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# The Fungal Endobiome of Medicinal Plants: A Prospective Source of Bioactive Metabolites

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## Abstract

Fungal endobiome has over the period of time evolved from being defined just as the microbes living within plants indicating not only their location but also the type of association that they have with the host. They are the organisms that live asymptotically within the internal tissues of the plant and exhibit a variety of relationships with their host ranging from symbiotic to pathogenic. They have a very intimate and also a co-evolutionary relationship with their host and therefore have the potential to influence the physiology of the plant. Endophytes from medicinal plants especially represent an important and potential source of bioactive compounds. Endophytic fungi under the influence of multiplexed interactions within its niche, viz., host plant, produce a plethora of secondary metabolites which belong to diverse chemical groups including terpenoids, alkaloids, phenylpropanoids, polyketides, peptides, flavonoids, steroids, lignans, etc. Terpenoids and polyketides are most commonly purified from endophytes, whereas flavonoids and lignans are rare. Due to chemical diversity of their secondary metabolites, endophytic fungi have been explored for medicinal, agricultural, and industrial uses. These are proven useful for novel drug discovery and can be used as potential sources of pharmaceutical leads. These metabolites are known to possess a wide variety of biological activities like antimicrobial, antioxidant, immunomodulatory, anticancerous, antidiabetic, acetylcholinesterase, etc. Endophytes are viewed as an outstanding source for bioprospecting new drugs, and their importance lies in the fact that they represent an ecological group which is less explored and develop in special and sequestered environments. Their diversity and specialized habituation make them an exciting field to search for novel bioactive compounds.

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Fungal endophytes • Endobiome • Bioactive compounds • Medicinal plants • Bioactivity

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**Abbreviations**

AgNPs	Silver nanoparticles
DPPH	1, 1-Diphenyl-2-picrylhydrazyl base pairs
IC <sub>50</sub>	Half maximal inhibitory concentration
MDR	Multidrug resistance
ml	Milliliter
MLR	Mixed lymphocyte reaction assay
sp.	Species
WHO	World Health Organization
α	Alpha
μg	Microgram

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**7.1 Introduction**
**7.1.1 Endobiome**

The plant endobiome consists of various microorganisms residing inside the endosphere, i.e., internal compartments of the plant. Various studies have significantly advanced the understanding of the composition and structure of plant microbiomes

which indicate that abiotic factors, as well as plant–microbe, microbe–microbe, and plant–plant interactions, contribute to plant endobiome composition and structure. The community structure of the endobiome depends on the combination of ability to colonize and the allocation of plant resources. The drivers for which include soil, host plant, and microbes. Factors such as latitude, elevation, temperature, and precipitation can also interact and influence the endobiome composition. The interactions between the endobiome and plant are highly complex and dynamic and can be beneficial (mutualistic), neutral (commensalism), or detrimental (parasitic). Consequently, the plant endobiome dramatically affects plant health and productivity (Turner et al. 2013; Berg et al. 2014; Schlaeppli and Bulgarelli 2015). The plant microbiome is known to induce or prime plant defenses against a broad range of pathogens and insect herbivores. Studies on the plant–microbe interaction involved in endosphere provide an alternative for the manipulations of different biosynthetic pathways responsible for the production of various bioactive and novel molecules of commercial importance. Additionally, the plant endobiome is a crucial player in global biogeochemical cycles, participating significantly in the biochemical cycling of the products of photosynthesis. Therefore, manipulation of plant endobiome is believed to have the potential to interfere with plant disease development, promote plant secondary metabolite production, and ease chemical inputs, leading to more sustainable agricultural practices and enhanced productivity.

Fungal endobiome can precisely be called as the endophytes and is defined functionally by their occurrence within tissues of plants without causing any immediate overt effects (Bacon and White 2000; Hyde and Soyong 2008). Endophytes are ubiquitous and have been found in all the species of plants studied to date. Host–endophyte interactions fall within a continuum ranging from mutualism to commensalism and ultimately pathogenicity. Mutualistic relationship provides benefit to both the partners. Endophytes get nutrition and shelter from the host, while endophyte contributes to the well-being of the host by providing adequate nutrient supply, improved growth, and resistance from herbivores, pathogens, drought, salinity, etc. Colonization of host plants by endophytic fungi can have a profound effect on the plant ecology, fitness, and plant community health (Gopalakrishnan et al. 2015). Enormous biological diversity coupled with their capability to biosynthesize secondary metabolites has provided the impetus for a number of investigations on endophytes. They are particularly interesting due to their easy biological utilization on the large commercial scale and have proven to be a promising source of novel and biologically active natural products, extracellular enzymes, and plant growth-promoting agents of biotechnological interest.

### 7.1.2 Fungal Endophytes

Over the period of time, the research on endophytes has taken a long leap, but the basic definition given by various researchers remains more or less the same, i.e., “Microbes that exist within the living tissues of plants intercellularly or

intracellularly, at least for a part of their life cycle without causing any harm to their host are known as endophytes” (Nisa et al. 2015). Endophytes in Greek means “within plant” (endo = within, phytes = plants). The term endophyte was first coined by De Bary in 1866 (Jain and Pundir 2015). The endophytic microorganisms which constitute the plant endobiome include bacteria, fungi, and actinomycetes which form a range of relationships with their host plant including symbiotic, mutualistic, commensalism, parasitic, etc. (Stepniewska and Kuzniar 2013; Swarnalatha et al. 2015). The infection caused by endophytes within the invading tissues of the host plant is inconspicuous and symptomless unless endophytes become pathogenic under stressful conditions. Some endophytes assume a quiescent state either for the whole lifetime or for an extended period of time. A harmonious symbiotic association generally exists between plant and endophyte in which both of them are benefitted. Most of the mycologists use the term “endophyte” strictly for those fungi that never cause any visible disease symptoms at any specific moment. Endophytes get nutrition and shelter inside the host and in return provide resistance to the concerned host against biotic and abiotic stresses. They can have a profound effect on the plant ecology, fitness, and plant community health. They reside entirely within host tissues and emerge during host senescence unlike mycorrhizal fungi that colonizes plant roots and grow into the rhizosphere.

The importance of endophytes lies in the fact that they represent the ecological group with less explored fungal species. Each plant, in turn, is host to one or more endophytes. As documented by Hawksworth (2001), only 1 lakh fungal species is presently known out of estimated 1.5 million fungal species. The remaining undiscovered fungi may be in the form of hidden endophytes. Endophytes develop in special and sequestered environments and represent a huge diversity of microbial adaptations. Their diversity and specialized habituation make them an exciting field of study in the search for new medicines or novel drugs. Endophytic fungal diversity is supposed to be high in tropics as compared to temperate regions. It is assumed that fungi within tropical tree leaves may be hyperdiverse. Their diversity even varies from one location to another.

Another interesting point in studying endophytes is because of the hope it brings, by synthesizing diverse and novel secondary metabolites. The new and bioactive compounds can provide assistance and relief in all aspects of human conditions like drug resistance, treatment of new emerging diseases, and safe bioactive compounds (Kaul et al. 2012). A list of all approved agents has been prepared from 1981 to 2006 in which microbes/endophytes were found to be the main source of a significant number of natural products (Pimentel et al. 2011). Synthesis of new active secondary metabolites by the endophytes may be induced by metabolic interaction between the host plant and endophytes.

The discovery of paclitaxel-producing endophytic fungus *Taxomyces andreanae* from *Taxus brevifolia* (yew trees) increased the importance of endophytes and created an impetus among the researchers for a more comprehensive and elaborative examination in this area (Selim et al. 2012). The presence of a microbial source of

the drug could eliminate the need to harvest and extract the slow growing and relatively rare yew trees, and the price for the drug would also be reduced. The drug would be available to cancer patients, since paclitaxel (taxol) could be produced via fermentation in the same way that penicillin is fermented (Strobel 2003). Fermentation of endophytic fungus producing bioactive compounds has several advantages like reproducible and dependable productivity. It can be grown in fermenters to provide an inexhaustible supply of bioactive compound and thus can be exploited commercially. Direct changes in the culture conditions can be explored as a method of optimizing various biosynthetic pathways that lead to the production of derivatives and analogs of novel compounds (Strobel et al. 2004). Later in the years to follow, taxol has been detected in numerous other endophytic fungi, isolated from diverse host plants. In addition to taxol, other medicinally important plant compounds have also been produced by endophytic fungi (Kaul et al. 2012).

There are many assumptions that endophytes uptake plant DNA into their own genome during their long co-evolution with host plants. This adaptation and genetic variation could have led to endophytes with the ability to synthesize phytochemicals originally associated with the host plant. Horizontal gene transfer makes the endophyte capable of producing associated plant compounds. For example, an endophyte *Fusarium solani* could produce precursors of camptothecin. However, in order to synthesize camptothecin, endophyte uses host's strictosidine synthase, an enzyme involved in the synthesis of camptothecin (Kusari et al. 2012). Initially, it was thought and hypothesized that the endophytic fungi do not synthesize paclitaxel independently but derived it from the host and later accumulates it in their cell walls (Heinig et al. 2013). On the contrary, in a recent study, a large set of potential genes involved in paclitaxel synthesis have been reported in *Penicillium aurantiogriseum* NRRL 62421, an endophyte of *Corylus avellana*. This study has however ruled out the possibility of horizontal gene transfer between endophytic fungus and host plant (Yang et al. 2014a).

Host–endophyte association is very important and influences the synthetic ability of endophytes. Under stressful conditions, endophytes affect the host plant in producing defense chemicals against invading pathogens. During this hostile environment, the ability to synthesize diverse and novel bioactive compounds by endophytes also increases. Thus, environment affects host plant which in turn affects endophyte to change its metabolite profile, increasing synthetic ability and overall biological activity of its secondary metabolites (Selim et al. 2012).

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## 7.2 Fungal Endophytes from Medicinal Plants

Since time immemorial, medicinal plants have been used as a source of medicine. They produce unique and divergent secondary metabolites and harbor a distinctive microbiome (Qi et al. 2012). The endophytes that live inside the plant have distinct but similar metabolic pathway for the production of secondary metabolites. Inducing

factors from both plants and endophytes affect the accumulation of secondary metabolites. Researchers around the globe have been prompted to explore the medicinal plants for isolation of endophytes. This is because of the importance of secondary metabolites from medicinal plants in pharmaceuticals and their influence on the synthetic ability of endophytes. Endophytes are also known to mimic the bioactive compounds as produced by the plant itself. Therefore, it is significant to bioprospect endophytes from medicinal plants which have been used for centuries as a source of important bioactive compounds. Endophytes isolated from medicinal plants may result in the inexhaustible and cost-effective production of desired compounds and therefore help to conserve the biodiversity. Endophytes from medicinal plants are believed to be the source of unique and novel compounds associated with diverse biological activities (Jain and Pundir 2015). Their capability to biosynthesize bioactive secondary metabolites has provided the impetus on bioprospection of endophytes.

Different workers have investigated and documented the isolation of endophytic fungi and their metabolites from Indian medicinal plants. Eight medicinal plants of Western Ghats of India were sampled for endophyte isolation. Fifteen species were recovered, out of which *Alternaria* sp., *Nigrospora oryzae*, and *Papulospora* sp. showed antimicrobial activity (Raviraja et al. 2006). Similarly, medicinal plants of Jammu and Kashmir, namely, *Digitalis lanata*, *Digitalis purpurea*, *Plantago ovata*, *Dioscorea bulbifera*, and *Crocus sativus*, have been sampled for the isolation of endophytes (Ahmed et al. 2012; Sharma et al. 2015). Furthermore, 30 species of endophytic *Pestalotiopsis* sp. have been isolated from four medicinal plants *Terminalia arjuna*, *T. chebula*, *Azadirachta indica*, and *Holarrhena antidysenterica* (Tejesvi et al. 2007). Likewise, endophytic fungi from *Garcinia atroviridis*, *G. dulcis*, *G. mangostana*, *G. nigrolineata*, and *G. scortechinii* have been documented as the potential source of antimicrobial agents (Phongpaichit et al. 2006). Fungal endophytes isolated from indigenous medicinal plants belonging to North Maharashtra region of India have been investigated for antimicrobial activity. The isolates from roots of *Aloe vera* possess strong antibacterial activity (Jalgaonwala et al. 2010).

Chinese traditional medicinal plants have also been immensely explored for isolation of endophytes. Li et al. (2005) have studied 12 Chinese medicinal plants for fungal endophyte isolation. One hundred thirty endophytic fungi were reported, out of which 9.2% of the isolates exhibited antitumor activity and 30% exhibited antifungal activity. Similarly, Huang et al. (2007) have reported 292 morphologically distinct endophytic fungi from 29 Chinese medicinal plants. The endophytes recovered from *Ginkgo biloba* have shown different bioactivities like antimicrobial, antioxidant, cytotoxic, etc. (Li et al. 2014c; Ye et al. 2013; Yuan et al. 2013). Bioprospecting of fungal endophytes from different medicinal plants for the period 2010–2016 has been tabulated (Table 7.1).

