# **Understanding the neurobiology of bipolar depression**

Jun-Feng Wang and L. Trevor Young

*Department of Psychiatry, University of British Columbia, 2255 Wesbrook Mall, Vancouver, BC, Canada*

## **Abstract**

Many studies have shown decreased brain volume and cell number in prefrontal and limbic regions of bipolar disorder subjects, suggesting presence of disturbed neuronal circuitry and impaired neuroplasticity in these regions. The serotonin system plays an important role in depression and is a target of antidepressants. Studies have shown that the major serotonin metabolite 5-HIAA and serotonin transporter activity are decreased in cerebrospinal fluid, brain or platelets of subjects with bipolar depression, indicating that an abnormal serotonin system also contributes significantly to this disease. Mitochondria regulate synthesis, release and uptake of neurotransmitters via energy production. Evidence has shown that glucose metabolic rate and cerebral blood flow are decreased in bipolar depression. Studies also suggest defects in mitochondrial electron transport chain and oxidative damage in bipolar disorder. These studies together indicate that bipolar depression may be associated with an abnormal serotonin system resulting from mitochondrial dysfunction-induced impaired neuroplasticity in neuronal circuitry related to mood regulation.

# **Introduction**

The depressed phase of bipolar disorder remains an enormous challenge for patients, their families and to clinicians. Depression is far more common in bipolar disorder than is mania, and is associated with high rates of suicide or suicide attempts and marked impairment in function [1–5]. There has long been debate about whether bipolar depression is similar to or different from unipolar depression, with recognition of different treatment responses and clinical courses suggesting a different neurobiology.

Mood stabilizing drugs are useful for treatment of different phases of the illness including treatment of mania and depression acutely and prophylaxis against relapse into either state. There is a general consensus that treating either acute depression or preventing relapses into depression is more challenging than treating mania and that the drugs we use to treat the disorder are less effective for these symptoms [6]. Therefore, a better understanding of the molecular mechanism of bipolar disorder, and particularly of bipolar depression, is critical to improving treatment for the disease.

Many studies using brain imaging and postmortem brain tissue have revealed reduced brain volume and cell density in prefrontal cortex and limbic regions from subjects with bipolar disorder. These findings suggest that critical neuronal circuits made up of regions that modulate emotion and mood are disturbed, which subsequently affects synthesis, release, reuptake and other functions of neurotransmitters and results in impairment of neuroplasticity. Serotoninergic neurotransmitter system has been extensively studied in the pathology of major depressive disorder, and is targeted by antidepressants. Studies also implicate the serotoninergic system in bipolar depression and as a target for many mood stabilizing drugs, suggesting that the system may also have a critical role in the pathology of bipolar disorder. It remains to be determined what causes or promotes the loss in neuroplasticity and cell loss in bipolar disorder. Among several plausible hypotheses, the possibility that altered energy metabolism and oxidative stress occurs in the illness lead to cellular damage is becoming increasing accepted. Mitochondria are enriched in both dendrites and synaptic terminals, produce energy for many neuronal functions including synthesis, release and reuptake of neurotransmitters, and are important in regulation of neuronal plasticity and surviving. Increasing evidence suggest impaired energy metabolism and mitochondrial dysfunction in brain of bipolar disorder patients.

In the following chapter, we review findings on the neurobiology of depression. Most studies in bipolar disorder have not focused only on the depressed phase of illness but have looked at patients in all phases of illness. There is a long debate about whether these may be a specific neurobiology of depression in bipolar disorder and how it relates to unipolar depression. We have focused on the three areas described above as they are related to approaches that have been successful in the study of major depression. We suggest in this review that cell loss and decreased neuroplasticity occurred in key limbic and cortical regions lead to monoaminergic dysfunction including the serotonin system, which could contribute to depressive symptoms. Finally we suggest that altered energy metabolism occurs in bipolar disorder which ultimately results in cell damage and altered neurotransmission.

