## **Chapter 6 Metastatic Dissemination**

**Stefania Staibano**

**Abstract** In spite of recent developments in diagnosis, staging and treatment, most patients with advanced prostate cancer will ultimately progress from androgensensitive to an irreversible castration-resistant disease. These androgen-independent cancers frequently give rise to widespread metastasis, dramatically reducing the median survival of patients (Tannock et al, N Engl J Med, 351(15):1502–1512, 2004) and accounting for more than 32, 000 deaths/year in USA (Jemal et al, CA Cancer J Clin, 60:277–300, 2010), which correspond to over 90 % of PC related mortality (Man, Gardner, Int J Biol Sci, 4(4):246–258, 2008).

It is a common belief that cancer metastasis result from a multi-stage nonrandom process characterized by intricate interactions between cancer cells and the host microenvironment, leading to the detachment of cancer cells from their tissue of origin, their dissemination through the bloodstream and to invasion of the target metastatic site (Patel et al, Future Oncol, 7(11):1285–1297, 2011).

Metastasis represents yet one of the most enigmatic aspects of prostate cancer pathogenesis, in which a cascade of proteolytic enzymes, inflammatory cytokines, growth factors, activated oncogenes, oxidative stress and hypoxia linked proteins and adhesion molecules, orchestrate a continuous loop that enable migrating cancer cells detached from the primary tumor bulk, to survive and proliferate in an adverse remote body microenvironment.

In this chapter, we discuss the nature and alterations of the signaling pathways involved in the development of prostate cancer metastasis, reporting the current status of knowledge on the changes occurring either in prostate cancer cells and in tumor-associated stromal tissue, with particular emphasis to the process of epithelial-mesenchymal transition ("phenotypic plasticity") and to the role of cancer stem cells in prostate cancer progression and metastasis.

111

S. Staibano  $(\boxtimes)$ 

Department of Advanced Biomedical Sciences, Pathology Section, Faculty of Medicine and Surgery, University of Naples "Federico II", via S. Pansini, n.5, Naples, Italy e-mail: [staibano@unina.it](mailto:staibano@unina.it)

We will highlight, also, the emerging data concerning new therapeutic targets for treatment of metastatic prostate cancer that, while deserving further inquiry, look very promising to improve our chances to successful approach the advanced disease or, even, primarily reduce the risk of metastasis from castration-resistant prostate cancer (Vashisht, Bagler, PLoS One, 7(11):e49401, 2012).

Metastases represent the most fearful evolution of advanced/systemic prostate cancer progressed into a castration-resistance state after first-instance deprivation therapy.

Before the onset of metastasis, prostate cancer is usually characterized by a long latency period, in which genetic (Nguyen and Massague [2007;](#page-11-0) Zhao et al. [2013\)](#page-14-0) and epigenetic (Rodenhiser [2009\)](#page-12-0) cellular alterations lead to changes in cancer cells molecular phenotype, with the gain of both cytoskeletal motility and the ability to detach from the tissue of origin. The acquired abilities of epithelial prostate cancer cells are critically boosted by activated prostatic stromal cells, as tumor-associated macrophages (TAMs), cancer-associated fibroblasts (CAFs) and endothelial cells (Wang et al. [2013\)](#page-13-0).

Besides their role in tumor-associated angiogenesis, CD31/CD34-positive endothelial cells lining microvessels decrease upon castration, increasing instead when prostate cancer progress to castration-resistance. Recently, it has shown *in vitro* that endothelial cells secrete high levels of IL-6. This cytokine down-regulates AR and activates the TGFbeta/MMP9 signaling pathway in prostate cancer cells, contributing then to their invasive and metastasizing ability (Wang et al. [2013\)](#page-13-0).

TAMs produce several migration-stimulating factors, as CXCL12, IL-6 and TNF (Allavena et al. [2008\)](#page-9-0). Activated CAFs mostly exhibit a myofibroblastic phenotype induced either by the direct physical contact with cancer cells and *via* the hyperstimulation, by several tumor- and hypoxia associated growth factors, as EGF, FGF, IGF, VEGF. CAFs overproduce TGFbeta (Roodman [2004\)](#page-12-1), which intervene in ECM remodeling (Allavena et al. [2008\)](#page-9-0) and in the induction of epithelial-mesenchymal transition (EMT) (Yilmaz and Christofori [2009\)](#page-13-1) of metastasizing cells.

Extracellular matrix proteins, facilitating either tumor growth and metastasis, continuously accumulate in tumor stroma. This is the case for versican, a large proteoglycan associated with metastasis and poor outcome of prostate cancer and several solid malignant cancers. It has been shown to regulate cancer cell adhesion, proliferation, migration, angiogenesis, invasion and metastasis mainly through physical interactions mediated by chondroitin and dermatan sulfate side chains; looking particularly attractive as a possible adjunctive therapeutic target for aggressive prostate cancers (Du et al. [2013\)](#page-10-0).

Even a disturbance of the interplay between the electrical and metabolic activity of prostate cells seems to play a role in the gain of propensity to metastasize of prostate cancer. It has been, in fact, recently reported that an altered expression of connexins, which form intercellular channels involved in gap-junction-mediated intercellular coupling, might be correlated with the invasive potential of cancer cells (Czy $\ddot{z}$  et al. [2012\)](#page-10-1). This finding, however, deserves further investigation.

## 6 Metastatic Dissemination 113

The acquired EMT ability of prostate cancer cells leading to the detachment from the bulk of primary cancer, is conditioned by the dramatic loss of adhesion proteins, as E-cadherin (Yates [2011;](#page-13-2) Lazari et al. [2013\)](#page-11-1) and their regulating transcriptional inducers, as the SAM Pointed Domain ETS transcription Factor (SPDEF) (Pal et al. [2013\)](#page-12-2), and by the increase of their transcriptional repressors, as the Wilms' tumor gene (WT1) (Brett et al. [2013\)](#page-10-2). Recently, the altered expression of the human metastasis-associated gene 1 (MTA1) has been found strictly associated with the pAkt/E-cadherin pathway regulation and with metastatic prostate cancer (Wang et al. [2012\)](#page-13-3), and, a combined testing strategy for detecting MTA1 and E-cadherin, has been proposed for selecting high-risk prostate cancer patients (Fan et al. [2012\)](#page-10-3).

