

# Chapter 12

## PET Radiopharmaceuticals in Oncology Beyond FDG

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**Abstract** Several imaging modalities to diagnose cancer, which include computed tomography (CT), magnetic resonance imaging (MRI), ultrasound, optical imaging, and gamma scintigraphy, have been previously used. For instance, CT and MRI provide considerable anatomic information about the location and the extent of tumors, but do not adequately differentiate residual or recurrent tumors from edema, radiation necrosis, or gliosis. On the other ultrasound images provide limited information about local and regional morphology with blood flow. Similarly optical imaging showed promising results, but did not demonstrate the ability to detect deep tissue penetration. Notably, radionuclide imaging modalities are diagnostic cross-sectional imaging techniques that map the location and concentration of radionuclide-labeled compounds. Moreover, molecular imaging agents are making it possible to “see” the molecular makeup of the tumor and its metabolic activity beyond tumor location, size, shape, and viability. Other technological limitations include nuclear images, which can provide a very accurate definition of metabolically active areas, but miss anatomic features. As a result, new imaging modalities have combined nuclear images with CT scans for treatment planning. The hybrid scanners combine anatomic and functional images taken during a single procedure, without having to reposition the patient between scans. In this chapter, multiple ligands beyond clinical standard  $^{18}\text{F}$ -fluorodeoxyglucose are reviewed.

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## 12.1 Molecular Imaging in Oncology

CT and MRI assess the effectiveness of cancer therapy (e.g., volumetric and morphological changes), whereas the endpoints rely almost exclusively on the analysis of biopsies by molecular and histopathological methods. While these methods provide microscopic data of the general heterogeneous process, nuclear imaging measures blood flow and metabolic patterns of the target organs. Therefore, a predictive nuclear biomarker is needed to assess clinical endpoints adequately and allow precise measurements of tumor targets on a whole-body image, upon administration of a functional radiolabeled agent. These mechanism-based agents provide image-guided therapy, which may help to decide whether it is better for the patient to discontinue an ineffective treatment at an earlier phase. Molecular imaging agents play a major role in drug discovery and development because of their ability to quantify drug properties *in vivo*. For example, positron emission tomography (PET) agents show high sensitivity and specific activities, since they are made through a nuclear transformation and use carrier-free forms of isotopes. Thus, PET agents do not produce detectable pharmacologic effects but provide important information concerning the characterization of various diseases such as the central nervous system (epilepsy, psychosis, dementia, Alzheimer's disease) [1–4] and cardiovascular system diseases (myocardial viability) [5]. At the same time it also helped determine cancer staging, cancer restaging, and treatment planning for malignant diseases [6]. In addition, molecular imaging helps to control and monitor dosage for increased safety and effectiveness. The trends for PET agent's development in oncology are to assist in determining optimal therapeutic dose, to delineate differential diagnosis between inflammation/infection and recurrence, to determine sensitive or resistant to treatment responses, and thus to select the patients who may be good candidates for therapy. Below are the molecular targets for pathway-activated imaging systems.

## 12.2 Glucose Transporter Target

2-Deoxy-2-[ $^{18}\text{F}$ ]fluoro-D-glucose ( $^{18}\text{F}$ -FDG)-PET has been used to diagnose [7–9] and stage tumors [10–20], myocardial infarction [21], and neurologic diseases [22, 23].  $^{18}\text{F}$ -FDG was developed in 1976 for the specific purpose of mapping brain glucose metabolism in living humans. After the first synthesis of  $^{18}\text{F}$ -FDG via an electrophilic fluorination with  $^{18}\text{F}$  gas, small-volume enriched water targets were developed, which made it possible to produce large quantities of [ $^{18}\text{F}$ ] fluoride ion via high yield  $^{18}\text{O}(\text{p},\text{n})^{18}\text{F}$  reaction. This was followed by a major milestone, resulting in the development of a nucleophilic fluorination method that produced  $^{18}\text{F}$ -FDG in very high yields. These advances and the remarkable properties of  $^{18}\text{F}$ -FDG have largely overcome the limitations of the 110-min half-life of  $^{18}\text{F}$ . Although  $^{18}\text{F}$ -FDG has been successfully used to image tumors with high glycolytic activity [24]

in the past two decades, it has several limitations that give rise to false-positive/false-negative diagnosis and poor predictive value of tumor response to chemoradiation therapy [25]. For instance, FDG has poor contrast in brain tumor due to the high uptake of glucose in normal brain tissue [26, 27], and exhibits poor differentiation between tumors and inflammatory tissue due to the high uptake of FDG by granulocytes and macrophages [28].

