

Maria C. Calomarde, Javier De Santiago,
and Ignacio Zapardiel

Abstract

Breast cancer is a clinically heterogeneous disease, which necessitates a variety of treatments and leads to different outcomes; in fact, only some women will benefit from chemotherapy. Identifying patients who will respond to chemotherapy and thereby improve their long-term survival has important implications to treatment protocols and outcomes, while identifying nonresponders may enable these patients to avail themselves of other investigational approaches or other potentially effective treatments.

Furthermore, prognostic tools in early breast cancer are inadequate. The evolving field of metabolomics may allow more accurate identification of patients with residual micrometastases.

Metabolomics is a new, rapidly expanding field dedicated to the global study of metabolites in biological systems. Many of the studies have focused on identifying altered metabolic levels in breast cancer cells or tissues and relating these changes to their associated metabolic pathways. Metabolomics provides a strong link between genotype and phenotype and may provide some insight into oncogenesis.

The relatively new approach using metabolomics has just begun to enter the mainstream of cancer diagnostics and therapeutics. As this field advances, metabolomics will take its well-deserved place next to genomics, transcriptomics, and proteomics in both clinical and basic research in oncology.

Results of these investigations show promise for larger studies that could result in more personalized treatment protocols for breast cancer patients.

Keywords

Breast cancer • Metabolomics • Therapy response • Prognosis

M.C. Calomarde, MD • J. De Santiago, MD, PhD
I. Zapardiel, MD, PhD (✉)
Department of Gynecologic Oncology,
La Paz University Hospital, Madrid, Spain
e-mail: ignaciozapardiel@hotmail.com

Introduction

Breast cancer, although histologically similar, is clinically a very heterogeneous and phenotypically diverse disease, which results in a range of

treatment effectiveness and outcomes [1]. It is composed of several biological subtypes that have distinct behavior and response to therapy. This heterogeneity was first noted over 100 years ago with the identification that simple removal of the ovaries was therapeutic in some breast cancer patients, but not others. Breast cancer characterization (profiling) has significantly advanced since the turn of the millennium due to the development of sophisticated technologies, such as gene expression arrays, which permit simultaneous measurement of thousands of genes to create a molecular portrait of the tumor.

As an alternative approach for biomarker discovery, metabolomics (or metabolite profiling) enables identification of small-molecule metabolites in biofluids and tissues that are sensitive to altered pathology [2–4]. High-throughput analytical techniques of nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry (MS) combined with multivariate statistical analyses provide information on a large number of metabolites, including those that have altered levels between healthy subjects and patients with various diseases, including cancer [5–7].

So far, the metabolomic-based approaches have been used in a large variety of applications, including early disease detection, drug response, toxicity and nutritional studies, and basic systems biology [8–11]. Compared with other biomarker discovery approaches for breast cancer, metabolomics provides a strong link between genotype and phenotype and may provide some insight into oncogenesis. Also, once established, tests based on metabolic profiles are relatively inexpensive and rapid and can be automated [12].

A growing number of metabolomic studies are contributing toward an improved understanding of breast cancer, and these advances have been reviewed [9, 13, 14]. Many of the studies have focused on identifying altered metabolic levels in breast cancer cells or tissues and relating these changes to their associated metabolic pathways [15–18]. A very recent study using metabolic profiling of numerous human cancer cell lines found a high correlation between breast cancer (and other cancer) proliferation and the glycine biosynthetic pathway [19]. Previously,

differences between normal and metastatic mammary epithelial cell lines—including upregulation of fatty acid synthesis and alterations in glycolysis, the TCA cycle, and others—were detected using ^{13}C stable isotopic label tracing by 2D NMR and GCeMS methods [18]. Breast cancer tumors could be separated from non-involved tissues based on intensities from spectra generated by high-resolution magic angle spinning (HR-MAS) NMR spectroscopy with a sensitivity of 83 % and a specificity of 100 %. Some metabolites, such as choline and glycine, were found to be significantly upregulated in tumors larger than 2 cm [20].

In another NMR study, a multivariate statistical model based on 67 urinary metabolites successfully identified all the breast cancer patients with high specificity (93 %) [21].

Breast cancer prognostic factors, such as estrogen and progesterone receptor status, could be predicted by HR-MAS NMR-based metabolomics on tissue samples [22].

Metastatic breast cancer patients could be differentiated from early-stage patients with 72 % prediction accuracy using serum samples detected by NMR-based metabolomics [14].

For identifying breast cancer recurrence, a predictive model built on 11 biomarkers detected by combining NMR and two-dimensional gas chromatography mass spectrometry (GC/MS) provided 86 % sensitivity and 84 % specificity [23].

For predicting the response to chemotherapy in the neoadjuvant setting, a metabolomic approach is used. Four metabolites that were identified from NMR and MS methods are well correlated with a pathological complete response (pCR). A statistical model built based on these metabolites predicts pCR with high sensitivity and specificity [24].

Predicting Response to Neoadjuvant Chemotherapy

Neoadjuvant chemotherapy can significantly benefit breast cancer patients; however, the varied response to such therapy means that a significant

proportion of the patient population is subjected to ineffective treatment while at the same time being exposed to the therapy's toxicities [25]. Pathological complete response (pCR), which is defined as the disappearance of the invasive cancer cells in the breast after chemotherapy, is used to evaluate patient response and is strongly associated with improved long-term survival rates [26–28]. Unfortunately, less than 30 % of patients overall show complete response to neoadjuvant chemotherapy [29]. An ability to predict response to chemotherapeutic agents should enable development of personalized treatment protocols, improving survival rates and reducing unnecessary exposure of patients to toxic drugs.

Research focused on finding useful molecular or clinical predictors of pCR to neoadjuvant chemotherapy in breast cancer is relatively sparse. Imaging studies, such as magnetic resonance imaging (MRI) [30] and scintimammography [31, 32], were proposed to predict pathological responses to neoadjuvant chemotherapy, but they are somewhat limited by low sensitivity combined with high costs.

High levels of MUC-1 antigen (CA 15.3) in pretreatment serum and its fall after chemotherapy can predict responses as well [33], but many patients do not exhibit elevation of this marker before treatment, and hence it is not helpful for such patients [34]. Approaches using genomics and immunohistochemistry have been explored to find serum and tissue biomarkers [26, 35–37]. It has been shown that gene signatures such as HER2 overexpression/amplification and lack of ER expression were associated with pCR and certain neoadjuvant chemotherapy regimens [38–40].

