# **Phycoremediation: Future Perspective of Green Technology**

Sonal Dixit and D.P. Singh

# **1 Introduction**

 The unrestricted developmental activities such as rapid industrialization and urbanization carried out during the past few decades have given rise to serious problems of environmental contamination. The load of pollutants including toxic metals is ever rising in the environment as a result of enhanced industrial activities. These heavy metals enter the environment through a variety of human activities, such as mining, refining, electroplating industries, etc. (Micheletti et al. 2007). Thus, the deposition of toxic metals in the environment and their speciation between abiotic and biotic components of ecosystem is posing toxicity in the latter group as it is impossible to degrade these pollutants by any means; the only way to overcome the effects of toxic heavy metals is their physical removal from the contaminated sites (Volesky [1997](#page-12-0)). The accumulation of toxic heavy metals in the aquatic environment has become a significant problem worldwide, and, therefore, it is a matter of great concern (Khoshmanesh et al. 1996; Dönmez et al. 1999; Gupta et al. [2006](#page-9-0)). At present, heavy metals are one of the most widespread pollutants, and their continuous accumulation in water bodies, soil and water sediments constitutes a serious hazard to both the environment and human health (Wase and Forster 1997). To avoid the adverse effects of metalcontaminated wastewater, it is necessary to treat them prior to their discharge into the environment.

 The conventional physicochemical techniques for the removal of inorganic pollutants from wastewater involve lime precipitation, chemical oxidation or reduction, ion exchange, electrochemical treatment, filtration, reverse osmosis, membrane technologies and evaporative recovery

 Department of Environmental Science , Babasaheb Bhimrao Ambedkar University, Vidhya Vihar, Raebareli Road, Lucknow, UP, India e-mail: [dpsingh\\_lko@yahoo.com](mailto:dpsingh_lko@yahoo.com)

(Barakat  $2011$ ). However, these techniques have significant shortcomings, for instance, low efficiency at lower concentrations of individual metal pollutants, high capital investment and operational costs and production of toxic sludge (Khoshmanesh et al.  $1996$ ). Therefore, it is imperative to have new technologies for reduced contamination of environmental components, which are not only environment friendly but also cost-effective.

 The removal of toxic inorganic environmental pollutants has received an ever-increasing attention in recent years, and various biomaterials such as bacteria, fungi, algae and plants have been employed as bioremedial agents to decontaminate the metal-polluted environment (Kotrba and Ruml [2000](#page-10-0); Kiran et al. 2008). This chapter mainly emphasizes the removal of certain deadly inorganic heavy metals by using algal strains and their phycoremediation potential.

# **2 Pollution in the Aquatic Environment**

 There are several ways in which we can classify contaminants of a water body. Broadly there are two classes of pollutants: organic pollutants and inorganic pollutants (Fig. [2.1 \)](#page-1-0).

# **2.1 Organic Pollutants**

 These are the compounds which consist of mainly carbon and hydrogen. The toxicity of organic pollutants depends upon the functional groups present in them. There are several subgroups of organic pollutants, as follows.

#### **2.1.1 Hydrocarbons**

 They can be divided into two classes: aliphatic hydrocarbons (alkanes, alkenes and alkynes) and aromatic hydrocarbons which contain carbon ring. Aromatic hydrocarbons such as poly aromatic hydrocarbons (PAHs) are much more reactive than any other class of aliphatic hydrocarbons.

S. Dixit • D.P. Singh  $(\boxtimes)$ 

<span id="page-1-0"></span>

**Fig. 2.1** Classification of the water pollutants

# **2.1.2 Polychlorinated Biphenyls (PCBs)**

These are stable and unreactive fluids, relatively insoluble in water. They are mostly used as hydraulic fluids, coolants/ insulation fluids and plasticizers in paints.

#### **2.1.3 Insecticides**

 Some of the insecticides are found to be highly dangerous for living tissues as they accumulate in fat tissues and enter in the food chain. Examples are DDT, lindane, carbamate, azadirachtin, etc.

#### **2.1.4 Detergents**

These are classified as phosphate detergents and surfactants. Phosphate detergents are used to soften the hard water and they have caustic property. Surfactants are used to enhance the foaming and emulsifying properties of detergents and are very toxic.

# **2.2 Inorganic Pollutants**

 Mostly inorganic pollutants include highly toxic metals (lead, cadmium, zinc, mercury, etc.), while some of these are nonmetallic inorganic substances such as nitrates and phosphates but still dangerous for the quality deterioration of aquatic system and partly toxic for the living system. An excessive input of nitrates and phosphates in aquatic bodies received through inorganic fertilizers is the major cause of algal blooms in surface waters, leading to eutrophication of water bodies.

#### **2.2.1 Heavy Metals**

The heavy metals, commonly defined as metals having a specific density of more than 5 g/cm<sup>-3</sup> (Hawkes [1997](#page-10-0)),

include Fe, Mn, Cu, Mo, Zn and Co, which are required in traces as nutrients by the living organisms, but they become toxic at higher concentrations. The other group of metals like Cd, Hg and Pb exert their potential toxic effects even at extremely lower concentrations. These metals are found in surface water bodies in their stable ionic forms. They mostly interfere with electron transfer reactions, and their interaction with oxygen often leads to the formation of toxic oxyradicals. Metalloids can bind with organic compounds, leading to the formation of lipophilic substances which are highly toxic, and can be stored in the fat tissues of animals including humans. Since the heavy metals cannot be broken down into less harmful components as they are non-biodegradable, they can only be remediated to reduce their toxic effects.

#### **2.2.2 Radioactive Isotopes**

 Radioactive isotopes are either present in nature or created by the humans in the nuclear industry. The decay of radioactive isotopes and their half-life determines the potential danger of these elements to humans. Different kinds of radiations can cause damage to the living tissues, depending upon the type of radiation and its energy level.

# **3 Bioremediaton**

 Bioremediation is a biological process used to clean up the hazardous chemicals present in the environment (Gianfreda and Rao 2004). It has several obvious advantages over physicochemical remediation methods in terms of costeffectiveness, convenience, complete removal of organic

pollutants and lack of collateral destruction of the site materials or its impact on indigenous flora and fauna (Timmis and Pieper 1999). With the advances in the field of biotechnology, bioremediation has become one of the major developing fields applied for environmental restoration. The bioremediation technique involves the use of microorganisms to reduce the concentration and toxicity of various chemical pollutants such as heavy metals, dyes, pesticides, etc. A considerable effort is devoted for developing a low-cost environmental friendly bioremediation technology that can effectively immobilize the dissolved toxic metals; a variety of living biomass has been tested (Say et al. [2001](#page-11-0); Adhiya et al. [2002](#page-8-0); Sheng et al. [2004](#page-11-0)) for the removal and/or recovery of metals for their probable reuse potential. Because of high metaladsorbing capacity, low cost and widespread abundance, the algal biomass has attracted the attention of scientists all over the world (Davis et al. [2003](#page-9-0)). Various types of either living or dead microalgal biomass have been employed to absorb the dissolved toxic metals.

# **3.1 Phycoremediation**

 Phycoremediation is a part of bioremediation where macroalgae or microalgae are being used for the removal or biotransformation of pollutants, including nutrients, xenobiotics and  $CO<sub>2</sub>$ . It simply offers cleanup technology, which is costeffective, nonintrusive and safe.

#### **3.1.1 Algae**

 Algae represent a large group of aquatic, most primitive photoautotrophic organisms that include around 30,000 species, ranging from unicellular (microalgae) to more complex multicellular organisms (macroalgae). Cyanobacteria (blue- green algae) were also included under the microalgae by some authors (Priyadarshani et al. [2011](#page-11-0) ). Algae possess chlorophyll and are able to transform light energy into chemical energy in a similar way to higher plants but lack true roots, stems and leaves. They grow comparatively faster, which results in fixation of  $CO<sub>2</sub>$  being 10–50 times faster than in plants (Subashchandrabose et al. [2013 \)](#page-11-0). When compared to plants, microalgae have a simple cell structure, and they are also often surrounded by fluid allowing easier uptake of water and nutrients (Chacoón-Lee and González-Mariño 2010). Algae are taxonomically divided based on their pigments, storage compounds and the main compounds present in their cell wall. The major classes are Chlorophyta (green algae), Rhodophyta (red algae), Phaeophyta (brown algae), Euglenophyta, Pyrrophyta, Chrysophyta and Cyanophyta (blue-green algae).

#### **Advantages of Using Algae**

• The blue-green alga (cyanobacteria) uses light energy source and  $CO<sub>2</sub>$  for its growth and survival. This way it helps in carbon sequestration and mitigation of global warming.