Before permeating blood vessels, detached tumor cells have to escape anoikis and gain survival benefits (Hu et al. [2012\)](#page-11-2). The anoikis-resistance and EMT properties of prostate tumor cells are mediated by several molecular players, including members of the Notch signaling pathway, as well of the Akt survival pathway including the early-recruited focal adhesion player tallin. Tallin mediates integrin activation and induces downstream survival pathways resulting in the promotion of cancer cells progression to metastasis (Desiniotis and Kyprianou [2011\)](#page-10-4).

Both Notch-related proteins and tallin appear, then, as promising candidate as either prognostic markers and therapeutic targets in metastasizing prostate cancers.

To survive in the bloodstream, prostate cancer cells activate multiple survival pathways, comprising the overexpression of several members of the anti-apoptotic Bcl-2 protein family, combined with the inactivation of the FADD death receptor pathway, or the lack of expression of pro-apoptotic effector proteins as Bax and caspases (Igney and Krammer [2002\)](#page-11-3).

Moreover, circulating prostate cancer cells may activate survivin expression and undergo to autophagy, to survive in the absence of sufficient extracellular nutrients (Roca et al. [2008\)](#page-12-3).

While metastasizing prostate cancer cells may optionally variously localize in several body sites, as lung and liver, they invariably hit the bone (Osanto and Van Poppel [2012\)](#page-12-4) (Fig. [6.1\)](#page-4-0).

In bone-metastasizing cancers, the CG-protein-coupled calcium sensing receptor (CaSR), which is primarily involved in the feedback regulation of extracellular free ionised calcium  $(Ca2+)$ , may act as an oncogene, associating also with cancer progression. In prostate cancer, its altered expression seems to facilitate bone metastasis (Brennan et al. [2012\)](#page-10-5).

Disseminated prostate cancer cells (DPCC) reaching the bone marrow occupy the same bone niche in which hematopoietic stem cells reside in a quiescent state (Taichman [2005\)](#page-13-4). This has led to the concept that DPCCs behave as "parasites" of the hematopoietic niche.

DPCCs which evade immune attack and/or chemotherapy cytotoxicity may outlive for a variable time in bone marrow of patients after radical prostatectomy or chemotherapy (Morgan et al. [2009;](#page-11-4) Pfitzenmaier et al. [2007\)](#page-12-5), in an auto-induced reversible state of growth arrest, the so-called "tumor-dormancy" (Townson and Chambers [2006;](#page-13-5) Aguirre-Ghiso [2007;](#page-9-1) Shiozawa et al. [2008b;](#page-12-6) Joyce and Pollard [2009\)](#page-11-5). This underlies the troublesome unresolved phenomenon of "minimal residual



 $\blacktriangleleft$ 

disease", responsible for most cases of prostate cancer recurrence and therapy failure. The overall regulation of this process is still under active investigation but it has been now accepted that it involves prostate cancer stem cells (PCSC). From the first appearance on the scenario of cancer metastasis of solid tumors, in 2003 (Al-Hajj et al. [2003\)](#page-9-2), a definitive consensus about their origin and specific markers has not been reached yet.

Several putative surface markers, in fact, are shared also by normal stem cells (Patrawala et al. [2007;](#page-12-7) Collins et al. [2005\)](#page-10-6) as well as by different solid tumors. This is the case for CD133/prominin-1 and CD44 (Patrawala et al. [2007\)](#page-12-7) that have been found expressed in CSC of lung, breast, colon, ovarian and head and neck squamous cell carcinomas (Cui et al. [2011;](#page-10-7) Chu et al. [2009;](#page-10-8) Shi et al. [2010\)](#page-12-8). As well, CD133, CD44, integrins, Sca-1, and breast cancer resistance protein (BRCP) are expressed either in PCSC and in normal prostate stem cells (Yu et al. [2012;](#page-13-6) Tang et al. [2007\)](#page-13-7); Oct-3/4, beta-catenin and SMO are stemness markers expressed by most of normal and neoplastic stem cells (Patrawala et al. [2006\)](#page-12-9). In addition, there is still a considerable variance among the different antibodies available for the detection of stem cells markers and this may explain, almost partially, the presence of some overlaps or discrepancies between the many existing studies on this topic.

Encouraging results indicate that ALDH1A1, a member of ALDH family of proteins involved in the intracellular production of retinoic acid, could be considered as a promising marker of stemness for prostate cancer cells (Li et al. [2010\)](#page-11-6). ALDH1A1 overexpressing prostate cancer cells display, in fact, high migration and clonogenic ability *in vitro* and metastatic ability *in vivo* (van den Hoogen et al. [2010\)](#page-13-8). As well, a second member of the ALDH family, ALDH7A1, seems to be involved in the bone metastasis formation (van den Hoogen et al. [2010\)](#page-13-8), since its knockdown results in inhibition of experimentally induced intra-bone metastasis.

<span id="page-4-0"></span>**Fig. 6.1 The signaling pathways involved in the development of prostate cancer bone metastasis**. Metastasis result from interactions between cancer cells and the host microenvironment that enable them to detach from the primary tumor bulk, disseminate through the bloodstream and invade of metastatic site. These steps are regulated by a cascade of proteolytic enzymes, inflammatory cytokines, growth factors, activated oncogenes, oxidative stress and hypoxia linked proteins, and adhesion molecules. Activated stromal cells, as tumor-associated macrophages (*TAMs*), cancer-associated fibroblasts (*CAFs*), and endothelial cells, favor the entire process: TAMs produce several migration-stimulating factors as CXCL12, IL-6, and TNF; CAFs intervene in ECM remodeling and in the induction of epithelial-mesenchymal transition (EMT) of metastasizing cells. Circulating cells activate multiple survival pathways, as the overexpression of several members of the anti-apoptotic Bcl2 protein family and the activation of survivin expression. Disseminated prostate cancer cells (DPCC) reaching the bone marrow occupy a bone niche and, when evade immune attack and/or chemotherapy cytotoxicity, may outlive for a variable time in an autoinduced reversible state of growth arrest, the so-called "tumor-dormancy". Dormant DPCCs give rise to bone metastatic lesions by re-entering the cell cycle and proliferating. The causative factors leading to this process are still a matter of investigation, but it seems involve prostate cancer stem cells (PCSC)

It has become clear, however, that the existence of a single reliable marker of PCSCs doesn't exist and a definite combination of markers expression may, instead, identify the metastatic profile of PCSC (Eaton et al. [2010\)](#page-10-9). To support this idea, it has been shown that the co-expression of CD166 (epithelial stem cell marker) (Dalerba et al. [2007\)](#page-10-10), CD151 (marker of stem-like tumor stromal cells) and the tumor rejection antigen/TRA-1-60 (Draper et al. [2002\)](#page-10-11) identifies prostate cancer cells with high ability in sphere formation *in vitro* and generating, *in vivo*, tumors capable of self-renewal and differentiation, consistent with stem cells properties.

