Chapter 8 Crossroads of Signaling Pathways

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Abstract As studies on PC progression continue to uncover a growing number of crosstalks and co-occurrences of mutations and epigenetic alterations, new drugs are getting approved bringing significant changes in the treatment paradigm of these tumors.

This chapter recapitulates the best known examples of molecular interactions potentially targetable to achieve these therapeutic evolutionary changes, to allow a better control of PC which, in 2012 alone, has still killed more than 28,000 men, in USA (Siegel et al, CA Cancer J Clin, 62:10–29, 2012; El-Amm, Aragon-Ching, Ther Adv Med Oncol, 5(1):25–40, 2013).

8.1 Background and Aims

Prostate cancer (PC) is a multifocal disease, composed by several independent tumor foci that may show different degrees of molecular alterations. The heterogeneous nature of PC which, at a molecular level, derives from the crosstalk of multiple signal transductions variously acting in promoting growth, survival and therapy-resistance of PC cells (Pittoni et al. 2011).

The development of new therapeutic strategies, particularly focused toward castration-resistant prostate cancer (CRPC), relies on a better understanding of the resistance pathways selectively adopted from prostate cancer cells (El-Amm and Aragon-Ching 2013).

Recently, it has been shown that androgens residual from androgen-deprivation therapy may indirectly favor cancer growth, with a progressive increase of the PSA levels, *via* the over-expression of many HIF-1 dependent, hypoxia-inducible

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genes. The interplay between hypoxia and AR, further cross-talks with several oxidative stress mediators, cytokines, growth factors, DNA-repair pathways, and epigenetic regulators, in a cooperative effort to ensure the survival of neoplastic cells in a highly adverse metabolic (and environmental) background. These interacting signaling mechanisms, indeed, may either potentiate or counteract each other, leading alternatively to cell death or adaptation and radio-chemoresistance. It thus becomes apparent that resistance to therapy can be overcome only through a proper therapeutic manipulation of the right factor(s) that, in turn, will influence the others, triggering PC cells death (Marignol et al. 2008).

As an example, it has been surprisingly shown that cell death can be induced in castration-resistant tumors "still" *via* AR, which can modulate apoptosis and autophagy if targeted in conjunction with PKA pathway members (Attar et al. 2009).

As well, considering that AR is largely expressed in tumor microenvironmental stromal cells, drugs targeting AR signaling in PC cells give rise to a therapeutic favourable effect also on the stromal compartments, as AR is largely expressed in tumor microenvironmental stromal cells (Mantalaris et al. 2001).

Thus, a favourable response to AR targeting will encompass both reductions in serum PSA and bone-specific, osteoblast derived, alkaline phosphatase.

Overall, mounting evidence suggest that the cell fate, in response to therapeutic attack, depends on a plethora of variable factors, ranging from metabolic stress, functional status of cells, the interaction level between the stress-response pathways, paracrine mediators produced by tumor microenvironment, and the epigenetic interactions on DNA damage response and DNA repair (Murr 2010).

During these last years, the efforts of the scientific community have been focused on the correct interpretation of the *complementary pathways* which could kill radioand chemoresistant cancer cells.

The end-point of such a program will require carefully designed clinical trials, a rigorous patient selection and retrospective analyses of clinical, pathological, and follow-up data.

In the rapidly evolving field of prostate cancer therapy, new drugs are being used; as well, new insights indicating possible different rational approaches to the treatment sequencing with "old" drugs are being proposed. The search for novel biomarkers, useful for the individualized prediction of treatment response and outcome of PC patients, is actively on. This is a particularly complex investigational field (considering that the malignant phenotype of prostate cancer cells results from a highly variable combination of functional, genetic and epigenetic defects in cell cycle metabolism, checkpoint control and DNA-repair pathways, working together to render PC a lethal disease). The real challenge lies not only in detecting all these alterations, but also in defining the multiple layers of their reciprocal intersection (Sarwar and Persson 2011).

We are thus requested to critically review our knowledge about the role of the plethora of molecular guests acting on the scenario of prostate cancer progression, which comprises both epithelial and stromal cells, both contributing to tumor heterogeneity and growth dynamics (Cho-Chung 1989, 1990; Camps et al. 1990; Cunha et al. 1996).

That prostate cancer development and growth is dependent on androgens and can be suppressed by androgen ablation monotherapy is an old concept (Zhu and Kyprianou 2008).

The appearance of androgen-independent prostate tumor growth, leading to cancer recurrence and highly metastatic disease, is a well-known phenomenon as well (Wang et al. 2007).

During the entire life-span of prostate cancer, the androgen axis actively crosstalks with a plethora of growth factors, driving the shift of prostate cancer cell toward survival and invasion advantage. Androgenic control of growth and differentiation is tightly regulated in both stromal and epithelial cells (Sar et al. 1990).

This explain why the successful treatment of PC with drugs targeting (AR) signaling (defined as *Epithelial-Stromal Targeting Agents*), leads to reduction in either serum PSA and bone-specific alkaline phosphatase (Mantalaris et al. 2001; Niu et al. 2010).

A poor clinical outcome for prostate cancer patients has been associated, instead, with low-AR levels in the stromal microenvironment (Henshall et al. 2001), and this finding has been proposed as one of the mechanisms involved in the emergence of androgen-independent cancer (Dayyani et al. 2011). The propulsive effect of androgens on prostate epithelial cell proliferation and survival are indirectly regulated by paracrine mediators produced by stromal cells, such as insulin-like growth factor (IGF), fibroblast growth factor (FGF), epidermal growth factor (EGF), (Cunha and Donjacour 1989), vascular endothelial growth factor (VEGF) and transforming growth factor- β (TGF- β) (Byrne et al. 1996).

