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# Development of Cancer Vaccine and Targeted Immune Checkpoint Therapies

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## 16.1 Introduction

The immune system's natural capacity to detect and destroy abnormal cells may prevent the development of many cancers [1, 2]. However, cancer cells are capable of evading detection and destruction by the immune system. They create a heterogeneous environment to favor or facilitate their progression, the so-called tumor microenvironment (TME) [3–5]. Besides tumor cells, the TME comprises many different stromal cells. These include vascular or lymphatic endothelial cells, supporting pericytes, fibroblasts, and infiltrating immune cells. These nonimmune stromal cells provide support to tumor cells, with growth factors and cytokines, and promote angiogenesis, tissue invasion, and metastasis [6]. In addition, the stroma provides a chemoresistant barrier to the tumor, preventing chemotherapeutics from reaching their targets [7].

The major immune cells at TME include myeloid-derived suppressor cells (MDSCs), tumor-associated macrophages (TAMs), dendritic cells (DCs), natural killer (NK) cells, and T and B lymphocytes [8, 9]. Generally, immune cells can exert both tumor suppressive and promoting effects [10]. T lymphocytes have a paramount role in tumor-specific cellular adaptive immunity. The main components of this population are CD8+ cytotoxic T lymphocytes (CTLs), CD4+ helper T cells, and regulatory T cells ( $T_{reg}$ ). CD8+ CTLs are the major cell type that can directly kill cancer cells, and their presence is associated with prolonged survival. However, most CD8+ T cells at tumor sites exhibit dysfunctional or exhausted phenotypes and are reluctant to proliferate [11]. The presence of Th1 and Th2 lymphocytes in the tumor microenvironment appears to have opposite prognostic significance in the setting of tumor progression [12]. DCs are important for

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antigen presentation and T cell activation during antitumor immunity. However, the immunosuppressive TME always turns DCs into a suppressive or regulatory DC phenotype [13]. T<sub>reg</sub> cells, which are positive for CD4+, CD25+, and Foxp3, are enriched in the tumor microenvironment [14]. T<sub>reg</sub> cells effectively suppress the adaptive immune response, and their presence in the tumor microenvironment leads to decreased anticancer immunity and often correlates with poor prognosis [14]. TAMs are polarized macrophages with a protumoral phenotype; they suppress antitumor T cell responses, and promote tumor angiogenesis and metastasis [15]. MDSCs are mobilized during tumorigenesis, and infiltrate developing tumors where they promote tumor vascularization and disrupt major mechanisms of immunosurveillance by T cells, DCs, and NK cells [16, 17]. Neutrophils can play both tumor-promoting and tumoricidal functions, depending on their differentiation status and the presence of TGF- $\beta$  [18]. The role of B cells in tumor immunity remains unclear: some reports showed that B cell depletion promotes antitumor immune responses while some studies found that activated B cells increase T cell activation and suppress tumor growth [19].

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## 16.2 A Unique Immunosuppressive Microenvironment of Pancreatic Cancer

Pancreatic cancers present an enormous challenge, as they are insensitive to traditional therapies. One prime contributing factor is the uniquely abundant tumor stromal content present in the microenvironment of pancreatic cancer [20–22]. The epithelial and stromal compartments interact and communicate to enhance the aggressive nature of this disease, ultimately culminating in an extremely effective immunosuppressive network [23]. Pancreatic cancer cells release various factors that stimulate the formation of stroma. Stromal cells, in turn, release mutagenic substances that stimulate tumor growth, invasion, and resistance to therapy. Structurally, the presence of an enormous number of stromal cells forms a physical shield, preventing immune cells from reaching and attacking cancer cells [24, 25]. Furthermore, pancreatic cancer cells utilize multiple pathways to create an immunosuppressive microenvironment and evade immune cell attack. Several cytokines appear to be dysregulated and contribute to cancer progression in pancreatic ductal adenocarcinoma (PDAC). In particular, higher levels of circulating interleukin-6 (IL-6) are identified in patients with PDAC and appear to promote cancer progression through enhancement of pro-tumorigenic Stat3 signaling [26, 27]. Furthermore, members of the IL-1 family (e.g., IL- $\alpha$ , IL- $\beta$ , and IL-1 receptor antagonist (IL-1ra)) seem to play a role in PDAC development [28, 29]. Immunosuppressive cytokine IL-10 is upregulated in PDAC, which leads to a reduction in effector cell function in the PDAC microenvironment and indicates a worse prognosis [30]. Finally, pancreatic cancer is a non-immunogenic cancer type with low frequency of mutations [31]. As a result, the frequency of tumor-specific T cells at cancer sites is relatively low, and intraepithelial CD8<sup>+</sup> T cells infiltration is very rare in PDAC [23]. This poses a great challenge to active immunotherapies, such as cancer vaccines and immune checkpoint inhibitors, which would rely on the existing anticancer immunity in cancer patients. Therefore, a better understanding of the

complex interactions between the cancer cells and their associated stromal cells could be key to the development of new therapeutic options for patients [32].

### 16.3 Principles for Cancer Immunotherapy

The immune system is capable of detecting carcinogenesis though the extent and efficiency of anticancer effect are generally not strong enough to eradicate established cancer [1]. Therefore, the strategies of cancer immunotherapy are to launch a strong anticancer response by mobilizing endogenous anticancer immunity or by infusing immune effector cells to combat cancer. Based on the reliance of the existing immune system, the approaches of immunotherapy can be classified into two types: passive and active (Table 16.1) [51]. Passive immunotherapy comprises antibodies and immune cells that are made outside of the body and are subsequently

**Table 16.1** Major immunotherapeutic approaches in pancreatic cancer

Type of immunotherapy		Passive or active	Example	Advantages	Disadvantage	References
Adaptive cellular therapy	TIL	Passive	N/A	Limited TILs for <i>in vitro</i> expansion	A personalized approach and costly	N/A
	CAR-T	Passive	Mesothelin	Can be produced in large scale	*A costly personalized approach *Tumor-specific targets yet to be found *Target limited tumor antigens	[33]
Cancer vaccine	Peptide	Active	MUC-1, survivin, telomerase, Ras mutant, VEGFR2	Low cost, high specificity	*Derived from weak antigen (TAA) *Neoantigen yet to be identified * Target limited tumor epitopes	[34]
	DC-based			A good inducer of tumor-specific T cells		[35–38]
	Whole Cell		GVAX, Algenpantucel-L	*Easy to manufacture *Multiple and unknown tumor antigens targeted.	Trigger weak anticancer immunity	[39–48]
Checkpoint inhibitor		Active	CTLA-4, PD-1	*Target a broad spectrum of tumor antigens *Does not need the knowledge of antigen identity	The anticancer effect relies on the immunogenicity of cancer	[49, 50]

inoculated into cancer patients, in an attempt to target and destroy cancer cells. It includes but it is not limited to antibody and adaptive cellular therapy (ACT) [52]. On the other end of the spectrum, active immunotherapy interventions aim to trigger or amplify anticancer immunity by mobilizing the host immune system and include at least cancer vaccines and immune checkpoint inhibitors.

The following sections will summarize current major immunotherapy development in research and clinical trials, and their progresses in pancreatic cancer therapy.