**Table 7.1** Fungal endophytes from medicinal plants showing different bioactivities (Period: 2012–2016)

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	References
1.	<i>Hugonia mystax</i>	<i>Aspergillus</i> sp.	Antimicrobial	Abirami and Boominathan (2016)
2.	<i>Corchorus olitorius</i>	<i>Aspergillus terreus</i>	Extracellular enzymes	Ahmed et al. (2016a)
3.	Marine habitat	<i>Aspergillus</i> sp.	L-asparaginase	Ahmed et al. (2016b)
4.	<i>Sapium ellipticum</i>	<i>Chaetomium</i> sp.	Antimicrobial	Akone et al. (2016)
5.	<i>Cymbopogon caesius</i>	<i>Curvularia lunata</i>	Antimicrobial	Avinash et al. (2016)
6.	<i>Glycyrrhiza glabra</i>	<i>Phoma</i> sp.	Antimicrobial	Arora et al. (2016)
7.	<i>Catharanthus roseus</i>	<i>Alternaria alternata</i>	Acetylcholinesterase inhibitory	Bhagat et al. (2016)
8.	<i>Cupressus torulosa</i>	<i>Penicillium oxalicum</i>	Antidiabetic and antimicrobial	Bisht et al. (2016)
9.	<i>Acanthospermum australe</i>	<i>Aspergillus calidoustus</i>	Antimicrobial	Carvalho et al. (2016)
10.	<i>Sommeratia ovata</i>	<i>Nectria</i> sp.	Antidiabetic	Cui et al. (2016)
11.	<i>Nymphaea nouchali</i>	<i>Chaetomium globosum</i>	Antimicrobial	Dissanayake et al. (2016)
12.	<i>Eichhornia crassipes</i>	<i>Aspergillus austroafricanus</i>	Antimicrobial	Ebrahim et al. (2016)
13.	<i>Hydrastis canadensis</i>	<i>Alternaria</i> sp., <i>Colletotrichum fioriniae</i> , <i>Diaporthe eres</i> , <i>Diaporthe</i> sp., <i>Phoma</i> sp., and <i>Pyrenochaeta cava</i>	Antimicrobial	Egan et al. (2016)
14.	<i>Tamarix nilotica</i>	<i>Aspergillus sydowii</i> , <i>Penicillium chrysogenum</i> , and <i>Eupenicillium crustaceum</i>	Not reported	Gashgari et al. (2016)
15.	<i>Juniperus procera</i>	<i>Aspergillus fumigatus</i> <i>Hypocrea lutea</i> <i>Penicillium oxalicum</i> and <i>Preussia</i> sp.	Antimicrobial	Gherbawy and Elhariry (2016)
16.	<i>Pteris pellucida</i>	<i>Emericella quadrilineata</i>	Antimicrobial	Goutam et al. (2016)
17.	<i>Silybum marianum</i>	<i>Talaromyces minioluteus</i>	Antimicrobial	Kaur et al. (2016)

(continued)

Table 7.1 (continued)

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	References
18.	<i>Calotropis procera</i> , <i>Catharanthus roseus</i> , <i>Euphorbia prostrata</i> , <i>Trigonella foenum-graecum</i> , and <i>Vernonia amygdalina</i>	<i>Alternaria</i> sp.	Antimicrobial	Khiralla et al. (2016)
19.	<i>Mentha viridis</i>	<i>Fusarium oxysporum</i>	Antimicrobial	Kumar et al. (2016)
20.	<i>Nicotiana tabacum</i>	<i>Rhizopycnis vagum</i>	Antimicrobial	Lai et al. (2016)
21.	<i>Panax notoginseng</i>	<i>Chaetomium globosum</i>	Acetylcholinesterase inhibitory	Lia et al. (2016)
22.	<i>Mahonia fortunei</i>	<i>Fusarium decemcellulare</i>	Antimicrobial	Li et al. (2016a, b)
23.	<i>Cephalotaxus hainanensis</i>	<i>Diaporthe</i> sp., <i>Phomopsis</i> sp., <i>Colletotrichum</i> sp., <i>Colletotrichum</i> sp., <i>Corynespora</i> sp., <i>Penicillium</i> sp. and <i>Nemania</i> sp.	Antimicrobial	Liu et al. (2016)
24.	<i>Salvia miltiorrhiza</i>	<i>Alternaria</i> sp.	Antimicrobial	Lou et al. (2016)
25.	<i>Schima wallichii</i>	<i>Penicillium simplicissimum</i> and <i>Talaromyces verruculosus</i>	Antimicrobial	Mishra et al. (2016b)
26.	<i>Rhizophora annamalayana</i>	<i>Trichoderma</i> sp.	Antimicrobial	Narendran and Kathiresan (2016)
27.	<i>Pisonia grandis</i>	<i>Aspergillus niger</i> , <i>Aspergillus fumigates</i> , and <i>Aspergillus japonicus</i>	Antimicrobial	Nishanthi et al. (2016)
28.	<i>Garcinia preussii</i>	<i>Penicillium</i> sp.	Antimicrobial	Jouda et al. (2016a)
29.	<i>Ficus carica</i>	<i>Aspergillus tamarii</i>	Antimicrobial	Ma et al. (2016)
30.	<i>Cinnamomum iners</i> , <i>Shorea siamensis</i> , <i>Fernandoa adenophylla</i> , <i>Quercus semiserrata</i>	<i>Xylaria</i> sp.	Antimicrobial	Orachaipunlap et al. (2016)
31.	<i>Cinnamomum malabarrum</i>	<i>Colletotrichum gloeosporioides</i>	Antimicrobial	Packiaraj et al. (2016)



32.	<i>Houttuynia cordata</i>	<i>Chaetomium globosum</i>	Antimicrobial	Pan et al. (2016)
33.	<i>Moringa oleifera</i>	<i>Aspergillus flavus</i>	Antimicrobial	Rajeshwari et al. (2016)
34.	<i>Cupressus torulosa</i>	<i>Alternaria alternata</i>	Extracellular enzyme	Rajput et al. (2016)
35.	<i>Hypocrea virens</i>	<i>Premna serratifolia</i>	Antimicrobial	Ratnaweera et al. (2016)
36.	<i>Rauwolfia serpentina</i>	<i>Colletotrichum</i> sp. <i>Fusarium</i> sp. <i>Cladosporium</i> sp. <i>Aspergillus fumigatus</i> , <i>Aspergillus niger</i> , <i>Aspergillus terreus</i> var. <i>africanus</i> , <i>Cladosporium cucumerinum</i> , <i>Cladosporium</i> <i>oxysporum</i> , <i>Penicillium aurantigriseum</i> , and <i>Penicillium chrysogenum</i>	Antimicrobial	El-Said et al. (2016)
38.	<i>Acalypha indica</i>	<i>Phoma</i> sp.	Antimicrobial	Sowparthani (2016)
39.	<i>Santalum album</i>	<i>Fusarium oxysporum</i> , <i>Fusarium solani</i> , <i>Histoplasma</i> sp., <i>Periconia</i> sp. and <i>Pestalotiopsis</i> sp.	Antimicrobial	Tapwal et al. (2016)
40.	<i>Picea maritima</i> and <i>Picea rubens</i>	<i>Diaporthe maritime</i>	Antimicrobial	Tanney et al. (2016)
41.	<i>Narcissus tazetta</i>	<i>Fusarium solani</i>	Antimicrobial	Wang et al. (2015)
42.	<i>Huperzia serrata</i>	<i>Colletotrichum</i> sp., <i>Ascomycoia</i> sp.	Acetylcholinesterase inhibitory	Wang et al. (2016b)
43.	<i>Eugenia jambolana</i>	<i>Aspergillus niger</i> and <i>Aspergillus terreus</i>	Antimicrobial	Yadav et al. (2016)
44.	<i>Lonicera japonica</i>	<i>Fusarium</i> sp.	Antimicrobial	Zhang et al. (2016b)
45.	<i>Edgeworthia chrysantha</i>	<i>Fusarium oxysporum</i>	Antimicrobial	Zhang et al. (2016a)
46.	<i>Sapium ellipticum</i>	<i>Penicillium tropicum</i>	Antimicrobial	Zeng et al. (2016)

(continued)

Table 7.1 (continued)

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	References
47.	<i>Panax notoginseng</i>	<i>Acremonium</i> sp. <i>Alternaria</i> sp. <i>Arthrinium</i> sp. <i>Aspergillus</i> sp. <i>Botryotinia</i> sp. <i>Chaetomium</i> sp. <i>Cladosporium</i> sp. <i>Colletotrichum</i> sp. <i>Dictyosporium</i> sp. <i>Fusarium</i> sp. <i>Humicola</i> sp. <i>Ilyonectria</i> sp. <i>Mucor</i> sp. <i>Myrothecium</i> sp. <i>Penicillium</i> sp. <i>Periconia</i> sp. <i>Pestalotiopsis</i> sp. <i>Phialophora</i> sp. <i>Phoma</i> sp. <i>Phomopsis</i> sp. <i>Plectosphaerella</i> sp. <i>Thielavia</i> sp. and <i>Trichoderma</i> sp.	Antimicrobial	Zheng et al. (2016)
48.	<i>Centaurea stoebe</i>	<i>Trichoderma</i> sp.	Antifungal, Cytotoxic	Abdou and Abdelhady (2015)

49.	<i>Rhododendron anthopogon</i>	<i>Stemphylium</i> sp.	Antimicrobial	Baral et al. (2015)
		<i>Alternaria</i> sp.		
		<i>Penicillium</i> sp.		
		<i>Aspergillus</i> sp.		
		<i>Trichoderma</i> sp.		
		<i>Papulaspora</i> sp.		
		<i>Hansfordia</i> sp.		
		<i>Wardomyces</i> sp. and <i>Geotrichum</i> sp.		
		<i>Acromonium curvulum</i>		
50.	<i>Bauhinia forficata</i>	<i>Aspergillus ochraceus</i>	Antibacterial, Enzymatic	Bezerra et al. (2015)
		<i>Gibberella fujikuroi</i>		
		<i>Myrothecium verrucaria</i> and <i>Trichoderma piluliferum</i>		
51.	<i>Curcuma longa</i>	44 endophytic fungal isolates	Antioxidant	Bustanussalam et al. (2015)
52.	<i>Amona crassiflora</i>	<i>Rhizoctonia</i> sp.	Antibacterial	De Mendonca et al. (2015)
53.	<i>Carapa guianensis</i>	35 distinct fungal taxa	Antibacterial	Ferreira et al. (2015)
54.	<i>Mallotus philippensis</i>	<i>Alternaria</i> sp., <i>Pestalotiopsis</i> sp. and <i>Phomopsis</i> sp.	Antimicrobial	Gangwar et al. (2015)
55.	<i>Curcuma xanthorrhiza</i>	<i>Xylaria</i> sp.	Cytotoxic	Hammerschmidt et al. (2015)
56.	<i>Kadsura angustifolia</i>	42 fungal taxa	Extracellular enzymatic	Huang et al. (2015)
57.	<i>Azadirachta indica</i>	<i>Chaetomium</i> sp.	Antioxidant	Kumaresan et al. (2015)
		<i>Colletotrichum</i> sp.		
58.	<i>Rauwolfia serpentina</i>	<i>Curvularia</i> sp. and <i>Trichoderma</i> sp.	Antibacterial	Nath et al. (2015)
		<i>Aspergillus awamori</i>		
		<i>Penicillium</i> sp. and <i>Colletotrichum gloeosporioides</i>		

(continued)

Table 7.1 (continued)

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	References
59.	<i>Brucea javanica</i>	<i>Trichoderma</i> sp. <i>Fusarium</i> sp. <i>Aspergillus</i> sp. and <i>Penicillium</i> sp.	Not reported	Nur and Muh Danial (2015)
60.	<i>Solanum xanthocarpum</i>	<i>Phomopsis vexans</i>	Lowering blood cholesterol (lovastatin)	Parthasarathya and Sathiyabama (2015)
61.	<i>Aegle marmelos</i>	<i>Aspergillus flavus</i>	Antioxidant Antimicrobial	Patil et al. (2015b)
62.	<i>Silybum marianum</i>	25 fungal taxa	Cytotoxic	Raja et al. (2015)
63.	<i>Cyperus rotundus</i>	<i>Rhizoctonia solani</i>	Antibacterial	Ratnaweera et al. (2015a)
64.	<i>Indigofera stueffertiana</i>	<i>Nigrospora sphaerica</i> and <i>Pestalotiopsis maculans</i>	Antibacterial	Santos et al. (2015)
65.	<i>Withania somnifera</i>	<i>Fusarium</i> sp.	Antibacterial	Singh et al. (2015a)
66.	<i>Limonium acidissima</i>	<i>Aspergillus</i> sp.	Cytotoxic	Siriwardane et al. (2015)
67.	<i>Huperzia serrata</i>	<i>Paecilomyces tenuis</i>	Anti-Alzheimer's	Su and Yang (2015)
68.	<i>Bacopa monnieri</i>	<i>Aspergillus fumigatus</i>	Antioxidant and antitubercular	Thakur et al. (2015)
69.	<i>Astonia boonei-Ahun</i> , <i>Enantia chlorantha-Awopa</i> , and <i>Kigelia africana-Pandoro</i>	<i>Aspergillus niger</i> <i>Macrophomina</i> sp. <i>Trichoderma</i> sp. and four different <i>Penicillium</i> sp.	Antibacterial	Tolulope et al. (2015)
70.	<i>Dracaena draco</i>	<i>Botryodiplodia theobromae</i>	Antibacterial	Zaher et al. (2015a, b)
71.	<i>Calotropis procera</i>	<i>Aspergillus niger</i> <i>Cladosporium herbarum</i> , <i>Aspergillus tamari</i> , <i>Drechslera nodulosa</i> , <i>Fusarium solani</i> <i>Aspergillus japonicus</i> , <i>Alternaria alternata</i> , <i>Alternaria tenuissima</i> , <i>Curvularia pallenscens</i> , and <i>Curvularia lunata</i>	Antibacterial	Aharwal et al. (2014)
72.	<i>Tabebuia argentea</i>	<i>Aspergillus niger</i>	Anticancer	Channabasava and Govindappa (2014)