The epidermal growth factor-1 (EGF) and its receptor (EGFR), (Russell et al. 1998) are frequently up-regulated in advanced stages of PC (Di Lorenzo et al. 2002) Targeting EGFR with monoclonal antibodies or with tyrosine kinase inhibitors, suppresses growth and invasion of androgen-dependent and -independent prostate cancer cells *in vitro*, leading to the conclusion that the multi-crossed signals between EGF/EGFR and androgen signaling is crucial for the acquisition and the maintenance of androgen sensitivity (Bonaccorsi et al. 2004; Festuccia et al. 2005; Leotoing et al. 2007).

Both AR and EGF can activate MAPK and, in a 'functional-symmetry', the EGF-activated MAPK/extracellular signaling-regulated kinase kinase-1 (MEKK1) cascade. This allows EGF to interfere with AR function, modulating the androgen response and blocking androgen-dependent transcription in differentiated cells. MAPK extracellular kinase (MEK) inhibition reverses the EGF-mediated AR down-regulation in differentiated cells (Peterziel et al. 1999).

The alteration of this EGF–AR interplay is an important contributor to prostate tumor progression. The modulation of AR signaling activity by ERBB2 (HER-2/neu), a lead member of the EGFR family of receptor tyrosine kinases, has been found correlated with prostate cancer progression to cell growth of androgen-independent metastatic disease (Heinlein and Chang 2004), *in vitro* and *in vivo* (Craft et al. 1999; Yeh et al. 1999; Mellinghoff et al. 2004; Liu et al. 2005).

Similar positive feedbacks with AR activity in prostate cancer cells have been described also for several other growth factors (Orio et al. 2002). Evidence supports a strict interaction between AR and the IGF signaling. The high IGF1 signaling in prostate cancer cells (HepG2 and LNCaP cells) (Wu et al. 2007) likely depends upon AR up-regulation of IGF1 receptor expression and/or, alternatively, upon the modulation of IGF-binding proteins (IGFBPs), which, in turn, are up-regulated by either androgens and IGF1 in androgen-responsive human fibroblasts (Yoshizawa and Ogikubo 2006) IGF1 enhances AR transactivation under low/absent androgen levels (Culig et al. 1994; Orio et al. 2002) and promotes prostate tumor cell proliferation (Burfeind et al. 1996). According to several reports, high IGF1 levels in the serum can be considered a marker of an increased risk of prostate cancer (Pollak et al. 1998; Wolk et al. 1998).

Even more data concern the cross-talks between AR and Transforming growth factor- β (TGF β). This ubiquitous cytokine, for instance, contributes to the regulation of proliferation, growth and differentiation of prostate stromal and epithelial cells.

Cofilin and prohibitin, two novel signaling effectors of TGFB1, that serve as potential intracellular effectors of its apoptotic action, were identified in human prostate cancer cells (Zhu et al. 2006). Androgens can inhibit TGFB1-induced apoptosis in prostate cancer cells (Chipuk et al. 2002) via the AR-associated protein 55 (ARA55/Hic-5; LIM protein superfamily). Overexpression of ARA55 inhibits TGFB-mediated up-regulation of SMAD transcriptional activity in rat prostate epithelial cells, as well as human prostate cells, via an interaction between ARA55 and SMAD3 (Wang et al. 2005). Cancer cells become refractory to the growth inhibitory activity of TGFB due to the loss (or mutation) of transmembrane receptors or intracellular TGFB signaling effectors during tumor initiation (Akhurst and Derynck 2001). In advanced prostate cancer and in PC bone metastasis, TGFB is over-expressed and TGFB1 ligand overexpression results in pro-oncogenic rather than growth suppressive effect (Coffey et al. 1986; Roberts et al. 1986; Derynck and Zhang 2003; Zhu and Kyprianou 2005).

The androgenic-mediated TGFB enhancement seems to play a role on the epithelial-mesenchymal transition (EMT) in metastasizing cancers (Zavadil and Bottinger 2005), with a further interplay with E-cadherin. The effects of TGFB1 expression on stromal cell proliferation and differentiation, depend on the specific stromal cell type, microenvironment, and interactions with other growth factor (Sporn and Roberts 1992). AR and TGFB1 levels significantly correlate in the stromal component of prostatic intraepithelial neoplasia (Cardillo et al. 2000). Very interestingly, TGFB1 triggers the AR translocation from nucleus to cytoplasm in prostate stromal cells underlying to myodifferentiation (Gerdes et al. 1998, 2004), while androgens enhance TGFB1-mediated proliferation of prostatic smooth muscle cells (Salm et al. 2000).

Prostate cancer progression toward androgen-independent disease has been linked also to changes in the expression of several members of the FGF family, characterized by a broad spectrum of functions on cell differentiation, migration, and angiogenesis (Ornitz and Itoh 2001). FGF2 can stimulate also the proliferation of prostate stromal cells, in a synergistic fashion with DHT (Niu et al. 2001).

The synthesis of FGF2 and FGF7 in prostate epithelial cells seems to be mainly regulated by estrogen receptors (ER), whereas ER act in coordination with AR to mediate the synthesis of these growth factors in stromal cells. Androgen depletion rapidly reduces stromal IGF1 expression, after castration, favoring PC cells apoptosis (Ohlson et al. 2007). AR can otherwise directly influence the expression of FGF1, FGF2, FGF8, and FGF10 in either prostate tumor epithelial cells and stromal cells (Saric and Shain 1998; Nakano et al. 1999; Rosini et al. 2002), while a paracrine secretion of FGF10 exert a positive feedback on AR, up-regulating its expression (Memarzadeh et al. 2007). In response to FGFs, AR potentiates FGF-induced survival of prostate cancer cells, possibly through BCL-2 induction, allowing the escape of selected clones from androgenic control (Rosini et al. 2002; Gonzalez-Herrera et al. 2006).