Other molecular markers such as tumor RNA [41], glucose-regulated protein (GRP78) [42], and hormone receptors [18, 43] have also been

identified as potential predictors of pCR. However, suboptimal performance is a major issue that limits their wide applicability. Circulating tumor cells (CTC) have also been established as providing outcome predictions from particular therapies; however, CTCs can be detected in less than 30 % of early-stage breast cancer patients, which limits their clinical applicability [44].

Study and Results

In this study, a metabolomic approach is used to predict the response to chemotherapy in the neoadjuvant setting. Serum samples from 28 patients obtained before preoperative chemotherapy have been studied using a combination of NMR, liquid chromatography mass spectrometry (LC-MS), and multivariate statistics methods. Four metabolites that were identified from NMR and MS methods are well correlated with pCR. A statistical model built based on these metabolites predicts pCR with high sensitivity and specificity.

Comparison of the NMR data between different groups of patients using the Student's *t*-test showed four metabolites to be statistically significant ($p < 0.05$) (Table 10.1). These *p*-values indicate that levels of three metabolites, isoleucine, threonine, and glutamine, were significantly different between pCR and stable disease (SD) groups and the levels of two metabolites, threonine and glutamine, were different between PR and SD. Only one metabolite, histidine, differed significantly between pCR and partial response (PR). The LC-MS data showed that the most statistically differentiating compounds found were long-chain lipids or fatty acids. The most interesting of these, linolenic acid, was validated using a pure, commercially obtained compound. This metabolite separated pCR from SD samples

Table 10.1 Summary of NMR metabolites having low *p*-values

Chemical shift	Multiplicity	Assignment	<i>p</i> -value (pCR vs. SD)	<i>p</i> -value (pCR vs. PR)	<i>p</i> -value (PR vs. SD)
4.24	m	Threonine	0.04	0.28	0.30
1.00	s	Isoleucine	0.04	0.01	0.02
2.09	m	Glutamine	0.01	0.10	0.01
7.07	s	Histidine	0.29	0.20	0.54

perfectly. Statistical analysis shows linolenic acid to be significantly different between pCR and SD groups ($p < 0.01$). The concentration distribution for all the metabolites except histidine showed a consistent trend from pCR to PR to SD; while threonine, glutamine, and linolenic acid increased, isoleucine decreased.

Further analysis focused on evaluating the performance of the metabolites in combination. Combining three NMR-derived markers (threonine, glutamine, and isoleucine) with LC-MS detected linolenic acid. The model provides 100 % selectivity and 80 % sensitivity for the prediction of pCR vs. SD with an AUROC of 0.95.

The results suggest that metabolites in the serum of breast cancer patients are indicators of tumor/host metabolism and that they can predict both sensitivity and resistance to chemotherapy a priori.

A prediction model for the outcome of breast cancer neoadjuvant chemotherapy based on metabolic profiling studies is presented. It combines NMR and LC-MS methods. A combination of four metabolites, three detected by NMR: threonine, glutamine, and isoleucine, and one by MS, linolenic acid, distinguishes groups of patients with no, partial, or complete response.

It clearly indicates that several blood-based metabolite markers are sensitive to response and that the approach is promising for predicting the response to chemotherapy. In addition, considering the strong performance as a biomarker, linolenic acid and possibly other fatty acids might be of particular interest for further validation studies.

Potential Early Diagnosis

For breast cancer, screening mammography is considered the gold standard for early detection; however, the sensitivity of this test is between 54 and 77 %, depending on the type of mammography [45]. Furthermore, mammography is uncomfortable for many patients and exposes them to radiation. As a result, many women do not obtain yearly mammograms. There is a need to find a

general screening test for all cancers that would ideally be noninvasive and have high sensitivity and specificity.

Monitoring of blood or urine for glucose and creatinine continues to be an integral part of diagnostic tests run today. Although these one- or two-component chemical tests provide a quick and inexpensive way to monitor health, what distinguishes metabolomics from clinical chemistry is that metabolomics measures tens to hundreds and potentially thousands of metabolites at once, rather than just one or two. Through urinary measurement, it has the potential to become a general screening test because it is convenient, easy to obtain, and noninvasive. In this study, metabolomics is applied to study urine from women with breast cancer.

Study and Results

Comparison of 67 metabolite concentrations from healthy subjects ($n:62$) and subjects with breast cancer ($n:38$) revealed significant differences. Application of multivariate statistical data analysis (OPLS-DA) to this dataset resulted in distinction between individuals with breast cancer and those without. Five of the healthy individuals overlapped with the breast cancer category. The model parameters and validation of the PLS-DA (multivariate statistical data analysis) suggested a good model. OPLS-DA class prediction was performed as for the EOC subjects, on a total of 20 subjects, 10 each of breast cancer and healthy. As may be observed, all breast cancer and healthy test subjects were correctly classified [21].

Analysis of urinary metabolite changes revealed that many metabolites decreased in relative concentration with a cancer phenotype when compared with healthy. That the majority of urinary metabolites appeared to decrease in concentration in cancer patients is a similar result to what has been seen in colon cancer tissue metabolomics. Interestingly, some metabolites that were shown to increase in cancer tissue (such as some of the amino acids) were lower in the urine of cancer patients. Concentrations of many amino

acids decrease in cancer patients relative to healthy. Decreases in tricarboxylic acid (TCA) cycle intermediates are suggestive of a suppressed TCA cycle. In a study of urinary markers of colorectal cancer, it was observed that several TCA cycle intermediates decrease in those with colorectal cancer as compared with those without [46]. The biological reason behind the metabolite changes is largely speculative at this point but likely involves a shift in energy production, as tumors rely primarily on glycolysis as their main source of energy. This phenomenon is known as the Warburg effect [47], and decreases in TCA cycle intermediates and glucose in the urine could be indicative of this phenomenon. Clearly, lower glucose concentrations were observed in women with ovarian cancer as compared with breast cancer. This could be because of the fact that more of the women with ovarian cancer were in an advanced stage of the disease. Furthermore, the use of amino acids by tumors requires the upregulation of amino acid transporters, [48] pulling these metabolites from the blood. Decreases in circulating glucose and amino acids could subsequently result in an overall decrease in energy metabolism elsewhere in the body, diminishing other metabolic pathways such as the urea cycle, resulting in lower concentrations of urea and creatine, and potentially affecting gut microbial population and/or metabolism.