	Three endophytic strains	Enzymatic, phytochemical screening	Desire et al. (2014)
73.	<i>Lantana camara</i>		
74.	<i>Capsicum annuum</i>	<i>Alternaria alternata</i>	Devart et al. (2014)
75.	<i>Gloriosa superba</i>	<i>Alternaria solani</i> and <i>Penicillium funiculosum</i>	Devi et al. (2014)
76.	<i>Tabebuia argentea</i>	<i>Alternaria alternata</i>	Govindappa et al. (2014)
77.	<i>Garcinia nobilis</i>	<i>Penicillium</i> sp.	Jouda et al. (2014)
78.	<i>Bacopa monnieri</i>	26 endophytes	Katoch et al. (2014a, b)
79.	<i>Xanthium sibiricum</i>	<i>Eupenicillium</i> sp.	Li et al. (2014a)
80.	<i>Ginkgo biloba</i>	<i>Chaetomium globosum</i>	Li et al. (2014c)
81.	<i>Datura stramonium</i>	<i>Aspergillus</i> sp.	Mahdi et al. (2014)
		<i>Curvularia</i> sp.	
	<i>Moringa oleifera</i> and <i>Prosopis chilensis</i>	<i>Emericella</i> sp. and <i>Chaetomium</i> sp.	
82.	<i>Terminalia arjuna</i>	<i>Aspergillus flavus</i> , <i>Diaporthe arengae</i> <i>Alternaria</i> sp. and <i>Lasioidiplodia theobromae</i>	Patil et al. (2014)
83.	<i>Polygala elongata</i>	<i>Colletotrichum</i> sp.	
84.	<i>Nothapodytes foetida</i> and <i>Hypericum mysorense</i>	<i>Bionectria ochroleuca</i> and <i>Chaetomium globosum</i>	Pawle and Singh (2014) Samaga and Rai (2014)
85.	<i>Camellia sinensis</i>	<i>Colletotrichum</i> sp. and <i>Gloeosporioides</i> sp.	Rabha et al. (2014)
86.	<i>Cynodon dactylon</i> and <i>Dactyloctenium aegyptium</i>	26 fungal endophytes 30 fungal endophytes	Rekha and Shivanna (2014)
87.	<i>Saraca indica</i>	<i>Phomopsis</i> sp. <i>Aspergillus terreus</i> <i>Phialophora</i> sp. <i>Alternaria alternata</i> and <i>Phyllosticta</i> sp.	Sandhu et al. (2014)

(continued)

Table 7.1 (continued)

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	References
88.	<i>Cinnamomum mollissimum</i>	<i>Phoma</i> sp.	Antibacterial, Antifungal, Cytotoxic	Santiago et al. (2014)
89.	<i>Adiantum capillus-veneris</i>	<i>Chaetomium globosum</i>	Cytotoxic Antioxidant Butyrylcholinesterase inhibitory	Selim et al. 2014
90.	<i>Allium sativum</i>	<i>Trichoderma brevicompactum</i>	Antifungal	Shentu et al. (2014)
91.	<i>Curcuma longa</i>	<i>Penicillium</i> sp.	Antibacterial	Singh et al. (2014)
92.	<i>Phyllanthus amarus</i>	30 endophytic fungi	Antifungal, Anticancer, Anti-metastatic	Taware et al. (2014)
93.	<i>Crotalaria pallida</i>	<i>Alternaria</i> sp.	Antioxidant	Umashankar et al. (2014)
94.	<i>Madhuca indica</i>	<i>Penicillium</i> sp. and <i>Aspergillus flavus</i> 40 taxa Dominant: <i>Phomopsis</i> sp.	Antimicrobial Antibacterial	Verma et al. (2014)
95.	<i>Baccharis trimera</i>	<i>Colletotrichum</i> sp. and <i>Gloeosporioides</i> sp. <i>Diaporthe phaseolorum</i> <i>Pestalotiopsis</i> sp. and <i>Preussia pseudominima</i> 14 fungal species Dominant: <i>Colletotrichum falcatum</i>	Antimicrobial	Vieira et al. (2014)
96.	<i>Boswellia ovalifoliolata</i>		Not reported	Anitha et al. (2013)
97.	<i>Pterocarpus Santalinus</i> <i>Shorea thumbugaia</i> and <i>Syzygium alternifolium</i>			
98.	<i>Bauhinia guianensis</i>	<i>Aspergillus</i> sp.	Antimicrobial	Pinheiro et al. (2013)
99.	<i>Pandanus amaryllifolius</i> <i>Camptotheca acuminata</i>	<i>Colletotrichum</i> sp. <i>Botryosphaeria</i> sp. and <i>Fusarium</i> sp.	Antibacterial Antimicrobial	Bunghan et al. (2013) Ding et al. (2013)

100.	<i>Moringa oleifera</i>	<i>Alternaria</i> sp. <i>Aspergillus</i> sp. <i>Bipolaris</i> sp. <i>Exophiala</i> sp. <i>Nigrospora</i> sp. and <i>Penicillium</i> sp. <i>Penicillium citrinum</i> <i>Aspergillus niger</i> <i>Aspergillus flavus</i> <i>Penicillium</i> sp. <i>Rhizopus</i> sp. and <i>Fusarium</i> sp. <i>Cladosporium</i> sp. <i>Aspergillus flavus</i> <i>Aspergillus</i> sp. and <i>Curvularia lunata</i> <i>Aspergillus</i> sp. <i>Guignardia</i> sp. <i>Fusarium</i> sp. <i>Penicillium</i> sp. <i>Pestalotiopsis</i> sp. and <i>Trichoderma</i> sp. <i>Chaetomium globosum</i> <i>Colletotrichum truncatum</i> <i>Nigrospora oryzae</i> <i>Fusarium proliferatum</i> <i>Guignardia cammilleae</i> <i>Alternaria destruens</i> and <i>Chaetomium</i> sp. 30 fungal species Dominant: <i>Penicillium copicicola</i> <i>Penicillium citrinum</i>	Not reported	Dhanalakshmi et al. (2013)
101.	<i>Ceratonia siliqua</i>		Cytotoxic	El-Neketi et al. (2013)
102.	<i>Tabebuia argentea</i>		Antioxidant	Govindappa et al. (2013)
103.	<i>Kigelia africana</i>		Antibacterial	Idris et al. (2013)
104.	<i>Anidesma madagascariense</i>		Not reported	Jeewon et al. (2013)
105.	<i>Withania somnifera</i>		Antifungal	Kumar et al. (2013)
106.	<i>Jatropha curcas</i>		Antifungal	Kumar and Kaushik (2013)
107.	<i>Cannabis sativa</i>		Antifungal	Kusari et al. (2013)
108.	<i>Ocimum tenuiflorum</i>		Antibacterial	Lai et al. (2013)

(continued)

Table 7.1 (continued)

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	References
109.	<i>Zingiber zerumbet</i>	<i>Fusarium oxysporum</i>	Antioxidant	Nongalleima et al. (2013)
110.	<i>Erythrina variegata</i>	<i>Alternaria</i> sp.	Antiangiogenic	Pompeng et al. (2013)
111.	<i>Garcinia</i> sp.	<i>Aspergillus fumigates</i> and <i>Fusarium</i> sp.	Antimicrobial	Ruma et al. (2013)
			Antioxidant,	
			Anti-inflammatory	
112.	<i>Viscum album</i>	<i>Aspergillus flavus</i>	Antioxidant	Sadananda et al. (2013)
		<i>Fusarium oxysporum</i>		
		<i>Fusarium moniliforme</i> and <i>Trichothecium</i> sp.		
113.	<i>Ocimum sanctum</i>	147 fungal endophytes	Antioxidant	Sharma and Kumar (2013)
114.	<i>Elaeis guineensis</i>	<i>Trichoderma</i> sp.	Antifungal	Sundram (2013)
115.	<i>Rhododendron tomentosum</i>	<i>Fusarium tricinatum</i>	Antibacterial	Tejesvi et al. (2013)
116.	<i>Glycine max</i>	<i>Alternaria alternata</i>	Not reported	Tenguria and Firodiya (2013)
		<i>Phoma</i> sp.		
		<i>Penicillium</i> sp. and <i>Fusarium</i> sp.		
117.	<i>Sauracia scaberrinae</i>	<i>Phoma</i> sp.	Antibacterial	Wijeratne et al. (2013)
118.	<i>Panax ginseng</i>	<i>Nectria</i> sp.	Antibacterial	Wu et al. (2013b)
		<i>Aspergillus</i> sp.		
		<i>Fusarium</i> sp.		
		<i>Verticillium</i> sp.		
		<i>Engyodontium</i> sp.		
		<i>Plectosphaerella</i> sp.		
		<i>Penicillium</i> sp. and <i>Cladosporium</i> sp.		
119.	<i>Ginkgo biloba</i>	<i>Chaetomium globosum</i>	Antioxidant	Ye et al. (2013)
120.	<i>Huperzia serrata</i>	<i>Ceriporia lacerate</i>	Cytotoxic (ceriponols A–K)	Ying et al. (2013)
121.	<i>Ginkgo biloba</i>	<i>Penicillium</i> sp.	Antioxidant	Yuan et al. (2013)
122.	<i>Taraxacum mongolicum</i>	<i>Phoma</i> sp.	Antibacterial	Zhang et al. (2013a)



123.	<i>Ammonia muricata</i>	<i>Periconia</i> sp.	Cytotoxic (periconiasins A-C)	Zhang et al. (2013b)
124.	<i>Terminalia brownii</i>	<i>Rhizophorus oryzae</i> <i>Aspergillus niger</i> and <i>Aspergillus flavus</i>	Antimicrobial	Basha et al. (2012)
125.	<i>Ocimum sanctum</i> and <i>Sapindus detergens</i>	63 endophytic fungal Isolates	Antibacterial Anticancer	Bhagat et al. (2012)
126.	<i>Nyctanthes arbor-tristis</i>	19 endophytic fungi Dominant: <i>Alternaria alternata</i> and <i>Cladosporium cladosporioides</i>	Antimicrobial	Gond et al. (2012)
127.	<i>Sapindus saponaria</i>	<i>Cochliobolus intermedius</i> and <i>Phomopsis</i> sp.	Antimicrobial	Garcia et al. (2012)
128.	<i>Cinnamomum camphora</i>	20 fungal species	Antimicrobial	Kharwar et al. (2012)
129.	<i>Dysoxylum binectariferum</i>	<i>Fusarium proliferatum</i>	Anticancer (rohitukine)	Kumara et al. (2012)
130.	<i>Ophiopogon japonicus</i>	30 fungal strains	Antimicrobial	Liang et al. (2012)
131.	<i>Embllica officinalis</i>	<i>Phomopsis</i> sp.	Antioxidant Antimicrobial	Nath et al. (2012)
132.	<i>Piper hispidum</i>	21 isolates belonging to 11 genera	Not reported	Orlandelli and Pamphile (2012)
133.	<i>Trichilia elegans</i>	<i>Cordyceps memorabilis</i> <i>Phomopsis longicolla</i> and <i>Dothideomycetes</i> sp.	Antimicrobial	Rhoden et al. (2012)
134.	<i>Arisaema erubescens</i>	<i>Phoma</i> sp.	Antimicrobial Antitumor	Wang et al. (2012a)
135.	<i>Curcuma wenyujin</i>	<i>Chaetomium globosum</i>	Antifungal Cytotoxic	Wang et al. (2012b)
136.	<i>Moringa oleifera</i>	<i>Nigrospora</i> sp.	Antifungal	Zhao et al. (2012a, b)

## 7.3 Fungal Endobiome as a Source of Bioactive Metabolites

Endophytic fungi exist within a niche where it communicates with diverse communities of microorganisms. Variegated cross talks take place among endophytic fungi, endophytes and host, endophytic fungi and endophytic bacteria, etc. Under the influence of such multiplexed interactions and environmental conditions, a plethora of secondary metabolites is synthesized by the fungal endophytes (Kusari et al. 2014). Secondary metabolites are defined as small molecules that are not necessary for normal growth or development. Although it is not possible to reproduce such an array of diverse metabolites by endophytes under in vitro conditions, however, it is interesting that using controlled fermentation condition, by altering the accessible culture and process parameters (media composition, aeration, pH, incubation period, shaking conditions, inoculum size, etc.), the endophyte can be optimized for the production of surplus biologically active secondary metabolites (Kusari et al. 2012).

Secondary metabolites from endophytes have a tremendous impact on the society and proven useful for novel drug discovery and can be used as a potential source of pharmaceutical leads. These belong to diverse chemical groups including terpenoids, alkaloids, phenylpropanoids, aliphatic, polyketides, peptides, flavonoids, steroids, lignans, etc. Terpenoids and polyketides are most commonly purified from endophytes, whereas flavonoids and lignans are rare (Mousa and Raizada 2013). Due to chemical diversity of their secondary metabolites, endophytic fungi have been explored for medicinal, agricultural, and industrial uses. These metabolites are known for a wide variety of biological activities like antimicrobial, antioxidant, immunomodulatory, anticancerous, antidiabetic, antiviral, etc.

Some of the important categories of bioactive secondary metabolites produced by fungal endophytes of medicinal plants are as follows.

### 7.3.1 Anticancer Compounds

Cancer is a killer disease affecting more than six million people every year. It is characterized by unregulated cell proliferation. Due to uncontrollable growth of cells, an abnormal mass of tissue is formed which is generally called as a tumor. Antitumor agents are the compounds that are capable of counteracting the formation of malignant. Plant-based compounds have played an important role in the development of several clinically useful anticancer drugs like taxol, vinblastine, vincristine, topotecan, and etoposide (Nirmala et al. 2011). Despite this, there is a need to explore alternative source with more diversity and novelty. Endophytes with their unique secondary metabolites provide tremendous diversity. Active metabolites isolated from endophytes provide anticancer action with minimum side effects. These compounds could be an alternative approach for discovery of novel anticancer drugs (Kharwar et al. 2011; Kaul et al. 2012; Chen et al. 2014). Various anticancer compounds have been reported from fungal endophytes. For the sake of convenience, some of the anticancer compounds from fungal endophytes have been tabulated (Table 7.2).