### 16.3.1 Adaptive Cellular Therapy

Adaptive cellular therapy (ACT) is a procedure that aims to first expand T cells *in vitro* and then re-infuse the expanded T cell pool back into patients for cancer treatment [53]. Compared to peripheral blood of cancer patients, tumor infiltrated lymphocytes (TILs) are enriched in tumor-specific T cells and can be easily expanded *in vitro* by tumor cells with the presence of growth factors like interleukin-2 [54, 55]. This practice can generate tumor-reactive T cells with a broad range of tumor reactivity, without the knowledge of tumor antigen identities. With the improvement of culturing technology, the degree of expansion and quality control has been greatly enhanced. Isolating and expanding TILs for ACT is a very efficacious treatment strategy in melanoma [56]. However, the number of TILs that can be successfully recovered from the vast majority of solid tumors is very limited, especially for those cancers with few TILs. In addition, the majority of TILs display exhausted or dysfunctional phenotypes, which might cause the poor persistence of expansion tumor-specific T cell clones upon intravenous infusion [57, 58]. Therefore, the current approach of expanding TILs for ACT is mainly practiced in melanoma patients.

Genetically, engineering of lymphocytes is a new approach that aims to eliminate the obstacle posed by many tumors with a limited number of tumor-reactive T cells for ACT [52, 59, 60]. This strategy involves transducing immune cells with genes that redirect T cells to recognize cancer cells. The specificity of T cells can be redirected by the incorporation of genes encoding either conventional alpha-beta TCRs or chimeric antigen receptors (CARs) [61]. In this case, T cells from patient blood can be directly used as a source for ACT. CARs are constructed by linking the variable regions of the antibody heavy and light chains to intracellular signaling chains, such as CD3-zeta. The new generation of CARs is also composed of costimulatory domains of CD28 and/or CD137 to promote T cell expansion [62]. Because CARs are derived from antibodies, recognition of tumor-associated antigens (TAAs) by CARs is strong and is not restricted by major histocompatibility complex (MHC) [62].

However, a major hurdle for CAR-T therapy in human cancer is the identification requirement of appropriate tumor antigens that are exclusively expressed on the cancer cells, but not normal self-tissues. Most of the currently identified tumor-specific antigens are self-antigens that are normally expressed in early fetal development and that are aberrantly expressed during malignancy [63]. Examples include

NY-ESO1 and the MAGE family antigens. The phenomenon known as “off-tumor, on-target,” where CAR-T cells recognize non-cancer cells expressing the tumor antigen, is responsible for severe immune-mediated toxicities that have limited the applicability of this treatment strategy [64, 65]. Therefore, careful monitoring and screening of targets for CAR-T therapy is extremely important. As a result, current successes of CAR-T therapy in clinic are mainly limited to certain types of lymphoma/leukemia [66]. Testing the feasibility of this approach can only be carried out in clinical trials, as preclinical models have proven to be insufficiently predictive of both efficacy and toxicity in humans. Whole-genome sequencing of cancer cells is generating abundant information about specific mutations in tumor cells, which may lead to the identification of tumor-specific antigens, also called neoantigens [67]. Innovative ways of generating antigen receptors that recognize these, including CARs that directly recognize intracellular molecules presented by MHC, may generate T cells with even greater specificities for tumor cells. It is worth noting that another potential obstacle for ACT is that the majority of the inoculated T cells die before reaching the cancer site, which can be a challenging obstacle for patients with solid cancers [68]. Therefore, selection of T cell subsets with better capacity for survival and proliferation is a critical step in ACT, and methods to selectively enrich central memory and stem cell memory T cell subsets from human lymphocytes may enable more effective anticancer responses in humans, similar to those observed in mouse models [68–70]. Although the range of CARs currently available is sufficient to cover most types of malignancy, tumor cells can lose the expression of TAAs to evade immune attack during ACT [60]. Therefore, using several CAR genes that target multiple TAAs simultaneously may be needed for future ACT to better accommodate the heterogeneity in human cancers.

Animal models of pancreatic cancer have shown encouraging results with the use of ACT [71, 72], and clinical trials using CAR engineered T cells for pancreatic cancer are currently ongoing in many cancer centers (NCT01897415, NCT02465983) [33]. The recently completed PDAC genomic analysis by Bailey et al. led to a deeper understanding of the molecular evolution of PDAC and to the identification of a specific immunogenic PDAC subtype [73]. This new and long awaited information may open the way to new and more accurate therapeutic targets for ACT.

### 16.3.2 Cancer Vaccines

Vaccine is an active therapeutic approach aiming to mobilize the immune system to generate or amplify tumor-specific immune response to combat cancer [74]. The primary mechanism for therapeutic cancer vaccines is to increase the presentation of tumor-associated antigens (TAAs) to the immune system, so as to mount a potent immune response against tumors. Cancer vaccines attempt to copy the achievements made in vaccinations against pathogens though more work is necessary to bring it to fruition. Based on the formats utilized, cancer vaccine can be classified into three major categories: protein/peptide vaccines, whole cell vaccines, and DNA vaccines [51].

### 16.3.2.1 Peptide Vaccines

Protein/peptide vaccines attempt to immunize patients with a peptide or a protein derived from cancer antigens in the formation of adjuvant or cellular vehicles. This strategy requires the identification of tumor-specific antigens or TAAs that are only expressed on cancer cells or overexpressed on cancer cells.

Peptide vaccine therapy for PDAC has been conducted in clinic for many years [34]. The most commonly used antigens in trials include telomerase, Wilms tumor gene, KIP20A, survivin, mutated Ras protein, mucin MUC1 protein, and vascular endothelial growth factor receptor 2 (VEGFR2). Though overall cancer vaccine is well tolerated, the outcomes of these vaccine trials have been disappointing with many lessons learned [34]. First, the presence of suppressive mechanisms at the cancer sites must be conquered. Immunoconditioning can eliminate some of these immunosuppressive mechanisms, but at the same time it also dampens endogenous anticancer immunity that is needed for cancer vaccines. Examples of cells responsible for this suppressive mechanism include  $T_{reg}$  cells, MDSCs, as well as the signal generated through the interaction between PD1 and PD-L1 at the cancer site [75, 76]. Second, the antigen/peptides used in trials are mainly tumor-associated antigens (TAAs), which may be well tolerated and thus incapable of triggering anticancer immunity strong enough to destroy PDAC [77]. Emerging data in clinical immunotherapy suggest that the recognition and response to neoantigens, which arise as a consequence of tumor-specific mutation, is the major player, and neoantigen loads correlate with overall response rates to therapy [67]. Recent technological advancements have made it possible to dissect the immune response to patient-specific neoantigens [78]. It remains to be seen whether a neoantigen-based vaccine is capable of triggering potent anticancer immunity for cancer therapy.

### 16.3.2.2 Whole Cell-based Vaccines

Whole cell vaccines are conceptually easy to understand as this strategy, as the name indicates, proposes to utilize the whole tumor cell to elicit a specific anticancer immune response [79]. The tumor cell can be either autologous (i.e., patient-specific tumor cell) or allogenic (i.e., established human tumor cell line). The rationale for this approach is that cancer cells express the entire spectrum of tumor antigens (i.e., for that specific tumor in that specific patient) as well as specific epitopes for CD8+ and CD4+ T cells that can be presented to the patient's immune system [80]. This approach is considered polyvalent (as it presents a wide range of tumor antigens to the immune system) and therefore, at least in theory, it is less susceptible to tumor immune evasion as seen in peptide-based vaccine (i.e., where mutation of TAAs under selective pressure leads to loss of the immune target). In addition, this approach is applicable to cancers even without the knowledge of antigen identity [80]. In the autologous approach, tumor cells are required to be isolated from the patient, irradiated, combined with an immunostimulating agent and ultimately infused back into the patient [79]. Therefore, this technique is limited by the availability of sufficient tumor sample that at times can be difficult to obtain, especially in certain cancer types. In this case, allogenic cell lines offer a valid alternative, as they are readily available and can be produced on a large scale [81].