So, it is suggested that a urine test is faster, easier to administer, less costly, and noninvasive and could be used as a prescreen to other forms of more invasive or uncomfortable screening.

Prediction of Prognostic Factors

There are few predictive and prognostic markers in breast cancer, but some specific markers are routinely being used for treatment planning and evaluating prognosis [49]. Estrogen receptor (ER) and progesterone receptor (PgR) status predict a possible endocrine responsive tumor, whereas human epidermal growth factor receptor 2 (HER-2)-positive tumors may be suitable for trastuzumab treatment. ER, PgR, and axillary lymph node status, together with tumor size and

lymphovascular invasion, are important for predicting the clinical outcome of breast cancer patients [49–51].

High-resolution magic angle spinning magnetic resonance spectroscopy (HR-MAS MRS) can be used to describe the metabolic profile of intact tissue samples. Metabolic profiles have been shown to correlate with characteristics of several malignant diseases such as breast [15, 17, 20], brain [52], colon [53], and cervical cancer [54]. More than 30 metabolites have been described by HR-MAS MRS analysis of breast cancer tissue [20].

The study of the metabolic profile of certain cell or tissue types in combination with multivariate and analytical statistics is referred to as metabolomics. In a study, Bathen et al. showed that hormone receptor and axillary lymph node status, as well as histological grade, could be predicted by MR metabolomics [17]. The study by Bathen et al. was, however, performed using spectra from a restricted number of patients ($n:77$) and verified on a small amount of blind samples ($n:12$).

The purpose of a recent study [22] was to further explore the potential of MR metabolomics to provide clinically useful prognostic factors for breast cancer patients. The use of HR-MAS MRS and chemometrics as tools for determining prognostic and predictive factors of breast cancer was evaluated. Several multivariate classification techniques exist, and in this study, partial least squares discriminant analysis (PLS-DA), probabilistic neural networks (PNNs), and Bayesian belief networks (BBNs) were used. The relationship between the metabolic profiles of breast cancer tissue and the status of ER, PgR, and axillary lymph nodes was examined, and blind samples were predicted for verification.

Study and Results

ER and PgR status were best predicted by PLS-DA (Tables 10.2 and 10.3). For ER status, the number of correctly classified blind samples were 44/50 and 42/50 for Kennard-Stone and SPXY sample selection, respectively, while PgR

Table 10.2 Results from prediction of ER status^a

	PLS-DA (1 LVs)	BBN	PNN
<i>Kennard-stone</i>			
Correct classification	44/50	39/50	40/50
Sensitivity (%)	90	95	82
Specificity (%)	82	18	73
<i>SPXY</i>			
Correct classification	42/50	41/50	42/50
Sensitivity (%)	87	97	90
Specificity (%)	73	38	64

Correct classification: number of samples in the test set predicted to have the correct ER status. Sensitivity: the proportion of ER-positive samples correctly classified. Specificity: the proportion of ER-negative samples correctly classified

^aThe best predictions are emphasized in bold

Table 10.3 Results from prediction of PgR status^a

	PLS-DA (1 LVs)	BBN	PNN
<i>Kennard-Stone</i>			
Correct classification	39/50	35/50	35/50
Sensitivity (%)	81	77	71
Specificity (%)	74	58	68
<i>SPXY</i>			
Correct classification	36/50	36/50	36/49 ^b
Sensitivity (%)	77	84	80
Specificity (%)	63	53	63

Correct classification: number of samples in the test set predicted to have the correct PgR status. Sensitivity: the number of PgR-positive samples correctly classified. Specificity: the number of PgR-negative samples correctly classified

^aThe best predictions are emphasized in bold

^bOne row not classified

status had a correct blind sample classification of 39/50 for the Kennard-Stone test set and 36/50 for SPXY. Similar results for both Kennard-Stone and SPXY sample selection indicate robust classification by PLS-DA. The sensitivity and specificity of classification were approximately equal; this is in contrast to the results of PNN and BBN where the sensitivity was higher than the specificity. The higher sensitivity may be due to the fact that, especially for ER status, there are more positive than negative samples. This could lead to networks that are more specialized in

recognizing positive than negative samples. Since the probability of a sample being positive is much higher than the probability of it being negative, the network achieves a greater number of total correct classified samples by classifying most of the samples as positives. In PNNs, this can be partly overcome by the customized fitness function, allowing the user to insert a penalty whenever a negative sample is classified incorrectly. In this study, the same penalty was used for both the Kennard-Stone and the SPXY training and test sets. Although this improved the classification ability of the networks compared to networks without penalty, the classification error was still higher than that achieved by PLS-DA.

A PLS-DA model of the whole dataset with three latent variables (LVs) explains 43.8 % of the *X*-variance and 42.7 % of the *Y*-variance. The score values for ER+ and ER- samples are significantly different for all three LVs (*t*-test, $p < 0.001$), and it is possible to discriminate between ER+ and ER- samples in a score plot of LV1, LV2, and LV3. ER+ and ER- samples are mainly separated on the first LV that represents 70 % of the *Y*-variance explained by the model, and ER- samples have higher score for LV1 than ER+ samples. The loading profile for LV1 reveals that samples with higher score for LV1 have more of the metabolites glycine (Gly), glycerophosphocholine (GPC), choline (Cho), and alanine (Ala) and less ascorbate (Asc), creatine (Cr), taurine (Tau), and phosphocholine (PC) than samples with lower LV1 scores. The regression vector of the PLS-DA model gives an indication of the overall influence of the variables based on all three LVs. The regression vector of ER- samples appears similar to LV1 and shows the same metabolic patterns. In addition, lactate (Lac) appears to be more expressed in ER- samples.

Axillary lymph node status was best predicted by BBN with 34 of 50 blind samples correctly classified. However, this was only true for the samples chosen by SPXY sample selection, and the same number of correctly classified samples was not achieved using Kennard-Stone sample selection. PLS-DA and BBN gave similar results, and overall, all three methods gave unacceptably high classification errors. However, the number

of correctly classified samples was better than expected by chance for all methods. This indicates that there is a difference between the MR spectra of lymph node-positive and lymph node-negative patients and that the metabolic profile is altered in patients with lymphatic spread compared to patients without spread.