**Table 7.2** Anticancer compounds from endophytic fungi of medicinal plants (2012–2016)

S. no.	Medicinal plants	Fungal endophytes	Anticancer compound	Reference
1.	<i>Piper hispidum</i>	<i>Diaporthe</i> sp.	(1 → 3,1 → 6)-D-glucans	Orlandelli et al. (2017)
2.	Medicinal plant	<i>Bipolaris setariae</i>	Ophiobolin A	Bhatia et al. (2016)
3.	<i>Acanthospermum australe</i>	<i>Aspergillus calidoustus</i>	Ophiobolin K and 6-epiophiobolin K	Carvalho et al. (2016)
4.	<i>Uncaria rhynchophylla</i>	<i>Colletotrichum gloeosporioides</i>	Colletotriactam A–D	Wei et al. (2016)
5.	<i>Acanthus ilicifolius</i>	<i>Aspergillus flavipes</i>	Meroterpenoids (guignardones)	Bai et al. (2015)
6.	<i>Hevea brasiliensis</i>	<i>Eutypella scoparia</i>	Cytochalasins	Kongprapan et al. (2015)
7.	<i>Diphyletia sinensis</i>	<i>Aspergillus fumigates</i>	Fumitremorgin and fumitremorgin D	Liang et al. (2015b)
8.	<i>Sinopodophyllum emodi</i>	<i>Alternaria tenuissima</i>	Podophyllotoxin	Liang et al. (2015a)
9.	Mangrove plant	<i>Lasiodiplodia</i> sp.	Lasiodiplodins	Li et al. (2015b)
10.	<i>Paris polyphylla</i> var. <i>yunnanensis</i>	<i>Aspergillus versicolor</i>	Versicolols A and B	Zhou et al. 2015
11.	<i>Tabebuia argentea</i>	<i>Aspergillus niger</i>	Lapachol	Channabasava and Govindappa (2014)
12.	<i>Tripterygium wilfordii</i>	<i>Penicillium</i> sp.	Penifupyrone	Chen et al. 2014
13.	<i>Ginkgo biloba</i>	<i>Chaetomium globosum</i>	Chaetoglobosins A, G, V, Vb, and C	Li et al. (2014c)
14.	<i>Ludwigia prostrata</i>	<i>Colletotrichum</i> sp.	Pyrenocines N–O	Yang et al. (2014b)
15.	<i>Ceratonia siliqua</i>	<i>Penicillium citrinum</i>	Tanzawaic acids G–H, 6-methylcurvulinic acid, 8-methoxy-3, 5-dimethylisoquinolin-6-ol, and 1,2,3,1b-tetrahydroquinolactacide	El-Neketi et al. (2013)
16.	<i>Ocimum tenuiflorum</i>	<i>Penicillium citrinum</i>	Two new alkaloids	Lai et al. (2013)
17.	<i>Miquelia dentata</i>	<i>Fomitopsis</i> sp.	Camptothecin	Shweta et al. (2013)
18.	<i>Tamarix chinensis</i>	<i>Penicillium</i> sp.	Arisagacins I and J	Sun et al. (2013)
19.	<i>Mentha pulegium</i>	<i>Stemphylium globuliferum</i>	Altersolanol A	Teiten et al. (2013)

(continued)

**Table 7.2** (continued)

S. no.	Medicinal plants	Fungal endophytes	Anticancer compound	Reference
20.	<i>Taxus chinensis</i>	<i>Perenniporia tephropora</i>	Perenniporin A	Wu et al. (2013a)
21.	<i>Astragalus lentiginosus</i>	<i>Emericella</i> sp.	Secoestrein D	Xu et al. (2013)
22.	<i>Avicennia</i> sp.	<i>Penicillium</i> sp.	4-(methoxymethyl)-7-methoxy-6-methyl-1(3H)-isobenzofuranone	Yang et al. (2013)
23.	<i>Bruguiera sexangula</i>	<i>Pestalotiopsis foedan</i>	(-)-(4S, 8S)-foedanolid and (+)-(4R, 8R)-foedanolid	Yang and Li (2013)
24.	<i>Huperzia serrata</i>	<i>Ceriporia lacerate</i>	Ceriponols A–K,	Ying et al. (2013)
25.	<i>Annona muricata</i>	<i>Periconia</i> sp.	Periconiasins A–C	Zhang et al. (2013)
26.	<i>Cajanus cajan</i>	<i>Hypocrea lixii</i>	Cajanol	Zhao et al. (2013)

### 7.3.2 Antioxidant Compounds

Oxidation is an essential process that utilizes oxygen and metabolizes macromolecules for energy production. Paradoxically, this vital mechanism may also lead to cell and tissue damage through production of free radicals and reactive oxygen species. These radicals get stabilized by reacting with cellular components including lipids, proteins, and DNA leading to impairment in their normal structure and function. This ultimately leads to the development of pathologies such as diabetes and cardiovascular and neurodegenerative diseases. An antioxidant is a molecule that slows down the oxidative damage caused by the free radicals and inhibits the deleterious effect caused by oxidation chain reaction. Antioxidant compounds can be obtained from plants, fruits, and vegetables. Since few antioxidants are approved for clinical application due to health safety issues, exploration of novel compounds from endophytes can be considered as an alternative source. Investigation of antioxidant compounds from endophytes gained importance after the discovery of pestacin and isopestacin as antioxidant compounds from endophyte *Pestalotiopsis microspore* residing in *Terminalia morobensis* (Harper et al. 2003). Since then, different studies on isolation of diverse antioxidant compounds have been reported from endophytes.

It is presumed that phenolic and flavonoid compounds are known to possess good antioxidant capacity. Huang et al. (2007) showed a positive correlation between the antioxidant capacity of *Chaetomium* sp., an endophyte of *Nerium oleander*, to phenolic and flavonoid compounds which were the major antioxidant constituents isolated from fungal extract (Huang et al. 2007). Similarly, *Xylaria* sp. isolated from *Ginkgo biloba* has been reported to show antioxidant activity. The activity was due to the presence of phenolic and flavonoid compounds present in the methanolic extract of the fungus (Liu et al. 2007). Similarly, *Chaetomium globosum* (CDW7), an endophyte of *Ginkgo biloba*, has been reported to synthesize antioxidant compound flavipin. The compound has been reported to be used in the therapy for free radical-associated diseases (Ye et al. 2013). Yuan et al. (2013) also isolated *Penicillium* sp. from roots of *Ginkgo biloba*. Six known metabolites have been obtained from this endophyte out of which three compounds, viz., adenosine, adenine, and 2-deoxyadenosine, exhibited potential DPPH scavenging activity.

Some new metabolites possessing antioxidant activity have also been reported from endophytes. In a recent study, phomopsidone A, a novel pentacyclic depsidone, has been reported from mangrove endophytic fungus *Phomopsis*. The compound exhibited antioxidant activity in addition to antifungal and cytotoxic activities (Zhang et al. 2014). Huang et al. (2012) also reported a new isobenzofuranone derivative 4, 6-dihydroxy-5-methoxy-7-methylphthalide from *Cephalosporium* sp. AL031 endophytic in *Sinarundinaria nitida*.

An exopolysaccharide, rhamnogalactan was obtained from endophyte *Fusarium solani* SD5 isolated from *Alstonia scholaris*. The compound showed the significant free radical scavenging effect on DPPH radicals with an IC<sub>50</sub> value of 578.5 µg/ml (Mahapatra and Banerjee 2014). In another study endophytes for antioxidant compounds glutaminase enzyme possessing free radical scavenging activity were

isolated from the endophytic fungus *Penicillium citrinum*. The IC<sub>50</sub> value of enzyme for DPPH, reducing power, nitric oxide, and hydroxyl radical scavenging activity were found to be 94.65, 117.73, 87.26, and 105.62, respectively (Sajitha et al. 2014). Antioxidant compounds palmarumycins C2 and C3 from endophyte *Berkleasium* sp. Dzf12 and terrain from *Aspergillus terreus* have been reported in different studies (Mou et al. 2012; Al-Trabolsy et al. 2014). An antioxidant compound graphis lactone A was obtained from endophyte *Cephalosporium* sp. IFB-E00, a resident of *Trachelospermum jasminoides*. The compound was confirmed to have stronger antioxidant activity in vitro as compared to butylated hydroxytoluene and ascorbic acid which were used as positive control (Song et al. 2005). Cajaninstilbene acid, a natural antioxidant, has been reported from *Fusarium*, an endophyte of pigeon pea *Cajanus cajan* (Zhao et al. 2012).

### 7.3.3 Antimicrobial Compounds

Secondary metabolites produced by fungal endophytes having antimicrobial activity are a promising way to overcome the increasing threat of drug-resistant microbes. These can also be used as food preservatives in the control of food spoilage and food-borne diseases (Liu et al. 2007). Antimicrobial metabolites (antibiotics) are low-molecular-weight organic compounds produced by microorganisms that are active at low concentrations against other microorganisms not required for its growth. They are produced as an adaptation for a specific function in nature and are the most frequent bioactive natural products isolated from endophytes.

*Penicillium* sp. has always been the important source of antimicrobial compounds. The literature reviewed revealed several examples where antimicrobial compounds have been reported from diverse species of *Penicillium*. Five new picolinic acid derivatives penicolinates A–E have been isolated from an endophytic fungus *Penicillium* sp. BCC16054. Penicolinates B and C have displayed activity against *Bacillus cereus* and *Candida albicans* (Intaraudom et al. 2013). Chemical investigation of *Penicillium citrinum*, a fungal endophyte of *Ocimum tenuifolium*, has led to the isolation of two new alkaloids along with 14 known polyketides and 4 known alkaloids. Perinadine A, alternariol, and citrinin were found to be moderately active against *Staphylococcus aureus* (Lai et al. 2013). Another *Penicillium* sp. isolated as endophyte of *Curcuma longa* has been reported to exhibit antimicrobial activity against *Staphylococcus aureus* and *Escherichia coli* (Singh et al. 2014). *Staphylococcus aureus* was also susceptible to antimicrobial compounds isolated from *Penicillium* sp., an endophyte of *Acrostichum aureum*. The compounds have been identified as cyclo(pro-Thr), cyclo(pro-Tyr), and liquiritigenin (Cui et al. 2008).

Various species of *Aspergillus* isolated as endophytes from different medicinal plants have been described as the promising source of antimicrobial compounds. *Aspergillus* sp. from *Bauhinia guianensis* yielded alkaloidal antimicrobial compounds pseurotin and fumigaclavine C. The latter was found to be active against *Bacillus subtilis* (Pinheiro et al. 2013). Nigerasterols A–B and malformins A–C

have been isolated from culture extract of *Aspergillus niger*, an endophyte of mangrove plant *Avicennia marina*. Malformins A–C displayed weak activity against *S. aureus* (Liu et al. 2013). Another mangrove endophyte *Aspergillus* sp. yielded antimicrobial compound asperterpenoid A. It exhibited strong inhibitory activity against *Mycobacterium tuberculosis* (Huang et al. 2013). *Aspergillus* sp. isolated from *Melia azedarach* afforded seven metabolites. All the isolated compounds have been evaluated against several phytopathogenic fungi and pathogenic bacteria. Compounds asperpyrone A, asperazine, and rubrofusarin B were found to inhibit fungal pathogen *Aspergillus solani*. Asperpyrone A also exhibited antibacterial activity against *Staphylococcus aureus* and *Bacillus subtilis* with MICs of 25  $\mu$ M (Xiao et al. 2014b).

A large number of diverse antimicrobial compounds have been reported from *Xylaria* residing in different host plants as endophytes. 7-Amino-4-methylcoumarin was obtained from *Xylaria* sp. YX-28, an endophyte of *Ginkgo biloba*, displayed antibacterial and antifungal activity against many pathogenic organisms (Liu et al. 2007). Three compounds chaetomugilin D, chaetomugilin A, and chaetoglobosin C were isolated from *Chaetomium globosum* endophytic in *Ginkgo biloba*. All of them exhibited significant activity against *Artemia salina* and *Mucor miehei* (Qin et al. 2009). Compounds 2-hexyl-3-methyl butanodioic acid and cytochalasin D possessing antifungal activity were recovered from endophytic *Xylaria* sp. The endophyte was isolated from *Palicourea marcgravi* (Cafeu et al. 2005). Likewise, *Xylaria* F0010, an endophyte of *Abies holophylla*, was found to be a potential producer of antifungal antibiotic agent griseofulvin. The compound has been used for the treatment of human and veterinary mycotic diseases (Park et al. 2005).

Endophytic *Phoma* sp. isolated from different medicinal plants has been reported to be a promising source of antimicrobial compounds. Santiago et al. (2012) have reported a polyketide compound 5-hydroxyramulosin from *Phoma* sp., an endophyte of *Cinnamomum mollissimum*. The isolated compound exhibited antifungal activity against *Aspergillus niger*. In another study mycelial extract of *Phoma* sp. NRRL 46751, inhabiting *Sauracia scaberrinae*, afforded three new alkaloids: phomapyrrolidones A–C out of which phomapyrrolidones B and C exhibited weak activity against *Mycobacterium tuberculosis* (Wijeratne et al. 2013). Similarly, a compound phomodione, an usnic acid derivative, was reported to be produced by *Phoma* sp. isolated from *Saurauia scaberrinae*. The compound displayed antibacterial activity against *Staphylococcus aureus* (Hoffman et al. 2008). Bioassay-guided fractionation of culture filtrate of fungal endophyte *Phoma*, isolated from *Taraxacum mongolicum*, led to the isolation of 2-hydroxy-6-methyl benzoic acid. The compound showed antibacterial activity against five bacterial test pathogens: *Escherichia coli*, *Staphylococcus aureus*, *Aeromonas hydrophila*, *Edwardsiella tarda*, and *Pasteurella multocida* (Zhang et al. 2013a). Four compounds, phomafuranol, phomalacton, (3R)-5-hydroxymellein, and emodin, were isolated from ethyl acetate extract of *Phoma* sp., a marine endophyte isolated from plant *Fucus serratus* (Hussain et al. 2014).