To improve the diagnosis, prognosis, planning, monitoring, and prediction of cancer treatment results, radiolabeled N-acetylglucosamine of the hexosamine biosynthetic pathway (HBP) has been used as an alternative to PET FDG [29–32]. Glucose and glutamine are substrates in the synthesis of GlcNAc via the HBP and key nutrients for cancer cells. Glutamine is a precursor amino acid for the synthesis of glucosamine, a prominent initiator in the HBP [33]. Briefly, fructose-6-phosphate from the glycolytic pathway combines with glutamine in the presence of the initiating enzyme glutamine–fructose-6-phosphate amidotransferase (GFAT) to synthesize glucosamine-6-phosphate. Subsequent enzymatic steps lead to the production of uridine diphosphate N-acetylglucosamine (GlcNAc), a substrate for O-linked glycosylation that is regulated by the terminating enzyme *O-linked* GlcNAc (*O*-GlcNAc) *transferase* (*OGT*). *OGT* is the enzyme responsible for the addition of a single GlcNAc residue to the hydroxyl groups of serine and/or threonine residues of target proteins. The HBP terminating in *O*-GlcNAc cycling (*O*-GlcNAcylation) has been implicated in cellular signaling cascades and regulation of transcription factors involved in cancer biology [34–37]. In fact, *O*-GlcNAc glycosylation on the serine or threonine residues of nuclear and cytosolic proteins is common and occurs frequently in all multicellular eukaryotes [38]. *O*-GlcNAc is part of posttranslational modification reactions and appears to modify many nucleocytoplasmic proteins, such as the transcription factors NF- $\kappa$ B and NFATc1. NF- $\kappa$ B and NFATc1 are both downstream targets of *O*-GlcNAc modification during T- and B-cell activation [39]. However, these transcription factors are constitutively activated in cancers and play an important role in growth and survival [40]. The chelator ethylenedicysteine (EC) was conjugated to glucosamine to mimic a GlcNAc analog called ethylenedicysteine-N-glucosamine (ECG). Radiolabeled ECG has been reported as a promising imaging agent in various tumor models in rodents [29, 31], as well as in humans [30].

### 12.3 Amino Acid Transporter Target

Radiolabeled amino acids are an alternative in characterizing tumors because of their much higher accumulation in tumors than in normal tissues and their rapid blood clearance. In keeping with the higher uptake, the expression of amino acid transporters is upregulated in cancer cells [41, 42], which indirectly measure cell proliferation [43], and assessment of their activities provides the potential of differential diagnosis and early prediction of treatment responses. Although  $^{11}\text{C}$ -methionine is useful for metabolic imaging of tumors by PET [44], it has too

many metabolic pathways, which makes it difficult to obtain a rate constant [20, 21]. Furthermore, it is difficult to image tumors that have slow uptake because of its short half-life. Among all radiolabeled amino acids, aromatic amino acids are more suitable due to the easier chemistry alteration and characterization. However, aromatic amino acids tend to decarboxylate [45], which reduce its ability to get into cells via amino acid transporters. To circumvent these potential limitations, placement of a methyl group at the alpha position could retard metabolism, and hence, greater effort could be then directed toward the  $^{18}\text{F}$ - and  $^{123/124}\text{I}$ -labeled amino acids such as 2- $^{18}\text{F}$  fluoromethyl-L-phenylalanine [46], position 2- and 3-L- $^{18}\text{F}$  flour- $\alpha$ -methyl tyrosine (FAMT) [47–49], and  $^{123}\text{I}$ -labeled L-3- $^{123}\text{I}$  iodo- $\alpha$ -methyltyrosine ( $^{123}\text{I}$ -AMT) [50, 51]. L-type amino acid transporter system (LAT), especially LAT1 subtype, is the only system that can transport large neutral amino acids with aromatic rings such as tyrosine, phenylalanine, and tryptophan [52, 53]. LAT family is known to form heterodimers, which contain a chaperone-like heavy chain 4F2hc, and a 12-time-folding transmembrane light chain, which is unique to each subtype [54]. Previous clinical studies showed that uptake of FAMT in the tumor was closely correlated with LAT1 expression and cellular proliferation [41, 48]. Its analog,  $^{123}\text{I}$ -AMT, has also been used for SPECT studies on brain and pancreatic tumors [55–57]. High accumulation of  $^{123}\text{I}$ -AMT in tumors was reported upon PET examination with  $^{124}\text{I}$ -AMT, which was carried out in patients with brain tumors [57]. Though AMT shows very promising clinical results, the cost of using such isotope is prohibitive. Therefore, it is desirable to develop a radiotracer with easier chemistry and more affordable isotope that can be used clinically in most major medical facilities. Few promising chelator–amino acid conjugates in animal studies have been reported [58–60].