- They are economically more viable and an eco-friendly tool.
- They are capable of not only photosynthesis but also fix up atmospheric nitrogen, and they can survive better under the nutrient-limited conditions.
- Microalgae cultures can be cultivated in open ponds or in large-scale water reservoirs. At the same time, the algal growth under laboratory conditions provides reliable and consistent supply of biomass.
- They have regenerative and metal recovery potentiality.
- They generate lesser volume of chemical and/or biological sludge to be disposed off.
- They have high efficiency in dilute effluents and have large surface area to volume ratio.
- They have the potential to treat sites polluted with more than one type of pollutant.

## **4 Removal of Heavy Metals by Algae**

 Microalgae are sensitive indicators of environmental changes, and their ubiquitous presence serves as the basis of most freshwater and marine ecosystems, widely being used in the assessment of risk and development of environmental regulations for metals (Levy et al. [2007](#page-10-0)). Algae are known to accumulate heavy metals and bind with metal ions in uncomplicated aquatic environment in a short period of time by bio-sorption without any problem of toxicity (Afkar et al. [2010](#page-8-0)). Algae have many features that make them an ideal tool for the selective removal and concentration of heavy metals, which include high tolerance to heavy metals, ability to grow both autotrophically and heterotrophically, large surface area/volume ratios, phototaxy, phytochelatin expression and potential for genetic manipulation (Cai et al. 1995). An important biochemical function of algae is their involvement in the shaping of proper ecological relationships and interaction between organisms in the aquatic environment (Wilde and Benemann [1993](#page-12-0); Sandau et al. 1996; Bajguz 2000) by way of accumulating high concentration of heavy metals depending on their concentration in the external environment. The threshold level of heavy metals varies greatly for different algal species, but it increases as the metal concentration in the water decreases (Kelly 1988; Sharma and Azeez [1988](#page-11-0)). However, little attention has been paid to metal removal and detoxification by algae in the natural environment.

 The studies on biosorption of metals by marine algae revealed an interesting adsorption potential of some algal species such as *Ascophyllum nodosum*, *Sargassum baccularia* (Volesky 1994; Chong and Volesky [1995](#page-9-0); Holan et al. [1998](#page-10-0)), *Scenedesmus abundans* (Terry and Stone [2002](#page-11-0)), *Ecklonia radiata* (Matheickal and Yu 1996) and *Sargassum fluitans* (Fourest and Volesky [1996](#page-9-0)). Marine alga *Dunaliella tertiolecta* has been shown to have high phytochelatin (PC) content attributed to its capability to hyperaccumulate Zn and Cd (Tsuji et al. [2002](#page-11-0), [2003](#page-11-0)). Similarly, a periphytic green alga *Stigeoclonium tenue* is also known to produce high amounts of PC-related peptides when adapted to high Zn concentrations (Pawlik-Skowronska 2003). Ettajani et al.  $(2001)$  reported the hyperaccumulation of Cd in microalgae *Skeletonema costatum* and *Tetraselmis suecica* . The use of both marine and freshwater algae for adsorption and elution of gold, silver and cobalt has been reported (Fujita et al. [1992](#page-9-0); Hamdy [2000](#page-10-0)).

 The growth of large amounts of algae due to eutrophication of water body is commonly seen in most of the water bodies. Such eutrophic algae may help to eliminate the toxicity of heavy metals and exert a major influence on the behaviour and fate of trace metals entering freshwaters. Metal accumulation capacity of algal biomass is either comparable or sometimes higher than chemical sorbents (Mehta and Gaur 2005). It has also been reported that metal uptake capacities of certain algae are much higher than the activated carbon, natural zeolite and synthetic ion-exchange resin (Volesky 1992). It provides a cost-effective solution for industrial wastewater management. Algae have been used for pharmaceutical reasons for detoxification of heavy metals in the human body due to a very efficient adsorption of the toxic ions (David and Volesky [1998](#page-9-0)). Moreover, algae possess high metal-binding capacities, because polysaccharides, proteins, or lipids on the surface of their cell walls contain

**Table 2.1** Some work on algae as heavy metal removing agent

some functional groups such as amino, hydroxyl, carboxyl and sulphate, which can act as binding sites for metals (Holan and Volesky [1994](#page-10-0)) (Table 2.1).

# **4.1 Factors Affecting the Removal of Heavy Metals**

 Biosorption of metals by algae may be affected by several factors, including concentration of metals and biomass, pH, temperature, the presence of competing ions, etc.

#### **4.1.1 Effect of pH**

 Most of the studies have shown that sorption of metal ions is a function of pH of the solution. Earlier studies have indicated that ambient pH condition is an important parameter affecting the biosorption of heavy metal ions (Matheickal et al. 1991; Fourest et al. 1994; Matheickal and Yu [1996](#page-10-0)). pH affects the chemistry of the metal, the activity of the metalbinding functional groups in the biomass and the competi-tion of metallic ions (Selatina et al. [2004](#page-11-0)). pH strongly influences the speciation and biosorption ability of the metal ions (Esposito et al. [2001](#page-9-0)). Since a majority of metal-binding groups of algal cell are acidic (e.g. carboxyl), their availability is pH dependent. These groups generate negatively charged surface groups at acidic pH, and electrostatic interactions



between cationic species and the cell surface are responsible for metal biosorption.

 There are numerous studies showing an increase in metal sorption with increasing pH of the solution (Gupta et al.  $2006$ ; Solisio et al.  $2008$ ; Liping et al.  $2008$ ). A decreased metal sorption by algae has been frequently observed at extremely acidic pH (<2) (Özer et al. 1994; Mehta and Gaur [2001a](#page-10-0) ). The marine algae *Sargassum* sp., *Padina* sp., *Ulva* sp. and *Gracilaria* sp. were investigated for their biosorption performance in the removal of lead, copper, cadmium, zinc and nickel from dilute aqueous solutions. Maximum biosorption was found to be at pH 5.0 for lead and copper and at pH 5.5 for cadmium, zinc and nickel (Sheng et al. 2004).

 There is also a great variability in optimum pH for sorption of a particular metal ion by different algal species. For example, the optimum pH for sorption of Cu by cyanobacteria ( *Microcystis aeruginosa* and *Spirulina platensis*) is far greater than that for Cu sorption by green algae ( *Cladophora prolifera* , *Chlorella vulgaris* , *C. kessleri* ). This variability may be related to the differences in chemical composition of cell surface of various algal species. Chojnacka et al. (2005) found a distinct relationship between pH of aqueous metal solution and involvement of functional group in binding of Pb onto *Spirulina maxima* ; for pH ranging between 2–5, 5–9 and 9–12, respectively, the functional groups involved in the binding of Pb were carboxyl, carboxyl and phosphate and carboxyl, phosphate and hydroxyl.

### **4.1.2 Effect of Contact Time**

 The contact time is of great importance in adsorption for the assessment of the suitability of microbes to serve as biosorbents in a continuous flow system. The biosorption consists of two phases: a primary rapid phase that accounts for the major part in the total metal biosorption and a second slow phase that contributes to a relatively small part. Metal uptake increases with an increase in contact time but remains con-stant after equilibrium time period (Murugesan et al. [2006](#page-10-0)). Equilibrium time varied with metals due to the difference in initial metal concentration and affinity of the biosorbent for a particular metal ion.

 The metal removal rates were rapid, with 90 % of the total adsorption taking place within 60 min (Sheng et al.  $2004$ ). Some authors observed that at the initial stage  $(0-12)$ min), the adsorption rate of Pb was so rapid that 74 % of the metal was biologically adsorbed by *Spirulina* (Chen and Pan  $2005$ ). Heavy metal ion ( $Cr^{3+}$ ,  $Cu^{2+}$  and  $Cd^{2+}$ ) removal from solutions by *Spirulina* species showed that the equilibrium reached after 10 min (Chojnacka et al. 2005). An increase in the biosorption of Cu by *Spirogyra* species was observed with increase in contact time from 0 to 120 min and after that becomes almost constant up to 180 min (Gupta et al. [2006](#page-9-0)).