Moreover, it has been reported that the signature for stem cell markers may also vary between metastasis and primitive tumors with different Gleason grade  $(Castellón et al. 2012).$  $(Castellón et al. 2012).$  $(Castellón et al. 2012).$ 

Traditionally, prostate CSC have been thought to derive from the basal cel layer, which express most of the known markers of stemness, as CD133, CD44, CD117, Tert, p63 (Tsujimura et al. [2002\)](#page-13-9). Several findings support this hypothesis. As an example, it emerged that normal basal cells of human prostate can initiate prostate cancer in immunocompromised mice (Goldstein et al. [2010\)](#page-11-7) and primary cells FACS-sorted confirmed the basal cell origin for prostate cancer (Goldstein et al. [2010;](#page-11-7) Lawson et al. [2010\)](#page-11-8).

On the other side, there are line of evidence that, in several instances, PCSC could have originated from prostate luminal-cells. For instance, a genetic lineagemarking study has shown that rare prostate luminal cells express the androgen/ARregulated transcriptional co-activator Nkx3-1 in absence of androgens (castrationresistant Nkx3-1-expressing cells, CARNs). CARNs show stem-cell properties, as they are self-renewing and reproduce prostate ducts in renal graft, and cause HGPIN and cancer following Pten deletion (Wang et al. [2009\)](#page-13-10).

Besides their origin, PCSC are considered as the guest actors in the bone marrow metastasis phase (Colombel et al. [2012\)](#page-10-13). They have, in fact, the necessary characteristics for survive and reproduce in the bone microenvironment.

The bone marrow niche, in turn, is critical for the progression from localized disease to distant metastases (Chung et al. [2005;](#page-10-14) Cher et al. [2006;](#page-10-15) Morrissey and Vessella [2007;](#page-11-9) Karlou et al. [2010\)](#page-11-10). The niche is composed by the endothelia of sinusoids (Kiel and Morrison [2008;](#page-11-11) Doan and Chute [2012\)](#page-10-16), osteoblasts, adipocytes, mesenchymal stem cells, and contains a soluble extracellular matrix rich in growth factors, cytokines (Bussard et al. [2008\)](#page-10-17) and nutrients, useful for cancer cell survival. In addition, it contains adhesion molecules (Taichman [2005;](#page-13-4) Yin and Li [2006;](#page-13-11) Arai et al. [2009\)](#page-9-3) as annexin II (Shiozawa et al. [2008a\)](#page-12-10), which interact with tumor cells and local osteoblasts and fibroblasts to provide the framework for the stable homing of prostate cancer cells (Shiozawa et al. [2008b\)](#page-12-6).

Among the several cytokines actively secreted by osteoblast, a pivotal role seems to be played by CXCL12, also known as stromal cell derived factor-1, with its receptors CXCR4 and CXCR7. These two receptors are strongly expressed by DPCC. The binding of CXCR4 and CXCR7 of prostate cancer cells with CXCL12 induces the expression of several adhesion molecules, which enhance their binding to the bone niche (Sun et al. [2005,](#page-13-12) [2007\)](#page-13-13).

This finding may have relevant implication on therapy, as it has been shown that molecular antagonists of the CXC4R, as the small molecule AMD3100 and the G-CSF analog Filgrastim is able to mobilize metastasizing prostate cancer cells from the bone marrow niche.

Another protein responsible for the reversible cell-cycle arrest of DPCC is the fibroblast secreted annexin II (Anxa2) which operates with its receptor Anxa2R in a manner similar to the CXCL12/CXCR4-CXCR7 pathway (Jung et al. [2007;](#page-11-12) Shiozawa et al. [2008a\)](#page-12-10).

The degree of expression of either CXCR4 and CXCR7 by prostate cancer cells has been found to correlate with a poor outcome of patients (Sun et al. [2003;](#page-13-14) Wang et al. [2008;](#page-13-15) Shiozawa et al. [2008b;](#page-12-6) Mai et al. [2000\)](#page-11-13). All these considerations have rendered the targeting of bone marrow niche molecules a particularly active and attractive research field.

Several reports indicate that the alteration of multiple other signaling pathways accounting for the tumorigenic potential of PCSC may be used to control them.

For instance, targeting NF-kB with small molecule inhibitors may block sphere generation *in vitro* and tumor-initiation *in vivo*, by purified naïve stem-like human prostatic cells (Rajasekhar et al. [2011\)](#page-12-11), thus supporting the reported adverse prognostic significance in terms of biochemical recurrence risk of the presence of NF-kB stained cells in positive margins of radical prostatectomy specimens (Ross et al. [2004\)](#page-12-12). Similarly, the therapeutic use of WNT inhibitors has been shown to reduce the self-renewal of PCSC and improve the outcome of patients harbouring tumors co-expressing Wnt3a, nuclear beta-catenin, keratin 18, CD133 and CD44 (Bisson and Prowse [2009\)](#page-10-18).

Moreover, the colonization of the skeleton by prostate cancer cells is mediated also by collagen type I, the most represented bone protein, mainly through the binding with the increased expression of integrin  $\alpha(2)\beta(1)$ . This integrin has been found elevated in PCa bone metastatic lesions compared to either primary tumors or their soft tissue metastases suggesting it is needed for the selective metastatization to the bone (Sottnik et al. [2013\)](#page-13-16).

Dormant DPCCs give rise to bone metastatic lesions by re-entering the cell cycle and proliferating. The causative factors leading to this process are still a matter of investigation.

The striking propensity to localize to the bone is shared also by other "big killers", as lung and breast cancer (Patel et al. [2011\)](#page-12-13). However, these other cancer types give rise to osteolytic (bone resorbing) bone marrow metastases, while prostate cancer can produce predominantly osteoblastic lesions (Zetter [1990;](#page-13-17) Jacobs [1983;](#page-11-14) Chappard et al. [2011\)](#page-10-19), via the inhibition of osteoblast apoptosis and the increase of osteoblast proliferation and metabolism, induced by parathyroid hormone (PTH), PTH-related protein and bone morphogenic proteins BMP (Keller et al. [2001\)](#page-11-15). In addition, the expression of BMP may lead to the osteoblastic differentiation of bone mesenchymal stem cells creating an autocrine and paracrine feedback loop between the prostate cancer epithelial cell and the bone microenvironment.