## 12.4 Estrogen Receptor Target

The presence of sex hormone receptors in both primary and secondary breast tumors is an important indicator for both prognosis and choice of therapy of the disease. Currently, receptors are determined by in vitro analysis of biopsy specimens and the use of antiestrogen antibodies. Tamoxifen is the therapy of choice for estrogen receptor-positive (ER+) tumors. The detection and measurement of ER+ tumors by the use of a radiolabeled ligand should provide a useful tool for the diagnosis of primary and secondary tumors. This approach may assist in selecting and following the most favorable therapy, as well as predicting its outcome. To this end a number of variations of substituted estradiol [61, 62] and tamoxifen [63] have been prepared. These compounds have been relatively successful in detecting ER-rich tissue in vivo, such as breast cancer, ovarian cancer, endometriosis, uterine carcinoma, and meningioma. These compounds may predict the response to anticancer therapy agents.

## 12.5 Tumor Hypoxia Target

The success to the noninvasive detection of the hypoxic fraction of tumors by nuclear molecular imaging is that it allows physicians to select patients for additional or alternative treatment regimens that may circumvent or overcome the ominous impact of tumor hypoxia and improve disease control [64]. Misonidazole (MISO) is a hypoxic cell sensitizer, and labeling it with different radioisotopes could be useful for differentiating a hypoxic but metabolically active tumor from a well-oxygenated active tumor by PET or planar scintigraphy. The nitro group of nitroimidazole (NIM) is enzymatically reduced by ribonucleoside reductase within viable hypoxic cells [65, 66].  $^{18}\text{F}$ -FMISO and clinically user-friendly chelator-based hypoxia tracers have been used to assess the hypoxic component in brain ischemia, myocardial infarction, and various tumors [51, 67]. Moreover, the assessment of tumor hypoxia prior to radiation therapy would provide a rational means to select patients for treatment with radiosensitizing or bioreductive drugs (e.g., mitomycin C). It is also possible to select proper modalities of radiotherapy (proton, neutron vs. photon) by correlating tumor uptake results with tumor response.

## 12.6 Lipid Metabolism Target

An elevated level of phosphatidylcholine has been found in tumors, which is the most abundant phospholipid in the cell membranes of all eukaryotic cells and provides a potential target for tumor imaging. This elevation is thought to be the result of increased uptake of choline, a precursor of the biosynthesis of phosphatidylcholine. Malignant tumors show a high proliferation and increased metabolism of cell membrane components that will lead to an increased uptake of choline [54]. Thus, radiolabeled choline can be used as a PET marker for imaging cell proliferation in prostate cancer, brain tumors, and many other types of tumors that lack the urinary radioactivity seen with  $^{18}\text{F}$ -FDG [68, 69].

