previously (Matsuzaki et al. 2004), *C. merolae* appears to contain the simplest nuclear genome of the nonsymbiotic species of eukaryotes. These unusually simple genomic features found in this 100%-complete genome sequence were considered very useful for further biological studies of cells of eukaryotes.

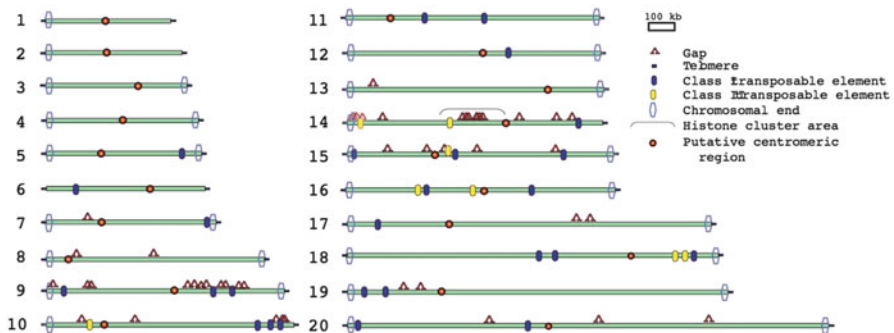


Fig. 5.3 Bird's-eye view of the 100%-complete structures of 20 chromosomes of *C. merolae* showing regions that were filled by Nozaki et al. (2007; “gap” and “chromosomal end”), the histone cluster area, telomere repeats, putative centromeric regions, and transposable elements (“class I” and “class II”) (From Nozaki et al. (2007))

5.5 Centromere Regions and Their Dynamics

The centromere is a conserved chromosomal site responsible for attachment of spindles and accurate segregation of chromosomes during mitosis and meiosis. Each chromosome in a eukaryotic cell has a unique centromere where the kinetochore complex assembles and captures the spindle (Cleveland et al. 2003). In addition, the centromere core contains the histone H3 variant, centromere protein A (CENP-A), which replaces canonical H3 at the centromere. The 100%-complete genome sequence of *C. merolae* clarified that each chromosome also possesses a single A + T-rich region, which is predicted to be the putative centromere region (Matsuzaki et al. 2004; Maruyama et al. 2008). By ChIP-on-chip analysis using an anti-CENP-A antibody and a whole-genome tiling array, core centromere sequences were determined at the predicted region (Kanesaki et al. 2015). The identified centromeres were of the regional type, 1–3 kb in length, and with no consensus sequences or repeat elements. The expression of the CENP-A protein rapidly increased during the S phase of the cell cycle; subsequently a drastic reconstitution into two discrete foci adjacent to the spindle poles occurred at metaphase (Maruyama et al. 2007). The dynamics of condensins I and II during the cell cycle were also well analyzed (Fujiwara et al. 2013a, b). In *C. merolae*, condensin II is enriched at centromere regions and is absent along chromosome arms during metaphase of the M phase, whereas condensin I is not enriched at centromere regions. Furthermore, *C. merolae* possesses the centromere proteins CENP-A, CENP-E, and the hypothetical CENP-C, but does not possess CENP-B or CENP-T. The characteristics of the centromere/kinetochore in lower plants and algae remain largely unknown. Thus, *C. merolae* is the most well-studied model system for clarifying the detailed mechanisms of centromere dynamics in primitive eukaryotic plant cells.