In contrast to the rapid progress being made in the development of anti-osteolytic therapies, the treatment of osteosclerotic MBD remains restricted to palliative radiotherapy for symptomatic solitary lesions and systemic taxane-based chemotherapy for widespread multiple lesions (Sturge et al. [2011\)](#page-13-18). Thus, new therapeutic strategies focused on the complex pathology of osteoblastic bone-forming metastases of prostate cancer are urgently needed and promising results start to emerge from current preclinical studies.

The "lethal phenotype" of metastatic castrate-resistant prostate cancer depends, then, from the bi-directional action of cancer epithelial cells in the bone and host stromal response to tumor cells (Loberg et al. [2005\)](#page-11-16).

Elucidating the bidirectional interactions between the cancer cell and host bone microenvironment is now an important area of prostate cancer research (Efstathiou and Logothetis [2010\)](#page-10-20).

By a clinical point-of-view, these osteoblastic metastases cause bone pain, and are constituted by disorganized neo-synthetized, unstructured "woven" bone which, similarly to that observed also for osteolytic lesions, frequently give rise to painful fractures (Roudier et al. [2003,](#page-12-14) [2008;](#page-12-15) Eastham [2007\)](#page-10-21).

The progressive filling of bone marrow by metastatic prostate cancer cells cause myelophtisis, leukoerythroblastic anemia (Eriksson et al. [1972;](#page-10-22) Shamdas et al. [1993\)](#page-12-16), up to bone marrow failure (Spivak [1994\)](#page-13-19). These phenomena are thought to be caused, at least in part, by the physical displacement out of their bone marrow niches of hematopoietic stem cells by prostate cancer cells. HPCs displaced in the bloodstream might then undergo to forced, but incomplete, differentiation into lineage-specific nonfunctional progenitors (Shiozawa et al. [2011\)](#page-12-17).

Patients with bone-metastatic prostate cancer experience a significative higher risk of death for disease when compared with patients without skeletal involvement (Norgaard et al. [2010\)](#page-12-18).

The rationale for this bone-forming activity could reside in its possible contribute to support availability of bone niches for the successful homing and expansion of metastasizing prostate cancer cells.

The last decade has registered significant advancement in the identification of the steps involved in the multilayered process of prostate cancer metastasis but further translational studies are needed, to shed new light on several fundamental questions:

– Do hormone receptors have a relevant role in the induction and establishment of prostate cancer metastasis?

Mounting evidence indicates that androgen receptor (AR) signaling continues to play a critical role in the growth of advanced PC despite androgen deprivation (Zheng et al. [2013\)](#page-14-1). Recent data indicate that convergence of oncogenic and hormone receptor pathways promotes the metastatic phenotype (Augello et al. [2013\)](#page-9-4). However, the downstream AR target genes involved in progression of castration-resistance are largely unknown. It has been reported that cyclin D1b, a splice variant of cyclin D1 exerting a highly oncogenic function in human cancers, promote AR-mediated activation of genes associated with metastatic phenotype in tumor xenograft models of prostate cancer (Augello et al. [2013\)](#page-9-4).

Moreover, Jin HJ and colleagues showed that the AR pathway induces prostate cell growth also via the induction of the synthesis of FoxA1 (Jin et al. [2013\)](#page-11-17). However, this protein, which is a transcription factor essential for the prostate lineage-specific gene expression, inhibits cell motility and epithelialto-mesenchymal transition (EMT) through AR-independent mechanism opposite to the action of AR signaling, thus behaving as an inhibitor of prostate cancer metastasis. In orthotopic mouse models, FoxA1 has been found up-regulated in localized prostate cancer and down-regulated in EMT bearing metastatic prostate cancer cells. Then, FoxA1 may be considered an AR-independent metastasis inhibitor that, following mutations, can contribute instead to prostate cancer progression.

WNT7B, as a direct AR target gene highly expressed in castration-resistant prostate cancer (CRPC), suggests that AR-regulated WNT7B signaling is critical for the growth of CRPC and development of the osteoblastic bone response characteristic of advanced PC (Zheng et al. [2013\)](#page-14-1). WNT7B is necessary for the growth of PC cells and this effect is enhanced under androgen-deprived conditions; it promotes the androgen-independent growth of CRPC cells likely through the activation of protein kinase C isozymes, induces osteoblast differentiation *in vitro* through a direct cell-cell interaction, and is upregulated in human PC xenografts that cause an osteoblastic reaction when grown in bone. Contrasting data still exist about the real significance of AR reactivation in castration-resistant prostate cancer cells and its relevance for prostate cancer stem cell biology (Miki et al. [2007;](#page-11-18) Collins et al. [2005;](#page-10-6) Rajasekhar et al. [2011;](#page-12-11) Patrawala et al. [2006\)](#page-12-9).

– MicroRNAs (miRs) function as either oncogenes or tumor suppressor genes in cancer (Zhu et al. [2013\)](#page-14-2). Early reports suggest that in androgen-dependent prostate cancer cells, they may play a role in tumor development, progression, evolution to metastasis, response to therapy, and prognosis (Qu et al. [2013\)](#page-12-19) In prostate epithelial EP156T cells, miR-182 and miR-203 have been really shown to induce MET features and growth factor independent cell growth.

On the opposite side, elevated serum levels of miR-141 have been found related with the presence of bone prostate cancer metastasis, without significant correspondence with either Gleason score of primary tumor or PSA value. By converse, miR-141 showed a positive correlation with serum alkaline phosphatase levels (Zhang et al. [2013\)](#page-14-3).

However, more data are required before we reach a comprehensive knowledge about their definite roles in androgen-independent, bone metastasizing prostate cancer (Brennan et al. [2012\)](#page-10-5).

The better understanding of the molecular phenotype of PCSC and DPCC could provide novel therapeutic strategies, allowing the targeting of bone metastatic prostate cancer cells, before they exit dormancy and become lethal (Patel et al. [2011\)](#page-12-13).

Early profiling studies have evidenced the role of miRNA expression in prostate CSC (Liu et al. [2011\)](#page-11-19), revealing that they specifically target several stem cells markers in prostate cancer. As an example, the overexpression of miR-34A leads to the decrease of  $CD44+$  prostate cancer cells, inhibiting tumor development and metastasis, thus appearing as a promising potential new therapeutic tool for neutralize the killing potential of PSCS.