## 12.7 Tumor Cell Proliferation Target

Noninvasive imaging assessment of tumor cell proliferation could be helpful in the evaluation of tumor growth potential and the degree of malignancy and treatment response in the early assessment, prior to changes in tumor size. Radiolabeled nucleoside/nucleotide analogs should provide proliferative imaging information of primary and secondary tumors. They may also assist in selecting and following the most favorable choice of nucleoside/nucleotide therapy and in following its outcome.  $^{18}\text{F}$ -Fluorothymidine (FLT) was developed in an attempt to improve the understanding of the biologic behavior of malignant tumors, which should lead to

better prognostic evaluation, treatment follow-up, and patient management. However,  $^{18}\text{F}$ -FLT shows a lower accumulation in tumors than  $^{18}\text{F}$ -FDG, since it only accumulates in the cells that are in the S phase of the cell cycle. This demonstrates a low sensitivity for nodal staging [70], which indicates that there is still room to develop better markers of cell proliferation biomarkers.

## 12.8 Gene Expression Target

Radiolabeled pyrimidine and purine probes to image herpes simplex virus type 1 thymidine kinase (HSV-1-*tk*) expression and other reporter genes by PET have been developed [71–73]. For example, pyrimidinenucleoside (e.g., FIAU, 2'-fluoro-2'-deoxy-5-iodo-1- $\beta$ -D-ribofuranosyl-uracil[FIRU], 2'-fluoro-2'-5-methyl-1- $\beta$ -D-arabinofuranosyl-uracil [FMAU], 2'-fluoro-2'-deoxy-5-iodovinyl-1- $\beta$ -D-ribofuranosyluracil [IVFRU]), acycloguanosine([9-[(2-hydroxy-1-(hydroxymethyl)ethoxy)methyl]-guanine (GCV) and 9-[4-hydroxy-3-(hydroxymethyl)butyl] guanine (PCV)]), and other  $^{18}\text{F}$ -labeled acycloguanosine analogs such as 8-fluoro-9-[(2-hydroxy-1-(hydroxymethyl)ethoxy)methyl]guanine (FGCV), 8-fluoro-9-[4-hydroxy-3-(hydroxymethyl)guanine (FHBG), were developed as reporter substrates to image wild-type and mutant HSV-1-*tk* expression. Recently, imaging, pharmacokinetics, and dosimetry of  $^{18}\text{F}$ -FHBG were reported in healthy volunteers as a first step to image the HSV-1-*tk* reporter expression in clinical gene therapy trials. The difficulty with these probes is that HSV-1-*tk* enzyme expression depends on HSV-1-*tk* gene transduction with adenoviral vectors. The level of HSV-1-*tk* enzyme expression is likely to be different in diverse types of transduced cells and tissues, thus limiting the application. Understanding the proliferative activity of tumors could aid in the selection of optimal therapy, by estimating patient prognosis and selecting the proper management.

In summary, in order to improve the diagnosis, prognosis, planning, and monitoring of the cancer treatment, it is important to characterize tumor tissue extensively by the development and application of more tumor-specific pharmaceuticals. Radiolabeled ligands as well as radiolabeled antibodies have opened a new era in scintigraphy detection of tumors and have undergone extensive preclinical evaluation.

## References

1. Rubi S, Costes N, Heckemann RA, et al. Positron emission tomography with alpha-[ $^{11}\text{C}$ ]methyl-L-tryptophan in tuberous sclerosis complex-related epilepsy. *Epilepsia*. 2013;54(12):2143–50.
2. Park HK, Kim JS, Im KC, Kim MJ, Lee JH, Lee MC, Kim J, Chung SJ. Visual hallucinations and cognitive impairment in Parkinson's disease. *Can J Neurol Sci* 2013;40(5):657–662.
3. Bertelson JA, Ajtai B. Neuroimaging of dementia. *Neurol Clin*. 2014;32(1):59–93.