# **Structural abnormality in frontal and limbic brain regions of subjects with bipolar disorder**

Mood regulation appears to be related to activity in the prefrontal cortex and anterior cingulate cortex, and in the hippocampal and amygdalar limbic brain regions, all of which together regulate behaviors such as emotion, attention, motivation and cognition. Prefrontal cortex plays a critical role in thought processes that control mood, cognition and motor behavior functions. Anterior cingulate cortex is important to attentional, emotional and cognitive mental functions, and in the initiation of motivated behavior. Hippocampal and amygdalar limbic regions are involved in production of emotion, in motivation, and in emotional association with memory  $[7-10]$ . In patients with bipolar disorder, compelling evidence for structural abnormalities, cell loss and reduced cell density in these key brain regions associated with mood regulation has been provided by neuroimaging and postmortem brain studies.

Using magnetic resonance imaging, a number of these studies have found a decrease in grey matter volume in prefrontal cortex from bipolar and unipolar depression patients (Fig. 1)  $[11-14]$ . However, the findings for changes in hippocampal and amygdalar volume in adult bipolar subjects are equivocal compared to those for prefrontal cortex [15–21]. The reasons for the discrepant data for structural hippocampal and amygdalar changes may reflect the varied histories of illness diagnosis and treatment that are characteristic of adult bipolar disorder populations. On the other hand, recent studies in children and adolescents with bipolar disorder have consistently reported decreased amygdalar volume [22–25]. Altshuler et al. [26] found that amygdalar volume in bipolar disorder subjects with a history of psychoses was positively correlated with the number of manic episodes, indicating that course of illness may play a role in exacerbating volumetric changes.

Postmortem brain studies have provided direct evidence suggesting that reduced cell number and cell density may contribute to structural changes in the brain that have been observed in bipolar disorder subjects. Rajkowska et al.



Figure 1. MRI-based volumes of the left subgenual prefrontal cortex (PFC) grey matter differed between the bipolar disordered, unipolar depressed and control groups (ANOVA,  $F = 9.8$ ;  $P < 0.0002$ , co-varying for gender, age and whole brain volume). *Post hoc* tests (Tukey-Kramer) showed significant volumetric decreases in the bipolar and unipolar groups relative to the control group. Wholebrain MRI volume and skull X-ray measures (product of anterior-posterior distance between the inner tables of the skull at the estimated bicommissural segment and perpendicular distance from the midpoint of the bicommissural segment to the vertex) were slightly smaller in the unipolar group compared with the bipolar and control groups, and did not differ between the bipolar and control groups. The left subgenual PFC/whole brain volume ratio was also reduced in the bipolar and unipolar groups, being  $1.8 \times 10^{-4} \pm 0.74 \times 10^{-4}$ ,  $1.1 \times 10^{-4} \pm 0.64 \times 10^{-4}$ , and  $1.1 \times 10^{-4} \pm 0.51 \times 10^{-4}$  in the control, bipolar, and unipolar groups, respectively  $(F = 8.4; P < 0.001)$  [11].

[27] found, in the prefrontal area, that pyramidal cell density in layers III and V were decreased in bipolar disorder subjects. Non-pyramidal neurons were also reported to be decreased in layer II of anterior cingulate cortex and in the CA2 region of hippocampus [28, 29]. Recent data have also indicated decreased neuronal size and number in the lateral nucleus of the amygdala in subjects with bipolar disorder [30, 31]. Aberrant mossy fiber sprouting has been observed in the hippocampus of bipolar disorder subjects [32], which is suggestive of altered neuroplasticity. Levels of the neuronal viability marker N-acetyl-aspartate was shown to be decreased in *post mortem* prefrontal cortex of bipolar disorder subjects [33, 34]. Glial number and density were also reduced in the same region [27, 35]. The glial fibrillary acidic protein isoforms were found to be decreased in the prefrontal cortex of both bipolar disorder and unipolar depression subjects, suggesting decreased astrocytes [36]. It has also been reported that dystrophic alterations and reduced density occur in oligodendroglial cells in layer VI of BA 9/10 in bipolar disorder subjects [37–39]. These results suggest that both astrocytes and oligodendroglial cells are involved in the changes observed in glial cells.