#### **4.1.3 Effect of Temperature**

 Owing to the dependence on metabolism, metal uptake by live cells is considerably affected by variation in temperature. There are reports showing altered metal uptake by live organisms with change in temperature regime (Skowron'ski 1986; Mehta and Gaur 2001a; Mehta et al. [2002a](#page-10-0)), with maximum uptake occurring at specific temperature optima. Higher temperature usually enhances sorption due to the increased surface activity and kinetic energy of the solute (Sag and Kutsal [2000](#page-11-0); Vijayaraghavan and Yun [2007](#page-12-0)). An increase in metal sorption with increasing temperature (Tsezos and Volesky [1981](#page-11-0); Kuyucak and Volesky [1989](#page-10-0); Aksu and Kutsal [1991](#page-8-0); Aksu [2002](#page-8-0)) suggests that metal biosorption by algae is an endothermic process. On the contrary, some studies indicate exothermic nature of metal sorption by algae (Cruz et al. [2004](#page-9-0); Aksu [2001](#page-8-0); Benquell and Benaissa [2002](#page-9-0)). Due to the exothermic nature of some adsorption processes, an increase in temperature has been found to reduce the biosorption capacity of the biomass (Mameri et al. [1999](#page-11-0); Suhasini et al. 1999). A temperature change affects the number of factors which are important in heavy metal biosorption. Some of the factors include (i) the stability of the metal ion species initially placed in solution, (ii) microorganism- metal complex depending on the biosorption sites, (iii) the effect of temperature on the cell wall of microorganisms and (iv) the ionization of the chemical moieties on the cell wall (Sag and Kutsal 2000). Increased biosorption of heavy metals with increasing temperature has been ascribed to bond rupture, which perhaps enhances the number of active sites involved in metal sorption or higher affinity of sites for metals. There are also some reports which show no effect of temperature on metal sorption (Norris and Kelly [1979](#page-11-0); Zhao et al. 1994). It is always desirable to conduct biosorption at room temperature, as this condition is easy to replicate.

#### **4.1.4 Effect of Biomass Concentration**

The dosage of a biosorbent strongly influences the extent of biosorption, and also the amount of metal ion recovered from a solution is affected by biomass concentration. Biomass concentration in solution seems to influence the specific uptake: for lower value of biomass concentrations, there is an increase in the specific uptake (Gadd et al. 1988; Fourest and Roux [1992](#page-9-0)). An increase in biomass concentration leads to interference between the binding sites (Gadd et al. [1988](#page-9-0)). Hence, this factor needs to be taken into consideration in any application of microbial biomass as biosorbent. Conversely the quantity of biosorbed solute per unit weight of biosorbent decreases with increasing biosorbent dosage, which may be due to the complex interaction of several factors.

 An increase in the biomass concentration generally increases the amount of solute biosorbed due to the increase in the surface area of biosorbent, which in turn increases the

availability of metal-binding sites (Esposito et al. [2001](#page-9-0); Mehta and Gaur [2001c](#page-10-0)). However, there is no straightforward relationship between biomass concentration and metal removal as some workers have noticed a decrease in sorption of heavy metals by different algae with increasing biomass concentration (Hamdy 2000; Nuhoglu et al. [2002](#page-11-0); Gong et al. 2005). This may be due to the limited availability of metal, increased electrostatic interactions, interference between binding sites and poor mixing at higher biomass concentrations (Meikle et al. 1990; Fourest et al. 1994).

## **4.1.5 Effect of Initial Metal Ion Concentration**

 Sorption and removal of heavy metals largely depend on the initial metal ion concentration in the solution. Several workers have reported that metal sorption initially increases with an increase in the metal ion concentration in solution and then becomes saturated after a certain concentration of metal (Da Costa and Leite [1991](#page-9-0); Aloysius et al. [1999](#page-8-0); Mehta and Gaur [2001a](#page-10-0), [b](#page-10-0), c; Mehta et al. 2002a, b). This is because at lower initial metal concentrations, the ratio of the initial moles of metal to the available surface area is low; subsequently, the fractional sorption becomes independent of the initial concentration. However, at higher concentrations, the sites available for sorption become fewer compared to the moles of metal present, and hence, the binding of the metal is strongly dependent upon the initial metal concentration. It is always necessary to identify the maximum saturation potential of a biosorbent, for which experiments should be conducted at the highest possible initial metal concentration. Algal cell surface has several kinds of functional groups with varying affinity for an ionic species. Low- and high-affinity functional groups are involved in sorption of metal ions at high and low concentration of metal ions, respectively.

# **4.1.6 Effect of the Presence of Anions and Cations**

 Actual industrial wastewaters contain different kinds of impurities, which may significantly affect metal biosorption (Ho and McKay [2000](#page-10-0)). Some studies indicated that cations and anions in addition to the ion of interest have a generally detrimental impact on metal accumulation (Suh and Kim  $2000$ ). Among such impurities, cations such as Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>+</sup> and  $Ca<sup>+</sup>$  and anions like sodium salts of chloride, nitrate, acetate and EDTA exist in most of the industrial effluents, and they greatly interfere with the metal sorption potential of biosorbents (Chen and Yiacoumi 1997; Lee and Volesky [1997](#page-10-0); Low et al. [2000](#page-10-0)). Earlier low level of accumulation of Co and Cu by algae in the presence of carbonate, orthophosphate, sulphate, nitrate, EDTA and chloride ions has been observed (Rai et al. [1981](#page-11-0)). However, the accumulation of ionic species like nitrate and ammonium also increased in the presence of Cu and Fe in *Anabaena doliolum* (Rai and Mallick [1992](#page-11-0)). The presence of other cations including metal

ions significantly affects metal sorption by algae (Mehta and Gaur [2001a](#page-10-0), [b](#page-10-0); Mehta et al. 2002a, [b](#page-10-0)). Reduced heavy metal uptake in the presence of light metals is attributed to competition for cellular-binding sites or precipitation or complexation by carbonates, bicarbonates, or hydroxides of Ca and Mg (Rai et al. [1981](#page-11-0)). High concentrations of salts like NaCl in solution can also decrease the rate of metal sorption by algae (Cho et al. [1994](#page-9-0)). The inhibitory effect of Na is more pronounced with weakly bound metals such as Zn or Ni. It is important to note that  $Na<sup>+</sup>$  and  $K<sup>+</sup>$ , being monovalent cations, do not compete directly with covalent binding of heavy metals by the biosorbent. Other compounds that could be considered as impurities in metal removal process are surfactants and some chelating agents. The nature of impurities differs depending on the type of effluent to be treated. Whereas most of the studies reported the inhibitory effect of light metal ions on sorption of heavy metals by biosorbent, a few of them showed no effect (Pawlik and Skowronski [1994](#page-11-0); Adhiya et al. [2002](#page-8-0); Axtell et al. [2003](#page-8-0); Jalali-Rad et al. [2004](#page-10-0)).

#### **4.2 Mechanism of Heavy Metal Removal**

 The mechanism of metal biosorption is a complicated process. The status of biomass (living or nonliving), types of biomaterials, properties of metal solution chemistry and environmental conditions such as pH influence the mechanism of metal biosorption. The uptake of heavy metal ions by algae was found to occur in two principal ways: passive uptake due to surface adsorption (metabolism independent) followed by cellular uptake (metabolism dependent) via intracellular transport and chelation (absorption) (Khummongkol et al. 1982; Cho et al. 1994; Yee et al. [2004](#page-12-0)). Some metals such as Pb and Sr may be passively adsorbed by charged polysaccharides in cell wall and intracellular matrix (El-Sheekh et al. [2003](#page-9-0); Osman et al. [2004](#page-11-0); Fathi et al. [2000](#page-9-0); [2005](#page-9-0)); other metals (e.g. Zn, Cd) are taken up actively against large intracellular concentration gradients. As passive biosorption mainly depends on the binding to functional surface ligands, the cell wall structure is the most important for rapid metal ion uptake.

 The probable sites of an algal cell for the binding of metal ions are shown in Fig. [2.2](#page-6-0) . Adsorption occurs directly onto the cell wall in some algae, but the presence of various amounts of mucilage or extracellular polymeric substances (EPS) (Leppard [1995](#page-10-0); Lee [1997](#page-10-0)) in others (e.g. Cyanophyta) may play a key role in metal binding (Weckesser et al. [1988](#page-12-0)). The algal cell wall contains many functional groups, such as hydroxyl (–OH), phosphoryl, amino (–NH<sub>2</sub>), carboxyl (– COOH), sulphydryl (–SH), etc., which confer a negative charge on the cell surface. Since metal ions in water are generally in the cationic forms, they are adsorbed onto the cell surface. The functional group involved in the metal sorption

<span id="page-6-0"></span>



by algae have been identified by FTIR spectroscopy, pH titration, potentiometric and conductimetric titration techniques and also after blocking of functional groups by certain chemicals. Each functional group has a specific dissociation constant (pKa), and it dissociates into corresponding anion and proton at a specific pH (Niu and Volesky [2000](#page-11-0)). The cell wall functional groups are found linked with various cell wall components, e.g. peptidoglycan, teichoic acid, polysaccharides and proteins. Among different cell wall constituents, polysaccharides and proteins have most of the metal-binding sites (Kuyucak and Volesky 1989). Since the distribution and abundance of the cell wall components vary among different algal groups, the number and kind of functional group also vary among them.