4. Chetelat G, La Joie R, Villain N, Perrotin A, de La Sayette V, Eustache F, Vandenberghe R. Amyloid imaging in cognitively normal individuals, at-risk populations and preclinical Alzheimer's disease. *Neuroimage Clin.* 2013;2:356–65.
5. Lehner S, Uebles C, Schussler F, et al. The amount of viable and dyssynchronous myocardium is associated with response to cardiac resynchronization therapy: initial clinical results using multiparametric ECG-gated [18F]FDG PET. *Eur J Nucl Med Mol Imaging.* 2013;40(12):1876–83.
6. Friedberg JW, Fischman A, Neuberger D, et al. FDG-PET is superior to gallium scintigraphy in staging and more sensitive in the follow-up of patients with de novo Hodgkin lymphoma: a blinded comparison. *Leuk Lymphoma.* 2004;45(1):85–92.
7. Bar-Shalom R, Valdivia AY, Blafox MD. PET imaging in oncology. *Semin Nucl Med.* 2000;30(3):150–85.
8. Plowman PN, Saunders CA, Maisey M. On the usefulness of brain PET scanning to the paediatric neuro-oncologist. *Br J Neurosurg.* 1997;11(6):525–32.
9. Weber WA, Avril N, Schwaiger M. Relevance of Positron Emission Tomography (PET) in oncology. *Strahlenther Onkol.* 1999;175(8):356–73.
10. Lau CL, Jr Harpole DH, Patz E. Staging techniques for lung cancer. *Chest Surg Clin N Am.* 2000;10(4):781–801.
11. Schulte M, Brecht-Krauss D, Heymer B, et al. Grading of tumors and tumorlike lesions of bone: evaluation by FDG PET. *J Nucl Med.* 2000;41(10):1695–701.
12. Yutani K, Shiba E, Kusuoka H, Tatsumi M, Uehara T, Taguchi T, Takai SI, Nishimura T. Comparison of FDG-PET with MIBI-SPECT in the detection of breast cancer and axillary lymph node metastasis. *J Comput Assist Tomogr.* 2000;24(2):274–80.
13. Franzius C, Sciuk J, Daldrup-Link HE, Jurgens H, Schober O. FDG-PET for detection of osseous metastases from malignant primary bone tumours: comparison with bone scintigraphy. *Eur J Nucl Med.* 2000;27(9):1305–11.
14. Folpe AL, Lyles RH, Sprouse JT, Conrad ER, Eary JF. (F-18) fluorodeoxyglucose positron emission tomography as a predictor of pathologic grade and other prognostic variables in bone and soft tissue sarcoma. *Clin Cancer Res.* 2000;6(4):1279–87.
15. Meyer PT, Spetzger U, Mueller HD, Zeggel T, Sabri O, Schreckenberger M. High F-18 FDG uptake in a low-grade supratentorial ganglioma: a positron emission tomography case report. *Clin Nucl Med.* 2000;25(9):694–7.
16. Franzius C, Sciuk J, Brinkschmidt C, Jurgens H, Schober O. Evaluation of chemotherapy response in primary bone tumors with F-18 FDG positron emission tomography compared with histologically assessed tumor necrosis. *Clin Nucl Med.* 2000;25(11):874–81.
17. Carretta A, Landoni C, Melloni G, Ceresoli GL, Compierchio A, Fazio F, Zannini P. 18-FDG positron emission tomography in the evaluation of malignant pleural diseases – a pilot study. *Eur J Cardiothorac Surg.* 2000;17(4):377–83.
18. Torre W, Garcia-Veloso MJ, Galbis J, Fernandez O, Richter J. FDG-PET detection of primary lung cancer in a patient with an isolated cerebral metastasis. *J Cardiovasc Surg.* 2000;41(3):503–5.
19. Brunelle F. Noninvasive diagnosis of brain tumours in children. *Childs Nerv Syst.* 2000;16(10–11):731–4.
20. Mankoff DA, Dehdashti F, Shields AF. Characterizing tumors using metabolic imaging: PET imaging of cellular proliferation and steroid receptors. *Neoplasia.* 2000;2(1–2):71–88.
21. Fitzgerald J, Parker JA, Danias PG. F-18 fluoro deoxyglucose SPECT for assessment of myocardial viability. *J Nucl Cardiol.* 2000;7(4):382–7.
22. Schwarz A, Kuwert T. Nuclear medicine diagnosis in diseases of the central nervous system. *Radiologe.* 2000;40(10):858–62.
23. Roelcke U, Leenders KL. PET in neuro-oncology. *J Cancer Res Clin Oncol.* 2001;127(1):2–8.
24. Buerkle A, Weber WA. Imaging of tumor glucose utilization with positron emission tomography. *Cancer Metastasis Rev.* 2008;27(4):545–54.