5.6 Conclusions

A histone gene cluster of the smallest known size, all chromosomal ends with a unique telomeric repeat, and an extremely low number of transposons in *C. merolae* (Nozaki et al. 2007), as well as other simple features of the *C. merolae* nuclear genome (Matsuzaki et al. 2004; Misumi et al. 2005), are extremely distinctive and represent the simplest set of features of genomes recognized in any nonsymbiotic eukaryote yet studied (Fig. 5.4). It is generally considered that such simple genomic features are the consequences of reductive evolution of a eukaryote of ultrasmall size (Derelle et al. 2006). However, none of these features are shared by *Ostreococcus*, a similarly ultrasmall green alga, in which histone genes are distributed among 6 or more chromosomes, 39% of the genes harbor introns, and 8166 protein genes and 417 transposable elements are dispersed across the chromosomes (Derelle et al. 2006). These characteristics suggest differences in modes of

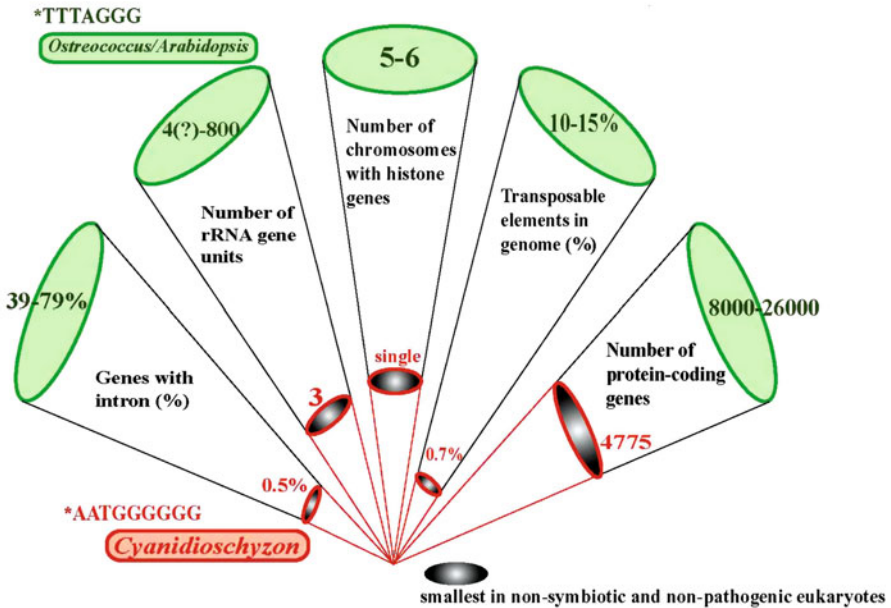


Fig. 5.4 Comparison of the three nuclear genomes of photosynthetic eukaryotes, *Cyanidioschyzon*, *Ostreococcus* (a green alga of ultrasmall size), and *Arabidopsis* (a seed plant). Telomere repeat sequences are indicated by asterisks above the generic names (Based on Nozaki et al. (2007))

reduction of genomes between the ancestors of *Cyanidioschyzon* (red algae) and *Ostreococcus* (Chloroplastida [green plants and algae]). On the other hand, algae growing in acidic hot springs (pH 1.5, 45 °C) may be candidates for retaining ancient or ancestral attributes of plants, because throughout Earth's history, volcanic activity is thought to have provided such an extreme environment. According to Cunningham et al. (2006), *C. merolae* has perhaps the simplest chlorophyll and carotenoid assortment found in any photosynthetic eukaryote. In addition, the *C. merolae* plastid and mitochondrial genomes contain large numbers of genes, which are thought to be ancestral features, because it is generally considered that reversal of plastid gene loss is impossible (Martin et al. 2002; Nozaki et al. 2003). Thus, we hypothesized that some of the unusual or simple characteristics of the *C. merolae* genome may represent ancestral features that have been conserved in the *Cyanidioschyzon* lineage but have become modified extensively during the evolution of other lineages of plants/algae (Nozaki et al. 2007). Alternatively, the unique features of the *C. merolae* genome may reflect adaptations to their extreme environment. However, many simple features found in the *C. merolae* genome, such as the rare presence of introns and few rRNA regions in the nuclear genome, are not present in the nuclear genome sequence of another hot spring red alga, *Galdieria* (Barbier et al. 2005; Schönknecht et al. 2013). Organellar genomic features are also different between *Cyanidioschyzon* and *Galdieria*, as discussed above. Based on the