Among the cross-actions between AR and the *multifunctional growth factor signaling pathways*, the interplay between the cellular responses to androgen and hypoxia is emerging as a further key phenomenon in the developing of androgen-independent, metastasizing prostate cancer cell clones (Marignol et al. 2008). In prostate cells, androgens and hypoxia share several regulatory molecular mechanisms: both androgens and hypoxia, in fact, due to the presence of a hypoxia-responsive region in the human PSA promoter, can induce in fact PSA expression (Horii et al. 2007).

It has been then hypothesized that hypoxia, mainly through the hypoxia-inducible factor (HIF1A), may facilitate PC progression through the cross-talk with AR. To further support this idea, it has been recently reported that residual androgens following androgen deprivation induce the expression of hypoxia-inducible genes and stimulate cancer re-growth (Marignol et al. 2008).

This is of particular interest if we consider that, as for most solid cancers, hypoxia is a common feature of prostate tumors. Then, targeting hypoxia looks as a very appealing complementary strategy for the management of aggressive prostate cancers.

The 'hypoxia-response' signaling system up-regulates the expression of a a wide spectrum of effectors that increase the ability of tumor cells to turn poor oxygenation into survival advantage (Anastasiadis et al. 2003) and radio- and chemoresistance (Zhou et al. 2006).

In response to the decrease in the micro-environmental oxygen (Ellis et al. 2009) HIF-1 α regulates gene expression of several genes involved in multiple physiological responses, such as erythropoiesis and glycolysis (short term solutions) and angiogenesis (long term solution) (Semenza 1998), via the expression of VEGF (Delongchamps et al. 2006).

VEGF, is "the" angiogenic cytokine, driving endothelial cell proliferation and migration, and vessel assembly (Fong et al. 1995). The expression of HIF1, AR and VEGF expression are tightly correlated (Boddy et al. 2005; Banham et al. 2007). So, AR regulates angiogenesis in androgen-sensitive PC through the HIF1-induced VEGF increase (Boddy et al. 2005). Following androgen-deprivation, the intracellular reactive oxygen species induce, instead, the direct up-regulation of VEGF-C, which favor AR transactivation mediated by the AR co-activator BAG-1L

(Rinaldo et al. 2007). Clonal selection for cells with higher expression of HIF-1a and/or apoptotic resistance pathways contributes to determine cell specific responses to hypoxia (Zhou et al. 2006).

HIF-1 α over-expression/hyperfunction may be induced by genetic loss of expression/function of pVHL (Ivan and Kaelin 2001, p 53; An et al. 1998) and/or PTEN (Zundel et al. 2000) leading to the activation of PI3K/AKT/mTOR pathway (Zundel et al. 2000; Stiehl et al. 2002) which, in turn, plays a well-known role in proliferation, survival and metastatic ability of hormone independent prostate cancers, as demonstrated by the correlation between elevated phosphorylated AKT and high Gleason grade of PC (Yuan et al. 2007). HIF-1 α may be induced, in addition, by several cytokines and growth factors including insulin (Zelzer et al. 1998), insulin like growth factors (Feldser et al. 1999), P42/44 mitogen activated kinase (MAPK) (Richard et al. 1999).

Furthermore, alternative mechanisms to those mentioned above have been identified which include the tumor microenvironment (Weidemann and Johnson 2008) and mutations within the ODD domain of HIF-1 α (Mabjeesh and Amir 2007). This eloquently explain why targeting only one member of the hypoxia-related angiogenetic pathway is insufficient to permanently inhibit tumor angiogenesis and why tumor cells, treated with a mono-drug therapy, develop resistance by selection of 'hypoxia resistant' cells or by activating alternate angiogenic pathways (Kerbel and Folkman 2002).

To further complicate this scenario, under severe hypoxia, radio-/chemoresistance and clonal selection may develop as a response of opposing signals delivered by survival and death pathways that allow selection of cells that have a growth advantage either genetically or epigenetically determined (Zhou et al. 2006).

Post-translational epigenetic modifications, including acetylation mediated by histone acetyltransferases (HATS) and deacetylation by histone deacetylases (HDACs), have been shown to be critical to HIF-1 α signaling (Ellis et al. 2009).

HIF-1 α signaling up-regulates the activity of HDACs. Then, HDAC inhibitors are emerging as a new class of anti-angiogenetic cancer therapeutics.

However, the anti-angiogenic properties of HDACI have been associated also with the alteration of numerous pro- and anti-angiogenic genes (Liu et al. 2006) other than HIF-1 α and VEGF. They encompass FGF, *angiopoietin*, tunica intima endothelial kinase 2 (*TIE2*), endothelial nitric oxide synthase (*eNOS*) (Qian et al. 2004; Rossig et al. 2002, p 53), pVHL, thrombospondin 1 (Kim et al. 2001; Kwon et al. 2002; Sasakawa et al. 2003; Kang et al. 2008, p 21)^{WAF1/CIP1}, and survivin (Qian et al. 2004). Even for HDACI, monotherapy has shown limited responses in the clinic, but it seems very promising, as a part of combination therapies. Pre-clinical and clinical studies indicate that HDACI have positive effects on the expression of pro- and anti-angiogenic genes, suggesting their useful role in reinforcing the actions of anti-VEGF therapies.

As it has been largely shown in the past decade, the intricate molecular cross-talks underlying the malignant phenotype and the emergence of androgenindependent prostate tumors encompass the expression and functional defects in HR, single-strand break- (DNA-ssb) repair, MMR and base-excision repair (BER). Prostate cancer cell lines (Chen et al. 2003; Yeh et al. 2001) have been found to be defective in mismatch–repair (MMR), and up to 23 % of prostate cancers display a high level of microsatellite instability associated with mutations in MMR genes and deficient MMR protein expression (Norris et al. 2007; Prtilo et al. 2005; Sun et al. 2006).

This, in turn, may lead to high mutation rates among microsatellites, ending in a mutator phenotype.