GVAX is an allogenic irradiated whole cell vaccine composed of two irradiated cancer cell lines (PANC 6.03 and PANC 10.05) engineered to express granulocyte macrophage-colony stimulating factor (GM-CSF) [39, 40]. GM-CSF is a potent cytokine that functions to promote the growth of granulocytes and monocytes and also to attract dendritic cells for better antigen presentation. GVAX alone or in combination with other therapies has been investigated in multiple phase I and II studies [41–43]. A phase I trial of GVAX in 14 patients with resectable pancreatic cancer showed a mean disease-free survival (DFS) of 13 months, with three patients disease-free from 25 to 30 months [44]. Though a following phase II trial of GVAX in combination with cyclophosphamide (CY) in patients with metastatic pancreatic cancer failed to show improvement of overall survival (OS), a higher rate of induced mesothelin-specific T cell responses could correlate with longer progression-free survival (PFS) and OS [41]. Similarly, a phase II study of patients with resected PDAC using GVAX plus chemoradiation displayed median DFS of 17.3 and median survival of 24.8 months. This demonstrated an association between mesothelin-specific T cell induction and improved overall survival [42]. GVAX also has been tested in combination with *Live-Attenuated Listeria monocytogenes* (CRS-207), in an attempt to use the ability of *Listeria* to stimulate both innate and adaptive immunity to ultimately boost the overall response to the cancer vaccine [45, 46]. In a recent phase II trial, the authors showed a 2-month improvement in overall survival in patients treated with GVAX–cyclophosphamide and CRS-207, compared with GVAX–cyclophosphamide (median 6 months vs. 4 months; HR 0.60;  $P = 0.02$ ) [46]. Based on that, it is anticipated that a larger study of the GVAX/CRS-207 combination on patient survival will launch soon.

Algenpantucel-L is another major whole cell cancer vaccine being developed for PDAC [47]. It is composed of two human pancreatic cancer cells expressing the enzyme alpha-1, 3-galactosyl transferase ( $\alpha$ GT) [48]. Humans lack a functional  $\alpha$ GT gene and are not tolerant to  $\alpha$ GT; therefore,  $\alpha$ GT-labeled tumor cells could lead to enhanced antitumor response, as has been demonstrated in mouse tumor models [82, 83, 84]. In an open labeled, phase II trial of algenpantucel-L with gemcitabine and 5-fluorouracil (FU) for patients with resected PDAC, 12-month DFS of 62% and OS of 86% were achieved as compared to historical controls (45% and 65%, respectively) [48]. Another positive sign was that patients with algenpantucel-L therapy experienced minimal side effects, mainly consisting of injection site pain and induration. Based upon these encouraging results, a phase III study in patients with surgically resected PDAC was launched in 2010 [NCT01072981]. Another ongoing phase III trial ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT01836432): NCT01836432) involving Algenpantucel-L in PDAC is to combine with FOLFIRINOX or gemcitabine/nab-paclitaxel, and results of the trial are expected to be released in June 2017.

### 16.3.2.3 DC and DNA Vaccine

Similar to peptide vaccine, DC and DNA vaccines require the knowledge of TAAs or neoantigens. Genetic vaccine consists of a DNA-based vaccine that aims to introduce genetic material into a live host [85]. This allows the chosen gene products to

be expressed and ultimately triggers a specific immune reaction to the gene-derived antigen. The advantage of a genetic vaccine is that it allows the expression of antigens that resemble native viral epitopes more closely than live-attenuated or killed vaccines that often alter the protein structure and antigenicity [85]. DCs are one of the most effective APCs which function to process and present antigens on MHC molecules to trigger T cell responses [86]. DC vaccines use DCs as a vehicle for peptide/DNA vaccine, and this strategy has the potential of bridging the gap between innate and adaptive immunity [87]. This approach requires isolation of patient's DC that are eventually pulsed with peptides, and finally injected back to the patient. A successful example of peptide vaccines is Sipuleucel-T, the first FDA-approved drug for the treatment of hormone refractory prostate cancer, which is capable of extending the overall survival of cancer patients [88].

Early clinical trials of PDAC patients demonstrated that DC vaccine is well tolerated and capable of inducing detectable antigen-specific immune response in patient blood though no clear clinical benefit is observed [35]. In a phase I/II study, Lepisto and colleagues evaluated the use of an MUC1 peptide pulsed autologous DC vaccine as adjuvant therapy in patients with resectable pancreatic and biliary tumor [36]. In this study, patients were followed for over 4 years and 4 out of the 12 enrolled patients (10 had pancreatic cancer) were alive and without any evidence of recurrence. Other TAAs, such as CEA and hTERT, were used for early clinical trials of DC vaccine, with only minor objective clinical responses reported [37, 38]. Because neoantigens are more immunogenic and trigger a more potent immune response in cancer patients, the future development of DC vaccine for PDAC will likely utilize neoantigen-based DC vaccine.

### 16.3.3 Immune Checkpoint Inhibitors

T cell response is largely controlled by an array of cellular surface signaling molecules, also known as cosignaling molecules [89]. Modulating these cosignaling pathways increases anticancer immunity, either through the amplification of costimulatory pathways or blockade of negative signals, also known as immune checkpoints [90]. The major immune checkpoints under clinical investigation include at least CTLA-4, PD-1, TIM-3, LAG3, and TIGIT [91, 92]. Many of the ligands for immune checkpoints are upregulated at cancer sites and contribute to the induction of tumor-specific T cell exhaustion/dysfunction at cancer sites [91, 93]. Using monoclonal antibodies or fusion proteins are the main strategy to block or amplify cosignaling pathways. The immunomodulation strategy strives to promote or liberate internal anticancer immunity in a patient with an established cancer [94]. One of the advantages of this therapeutic strategy is that immunomodulation does not require the knowledge of specific cancer antigens but rather focuses on the manipulation of known leukocyte receptors. These provide several potential therapeutic targets that are characterized by a broad spectrum of antigen diversity that could ultimately avoid the mechanism of cancer immune evasion, caused by mutation of cancer-specific antigens [95].