A broad diversity of endophytic fungi exists in the rhizome of *Paris polyphylla* var. *yunnanensis*, a medicinal plant used in traditional Chinese medicine. *Fusarium*

sp. Ppf4 from this plant yielded two sterols and one fatty acid by bioassay-guided fractionation. The compounds were elucidated as 5 $\alpha$ , 8 $\alpha$ -epidioxyergosta-6, 22-dien-3 $\beta$ -ol and ergosta-8(9), 22-dien-3 $\beta$ , 5 $\alpha$ , 6 $\beta$ , 7 $\alpha$ -tetraol and displayed antimicrobial activity (Huang et al. 2009). *Fusarium redolens* DzF2 was isolated from Chinese medicinal plant *Dioscorea zingiberensis*. Beauvericin was obtained using bioautographic antibacterial assay. The compound displayed activity against six test bacteria: *Bacillus subtilis*, *Staphylococcus haemolyticus*, *Pseudomonas lachrymans*, *Agrobacterium tumefaciens*, *Escherichia coli*, and *Xanthomonas vesicatoria* (Xu et al. 2010). Taynung et al. (2011) have reported four compounds, 1-tetradecene, 8-octadecanone, 8-pentadecanone, and octylcyclohexane and 10-nonadecanone, from *Fusarium solani* isolated from *Taxus baccata*. All the compounds showed antibacterial as well as antifungal activity. *Fusarium tricinctum* was isolated from *Rhododendron tomentosum*. Transcriptome of this endophyte was sequenced; 12,006 contigs were assembled. On analyzing transcriptomic library, it yielded a peptide resin. The compound was found to be active against *Staphylococcus carnosus*, *Candida albicans*, and *Candida utilis* (Tejesvi et al. 2013).

*Colletotrichum* is one of the important genus frequently isolated as an endophyte from different hosts. The endophyte has been investigated during different studies for the isolation of antimicrobial compounds. Colletotriolide a new macrolide was obtained from *Colletotrichum* sp. residing in *Pandanus amaryllifolius*. The compound showed low activity against *E. coli* (Bungihan et al. 2013). Similarly, Chithra et al. (2014) have reported the ability of *Colletotrichum gloeosporioides* to produce piperine, a compound originally synthesized by the host plant *Piper nigrum*. The compound has antimicrobial activity. One new compound 2-phenylethyl 1H-indol-3-yl-acetate was obtained from endophyte *Colletotrichum gloeosporioides* isolated from *Michelia champaca*. The compound possessed antifungal activity against *Cladosporium cladosporioides* and *C. sphaerospermum* (Chapla et al. 2014). Some of the antimicrobial compounds isolated from fungal endobiome have been discussed, and the rest of the data has been tabulated (Table 7.3).

### 7.3.4 Immunomodulatory Compounds

Immunomodulatory compounds are those compounds that help in modulating the immune system either by stimulating it or suppressing it. Immunosuppressive compounds are required to deal with autoimmune disorders and allograft rejection in transplant patients. Immunomodulatory drugs play a key role in the treatment of cancer. Due to the emergence of new autoimmune disorders and their role in the treatment of cancer, an intensive search is going on for more effective agents that provide aid in this regard. Since fungal endophytes have the capacity to produce novel compounds, these could prove a useful source for potentially active immunomodulatory compounds (Kaul et al. 2012). Several immunomodulatory compounds have been reported from fungal endophytes in the recent past.



**Table 7.3** Antimicrobial compounds from fungal endophytes comprising the endobiome of some medicinal plants

S. no.	Medicinal plants	Fungal endophytes	Extract/compound isolated	Reference
1.	<i>Hugonia mystax</i>	<i>Aspergillus</i> sp.	Ethanol	Abirami and Boominath (2016)
2.	<i>Sapium ellipticum</i>	<i>Chaetomium</i> sp.	Polyketides	Akone et al. (2016)
3.	<i>Cymbopogon caesius</i>	<i>Curvularia lunata</i>	Ethyl acetate	Avinash et al. (2016)
4.	<i>Glycyrrhiza glabra</i>	<i>Phoma</i> sp.	Thiodiketopiperazine derivatives	Arora et al. (2016)
5.	<i>Cupressus torulosa</i>	<i>Penicillium oxalicum</i>	Methanol and chloroform	Bisht et al. (2016)
6.	<i>Acanthospermum australe</i>	<i>Aspergillus calidoustus</i>	Ophiobolin K and 6-epiophiobolin K	Carvalho et al. (2016)
7.	Mangrove plants	<i>Talaromyces amestolkiae</i>	Isocoumarins and benzofurans	Chen et al. (2016)
8.	<i>Nymphaea nouchali</i>	<i>Chaetomium globosum</i>	Chaetoglobosin A and C	Dissanayake et al. (2016)
9.	<i>Hydrastis canadensis</i>	<i>Alternaria</i> sp., <i>Colletotrichum fioriniae</i> , <i>Diaporthe eres</i> , <i>Diaporthe</i> sp., <i>Sordariomycetes</i> sp., <i>Magnaportheales</i> sp., <i>Phoma</i> sp., and <i>Pyrenochaeta cava</i>	Alternariol, alternariol monomethyl ether, 50 epi-equisetin, equisetin, 10–11 dehydrocurvularin, macrospheptide A, cordipyridone A verticillin A, aurofusarin	Egan et al. (2016)
10.	<i>Eichhornia crassipes</i>	<i>Aspergillus australoffricanus</i>	Diphenyl ether	Ebrahim et al. (2016)
11.	<i>Datura innoxia</i> and <i>Hyoscyamus muticus</i>	<i>Aspergillus fumigatus</i> , <i>A. niger</i> , <i>A. terreus</i> var. <i>affricanus</i> , <i>Cladosporium cucumerinum</i> , <i>C. oxysporum</i> , <i>Penicillium aurantiogriseum</i> , and <i>P. chrysogenum</i>	Chloroform	El-Said et al. (2016)
12.	<i>Juniperus procera</i>	<i>Aspergillus fumigatus</i> , <i>Hypocrea lutea</i> , <i>Penicillium oxalicum</i> , and <i>Preussia</i> sp.	Methanol	Gherbawy and Elharriry (2016)
13.	<i>Pteris pellucida</i>	<i>Emericella quadrilineata</i>	Benzyl benzoate	Goutam et al. (2016)
14.	<i>Glycosmis mauritiana</i>	<i>Penicillium</i> sp.	AgNP	Govindappa et al. (2016a)

(continued)

Table 7.3 (continued)

S. no.	Medicinal plants	Fungal endophytes	Extract/compound isolated	Reference
15.	<i>Curcuma longa</i>	<i>Phoma herbarum</i>	Gentisyl alcohol	Gupta et al. (2016)
16.	<i>Garcinia prussii</i>	<i>Aspergillus japonicus</i>	Variecolin and neovasifuranone B	Jouda et al. (2016b)
17.	<i>Silybum marianum</i>	<i>Talaromyces mintoluteus</i>	Talarolutins A–D; Meroterpenoids	Kaur et al. (2016)
18.	<i>Calotropis procera</i> , <i>Catharanthus roseus</i> , <i>Euphorbia prostrata</i> , <i>Trigonella foenum-graecum</i> , and <i>Vernonia amygdalina</i>	<i>Byssochlamys spectabilis</i> and <i>Alternaria</i> sp.	Ethyl acetate	Khiralla et al. (2016)
19.	<i>Menthe viridis</i>	<i>Fusarium oxysporum</i>	Broth	Kumar et al. (2016)
20.	<i>Nicotiana tabacum</i>	<i>Rhizopycnis vagum</i>	Dibenzo- $\alpha$ -pyrone derivatives	Lai et al. (2016)
21.	<i>Mahonia fortune</i>	<i>Fusarium decemcellulare</i>	Pentapeptides and lipopeptide	Li et al. (2016b)
22.	<i>Cephalotaxus hainanensis</i>	<i>Diaporthe</i> sp., <i>Phomopsis</i> sp., <i>Colletotrichum</i> sp., <i>Corynespora</i> sp., <i>Penicillium</i> sp., and <i>Nemania</i> sp.	Ethyl acetate	Liu et al. (2016)
23.	<i>Salvia miltiorrhiza</i>	<i>Alternaria</i> sp.	Alternariol 9-methyl ether	Lou et al. (2016)
24.	<i>Ficus carica</i>	<i>Aspergillus tamarii</i>	Cyclic pentapeptide: malformin E	Ma et al. (2016)
25.	<i>Melastoma malabathricum</i>	<i>Diaporthe phaseolorum</i>	Ethyl acetate	Mishra et al. (2016a)
26.	<i>Schima wallichii</i>	<i>Penicillium simplicissimum</i> and <i>Talaromyces verruculosus</i>	Ethyl acetate	Mishra et al. (2016b)
27.	<i>Rhizophora annamalayana</i>	<i>Trichoderma</i> sp.	Ethyl acetate	Narendran and Kathiresan (2016)
28.	<i>Cinnamomum iners</i> , <i>Shorea</i> <i>siamensis</i> , <i>Fernandoa</i> <i>adenophylla</i> , and <i>Quercus</i> <i>semiserrata</i>	<i>Xylaria</i> sp.	Ethyl acetate	Orachaijapunlap et al. (2016)

29.	<i>Cinnamomum malabattrum</i>	<i>Colletotrichum gloeosporioides</i>	Phenol 3, 5- dimethoxy acetate, 4'-isopropylidene-bis-(2-cyclohexyl) phenol, N-didehydrohexacarboxyl-2, 4, 5-trimethylpiperazine and 1, 2, 4-triazolium ylide	Packiaraj et al. (2016)
30.	<i>Houttuynia cordata</i>	<i>Chaetomium globosum</i>	Ethyl acetate	Pan et al. (2016)
31.	<i>Moringa oleifera</i>	<i>Aspergillus flavus</i>	Fenacalone	Rajeshwari et al. (2016)
32.	<i>Hypocrea virens</i>	<i>Premna serratifolia</i>	Epidithiodioxopiperazine, gliotoxin, bisdethiobis(methylthio)gliotoxin	Ramaweera et al. (2016)
33.	<i>Rauwolfia serpentina</i>	<i>Colletotrichum</i> sp., <i>Fusarium</i> sp., and <i>Cladosporium</i> sp.	Methanol	Singh et al. (2016)
34.	<i>Acalypha indica</i>	<i>Phoma</i> sp.	Terpenoids	Sowparthani (2016)
35.	<i>Santalum album</i>	<i>Fusarium oxysporum</i> , <i>Fusarium solani</i> , <i>Histoplasma</i> sp., <i>Periconia</i> sp., and <i>Pestalotiopsis</i> sp.	Distilled water, ethanol	Tapwal et al. (2016)
36.	<i>Picea mariana</i> and <i>Picea rubens</i>	<i>Diaporthe maritima</i>	Dihydroxyrones, phomopsolides, alpha-pyrone	Tanney et al. (2016)
37.	<i>Schinus terebinthifolius</i>	<i>Alternaria</i> sp.	E-2-hexyl-cinnamaldehyde and two compounds of the pyrrolopyrazine alkaloids	Tonial et al. (2016)
38.	<i>Buxus sinica</i>	<i>Colletotrichum</i> sp.	Colletotrichone A	Wang et al. (2016a, b)
39.	<i>Eugenia jambolana</i>	<i>Aspergillus niger</i> and <i>A. terreus</i>	Ethyl acetate	Yadav et al. (2016)
40.	<i>Lonicera japonica</i>	<i>Fusarium</i> sp.	Methanol	Zhang et al. (2016b)
41.	<i>Sapium ellipticum</i>	<i>Penicillium tropicum</i>	Cyclohexapeptide, penitropeptide, and a new polyketide, penitropone	Zeng et al. (2016)

(continued)

Table 7.3 (continued)

S. no.	Medicinal plants	Fungal endophytes	Extract/compound isolated	Reference
42.	<i>Panax notoginseng</i>	<p><i>Acremonium</i> sp.</p> <p><i>Alternaria</i> sp.</p> <p><i>Arthrinium</i> sp.</p> <p><i>Aspergillus</i> sp.</p> <p><i>Botryotinia</i> sp.</p> <p><i>Chaetomium</i> sp.</p> <p><i>Cladosporium</i> sp.</p> <p><i>Colletotrichum</i> sp.</p> <p><i>Dictyosporium</i> sp.</p> <p><i>Fusarium</i> sp.</p> <p><i>Humicola</i> sp.</p> <p><i>Ilyonectria</i> sp.</p> <p><i>Mucor</i> sp.</p> <p><i>Myrothecium</i> sp.</p> <p><i>Penicillium</i> sp.</p> <p><i>Periconia</i> sp.</p> <p><i>Pestalotiopsis</i> sp.</p> <p><i>Phialophora</i> sp.</p> <p><i>Phoma</i> sp.</p> <p><i>Phomopsis</i> sp.</p> <p><i>Plectosphaerella</i> sp.</p> <p><i>Thielavia</i> sp. and <i>Trichoderma</i> sp.</p>	Ethyl acetate	Zheng et al. (2016)
43.	<i>Edgeworthia chrysantha</i>	<i>Fusarium oxysporum</i>	Beauvericin	Zhang et al. (2016a)
44.	<i>Mallotus philippensis</i>	<i>Alternaria</i> sp., <i>Pestalotiopsis</i> sp., and <i>Phomopsis</i> sp.	Ethyl acetate	Gangwar et al. (2015)