As well, DNA polymorphisms in BER- or DNA-ssb repair associated Xrcc1, Ogg1 and DNA polymerase-b genes have been associated with higher risk for prostate cancer (Chen et al. 2003; Rybicki et al. 2004; Xu et al. 2002).

Most of the molecular therapies targeting the key control pathways involved in prostate cancerogenesis and progression may indirectly influence the DNA-dsb repair activity of neoplastic cells. This has been observed for therapies inhibiting EGFR, IGFR, HDAC and proteasome pathways (Ma et al. 2003; Chinnaiyan et al. 2005; Dittmann et al. 2005a, b) have documented that the radiosensitization mediated by inhibiting EGFR can be related with the altered DNA-PKcs expression, function and cytoplasmic sequestration; as well, an increased DNA-dsbs (Rochester et al. 2005) has been shown to be induced by the inhibition of IGF-1R, via the altered ATM activation, and could be used in combination with radiotherapy (Choudhury et al. 2006) in hypoxic cancers. By converse, the RAS-mediated tumor cell radioresistance could be linked to the augmented DNA-dsb repair (Chang et al. 2005) induced by the use of farnesyl transferase inhibitors (FTIs) via the increase of Ku80 expression.

An additional crosstalk involves DNA-repair genes in human cancers. It concerns the occurrence of silencing DNA repair genes such as *MLH1* and *O*-6-methylguanine-DNA methyltransferase (*MGMT*) leading to microsatellite instability and a failure to repair DNA lesions (Jones and Baylin 2002). This phenomenon is still a matter of investigation in PC.

Another intriguing example of molecular co-sharing in PC is represented by the c-kit receptor (Pittoni et al. 2011). c-Kit receptor is normally expressed by prostate stem cells, that apparently require the c-Kit signaling for prostate regeneration (Leong et al. 2008) in humans, after hormone ablation, c-Kit expression may be observed in a considerable percentage of high-risk prostate cancer cells (Di Lorenzo et al. 2004) and in 10–30 % of this subset of prostate cancers NE differentiation occurs.

In normal prostate, a resident stromal population of mast-cells (MCs), also express the c-kit receptor (Leong et al. 2008).

It was initially thought that MCs can promote tumor growth of WD adenocarcinoma synthetizing MMP-9. MMP-9 has been indicated to correlate with progression of prostate tumor in humans (Castellano et al. 2008). As an ECM-degrading enzyme, it facilitates cell migration and invasion of tumor cells, allowing also the cleavage and activation of growth factors concurrently acting in tumor progression. In addition, peritumoral MCs were shown to stimulate prostate tumor growth in rats by providing proangiogenic factors (Johansson et al. 2010). However, it was demonstrated, both in mouse and in humans, that poorlydifferentiated prostate tumors with features of EMT show an autocrine production of MMP-9 and are devoid of infiltrating MCs.

This implies that MC inactivation would be ineffective when used in therapy for advanced and poorly-differentiated PC, and lead to the intriguing consideration that MC may contribute to the maintenance of prostate stem cell homeostasis and counteract NE tumor formation, serving as "natural decoys" that sequester stimulating growth factors, thus limiting c-Kit signaling in prostate cancer stem cells (Pittoni et al. 2011).

This hypothesis strongly discourages the idea of MC inhibition in PC. Otherwise, the therapeutic use of c-Kit tyrosine kinase inhibitors, such as imatinib, would instead offer the advantage of targeting both adenocarcinoma-promoting MCs (stroma targeting) and NE tumor variants (tumor targeting (Pittoni et al. 2010).

Overall, prostate cancerogenesis emerges as an extremely complex field, involving genetic and epigenetic alterations with multiple layers of merging.

We are still far away to use molecular classifications to unequivocally define different prognostic subcategories of prostate cancer. Key questions remain to be answered before the full range of mutations and genetic alterations will be elucidated.

Nevertheless, it is likely that, as for most of human cancers, also in PC the genes with a high incidence of mutation frequently participate to the evenience of abnormal epigenetic events, and this co-occurrence may be related, for instance, to the abnormal expansion of neoplastic stem cell population (Coussens and Werb 2002; Meng and Riordan 2006) which, in turn, may further select the addiction of oncogenic gene mutations, which drive PC cells to invasion, metastasis, and resistance to therapy.

Our knowledge of the intricated cross-links between genetic and epigenetic events occurring in PC has registered exciting progresses during the last decade.

The role of the three-dimensional texture and regulation of chromatin function in PC has been partially uncovered, and this has led to hypothesize the therapeutic use of drugs and small molecules such as HDAC inhibitors or DNA methylating and demethylating agents, acting as epigenetic modulators, as alternative or complementary tools for fighting aggressive PC (Murr 2010). The emerging data confirm that prostate carcinogenesis bases upon a definite group of interconnecting key signaling pathways.

Large scales of studies and carefully designed clinical trials will be required to validate novel effective therapeutic strategies for the treatment of PC.

The availability of next-generation sequencing will provide us with a broad genotyping platform which contribute to further refine our notions, shortening the time occurring to set-up multi-faceted molecular strategies tailored against the multiple molecular alterations responsible for the killing ability of advanced, androgen-independent, prostate cancers (Dias-Santagata et al. 2010) (Fig. 8.1).