Targeting immune checkpoints has been a major breakthrough in cancer treatment in recent years [96]. CTLA-4 is transiently expressed on the T cell surface

upon activation and attenuates ongoing T cell response by competing ligands B7-1 (CD80) and B7-2 (CD86) with the costimulatory receptor CD28 [97, 98]. In addition, CTLA-4 also transduces a suppressive signal to T cell via the recruitment of phosphatases SHP-2 and PP2A [90]. Ipilimumab, an anti-CTLA-4 mAb, is the first FDA-approved immunotherapy drug to treat patients with late-stage melanoma [99, 100]. Administration of ipilimumab activates T cells systemically, leading to extensive antitumor immunity and therefore a survival benefit in 10–15% of patients with advanced metastatic melanoma. Furthermore, this antitumor response significantly increases overall patient survival in advanced melanoma cases [99]. However, antitumor activity is frequently accompanied by significant immune-related adverse events. PD-1 is another inducible immune checkpoint on T cells that suppresses T cell response upon interaction with its two ligands, PD-L1 (B7-H1) and PD-L2 (B7-DC) [36, 101]. The PD-1 pathway is heavily involved in the immunosuppressive cancer microenvironment: PD-1 is highly expressed in TILs while the ligand PD-L1 is found on tumor cells, tumor-associated DCs, macrophages, and fibroblasts [94, 102]. Targeting the PD-1 pathway has elicited durable antitumor responses and long-term remissions in patients with a broad spectrum of cancers. The objective response rates varies in different cancer types, with bladder cancer, melanoma, mismatch repair-deficient colorectal cancer and certain hematopoietic malignancies among the most responsive cancer types [102]. Compared to CTLA-4 blockade, the antitumor efficacy of PD-1 blockade is higher, and the side effect is significantly milder and manageable [49, 103–105]. Currently, PD-1/PD-L1 mAbs have been approved by FDA to treat late-stage melanoma, non-small cell lung cancer, and kidney cancer [102]. It is anticipated that PD-1 mAb will be approved for treating more cancer types and become the frontline therapy for future cancer treatment.

Ipilimumab alone, or in combination with peptide vaccine, did not have any clinical benefit in treating PDAC patients. In a phase II trial of 27 patients with advanced PDAC, single-agent ipilimumab failed to detect any responder by response evaluation criteria in solid tumors [50]. However, a significant delayed response in one subject of this trial suggests that this immunotherapeutic approach to PDAC deserves further exploration [50]. With tremendous success in many cancer types, early trials of anti-PD-1 mAb alone showed no effect in treating patients with advanced PDAC, though the number of patients in the study was small [49]. PD-1 mAb alone is ineffective in treating cancers with few neoantigen loads [49]. PDAC happens to be a low immunogenic cancer [31]. It is not surprising that targeting immune checkpoints alone is incapable of launching an effective anticancer immunity in PDAC patients. Therefore, additional procedures are needed to increase the number of TILs surrounding PDAC cancers, so as to prime PDAC for immune checkpoint therapy [106].

### 16.3.4 Combined Therapy

The low immunogenicity and unique stromal structure of PDAC cancer poses a great challenge for immunotherapy [22]. The disappointing outcomes in clinical trials using single-agent immunotherapy propel the launch of combinatory

approaches. Combination therapy targets more than one aspect and can be classified as the combination of two different arms of immunotherapeutic approaches, or the combination of immunotherapy with traditional therapy (chemotherapy or radiotherapy).

Examples of combinatory approaches that combine two different arms of immunotherapy could include the combination of cancer vaccine with immune checkpoint blockade or the simultaneous use of both active and passive immune therapy. Immune checkpoint inhibitors alone are not effective in the treatment of PDAC, much due to the lack of tumor-infiltrating T cells at tumor sites [23]. Cancer vaccine is known to be a very efficient method of expanding tumor-reactive T cells, while blockade of immune checkpoints will further promote antitumor immune responses at tumor sites. In fact, several preclinical studies exist that demonstrate the synergistic role of cancer vaccine therapy, which is responsible for stimulation of the immune system, and the use of immune checkpoint blockade, which allows for the unopposed effector function of cytotoxic T cells [107–109]. Consistently, clinical examination of resected PDAC tumors demonstrated that vaccine therapy can alter the immunosuppressive cancer microenvironment [106]. The majority of PDAC patients receiving GVAX vaccine had vaccine-induced intratumoral tertiary lymphoid aggregates in resected tumors, accompanied with increased intratumoral  $T_{\text{eff}}/T_{\text{reg}}$  ratios [106]. As such, a phase Ib, open-label randomized study demonstrated the feasibility and safety of an approach based on the combination of Ipilimumab with GVAX in patients with previously treated PDAC [47]. One of the most interesting aspects of this study was the difference in 12-month OS: 27% vs. 7% and the median OS: 5.7 vs. 3.6 months (HR = 0.51;  $P = 0.072$ ), respectively, for combination therapy vs. monotherapy (i.e., Ipilimumab alone).

Given that PD-1/PD-L1 blockade is safer and more effective than CTLA-4 blockade in the treatment of many cancers, it is interesting to see how the combination of GVAX with PD-1/PD-L1 blockade performs in the treatment of PDAC [104]. Interestingly, PD-L1 expression was observed in all these lymphoid aggregates in GVAX-treated PDAC patients [106]. Currently, a phase I/II study with GVAX and anti-PD-1 mAb (nivolumab) has started to recruit patients with PDAC (NCT02451982). Similarly, a randomized phase II trial of GVAX and CRS-207 with or without nivolumab has also launched (NCT02243371).

Combinational therapy involving PD-1/PD-L1 blockade has also been investigated with chemotherapy or radiotherapy in PDAC [110]. These approaches are based on the observation that chemotherapy or radiotherapy can kill cancer cells to increase the supply of tumor antigens for presentation, so as to promote tumor-reactive immune responses [111–113]. In addition, many conventional cancer treatments in chemotherapy and radiotherapy have immune potentiating mechanisms of action, such as the elimination of immunosuppressive cells, including  $T_{\text{reg}}$  and MDSC (Zitvogel L, JCI 2008). A phase I trial (NCT02303990) of pembrolizumab (anti-PD-1 mAb) with the combination of hypofractionated radiotherapy has started to treat patients with metastatic pancreatic cancer. In another phase I study of PDAC (NCT02546531), PD-1 blockade (pembrolizumab) is proposed to be combined with gemcitabine and defactinib, an inhibitor of focal adhesion kinase (FAK), which

promotes stromal fibrosis. Because immune effector cells are also sensitive to chemotherapy and radiotherapy, early phase clinical investigations into optimizing dose and schedule in patients are necessary.

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## 16.4 Perspective

Despite recent advancements in PDAC treatment modalities, modest success has been achieved and the curative goal remains unmet. Surgical resection remains to be the only potential cure for early-stage PDAC. Immunotherapy emerges as a promising treatment for metastatic PDAC, with the potential of targeting disseminated disease as well as preventing cancer recurrence. With the technological advancement in genome sequencing, neoantigens in PDAC will be identified as better targets for vaccine therapy or ACT. Together with further interrogation of the PDAC microenvironment, it is promising that more PDAC immunosuppressive mechanisms, by which PDAC evades immune attack, will be revealed for future immune interventions.

Early clinical trials in immunotherapy also demonstrated that the complexity of the PDAC microenvironment and the formidable immunosuppressive nature of this cancer might require a combination of different therapeutic strategies [110]. These therapies need to be able to simultaneously target the stroma-cell population, where the tumor cells locate, as well as the cytotoxic T lymphocytes (manipulating different immune checkpoint inhibitors) or directly the tumor cells (traditional chemotherapeutic, vaccination, ACT). For instance, besides cancer vaccine, other therapeutic approaches, including chemotherapy, radiotherapy, and ACT may prime PDAC to become susceptible for immune checkpoint inhibitor therapy. Moving forward, the focus of modern clinical immunotherapy will be to identify the most efficacious, synergistic therapy that is able to obtain the maximum antitumor activity with the least systemic toxicity. Finally, it is imperative to identify reliable biomarkers to predict tumor susceptibility to immunotherapy in clinic, to identify those patients that are more likely to benefit from this unique therapeutic approach.