– Recently, it has been suggested that infiltrating immune cells facilitate tumor stem cell proliferation. Moreover, it has been proposed that aberrant immune cell infiltration preferentially associates with tumor capsular areas showing distinct degenerative alterations. Tumor-associated lymphocytes might cause focal disruption of prostate cancer capsule, favoring, then, tumor cell budding and metastasis (Jiang et al. [2013\)](#page-11-20).

This finding deserves further evaluation, as it may have a relevant impact on our knowledge of the prostate cancer metastasis causative events. It suggests, in fact, that the aberrant immune cell infiltration may have the same destructive impact of cancer cells on the lining capsule, offering in turn a selective proliferative advantage to prostate cancer stem cells proximal to these focal disruptions.

Moreover, it will be also clarified if the selective tumor-associated immunoreactive infiltrate may have a causative role even for the early onset of aggressive prostate cancer at young ages, typically originating in healthy men with morphologically normal prostate (Man and Gardner [2008\)](#page-11-21).

Overall, the understanding of the molecular background of prostate cancerogenesis has already changed our way to look at prostate cancer.

The growing flow of information concerning the bidirectional interactions between the epithelial cancer cells, tumor-associated stroma, and host bone microenvironment has become an impressively active area of prostate cancer research (Efstathiou and Logothetis [2010\)](#page-10-20). The stromal-interacting pathways represent exciting targets for new molecular niche-directed therapies, which in the next future will guide our efforts to fight metastatic prostate cancer.

