

# Residue-Derived Amino Sugar Formation and Its Carbon Use Efficiency

Zhen Bai, Samuel Bodé, Pascal Boeckx, and Xudong Zhang

**Abstract** A 21-day lab incubation experiment was carried out with arable soil and uniform  $^{13}\text{C}$ -labeled wheat residues. Residue-derived amino sugars ( $\text{AS}_\text{R}$ ) accumulated exponentially and were fitted with a first-order model  $C_t = C_{\text{Max}}(1 - e^{-kt})$  well. Their maxima were 88–381  $\text{nmol g}^{-1}$  for glucosamine ( $\text{GluN}_\text{R}$ ), 52–105  $\text{nmol g}^{-1}$  for galactosamine ( $\text{GalN}_\text{R}$ ), and 107–137  $\text{nmol g}^{-1}$  for muramic acid ( $\text{MurN}_\text{R}$ ), respectively. Their half-lives ( $T_{1/2}$ ) separately varied between 1.2 and 3.3 days for  $\text{GluN}_\text{R}$ , 1.6 and 4.7 days for  $\text{GalN}_\text{R}$ , and less than 0.3 day for  $\text{MurN}_\text{R}$ . A close relationship was observed between carbon mineralization and  $\text{AS}_\text{R}$  ( $P < 0.05$ ), and carbon use efficiency of cumulative  $\text{AS}_\text{R}$  was calculated and compared.

**Keywords** Amino sugar • Carbon use efficiency • Organic residue • Carbon-13 • LC-c-IRMS

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## Introduction

Though amino sugars (AS) have widely been used to investigate microbial residues and correlated organic compounds sequestration, little is known about their formation dynamics in soil (Zhang and Amelung 1996; He et al. 2011). Difficulties are probably ascribed to the definition and differentiation of varied ASs pools, substrate availability, and time scale of the study. With  $^{13}\text{C}$ -labeled residues' incubation, LC-c-IRMS was utilized to investigate AS formation kinetics and its carbon use efficiency for the first time in this study.

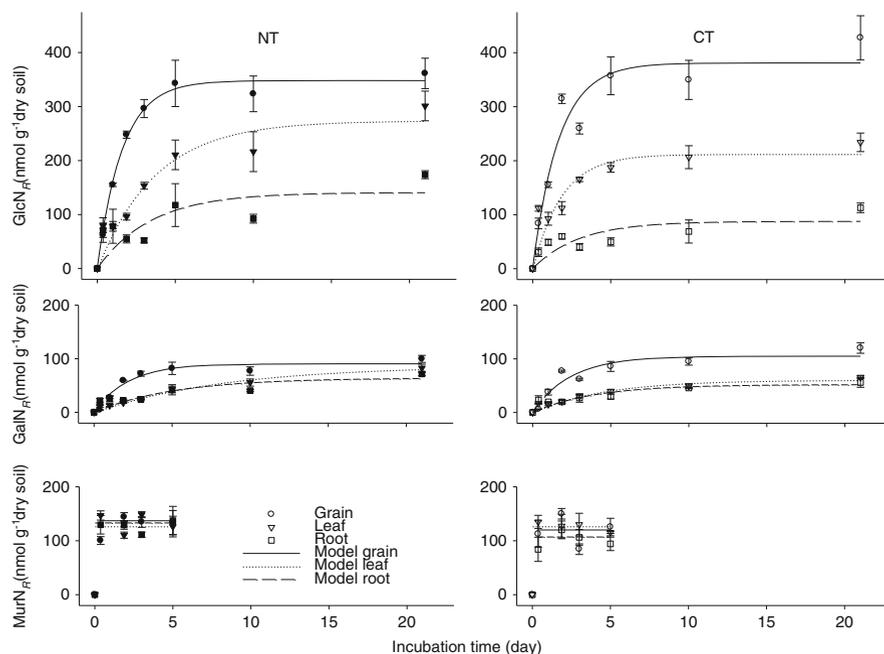
## Materials and Methods

0–10-cm top soil from conventional till (CT) and no till (NT) was amended with  $^{13}\text{C}$ -labeled wheat grain (G), leaf (L), and root (R) at 6-mg substrate  $\text{Cg}^{-1}$  for a 21-day incubation, including six treatments: **NG** (NT + G), **NL** (NT + L), **NR** (NT + R), **CG** (CT + G), **CL** (CT + L), and **CR** (CT + R). Concentrations and  $\delta^{13}\text{C}$  of AS were determined by LC-c-IRMS. Residue-derived AS ( $\text{AS}_R$ ) formation was described in  $\text{AS}_{R,t} = \text{AS}_{R,\text{Max}} \cdot (1 - e^{-kt})$ , where  $\text{AS}_{R,\text{Max}}$  is potentially accumulative  $\text{AS}_{R,t}$ , and  $k$  is  $\text{AS}_{R,t}$  the formation rate constant, and  $t$  is the incubation time.