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## References

1. Finn OJ. Immuno-oncology: understanding the function and dysfunction of the immune system in cancer. *Ann Oncol.* 2012;23(Suppl 8):viii6–9. PubMed PMID: 22918931. Pubmed Central PMCID: [4085883](#)
2. Disis ML. Immune regulation of cancer. *J Clin Oncol.* 2010;28(29):4531–8. PubMed PMID: 20516428. Pubmed Central PMCID: [3041789](#)
3. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med.* 2013;19(11):1423–37. PubMed PMID: 24202395. Pubmed Central PMCID: [3954707](#)
4. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell.* 2011;144(5):646–74. PubMed PMID: 21376230
5. Turley SJ, Cremasco V, Astarita JL. Immunological hallmarks of stromal cells in the tumour microenvironment. *Nat Rev Immunol.* 2015;15(11):669–82. PubMed PMID: 26471778
6. Pietras K, Ostman A. Hallmarks of cancer: interactions with the tumor stroma. *Exp Cell Res.* 2010;316(8):1324–31. PubMed PMID: 20211171

7. Cairns R, Papandreou I, Denko N. Overcoming physiologic barriers to cancer treatment by molecularly targeting the tumor microenvironment. *Mol Cancer Res.* 2006;4(2):61–70. PubMed PMID: 16513837
8. Gajewski TF, Schreiber H, Fu YX. Innate and adaptive immune cells in the tumor microenvironment. *Nat Immunol.* 2013;14(10):1014–22. PubMed PMID: 24048123. Pubmed Central PMCID: [4118725](#)
9. Shiao SL, Ganesan AP, Rugo HS, Coussens LM. Immune microenvironments in solid tumors: new targets for therapy. *Genes Dev.* 2011;25(24):2559–72. PubMed PMID: 22190457. Pubmed Central PMCID: [3248678](#)
10. Chew V, Toh HC, Abastado JP. Immune microenvironment in tumor progression: characteristics and challenges for therapy. *J Oncol.* 2012;2012:608406. PubMed PMID: 22927846. Pubmed Central PMCID: [3423944](#)
11. Joyce JA, Fearon DT. T cell exclusion, immune privilege, and the tumor microenvironment. *Science.* 2015;348(6230):74–80. PubMed PMID: 25838376
12. Tatsumi T, Kierstead LS, Ranieri E, Gesualdo L, Schena FP, Finke JH, et al. Disease-associated bias in T helper type 1 (Th1)/Th2 CD4(+) T cell responses against MAGE-6 in HLA-DRB10401(+) patients with renal cell carcinoma or melanoma. *J Exp Med.* 2002;196(5):619–28. PubMed PMID: 12208877. Pubmed Central PMCID: [2193999](#)
13. Tran Janco JM, Lamichhane P, Karyampudi L, Knutson KL. Tumor-infiltrating dendritic cells in cancer pathogenesis. *J Immunol.* 2015;194(7):2985–91. PubMed PMID: 25795789. Pubmed Central PMCID: [4369768](#)
14. Zou W. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol.* 2006;6(4):295–307. PubMed PMID: [16557261](#)
15. Noy R, Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. *Immunity.* 2014;41(1):49–61. PubMed PMID: 25035953. Pubmed Central PMCID: [4137410](#)
16. Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol.* 2009;9(3):162–74. PubMed PMID: 19197294. Pubmed Central PMCID: [2828349](#)
17. Li H, Han Y, Guo Q, Zhang M, Cao X. Cancer-expanded myeloid-derived suppressor cells induce anergy of NK cells through membrane-bound TGF-beta 1. *J Immunol.* 2009;182(1):240–9. PubMed PMID: 19109155
18. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell.* 2009;16(3):183–94. PubMed PMID: 19732719. Pubmed Central PMCID: [2754404](#)
19. He Y, Qian H, Liu Y, Duan L, Li Y, Shi G. The roles of regulatory B cells in cancer. *J Immunology Res.* 2014;2014:215471. PubMed PMID: 24991577. Pubmed Central PMCID: [4060293](#)
20. Tjomsland V, Niklasson L, Sandstrom P, Borch K, Druid H, Bratthall C, et al. The desmoplastic stroma plays an essential role in the accumulation and modulation of infiltrated immune cells in pancreatic adenocarcinoma. *Clin Dev Immunol.* 2011;2011:212810. PubMed PMID: 22190968. Pubmed Central PMCID: [3235447](#)
21. Zischek C, Niess H, Ischenko I, Conrad C, Huss R, Jauch KW, et al. Targeting tumor stroma using engineered mesenchymal stem cells reduces the growth of pancreatic carcinoma. *Ann Surg.* 2009;250(5):747–53. PubMed PMID: 19826249
22. Feig C, Gopinathan A, Neesse A, Chan DS, Cook N, Tuveson DA. The pancreas cancer microenvironment. *Clin Cancer Res.* 2012;18(16):4266–76. PubMed PMID: 22896693. Pubmed Central PMCID: [3442232](#)
23. von Bernstorff W, Voss M, Freichel S, Schmid A, Vogel I, Johnk C, et al. Systemic and local immunosuppression in pancreatic cancer patients. *Clin Cancer Res.* 2001;7(3 Suppl):925s–32s. PubMed PMID: 11300493
24. Merika EE, Syrigos KN, Saif MW. Desmoplasia in pancreatic cancer. Can we fight it? *Gastroenterol Res Pract.* 2012;2012:781765. PubMed PMID: 23125850. Pubmed Central PMCID: [3485537](#)