## **References**

- <span id="page-9-1"></span>Aguirre-Ghiso JA (2007) Models, mechanisms and clinical evidence for cancer dormancy. Nat Rev Cancer 7(11):834–846
- <span id="page-9-2"></span>Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF (2003) Prospective identification of tumorigenic breast cancer cells. Proc Natl Acad Sci U S A 100:3983–3988
- <span id="page-9-0"></span>Allavena P, Sica A, Solinas G, Porta C, Mantovani A (2008) The inflammatory microenvironment in tumor progression: the role of tumor-associated macrophages. Crit Rev Oncol Hematol 66(1):1–9
- <span id="page-9-3"></span>Arai F, Yoshihara H, Hosokawa K, Nakamura Y, Gomei Y et al (2009) Niche regulation of hematopoietic stem cells in the endosteum. Ann N Y Acad Sci 1176:36–46
- <span id="page-9-4"></span>Augello MA, Burd CJ, Birbe R, McNair C, Ertel A, Magee MS, Frigo DE, Wilder-Romans K, Shilkrut M, Han S, Jernigan DL, Dean JL, Fatatis A, McDonnell DP, Visakorpi T, Feng FY, Knudsen KE (2013) Convergence of oncogenic and hormone receptor pathways promotes metastatic phenotypes. J Clin Invest 123(1):493–508
- <span id="page-10-18"></span>Bisson I, Prowse DM (2009) WNT signaling regulates self-renewal and differentiation of prostate cancer cells with stem cell characteristics. Cell Res 19:683–697
- <span id="page-10-5"></span>Brennan SC, Thiem U, Roth S, Aggarwal A, Fetahu IS, Tennakoon S, Gomes AR, Brandi ML, Bruggeman F, Mentaverri R, Riccardi D, Kallay E (2013) Calcium sensing receptor signalling in physiology and cancer. Biochim Biophys Acta 1833(7):1732–1744
- <span id="page-10-2"></span>Brett A, Pandey S, Fraizer G (2013) The Wilms' tumor gene (WT1) regulates E-cadherin expression and migration of prostate cancer cells. Mol Cancer 12:3
- <span id="page-10-17"></span>Bussard KM, Gay CV, Mastro AM (2008) The bone microenvironment in metastasis; what is special about bone? Cancer Metastasis Rev 27:41–55
- <span id="page-10-12"></span>Castellón EA, Valenzuela R, Lillo J, Castillo V, Contreras HR, Gallegos I, Mercado A, Huidobro C (2012) Molecular signature of cancer stem cells isolated from prostate carcinoma and expression of stem markers in different Gleason grades and metastasis. Biol Res 45(3):297–305
- <span id="page-10-19"></span>Chappard D, Bouvard B, Basle MF, Legrand E, Audran M (2011) Bone metastasis: histological ´ changes and pathophysiological mechanisms in osteolytic or osteosclerotic localizations. A review. Morphologie 95(309):65–75. Epub 2011 May 28
- <span id="page-10-15"></span>Cher ML, Towler DA, Rafii S et al (2006) Cancer interaction with the bone microenvironment: a workshop of the National Institutes of Health Tumor Microenvironment Study Section. Am J Pathol 168:1405–1412
- <span id="page-10-8"></span>Chu P, Clanton DJ, Snipas TS, Lee J, Mitchell E, Nguyen ML et al (2009) Characterization of a subpopulation of colon cancer cells with stem cell-like properties. Int J Cancer 124:1312–1321
- <span id="page-10-14"></span>Chung LW, Baseman A, Assikis V, Zhau HE (2005) Molecular insights into prostate cancer progression: the missing link of tumor microenvironment. J Urol 173:10–20
- <span id="page-10-6"></span>Collins AT, Berry PA, Hyde C, Stower MJ, Maitland NJ (2005) Prospective identification of tumorigenic prostate cancer stem cells. Cancer Res 65:10946–10951
- <span id="page-10-13"></span>Colombel M, Eaton CL, Hamdy F, Ricci E, van der Pluijm G, Cecchini M, Mege-Lechevallier F, Clezardin P, Thalmann G (2012) Increased expression of putative cancer stem cell markers in primary prostate cancer is associated with progression of bone metastases. Prostate 72(7): 713–720
- <span id="page-10-7"></span>Cui F, Wang J, Chen D, Chen YJ (2011) CD133 is a temporary marker of cancer stem cells in small cell lung cancer, but not in non-small cell lung cancer. Oncol Rep 25:701–708
- <span id="page-10-1"></span>Czyż J, Szpak K, Madeja Z  $(2012)$  The role of connexins in prostate cancer promotion and progression. Nat Rev Urol 9(5):274–282
- <span id="page-10-10"></span>Dalerba P, Dylla SJ, Park IK, Liu R, Wang X, Cho RW et al (2007) Phenotypic characterization of human colorectal cancer stem cells. Proc Natl Acad Sci U S A 104:10158–10163
- <span id="page-10-4"></span>Desiniotis A, Kyprianou N (2011) Significance of talin in cancer progression and metastasis. Int Rev Cell Mol Biol 289:117–147
- <span id="page-10-16"></span>Doan PL, Chute JP (2012) The vascular niche: home for normal and malignant hematopoietic stem cells. Leukemia 26(1):54–62
- <span id="page-10-11"></span>Draper JS, Pigott C, Thomson JA, Andrews PW (2002) Surface antigens of human embryonic stem cells: changes upon differentiation in culture. J Anat 200:249–258
- <span id="page-10-0"></span>Du WW, Yang W, Yee AJ (2013) Roles of versican in cancer biology – tumorigenesis, progression and metastasis. Histol Histopathol 28(6):701–713
- <span id="page-10-21"></span>Eastham JA (2007) Bone health in men receiving androgen deprivation therapy for prostate cancer. J Urol 177(1):17–24
- <span id="page-10-9"></span>Eaton CL, Colombel M, van der Pluijm G, Cecchini M, Wetterwald A, Lippitt J et al (2010) Evaluation of the frequency of putative prostate cancer stem cells in primary and metastatic prostate cancer. Prostate 70:875–882
- <span id="page-10-20"></span>Efstathiou E, Logothetis CJ (2010) A new therapy paradigm for prostate cancer founded on clinical observations. Clin Cancer Res 16:1100–1107
- <span id="page-10-22"></span>Eriksson S, Killander J, Wadman B (1972) Leuco-erythroblastic anaemia in prostatic cancer. Report of two cases with complete haematological remission. Scand J Haematol 9(6):648–653
- <span id="page-10-3"></span>Fan L, Wang H, Xia X, Rao Y, Ma X, Ma D, Wu P, Chen G (2012) Loss of E-cadherin promotes prostate cancer metastasis via upregulation of metastasis-associated gene 1 expression. Oncol Lett 4(6):1225–1233. Epub 2012 Sep 21
- <span id="page-11-7"></span>Goldstein AS, Huang J, Guo C, Garraway IP, Witte ON (2010) Identification of a cell of origin for human prostate cancer. Science 329:568–571
- <span id="page-11-2"></span>Hu YY, Zheng MH, Zhang R, Liang YM, Han H (2012) Notch signaling pathway and cancer metastasis. Adv Exp Med Biol 727:186–198
- <span id="page-11-3"></span>Igney FH, Krammer PH (2002) Death and anti-death: tumour resistance to apoptosis. Nat Rev Cancer 2(4):277–288. A comprehensive review of cell death pathways and survival mechanisms exploited by cancer
- <span id="page-11-14"></span>Jacobs SC (1983) Spread of prostatic cancer to bone. Urology 21(4):337–344
- Jemal A, Siegel R, Ward E (2010) Cancer statistics. CA Cancer J Clin 60:277–300