25. Delbeke D, Coleman RE, Guiberteau MJ, et al. Procedure guideline for tumor imaging with 18F-FDG PET/CT 1.0. *J Nucl Med.* 2006;47(5):885–95.
26. Rosenbaum SJ, Lind T, Antoch G, Bockisch A. False-positive FDG PET uptake—the role of PET/CT. *Eur Radiol.* 2006;16(5):1054–65.
27. Brock CS, Meikle SR, Price P. Does fluorine-18 fluorodeoxyglucose metabolic imaging of tumours benefit oncology? *Eur J Nucl Med.* 1997;24(6):691–705.
28. Chang JM, Lee HJ, Goo JM, Lee HY, Lee JJ, Chung JK, Im JG. False positive and false negative FDG-PET scans in various thoracic diseases. *Korean J Radiol.* 2006;7(1):57–69.
29. Yang DJ, Kim CG, Schechter NR, et al. Imaging with 99mTc ECDG targeted at the multifunctional glucose transport system: feasibility study with rodents. *Radiology.* 2003;226(2):465–73.
30. Schechter NR, Erwin WD, Yang DJ, et al. Radiation dosimetry and biodistribution of (99 m) Tc-ethylene dicysteine-deoxyglucose in patients with non-small-cell lung cancer. *Eur J Nucl Med Mol Imaging.* 2009;36(10):1583–91.
31. Zhang YH, Bryant J, Kong FL, Yu DF, Mendez R, Edmund KE, Yang DJ. Molecular imaging of mesothelioma with (99 m)Tc-ECG and (68)Ga-ECG. *J Biomed Biotechnol.* 2012;2012:232863.
32. Yang DJ, Kong FL, Oka T, Bryant JL. Molecular imaging kits for hexosamine biosynthetic pathway in oncology. *Curr Med Chem.* 2012;19(20):3310–4.
33. Hanover JA, Krause MW, Love DC. The hexosamine signaling pathway: O-GlcNAc cycling in feast or famine. *Biochim Biophys Acta.* 2010;1800(2):80–95.
34. Hart GW, Slawson C, Ramirez-Correa G, Lagerlof O. Cross talk between O-GlcNAcylation and phosphorylation: roles in signaling, transcription, and chronic disease. *Annu Rev Biochem.* 2011;80:825–58.
35. Lynch TP, Ferrer CM, Jackson SR, Shahriari KS, Vosseller K, Reginato MJ. Critical role of O-Linked beta-N-acetylglucosamine transferase in prostate cancer invasion, angiogenesis, and metastasis. *J Biol Chem.* 2012;287(14):11070–81.
36. Krzeslak A, Forma E, Bernaciak M, Romanowicz H, Brys M. Gene expression of O-GlcNAc cycling enzymes in human breast cancers. *Clin Exp Med.* 2012;12(1):61–5.
37. Ozcan S, Andrali SS, Cantrell JE. Modulation of transcription factor function by O-GlcNAc modification. *Biochim Biophys Acta.* 2010;1799(5–6):353–64.
38. Lewis BA, Hanover JA. O-GlcNAc and the epigenetic regulation of gene expression. *J Biol Chem.* 2014;289(50):34440–8.
39. Golks A, Tran TT, Goetschy JF, Guerini D. Requirement for O-linked N-acetylglucosaminyltransferase in lymphocytes activation. *EMBO J.* 2007;26(20):4368–79.
40. Pham LV, Tamayo AT, Yoshimura LC, Lin-Lee YC, Ford RJ. Constitutive NF-kappaB and NFAT activation in aggressive B-cell lymphomas synergistically activates the CD154 gene and maintains lymphoma cell survival. *Blood.* 2005;106(12):3940–7.
41. Urakami T, Sakai K, Asai T, Fukumoto D, Tsukada H, Oku N. Evaluation of O-[(18)F] fluoromethyl-D-tyrosine as a radiotracer for tumor imaging with positron emission tomography. *Nucl Med Biol.* 2009;36(3):295–303.
42. Langen KJ, Muhlensiepen H, Holschbach M, Hautzel H, Jansen P, Coenen HH. Transport mechanisms of 3-[123I]iodo-alpha-methyl-L-tyrosine in a human glioma cell line: comparison with [3H]methyl-L-methionine. *J Nucl Med.* 2000;41(7):1250–5.
43. Miyagawa T, Oku T, Uehara H, Desai R, Beattie B, Tjuvajev J, Blasberg R. “Facilitated” amino acid transport is upregulated in brain tumors. *J Cereb Blood Flow Metab.* 1998;18(5):500–9.
44. Syrota A, Comar D, Cerf M, Plummer D, Maziere M, Kellershohn C. [11C]methionine pancreatic scanning with positron emission computed tomography. *J Nucl Med.* 1979;20(7):778–81.
45. Lindstrom LH, Gefvert O, Hagberg G, Lundberg T, Bergstrom M, Hartvig P, Langstrom B. Increased dopamine synthesis rate in medial prefrontal cortex and striatum in schizophrenia indicated by L-(beta-11C) DOPA and PET. *Biol Psychiatry.* 1999;46(5):681–8.