25. Whatcott CJ, Posner RG, Von Hoff DD, Han H. Desmoplasia and chemoresistance in pancreatic cancer. In: Grippo PJ, Munshi HG, editors. *Pancreatic cancer and tumor microenvironment*. Trivandrum (India); 2012
26. Lesina M, Kurkowski MU, Ludes K, Rose-John S, Treiber M, Kloppel G, et al. Stat3/Socs3 activation by IL-6 transsignaling promotes progression of pancreatic intraepithelial neoplasia and development of pancreatic cancer. *Cancer Cell*. 2011;19(4):456–69. PubMed PMID: 21481788
27. Okada S, Okusaka T, Ishii H, Kyogoku A, Yoshimori M, Kajimura N, et al. Elevated serum interleukin-6 levels in patients with pancreatic cancer. *Jpn J Clin Oncol*. 1998;28(1):12–5. PubMed PMID: 9491135
28. Ling J, Kang Y, Zhao R, Xia Q, Lee DF, Chang Z, et al. KrasG12D-induced IKK2/beta/NF-kappaB activation by IL-1alpha and p62 feedforward loops is required for development of pancreatic ductal adenocarcinoma. *Cancer Cell*. 2012;21(1):105–20. PubMed PMID: 22264792. Pubmed Central PMCID: [3360958](#)
29. Muerkoster S, Wegehenkel K, Arlt A, Witt M, Sipos B, Kruse ML, et al. Tumor stroma interactions induce chemoresistance in pancreatic ductal carcinoma cells involving increased secretion and paracrine effects of nitric oxide and interleukin-1beta. *Cancer Res*. 2004;64(4):1331–7. PubMed PMID: 14973050
30. Poch B, Lotspeich E, Ramadani M, Gansauge S, Beger HG, Gansauge F. Systemic immune dysfunction in pancreatic cancer patients. *Langenbeck Arch Surg*. 2007;392(3):353–8. PubMed PMID: 17235586
31. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, et al. Signatures of mutational processes in human cancer. *Nature*. 2013;500(7463):415–21. PubMed PMID: 23945592. Pubmed Central PMCID: [3776390](#)
32. Swartz MA, Iida N, Roberts EW, Sangaletti S, Wong MH, Yull FE, et al. Tumor microenvironment complexity: emerging roles in cancer therapy. *Cancer Res*. 2012;72(10):2473–80. PubMed PMID: 22414581. Pubmed Central PMCID: [3653596](#)
33. Beatty GL, Haas AR, Maus MV, Torigian DA, Soulen MC, Plesa G, et al. Mesothelin-specific chimeric antigen receptor mRNA-engineered T cells induce anti-tumor activity in solid malignancies. *Canc Immunol Res*. 2014;2(2):112–20. PubMed PMID: 24579088. Pubmed Central PMCID: [3932715](#)
34. Gaudernack G. Prospects for vaccine therapy for pancreatic cancer. *Best Pract Res Clin Gastroenterol*. 2006;20(2):299–314. PubMed PMID: 16549329
35. Mule JJ. Dendritic cell-based vaccines for pancreatic cancer and melanoma. *Ann N Y Acad Sci*. 2009;1174:33–40. PubMed PMID: 19769734
36. Lepisto AJ, Moser AJ, Zeh H, Lee K, Bartlett D, McKolanis JR, et al. A phase I/II study of a MUC1 peptide pulsed autologous dendritic cell vaccine as adjuvant therapy in patients with resected pancreatic and biliary tumors. *Canc Ther*. 2008;6(B):955–64. PubMed PMID: 19129927. Pubmed Central PMCID: [2614325](#)
37. Kimura Y, Tsukada J, Tomoda T, Takahashi H, Imai K, Shimamura K, et al. Clinical and immunologic evaluation of dendritic cell-based immunotherapy in combination with gemcitabine and/or S-1 in patients with advanced pancreatic carcinoma. *Pancreas*. 2012;41(2):195–205. PubMed PMID: 21792083
38. Suso EM, Dueland S, Rasmussen AM, Vethrus T, Aamdal S, Kvalheim G, et al. hTERT mRNA dendritic cell vaccination: complete response in a pancreatic cancer patient associated with response against several hTERT epitopes. *Canc Immunol Immunother*. 2011;60(6):809–18. PubMed PMID: 21365467. Pubmed Central PMCID: [3098983](#)
39. Jaffee EM, Thomas MC, Huang AY, Hauda KM, Levitsky HI, Pardoll DM. Enhanced immune priming with spatial distribution of paracrine cytokine vaccines. *J Immunother Emp Tumor Immunol*. 1996;19(3):176–83. PubMed PMID: 8811492
40. Dranoff G, Jaffee E, Lazenby A, Golumbek P, Levitsky H, Brose K, et al. Vaccination with irradiated tumor cells engineered to secrete murine granulocyte-macrophage colony-stimulating factor stimulates potent, specific, and long-lasting anti-tumor immunity. *Proc*

- Natl Acad Sci U S A. 1993;90(8):3539–43. PubMed PMID: 8097319. Pubmed Central PMCID: 46336
41. Laheru D, Lutz E, Burke J, Biedrzycki B, Solt S, Onners B, et al. Allogeneic granulocyte macrophage colony-stimulating factor-secreting tumor immunotherapy alone or in sequence with cyclophosphamide for metastatic pancreatic cancer: a pilot study of safety, feasibility, and immune activation. *Clin Canc Res*. 2008;14(5):1455–63. PubMed PMID: 18316569. Pubmed Central PMCID: 2879140
  42. Lutz E, Yeo CJ, Lillemoed KD, Biedrzycki B, Kobrin B, Herman J, et al. A lethally irradiated allogeneic granulocyte-macrophage colony stimulating factor-secreting tumor vaccine for pancreatic adenocarcinoma. A Phase II trial of safety, efficacy, and immune activation. *Ann Surg*. 2011;253(2):328–35. PubMed PMID: 21217520. Pubmed Central PMCID: 3085934
  43. Chawla S, Henshaw R, Seeger L, Choy E, Blay JY, Ferrari S, et al. Safety and efficacy of denosumab for adults and skeletally mature adolescents with giant cell tumour of bone: interim analysis of an open-label, parallel-group, phase 2 study. *Lancet Oncol*. 2013;14(9):901–8. PubMed PMID: 23867211
  44. Jaffee EM, Hruban RH, Biedrzycki B, Laheru D, Schepers K, Sauter PR, et al. Novel allogeneic granulocyte-macrophage colony-stimulating factor-secreting tumor vaccine for pancreatic cancer: a phase I trial of safety and immune activation. *J Clin Oncol Off J Am Soc Clin Oncol*. 2001;19(1):145–56. PubMed PMID: 11134207
  45. Le DT, Brockstedt DG, Nir-Paz R, Hampl J, Mathur S, Nemunaitis J, et al. A live-attenuated *Listeria* vaccine (ANZ-100) and a live-attenuated *Listeria* vaccine expressing mesothelin (CRS-207) for advanced cancers: phase I studies of safety and immune induction. *Clin Canc Res*. 2012;18(3):858–68. PubMed PMID: 22147941. Pubmed Central PMCID: 3289408
  46. Le DT, Wang-Gillam A, Picozzi V, Greten TF, Crocenzi T, Springett G, et al. Safety and survival with GVAX pancreas prime and *Listeria Monocytogenes*-expressing mesothelin (CRS-207) boost vaccines for metastatic pancreatic cancer. *J Clin Oncol Off J Am Soc Clin Oncol*. 2015;33(12):1325–33. PubMed PMID: 25584002. Pubmed Central PMCID: 4397277
  47. Coveler AL, Rossi GR, Vahanian NN, Link C, Chiorean EG. Algenpantucel-L immunotherapy in pancreatic adenocarcinoma. *Immunotherapy*. 2016;8(2):117–25. PubMed PMID: 26787078
  48. Hardacre JM, Mulcahy M, Small W, Talamonti M, Obel J, Krishnamurthi S, et al. Addition of algenpantucel-L immunotherapy to standard adjuvant therapy for pancreatic cancer: a phase 2 study. *J Gastrointest Sur*. 2013;17(1):94–100. discussion p -1. PubMed PMID: 23229886
  49. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med*. 2012;366(26):2443–54. PubMed PMID: 22658127. Pubmed Central PMCID: 3544539
  50. Royal RE, Levy C, Turner K, Mathur A, Hughes M, Kammula US, et al. Phase 2 trial of single agent Ipilimumab (anti-CTLA-4) for locally advanced or metastatic pancreatic adenocarcinoma. *J Immunother*. 2010;33(8):828–33. PubMed PMID: 20842054
  51. Panizza A, Merkow J, Edil BH, Zhu Y. Immunotherapy for pancreatic ductal adenocarcinoma: an overview of clinical trials. *Chinese J Can Res*. 2015;27(4):376–91. PubMed PMID: 26361407. Pubmed Central PMCID: 4560736
  52. Hinrichs CS, Rosenberg SA. Exploiting the curative potential of adoptive T-cell therapy for cancer. *Immunol Rev*. 2014;257(1):56–71. PubMed PMID: 24329789. Pubmed Central PMCID: 3920180
  53. Rosenberg SA, Restifo NP, Yang JC, Morgan RA, Dudley ME. Adoptive cell transfer: a clinical path to effective cancer immunotherapy. *Nat Rev Cancer*. 2008;8(4):299–308. PubMed PMID: 18354418. Pubmed Central PMCID: 2553205
  54. Restifo NP, Dudley ME, Rosenberg SA. Adoptive immunotherapy for cancer: harnessing the T cell response. *Nat Rev Immunol*. 2012;12(4):269–81. PubMed PMID: 22437939
  55. Besser MJ, Shapira-Frommer R, Treves AJ, Zippel D, Itzhaki O, Hershkovitz L, et al. Clinical responses in a phase II study using adoptive transfer of short-term cultured tumor infiltration lymphocytes in metastatic melanoma patients. *Clin Cancer Res*. 2010;16(9):2646–55. PubMed PMID: 20406835