In conclusion, ER and PgR status were successfully predicted by MR metabolomics. There is also a relationship between metabolic profile and lymph node status, although prediction of lymph node status based on MR spectra did not reach a reliable level of correctly classified samples. By combining MR spectroscopy with multivariate modeling, the biological differences between different metabolic profiles could be revealed. Here hormone receptor-negative patients appear to have more of the metabolites glycine (Gly), glycerophosphocholine (GPC), and choline (Cho) than receptor-positive patients. The data also indicate different metabolic profiles between ER status and PgR status. Thus, this study has shown that MR profiles contain prognostic information that may be of benefit in treatment planning and patient follow-up, and MR metabolomics may become an important tool for clinical decision-making in breast cancer patients.

Identification of the Presence of Micrometastasis

Current approaches, using traditional clinicopathological features or gene profiling, assess the primary tumor and estimate the risk of recurrence based on the presumption of micrometastatic disease. These tools have limitations. Consequently, an individual's risk may be over- or underestimated.

The 21-gene Oncotype Dx assay was assessed in 355 placebo-treated patients from the NSABP-B14 trial in node-negative ER-positive disease. Ten-year distant recurrence-free survival for these patients treated with surgery alone was 86, 62, and 69 % for low, intermediate, and high recurrence scores, respectively [1]. The 70-gene MammaPrint applied to 151 lymph node-negative patients, only ten of whom received any adjuvant

therapy, showed differential 10-year distant metastases-free survival between good and poor prognosis signatures at 87 and 44 %, respectively [55]. A striking feature of these studies is that some individuals, despite apparent high-risk disease, clearly have excellent long-term outcomes. This reflects heterogeneity of disease, host, and risk and highlights overestimation of risk by current prognostic tools.

An alternative to presuming residual disease is actual measurement of micrometastases. Studies of micrometastatic disease are intriguing, particularly those of isolated tumor cells (ITC) in the bone marrow and circulating tumor cells (CTC) [56–58]. Of particular interest is that not all patients with ITC or CTC develop clinically detectable metastatic disease. Thus, tumor survival depends on both favorable tumor and host characteristics. Indeed, assessment of this dynamic multifactorial interaction is a strength of the evolving field of metabolomics.

Transformed human cells exhibit profound metabolic shifts, particularly reflecting the induction of cell membrane phospholipids biosynthesis and breakdown, and preferential use of glucose through non-oxidative pathways. Metabolomic analyses of patient serum and urine samples have been shown to delineate between healthy, benign, and malignant conditions. Specifically with breast cancer, there is cell line evidence of metabolomic distinction between normal and malignant and, even more specifically, identification of malignant breast cell lines with greater metastatic potential. With breast tissue, metabolomic analyses distinguish normal tissue, benign disease, carcinoma in situ, and invasive carcinoma. The subsequent challenge is to capture the malignant metabolomic signal among the complex serum metabolomic fingerprint for an individual [59].

Information on the metabolite pattern alterations that can be significantly associated to the pathology is directly obtained through statistical analysis of the NMR profiles. A metabolomic fingerprint may exist for micrometastatic disease. More specifically, a fingerprint may exist which identifies the interaction between host and any residual disease.

Metabolomic analyses in breast cancer patients with early and metastatic disease have been carried out and compared. Prognostic ability of the fingerprint has been explored by comparison with 10-year mortality rates determined by the current prognostic tool Adjuvantionline. The pilot model, developed in 44 early breast cancer patients, was then validated in a second cohort of 45 early breast cancer patients.

Study and Results

The appeal of metabolomics is concurrent assessment of tumor and host. Indeed, survival of a specific tumor in a specific host relies on a dynamic interaction, with evasion of normal host immunity and favorable stromal environment for metastatic deposits as key factors. In this recent study, metastatic subjects were characterized by higher values of phenylalanine, glucose, proline, lysine, and N-acetyl cysteine and lower values of lipids, when compared to the spectra of both post- and preoperative patients [60].

A strength of metabolomics, as compared with current prognostic tools, may be confirmation rather than assumption of micrometastatic disease. Results reveal differential metabolomic fingerprints for most early and metastatic breast cancer patients. Among the normal noise of the metabolomic fingerprint, most patients were distinguished based on metabolomic analysis of one serum sample [60].

Metabolomic analysis assigns more patients to low risk than are assigned by Adjuvantionline. Similarly, when compared with conventional clinical and pathological factors, prognostic gene expression signatures generally identify more patients of low risk. The 21-gene Oncotype Dx shows direct concordance of 36 % in relapse risk stratification compared with an adjusted Adjuvantionline [61]. The 70-gene MammaPrint, when compared with Adjuvantionline, had stronger predictive power and provided lower-risk estimates for more patients [62]. These low-risk patients may be spared or receive less intensive adjuvant treatment.

In conclusion, the benefit of metabolomics is the incorporation of both a specific tumor profile with metastatic features and a specific host profile conducive to tumor growth. A preliminary exploration in a limited number of patients of a potential role for the evolving field of metabolomics in assessment of micrometastatic disease in early breast cancer has been presented. Clearly, this approach requires refinement and validation, but the distinction identified between early and late disease and the prognostic role of the metabolomic fingerprint provide an exciting platform for further work.

Early Detection of Recurrence

Common methods of routine surveillance for recurrent breast cancer include periodic mammography, self- or physician-performed physical examination, and blood tests. The performance of such tests is lacking and extensive investigations for surveillance have not proven effective [63]. Often, mammography misses small local recurrences or leads to false positives, resulting in suboptimal sensitivity and specificity and unnecessary biopsies. In view of the unmet need for more sensitive and earlier detection methods, the last decade or so has witnessed the development of a number of new approaches for detecting recurrent breast cancer and monitoring disease progression using blood-based tumor markers or genetic profiles. The *in vitro* diagnostic (IVD) markers include carcinoembryonic antigen (CEA), cancer antigen (CA 15–3, CA 27.29), tissue polypeptide antigen (TPA), and tissue polypeptide specific antigen (TPS). Such molecular markers are thought to be promising since the outcome of the diagnosis based on these markers is independent of expertise and experience of the clinician, and their use potentially avoids sampling errors commonly associated with conventional pathological tests such as histopathology. However, currently, these markers lack the desired sensitivity and/or specificity, and often respond late to recurrence, underscoring the need for alternative approaches [64].