Changes of volume and cell density in prefrontal and limbic brain regions suggest a possible disruption of neuronal circuits which regulate mood and emotion. Brain derived neurotrophic factor (BDNF) is abundantly and widely distributed in brain. It regulates synapse formation, differentiation and neuronal survival, and plays an important role in regulation of neuroplasticity and behaviors. Recent studies have shown that altered expression of BDNF occurs in brain in bipolar disorder, suggesting that it is an important factor in causing or promoting changes in neuronal structure or density in these brain regions. For example, it has been reported that levels of BDNF protein were lower in postmortem hippocampus of subjects with bipolar disorder than controls [40]. The val66met BDNF polymorphism has also shown to be associated with bipolar disorder [41–44]. Recently, a number of studies also found abnormal expression of BDNF in peripheral blood from bipolar patients. Cunha et al. [45] investigated serum BDNF levels in manic, depressive and euthymic states of BD patients and found that serum BDNF levels in manic and depressive states, but not in euthymic state, were decreased when compared with healthy controls. Decreased serum BDNF levels have also been found in euthymic patients with bipolar disorder, and depressed and euthymic patients with unipolar depression [46]. Our laboratory also found that BDNF levels were significantly lower in lymphoblasts from bipolar disorder subjects when compared with matched healthy controls [47] (Fig. 2). These findings together implicate BDNF may be one of the critical factors which ultimately contribute to a loss of neuroplasticity and cell loss in bipolar disorder.

There are many consequences to disruption of these neurons and impairment of neuroplasticity, one of which is the possibility that these changes may lead to a predisposition for depressive and manic affective responses. The serotoninergic system has long been identified as important in mood and in particular depression. Indeed, as reviewed below both older and very recent data



Figure 2. BDNF protein levels in transformed lymphoblasts from non-psychiatric controls and lithium-responsive bipolar subjects, with and without 1 mM lithium treatment for 7 days, and unaffected relatives. BDNF protein levels were 36% lower in subjects with bipolar disorder compared to nonpsychiatric controls ( $*, p = 0.02$ ), and this decrease remained (33%,  $\hat{\xi}, p = 0.02$ ) after lithium treatment. Lithium treatment decreased BDNF levels in both bipolar and control populations  $(\dagger, p = 0.05)$ . All measures are expressed as mean  $\pm$  standard deviation [47].

suggest the importance of this same neurotransmitter in bipolar depression. We speculate that this may be one of the consequences of the cellular changes in brain of patients with bipolar disorder which is critical to the development of depressive symptoms and episodes.

#### **Serotonin neurotransmitter system in subjects with bipolar depression**

The serotonergic neurons project to many cortical brain regions and regulate many brain functions such as emotion, cognition, motor function, pain sensitivity and many others. Based upon evidence from neuroimaging, postmortem brain, peripheral blood cell, pharmacological and genetic studies, an abnormal serotoninergic system is well acknowledged to play an important role in the pathophysiology of unipolar depression. Although whether molecular and cellular mechanisms are distinct in unipolar depression and bipolar depression remains unclear, a number of studies have also suggested that the serotoninergic system, particularly serotonin level and serotonin transporter reuptake activity, is altered in bipolar depression.

Studies have shown that the turnover of serotonin is altered in bipolar depression. 5-hydroxyindoleacetic acid (5-HIAA) is the main metabolite of serotonin. Decreased concentrations of this serotonin metabolite have been

consistently reported in cerebrospinal fluid of bipolar depressed patients. It has been reported that the concentration of 5-HIAA in cerebrospinal fluid of depressed subjects, including both bipolar and unipolar subjects, was significantly lower than that of healthy controls [48]. The concentration of 5-HIAA in cerebrospinal fluid has also been found to be decreased in bipolar depression subjects with suicide attempts when compared with controls [49]. It is interesting that 5-HIAA levels of cerebrospinal fluid were significantly increased in female manic patients when compared with gender matched controls [50]. Decreased 5-HIAA levels in depression, and increased 5-HIAA levels in mania, suggest an important relevance of 5-HIAA in manic and depressive symptoms of bipolar disorder. In order to determine if monoamine metabolites are critical to lethality of suicide attempts in bipolar patients, Sher et al. [51] recently using high-performance liquid chromatography to analyze the relationship between 5-HIAA, dopamine metabolite homovanillic acid (HVA) and norepinephrine metabolite 3-methoxy-4-hydroxyphenylglycol (MHPG) levels in cerebrospinal fluid, and maximum lethality of suicide attempts during a 2-year follow up in 27 bipolar depressed patients. They found that 5-HIAA, HVA, and MHPG levels were negatively correlated with the maximum lethality of suicide attempts during this follow-up period. Decreased levels of monoamine metabolites have also been found in postmortem brain tissue in subjects with bipolar depression. Young et al. [52] found that in bipolar disorder subjects who died while depressed, 5-HIAA levels were decreased in postmortem frontal and parietal cortex, and 5-HIAA/serotonin ratio was decreased in postmortem temporal cortex, while HVA levels were decreased in postmortem parietal cortex and HVA/dopamine ratios were decreased in postmortem occipital cortex when compared with controls. These studies suggest that decreased production or increased removal of 5-HIAA in cerebrospinal fluid and brain play an important role in bipolar depression, and also suggest that levels of monoamine metabolites may be used as predictors of lethality of suicide attempts in subjects with bipolar disorder.