- <span id="page-11-20"></span>Jiang B, Mason J, Jewett A, Liu ML, Chen W, Qian J, Ding Y, Ding S, Ni M, Zhang X, Man YG (2013) Tumor-infiltrating immune cells: triggers for tumor capsule disruption and tumor progression? Int J Med Sci 10(5):475–497
- <span id="page-11-17"></span>Jin HJ, Zhao JC, Ogden I, Bergan RC, Yu J (2013) Androgen receptor-independent function of FoxA1 in prostate cancer metastasis. Cancer Res 73(12):3725–3736
- <span id="page-11-5"></span>Joyce JA, Pollard JW (2009) Microenvironmental regulation of metastasis. Nat Rev Cancer 9(4):239–252
- <span id="page-11-12"></span>Jung Y, Wang J, Song J, Shiozawa Y, Havens A et al (2007) Annexin II expressed by osteoblasts and endothelial cells regulates stem cell adhesion, homing, and engraftment following transplantation. Blood 110(1):82–90
- <span id="page-11-10"></span>Karlou M, Tzelepi V, Efstathiou E (2010) Therapeutic targeting of the prostate cancer microenvironment. Nat Rev Urol 7:494–509
- <span id="page-11-15"></span>Keller ET, Zhang J, Cooper CR, Smith PC, McCauley LK et al (2001) Prostate carcinoma skeletal metastases: cross-talk between tumor and bone. Cancer Metastasis Rev 20(3–4):333–349
- <span id="page-11-11"></span>Kiel MJ, Morrison SJ (2008) Uncertainty in the niches that maintain haematopoietic stem cells. Nat Rev Immunol 8(4):290–301
- <span id="page-11-8"></span>Lawson DA, Zong Y, Memarzadeh S, Xin L, Huang J, Witte ON (2010) Basal epithelial stem cells are efficient targets for prostate cancer initiation. Proc Natl Acad Sci U S A 107:2610–2615
- <span id="page-11-1"></span>Lazari P, Poulias H, Gakiopoulou H, Thomopoulou GH, Barbatis C, Lazaris AC (2013) Differential immunohistochemical expression of CD44s, E-cadherin and  $\beta$ -catenin among hyperplastic and neoplastic lesions of the prostate gland. Urol Int 90(1):109–116. Epub 2012 Dec 5 Leukemia. 2011
- <span id="page-11-6"></span>Li T, Su Y, Mei Y, Leng Q, Leng B, Liu Z et al (2010) ALDH1A1 is a marker for malignant prostate stem cells and predictor of prostate cancer patients' outcome. Lab Invest 90:234–244
- <span id="page-11-19"></span>Liu C, Kelnar K, Liu B, Chen X, Calhoun-Davis T, Li H et al (2011) The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. Nat Med 17:211–215
- <span id="page-11-16"></span>Loberg RD, Logothetis CJ, Keller ET, Pienta KJ (2005) Pathogenesis and treatment of prostate cancer bone metastases: targeting the lethal phenotype. J Clin Oncol 23:8232–8241
- <span id="page-11-13"></span>Mai J, Waisman DM, Sloane BF (2000) Cell surface complex of cathepsin B/annexin II tetramer in malignant progression. Biochim Biophys Acta 1477(1–2):215–230
- <span id="page-11-21"></span>Man YG, Gardner WA (2008) Bad seeds produce bad crops: a single stage-process of prostate tumor invasion. Int J Biol Sci 4(4):246–258
- <span id="page-11-18"></span>Miki J, Furusato B, Li H, Gu Y, Takahashi H, Egawa S et al (2007) Identification of putative stem cell markers, CD133 and CXCR4, in hTERT-immortalized primary nonmalignant and malignant tumor-derived human prostate epithelial cell lines and in prostate cancer specimens. Cancer Res 67:3153–3161
- <span id="page-11-4"></span>Morgan TM, Lange PH, Porter MP, Lin DW, Ellis WJ et al (2009) Disseminated tumor cells in prostate cancer patients after radical prostatectomy and without evidence of disease predicts biochemical recurrence. Clin Cancer Res 15(2):677–683
- <span id="page-11-9"></span>Morrissey C, Vessella RL (2007) The role of tumor microenvironment in prostate cancer bone metastasis. J Cell Biochem 101:873–886
- <span id="page-11-0"></span>Nguyen DX, Massague J (2007) Genetic determinants of cancer metastasis. Nat Rev Genet 8(5):341–352
- <span id="page-12-18"></span>Norgaard M, Jensen AO, Jacobsen JB, Cetin K, Fryzek JP, Sorensen HT (2010) Skeletal related events, bone metastasis and survival of prostate cancer: a population based cohort study in Denmark (1999 to 2007). J Urol 184(1):162–167
- <span id="page-12-4"></span>Osanto S, Van Poppel H (2012) Emerging novel therapies for advanced prostate cancer. Ther Adv Urol 4(1):3–12
- <span id="page-12-2"></span>Pal M, Koul S, Koul HK (2013) The transcription factor sterile alpha motif (SAM) pointed domaincontaining ETS transcription factor (SPDEF) is required for E-cadherin expression in prostate cancer cells. J Biol Chem 288(17):12222–12231
- <span id="page-12-13"></span>Patel LR, Camacho DF, Shiozawa Y, Pienta KJ, Taichman RS (2011) Mechanisms of cancer cell metastasis to the bone: a multistep process. Future Oncol 7(11):1285–1297
- <span id="page-12-9"></span>Patrawala L, Calhoun T, Schneider-Broussard R, Li H, Bhatia B, Tang S et al (2006) Highly purified CD44+ prostate cancer cells from xenograft human tumors are enriched in tumorigenic and metastatic progenitor cells. Oncogene 25:1696–1708
- <span id="page-12-7"></span>Patrawala L, Calhoun-Davis T, Schneider-Broussard R, Tang DG (2007) Hierarchical organization of prostate cancer cells in xenograft tumors: the  $CD44+alpha2beta+$  cell population is enriched in tumor-initiating cells. Cancer Res 67:6796–6805
- <span id="page-12-5"></span>Pfitzenmaier J, Ellis WJ, Hawley S, Arfman EW, Klein JR et al (2007) The detection and isolation of viable prostate-specific antigen positive epithelial cells by enrichment: a comparison to standard prostate-specific antigen reverse transcriptase polymerase chain reaction and its clinical relevance in prostate cancer. Urol Oncol 25(3):214–220
- <span id="page-12-19"></span>Qu Y, Li WC, Hellem MR, Rostad K, Popa M, McCormack E, Oyan AM, Kalland KH, Ke XS (2013) MiR-182 and miR-203 induce mesenchymal to epithelial transition and self-sufficiency of growth signals via repressing SNAI2 in prostate cells. Int J Cancer 133(3):544–555
- <span id="page-12-11"></span>Rajasekhar VK, Studer L, Gerald W, Socci ND, Scher HI (2011) Tumour-initiating stem-like cells in human prostate cancer exhibit increased NF-kappaB signalling. Nat Commun 2:162
- <span id="page-12-3"></span>Roca H, Varsos ZS, Mizutani K, Pienta KJ (2008) CCL2, survivin and autophagy: new links with implications in human cancer. Autophagy 4(7):969–971
- <span id="page-12-0"></span>Rodenhiser DI (2009) Epigenetic contributions to cancer metastasis. Clin Exp Metastasis 26(1):5–18
- <span id="page-12-1"></span>Roodman GD (2004) Mechanisms of bone metastasis. N Engl J Med 350(16):1655–1664
- <span id="page-12-12"></span>Ross JS, Kallakury BV, Sheehan CE, Fisher HA, Kaufman RP Jr, Kaur P et al (2004) Expression of nuclear factor-kappa B and I kappa B alpha proteins in prostatic adenocarcinomas: correlation of nuclear factor-kappa B immunoreactivity with disease recurrence. Clin Cancer Res 10:2466–2472
- <span id="page-12-14"></span>Roudier MP, True LD, Higano CS et al (2003) Phenotypic heterogeneity of end-stage prostate carcinoma metastatic to bone. Hum Pathol 34:646–653