A new approach is to use metabolite profiling (or metabolomics), which can detect disease based on a panel of small molecules derived from the global or targeted analysis of metabolic profiles of samples such as blood and urine, and this approach is increasingly gaining interest. Metabolite profiling utilizes high-resolution analytical methods such as nuclear magnetic resonance (NMR) spectroscopy and mass spectroscopy (MS) for the quantitative analysis of hundreds of small molecules (less than 1,000 Da) present in biological samples. Owing to the complexity of the metabolic profile, multivariate statistical methods are extensively used for data analysis. The high sensitivity of metabolite profiles to even subtle stimuli can provide the means to detect the early onset of various biological perturbations in real time. Metabolite profiling has applications in a growing number of areas, including early disease diagnosis, investigation of metabolic pathways, pharmaceutical development, toxicology, and nutritional studies. Moreover, the ability to link the metabolome, which constitutes the downstream products of cellular functions, to genotype and phenotype can provide a better understanding of complex biological states that promises routes to new therapy development.

Metabolite profiling methods are applied to investigate blood serum metabolites that are sensitive to recurrent breast cancer. We utilize a combination of NMR and two-dimensional gas chromatography resolved MS (GCxGC-MS) methods to build and verify a model for early breast cancer recurrence detection based on a set of 257 retrospective serial samples. Performance of the derived 11-metabolite biomarker model is compared with that of the currently used molecular marker, CA 27.29, in particular, for providing a sensitive test for follow-up surveillance of treated breast cancer patients.

This is the first metabolomic study that combines the information-rich analytical methods of NMR and MS to derive a sensitive and specific model for the early detection of recurrent breast cancer. The results indicate that such an approach may provide a new window for earlier treatment and its benefits.

Study and Results

The development of a metabolomic-based profile for the early detection of breast cancer recurrence is presented in a recent study [23]. The investigation makes use of a combination of analytical techniques, NMR and MS, and advanced statistics to identify a group of metabolites that are sensitive to the recurrence of breast cancer.

The new method distinguishes recurrence from no evidence of disease (NED) patients with significantly improved sensitivity compared to CA 27.29. Using the predictive model, the recurrence in over 55 % of the patients was detected as early as 13 months before the recurrence was diagnosed based on the conventional methods.

Breast cancer recurs in over 20 % of patients after treatment. Up to nearly 50 % improvement in the relative survival of patients can be achieved by detecting at least local recurrence at asymptomatic phase, underscoring the need to develop reliable markers indicative of secondary tumor cell proliferation [65]. Currently, a number of rapid and noninvasive tests based on circulating tumor markers such as carcinoembryonic antigen and cancer antigens are commercially available. However, the performance of these markers may be too poor to be of significant value for improving early detection because the levels of these markers are also elevated in numerous other malignant and nonmalignant conditions unconnected with breast cancer. Considering such limitations, the American Society of Clinical Oncologists (ASCO) guidelines recommend the use of these markers only for monitoring patients with metastatic disease during active therapy in conjunction with numerous other examinations and investigations [66]. The results presented in a recent study [23] based on the detection of multiple metabolites in the patients' blood provide a new approach for earlier detection.

Although perturbation in the metabolite levels were detected for nearly all the 40 metabolites that were used in the initial analysis (Table 10.4), the use of smaller numbers of metabolites provided improved models. Particularly, the group of 11 metabolites (7 from NMR and 4 from GC;

Table 10.4 Summary of clinical and demographic characteristics of the patients used in the Asiago et al. study

Clinical diagnosis	Control		Recurrence	
	Samples	(Patients)	Samples	(Patients)
No evidence of disease (NED)	141	(35)		
Pre-recurrence (pre)	–		67	(20)
Within recurrence (within)	–		18	(18)
Post-recurrence (post)	–		31	(20)
Age mean (range)	53	(37–75)	55	(36–69)
<i>Breast cancer stage</i>				
Stage I	47	(11)	7	(1)
Stage II	53	(16)	21	(5)
Stage III	10	(3)	34	(6)
Unknown	25	(6)	54	(8)
<i>Estrogen receptor status</i>				
Positive	65	(15)	67	(11)
Negative	64	(18)	33	(7)
Unknown	12	(3)	16	(2)
<i>Progesterone receptor status</i>				
Positive	52	(13)	71	(11)
Negative	77	(20)	29	(7)
Unknown	12	(3)	16	(2)
CA27.29	140	(36)	92	(19)
<i>Site of recurrence</i>				
Bone	–		37	(6)
Breast	–		13	(2)
Liver	–		11	(2)
Lung	–		10	(2)
Skin	–		6	(2)
Brain	–		15	(2)
Lymph	–		6	(1)
Multiple sites	–		18	(3)

Table 10.5) contributed significantly to distinguishing recurrence from NED. Further, the predictive model derived from these 11 metabolites performed significantly better in terms of both sensitivity and specificity when compared to those derived using individual metabolites or a group of metabolites derived from a single analytical method, NMR or MS, alone. Evaluation of other models with fewer metabolites indicated that they could also provide useful profiles. The AUROC for an 8-metabolite profile (4 detected by NMR and 4 by GC-MS) was 0.86, while a 7-marker model detected by NMR alone had an AUROC of 0.80. Nevertheless, the model based on 11 metabolites had the best performance and clearly outperformed the accepted monitoring assay CA 27.29 currently used for monitoring

patients. These results promise a significant improvement for early detection and potentially better treatment options for recurring patients.

A number of studies to date have used NMR or MS methods to detect altered metabolic profiles in different types of malignancy owing to the ability of the analytical techniques to analyze a large number of metabolites in a single experiment. In particular, several investigations have focused on establishing breast cancer biomarkers using a metabolomic approach, and numerous metabolites including glucose, lactate, lipids, choline, and amino acids are shown to correlate with breast cancer [20, 67]. A sensitivity of 100 % and specificity of 82 % in the classification of tumor and non-involved tissues was achieved from the analysis of NMR data [20].