Hamdy (2000) reported that metal uptake is dependent on the type of biosorbent, with different accumulation affinities towards the tested elements, and the amount of metal uptake increased steeply by increasing the weight of the biomass. Fathi et al. (2005) reported that the uptake of an element from the surrounding medium is seldom exactly proportional to the amount present in the water. Verma and Singh (1990) reported Cu uptake in a diazotrophic cyanobacterium *Nostoc calcicola* to be biphasic. During the first 10 min, there was a rapid binding of cations to the cell wall, followed by subsequent metabolism-dependent intracellular uptake for at least 1 h.

# **5 Potential Applications of Algae in Biotechnology**

 Algae are one of the potential organisms, which are useful to mankind in several ways. Algal cells constitute a vast potential resource in various applications as follows.

# **5.1 Food and Feed**

 Blue-green algal protein has received worldwide attention either as a supplement or as an alternative source of food. Some strains of *Spirulina* and *Nostoc* are consumed as human food in Chile, Mexico, Peru and the Philippines. *Spirulina* is used as a food supplement because of its excellent nutrient composition (60–70 % protein, 20 % carbohydrate, 5 % lipids, 7 % minerals and 6 % moisture) and digestibility. It is also a rich source of β-carotene, thiamine and riboflavin and is one of the richest sources of vitamin B12. *Nostoc commune* has high amount of fibre and moderate protein; therefore, this species is used as a new dietary fibre source in human diet (Jeraci and Vansoest [1986](#page-10-0)). Halotolerant marine algal species *Dunaliella* is also a rich source of β-carotenoid (Gudin and Chaumont [1991](#page-9-0) ). Green alga *Chlorella vulgaris* is used as a food supplement in many countries including China, Japan, Europe and the USA (Yamaguchi [1997](#page-12-0)).

# **5.2 Fine Chemicals**

A variety of fine chemicals such as pigments, vitamins and enzymes with various applications can be obtained commercially from different algal strains. Some marine algae are a potential source for commercial production of vitamins such as vitamin B complex and vitamin E (Borowitzka 1988). *Haematococcus pluvialis* accumulate the highest level of astaxanthin which is a potent radical scavenger and a singlet oxygen quencher that surpasses the antioxidant benefits of  $\beta$ -carotene, vitamin C and vitamin E (Lorenz and Cysewski [2000](#page-10-0)). A number of cyanobacteria are also rich in vitamins and many excrete them into the surrounding environment (Borowitzka [1988](#page-9-0)). The products obtained from cyanobacteria like carotenoids and phycobiliproteins are used as natural food colourants and also as food additives and have high commercial value (Emodi [1978](#page-9-0)). *Dunaliella salina* , a halotolerant microalga, is able to accumulate very large amount of β-carotene and also a valuable chemical, glycerol (Avron [1992 ;](#page-8-0) Oren [2005 \)](#page-11-0). Feed grade *Phormidium valderianum* is an excellent source of phycocyanin, a blue natural colourant useful as a phycofluor in diagnostics. Cyanobacteria being photoautotrophs have the ability to photosynthetically transform simple, labelled compounds such as  $14CO<sub>2</sub>$ ,  $13CO<sub>2</sub>$ ,  $33H<sub>2</sub>O$  and  $15NO<sub>3</sub>$  into complex organic compounds. In addition, cyanobacteria are a rich source of polysaccharides, lipids, amino acids, fatty acids, halogenated compounds, etc., which are used as flocculants, surfactants and others (Patterson 1996).

 Enzymes that can be exploited commercially such as chitinase, L-asparaginase, L-glutaminase, amylase, protease, lipase, cellulase, urease and superoxide dismutase have been reported from several algal strains (Prabhakaran et al. [1994](#page-11-0); Wikstrom et al. 1997). Several common and unique sequence-specific endonucleases are known from *Anabaena cylindrica* (Acy I), *A. flos-aquae* (Afl I and Afl III), *A. variabilis* (Ava I and Ava II), *A. variabilis UW* (Avr II), *Microcoleus* sp. *UFEX* 2220 (Mst II) and *Nostoc* sp. *PCC* 7524 (Nsp C I), which can be marketed at low cost since the relative biomass production of cyanobacteria is much less expensive than bacteria (Elhai and Wolk [1988](#page-9-0) ). Marine microalgae *Isochrysis galbana* and *Diacronema vlkianum* produce long-chain fatty acids, mainly eicosapentaenoic acid (EPA, 20:53) and doco-sahexaenoic acid (DHA, 22:6ω3) (Liu and Lin [2001](#page-10-0)).

# **5.3 Pharmaceuticals**

 Algae are one of the richest sources of known and novel bioactive compounds with wide pharmaceutical applications (Raghavan et al. [2002](#page-11-0)). The reported biological activities comprise cytotoxic, antitumor, antibiotic, antimicrobial, antiviral (e.g. anti-HIV) activities as well as biomodulatory effects like immunosuppressive and anti-inflammatory properties (Burja et al. [2001 \)](#page-9-0). *Chlorella vulgaris* has been used as an alternative medicine in the Far East. It is considered an important curing agent for many kinds of health disorders such as gastric ulcers, wounds, constipation, anaemia, hypertension, diabetes, infant malnutrition and neurosis (Yamaguchi [1997](#page-12-0)). A preventive action of *Chlorella* against atherosclerosis and hypercholesterolemia is attributed to glycolipids and phospholipids and antitumor actions assigned to glycoproteins, peptides and nucleotides (Yamaguchi [1997](#page-12-0)). However, the most important substance in *Chlorella*, i.e. beta-1,3-glucan, is an active immunostimulator, a free radical scavenger and a reducer of blood lipids (Spolaore et al. 2006). The anti-HIV activity of marine cyanobacterial

compounds from *Lyngbya lagerheimii* and *Phormidium tenue* was also reported (Gustafson et al. 1989). Halophilic marine algae *Dunaliella* has anticancerous, anti-atherosclerosis, antiinflammatory, anti-allergic, antidiabetic, antibacterial and antiviral properties (Hennekens et al. 1996; Fujitani et al. [2001](#page-9-0); Ayelet et al. [2008](#page-8-0); Francisco et al. 2009; Nakazawa et al. [2009](#page-11-0)). Medically important gamma-linolenic acid (GLA) is relatively rich in cyanobacteria *Spirulina platensis* which is easily converted into arachidonic acid in the human body and then into prostaglandin E2, which has lowering action on blood pressure and contracting function of the smooth muscle, thus playing an important role in lipid metabolism (Thajuddin and Subramanian [2005](#page-11-0)).

# **5.4 Biofertilizer**

Several cyanobacterial strains colonize paddy fields where heterocystous species are capable to fix atmospheric nitro-gen (Mishra and Pabbi [2004](#page-10-0)). However, a variety of nonheterocystous cyanobacteria are also able to fix atmospheric nitrogen under microaerophilic conditions. The role of N2-fixing cyanobacteria in maintenance of the fertility of rice fields has been well substantiated and documented all over the world (Saadatnia and Riahi [2009](#page-11-0)). In India alone, the beneficial effects of cyanobacteria on yield of many rice varieties have been demonstrated in a number of field locations (Venkataraman [1981](#page-12-0)). Beneficial effects of cyanobacterial inoculation have also been reported on a number of other crops such as barley, oats, tomato, radish, cotton, sugarcane, maize, chilli and lettuce (Kaushik and Venkataraman [1979](#page-10-0); Thajuddin and Subramanian [2005](#page-11-0)).

 Red marine algae *Laurencia obtusa* , *Corallina elongata* and *Jania rubens* were also used as biofertilizers by some workers to enhance the growth of maize (Zea mays L.) plants, and it was reported that the mixture of these algae is more suitable for the growth of maize in the field (Safinaz and Ragaa [2013](#page-11-0)). Lozano et al. (1999) stated that the application of an extract from algae to soil or foliage increased ash, protein and carbohydrate content of potatoes.