## Results and Discussion

This study showed that  $T_{1/2}$  of  $\text{AS}_R$  formation increased as  $\text{MurN} (\leq 0.3 \text{ day}) < \text{GluN} (1.2\text{--}3.3 \text{ days}) < \text{GalN} (1.6\text{--}4.7 \text{ days})$ ; their maxima were 88–381  $\text{nmol g}^{-1}$  for  $\text{GluN}$ , 52–105  $\text{nmol g}^{-1}$  for  $\text{GalN}$ , and 107–137  $\text{nmol g}^{-1}$  for  $\text{MurN}$ ; and  $\text{AS}_{R,\text{Max}}/\text{AS}_{\text{SOM},0h}$  were separately 14.9–17.8% for  $\text{MurN}$ , 5.5–13.0% for  $\text{GluN}$ , and 2.7–6.3% for  $\text{GalN}$  ( $\text{AS}_{\text{SOM}}$  data not shown) (Fig. 1, Table 1), which agrees with previous studies (Glaser and Gross 2005).  $\text{MurN}$  is exclusively originated from bacteria, and its faster formation suggested the important role of bacteria at the early stage of mineralization of wheat residues.

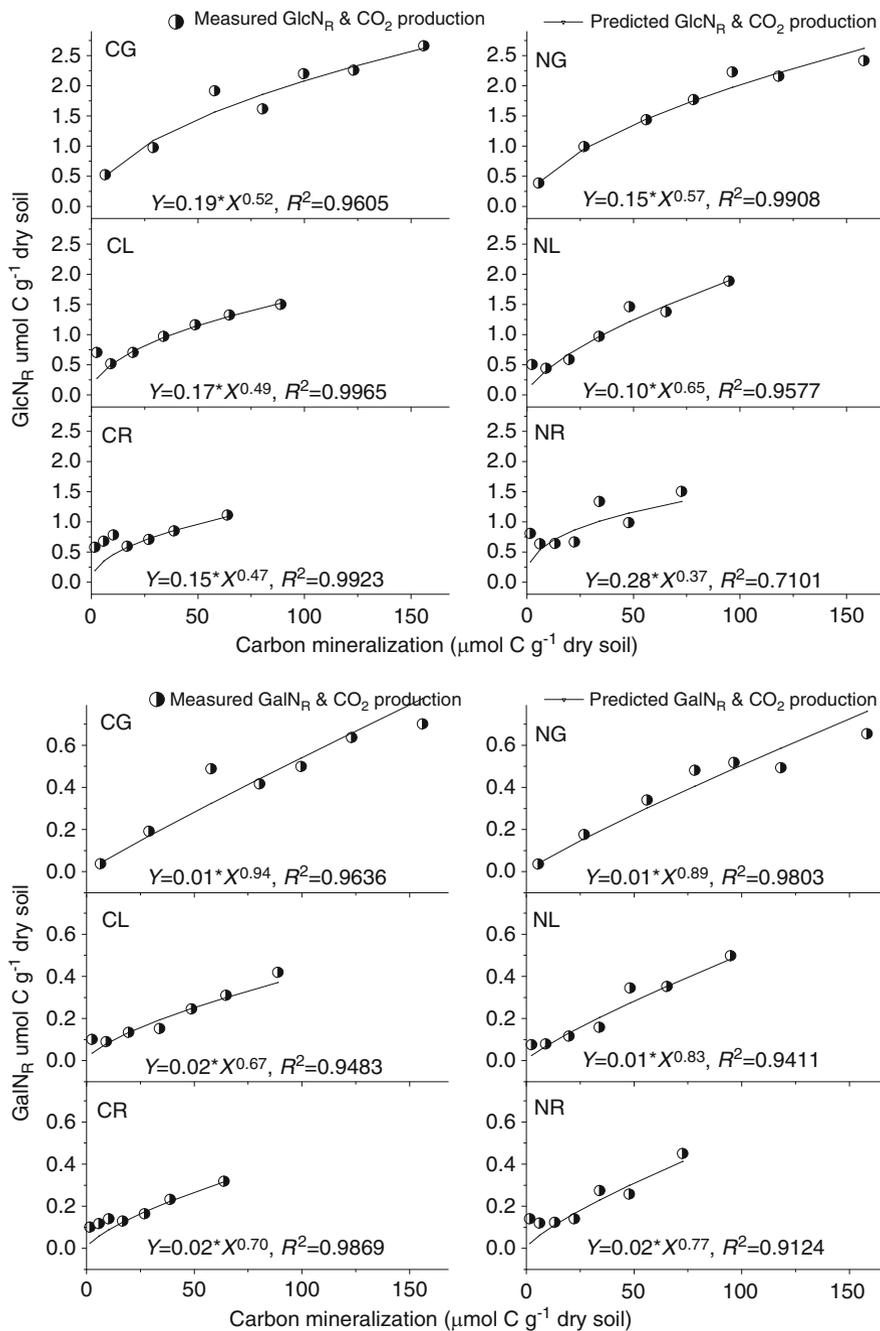
The cumulative  $\text{GluN}_R$  or  $\text{GalN}_R$  was significantly correlated to the mineralized carbon ( $C_M$ ) across all treatments throughout the whole incubation, which strongly suggests that a certain proportion of  $C_M$  is allocated into newly formed  $\text{AS}_R$  (except for  $\text{MurN}_R$ , Fig. 2). The intercepts and slopes of linear functions between  $\text{GluN}_R$  and  $C_M$  were evidently different from their counterparts between  $\text{GalN}_R$  and  $C_M$ , which clearly signifies a much higher carbon use efficiency of accumulative  $\text{GluN}_R$  than  $\text{GalN}_R$ . The stoichiometric relationship between  $C_M$  and  $\text{AS}_R$  could be calculated: an increase of 0.1  $\mu\text{mol GluN}_R$  needed 0.38  $\mu\text{mol } C_M$ , which was 13.61 lower than  $\text{GalN}_R$ , while after 1  $\mu\text{mol } C_M$ , 0.16  $\mu\text{mol GluN}_R$  would be synthesized, which was nearly 16 times the  $\text{GalN}_R$ .