Table 10.5 Smaller numbers of metabolites provided improved models

Metabolites	Within and post vs. NED <i>p</i> -value	Pre-recurrence vs. NED <i>p</i> -value
1. Formate	0.0022	0.2
2. Histidine	0.000041	0.18
3. Proline	0.018	0.9
4. Choline	0.000022	0.77
5. Tyrosine	0.25	0.1
6. 3-hydroxybutyrate	0.86	0.96
7. Lactate	0.96	0.54
8. Glutamic acid	0.000018	0.74
9. N-acetylglycine	0.01	0.96
10. 3-hydroxy-2-methyl-butanoic acid	0.00004	0.35
11. Nonanedioic acid	0.4	0.089

p-values for 11 markers, 7 NMR (numbers 1–7) and 4 GCxGC-MS markers (numbers 8–11) for different groups using all samples; within and post-recurrence vs. NED, pre-recurrence vs. NED as determined from the univariate Student's *t*-test

NED no evidence of disease, *Within* within recurrence, *Post* post-recurrence

A majority of these investigations focused on either breast cancer tumors or cell lines and all used NMR methods alone, except for a recent study that utilized a combination of NMR and MS methods [18].

The 11-serum metabolites represent some of the changes in metabolic activity of several pathways associated with breast cancer, including amino acids metabolism (glutamic acid, histidine, proline, and tyrosine), glycolysis (lactate), phospholipid metabolism (choline), and fatty acid metabolism (nonanedioic acid). Choline is one of the most prominent metabolites in cell biology and is invariably associated with increased activity of tumor cell proliferation in breast cancer. Increased lactate is one of the early findings of metabolic changes reported for breast tumors. Similarly, association of a number of amino acids, fatty acids, and organic acids with breast cancer has been established earlier. Correlation of the metabolites with clinical parameters, such as the cancer stage and estrogen and progesterone receptor status, contributes to the extent by which the disease can be detected early. Recently, a link between tumor metabolites and estrogen

and progesterone receptor status was shown with a prediction accuracy of 88 and 78 %, respectively, indicating the metabolic profile does vary with estrogen and progesterone receptor status of the patient [22]. These results support our observations and suggest that inclusion of such parameters may help advance further development of early-detection metabolite profiles.

Therefore, the development of a new tool for the surveillance of breast cancer recurrence based on the metabolic profiling of blood samples from patients obtained serially is recently showed.

The performance of the model was optimal when metabolites detected by both NMR and MS were combined. This multiple metabolite model outperforms the current diagnostic methods employed for breast cancer patients, including the tumor marker CA 27.29, for which comparison data on the same samples was available for direct comparison. Metabolic profiling of blood serum by NMR and mass spectroscopy can detect breast cancer relapse before it occurs, opening a window of opportunity for patients and oncologists to improve treatment.

Conclusion and Future Perspective

The study of all metabolites produced in the body, called metabolomics, which often includes flora and drug metabolites, is the omics approach that can be considered most closely related to a patient's phenotype. Metabolomics has a great and largely untapped potential in the field of oncology, and the analysis of the cancer metabolome to identify biofluid markers and novel drug-gable targets can now be undertaken in many research laboratories

The cancer metabolome has been used to identify and begin to evaluate potential biomarkers and therapeutic targets in a variety of malignancies, including breast, prostate, and kidney cancer. We discuss the several standard techniques for metabolite separation, identification, and usefulness in breast cancer, with their potential problems and drawbacks. Validation of biomarkers and targets may entail intensive use of labor and technology and generally requires a

large number of study participants as well as laboratory validation studies. The field of pharmacometabolomics, in which specific therapies are chosen on the basis of a patient's metabolomic profile, has shown some promise in the translation of metabolomics into the arena of personalized medicine.

The relatively new approach to using metabolomics has just begun to enter the mainstream of cancer diagnostics and therapeutics. As this field advances, metabolomics will take its well-deserved place next to genomics, transcriptomics, and proteomics in both clinical and basic research in oncology.

References

- Paik S, Shak S, Tang G. Expression of the 21 genes in the recurrence score assay and tamoxifen clinical benefit in the NSABP study B-14 of node negative, estrogen receptor positive breast cancer. *J Clin Oncol*. 2005;23:S(Abstr 510).
- Lindon JC, Holmes E, Nicholson JK. Metabolomics and its role in drug development and disease diagnosis. *Expert Rev Mol Diagn*. 2004;4:189–99.
- Nicholson JK, Lindon JC, Holmes E. 'Metabolomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica*. 1999;29:1181–9.
- Nicholson JK, Wilson ID. Understanding 'global' systems biology: metabolomics and the continuum of metabolism. *Nat Rev Drug Discov*. 2003;2:668–76.
- Lanza IR, Zhang SC, Ward LE, Karakelides H, Raftery D, Nair KS. Quantitative metabolomics by (1) H-NMR and LC-MS/MS confirms altered metabolic pathways in diabetes. *PLoS One*. 2010;5:e10538.
- Pan ZZ, Raftery D. Comparing and combining NMR spectroscopy and mass spectrometry in metabolomics. *Anal Bioanal Chem*. 2007;387:525–7.
- Zhang S, Gowda GAN, Asiago V, Shanaiah N, Barbas C, Raftery D. Correlative and quantitative H-1 NMR-based metabolomics reveals specific metabolic pathway disturbances in diabetic rats. *Anal Biochem*. 2008;383:76–84.
- Clayton TA, Lindon JC, Cloarec O, Antti H, Charuel C, Hanton G, et al. Pharmacometabonomic phenotyping and personalized drug treatment. *Nature*. 2006;440:1073–7.
- Gowda GAN, Zhang SC, Gu HW, Asiago V, Shanaiah N, Raftery D. Metabolomics-based methods for early disease diagnostics. *Expert Rev Mol Diagn*. 2008;8:617–33.
- Griffin JL, Kauppinen RA. Tumour metabolomics in animal models of human cancer. *J Proteome Res*. 2007;6:498–505.
- Zhang J, Bowers J, Liu L, Wei S, Gowda GA, Hammoud Z, et al. Esophageal cancer metabolite biomarkers detected by LC-MS and NMR methods. *PLoS One*. 2012;7:e30181.
- Sprattlin JL, Serkova NJ, Eckhardt SG. Clinical applications of metabolomics in oncology: a review. *Clin Cancer Res*. 2009;15:431–40.
- Claudino WM, Quattrone A, Biganzolim L, Pestrin M, Bertini I, Di Leo A. Metabolomics: available results, current research projects in breast cancer, and future applications. *J Clin Oncol*. 2007;25:2840–6.
- Oakman C, Tenori L, Biganzoli L, Santarpia L, Cappadona S, Luchinat C, et al. Uncovering the metabolic fingerprint of breast cancer. *Int J Biochem Cell Biol*. 2011;43:1010–20.
- Cheng LL, Chang IW, Smith BL, Gonzalez RG. Evaluating human breast ductal carcinomas with high resolution magic-angle spinning proton magnetic resonance spectroscopy. *J Magn Reson*. 1998;135:194–202.
- Sitter B, Sonnewald U, Spraul M, Fjösne HE, Gribbestad IS. High-resolution magic angle spinning MRS of breast cancer tissue. *NMR Biomed*. 2002;15(5):327–37.
- Bathen TF, Jensen LR, Sitter B, Fjosne HE, Halgunset J, Axelson DE, et al. SMR-determined metabolic phenotype of breast cancer in prediction of lymphatic spread, grade, and hormone status. *Breast Cancer Res Treat*. 2007;104(2):181–9.
- Yang C, Richardson AD, Smith JW, Osterman A. Comparative metabolomics of breast cancer. *Pac Symp Biocomput*. 2007;12:181–92.
- Jain M, Nilsson R, Sharma S, Madhusudhan N, Kitami T, Souza AL, et al. Metabolite profiling identifies a key role for glycine in rapid cancer cell proliferation. *Science*. 2012;336:1040–4.
- Sitter B, Lundgren S, Bathen TF, Halgunset J, Fjösne HE, Gribbestad IS. Comparison of HR MAS MR spectroscopic profiles of breast cancer tissue with clinical parameters. *NMR Biomed*. 2006;19:30–40.
- Slupsky CM, Steed H, Wells TH, Dabbs K, Schepansky A, Capstick V, et al. Urine metabolite analysis offers potential early diagnosis of ovarian and breast cancers. *Clin Cancer Res*. 2010;16:5835–41.
- Giskeodegard GF, Grinde MT, Sitter B, Axelson DE, Lundgren S, Fjösne HE, et al. Multivariate modeling and prediction of breast cancer prognostic factors using MR metabolomics. *J Proteome Res*. 2010;9:972–9.
- Asiago VM, Alvarado LZ, Shanaiah N, Gowda GAN, Owusu-Sarfo K, Ballas RA, et al. Early detection of recurrent breast cancer using metabolite profiling. *Cancer Res*. 2010;70:8309–18.
- Wei S, Liu L, Zhang J, Bowers J, Gowda GA, Seeger H, et al. Metabolomics approach for predicting response to neoadjuvant chemotherapy for breast cancer. *Mol Oncol*. 2013;7:297–307.
- Wolmark N, Wang J, Mamounas E, Bryant J, Fisher B. Preoperative chemotherapy in patients with operable breast cancer: nine-year results from National