56. Besser MJ. Is there a future for adoptive cell transfer in melanoma patients? *Oncoimmunology*. 2013;2(10):e26098. PubMed PMID: 24353909. Pubmed Central PMCID: [3862631](#)
57. Robbins PF, Dudley ME, Wunderlich J, El-Gamil M, Li YF, Zhou J, et al. Cutting edge: persistence of transferred lymphocyte clonotypes correlates with cancer regression in patients receiving cell transfer therapy. *J Immunol*. 2004;173(12):7125–30. PubMed PMID: 15585832. Pubmed Central PMCID: [2175171](#)
58. Hinrichs CS, Borman ZA, Gattinoni L, Yu Z, Burns WR, Huang J, et al. Human effector CD8+ T cells derived from naive rather than memory subsets possess superior traits for adoptive immunotherapy. *Blood*. 2011;117(3):808–14. PubMed PMID: 20971955. Pubmed Central PMCID: [3035075](#)
59. Wang M, Yin B, Wang HY, Wang RF. Current advances in T-cell-based cancer immunotherapy. *Immunotherapy*. 2014;6(12):1265–78. PubMed PMID: 25524383. Pubmed Central PMCID: [4372895](#)
60. Kershaw MH, Westwood JA, Darcy PK. Gene-engineered T cells for cancer therapy. *Nat Rev Cancer*. 2013;13(8):525–41. PubMed PMID: 23880905
61. Barrett DM, Singh N, Porter DL, Grupp SA, June CH. Chimeric antigen receptor therapy for cancer. *Annu Rev Med*. 2014;65:333–47. PubMed PMID: 24274181. Pubmed Central PMCID: [4120077](#)
62. Jena B, Dotti G, Cooper LJ. Redirecting T-cell specificity by introducing a tumor-specific chimeric antigen receptor. *Blood*. 2010;116(7):1035–44. PubMed PMID: 20439624. Pubmed Central PMCID: [2938125](#)
63. Blankenstein T, Coulie PG, Gilboa E, Jaffee EM. The determinants of tumour immunogenicity. *Nat Rev Cancer*. 2012;12(4):307–13. PubMed PMID: 22378190. Pubmed Central PMCID: [3552609](#)
64. Morgan RA, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. *Mol Ther* 2010;18(4):843–851. PubMed PMID: 20179677. Pubmed Central PMCID: [2862534](#)
65. Curran KJ, Brentjens RJ. Chimeric antigen receptor T cells for cancer immunotherapy. *J Clin Oncol Off J Am Soc Clin Oncol*. 2015;33(15):1703–6. PubMed PMID: 25897155
66. Kakarla S, Gottschalk S. CAR T cells for solid tumors: armed and ready to go? *Cancer J*. 2014;20(2):151–5. PubMed PMID: 24667962. Pubmed Central PMCID: [4050065](#)
67. Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. *Science*. 2015;348(6230):69–74. PubMed PMID: 25838375
68. Fousek K, Ahmed N. The evolution of T-cell therapies for solid malignancies. *Clin Canc Res*. 2015;21(15):3384–92. PubMed PMID: 26240290. Pubmed Central PMCID: [4526112](#)
69. Casati A, Varghaei-Nahvi A, Feldman SA, Assenmacher M, Rosenberg SA, Dudley ME, et al. Clinical-scale selection and viral transduction of human naive and central memory CD8+ T cells for adoptive cell therapy of cancer patients. *Canc Immunol Immunother*. 2013;62(10):1563–73. PubMed PMID: 23903715
70. Gattinoni L, Klebanoff CA, Restifo NP. Paths to stemness: building the ultimate antitumour T cell. *Nat Rev Cancer*. 2012;12(10):671–84. PubMed PMID: 22996603
71. Chmielewski M, Hahn O, Rapp G, Nowak M, Schmidt-Wolf IH, Hombach AA, et al. T cells that target carcinoembryonic antigen eradicate orthotopic pancreatic carcinomas without inducing autoimmune colitis in mice. *Gastroenterology*. 2012;143(4):1095–107. e2. PubMed PMID: 22750462
72. Stromnes IM, Schmitt TM, Hulbert A, Brockenbrough JS, Nguyen HN, Cuevas C, et al. T cells engineered against a native antigen can surmount immunologic and physical barriers to treat pancreatic ductal adenocarcinoma. *Cancer Cell*. 2015;28(5):638–52. PubMed PMID: 26525103. Pubmed Central PMCID: [4724422](#)
73. Bailey P, Chang DK, Nones K, Johns AL, Patch AM, Gingras MC, et al. Genomic analyses identify molecular subtypes of pancreatic cancer. *Nature*. 2016;531(7592):47–52. PubMed PMID: 26909576