46. Kersemans K, Bauwens M, Mertens J. Method for stabilizing non carrier added 2-[(18)F] fluoromethyl-L-phenylalanine, a new tumour tracer, during radiosynthesis and radiopharmaceutical formulation. *Nucl Med Biol.* 2008;35(4):425–32.
47. Inoue T, Shibasaki T, Oriuchi N, et al. 18F alpha-methyl tyrosine PET studies in patients with brain tumors. *J Nucl Med.* 1999;40(3):399–405.
48. Kaira K, Oriuchi N, Otani Y, et al. Fluorine-18-alpha-methyltyrosine positron emission tomography for diagnosis and staging of lung cancer: a clinicopathologic study. *Clin Cancer Res.* 2007;13(21):6369–78.
49. Yamaura G, Yoshioka T, Fukuda H, Yamaguchi K, Suzuki M, Furumoto S, Iwata R, Ishioka C. O-[18F]fluoromethyl-L-tyrosine is a potential tracer for monitoring tumour response to chemotherapy using PET: an initial comparative in vivo study with deoxyglucose and thymidine. *Eur J Nucl Med Mol Imaging.* 2006;33(10):1134–9.
50. Langen KJ, Pauleit D, Coenen HH. 3-[(123)I]Iodo-alpha-methyl-L-tyrosine: uptake mechanisms and clinical applications. *Nucl Med Biol.* 2002;29(6):625–31.
51. Ali MS, Kong FL, Rollo A, Mendez R, Kohanim S, Smith DL, Yang DJ. Development of (99 m)Tc-N4-NIM for molecular imaging of tumor hypoxia. *J Biomed Biotechnol.* 2012;2012:828139.
52. Uchino H, Kanai Y, Kim DK, Wempe MF, Chairoungdua A, Morimoto E, Anders MW, Endou H. Transport of amino acid-related compounds mediated by L-type amino acid transporter 1 (LAT1): insights into the mechanisms of substrate recognition. *Mol Pharmacol.* 2002;61(4):729–37.
53. Yanagida O, Kanai Y, Chairoungdua A, et al. Human L-type amino acid transporter 1 (LAT1): characterization of function and expression in tumor cell lines. *Biochim Biophys Acta.* 2001;1514(2):291–302.
54. del AE, Urti A, Yliperttula M. Pharmacokinetic role of L-type amino acid transporters LAT1 and LAT2. *Eur J Pharm Sci* 2008;35(3):161–174.
55. Tisljar U, Kloster G, Ritzl F, Stocklin G. Accumulation of radioiodinated L-alpha-methyltyrosine in pancreas of mice: concise communication. *J Nucl Med.* 1979;20(9):973–6.
56. Kloss G, Leven M. Accumulation of radioiodinated tyrosine derivatives in the adrenal medulla and in melanomas. *Eur J Nucl Med.* 1979;4(3):179–86.
57. Langen KJ, Coenen HH, Roosen N, et al. SPECT studies of brain tumors with L-3-[123I] iodo-alpha-methyl tyrosine: comparison with PET, 124IMT and first clinical results. *J Nucl Med.* 1990;31(3):281–6.
58. Kong FL, Zhang Y, Young DP, Yu DF, Yang DJ. Development of (99 m)Tc-EC-tyrosine for early detection of breast cancer tumor response to the anticancer drug melphalan. *Acad Radiol.* 2013;20(1):41–51.
59. Kong FL, Zhang Y, Ali MS, et al. Synthesis of (99 m)Tc-EC-AMT as an imaging probe for amino acid transporter systems in breast cancer. *Nucl Med Commun.* 2010;31(8):699–707.
60. Kong FL, Ali MS, Zhang Y, Oh CS, Yu DF, Chanda M, Yang DJ. Synthesis and evaluation of amino acid-based radiotracer 99mTc-N4-AMT for breast cancer imaging. *J Biomed Biotechnol.* 2011;2011:276907.
61. Yoshida Y, Kurokawa T, Sawamura Y, Shinagawa A, Okazawa H, Fujibayashi Y, Kotsuji F. The positron emission tomography with F18 17beta-estradiol has the potential to benefit diagnosis and treatment of endometrial cancer. *Gynecol Oncol.* 2007;104(3):764–6.
62. Takahashi N, Yang DJ, Kurihara H, Borne A, Kohanim S, Oh CS, Mawlawi O, Kim EE. Functional imaging of estrogen receptors with radiolabeled-GAP-EDL in rabbit endometriosis model. *Acad Radiol.* 2007;14(9):1050–7.
63. Yang DJ, Li C, Kuang LR, et al. Imaging, biodistribution and therapy potential of halogenated tamoxifen analogues. *Life Sci.* 1994;55(1):53–67.
64. Brown JM. The hypoxic cell: a target for selective cancer therapy—eighteenth Bruce F. Cain Memorial Award lecture. *Cancer Res.* 1999;59(23):5863–70.