45.	<i>Bauhinia forficata</i>	<i>Aspergillus ochraceus</i> , <i>Gibberella baccata</i> , <i>Penicillium commune</i> , and <i>P. glabrum</i> <i>Xylaria</i> sp. <i>Nectria</i> sp. <i>Fusarium</i> sp., <i>Epicoccum</i> sp. <i>Talaromyces</i> sp. and <i>Aspergillus</i> sp.	Ethyl acetate	Bezerra et al. (2015)
46.	<i>Caesalpinia echinata</i>		Ethyl acetate	Campos et al. (2015)
47.	<i>Carapa guianensis</i>	<i>Diaporthe mayeni</i> , <i>Endomelanconitopsis</i> , <i>Colletoirichum</i> sp., <i>Guignardia mangiferae</i> , <i>Pestalotiopsis</i> sp., and <i>Diaporthe melonis</i>	Ethanol	Ferreira et al. (2015)
48.	<i>Asclepias sinatica</i>	<i>Penicillium chrysogenum</i> and <i>Alternaria alternata</i>	Ethyl acetate	Fouda et al. (2015)
49.	<i>Dioscorea composita</i>	<i>Fusarium</i> sp. and <i>Alternaria</i> sp.	Steroidal saponins	Gupta et al. (2015)
50.	<i>Opuntia humifusa</i>	<i>Biscogniauxia mediterranea</i>	5-Methylmellein	Silva-Hughes et al. (2015)
51.	<i>Senecio kleinia</i>	<i>Phoma</i> sp.	Sclerodione, atrovenetinone	Hussain et al. (2015)
52.	<i>Avicennia officinalis</i>	<i>Acremonium</i> sp., <i>Cladosporium</i> sp., <i>Curvularia</i> sp., and <i>Saccharomyces</i> sp.	Ethyl acetate	Job et al. (2015)
53.	<i>Tridax procumbens</i>	<i>Alternaria</i> sp.	Methanol, chloroform, ethyl acetate, and petroleum ether	Kumar et al. (2015)
54.	<i>Tectona grandis</i>	<i>Diaporthe phaseolorum</i>	Ethyl acetate	Kumala et al. (2015)
55.	<i>Mahonia fortune</i>	<i>Diaporthe</i> sp.	Tetraacyclic Triterpenoid	Li et al. (2015a)
56.	<i>Taxus chinensis</i>	<i>Pestalotiopsis microspora</i>	$\alpha$ -Pyrone derivative	Li et al. (2015c)
57.	<i>Tephrosia purpurea</i>	<i>Penicillium griseofulvum</i> and <i>Aspergillus oryzae</i>	Broth	Luo et al. (2015)

(continued)

Table 7.3 (continued)

S. no.	Medicinal plants	Fungal endophytes	Extract/compound isolated	Reference
58.	<i>Avicennia marina</i>	<i>Penicillium brocae</i>	Sulfide diketopiperazines	Meng et al. (2015)
59.	<i>Rauwolfia serpentina</i>	<i>Colletotrichum gloeosporioides</i> , <i>Penicillium</i> sp., and <i>Aspergillus awamori</i>	Ethanol	Nath et al. (2015)
60.	<i>Panax ginseng</i>	<i>Phoma terrestris</i>	N-amino-3-hydroxy-6-meth oxyphthalimide and 5H-dibenz [B, F] azepine	Park et al. (2015)
61.	<i>Mikania glomerata</i>	<i>Diaporthe citri</i>	Ethyl acetate	Polonio et al. (2015)
62.	<i>Crescentia cujete</i>	<i>Nigrospora sphaerica</i> , <i>Fusarium oxysporum</i> , <i>Gibberella moniliformis</i> , and <i>Beauveria bassiana</i>	Aspirin and diethyl phthalate	Prabukumar et al. (2015)
63.	<i>Artemisia annua</i>	<i>Cladosporium</i> sp.	<i>Ethyl acetate</i>	Purwantini et al. (2015)
64.	<i>Aegle marmelos</i> , <i>Coccinia indica</i> , <i>Moringa oleifera</i>	<i>Cladosporium oxysporum</i>	Taxol	Raj et al. (2015)
65.	<i>Combretum latifolium</i>	<i>Gliomastix polychroma</i>	Ethyl acetate	Rao et al. (2015b)
66.	<i>Cryptolepis buchanani</i>	<i>Phomopsis liquidambaris</i>	Ethyl acetate	Rao et al. (2015a)
67.	<i>Opuntia dillenii</i>	<i>Fusarium</i> sp.	Equisetin	Ratnaweera et al. (2015b)
68.	<i>Cyperus rotundus</i>	<i>Rhizoctonia solani</i>	Solanoic acid	Ratnaweera et al. (2015a)
69.	<i>Abies</i> sp., <i>Cedrus</i> sp., <i>Juniperus</i> sp., <i>Larix</i> sp., <i>Metasequoia</i> sp., <i>Picea</i> sp., <i>Pinus</i> sp., <i>Taxus</i> sp., <i>Sambucus</i> sp., <i>Calluna</i> sp., and <i>Centaurea</i> sp.	<i>Lophodermium pinastri</i> , <i>L. sedditosum</i> , and <i>Phoma herbarum</i>	Methanol, ethyl acetate, and dichloromethane	Ravnikar et al. (2015)
70.	<i>Calophyllum apetalum</i> <i>Garcinia morella</i>	<i>Myrothecium</i> sp.	Methanol	Ruma et al. (2015)

71.	<i>Rhizophora mucronata</i> , <i>Excoecaria agallocha</i>	<i>Fusarium proliferatum</i>	Ethyl acetate	Salimi et al. (2015)
72.	<i>Indigofera suffruticosa</i>	<i>Nigrospora sphaerica</i> and <i>Pestalotiopsis maculans</i>	Methanol, ethyl acetate	Santos et al. (2015)
73.	<i>Cinnamomum camphora</i>	<i>Muscodora tigerii</i>	4-Octadecylmorpholine, 1-tetradecanamine, N,N-dimethyl, and 1,2-benzenedicarboxylic acid, mono(2-ethylhexyl) ester.	Saxena et al. (2015)
74.	<i>Tsuga heterophylla</i>	<i>Gloeosporium</i> sp.	6-Pentyl-2H-pyran-2-one	Schaible et al. (2015)
75.	<i>Caesalpinia sappan</i> , <i>Alternanthera sessilis</i> <i>Sapindus laurifolius</i> <i>Basella alba</i> and <i>Acalypha</i> <i>indica</i>	<i>Trichoderma</i> sp., <i>Aspergillus</i> sp., <i>Fusarium</i> sp., and <i>Trichoderma</i> sp.	Ethyl acetate	Srimivas et al. (2015)
76.	<i>Proxopsis juliflora</i>	<i>Colletotrichum gloeosporioides</i> and <i>Fusiclomyces lilacinus</i>	Ethyl acetate	Srivastava and Anandrao (2015)
77.	<i>Phragmites communis</i>	<i>Phoma</i> sp.	Barceloneic acid C	Xia et al. (2015)
78.	<i>Cephalotaxus hainanensis</i>	<i>Neonectria macroconidia</i> , <i>Xylaria</i> sp., and <i>Verticillium bulbillosum</i>	Ethyl acetate	Yang et al. (2015)
79.	<i>Swietenia macrophylla</i>	<i>Aspergillus terreus</i>	Di-n-octyl phthalate	Yin et al. (2015)
80.	<i>Dracaena draco</i>	<i>Botryodiplodia theobromae</i>	Dipeptides (maculosin and L,L- cyclo(leucylprolyl)), alkaloid (norharman), coumarin and isocoumarin (bergapten, meranzin, and monocerin), sesquiterpene (dihydrocumambrin A), aldehyde (formyl indanone), fatty alcohol (halaminol A), and fatty acid amide (palmitoleamide, palmitamide, capsi-amide and oleamide)	Zaher et al. (2015a)
81.	<i>Ginkgo biloba</i>	<i>Aspergillus</i> sp.	Xanthoascin	Zhang et al. (2015a)
82.	<i>Acanthus ilicifolius</i>	<i>Aspergillus flavipes</i>	Phenyl derivatives: aromatic butyrolactones, flavipesins A and B	Bai et al. (2014)

(continued)

Table 7.3 (continued)

S. no.	Medicinal plants	Fungal endophytes	Extract/compound isolated	Reference
83.	<i>Xanthium sibiricum</i>	<i>Eupenicillium</i> sp.	Eupenicinols A and B, butyloitaconic acid, and (2S)-hexylitaconic acid	Li et al. (2014a)
84.	Australian dry rainforests	<i>Preussia</i> sp.	Ethyl acetate	Mapperson et al. (2014)
85.	<i>Hyptis dilatata</i>	<i>Pestalotiopsis mangiferae</i>	Polyhydroxylated macrolide: mangiferaelactone	Ortega et al. (2014)
86.	<i>Tribulus terrestris</i>	<i>Aspergillus fumigatiifinis</i>	Neosartorin	Ola et al. (2014)
87.	<i>Vitex negundo</i>	<i>Pestalotiopsis</i> sp., <i>Fusarium</i> sp., <i>Fusarium</i> sp., and <i>Alternaria</i> sp.	Ethyl acetate	Palanichamy et al. (2014)
	<i>Justicia gendarussa</i> , <i>Ocimum basilicum</i>			
	<i>Costus spicatus</i> and <i>Glycosmis pentaphylla</i>			
88.	<i>Aloe vera</i>	<i>Talaromyces wortmannii</i>	Methanol	Pretsch et al. (2014)
89.	<i>Anoectochilus setaceus</i>	<i>Xylaria</i> sp.	Helvolic acid	Ratnaweera et al. (2014)
90.	<i>Plumeria acuminata</i> and <i>Plumeria obtusifolia</i>	<i>Colletotrichum gloeosporioides</i> and <i>Fusarium oxysporum</i>	Ethyl acetate	Ramesha and Srinivas (2014)
91.	<i>Nothapodytes foetida</i>	<i>Bionectria ochroleuca</i>	Ethyl acetate	Samaga et al. (2014)
92.	<i>Cupressus arizonica</i> , <i>C. sempervirens</i> var. <i>cereiformis</i> , and <i>Thuja orientalis</i>	<i>Alternaria alternata</i> , <i>A. pellucida</i> , and <i>A. tangelonis</i>	Methanol	Soltani and Moghaddam (2014)
93.	<i>Allium sativum</i>	<i>Trichoderma brevicompactum</i>	Extract	Shentu et al. (2014)
94.	<i>Madhuca indica</i>	<i>Aschersonia</i> sp.	Ethyl acetate	Verma et al. (2014)
95.	<i>Pinus walllichiana</i>	<i>Tritirachium oryzae</i> , <i>Truncatella spadicea</i> , and <i>Fusarium larvarum</i>	Methanol	Qadri et al. (2014)
96.	<i>Melia azedarach</i>	<i>Botryosphaeria dothidea</i>	Pycnophorin, stemphyrylenol	Xiao et al. (2014a)
97.	<i>Brguiera sexangula</i> var. <i>rhynchopetala</i>	<i>Stemphylium</i> sp.	Pyrone derivatives, infectopyrones A and B	Zhou et al. (2014b)
98.	<i>Bruguiera gymnorhiza</i>	<i>Penicillium</i> sp.	Penibrugueramine A; Pyrrolizidine alkaloid	Zhou et al. (2014a)
99.	<i>Rhizophora stylosa</i>	<i>Aspergillus nidulans</i>	Aniquinazolines A–D	An et al. (2013)



100.	Amazon rainforest biome	<i>Chaetomium globosum</i> , <i>Xylaria cubensis</i> and <i>Lewia infectoria</i>	Pyrocidine C	Casella et al. (2013)
101.	Amazon forests	<i>Xylaria Jeejeensis</i>	Xyolide	Baraban et al. (2013)
102.	<i>Campotheca acuminata</i>	<i>Botryosphaeria dothidea</i>	9-Methoxycamptothecin	Ding et al. (2013)
103.	<i>Vitex negundo</i>	<i>Phomopsis</i> sp.	Ethyl acetate, methanol, hexane	Desale and Bodhankar (2013)
104.	<i>Ceratonia siliqua</i>	<i>Penicillium citrinum</i>	Alkaloids and polyketides	El-Neketi et al. (2013)
105.	<i>Trichilia elegans</i>	<i>Phomopsis longicolla</i>	3-Nitropropionic acid	Flores et al. (2013)
106.	<i>Artabotrys odoratissimus</i> , <i>Cassia auriculata</i> , <i>Guazuma ulmifolia</i> , and <i>Terminalia catappa</i>	<i>Phomopsis</i> sp.	Ethyl acetate	Gopinath et al. (2013)
107.	<i>Cannabis sativa</i>	<i>Aspergillus niger</i> , <i>A. flavus</i> , <i>A. nidulans</i> , <i>Penicillium chrysogenum</i> , <i>P. citrinum</i> , <i>Phoma</i> sp., <i>Rhizopus</i> sp., <i>Colletotrichum</i> sp., <i>Cladosporium</i> sp., and <i>Curvularia</i> sp.	Ethanol	Gautam et al. (2013)
108.	<i>Rhizophora stylosa</i>	<i>Alternaria tenuissima</i>	Tricycloaltemarene 3, djalonensone	Hong et al. (2013)
109.	<i>Ocimum tenuiflorum</i>	<i>Penicillium citrinum</i>	Polyketides and alkaloids	Lai et al. (2013)
110.	<i>Ulmus macrocarpa</i>	<i>Microsphaeropsis arundinis</i>	Arundinols A–C and Arundinones A and B	Luo et al. (2013)
111.	<i>Bauhinia guianensis</i>	<i>Aspergillus</i> sp.	Alkaloids: fumigaclavine C and pseurotin A	Pinheiro et al. (2013)
112.	Forests of Western Ghats	<i>Xylaria</i> sp.	Ethyl acetate	Rajulu et al. (2013)
113.	<i>Cymodocea serrulata</i> , <i>Halophila ovalis</i> , and <i>Thalassia hemprichii</i>	<i>Hypocreales</i> sp., <i>Trichoderma</i> sp., and <i>Penicillium</i> sp., <i>Fusarium</i> sp. and <i>Stephanonectria</i> sp.	Ethyl acetate	Supaphon et al. (2013)

(continued)

Table 7.3 (continued)

S. no.	Medicinal plants	Fungal endophytes	Extract/compound isolated	Reference
114.	<i>Triticum durum</i>	<i>Aspergillus</i> sp., <i>Alternaria</i> sp., <i>Penicillium</i> sp., <i>Cladosporium</i> sp., <i>Chaetomium</i> sp., and <i>Phoma</i> sp.	Ethyl acetate	Sadrati et al. (2013)
115.	<i>Dioscorea zingiberensis</i>	<i>Berkleasmiium</i> sp.	Palmarumycins C3 and C4	Mou et al. (2013)
116.	<i>Cedrus deodara</i> , <i>Pinus roxburghii</i> , and <i>Abies pindrow</i>	<i>Trichophaea abundans</i> , <i>Diaporthe phaseolorum</i> and <i>Fusarium redolens</i>	Methanol	Qadri et al. (2013)
117.	<i>Ficus pumila</i>	<i>Phomopsis</i> sp.	Ethyl acetate	Rakshith et al. (2013)
118.	<i>Theobroma cacao</i>	<i>Epicoccum</i> sp.	Polyketides: Epicoccolides	Talontsi et al. (2013)
119.	<i>Rhododendron tomentosum</i>	<i>Fusarium tricinatum</i>	Polypeptides	Tejesvi et al. (2013)
120.	<i>Panax ginseng</i>	<i>Fusarium</i> sp.	Triterpenoid saponin	Wu et al. (2013b)
121.	<i>Rheum palmatum</i>	<i>Fusarium solani</i>	Rhein	You et al. (2013)
122.	<i>Taraxacum mongolicum</i>	<i>Phoma</i> sp.	2-Hydroxy-6-methylbenzoic acid	Zhang et al. (2013a)
123.	<i>Clidemia hirta</i>	<i>Cryptosporiopsis</i> sp.	1-(2,6-Dihydroxyphenyl)pentan-1-one (2) and (Z)-1-(2-(2-butaryl-3-hydroxyphenoxy)-6-hydroxyphenyl)-3-hydroxybut-2-en-1-one	Zilla et al. (2013)

*Pestalotiopsis* sp. isolated from *Taxus brevifolia* (Yew tree) has been reported for immunosuppressive pestalotiopsins A and B (Pulici et al. 1996). Another immunosuppressive compound cytochalasin U has been produced by *Pestalotia* sp. isolated from *Cassia fistula* (Burres et al. 1992).