Fig. 8.1 Crossroads of molecular pathways involved in PCa. In this picture the most frequent intersections between the major molecular pathways promoting growth, survival, invasiveness, metastasis and therapy-resistance of PC cells, potentially targetable for therapeutic strategies are summarized

References

- Akhurst RJ, Derynck R (2001) TGF-beta signaling in cancer a double-edged sword. Trends Cell Biol 11:S44–S51
- An WG et al (1998) Stabilization of wild-type p53 by hypoxia-inducible factor 1alpha. Nature 392(6674):405–408
- Anastasiadis AG, Bemis DL, Stisser BC, Salomon L, Ghafar MA, Buttyan R (2003) Tumor cell hypoxia and the hypoxia-response signaling system as a target for prostate cancer therapy. Curr Drug Target 4:191–196
- Attar RM, Takimoto CH, Gottardis MM (2009) Castration-resistant prostate cancer: locking up the molecular escape routes. Clin Cancer Res 15:3251–3255
- Banham AH, Boddy J, Launchbury R, Han C, Turley H, Malone PR, Harris AL, Fox SB (2007) Expression of the forkhead transcription factor FOXP1 is associated both with hypoxia inducible factors (HIFs) and the androgen receptor in prostate cancer but is not directly regulated by androgens or hypoxia. Prostate 67:1091–1098
- Boddy JL, Fox SB, Han C, Campo L, Turley H, Kanga S, Malone PR, Harris AL (2005) The androgen receptor is significantly associated with vascular endothelial growth factor and hypoxia sensing via hypoxia-inducible factors HIF-1a, HIF-2a, and the prolyl hydroxylases in human prostate cancer. Clin Cancer Res 11:7658–7663
- Bonaccorsi L, Marchiani S, Muratori M, Forti G, Baldi E (2004) Gefitinib ('IRESSA', ZD1839) inhibits EGFinduced invasion in prostate cancer cells by suppressing PI3 K/AKT activation. J Cancer Res Clin Oncol 130:604–614
- Burfeind P, Chernicky CL, Rininsland F, Ilan J (1996) Antisense RNA to the type I insulin-like growth factor receptor suppresses tumor growth and prevents invasion by rat prostate cancer cells *in vivo*. PNAS 93:7263–7268
- Byrne RL, Leung H, Neal DE (1996) Peptide growth factors in the prostate as mediators of stromal epithelial interaction. Br J Urol 77:627–633
- Camps JL, Chang SM, Hsu TC, Freeman MR, Hong SJ, Zhau HE, von Eschenbach AC, Chung LW (1990) Fibroblastmediated acceleration of human epithelial tumor growth *in vivo*. PNAS 87:75–79
- Cardillo MR, Petrangeli E, Perracchio L, Salvatori L, Ravenna L, Di Silverio F (2000) Transforming growth factor-beta expression in prostate neoplasia. Anal Quant Cytol Histol 22:1–10
- Castellano G, Malaponte G, Mazzarino MC, Figini M, Marchese F, Gangemi P et al (2008) Activation of the osteopontin/matrix metalloproteinase-9 pathway correlates with prostate cancer progression. Clin Cancer Res 14:7470–7480
- Chang IY, Youn CK, Kim HB et al (2005) Oncogenic H-Ras upregulates expression of Ku80 to protect cells from gamma-ray irradiation in NIH3T3 cells. Cancer Res 65:6811–6819
- Chen L, Elahi A, Pow-Sang J, Lazarus P, Park J (2003) Association between polymorphism of human oxoguanine glycosylase 1 and risk of prostate cancer. J Urol 170:2471–2474
- Chinnaiyan P, Vallabhaneni G, Armstrong E, Huang SM, Harari PM (2005) Modulation of radiation response by histone deacetylase inhibition. Int J Radiat Oncol Biol Phys 62:223–229
- Chipuk JE, Cornelius SC, Pultz NJ, Jorgensen JS, Bonham MJ, Kim SJ, Danielpour D (2002) The androgen receptor represses transforming growth factor-beta signaling through interaction with Smad3. J Biol Chem 277:1240–1248
- Cho-Chung YS (1989) Site-selective 8-chloro- cyclic adenosine 3',5'-monophosphate as a biologic modulator of cancer: restoration of normal control mechanisms. J Natl Cancer Inst 81:982–987
- Cho-Chung YS (1990) Role of cyclic AMP receptor proteins in growth, differentiation, and suppression of malignancy: new approaches to therapy. Cancer Res 50:7093–7100
- Choudhury A, Cuddihy A, Bristow RG (2006) Radiation and new molecular agents part I: targeting ATM-ATR checkpoints, DNA repair, and the proteasome. Semin Radiat Oncol 16:51–58
- Coffey RJ Jr, Shipley GD, Moses HL (1986) Production of transforming growth factors by human colon cancer lines. Cancer Res 46:1164–1169
- Coussens LM, Werb Z (2002) Inflammation and cancer. Nature 420:860-867