Serotonin transporter in presynaptic terminals reuptakes the neurotransmitter serotonin from synaptic cleft into presynaptic neurons, and is a key molecule for control of synaptic serotonin levels. Serotonin transporter is also primary target for many antidepressants [53, 54]. A number of studies indicate abnormal serotonin transporter activity in subjects with bipolar depression. Among these studies, most reported that the serotonin transporter activity is decreased in this disorder, but some also reported that the serotonin transporter activity is not changed or increased.

Serotonin transporter activity has been measured by analyzing the binding of selective serotonin reuptake inhibitor with the transporter uptake site. Leake et al. [55] studied 15 subjects with depression, 9 with major depressive disorder and 6 with bipolar disorder, and found that the concentration of  ${}^{3}$ H-citalopram binding to serotonin transporter was lower in postmortem frontal cortex from depressed subjects when compared with controls. They also found that serotonin uptake sites were attenuated in a trend in subjects with bipolar depression. Human platelets are able to uptake serotonin and resemble presynaptic serotoninergic neurons [56], which provide a convenient model to investigate involvement of the serotonin system in the pathophysiology of mood disorders. Stahl et al. [57] found that serotonin uptake was significantly reduced in subjects with bipolar depression, but not in subjects with schizophrenia when compared to controls. It has also been found that the maximum rate for serotonin reuptake into platelets was reduced in subjects with bipolar depression and unipolar depression [58–60]. However, serotonin transporter activity in platelets has also been reported to be no change between subjects with controls and bipolar depression [61]. Although no difference of serotonin transporter activity between subjects with depression and healthy controls has also been found, serotonin transporter activity has been found to be significantly lower in subjects with bipolar depression than unipolar depression [62]. A difference in this transporter activity between subjects with bipolar depression and unipolar depression may be useful to classify clinically defined subtypes of depression in affective disorders.

Decreased serotonin transporter activity in bipolar depression was also confirmed by a study using positron emission tomography. Oquendo et al. [63] recently studied brain serotonin transporter binding using positron emission tomography and radiolabeled *trans*-1,2,3,5,6,10-β-hexahydro-6-[4-(methylthio) phenyl] pyrrolo-[2,1-a]-isoquinoline  $({}^{11}C(+)$ -McN5652) in 18 medicationfree patients with bipolar depression compared with 41 controls. They found that serotonin transporter binding activity was reduced by 16% to 26% in the midbrain, amygdala, hippocampus, thalamus, putamen, and anterior cingulate cortex of patients with bipolar depression (Fig. 3). Using the same technology, serotonin transporter binding activity has also been found to be decreased in subjects with unipolar depression [64–66]. Low serotonin transporter binding activity may represent serotonin transporter internalization that is facilitated by decreased serotonin levels in synaptic cleft in subjects with bipolar depression. Because serotonin transporter binding has been used as is a marker for serotoninergic neurons, low serotonin transporter binding activity may also indicate fewer serotoninergic neurons in subjects with bipolar depression [63, 67, 68].  $\left[ {}^{11}C \right]$ -3-amino-4-(2-dimethylaminomethyl-phenylsulfanyl)-benzonitrile ( $\left[ {}^{11}C \right]$ DASB), a recently developed diphenyl sulfide radioligands, is able to higher selectively bind serotonin transporter *in vivo* than previously available radioligands. Cannon et al. [69, 70] analyzed the serotonin transporter binding potential in 18 unmedicated subjects with bipolar depression and 37 healthy controls using positron emission tomography and  $\lceil$ <sup>11</sup>C $\rceil$  DASB. They found that the serotonin binding activity was decreased in pontine raphe-nuclei of the brainstem, but increased in the thalamus, dorsal cingulate cortex, medial prefrontal cortex and insula when compared with controls. They also found that serotonin transporter binding was lower in the midbrain and higher in anterior cingulate cortex of bipolar subjects with a history of suicide attempts when compared to controls or bipolar subjects without suicide attempts. It is interesting that subjects with bipolar depression have shown lower serotonin transporter binding poten-