Subglutinols A and B, two immunosuppressive compounds, have been obtained from *Fusarium subglutinans*, inhabiting *Tripterygium wilfordii*. Both the compounds were nontoxic and very potent in the thymocyte proliferation (TP) assays and mixed lymphocyte reaction (MLR) (Lee et al. 1995). Cyclosporine-A, an immunosuppressive drug isolated from endophyte, was found to be 104 times more potent in the TP assay and roughly as potent in the MLR assay (Bentley et al. 2000). Similarly, *Pestalotiopsis leucothoe* from *Tripterygium wilfordii* has been documented to produce three compounds designated as BS, GS, and YS. All the compounds showed variable effects on T and B cells and monocytes. Hence, these represent a new source of immunomodulatory compounds for the treatment of human immune-mediated diseases. However, the structure of the compounds has not been elucidated yet (Kumar et al. 2005). Another potent immunosuppressive fungal metabolite used for the treatment of autoimmune diseases and organ transplantations has been documented to be produced by fungal endophytes *Penicillium*, *Aspergillus*, *Byssochlamys*, and *Septoria* species. The compound was identified as mycophenolic acid (Larsen et al. 2005).

Ren et al. (2008) have recorded collutelin-A and cyclosporine-A from *Colletotrichum dematium* inhabiting *Pteromischum* sp. growing in the tropical forests of Costa Rica. The compound displayed strong immunosuppressive activity by inhibiting CD4 T-cell activation of interleukin-2 production, whereas cyclosporine-A showed moderate activity in the same experiment.

Recently, an endophyte *Phomopsis longicolla* yielded four tetrahydroxanthone dimers, of which phomoxanthone A showed immunostimulation and pro-apoptotic activity. The compound exhibited immunostimulation by activating T lymphocytes, NK cells, and macrophages (Ronsberg et al. 2013). Similarly, *Botryosphaeria dothidea* isolated from *Kigelia africana* has been evaluated for its immunomodulatory potential. It was found to suppress T-cell proliferation by 50% and also inhibited TNF- $\alpha$  production (Katoch et al. 2014). These examples depict the potentiality of endophytes for exploring rare and uncommon immunomodulatory compounds.

### 7.3.5 Antidiabetic Compounds

Diabetes mellitus is the highest cause of death among other chronic diseases. It cannot be cured but controlled. In 2015, about 415 million people had diabetes worldwide, with type II diabetes accounting for about 90% of the cases (Cui et al. 2016). It can cause complications such as cardiovascular disorders, kidney failure, impotency, blindness, and gangrene. One of the strategies used to cure this is by inhibiting digestion of complex carbohydrates in the small intestine into glucose, resulting in the reduction of intake of glucose into the blood. Alpha-glucosidase and

alpha-amylase inhibitors are known to possess such activity. Medicinal plants for diabetes are a potential source of microbes producing alpha-glucosidase inhibitors.

$\alpha$ -Glucosidase is an important enzyme for breaking down complex carbohydrates for absorption;  $\alpha$ -glucosidase inhibitors such as acarbose, miglitol, and voglibose, all originating from natural products, are widely used to treat type II diabetes, indicating that natural products are an important source of antidiabetic drugs.

*Syncephalastrum* sp. isolated from *Adhatoda beddomei* exhibits antidiabetic activity by inhibiting  $\alpha$ -amylase (Prabavathy and Valli 2013). The alpha-glucosidase inhibitory activity of endophytic fungi isolated from *Cassia siamea* has also been reported (Munim et al. 2013).  $\alpha$ -Amylase inhibitor from endophytic fungi of antidiabetic medicinal plants of the Western Ghats retards the liberation of glucose from dietary complex carbohydrates and delays the absorption of glucose. Antidiabetic activity of ethanolic and acetone extracts of endophytic fungi *Syncephalastrum racemosum* isolated from the seaweed *Gracilaria corticata* by alpha-amylase inhibition has been reported (Ushasri and Anusha 2015). Similarly, endophytic *Alternaria* sp. isolated from *Viscum album* exhibited strong antidiabetic activity on alloxan-induced diabetic rats (Govindappa et al. 2015). The additional examples of antidiabetic activity of fungal endophytes have been tabulated (Table 7.4).

### 7.3.6 Acetylcholinesterase Inhibitory Activity of Fungal Endophytes

Alzheimer's disease is a neurodegenerative disease of the central nervous system. The first clinical manifestation is recent memory dysfunction, which is followed by persistent intellectual impairment, loss of judgment and reasoning abilities, aphasia, and movement dysfunction. A study found that of the 10–15% of elderly people with different degrees of dementia, approximately 60–70% of the cases are due to Alzheimer's disease. However, the pathogenesis of senile dementia is not clear. Cholinergic nerve injury is the most accepted hypothesis of Alzheimer's disease pathogenesis, and if this is true, acetylcholinesterase inhibitors could be developed to effectively improve Alzheimer's disease treatment.

The use of acetylcholinesterase inhibitors is the most effective approach to treating the cognitive symptoms of Alzheimer's disease (Zhang et al. 2011) and has other possible therapeutic applications in the treatment of Parkinson's disease, senile dementia, and ataxia (Zhang et al. 2011; Singh et al. 2012). Acetylcholinesterase inhibitors such as eserine, tacrine, donepezil, rivastigmine, and galantamine are the drugs currently approved for the treatment of Alzheimer's disease (Anand and Singh 2013). The additional examples of acetylcholinesterase inhibitory activities of endophytic fungal isolates have been tabulated (Table 7.5).

**Table 7.4** Antidiabetic activity of endophytic fungi isolated from medicinal plants

S. no.	Medicinal plants	Fungal endophytes	Bioactivity	Extract/compounds isolated	References
1.	<i>Cupressus torulosa</i>	<i>Penicillium oxalicum</i>	Alpha-amylase inhibitory activity	Chloroform and methanol	Bisht et al. (2016)
2.	<i>Mangrove plants</i>	<i>Talaromyces amestolkiae</i>	$\alpha$ -Glucosidase inhibitory and antibacterial	Isocoumarins and benzofurans	Chen et al. (2016)
3.	<i>Sonneratia ovate</i>	<i>Nectria</i> sp.	$\alpha$ -Glucosidase inhibitory activity	Polyketides: Nectriacid B, nectriacid C	Cui et al. (2016)
4.	<i>Hintonia latiflora</i>	<i>Xylaria feejeensis</i>	$\alpha$ -Glucosidase inhibitors	Pestalotin 4'-O-methyl- $\beta$ -mannopyranoside and 3S,4R-(+)-4-hydroxymellein	Chavez et al. (2015)
5.	<i>Viscum album</i>	<i>Alternaria</i> sp.	Antidiabetic	Lectin(N-acetylglactosamine)	Govindappa et al. (2015)
6.	<i>Sonneratia apetala</i>	<i>Aspergillus</i> sp.	Antidiabetic	Methanol Isocoumarin derivatives Aspergiferanone	Liu et al. (2015b)
7.	<i>Cerbera manghas</i>	<i>Penicillium</i> sp.	$\alpha$ -Glucosidase inhibitory	( $\pm$ )-penifuprone and phenolic compounds	Liu et al. (2015a)
8.	<i>Acacia nilotica</i>	<i>Aspergillus awamori</i>	Antidiabetic	Peptide	Singh and Kaur (2016)
9.	<i>Tinospora cordifolia</i>	<i>Cladosporium</i> sp.	$\alpha$ -Glucosidase inhibitors	Phenolic compound	Singh et al. (2015b)
10.	<i>Gracilaria corticata</i>	<i>Synechalastrum racemosum</i>	Antidiabetic	Ethanol, acetone	Ushasri and Anusha (2015)
11.	<i>Momordica charantia</i> and <i>Trigonella foenumgraceum</i>	<i>Trichoderma atroviride</i> and <i>Stemphylium globuliferum</i>	Antidiabetic	Ethyl acetate	Pavithra et al. (2014)
12.	<i>Morus alba</i>	<i>Alternaria</i> sp.	Antidiabetic	Ethyl acetate	Zheng et al. (2014)
13.	<i>Salvadora oleoides</i>	<i>Aspergillus</i> sp.	Antidiabetic	2, 6-Di-tert-butyl-p-cresol and phenol, 2, 6-bis (1, 1-dimethylethyl)-4-methyl	Dhankar et al. (2013)
14.	<i>Ficus religiosa</i>	<i>Dendrophion nanum</i>	Antidiabetic	Herbarin (naphthoquinones)	Mishra et al. (2013)
15.	<i>Catharanthus roseus</i>	Fungal endophytes	Antidiabetic	Ethyl acetate	Rosaline and Agastian (2013)
16.	<i>Coscinium jensestratum</i>	<i>Fusarium solani</i>	Antidiabetic	Berberine	Vinodhini and Agastian (2013)

**Table 7.5** Acetylcholinesterase inhibitory activity of endophytic fungi isolated from medicinal plants

S. no.	Medicinal plants	Fungal endophytes	Activity	Extract/compounds isolated	References
1.	<i>Catharanthus roseus</i>	<i>Alternaria alternata</i>	Acetylcholines terase inhibitory	Altenune	Bhagat et al. (2016)
2.	<i>Panax notoginseng</i>	<i>Chaetomium globosum</i>	Acetylcholines terase inhibitory	3-Methoxy epicoccone, epicoccolides B	Li et al. (2016a)
3.	<i>Huperzia serrata</i>	<i>Colletotrichum</i> sp.,	Acetylcholines terase inhibitory	Ethanol	Wang et al. (2016b)
4.	<i>Phlegmariurus phlegmaria</i>	<i>Ceriporia lacerate</i>	Acetylcholines terase inhibitory	Chloroform	Zhang et al. (2015b)
5.	<i>Huperzia serrata</i>	<i>Paecilomyces tenuis</i> YS-13	Acetylcholines terase inhibitory	Huperzine A	Su and Yang (2015)

### 7.3.7 Endophytes as a Source of Silver Nanoparticles

Nanotechnology is the ability to work at atomic, molecular, and supramolecular levels. It involves production, manipulation, and use of material ranging from less than a micron. Nanoparticles have a wide range of applications in diverse fields like catalysis, sensors, medicine, etc., and these depend on the physical and optical properties of the particles. As the field of nanotechnology is progressing, the knowledge of physical and chemical characteristics of nanoparticles has greatly increased. The most well-known nanoparticles are made from silver metal. Silver is used in medical fields as a topical bactericide. Silver nanoparticles possess broad-spectrum multifunctional activities and have the promising therapeutic potential to be used for the treatment of burns and variety of infections. The emphasis is being given to their use in prophylaxis and treatment of different types of cancers and microbial infections. The silver nanoparticles can change the 3D structure of proteins by interfering with S–S bond and block the functional operations of microorganisms (Sunkar and Nachiyar 2013). In the recent years apart from silver, gold nanoparticles have also been the focus of interest because of their emerging applications in the areas such as bioimaging and biosensors (Alappat et al. 2012).

Silver nanoparticles can be synthesized using chemical approaches, but it leads to the presence of traces of toxic chemicals absorbed on the surface which is undesirable in the medical applications (Bharathidasan and Panneerselvam 2012). Moreover, a lot of hazardous by-products are generated using this approach. Considering these facts, an alternative approach for nanomaterial synthesis has to be thought of. An important aspect in the field of nanotechnology is to develop a reliable and eco-friendly process for the synthesis of nanoscale materials.