100%-complete nuclear, mitochondrial, and plastid genome sequences (Ohta et al. 1998, 2003; Matsuzaki et al. 2004; Nozaki et al. 2007), all of the major types of genetic information in eukaryotes are present in *C. merolae*. Furthermore, *C. merolae* contains unusually simple sets of genes and sequences as revealed by the 100%-complete genome. Because introns are lacking in almost all protein-coding nuclear genes of this ultrasmall red alga, the 100%-complete genome can be used to directly deduce the sequences of all red algal proteins, which will be very valuable in future research of proteomics. Thus, *C. merolae* represents an ideal model for studying the fundamental relationships among the chloroplast, mitochondrial, and nuclear genomes. The 100%-complete nuclear genome sequence (Nozaki et al. 2007) has greatly improved the precision of biological analyses of *C. merolae*, including studies of the dividing machineries of plastids (Yoshida et al. 2010) and peroxisomes (Imoto et al. 2013). The 100%-complete genome sequence also enables us to design various tools for genome-wide analyses, such as an organellar DNA microarray (Minoda et al. 2005; Kanesaki et al. 2012), nuclear DNA microarray (Fujiwara et al. 2009; Kanesaki et al. 2009; Imamura et al. 2009), and high-density whole-genome tiling array (Kanesaki et al. 2015). Furthermore, the 100%-complete genome sequence greatly contributed to the establishment of homologous recombination techniques (Minoda et al. 2004; Fujiwara et al. 2013a, b; Taki et al. 2015) and conditional gene expression systems (Sumiya et al. 2014; Fujiwara et al. 2015). The establishment of these advanced molecular biological techniques has made *C. merolae* one of the most exceptional model organisms among the eukaryotic algae.

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References

- Barbier G, Oesterhelt C et al (2005) Comparative genomics of two closely related unicellular thermo-acidophilic red algae, *Galdieria sulphuraria* and *Cyanidioschyzon merolae*, reveals the molecular basis of the metabolic flexibility of *Galdieria sulphuraria* and significant differences in carbohydrate metabolism of both algae. *Plant Physiol*, 137:460–474. <https://doi.org/10.1104/pp.104.051169>
- Cleveland DW, Mao Y et al (2003) Centromeres and kinetochores: from epigenetics to mitotic checkpoint signaling. *Cell* 112:407–421. [https://doi.org/10.1016/S0092-8674\(03\)00115-6](https://doi.org/10.1016/S0092-8674(03)00115-6)
- Cunningham FX Jr, Lee H et al (2006) Carotenoid biosynthesis in the primitive red alga *Cyanidioschyzon merolae*. *Eukaryot Cell* 6:533–545. <https://doi.org/10.1128/EC.00265-06>
- Curtis BA, Tanifuji G et al (2012) Algal genomes reveal evolutionary mosaicism and the fate of nucleomorphs. *Nature* 492:59–65. <https://doi.org/10.1038/nature11681>
- Derelle E, Ferraz C et al (2006) Genome analysis of the smallest free-living eukaryote *Ostreococcus tauri* unveils many unique features. *Proc Natl Acad Sci U S A* 103:11647–11652. <https://doi.org/10.1073/pnas.0604795103>