65. Chu T, Li R, Hu S, Liu X, Wang X. Preparation and biodistribution of technetium-99 m-labeled 1-(2-nitroimidazole-1-yl)-propanhydroxyiminoamide (N2IPA) as a tumor hypoxia marker. *Nucl Med Biol.* 2004;31(2):199–203.
66. Seddon BM, Maxwell RJ, Honess DJ, Grimshaw R, Raynaud F, Tozer GM, Workman P. Validation of the fluorinated 2-nitroimidazole SR-4554 as a noninvasive hypoxia marker detected by magnetic resonance spectroscopy. *Clin Cancer Res.* 2002;8(7):2323–35.
67. Horsman MR, Mortensen LS, Petersen JB, Busk M, Overgaard J. Imaging hypoxia to improve radiotherapy outcome. *Nat Rev Clin Oncol.* 2012;9(12):674–87.
68. Hara T, Kosaka N, Kishi H. Development of (18)F-fluoroethylcholine for cancer imaging with PET: synthesis, biochemistry, and prostate cancer imaging. *J Nucl Med.* 2002;43(2):187–99.
69. DeGrado TR, Baldwin SW, Wang S, et al. Synthesis and evaluation of (18)F-labeled choline analogs as oncologic PET tracers. *J Nucl Med.* 2001;42(12):1805–14.
70. Szyszko TA, Yip C, Szlosarek P, Goh V, Cook GJ. The role of new PET tracers for lung cancer. *Lung Cancer-J Iaslc.* 2016;94:7–14.
71. Gambhir SS, Barrio JR, Phelps ME, et al. Imaging adenoviral-directed reporter gene expression in living animals with positron emission tomography. *Proc Natl Acad Sci U S A.* 1999;96(5):2333–8.
72. Alauddin MM, Shahinian A, Kundu RK, Gordon EM, Conti PS. Evaluation of 9-[(3-18F-fluoro-1-hydroxy-2-propoxy)methyl]guanine ([18F]-FHPG) in vitro and in vivo as a probe for PET imaging of gene incorporation and expression in tumors. *Nucl Med Biol.* 1999;26(4):371–6.
73. Yaghoubi S, Barrio JR, Dahlbom M, et al. Human pharmacokinetic and dosimetry studies of [(18)F]FHBG: a reporter probe for imaging herpes simplex virus type-1 thymidine kinase reporter gene expression. *J Nucl Med.* 2001;42(8):1225–34.