#### **5.5 Wastewater Treatment**

 Algal species have been used for many decades in wastewater treatment because of its high capacity to uptake inorganic nutrients (Talbot and De la Noue [1993](#page-11-0); Bajguz [2000](#page-8-0); Ettajani et al. 2001; Tsuji et al. [2002](#page-11-0), 2003; Afkar et al. 2010). The importance of microalgae in wastewater treatment has increased in recent years due to the biotechnological potential for producing valuable substances for biofuel production and animal feed (Pulz and Gross [2004](#page-11-0); Spolaore et al. [2006](#page-11-0)). Marine cyanobacteria *Oscillatoria* sp. BDU 10742 and

<b>Species</b>	Product	Application areas	References
Spirulina sp.	Phycocyanin, biomass	Health food, cosmetics, wastewater treatment	Lee $(2001)$ and Costa et al. $(2003)$
Chlorella vulgaris	<b>Biomass</b>	Health food, food supplement, feed surrogates, wastewater treatment	Lee $(2001)$
Dunaliella salina	Carotenoids, $\beta$ -carotene	Health food, food supplement, feed	Jin and Melis $(2003)$ and Del Campo et al. $(2007)$
Haematococcus pluvialis	Carotenoids, astaxanthin	Health food, feed additives, pharmaceuticals	Del Campo et al. (2007)
Odontella aurita	Fatty acids	Pharmaceuticals, cosmetics, baby food	Pulz and Gross (2004)
Porphyridium cruentum	Polysaccharides	Pharmaceuticals, cosmetics, nutrition	Fuentes et al. (1999)
Isochrysis galbana	Fatty acids	Animal nutrition	Molina Grima et al. (2003) and Pulz and Gross $(2004)$
Phaeodactylum tricornutum	Lipids, fatty acids	Nutrition, fuel production	Yongmanitchai and Ward (1991)
Lyngbya majuscula	Immune modulators	Pharmaceuticals, nutrition	Singh et al. $(2005)$
Muriellopsis sp.	Carotenoids, lutein	Health food, food supplement, feed	Blanco et al. (2007) and Del Campo et al. $(2007)$

<span id="page-8-0"></span> **Table 2.2** Important microalgal species, their products and applications

*Aphanocapsa* sp. BDU 16 were able to treat a factory effluent rich in calcium and chloride and enabled 100 % survival of *Tilapia* fish with only cyanobacteria as the feed source (Uma and Subramanian 1990). Shashirekha et al. (1997) found that *Phormidium valderianum* BDU 30501 was able to treat phenol-containing effluents. Studies at the National Facility for Marine Cyanobacteria (NFMC) have identified suitable cyanobacteria for treating a number of noxious effluents containing organophosphorus pesticides, detergents, antibiotics and other molecules (Subramaninan and Uma [1996](#page-11-0)) and also for the degradation of solid wastes like coir pith by their lignolytic action (Malliga et al. [1996](#page-10-0)). There are several reports for the treatment of heavy metalcontaminated wastewater by using marine and freshwater algae and also sorption/desorption of heavy metal for the recovery of this valuable resource (Hamdy 2000; Ettajani et al.  $2001$ ; Terry and Stone  $2002$ ; Dixit and Singh  $2013$ , [2014](#page-9-0)) (Table 2.2).

# **6 Conclusion**

 During the last few decades, the researchers have started viewing the algal cells as photobioreactor which can be exploited by many ways in different spheres of biotechnology at minimal cost. The algal cells require only water, minimum quantity of nutrients, sunlight and  $CO<sub>2</sub>$  for their growth and survival. Phycoremediation using these algal cells is finding favour for the treatment of variety of wastewaters due to their minimal need for space and easy-to-grow characteristics. The resulting biomass after bioremediation (phycoremediation) has the potential to be used as animal feed, production of biofuel and other industrially relevant bioproducts. Thus, phycoremediation technology of wastewater treatment is considered as a low-cost technology which

promises a more sustainable and environment-friendly way of life.

# **References**

- Adhiya J, Cai X, Sayre RT, Traina SJ (2002) Binding of aqueous cadmium by the lyophilized biomass of *Chlamydomonas reinhardtii* . Colloids Surf A Physicochem Eng Asp 210:1–11
- Afkar E, Ababna H, Fathi AA (2010) Toxicological response of the green alga *Chlorella vulgaris* to some heavy metals. Am J Environ Sci 6(3):230–237
- Aksu Z (2001) Biosorption of reactive dyes by dried activated sludge. Equilibrium and kinetic modelling. Biochem Eng J 7:79–84
- Aksu Z (2002) Determination of the equilibrium, kinetic and thermodynamic parameters of the batch biosorption of nickel (II) ions onto *Chlorella vulgaris* . Process Biochem 38:89–99
- Aksu Z, Kutsal T (1991) A bioseparation process for removing lead (II) ions from waste water by using *C. vulgaris* . J Chem Technol Biotechnol 52:109–118
- Aloysius R, Karim MIA, Arif AB (1999) The mechanism of cadmium removal from aqueous solution by non-metabolizing free and immobilized live biomass of *Rhizopus oligosporus* . World J Microbiol Biotechnol 15:571–578
- Anjana K, Kaushik A, Kiran B, Nisha R (2007) Biosorption of Cr (VI) by immobilized biomass of two indigenous strains of cyanobacteria isolated from metal contaminated soil. J Hazard Mater 148:383–386
- Avron M (1992) Osmoregulation. In: Avron M, Ben-Amotz A (eds) *Dunaliella* : physiology, biochemistry and biotechnology. CRC Press, Boca Raton
- Axtell NR, Sternberg SP, Claussen K (2003) Lead and nickel removal using *Microspora* and *Lemna minor* . Bioresour Technol 89:41–48
- Ayelet H, Dror H, Daniella M, Hofit C, Iris B, Yehuda K, Ayelet G, Yariv G, Ami BA, Aviv SA (2008) 9-cis b-carotene-enriched diet inhibits atherogenesis and fatty liver formation in LDL receptor knockout Mice. J Nutr 138:1923–1930
- Bajguz A (2000) Blockade of heavy metals accumulation in *Chlorella vulgaris* cells by 24-epibrassinolide. Plant Physiol Biochem 38:797–801
- Banerjee M, Mishra S, Chatterjee J (2004) Scavenging of nickel and chromium toxicity in Aulosira fertilissima by immobilization: effect

<span id="page-9-0"></span>on nitrogen assimilating enzymes. Electron J Biotechnol 7:302–309