- Fujiwara T, Misumi O et al (2009) Periodic gene expression patterns during the highly synchronized cell nucleus and organelle division cycles in the unicellular red alga *Cyanidioschyzon merolae*. *DNA Res* 16:59–72. <https://doi.org/10.1093/dnares/dsn032>
- Fujiwara T, Ohnuma M et al (2013a) Gene targeting in the red alga *Cyanidioschyzon merolae*: single- and multi-copy insertion using authentic and chimeric selection markers. *PLoS One* 8: e73608. <https://doi.org/10.1371/journal.pone.0073608>
- Fujiwara T, Tanaka K et al (2013b) Spatiotemporal dynamics of condensins I and II: evolutionary insights from the primitive red alga *Cyanidioschyzon merolae*. *Mol Biol Cell* 24:2515–2527. <https://doi.org/10.1091/mbc.E13-04-0208>
- Fujiwara T, Kanesaki Y et al (2015) A nitrogen source-dependent inducible and repressible gene expression system in the red alga *Cyanidioschyzon merolae*. *Front Plant Sci* 6:657. <https://doi.org/10.3389/fpls.2015.00657>
- Hanschen ER, Marriage TN et al (2016) The *Gonium pectorale* genome demonstrates co-option of cell cycle regulation during the evolution of multicellularity. *Nat Commun* 7:11370. <https://doi.org/10.1038/ncomms11370>
- Imamura S, Kanesaki Y et al (2009) R2R3-type MYB transcription factor, CmMYB1, is a central nitrogen assimilation regulator in *Cyanidioschyzon merolae*. *Proc Natl Acad Sci U S A* 106:12548–12553. <https://doi.org/10.1073/pnas.0902790106>
- Imoto Y, Kuroiwa H et al (2013) Single-membrane-bounded peroxisome division revealed by isolation of dynamin-based machinery. *Proc Natl Acad Sci U S A* 110:9583–9588. <https://doi.org/10.1073/pnas.1303483110>
- Janouškovec J, Liu S-L et al (2013) Evolution of red algal plastid genomes: ancient architectures, introns, horizontal gene transfer, and taxonomic utility of plastid markers. *PLoS One* 8:e59001. <https://doi.org/10.1371/journal.pone.0059001>
- Kaneko T, Sato S et al (1996) Sequence analysis of the genome of the unicellular cyanobacterium *Synechocystis* sp. strain PCC6803. II. Sequence determination of the entire genome and assignment of potential protein-coding regions. *DNA Res* 3:109–136
- Kanesaki Y, Kobayashi Y et al (2009) Mg-protoporphyrin IX signaling in *Cyanidioschyzon merolae*: multiple pathways may involve the retrograde signaling in plant cells. *Plant Signal Behav* 4:1190–1192. <https://doi.org/10.4161/psb.4.12.10061>
- Kanesaki Y, Imamura S et al (2012) External light conditions and internal cell cycle phases coordinate accumulation of chloroplast and mitochondrial transcripts in the red alga *Cyanidioschyzon merolae*. *DNA Res* 19:289–303. <https://doi.org/10.1093/dnares/dss013>
- Kanesaki Y, Imamura S et al (2015) Identification of centromere regions in chromosomes of a unicellular red alga, *Cyanidioschyzon merolae*. *FEBS Lett* 589:1219–1224. <https://doi.org/10.1016/j.febslet.2015.04.009>
- Kuroiwa T (1982) Mitochondrial nuclei. *Int Rev Cytol* 75:1–59
- Martin W, Rujan T et al (2002) Evolutionary analysis of *Arabidopsis*, cyanobacterial, and chloroplast genomes reveals plastid phylogeny and thousands of cyanobacterial genes in the nucleus. *Proc Natl Acad Sci U S A* 99:12246–11251. <https://doi.org/10.1073/pnas.182432999>
- Maruyama S, Kuroiwa H et al (2007) Centromere dynamics in the primitive red alga *Cyanidioschyzon merolae*. *Plant J* 49:1122–1129. <https://doi.org/10.1111/j.1365-313X.2006.03024.x>
- Maruyama S, Matsuzaki M et al (2008) Centromere structures highlighted by the 100%-complete *Cyanidioschyzon merolae* genome. *Plant Signal Behav* 3:140–141. <https://doi.org/10.4161/psb.3.2.5066>
- Matsuzaki M, Misumi O et al (2004) Genome sequence of the ultrasmall unicellular red alga *Cyanidioschyzon merolae* 10D. *Nature* 428:653–657. <https://doi.org/10.1038/nature02398>
- Minoda A, Sakagami R et al (2004) Improvement of culture conditions and evidence for nuclear transformation by homologous recombination in a red alga, *Cyanidioschyzon merolae* 10D. *Plant Cell Physiol* 45:667–671. <https://doi.org/10.1093/pcp/pch087>