- Surgical Adjuvant Breast and Bowel Project B-18. *J Natl Cancer Inst Monogr.* 2001;30:96–102.
26. Bear HD, Anderson S, Smith RE, Robidoux A, Kahlenberg MS, Margolese RG, et al. A randomized trial comparing preoperative (preop) doxorubicin/cyclophosphamide (AC) to preop AC followed by preop docetaxel (T) and to preop AC followed by postoperative (postop) T in patients (pts) with operable carcinoma of the breast: results of NSABP B-27. *Breast Cancer Res Treat.* 2004;88:S16.
 27. Fisher B, Bryant J, Wolmark N, Mamounas E, Brown A, Fisher ER, et al. Effect of preoperative chemotherapy on the outcome of women with operable breast cancer. *J Clin Oncol.* 1998;16:2672–85.
 28. Kuerer HM, Newman LA, Smith TL, Ames FC, Hunt KK, Dhingra K, et al. Clinical course of breast cancer patients with complete pathologic primary tumor and axillary lymph node response to doxorubicin based neoadjuvant chemotherapy. *J Clin Oncol.* 1999;17:460–9.
 29. Jones RL, Smith IE. Neoadjuvant treatment for early stage breast cancer: opportunities to assess tumour response. *Lancet Oncol.* 2006;7:869–74.
 30. Padhani AR, Hayes C, Assersohn L, Powles T, Makris A, Suckling J, et al. Prediction of clinicopathologic response of breast cancer to primary chemotherapy at contrast-enhanced MR imaging: initial clinical results. *Radiology.* 2006;239:361–74.
 31. Marshall C, Eremin J, El-Sheemy M, Eremin O, Griffiths PA. Monitoring the response of large (>3 cm) and locally advanced (T3-4, N0-2) breast cancer to neoadjuvant chemotherapy using Tc-99m-Sestamibi uptake. *Nucl Med Commun.* 2005;26:9–15.
 32. Sciuto R, Pasqualoni R, Bergomi S, Petrilli G, Vici P, Belli F, et al. Prognostic value of Tc-99m-sestamibi washout in predicting response of locally advanced breast cancer to neoadjuvant chemotherapy. *J Nucl Med.* 2002;43:745–51.
 33. Al-Azawi D, Kelly G, Myers E, McDermott EW, Hill ADK, Duffy MJ, et al. CA 15–3 is predictive of response and disease recurrence following treatment in locally advanced breast cancer. *BMC Cancer.* 2006;6:220.
 34. Kurebayashi J, Yamamoto Y, Tanaka K, Kohno N, Kurosumi M, Moriya T, et al. Significance of serum carcinoembryonic antigen and CA 15–3 in monitoring advanced breast cancer patients treated with systemic therapy: a large-scale retrospective study. *Breast Cancer.* 2003;10:38–44.
 35. Guarneri V, Broglio K, Kau SW, Cristofanilli M, Buzdar AU, Valero V, et al. Prognostic value of pathologic complete response after primary chemotherapy in relation to hormone receptor status and other factors. *J Clin Oncol.* 2006;24:1037–44.
 36. Rouzier R, Perou CM, Symmans WF, Ibrahim N, Cristofanilli M, Anderson K, et al. Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clin Cancer Res.* 2005;11:5678–85.
 37. van't Veer LJ, Dai HY, van de Vijver MJ, He YDD, Hart AAM, Mao M, et al. Gene expression profiling predicts clinical outcome of breast cancer. *Nature.* 2002;415:530–6.
 38. Chen YZ, Chen CM, Yang BL, Xu QH, Wu F, Liu F, et al. Estrogen receptor-related genes as an important panel of predictors for breast cancer response to neoadjuvant chemotherapy. *Cancer Lett.* 2011;302:63–8.
 39. Gianni L, Zambetti M, Clark K, Baker J, Cronin M, Wu J, et al. Gene expression profiles in paraffin-embedded core biopsy tissue predict response to chemotherapy in women with locally advanced breast cancer. *J Clin Oncol.* 2005;23:7265–77.
 40. Thuerigen O, Schneeweiss A, Toedt G, Warnat P, Hahn M, Kramer H, et al. Gene expression signature predicting pathologic complete response with gemcitabine, epirubicin, and docetaxel in primary breast cancer. *J Clin Oncol.* 2006;24:1839–45.
 41. Parissenti AM, Chapman JAW, Kahn HJ, Guo BQ, Han L, O'Brien P, et al. Association of low tumor RNA integrity with response to chemotherapy in breast cancer patients. *Breast Cancer Res Treat.* 2010;119:347–56.
 42. Lee E, Nichols P, Groshen S, Spicer D, Lee AS. GRP78 as potential predictor for breast cancer response to adjuvant taxane therapy. *Int J Cancer.* 2011;128:726–31.
 43. van Poznak C, Tan L, Panageas KS, Arroyo CD, Hudis C, Norton L, et al. Assessment of molecular markers of clinical sensitivity to single-agent taxane therapy for metastatic breast cancer. *J Clin Oncol.* 2002;20:2319–26.
 44. Hayes DF, Smerage J. Is there a role for circulating tumor cells in the management of breast cancer? *Clin Cancer Res.* 2008;14:3646–50.
 45. Skaane P. Studies comparing screen-film mammography and full field digital mammography in breast cancer screening: updated review. *Acta Radiol.* 2009;50:3–14.
 46. Qiu Y, Cai G, Su M, Chen T, Liu Y, Xu Y, et al. Urinary metabolomic study on colorectal cancer. *J Proteome Res.* 2010;9:1627–34.
 47. Garber K. Energy boost: the Warburg effect returns in a new theory of cancer. *J Natl Cancer Inst.* 2004;96:1805–6.
 48. Ganapathy V, Thangaraju M, Prasad PD. Nutrient transporters in cancer: relevance to Warburg hypothesis and beyond. *Pharmacol Ther.* 2009;121:29–40.
 49. Payne SJ, Bowen RL, Jones JL, Wells CA. Predictive markers in breast cancers—the present. *Histopathology.* 2008;52(1):82–90.
 50. Bentzon N, During M, Rasmussen BB, Mouridsen H, Kroman N. Prognostic effect of estrogen receptor status across age in primary breast cancer. *Int J Cancer.* 2008;122(5):1089–94.
 51. Liu S, Chia SK, Mehl E, Leung S, Rajput A, Cheang MC, et al. Progesterone receptor is a significant factor associated with clinical outcomes and effect of adjuvant tamoxifen therapy in breast cancer patients. *Breast Cancer Res Treat.* 2009;122(5):1089–94.