74. Lollini PL, Cavallo F, Nanni P, Forni G. Vaccines for tumour prevention. *Nat Rev Cancer*. 2006;6(3):204–16. PubMed PMID: 16498443
75. Laheru D, Jaffee EM. Immunotherapy for pancreatic cancer—science driving clinical progress. *Nat Rev Cancer*. 2005;5(6):459–67. PubMed PMID: 15905855
76. Wachsmann MB, Pop LM, Vitetta ES. Pancreatic ductal adenocarcinoma: a review of immunologic aspects. *J Investig Med*. 2012;60(4):643–63. PubMed PMID: 22406516. Pubmed Central PMCID: [3319488](#)
77. Melief CJ, van Hall T, Arens R, Ossendorp F, van der Burg SH. Therapeutic cancer vaccines. *J Clin Invest*. 2015;125(9):3401–12. PubMed PMID: 26214521. Pubmed Central PMCID: [4588240](#)
78. Rajasagi M, Shukla SA, Fritsch EF, Keskin DB, DeLuca D, Carmona E, et al. Systematic identification of personal tumor-specific neoantigens in chronic lymphocytic leukemia. *Blood*. 2014;124(3):453–62. PubMed PMID: 24891321. Pubmed Central PMCID: [4102716](#)
79. de Gruijl TD, van den Eertwegh AJ, Pinedo HM, Scheper RJ. Whole-cell cancer vaccination: from autologous to allogeneic tumor- and dendritic cell-based vaccines. *Cancer Immunol Immunother*. 2008;57(10):1569–77. PubMed PMID: 18523771. Pubmed Central PMCID: [2491427](#)
80. Keenan BP, Jaffee EM. Whole cell vaccines—past progress and future strategies. *Semin Oncol*. 2012;39(3):276–86. PubMed PMID: 22595050. Pubmed Central PMCID: [3356993](#)
81. Srivatsan S, Patel JM, Bozeman EN, Imasuen IE, He S, Daniels D, et al. Allogeneic tumor cell vaccines: the promise and limitations in clinical trials. *Human Vaccin Immunother*. 2014;10(1):52–63. PubMed PMID: 24064957. Pubmed Central PMCID: [4181031](#)
82. Galili U. Anti-Gal: an abundant human natural antibody of multiple pathogeneses and clinical benefits. *Immunology*. 2013;140(1):1–11. PubMed PMID: 23578170. Pubmed Central PMCID: [3809700](#)
83. Rossi GR, Mautino MR, Unfer RC, et al. Effective treatment of preexisting melanoma with whole cell vaccines expressing  $\alpha(1,3)$ -galactosyl epitopes. *Cancer Res*. 2005;65:10555–61
84. Rossi GR, et al. Allogeneic melanoma vaccine expressing alphaGal epitopes induces antitumor immunity to autologous antigens in mice without signs of toxicity. *Journal of Immunotherapy*. 2008;31(6):545–54
85. Rice J, Ottensmeier CH, Stevenson FK. DNA vaccines: precision tools for activating effective immunity against cancer. *Nat Rev Cancer*. 2008;8(2):108–20. PubMed PMID: 18219306
86. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature*. 1998;392(6673):245–52. PubMed PMID: 9521319
87. Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. *Nat Rev Cancer*. 2012;12(4):265–77. PubMed PMID: 22437871. Pubmed Central PMCID: [3433802](#)
88. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med*. 2010;363(5):411–22. PubMed PMID: 20818862
89. Zhu Y, Yao S, Chen L. Cell surface signaling molecules in the control of immune responses: a tide model. *Immunity*. 2011;34(4):466–78. PubMed PMID: 21511182. Pubmed Central PMCID: [3176719](#)
90. Chen L. Co-inhibitory molecules of the B7-CD28 family in the control of T-cell immunity. *Nat Rev Immunol*. 2004;4(5):336–47. PubMed PMID: 15122199
91. Zou W, Chen L. Inhibitory B7-family molecules in the tumour microenvironment. *Nat Rev Immunol*. 2008;8(6):467–77. PubMed PMID: 18500231
92. Jiang Y, Li Y, Zhu B. T-cell exhaustion in the tumor microenvironment. *Cell Death Dis*. 2015;6:e1792. PubMed PMID: 26086965. Pubmed Central PMCID: [4669840](#)
93. Wherry EJ. T cell exhaustion. *Nat Immunol*. 2011;12(6):492–9. PubMed PMID: 21739672
94. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* 2012;12(4):252–264. PubMed PMID: 22437870
95. Naidoo J, Page DB, Wolchok JD. Immune modulation for cancer therapy. *Br J Cancer*. 2014;111(12):2214–9. PubMed PMID: 25211661. Pubmed Central PMCID: [4264429](#)

96. Couzin-Frankel J. Breakthrough of the year 2013. *Canc Immunother Sci*. 2013;342(6165):1432–3. PubMed PMID: 24357284
97. Greenwald RJ, Freeman GJ, Sharpe AH. The B7 family revisited. *Annu Rev Immunol*. 2005;23:515–48. PubMed PMID: 15771580
98. Lenschow DJ, Walunas TL, Bluestone JA. CD28/B7 system of T cell costimulation. *Annu Rev Immunol*. 1996;14:233–58. PubMed PMID: WOS:A1996UH42900011. English
99. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med*. 2010;363(8):711–23. PubMed PMID: 20525992. Pubmed Central PMCID: [3549297](#)
100. Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. *N Engl J Med*. 2011;364(26):2517–26. PubMed PMID: 21639810
101. Chen L, Han X. Anti-PD-1/PD-L1 therapy of human cancer: past, present, and future. *J Clin Invest*. 2015;125(9):3384–91. PubMed PMID: 26325035. Pubmed Central PMCID: [4588282](#)
102. Zou W, Wolchok JD, Chen L. PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: mechanisms, response biomarkers, and combinations. *Sci Trans Med*. 2016;8(328):328rv4. PubMed PMID: 26936508
103. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, et al. Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. *N Engl J Med*. 2013;369(2):134–44. PubMed PMID: 23724846. Pubmed Central PMCID: [4126516](#)
104. Robert C, Schachter J, Long GV, Arance A, Grob JJ, Mortier L, et al. Pembrolizumab versus Ipilimumab in Advanced Melanoma. *N Engl J Med*. 2015;372(26):2521–32. PubMed PMID: 25891173
105. Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med*. 2012;366(26):2455–65. PubMed PMID: 22658128. Pubmed Central PMCID: [3563263](#)
106. Lutz ER, Kinkead H, Jaffee EM, Zheng L. Priming the pancreatic cancer tumor microenvironment for checkpoint-inhibitor immunotherapy. *Oncoimmunology*. 2014;3(11):e962401. PubMed PMID: 25941589. Pubmed Central PMCID: [4292514](#)
107. Soares KC, Rucki AA, Wu AA, Olino K, Xiao Q, Chai Y, et al. PD-1/PD-L1 blockade together with vaccine therapy facilitates effector T-cell infiltration into pancreatic tumors. *J Immunother*. 2015;38(1):1–11. PubMed PMID: 25415283. Pubmed Central PMCID: [4258151](#)
108. Duraiswamy J, Kaluza KM, Freeman GJ, Coukos G. Dual blockade of PD-1 and CTLA-4 combined with tumor vaccine effectively restores T-cell rejection function in tumors. *Cancer Res*. 2013;73(12):3591–603. PubMed PMID: 23633484. Pubmed Central PMCID: [3686913](#)
109. Zamarin D, Holmgaard RB, Subudhi SK, Park JS, Mansour M, Palese P, et al. Localized oncolytic virotherapy overcomes systemic tumor resistance to immune checkpoint blockade immunotherapy. *Sci Trans Med*. 2014;6(226):226ra32. PubMed PMID: 24598590. Pubmed Central PMCID: [4106918](#)
110. Arslan C, Yalcin S. Current and future systemic treatment options in metastatic pancreatic cancer. *J Gastrointest Oncol*. 2014;5(4):280–95. PubMed PMID: 25083302. Pubmed Central PMCID: [4110498](#)
111. Bernstein MB, Krishnan S, Hodge JW, Chang JY. Immunotherapy and stereotactic ablative radiotherapy (ISABR): a curative approach? *Nat Rev Clin Oncol*. 2016;13:516–24. PubMed PMID: 26951040
112. Formenti SC, Demaria S. Combining radiotherapy and cancer immunotherapy: a paradigm shift. *J Natl Cancer Inst*. 2013;105(4):256–65. PubMed PMID: 23291374. Pubmed Central PMCID: [3576324](#)
113. Lake RA, Robinson BW. Immunotherapy and chemotherapy—a practical partnership. *Nat Rev Cancer*. 2005;5(5):397–405. PubMed PMID: 15864281