- Craft N, Shostak Y, Carey M, Sawyers CL (1999) A mechanism for hormone-independent prostate cancer through modulation of androgen receptor signaling by the HER-2/neu tyrosine kinase. Nat Med 5:280–285
- Culig Z, Hobisch A, Cronauer MV, Radmayr C, Trapman J, Hittmair A, Bartsch G, Klocker H (1994) Androgen receptor activation in prostatic tumor cell lines by insulinlike growth factor-I, keratinocyte growth factor, and epidermal growth factor. Cancer Res 54:5474–5478
- Cunha GR, Donjacour AA (1989) Mesenchymal–epithelial interactions in the growth and development of the prostate. Cancer Treat Res 46:159–175
- Cunha GR, Hayward SW, Dahiya R, Foster BA (1996) Smooth muscle–epithelial interactions in normal and neoplastic prostatic development. Acta Anat 155:63–72
- Dayyani F, Gallick GE, Logothetis CJ, Corn PG (2011) Novel therapies for metastatic castrateresistant prostate cancer. J Natl Cancer Inst 103(22):1665–1675
- Delongchamps NB, Peyromaure M, Dinh-Xuan AT (2006) Role of vascular endothelial growth factor in prostate cancer. Urology 68:244–248
- Derynck R, Zhang YE (2003) Smad-dependent and Smadindependent pathways in TGF-beta family signalling. Nature 425:577–584
- Di Lorenzo G, Tortora G, D'Armiento FP, De Rosa G, Staibano S, Autorino R, D'Armiento M, De Laurentiis M, De Placido S, Catalano G et al (2002) Expression of epidermal growth factor receptor correlates with disease relapse and progression to androgen-independence in human prostate cancer. Clin Cancer Res 8:3438–3444
- Di Lorenzo G, Autorino R, D'Armiento FP, Mignogna C, De Laurentiis M, De Sio M et al (2004) Expression of proto-oncogene c-kit in high risk prostate cancer. Eur J Surg Oncol 30:987–992
- Dias-Santagata D, Akhavanfard S, David SS, Vernovsky K, Kuhlmann G, Boisvert SL, Stubbs H, McDermott U, Settleman J, Kwak EL, Clark JW, Isakoff SJ, Sequist LV, Engelman JA, Lynch TJ, Haber DA, Louis DN, Ellisen LW, Borger DR, Iafrate AJ (2010) Rapid targeted mutational analysis of human tumours: a clinical platform to guide personalized cancer medicine. EMBO Mol Med 2(5):146–158
- Dittmann K, Mayer C, Fehrenbacher B et al (2005a) Radiationinduced epidermal growth factor receptor nuclear import is linked to activation of DNA-dependent protein kinase. J Biol Chem 280:31182–31189
- Dittmann K, Mayer C, Rodemann HP (2005b) Inhibition of radiationinduced EGFR nuclear import by C225 (Cetuximab) suppresses DNA-PK activity. Radiother Oncol 76:157–161
- El-Amm J, Aragon-Ching JB (2013) The changing landscape in the treatment of metastatic castration resistant prostate cancer. Ther Adv Med Oncol 5(1):25–40
- Ellis L, Hammers H, Pili R (2009) Targeting tumor angiogenesis with histone deacetylase inhibitors. Cancer Lett 280(2):145–153. Review
- Feldser D et al (1999) Reciprocal positive regulation of hypoxia-inducible factor 1alpha and insulin-like growth factor 2. Cancer Res 59(16):3915–3918
- Festuccia C, Muzi P, Millimaggi D, Biordi L, Gravina GL, Speca S, Angelucci A, Dolo V, Vicentini C, Bologna M (2005) Molecular aspects of gefitinib antiproliferative and proapoptotic effects in PTEN-positive and PTENnegative prostate cancer cell lines. Endocr-Relat Cancer 12:983–998
- Fong GH, Rossant J, Gertsenstein M, Breitman ML (1995) Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium. Nature 376:66–70
- Gerdes MJ, Dang TD, Larsen M, Rowley DR (1998) Transforming growth factor-beta1 induces nuclear to cytoplasmic distribution of androgen receptor and inhibits androgen response in prostate smooth muscle cells. Endocrinology 139:3569–3577
- Gerdes MJ, Larsen M, Dang TD, Ressler SJ, Tuxhorn JA, Rowley DR (2004) Regulation of rat prostate stromal cell myodifferentiation by androgen and TGF-beta1. Prostate 58:299–307
- Gonzalez-Herrera IG, Prado-Lourenco L, Pileur F, Conte C, Morin A, Cabon F, Prats H, Vagner S, Bayard F, Audigier S et al (2006) Testosterone regulates FGF-2 expression during testis maturation by an IRES-dependent translational mechanism. FASEB J 20:476–478
- Heinlein CA, Chang C (2004) Androgen receptor in prostate cancer. Endocr Rev 25:276-308

- Henshall SM, Quinn DI, Lee CS, Head DR, Golovsky D, Brenner PC, Delprado W, Stricker PD, Grygiel JJ, Sutherland RL (2001) Altered expression of androgen receptor in the malignant epithelium and adjacent stroma is associated with early relapse in prostate cancer. Cancer Res 61:423–427
- Horii K, Suzuki Y, Kondo Y, Akimoto M, Nishimura T, Yamabe Y et al (2007) Androgendependent gene expression of prostatespecific antigen is enhanced synergistically by hypoxia in human prostate cancer cells. Mol Cancer Res 5:383–391
- Ivan M, Kaelin WG Jr (2001) The von Hippel–Lindau tumor suppressor protein. Curr Opin Genet Dev 11(1):27–34
- Johansson A, Rudolfsson S, Hammarsten P, Halin S, Pietras K, Jones J et al (2010) Mast cells are novel independent prognostic markers in prostate cancer and represent a target for therapy. Am J Pathol 177:1031–1041
- Jones PA, Baylin SB (2002) The fundamental role of epigenetic events in cancer. Nat Rev Genet 3:415–428
- Kang JH et al (2008) CCAAT box is required for the induction of human thrombospondin-1 gene by trichostatin A. J Cell Biochem 104(4):1192–1203
- Kerbel R, Folkman J (2002) Clinical translation of angiogenesis inhibitors. Nat Rev Cancer 2(10):727–739
- Kim MS et al (2001) Histone deacetylases induce angiogenesis by negative regulation of tumor suppressor genes. Nat Med 7(4):437–443
- Kwon HJ et al (2002) Histone deacetylase inhibitor FK228 inhibits tumor angiogenesis. Int J Cancer 97(3):290–296
- Leong KG, Wang BE, Johnson L, Gao WQ (2008) Generation of a prostate from a single adult stem cell. Nature 456:804–808
- Leotoing L, Manin M, Monte D, Baron S, Communal Y, Lours C, Veyssiere G, Morel L, Beaudoin C (2007) Crosstalk between androgen receptor and epidermal growth factor receptorsignalling pathways: a molecular switch for epithelial cell differentiation. J Mol Endocrinol 39:151–162
- Liu Y, Majumder S, McCall W, Sartor CI, Mohler JL, Gregory CW, Earp HS, Whang YE (2005) Inhibition of HER-2/neu kinase impairs androgen receptor recruitment to the androgen responsive enhancer. Cancer Res 65:3404–3409
- Liu T et al (2006) Histone deacetylase inhibitors: multifunctional anticancer agents. Cancer Treat Rev 32(3):157–165
- Ma BB, Bristow RG, Kim J, Siu LL (2003) Combined-modality treatment of solid tumors using radiotherapy and molecular targeted agents. J Clin Oncol 21:2760–2776
- Mabjeesh NJ, Amir S (2007) Hypoxia-inducible factor (HIF) in human tumorigenesis. Histol Histopathol 22(5):559–572
- Mantalaris A, Panoskaltsis N, Sakai Y et al (2001) Localization of androgen receptor expression in human bone marrow. J Pathol 193:361–366
- Marignol L, Coffey M, Lawler M, Hollywood D (2008) Hypoxia in prostate cancer: a powerful shield against tumour destruction? Cancer Treat Rev 34(4):313–327
- Mellinghoff IK, Vivanco I, Kwon A, Tran C, Wongvipat J, Sawyers CL (2004) HER2/neu kinasedependent modulation of androgen receptor function through effects on DNA binding and stability. Cancer Cell 6:517–527
- Memarzadeh S, Xin L, Mulholland DJ, Mansukhani A, Wu H, Teitell MA, Witte ON (2007) Enhanced paracrine FGF10 expression promotes formation of multifocal prostate adenocarcinoma and an increase in epithelial androgen receptor. Cancer Cell 12:572–585
- Meng X, Riordan NH (2006) Cancer is a functional repair tissue. Med Hypotheses 66:486-490
- Murr R (2010) Interplay between different epigenetic modifications and mechanisms. Adv Genet 70:101–141
- Nakano K, Fukabori Y, Itoh N, Lu W, Kan M, McKeehan WL, Yamanaka H (1999) Androgenstimulated human prostate epithelial growth mediated by stromal-derived fibroblast growth factor-10. Endocr J 46:405–413