- Barakat MA (2011) New trends in removing heavy metals from industrial wastewater. Arab J Chem 4(4):361–377
- Benquell B, Benaissa H (2002) Cadmium removal from aqueous solution by chitin: kinetic and equilibrium studies. Water Res 36:2463–2474
- Blanco AM, Moreno J, Del Campo JA, Rivas J, Guerrero JLG (2007) outdoor cultivation of lutein rich cells of *Muriellopsis* sp in open ponds. Appl Microbiol Biotechnol 73:1259–1266
- Borowitzka MA (1988) Vitamins and fine chemicals from microalgae. In: Borowitzka MA, Borowitzka LJ (eds) Microalgal biotechnology. Cambridge University Press, Cambridge, pp 153–196
- Burja AM, Banaigs B, Abou-Mansour E, Burgess JG, Wright PC (2001) Marine Cyanobacteria-a prolific source of natural products. Tetrahedron 57:9347–9377
- Cai XH, Logan T, Gustafson T, Traina S, Sayre RT (1995) Applications of eukaryotic algae for the removal of heavy metals from water. Mol Mar Biol Biotechnol 4:338–344
- Cain A, Vannela R, Woo LK (2008) Cyanobacteria as a biosorbent for mercuric ion. Bioresour Technol 99:6578–6586
- Chacoón-Lee TL, González-Mariño GE (2010) Microalgae for "Healthy" foods-possibilities and challenges. Compr Rev Food Sci Food Saf 9:655–675
- Chen H, Pan S (2005) Bioremediation potential of *Spirulina*: toxicity and biosorption studies of lead. J Zhejiang Univ Sci 6B(3):171–174
- Chen JP, Yiacoumi S (1997) Biosorption of metal ions from aqueous solutions. Sep Sci Technol 32:51–69
- Cho DY, Lee ST, Park SW, Chung AS (1994) Studies on the biosorption of heavy metals into *Chlorella vulgaris* . J Environ Sci Health Part A 29:389–409
- Chojnacka K, Chojnacki A, Górecka H (2005) Trace element removal by Spirulina sp. from copper smelter and refinery effluents. Hydrometallurgy 73:147–153
- Chojnacka K (2010) Biosorption and bioaccumulation-the prospects for practical applications. Environ Int 36:299–307
- Chojnacka K, Chojnacki A, Górecka H (2005) Biosorption of Cr<sup>3+</sup>, Cd<sup>2+</sup> and Cu<sup>2+</sup> ions by blue-green algae *Spirulina* sp.: kinetics, equilibrium and the mechanism of the process. Chemosphere 59:75–84
- Chong KH, Volesky B (1995) Description of two-metal biosorption equilibria by Langmuir-type models. Biotechnol Bioeng 47:451–460
- Costa JAV, Colla LM, Duarte P (2003) *Spirulina platensis* growth in open raceway ponds using freshwater supplemented with carbon, nitrogen and metal ions. Zeitschrift fur naturforschung C-A J Biosci 58:76–80
- Cruz CCV, Da Costa ACA, Henriques CA, Luna AS (2004) Kinetic modeling and equilibrium studies during cadmium biosorption by dead *Sargassum* sp. biomass. Bioresour Technol 91:249–257
- Da Costa ACA, Leite SGF (1991) Metal biosorption by sodium alginate immobilized *Chlorella homosphaera* cells. Biotechnol Lett 13:359–362
- David K, Volesky B (1998) Advances in biosorption of heavy metals. Trends Biotechnol 16:291–300
- Davis TA, Volesky B, Mucci A (2003) A review of biochemistry of heavy metal biosorption by brown algae. Water Res 37:4311–4330
- Del Campo JA, Garcia-Gonzale M, Guerrero MG (2007) Outdoor cultivation of microalgae for carotenoid production: current state and perspectives. Appl Microbiol Biotechnol 74:1163–1174
- Dixit S, Singh DP (2013) Phycoremediation of lead and cadmium by employing *Nostoc muscorum* as biosorbent and optimization of its biosorption potential. Int J Phytoremediation 15:801–813
- Dixit S, Singh DP (2014) An evaluation of Phycoremediation potential of cyanobacterium *Nostoc muscorum*: characterization of heavy metal removal efficiency. J Appl Phycol 26:1331-1342
- Dönmez GC, Aksu Z, Ozturk A, Kutsal T (1999) A comparative study on heavy metal biosorption characteristics of some algae. Process Biochem 34:885–892
- Elhai J, Wolk CP (1988) Conjugal transfer of DNA to cyanobacteria. Methods Enzymol 167:747
- El-Enany AE, Issa AA (2000) Cyanobacteria as a biosorbent of heavy metals in sewage water. Environ Toxicol Pharmacol 8:95–101
- El-Sheekh MM, El-Naggar AH, Osman MEH, El-Mazaly E (2003) Effect of cobalt on growth, pigments and the photosynthetic electron transport in *Monoraphidium minutum* and *Nitzschia perminuta* . Braz J Plant Physiol 15:159–166
- El-Sheekh MM, El-Shouny WA, Osman MEH, El-Gammal EWE (2005) Growth and heavy metals removal efficiency of Nostoc muscorum and Anabaena subcylindrica in sewage and industrial wastewater effluents. Environ Toxicol Pharmacol 19:357-365
- Emodi A (1978) Carotenoids: properties and applications. Food Technol 32:38–42
- Esposito A, Pagnanelli F, Lodi A, Solisio C, Vegliό F (2001) Biosorption of heavy metals by *Sphaerotilus natans*: an equilibrium study at different pH and biomass concentrations. Hydrometallurgy 60(2):129–141
- Ettajani H, Berthet B, Amiard JC, Chevolot L (2001) Determination of cadmium partitioning in microalgae and oysters: contribution to the assessment of trophic transfer. Arch Environ Contam Toxicol 40:209–221
- Fathi AA, Zaki FT, Fathy AA (2000) Bioaccumulation of some heavy metals and their influence on the metabolism of *Scenedesmus bijuga* and *Anabaena spiroides* . Egypt J Biotechnol 7:293–307
- Fathi AA, Zaki FT, Ibraheim HA (2005) Response of tolerant and wild type strains of *Chlorella vulgaris* to copper with special references to copper uptake system. Protistology 4:73–78
- Fourest E, Roux JC (1992) Heavy metal biosorption by fungal mycelial by-products: mechanism and influence of pH. Appl Microbiol Biotechnol 37(3):399–403
- Fourest E, Volesky B (1996) Contribution of sulfonate groups and alginate to heavy metal biosorption by the dry biomass of *Sargassum fl uitans* . Environ Sci Technol 30:277–282
- Fourest E, Canal C, Roux JC (1994) Improvement of heavy metal biosorption by fungal mycelial by-products: mechanism and influence of pH. Appl Microbiol Biotechnol 37:399–403
- Francisco JS, Alejandro C, Coral B (2009) *Dunaliella salina* extract effect on diabetic rats: metabolic fingerprinting and target metabolites analysis. J Pharm Biomed Anal 49:786–792
- Fuentes MMR, Sanchez JLG, Sevilla JMF, Fernandez FGA, Perez JAS, Grima EM (1999) Outdoor continuous culture of *Porphyridium cruentum* in a tubular photobioreactor: quantitative analysis of the daily cyclic variation of culture parameters. J Biotechnol 70:271–288
- Fujita T, Kuzuno E, Mamiya M (1992) Adsorption of metal ions by river algae. Bunseki Kagaku 108:123–128
- Fujitani N, Sakaki S, Yamaguchi Y, Takenaka H (2001) Inhibitory effects of microalgae on the activation of hyaluronidase. J Appl Phycol 13:489–492
- Gadd GM, White C, DeRome L (1988) Heavy metal and radionuclide uptake by fungi and yeasts. In: Norri PR, Kelly DP (eds) Biohydrometallurgy. A. Rowe, Chippenham
- Gianfreda L, Rao MA (2004) Potential of extracellular enzymes in remediation of polluted soils: a review. Enzyme Microb Technol 35:339–354
- Gong R, Ding Y, Liu H, Chen Q, Liu Z (2005) Lead biosorption and desorption by intact and pretreated *Spirulina maxima* biomass. Chemosphere 58:125–130
- Gudin C, Chaumont D (1991) Les microalgues, de nouvelles sorces de metabolites. Biofuture 106:27–30
- Gupta VK, Rastogi A, Saini VK, Jain N (2006) Biosorption of copper (II) from aqueous solutions by *Spirogyra* species. J Colloid Interface Sci 296:59–63
- <span id="page-10-0"></span> Gustafson KR, Cardellina JH, Fuller RW, Weislon OS, Kiser RF, Snader KM (1989) Antiviral sulfolipids from cyanobacteria (blue–green algae). J Natl Cancer Inst 81:1254
- Guzman MC, Cao EP (2010) Cadmium binding ability of the blue green alga Hapalosiphon welwitschii Nägel under controlled conditions. Phillipp Sci Lett 3(1):76–86
- Hamdy AA (2000) Biosorption of heavy metals by marine algae. Curr Microbiol 41:232–238
- Hawkes JS (1997) What is a heavy metal? J Chem Educ 74(11):1374–1378
- Hennekens CH, Buring JE, Manson JE, Stampfer M, Rosner B, Cook NR, Belanger C, LaMotte F, Gaziano JM et al (1996) Lack of effect of long term supplementation with beta carotene on the incidence of malignant neoplasms and cardiovascular disease. N Engl J Med 334:1145–1149
- Ho YS, McKay G (2000) Correlative biosorption equilibria model for a binary batch system. Chem Eng Sci 55:817–825