Figure 3. Lower radiolabeled trans-1,2,3,5,6,10-hexahydro-6-[4-(methylthio)-phenyl]pyrrolo-[2,1-a] isoquinoline  $([<sup>11</sup>C](+)$ -McNeil 5652) binding potential (BP1) to the serotonin transporter (5-HTT) in nonmedicated patients with bipolar disorder in six regions of interest compared with controls  $(F1,58 = 5.41; P = .02)$ . ACC indicates anterior cingulate cortex; AMY, amygdala; HIP, hippocampus; MID, midbrain; PUT, putamen; and THA, thalamus. Vertical bars represent the mean likelihood approach to the graphical method modeled BP1 (VT[region] – VT[cerebellum]) for each region; error bars,  $\pm 1$  SD [63].

tial in pontine raphe-nuclei region than subjects with unipolar depression and controls, while subjects with unipolar depression have shown higher serotonin binding potential in periaqueductal gray of middle brain than subjects with bipolar and controls. This difference of serotonin binding potential in brainstem between unipolar depression and bipolar depression may be relevant to patterns of illness symptoms and pharmacological sensitivity for these disorders.

In summary, the levels of the major metabolite of serotonin 5-HIAA may be decreased in cerebrospinal fluid and brain of subjects with bipolar depression, suggesting that intra-synaptic serotonin levels are low in this disorder. The findings of serotonin transporter binding activity in bipolar depression however are inconsistent. Most studies have suggested that serotonin transporter binding activity is decreased in bipolar depression, while others have reported that serotonin transporter binding activity is increased. The inconsistency may result from small sample sizes, differing ligands, confounding medications, and differing reference tissues, among other reasons.

Serotonin reuptake appears to be dysregulated in bipolar depression quite similar to which is seen in unipolar depression [71]. Many studies suggest that

BDNF regulates the development and function of serotoninergic neurons [72–75]. Recent studies have also shown that administration of selective serotonin reuptake inhibitor antidepressants increase gene expression of BDNF [76–80]. As described above, decreased BDNF levels have also been shown in bipolar disorder. Since both serotoninergic system and BDNF play an important role in brain development via regulation of neurite outgrowth, synaptogenesis, and cell survival, dysregulation in serotoninergic system and BDNF imply impaired neuroplasticity in bipolar depression, resulting in the cell loss and damage found in brain imaging and *post mortem* brain studies.

## **Abnormal energy metabolism and dysfunctional mitochondria**

While the factors that cause or promote cell loss and change in brain of patients with bipolar disorder, ultimately leading to altered neurotransmission, remain to be fully established, altered energy metabolism resulting in oxidative stress may be one important contributing factor. Mitochondria are enriched in both dendrites and synaptic terminals, and play important roles in neuroplasticity and cell viability via energy production. Therefore, abnormal regulation of serotonin system in bipolar depression may result from impaired neuroplasticity induced by abnormal energy metabolism and dysfunctional mitochondria.