- Niu Y, Xu Y, Zhang J, Bai J, Yang H, Ma T (2001) Proliferation and differentiation of prostatic stromal cells. BJU Int 87:386–393
- Niu Y, Chang TM, Yeh S, Ma WL, Wang YZ, Chang C (2010) Differential androgen receptor signals in different cells explain why androgen-deprivation therapy of prostate cancer fails. Oncogene 29:3593–3604
- Norris AM, Woodruff RD, D'Agostino RB Jr, Clodfelter JE, Scarpinato KD (2007) Elevated levels of the mismatch repair protein PMS2 are associated with prostate cancer. Prostate 67:214–225
- Ohlson N, Bergh A, Stattin P, Wikstrom P (2007) Castrationinduced epithelial cell death in human prostate tissue is related to locally reduced IGF-1 levels. Prostate 67:32–40
- Orio F Jr, Terouanne B, Georget V, Lumbroso S, Avances C, Siatka C, Sultan C (2002) Potential action of IGF-1 and EGF on androgen receptor nuclear transfer and transactivation in normal and cancer human prostate cell lines. Mol Cell Endocrinol 198:105–114
- Ornitz DM, Itoh N (2001) Fibroblast growth factors. Genome Biol 2(3):REVIEWS3005. Epub 2001 Mar 9. Review. PubMed PMID: 11276432; PubMed Central PMCID: PMC138918
- Peterziel H, Mink S, Schonert A, Becker M, Klocker H, Cato AC (1999) Rapid signalling by androgen receptor in prostate cancer cells. Oncogene 18:6322–6329
- Pittoni P, Piconese S, Tripodo C, Colombo MP (2010) Tumor-intrinsic and -extrinsic roles of c-Kit: mast cells as the primary off-target of tyrosine kinase inhibitors. Oncogene 30:757–769
- Pittoni P, Tripodo C, Piconese S, Mauri G, Parenza M, Rigoni A, Sangaletti S, Colombo MP (2011) Mast cell targeting hampers prostate adenocarcinoma development but promotes the occurrence of highly malignant neuroendocrine cancers. Cancer Res 71(18):5987–5997
- Pollak M, Beamer W, Zhang JC (1998) Insulin-like growth factors and prostate cancer. Cancer Metastasis Rev 17:383–390
- Prtilo A, Leach FS, Markwalder R et al (2005) Tissue microarray analysis of hMSH2 expression predicts outcome in men with prostate cancer. J Urol 174:1814–1818, discussion p. 18
- Qian DZ et al (2004) The histone deacetylase inhibitor NVP-LAQ824 inhibits angiogenesis and has a greater antitumor effect in combination with the vascular endothelial growth factor receptor tyrosine kinase inhibitor PTK787/ZK222584. Cancer Res 64(18):6626–6634
- Richard DE et al (1999) P42/p44 mitogen-activated protein kinases phosphorylate hypoxiainducible factor 1alpha (HIF-1alpha) and enhance the transcriptional activity of HIF-1. J Biol Chem 274(46):32631–32637
- Rinaldo F, Li J, Wang E, Muders M, Datta K (2007) RalA regulates vascular endothelial growth factor-C (VEGF-C) synthesis in prostate cancer cells during androgen ablation. Oncogene 26:1731–1738
- Roberts AB, Sporn MB, Assoian RK, Smith JM, Roche NS, Wakefield LM, Heine UI, Liotta LA, Falanga V, Kehrl JH et al (1986) Transforming growth factor type beta: rapid induction offibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. PNAS 83:4167–4171
- Rochester MA, Riedemann J, Hellawell GO, Brewster SF, Macaulay VM (2005) Silencing of the IGF1R gene enhances sensitivity to DNA-damaging agents in both PTEN wild-type and mutant human prostate cancer. Cancer Gene Ther 12:90–100
- Rosini P, Bonaccorsi L, Baldi E, Chiasserini C, Forti G, De Chiara G, Lucibello M, Mongiat M, Iozzo RV, Garaci E et al (2002) Androgen receptor expression induces FGF2, FGF-binding protein production, and FGF2 release in prostate carcinoma cells: role of FGF2 in growth, survival, and androgen receptor down-modulation. Prostate 53:310–321
- Rossig L et al (2002) Inhibitors of histone deacetylation downregulate the expression of endothelial nitric oxide synthase and compromise endothelial cell function in vasorelaxation and angiogenesis. Circ Res 91(9):837–844
- Russell PJ, Bennett S, Stricker P (1998) Growth factor involvement in progression of prostate cancer. Clin Chem 44:705–723
- Rybicki BA, Conti DV, Moreira A, Cicek M, Casey G, Witte JS (2004) DNA repair gene XRCC1 and XPD polymorphisms and risk of prostate cancer. Cancer Epidemiol Biomark Prev 13:23–29