- Holan ZR, Volesky B (1994) Biosorption of lead and nickel by biomass of marine algae. Biotechnol Bioeng 43:1001–1009
- Holan ZR, Volesky B, Prasetyo I (1998) Biosorption of cadmium by biomass of marine algae. Biotechnol Bioeng 41(8):819–825
- Inthorn D, Nagase H, Isaji Y, Hirata K, Miyamoto K (1996) Removal of cadmium from aqueous solution by the filamentous cyanobacterium Tolypothrix tenuis. J Ferment Bioeng 6:580–584
- Jalali-Rad R, Ghalocerian H, Asef Y, Dalir ST, Sahaftipour MH, Gharanjik BM (2004) Biosorption of cesium by native and chemically modified biomass of marine algae: introduce the new biosorbent for biotechnology application. J Hazard Mater 116:125–134
- Jeraci J, Vansoest P (1986) In: Spiller G (ed) Handbook of dietary fiber in human nutrition. CRC Press, Boca Raton, pp 299–303
- Jin ES, Melis A (2003) Microalgal biotechnology: carotenoid production by the green algae *Dunaliella salina* . Biotechnol Bioprocess Eng 8:331–337
- Kamaraj R, Muthukannan SK, Nooruddin T (2011) Adsorption isotherms for Cr (VI) by two immobilized marine cyanobacteria. Ann Microbiol. doi[:10.1007/s13213-11-0252-3](http://dx.doi.org/10.1007/s13213-11-0252-3)
- Kaushik BD, Venkataraman GS (1979) Effect of algal inoculation on the yield and vitamin C content of two varieties of tomato. Plant Soil 52:135–137
- Kelly M (1988) Mining and the freshwater environment. BP Elsevier Applied Science, London
- Khoshmanesh A, Lawson F, Prince IG (1996) Cadmium uptake by unicellular green microalgae. Chem Eng J 62:81–88
- Khummongkol D, Canterford GS, Freyer C (1982) Accumulation of heavy metals in unicellular algae. Biotechnol Bioeng 12:2643–2660
- Kiran B, Kaushik A, Kaushik CP (2008) Metal–salt co-tolerance and metal removal by indigenous cyanobacterial strains. Process Biochem 43:598–604
- Kotrba P, Ruml T (2000) Bioremediation of heavy metal pollution exploiting constituents, metabolites and metabolic pathways of livings. A review collect. Czech Chem Commun 65:1205–1247
- Kuyucak N, Volesky B (1989) Accumulation of cobalt by marine alga. Biotechnol Bioeng 33:809–814
- Lee RE (1997) Phycology, 2nd edn. Cambridge University Press, Cambridge
- Lee YK (2001) Microalgal mass culture systems and methods: their limitation and potential. J Appl Phycol 13:307–315
- Lee HS, Volesky B (1997) Interaction of light metals and protons with seaweed biosorbent. Water Res 31:3082–3088
- Leppard GG (1995) The characterization of algal and microbial mucilages and their aggregates in aquatic systems. Sci Total Environ 165:103–131
- Levy JL, Stauberand JL, Jolley DF (2007) Sensitivity of marine microalgae to copper: the effect of biotic factors on copper adsorption and toxicity. Sci Total Environ 387:141–154
- Liping D, Yingying S, Hua S, Xinting W, Xiaobin Z (2006) Biosorption of copper (II) and lead (II) from aqueous solutions by nonliving green algae *Cladophora fascicularis* : equilibrium, kinetics and environmental effects. Adsorption 12:267–277
- Liping DB, Xiaobin Z, Yingying SB, Hua SB, Xinting WA (2008) Biosorption and desorption of  $Cd^{2+}$  from wastewater by dehydrated shreds of *Cladophora fascicularis* . Chinese J Oceanol Limnol 26(1):45–49
- Liu CP, Lin LP (2001) Ultrastructural study and lipid formation of *Isochrysis* sp. CCMP1324. Bot Bull Acad Sin 42:207–214
- Lorenz RT, Cysewski GR (2000) Commercial potential for *Haematococcus* microalgae as a natural source of astaxanthin. Trends Biotechnol 18:160–167
- Low KS, Lee CK, Liew SC (2000) Sorption of cadmium and lead from aqueous solutions by spent grain. Process Biochem 36:59–64
- Lozano MS, Verde Star J, Maitic PK, Orandy CA, Gaona RH, Aranada HE, Rojas GM (1999) Effect of an algal extract and several plant growth regulators on the nutritive value of Potatoes ( *Solanum tuberosum* L. Var. Gigant). archives hat in oamericanos de Nuticion 49:166–170
- Mallick N (2003) Biotechnological potential of *Chlorella vulgaris* for accumulation of Cu and Ni from single and binary metal solutions. W J Microbiol Biotechnol 19:695–701
- Malliga P, Uma L, Subramanian G (1996) Lignolytic activity of the cyanobacterium *Anabaena azollae* ML 2 and the value of coir waste as a carrier for BGA biofertilizer. Microbios 86:175–183
- Mameri N, Boudries N, Addour L, Belhocine D, Lounici H, Grib H, Pauss A (1999) Batch zinc biosorption by a bacterial nonliving *Streptomyces rimosus* biomass. Water Res 33(6):1347–1354
- Matheickal JT, Yu Q (1996) Biosorption of lead from aqueous solutions by marine alga *Ecklonia radiata* . Water Sci Technol 34:1–7
- Matheickal JT, Iyengar L, Venkobachar C (1991) Sorption and desorption of Cu (II) by *Ganoderma lucidum*. Water Poll Res J Canada 26:187–200
- Mehta SK, Gaur JP (2001a) Characterization and optimization of Ni and Cu sorption from aqueous solution by *Chlorella vulgaris* . Ecol Eng 18:1–13
- Mehta SK, Gaur JP (2001b) Concurrent sorption of  $Ni^{2+}$  and  $Cu^{2+}$  by *Chlorella vulgaris* from a binary metal solution. Appl Microbiol Biotechnol 55:379–382
- Mehta SK, Gaur JP (2001c) Removal of Ni and Cu from single and binary metal solutions by free and immobilized *Chlorella vulgaris* . Eur J Protistol 37:261–271
- Mehta SK, Gaur JP (2005) Use of algae for removing heavy metal ions from wastewater: progress and prospects. Crit Rev Biotechnol 25:113–152
- Mehta SK, Singh A, Gaur JP (2002a) Kinetics of adsorption and uptake of Cu<sup>2+</sup> by *Chlorella vulgaris*: influence of pH, temperature, culture age and cations. J Environ Sci Health Part A 37:399–414
- Mehta SK, Tripathi BN, Gaur JP (2002b) Enhanced sorption of Ni<sup>2+</sup> and Cu 2+ by acid pretreated *Chlorella vulgaris* from single and binary metal solutions. J Appl Phycol 14:267–273
- Meikle AJ, Gadd GM, Reed RH (1990) Manipulation of yeast for transport studies: critical assessment of cultural and experimental procedures. Enzyme Microb Technol 12:865–872
- Micheletti E, Colica G, Viti C, Tamagnini P, De Philippis R (2008) Selectivity in the heavy metal removal by exopolysaccharideproducing cyanobacteria. J Appl Microbiol 105:88–94
- Mishra U, Pabbi S (2004) Cyanobacteria: a potential biofertilizer for rice. Resonance 9(6):6–10
- Molina Grima EM, Belarbi EH, Fernandez FGA, Medina AR, Chisti Y (2003) Recovery of microalgal biomass and metabolites: process options and economics. Biotechnol Adv 20:491–515
- Murugesan GS, Sathiskumar M, Swaminathan K (2006) Arsenic from groundwater by pretreated waste tea fungal biomass. Bioresour Technol 97(3):483–487
- <span id="page-11-0"></span> Nakazawa Y, Sashima T, Hosokawa M, Miyashita K (2009) Comparative evaluation of growth inhibitory effect of stereoisomers of fucoxanthin in human cancer cell lines. J Funct Foods 1:88–97
- Niu H, Volesky B (2000) Gold-cyanide biosorption with L-cysteine. J Chem Technol Biotechnol 75:436–442
- Norris PR, Kelly DP (1979) Accumulation of cadmium and cobalt by *Saccharomyces cerevisiae* . J Gen Microbiol 99:317–324
- Nuhoglu Y, Malkoc E, Gürses A, Canpolat N (2002) The removal of Cu (II) from aqueous solution by *Ulothrix zonata* . Bioresour Technol 85:331–333
- Oren A (2005) A hundred years of *Dunaliella* research: 1905–2005. Saline Syst 1:2
- Osman MEH, El-Naggar AH, El-Sheekh MM, El-Mazally E (2004) Differential effects of  $Co^{+2}$  and  $Ni^{+2}$  on protein metabolism in *Scenedesmus obliquus* and *Nitzschia perminuta* . Environ Toxicol Pharmacol 16:169–178
- Özer D, Aksu Z, Kutsal T, Caglar A (1994) Adsorption isotherms of lead (II) and chromium (VI) on *Cladophora crispate*. Environ Technol 15:439–448
- Patterson GML (1996) Biotechnological applications of cyanobacteria. J Sci Ind Res 55:669–684
- Pawlik B, Skowronski T (1994) Transport and toxicity of cadmium: its regulation in the cyanobacterium *Synechocystis aquatilis* . Environ Exp Bot 34(2):225–233
- Pawlik-Skowronska B (2003) When adapted to high concentration the periphytic green alga *Stigeoclonium tenue* produces high amounts of novel Phytochelatin-related peptides. Aquat Toxicol 62:155–163