- <span id="page-12-15"></span>Roudier MP, Morrissey C, True LD, Higano CS, Vessella RL et al (2008) Histopathological assessment of prostate cancer bone osteoblastic metastases. J Urol 180(3):1154–1160
- <span id="page-12-16"></span>Shamdas GJ, Ahmann FR, Matzner MB, Ritchie JM (1993) Leukoerythroblastic anemia in metastatic prostate cancer. Clinical and prognostic significance in patients with hormonerefractory disease. Cancer 71(11):3594–3600
- <span id="page-12-8"></span>Shi MF, Jiao J, Lu WG, Ye F, Ma D, Dong QG et al (2010) Identification of cancer stem cell-like cells from human epithelial ovarian carcinoma cell line. Cell Mol Life Sci 67:3915–3925
- <span id="page-12-10"></span>Shiozawa Y, Havens AM, Jung Y, Ziegler AM, Pedersen EA et al (2008a) Annexin II/annexin II receptor axis regulates adhesion, migration, homing, and growth of prostate cancer. J Cell Biochem 105(2):370–380
- <span id="page-12-6"></span>Shiozawa Y, Havens AM, Pienta KJ, Taichman RS (2008b) The bone marrow niche: habitat to hematopoietic and mesenchymal stem cells, and unwitting host to molecular parasites. Leukemia 22(5):941–950
- <span id="page-12-17"></span>Shiozawa Y, Pedersen EA, Havens AM, Jung Y, Mishra A et al (2011) Human prostate cancer metastases target the hematopoietic stem cell niche to establish footholds in mouse bone marrow. J Clin Invest 121(4):1298–1312
- <span id="page-13-16"></span>Sottnik JL, Daignault-Newton S, Zhang X, Morrissey C, Hussain MH, Keller ET, Hall CL (2013) Integrin alpha2beta 1 ( $\alpha$  2 $\beta$  1) promotes prostate cancer skeletal metastasis. Clin Exp Metastasis 30(5):569–578
- <span id="page-13-19"></span>Spivak JL (1994) Cancer-related anemia: its causes and characteristics. Semin Oncol 21(2 Suppl 3):3–8
- <span id="page-13-18"></span>Sturge J, Caley MP, Waxman J (2011) Bone metastasis in prostate cancer: emerging therapeutic strategies. Nat Rev Clin Oncol 8(6):357–368. doi[:10.1038/nrclinonc.2011.67.](http://dx.doi.org/10.1038/nrclinonc.2011.67) Epub 2011 May 10
- <span id="page-13-14"></span>Sun YX, Wang J, Shelburne CE, Lopatin DE, Chinnaiyan AM et al (2003) Expression of CXCR4 and CXCL12 (SDF-1) in human prostate cancers (PCa) *in vivo*. J Cell Biochem 89(3):462–473
- <span id="page-13-12"></span>Sun YX, Schneider A, Jung Y, Wang J, Dai J et al (2005) Skeletal localization and neutralization of the SDF-1(CXCL12)/CXCR4 axis blocks prostate cancer metastasis and growth in osseous sites *in vivo*. J Bone Miner Res 20(2):318–329
- <span id="page-13-13"></span>Sun YX, Fang M, Wang J, Cooper CR, Pienta KJ et al (2007) Expression and activation of alpha v beta 3 integrins by SDF-1/CXC12 increases the aggressiveness of prostate cancer cells. Prostate 67(1):61–73
- <span id="page-13-4"></span>Taichman RS (2005) Blood and bone: two tissues whose fates are intertwined to create the hematopoietic stem-cell niche. Blood 105(7):2631–2639
- <span id="page-13-7"></span>Tang DG, Patrawala L, Calhoun T, Bhatia B, Choy G, Schneider-Broussard R et al (2007) Prostate cancer stem/progenitor cells: identification, characterization, and implications. Mol Carcinog 46:1–14
- Tannock IF, de Wit R, Berry WR, Horti J, Pluzanska A, Chi KN, Oudard S, Theodore C, James ´ ND, Turesson I, Rosenthal MA, Eisenbergee MA (2004) TAX 327 investigators. Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer. N Engl J Med 351(15):1502–1512
- <span id="page-13-5"></span>Townson JL, Chambers AF (2006) Dormancy of solitary metastatic cells. Cell Cycle 5(16): 1744–1750
- <span id="page-13-9"></span>Tsujimura A, Koikawa Y, Salm S, Takao T, Coetzee S, Moscatelli D et al (2002) Proximal location of mouse prostate epithelial stem cells: a model of prostatic homeostasis. J Cell Biol 157:1257–1265
- <span id="page-13-8"></span>van den Hoogen C, van der Horst G, Cheung H, Buijs JT, Lippitt JM, Guzman-Ramirez N et al (2010) High aldehyde dehydrogenase activity identifies tumor-initiating and metastasisinitiating cells in human prostate cancer. Cancer Res 70:5163–5173
- Vashisht S, Bagler G (2012) An approach for the identification of targets specific to bone metastasis using cancer genes interactome and gene ontology analysis. PLoS One 7(11):e49401
- <span id="page-13-15"></span>Wang J, Shiozawa Y, Wang Y, Jung Y, Pienta KJ et al (2008) The role of CXCR7/RDC1 as a chemokine receptor for CXCL12/SDF-1 in prostate cancer. J Biol Chem 283(7):4283–4894
- <span id="page-13-10"></span>Wang X, Kruithof-de Julio M, Economides KD, Walker D, Yu H, Halili MV et al (2009) A luminal epithelial stem cell that is a cell of origin for prostate cancer. Nature 461:495–500
- <span id="page-13-3"></span>Wang H, Fan L, Wei J, Weng Y, Zhou L, Shi Y, Zhou W, Ma D, Wang C (2012) Akt mediates metastasis-associated gene 1 (MTA1) regulating the expression of E-cadherin and promoting the invasiveness of prostate cancer cells. PLoS One 7(12):e46888. Epub 2012 Dec 5
- <span id="page-13-0"></span>Wang X, Lee SO, Xia S, Jiang Q, Luo J, Li L, Yeh S, Chang C (2013) Endothelial cells enhance prostate cancer metastasis via IL-6->Androgen Receptor->TGF- $\beta$ ->MMP-9 signals. Mol Cancer Ther 12(6):1026–1037
- <span id="page-13-2"></span>Yates C (2011) Prostate tumor cell plasticity: a consequence of the microenvironment. Adv Exp Med Biol 720:81–90
- <span id="page-13-1"></span>Yilmaz M, Christofori G (2009) EMT, the cytoskeleton, and cancer cell invasion. Cancer Metastasis Rev 28(1–2):15–33
- <span id="page-13-11"></span>Yin T, Li L (2006) The stem cell niches in bone. J Clin Invest 116(5):1195–1201
- <span id="page-13-6"></span>Yu C, Yao Z, Jiang Y, Keller ET (2012) Prostate cancer stem cell biology. Minerva Urol Nefrol 64(1):19–33
- <span id="page-13-17"></span>Zetter BR (1990) The cellular basis of site-specific tumor metastasis. N Engl J Med 322(9): 605–612
- <span id="page-14-3"></span>Zhang HL, Qin XJ, Cao DL, Zhu Y, Yao XD, Zhang SL, Dai B, Ye DW (2013) An elevated serum miR-141 level in patients with bone-metastatic prostate cancer is correlated with more bone lesions. Asian J Androl 15(2):231–235. Epub 2013 Feb 4
- <span id="page-14-0"></span>Zhao J, Wu XY, Ling XH, Lin ZY, Fu X, Deng YH, He HC, Zhong W (2013) Analysis of genetic aberrations on chromosomal region 8q21-24 identifies E2F5 as an oncogene with copy number gain in prostate cancer. Med Oncol 30(1):465
- <span id="page-14-1"></span>Zheng D, Decker KF, Zhou T, Chen J, Qi Z, Jacobs K, Weilbaecher KN, Corey E, Long F, Jia L (2013) Role of WNT7B-induced non-canonical pathway in advanced prostate cancer. Mol Cancer Res 11(5):482–493
- <span id="page-14-2"></span>Zhu KC, Lu JJ, Xu XL, Sun JM (2013) MicroRNAs in androgen-dependent PCa. Front Biosci 18:748–755