52. Sjobakk TE, Johansen R, Bathen TF, Sonnewald U, Juul R, Torp SH, et al. Characterization of brain metastases using high-resolution magic angle spinning MRS. *NMR Biomed.* 2008;21(2):175–85.
53. Righi V, Durante C, Cocchi M, Calabrese C, Di Febo G, Lecce F, et al. Discrimination of healthy and neoplastic human colon tissues by ex vivo HR-MAS NMR spectroscopy and chemometric analyses. *J Proteome Res.* 2009;8(4):1859–69.
54. Lyng H, Sitter B, Bathen TF, Jensen LR, Sundfor K, Kristensen GB, et al. Metabolic mapping by use of high-resolution magic angle spinning 1H MR spectroscopy for assessment of apoptosis in cervical carcinomas. *BMC Cancer.* 2007;7:11.
55. van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW, et al. A gene-expression signature as a predictor of survival in breast cancer. *N Engl J Med.* 2002;347:1999–2009.
56. Braun S, Vogl FD, Naume B, Janni W, Osborne MP, Coombes RC, et al. A pooled analysis of bone marrow micrometastasis in breast cancer. *N Engl J Med.* 2005;353:793–802.
57. Janni W, Rack B, Schindlbeck C, Strobl B, Rjosk D, Braun S, et al. The persistence of isolated tumor cells in bone marrow from patients with breast carcinoma predicts an increased risk for recurrence. *Cancer.* 2005;103:884–91.
58. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, et al. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med.* 2004;351:781–91.
59. Bollard ME, Stanley EG, Lindon JC, Nicholson JK, Holmes E. NMR-based metabolomic approaches for evaluating physiological influences on biofluid composition. *NMR Biomed.* 2005;18:143–62.
60. Oakman C, Tenori L, Claudino WM, Cappadona S, Nepi S, Battaglia A, et al. Identification of a serum-detectable metabolomic fingerprint potentially correlated with the presence of micrometastatic disease in early breast cancer patients at varying risks of disease relapse by traditional prognostic methods. *Ann Oncol.* 2011;22:1295–301.
61. Goldstein L, Gray R, Badve S, et al. Prognostic utility of the 21-gene assay in hormone receptor-positive operable breast cancer compared with classical clinicopathologic features. *J Clin Oncol.* 2008;26:4063–71.
62. Buyse M, Loi S, van't Veer L, Viale G, Delorenzi M, Glas AM, et al. Validation and clinical utility of a 70-gene prognostic signature for women with node-negative breast cancer. *J Natl Cancer Inst.* 2006;98:1183–92.
63. Pivot X, Asmar L, Hortobagyi GN, Theriault R, Pastorini F, Buzzdar A. Retrospective study of first indicators of breast cancer recurrence. *Oncology.* 2000;58:185–90.
64. Lumachi F, Ermani M, Brandes AA, Basso S, Basso U, Boccagni P. Predictive value of different prognostic factors in breast cancer recurrences: multivariate analysis using a logistic regression model. *Anticancer Res.* 2001;6:4105–8.
65. Houssami N, Ciatto S, Martinelli F, Bonardi R, Duffy SW. Early detection of second breast cancers improves prognosis in breast cancer survivors. *Ann Oncol.* 2009;20:1505–10.
66. Harris L, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, et al. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. *J Clin Oncol.* 2007;25:5287–312.
67. Gribbestad IS, Sitter B, Lundgren S, Krane J, Axelson D. Metabolite composition in breast tumors examined by proton nuclear magnetic resonance spectroscopy. *Anticancer Res.* 1999;19:1737–46.