- Prabhakaran D, Sumathi M, Subramanian G (1994) Ability to use ampicillin as nitrogen source by marine cyanobacterium *Phormidium valderianum* BDU 30501. Curr Microbiol 28:315–320
- Prasad BB, Pandey UC (2000) Separation and preconcentration of copper and cadmium ions from multimetal solutions using Nostoc muscorum-based biosorbents. World J Microbiol Biotechnol 16:819–827
- Priyadarshani I, Sahu D, Rath B (2011) Microalgal bioremediation: current practices and perspectives. J Biochem Technol 3(3):299–304
- Pulz O, Gross W (2004) Valuable products from biotechnology of microalgae. Appl Microbiol Biotechnol 65:635–648
- Raghavan C, Kadalmani B, Thirunalasundari T, Subramanian G, Akbarsha MA (2002) A study of the male antifertility properties of the marine cyanobacterium *Oscillatoria willei* BDU 135011 – a preliminary report. In: Proceedings of the XX symposium. Reporting of biology comparative endocrinology, Bharathidasan University, Tiruchirapalli, pp 137–138
- Rai LC, Mallick N (1992) Removal and assessment of toxicity of Cu and Fe to *Anabaena doliolum* and *Chlorella vulgaris* using free and immobilized cells. World J Microbiol Biotechnol 8:110–114
- Rai LC, Gaur JP, Kumar HD (1981) Phycology and heavy-metal pollution. Biol Rev Philos Soc 56:99–151
- Raungsomboon S, Chidthaisong A, Bunnag B, Inthorn D, Harveya NW (2008) Removal of lead (Pb2+) by the cyanobacterium Gloeocapsa sp. Bioresour Technol 99:5650–5658
- Saadatnia H, Riahi H (2009) Cyanobacteria from paddy fields in Iran as a biofertilizer in rice plants. Plant Soil Environ 55(5):207–212
- Safinaz AF, Ragaa AH (2013) Effect of some red marine algae as biofertilizers on growth of maize (Zea mays L.) plants. Int Food Res J 20(4):1629–1632
- Sag YI, Kutsal T (2000) Determination of biosorption heats of heavy metal ions on *Zoogloea ramigera* and *Rhizopus arrhizus* . Biochem Eng J 6(2):145–151
- Sandau E, Sandau P, Pulz O (1996) Heavy metal sorption by microalgae. Acta Biotechnol 16:227–235
- Say R, Denizli AM, Arica MY (2001) Biosorption of cadmium (II), lead (II) and copper (II) with the filamentous fungus *Phanerochaete chrysosporium* . Bioresour Technol 76(1):67–70
- Selatina A, Boukazoula A, Kechid N, Bakhti MZ, Chergui A, Kerchich Y (2004) Biosorption of lead (II) from aqueous solution by a bacterial dead *Streptomyces rimosus* biomass. Biochem Eng J 19(2):127–135
- Sharma RM, Azeez PA (1988) Accumulation of copper and cobalt by blue-green algae at different temperatures. Int J Environ Anal Chem 32:87–95
- Shashirekha S, Uma L, Subramanian G (1997) Phenol degradation by the marine cyanobacterium *Phormidium valderianum* BDU 30501. J Ind Microbiol Biotechnol 19:130–133
- Sheng PX, Ting YP, Chen JP, Hong L (2004) Sorption of lead, copper, cadmium, zinc, and nickel by marine algal biomass: characterization of biosorptive capacity and investigation of mechanisms. Colloid Interface Sci 275:131–141
- Singh SP, Yadava V (1986) Cadmium tolerance in the Cyanobacterium Anacysis nidulans. Biol Zentralbl 105:539–542
- Singh S, Kate BN, Banerjee UC (2005) Bioactive compounds from cyanobacteria and microalgae: an overview. Crit Rev Biotechnol 25:73–95
- Skowron'ski T (1986) Influence of some physico-chemical factors on cadmium uptake by the green alga *Stichococcus bacillaris* . Appl Microbiol Biotechnol 24:423–425
- Solisio C, Lodi A, Soletto D, Converti A (2008) Cadmium biosorption on *Spirulina platensis* biomass. Bioresour Technol 99:5933–5937
- Spolaore P, Joannis-Cassan C, Durnan E, Isambert A (2006) Commercial applications of microalgae-review. J Biosci Bioeng 101:87–96
- Subashchandrabose SR, Ramakrishnan B, Megharaj M, Venkateswarlu K, Naidu R (2013) Mixotrophic cyanobacteria and microalgae as distinctive biological agents for organic pollutant degradation. Environ Int 51:59–72
- Subramaninan G, Uma L (1996) Cyanobacteria in pollution control. J Sci Ind Res 55:685–692
- Suh JH, Kim DS (2000) Comparison of different sorbents (inorganic and biological) for the removal of  $Pb<sup>2+</sup>$  from aqueous solutions. J Chem Technol Biotechnol 75:279–284
- Suhasini IP, Sriram G, Asolekar SR, Sureshkumar GK (1999) Biosorptive removal and recovery of cobalt from aqueous systems. Process Biochem 34(3):239–247
- Talbot P, De la Noue J (1993) Tertiary treatment of wastewater with *Phormidium bohneri* (Schmidle) under various light and temperature conditions. Water Resour 27(1):153–159
- Tamilselvan N, Saurav K, Kannabiran K (2012) Biosorption of Cr (VI), Cr (III), Pb (II) and Cd (II) from aqueous solutions by *Sargassum wightii* and *Caulerpa racemosa* algal biomass. J Ocean Univ China 11(1):52–58
- Terry PA, Stone W (2002) Biosorption of cadmium and copper contaminated water by *Scenedesmus abundans* . Chemosphere 47:249–255
- Thajuddin N, Subramanian G (2005) Cyanobacterial biodiversity and potential applications in biotechnology. Curr Sci 89(1):47–57
- Timmis KN, Pieper DH (1999) Bacteria designed for bioremediation. Trends Biotechnol 17:200–204
- Tsezos M, Volesky B (1981) Biosorption of uranium and thorium. Biotechnol Bioeng 23:583–604
- Tsuji N, Hirayanagi N, Okada M, Miyasaka H, Hirata K, Zenk MH, Miyamoto K (2002) Enhancement of tolerance to heavy metals and oxidative stress in *Dunaliella tertiolecta* by Zn-induced phytochelatin synthesis. Biochem Biophys Res Commun 293:653–659
- Tsuji N, Hirayanagi N, Iwabe O, Namba T, Tagawa M, Miyamoto S, Miyasaka H, Takagi M, Hirata K, Miyamoto K (2003) Regulation of phytochelatin synthesis by zinc and cadmium in marine green alga *Dunaliella tertiolecta* . Phytochemistry 62:453–459
- Uma L, Subramanian G (1990) Effective use of cyanobacteria in effluent treatment. In: Proceedings of the national symposium on cyanobacterial N2 fixation, IARI, New Delhi, pp 437-444
- <span id="page-12-0"></span>Vannela R, Verma SK (2006)  $Co^{2+}$ ,  $Cu^{2+}$  and  $Zn^{2+}$  accumulation by cyanobacterium Spirulina platensis. Biotechnol Prog 22:1282–1293
- Venkataraman GS (1981) Blue–green algae for rice production. FAO Soil Bull 16:33–42
- Verma SK, Singh SP (1990) Factors regulating copper uptake in cyanobacterium. Curr Microbiol 21:33–37
- Vijayaraghavan K, Yun YS (2007) Utilization of fermentation waste ( *Corynebacterium glutamicum* ) for biosorption of Reactive Black 5 from aqueous solution. J Hazard Mater 141(1):45–52
- Volesky B (1992) Removal of heavy metals by biosorption. American Chemical Society, Washington, DC, pp 462–466
- Volesky B (1994) Advances in biosorption of metals: selection of biomass types. FEMS Microbiol Rev 14:291–302
- Volesky B (1997) Removal and recovery of heavy metals by biosorption. In: Volesky B (ed) Biosorption of heavy metals. CRC Press, Boca Raton, pp 629–635
- Wase DAJ, Forster CF (1997) Biosorbents for metal ions. Taylor & Francis, London
- Weckesser J, Hofmann K, Jürgens UJ, Whitton BA, Raffelsberger B (1988) Isolation and chemical analysis of the sheaths of the filamen-

tous cyanobacteria *Calothrix parietina* and *C. scopulorum* . J Gen Microbiol 134:629–634

- Wikstrom P, Szwajeer E, Brodelius P, Nilsson KN, Mosbach K (1997) Formation of alpha keto acids from amino acids using immobilized bacteria and algae. Biotechnol Lett 4:153
- Wilde EW, Benemann JR (1993) Bioremoval of heavy metals by the use of microalgae. Biotechnol Adv 11:781–812
- Yamaguchi K (1997) Recent advances in microalgal bioscience in Japan, with special reference to utilization of biomass and metabolites: a review. J Appl Phycol 8:487–502
- Yee N, Benning LG, Phoenix VR, Ferris FG (2004) Characterization of metal-cyanobacteria sorption reactions: a combined macroscopic and infrared spectroscopic investigation. Environ Sci Technol 38:775–782
- Yongmanitchai W, Ward OP (1991) Growth of the omega-3-fatty-acid production by Phaeodactylum-Tricornutum under different culture conditions. Appl Environ Microbiol 57:419–425
- Zhao Y, Hao Y, Ramelow GJ (1994) Evaluation of treatment techniques for increasing the uptake of metal ions from solution by non-living seaweed algal biomass. Environ Monit Assess 33:61–70