Green technology is emerging nowadays and involves the use of microorganisms in the synthesis of nanoparticles (Razavi et al. 2015). The synthesis of nanoparticles using biological systems provides new routes to develop nanoparticles with desired

**Table 7.6** Nanoparticle producing endophytic fungi isolated from medicinal plants

S. no.	Endophyte	Plant	Nanoparticle	Activity	References
1.	<i>Penicillium</i> sp.	<i>Calophyllum apetalum</i>	Ag-Nps	Not reported	Chandrappa et al. (2016)
2.	<i>Aspergillus versicolor</i>	<i>Centella asiatica</i>	Ag-Nps	Antimicrobial Antioxidant	Netala et al. (2016)
3.	<i>Fusarium solani</i>	<i>Withania somniafera</i>	Ag-Nps	Antibacterial Cytotoxic	Vijayan et al. (2016)
4.	<i>Colletotrichum</i> sp.	<i>Andrographis paniculata</i>	Ag-Nps	Antibacterial	Azmath et al. (2016)
5.	<i>Fusarium</i> sp.	<i>Withania somniafera</i>	Ag-Nps	Antibacterial	Singh et al. (2015a)
6.	<i>Penicillium</i> sp. and <i>Alternaria</i> sp.	<i>Gloriosa superba</i>	Ag-Nps	Antibacterial	Devi et al. (2014)
7.	<i>Cryptosporiopsis ericace</i>	<i>Potentilla fulgens</i>	Ag-Nps	Antimicrobial	Devi and Joshi (2014)
8.	<i>Penicillium</i> sp.	<i>Curcuma longa</i>	Ag-Nps	Antimicrobial	Singh et al. (2014)
9.	<i>Epicoccum nigrum</i>	<i>Phellodendron amurense</i>	Ag-Nps	Antifungal	Qian et al. (2013)
10.	<i>Penicillium</i> sp.	<i>Centella asiatica</i>	Ag-Nps	Antimicrobial	Devi et al. (2012)

properties for making their exploitation possible in diverse fields (Pugazhenthiran et al. 2009). The nonpathogenic and eco-friendly behavior of endophytes makes them as good candidates for the synthesis of nanoparticles (Kaul et al. 2014). Fungal endophytes can be exploited for large-scale extracellular synthesis of nanoparticles which makes the downstream processing easier (Verma et al. 2011). Various studies on fungal endobiome described their ability to synthesize nanoparticles particularly silver nanoparticles (Ag-Nps) (Table 7.6).

### 7.3.8 Antitubercular Compounds

Tuberculosis is currently a major public health problem due to the advent of multidrug-resistant (MDR) forms of bacilli as well as human immunodeficiency virus epidemics. The World Health Organization (WHO) estimated that currently 50 million people are infected and 1,500 people die each hour from tuberculosis worldwide. After emergence and spread of *Mycobacterium tuberculosis*-resistant strains to multiple drugs, the search for new antimycobacterial agents is timely. The globe recognized medicinal plants as a repository for fungal endophytes with metabolites containing the novel molecular structure and biologically active compounds against various human pathogenic diseases for potential use in modern medicine. Endophytic fungi are a good source for exploring the possibility of new antimycobacterial drugs. Recently, polyketide such as penialidin C has been isolated from endophytic

*Penicillium* sp. of *Garcinia nobilis* that exhibits significant activity against *Mycobacterium tuberculosis* (Jouda et al. 2016a).

A new isofuranonaphthalenone isolated from endophytic fungus *Nodulisporium* sp. of *Antidesma ghaesembilla* displayed antimycobacterial activity with IC<sub>50</sub> values of 3.125 µg/mL (Prabpai et al. 2015). Asperlones A and B, dinaphthalenone derivatives, asperterpenoid A, and alterporriol-type dimmers from mangrove endophytic fungus *Aspergillus* sp. and *Alternaria* sp., respectively, exhibited potent inhibitory effects against *Mycobacterium tuberculosis* protein tyrosine phosphatase B (MtpB) with IC<sub>50</sub> values of 4.24 ± 0.41, 8.70, and 2.2 µM, respectively (Xia et al. 2014, 2015; Huang et al. 2013). Peniphenones A–D from endophytic *Penicillium dipodomycicola* of *Acanthus ilicifolius* exhibited strong inhibitory activity against *Mycobacterium tuberculosis* protein tyrosine phosphatase B (MtpB) (Li et al. 2014b).

Numerous secondary metabolites are known from endophytic strains of *Phoma* sp. Phomapyrrolidones A–C, alkaloids from the endophytic fungus *Phoma* sp. of *Saurauia scaberrinae*, possess significant antitubercular activity (Wijeratne et al. 2013). Penicolinates A–E from endophytic *Penicillium* sp. of grasses belonging to Poaceae family were found to possess antitubercular activity (Intaraudom et al. 2013). Colletotriolide, a macrolide isolated from *Colletotrichum* sp. endophytic to *Pandanus amaryllifolius*, exhibited an inhibition of greater than 90% at 128 µg/mL for *M. tuberculosis* (Bungihan et al. 2013).

### 7.3.9 Antihelminthic, Antiplasmodial, and Antileishmanial Compounds

Parasitic diseases such as malaria and leishmaniasis affect millions of people worldwide and pose a major health problem in developing countries. Malaria and leishmaniasis have affected major population with increasing number of new cases each year. Leishmaniasis is caused by protozoan parasites that belong to the genus *Leishmania* and is transmitted by the bite of certain species of sand fly (subfamily *Phlebotominae*). Most of the current drugs used to treat parasitic diseases are decades old and have many limitations, including the emergence of drug resistance. For leishmaniasis, either the first-line pentavalent antimonials or second-line drugs such as amphotericin B are available, which are costly and have serious side effects and are getting resistant to pathogens after treatment for several weeks, and hence there is a need for new antileishmanial agents with improved efficacy and fewer side effects for both visceral and cutaneous leishmaniasis.

Malaria remains the world's most devastating human parasitic infection, afflicting more than 500 million people and causing about 2.5 million deaths each year. It is an infectious disease caused by the main four protozoan species of the genus *Plasmodium* (*Plasmodium falciparum*, *P. malariae*, *P. ovale*, and *P. vivax*) (Mendis et al. 2009). The increasing resistance to existing antimalarial drugs demands the exploration of novel drugs and treatment efforts to eliminate this deadly disease. Natural products contain a great variety of chemical structures and have been



screened for antiplasmodial activity as potential sources of new antimalarial drugs (De Silva et al. 2013).

The development of anthelmintic resistance in helminths reported in a number of countries gives a clear indication that control programs based exclusively on their use are not sustainable. The development of integrated programs to control helminths is vital, but such control programs require viable alternatives to the use of anthelmintics. The history of herbal medicine is almost as old as human civilization. Medicinal plants have served through ages, as a constant source of medicaments for the exposure of a variety of diseases. The endophytic fungi act as an alternative to provide a rich source of anthelmintics, antibacterials, and insecticides.

Screening of mangrove endophytic fungi for antimalarial natural products displays the most favorable bioactivity profile (Calcul et al. 2013). Two unusual dibenzofurans, preussiafurans A–B, isolated from the fungus *Preussia* sp. occurring in *Enantia chlorantha* possessed antiplasmodial activity against erythrocytic stages of chloroquine-resistant *Plasmodium falciparum* (Talontsi et al. 2014). Reduced perylenequinone derivatives from an endophytic *Alternaria* sp. isolated from *Pinus ponderosa* were found to have antileishmanial and antimalarial activities (Idris et al. 2015). Meroterpenoid, isocoumarin, and phenol derivatives isolated from seagrass endophytic fungi *Pestalotiopsis* sp. exhibited antimalarial activity (Arunpanichlert et al. 2015).

Chemical and biological investigation for the endophytic fungus *Nigrospora sphaerica* led to the isolation of nigrosphaerin A, a new isochromene derivative with moderate antileishmanial activity having IC<sub>50</sub> values of 30.2, 26.4, and 36.4 µg/ml, respectively (Metwaly et al. 2013). Enniatins (ENs), a group of antibiotics commonly produced by various strains of *Fusarium*, are six-membered cyclic depsipeptides formed by the union of three molecules of D- $\alpha$ -hydroxyisovaleric acid and three N-methyl-L-amino acids. The endophyte *Fusarium tricinctum* isolated from the fruits of *Hordeum sativum* showed antileishmanial activities (Zaher et al. 2015b).

The genus *Aspergillus* represents a diverse group of fungi, which are among the most abundant fungi in the world. Biologically active metabolites from endophytic fungus *A. terreus* isolated from the roots of *Carthamus lanatus* were found to have antileishmanial activity. Terrenolide S, a new butenolide derivative, together with (22E,24R)-stigmasta-5,7,22-trien-3- $\beta$ -ol and stigmast-4-ene-3-one exhibited antileishmanial activity toward *Leishmania donovani* with IC<sub>50</sub> values of 27.27, 15.32, and 27.27 µM, respectively, and IC<sub>90</sub> values of 167.03, 40.56, and 14.68 µM, respectively (Elkhayat et al. 2016).

### 7.3.10 Extracellular Enzymes

Among a large number of microorganisms capable of producing useful enzymes, filamentous fungi are of particular interest due to their easy cultivation and high production of extracellular enzymes. Fungal enzymes are gaining importance in agriculture, industry, and human health as they are often more stable (at high temperature and extreme pH) than the enzymes derived from plants and animals (Maria

et al. 2005). Fungal endophytes known to be the treasure of new compounds represent an interesting alternative to be explored for enzyme production with different potentialities (Bhagobaty and Joshi 2012). Diverse array of extracellular enzymes produced by endophytes include cellulases, chitinases, amylases, lipases and proteases, pectinases, laccase, etc., having wide application in various industrial processes such as baking, brewing, textile, confectionaries, paper, pulp and leather, manufacturing corn syrup, hydrolyzing milk proteins, removing stains, separating racemic mixtures of amino acids, bioremediation, and biosensing (Kaul et al. 2014). The extracellular enzyme production varies among the fungal isolates. By optimizing the conditions, these isolates can prove to be a novel source of industrially relevant enzymes.

Different studies have been carried out on the screening of endophytes for enzyme production. For example, 50 fungal strains isolated from medicinal plants (*Alpinia calcarata*, *Bixa orellana*, *Calophyllum inophyllum*, and *Catharanthus roseus*) were screened for their ability to produce extracellular enzymes such as amylase, cellulase, laccase, lipase, pectinase, and protease on solid media. Variation in the enzyme production was recorded among the isolates. The array of enzymes produced by different fungal isolates often depends on the host and their ecological factors (Sunitha et al. 2013).

Endophytic fungi isolated from *Opuntia ficus-indica* were analyzed for preliminary screening for enzyme production. Among the 24 isolates which were studied, *Aspergillus japonicus* presented pectinolytic activity, and cellulase activity was exhibited by *Xylaria* (Bezerra et al. 2012).

Similarly, 30 fungal endophytes isolated from indigenous monocotyledonous and dicotyledonous plants have been evaluated for amylase, cellulase, protease, lipase, and laccase activity, and most of them showed positive results (Patel et al. 2013). Endophytes from *Lantana camara* have been screened for amylase, lipase, and laccase production, and three isolates were shown to produce three enzymes (Desire et al. 2014). Fungal endophytes isolated from *Butea monosperma*, a tropical medicinal plant, and *Bacopa monnieri* were found to be potential producers of industrial enzymes such as amylase, cellulase, pectinase, protease, and lipase (Tuppad and Shishupala 2014; Katoch et al. 2014a, b). The extracellular enzymatic activity of endophytic fungi *Cladosporium* sp., *Rhizoctonia* sp., *Aspergillus* sp., *Chaetomium* sp., *Biosporous* sp., *Fuzarium* sp., *Curvularia* sp., *Cladosporium* sp., and *Colletotrichum* sp. isolated from medicinal plants *Azadirachta indica*, *Citrus limon*, *Gossypium hirsutum*, *Magnolia champaca*, *Datura stramonium*, *Piper betle*, and *Phyllanthus emblica* has been reported (Patil et al. 2015a). Endophytic fungi from leaves of *Calophyllum inophyllum* produce extracellular enzymes such as amylase, protease, lipase, and cellulase (Patil et al. 2015a). Endophytic microbial resources producing extracellular enzymes can establish a unique niche for ecological adaptation during symbiosis with the host frankincense tree *Boswellia sacra* (Khan et al. 2016). A few more examples of common enzymes isolated from fungal endobiome of medicinal plants are enlisted below (Table 7.7).

**Table 7.7** Extracellular enzymes produced by fungal endophytes of medicinal plants

S. no.	Fungal endophytes	Medicinal plants	Enzyme	References
1.	<i>Corchorus olitorius</i>	<i>Aspergillus terreus</i>	Xylanase	Ahmed et al. (2016a)
2.	Marine habitat	<i>Aspergillus</i> sp.	L-asparaginase	Ahmed et al. (2016b)
3.	<i>Cupressus torulosa</i>	<i>Alternaria alternata</i>	Protease	Rajput et al. (2016)
4.	<i>Artemisia annua</i>	<i>Aspergillus</i> sp.	Amylolytic	Ogbonna et al. (2015)
5.	<i>Eurotium</i> sp.	<i>Curcuma longa</i>	Asparaginase	Jalgaonwala and Mahajan (2014)
6.	<i>Preussia minima</i>	<i>Eremophila longifolia</i>	Amylase	Zaferanloo et al. (2014a)
7.	<i>Alternaria alternata</i> and <i>Phoma herbarum</i>	<i>Eremophila longifolia</i>	Protease	Zaferanloo et al. (2014b)
8.	<i>Sordaria humana</i>	<i>Cedrus deodara</i> and <i>Pinus roxburghii</i>	Cellulase	Syed et al. (2013)

## 7.4 Conclusions

Fungal endobiome of medicinal plants is considered as an important and viable component of microbial biodiversity that offers a plethora of advantages to its host plant by producing bioactive secondary metabolites. In the continuous search for novel drug sources, endophytic fungi have proven to be a promising, largely untapped reservoir of natural products. A perusal of the literature indicates many ethnomedicinal plant species known to harbor potential endophytes that produce bioactive metabolites. Therefore, it is significant to bioprospect endophytes from medicinal plants for bioactive secondary metabolites. Bioactive metabolites from fungal endobiome could be a resolute solution to the present-day problems like the emergence of new diseases and resistance to existing drugs. The ability of endophytes to produce bioactive metabolites is influenced by its interaction with the host plant and its cross talk with other microbiota associated with the host. So it is very much significant to understand the mechanisms underlying the plant–microbe interaction. Improvization of isolation and purification methods needs to be done for commercial production of bioactive metabolites. Many novel and valuable compounds with antioxidant, anticancer, antimicrobial, immunomodulatory, and anti-diabetic activities have been reported from fungal endophytes. This proves that fungal endobiome of medicinal plants certainly holds in them great potential to improve future in medicinal cure along with various industrial applications.

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