- Minoda A, Nagasawa K et al (2005) Microarray profiling of plastid gene expression in a unicellular red alga, *Cyanidioschyzon merolae*. *Plant Mol Biol* 59:375–385. <https://doi.org/10.1007/s11103-005-0182-1>
- Misumi O, Matsuzaki M et al (2005) *Cyanidioschyzon merolae* genome. A tool for facilitating comparable studies on organelle biogenesis in photosynthetic eukaryotes. *Plant Physiol* 137:567–585. <https://doi.org/10.1104/pp.104.053991>
- Nozaki H, Ohta N et al (2003) Phylogeny of plastids based on cladistic analysis of gene loss inferred from complete plastid genome sequences. *J Mol Evol* 57:377–382. <https://doi.org/10.1007/s00239-003-2486-6>
- Nozaki H, Takano H et al (2007) A 100%-complete sequence reveals unusually simple genomic features in the hot-spring red alga *Cyanidioschyzon merolae*. *BMC Biol* 5:28. <https://doi.org/10.1186/1741-7007-5-28>
- Ohta N, Sato N et al (1998) Structure and organization of the mitochondrial genome of the unicellular red alga *Cyanidioschyzon merolae* deduced from the complete nucleotide sequence. *Nucleic Acids Res* 26:5190–5198
- Ohta N, Matsuzaki M et al (2003) Complete sequence and analysis of the plastid genome of the unicellular red alga *Cyanidioschyzon merolae*. *DNA Res* 10:67–77
- Ohyama K, Fukuzawa H et al (1986) Chloroplast gene organization deduced from complete sequence of liverwort *Marchantia polymorpha* chloroplast DNA. *Nature* 322:572–574
- Schönknecht G, Chen WH et al (2013) Gene transfer from bacteria and archaea facilitated evolution of an extremophilic eukaryote. *Science* 339:1207–1210. <https://doi.org/10.1126/science.1231707>
- Session AM, Uno Y et al (2016) Genome evolution in the allotetraploid frog *Xenopus laevis*. *Nature* 538:336–343. <https://doi.org/10.1038/nature19840>
- Shinozaki K, Ohme M et al (1986) The complete nucleotide sequence of the tobacco chloroplast genome: its gene organization and expression. *EMBO J* 5:2043–2049
- Sumiya N, Fujiwara T et al (2014) Development of a heat-shock inducible gene expression system in the red alga *Cyanidioschyzon merolae*. *PLoS One* 9:e111261. <https://doi.org/10.1371/journal.pone.0111261>
- Tajima N, Sato S et al (2014) Analysis of the complete plastid genome of the unicellular red alga *Porphyridium purpureum*. *J Plant Res* 127:389–397. <https://doi.org/10.1007/s10265-014-0627-1>
- Taki K, Sone T, Kobayashi Y, Watanabe S, Imamura S, Tanaka K (2015) Construction of a URA5.3 deletion strain of the unicellular red alga *Cyanidioschyzon merolae*: a background less host strain for transformation experiments. *J Gen Appl Microbiol* 61:211–214. <https://doi.org/10.2323/jgam.61.211>
- Yang EC, Kim KM et al (2015) Highly conserved mitochondrial genomes among multicellular red algae of the Florideophyceae. *Genome Biol Evol* 7:2394–23406. <https://doi.org/10.1093/gbe/evv147>
- Yoshida Y, Kuroiwa H et al (2010) Chloroplasts divide by contraction of a bundle of nanofilaments consisting of polyglucan. *Science* 329:949–953. <https://doi.org/10.1126/science.1190791>