- Salm SN, Koikawa Y, Ogilvie V, Tsujimura A, Coetzee S, Moscatelli D, Moore E, Lepor H, Shapiro E, Sun TT et al (2000) Generation of active TGF-beta by prostatic cell cocultures using novel basal and luminal prostatic epithelial cell lines. J Cell Physiol 184:70–79
- Sar M, Lubahn DB, French FS, Wilson EM (1990) Immunohistochemical localization of the androgen receptor in rat and human tissues. Endocrinology 127:3180–3186
- Saric T, Shain SA (1998) Androgen regulation of prostate cancer cell FGF-1, FGF-2, and FGF-8: preferential downregulation of FGF-2 transcripts. Growth Fact 16:69–87
- Sarwar M, Persson JL (2011) The protein kinase a (PKA) intracellular pathway and androgen receptor: a novel mechanism underlying the castration-resistant and metastatic prostate cancer. J Cancer Sci Ther S5:003
- Sasakawa Y et al (2003) Antitumor efficacy of FK228, a novel histone deacetylase inhibitor, depends on the effect on expression of angiogenesis factors. Biochem Pharmacol 66(6):897–906
- Semenza GL (1998) Hypoxia-inducible factor 1: master regulator of O2 homeostasis. Curr Opin Genet Dev 8(5):588–594
- Siegel R, Naishadham D, Jemal A (2012) Cancer statistics, 2012. CA Cancer J Clin 62(10):29
- Sporn MB, Roberts AB (1992) Transforming growth factorbeta: recent progress and new challenges. J Cell Biol 119:1017–1021
- Stiehl DP et al (2002) Normoxic induction of the hypoxia-inducible factor lalpha by insulin and interleukin-1beta involves the phosphatidylinositol 3-kinase pathway. FEBS Lett 512(1-3):157-162
- Sun X, Chen C, Vessella RL, Dong JT (2006) Microsatellite instability and mismatch repair target gene mutations in cell lines and xenografts of prostate cancer. Prostate 66:660–666
- Wang H, Song K, Sponseller TL, Danielpour D (2005) Novel function of androgen receptorassociated protein 55/Hic-5 as a negative regulator of Smad3 signaling. J Biol Chem 280:5154–5162
- Wang X, Yin L, Rao P, Stein R, Harsch KM, Lee Z, Heston WD (2007) Targeted treatment of prostate cancer. J Cell Biochem 102:571–579
- Weidemann A, Johnson RS (2008) Biology of HIF-1alpha. Cell Death Differ 15(4):621-627
- Wolk A, Mantzoros CS, Andersson SO, Bergstrom R, Signorello LB, Lagiou P, Adami HO, Trichopoulos D (1998) Insulin-like growth factor 1 and prostate cancer risk: a populationbased, case–control study. J Natl Cancer Inst 90:911–915
- Wu Y, Zhao W, Zhao J, Pan J, Wu Q, Zhang Y, Bauman WA, Cardozo CP (2007) Identification of androgen response elements in the insulin-like growth factor I upstream promoter. Endocrinology 148:2984–2993
- Xu J, Zheng SL, Turner A et al (2002) Associations between hOGG1 sequence variants and prostate cancer susceptibility. Cancer Res 62:2253–2257
- Yeh S, Lin HK, Kang HY, Thin TH, Lin MF, Chang C (1999) From HER2/Neu signal cascade to androgen receptor and its coactivators: a novel pathway by induction of androgen target genes through MAP kinase in prostate cancer cells. PNAS 96:5458–5463
- Yeh CC, Lee C, Dahiya R (2001) DNA mismatch repair enzyme activity and gene expression in prostate cancer. Biochem Biophys Res Commun 285:409–413
- Yoshizawa A, Ogikubo S (2006) IGF binding protein-5 synthesis is regulated by testosterone through transcriptional mechanisms in androgen responsive cells. Endocr J 53:811–818
- Yuan TC, Veeramani S, Lin MF (2007) Neuroendocrine-like prostate cancer cells: neuroendocrine transdifferentiation of prostate adenocarcinoma cells. Endocr Relat Cancer 14:531–547
- Zavadil J, Bottinger EP (2005) TGF-beta and epithelial-to-mesenchymal transitions. Oncogene 24:5764–5774
- Zelzer E et al (1998) Insulin induces transcription of target genes through the hypoxia-inducible factor HIF-1alpha/ARNT. EMBO J 17(17):5085–5094
- Zhou J, Schmid T, Schnitzer S, Brüne B (2006) Tumor hypoxia and cancer progression. Cancer Lett 237(1):10–21
- Zhu B, Kyprianou N (2005) Transforming growth factor beta and prostate cancer. Cancer Treat Res 126:157–173

- Zhu ML, Kyprianou N (2008) Androgen receptor and growth factor signaling cross-talk in prostate cancer cells. Endocr Relat Cancer 15(4):841–849
- Zhu B, Fukada K, Zhu H, Kyprianou N (2006) Prohibitin and cofilin are intracellular effectors of transforming growth factor beta signaling in human prostate cancer cells. Cancer Res 66:8640–8647
- Zundel W et al (2000) Loss of PTEN facilitates HIF-1-mediated gene expression. Genes Dev 14(4):391-396