**Fig. 1** Modeled and measured residue-derived amino sugar concentration ( $AS_{R,t}$ ) as a function of time

**Table 1** Parameters description of  $AS_R$  formation

$AS_R$	Treats	$AS_{R,max}$ (nmol g <sup>-1</sup> )	$k$ (day <sup>-1</sup> )	$T_{1/2}$ (day)	$R^2$
GluN <sub>R</sub>	NG	348 ± 13a	0.5 ± 0.0a	1.3 ± 0.1c	≥0.967
	NL	274 ± 20b	0.3 ± 0.0b	2.4 ± 0.4b	≥0.852
	NR	141 ± 22bc	0.6 ± 0.0a	1.3 ± 0.1c	≥0.474
	CG	381 ± 19a	0.6 ± 0.0a	1.2 ± 0.1c	≥0.900
	CL	212 ± 13bc	0.5 ± 0.0a	1.5 ± 0.1c	≥0.820
	CR	88 ± 11c	0.2 ± 0.0c	3.3 ± 0.4a	≥0.849
GalN <sub>R</sub>	NG	90 ± 5a	0.4 ± 0.0a	1.6 ± 0.1c	≥0.917
	NL	87 ± 7b	0.2 ± 0.0c	4.3 ± 0.0a	≥0.922
	NR	64 ± 7b	0.2 ± 0.0c	4.7 ± 1.2a	≥0.734
	CG	105 ± 6a	0.4 ± 0.0a	1.7 ± 0.0c	≥0.903
	CL	61 ± 3b	0.2 ± 0.0c	4.0 ± 0.5a	≥0.924
	CR	52 ± 6c	0.3 ± 0.0b	2.7 ± 0.1b	≥0.670
MurN <sub>R</sub>	NG	137 ± 6a	2.7 ± 0.7	0.3 ± 0.1	≥0.875
	NL	133 ± 40a	–	–	–
	NR	127 ± 45a	–	–	–
	CG	120 ± 11a	–	–	–
	CL	127 ± 43a	–	–	–
	CR	107 ± 8a	–	–	–



**Fig. 2** Correlation between C mineralization and AS<sub>R</sub>

## References

- Glaser, B., and S. Gross. 2005. Compound-specific  $\delta^{13}\text{C}$  analysis of individual amino sugars—a tool to quantify timing and amount of soil microbial residue stabilization. *Rapid Communications in Mass Spectrometry* 19: 1409–1416.
- He, H.B., X.B. Li, W. Zhang, and X.D. Zhang. 2011. Differentiating the dynamics of native and newly immobilized amino sugars in soil frequently amended with inorganic nitrogen and glucose. *European Journal of Soil Science* 62: 144–151.
- Zhang, X.D., and W. Amelung. 1996. Gas chromatographic determination of muramic acid, glucosamine, mannosamine, and galactosamine in soils. *Soil Biology and Biochemistry* 28: 1201–1206.