Metabolic pathways for energy production in humans include glycolysis, the citric acid cycle and the electron transport chain. The primary source for energy production in the brain is glucose. During the glycolysis process, one molecule of glucose is converted into two molecules of pyruvate that are then used for making more molecules of ATP in the citric acid cycle and in the electron transport chain in mitochondria [81, 82]. A number of studies have indicated presence of impaired energy metabolism in bipolar depression. Baxter et al. [83] using positron emission tomography and fluorodeoxyglucose F18 in a small group of subjects (11 with unipolar depression, 5 with bipolar depression, 5 with mania, 3 with bipolar mixed states and 9 normal controls) found that subjects with bipolar depression have significant lower cerebral metabolic rates for glucose when compared with other groups. The ranking for glucose metabolic rates in the whole brain from low to high are bipolar depression, unipolar depression or a mixed state, manic state respectively. Later this research group [84] also analyzed cerebral glucose metabolism in a larger group of subjects with depression that included bipolar depression, unipolar depression and obsessive-compulsive disorder with secondary depression. They found that the glucose metabolic rate was significantly lower in subjects with bipolar depression and unipolar depression than in controls in the left dorsal anterolateral prefrontal cortex. The depressive subjects with medication showed increased glucose metabolic rates in this brain region, and the percentage of change was correlated with the Hamilton Rating Scale for Depression score. The rate of glucose metabolism was also found to be

decreased in the prefrontal cortex of subjects with familial bipolar depression and familial unipolar depression, but increased in subjects with mania [11]. However, it has been also reported that the rate of glucose metabolism was higher in subjects with either unipolar depression or bipolar depression than in healthy controls in the left amygdale [85]. These studies suggested that change of glucose metabolism represents changes in mood state, has brain regional difference and contributes a significant role in mood disorders. Glucose metabolic process in brain may also be targeted by pharmacological treatment.

Efficient cerebral blood flow is required to meet the brain's metabolic demands. Studies have shown lower cerebral blood flow in subjects with bipolar depression. Ito et al. [86] using  $^{99m}$ Tc-hexamethylpropyleneamineoxime as a cerebral blood flow tracer for single photon emission computed tomography scan in 11 patients with unipolar depression, 6 patients with bipolar depression and 9 age-matched normal control subjects. They found that cerebral blood flow was significantly decreased in the prefrontal cortices, limbic systems and paralimbic areas of subjects with unipolar depression and bipolar depression, and there was no difference observed between these two types of depression. Using positron emission tomographic images, Drevets et al. [11] also found that cerebral blood flow was decreased in the prefrontal cortex of subjects with unipolar depression and bipolar depression. Rubin et al. [87] analyzed young adults in episodes of either acute mania or major depression. They found that cerebral blood flow was significantly lower in subjects with mania and depression than in controls in anterior cortical areas. In contrast to these studies, Tutus et al. [88] found that cerebral blood flow was decreased only in the left frontal lobe of subjects with unipolar depression but not bipolar depression when compared to subjects with and controls. Recently, Krüger et al. [89] found that cerebral blood flow was decreased in the orbitofrontal and inferior temporal cortices but increased in the dorsal/rostral anterior cingulate and anterior insula after induction of transient sadness in euthymic lithium responder bipolar patients and their healthy siblings. These findings suggest that energy metabolism is decreased in depressive mood states, but clear understanding of cerebral blood flow in bipolar depression requires further investigation.

Phosphorus-31 magnetic resonance spectroscopy allows detecting highenergy phosphates and the intracellular pH [90, 91]. Using this technology, Kato et al. [92, 93] found that subjects with bipolar disorder have shown increased phosphomonoester and intracellular pH in brain in the depressive mood state when compared with the euthymic state, while those values in the euthymic state were significantly lower as compared to age-matched normal controls. Increased phosphomonoester has also been found in the frontal lobe of subjects with unipolar depression when compared to age-matched controls. Phosphomonoester levels are correlated negatively with the degree of depression [94]. These studies suggest that disturbed phospholipid and intracellular high-energy phosphate metabolism may be common between bipolar depression and unipolar depression. Phosphocreatine is a high-energy compound

made from creatine and ATP. It has been found that phosphocreatine in brain was significantly decreased in subjects with Bipolar II in all hypomanic, euthymic and depressive states when compared to controls. Phosphocreatine was significantly lower in subjects with severe depression than subjects with mild depression [92, 95]. Further, the phosphocreatine levels were significantly lower in the left frontal lobe of subjects with bipolar depression than the normal controls, while phosphocreatine levels were lower in the right frontal lobe of the subjects in the manic and the euthymic states than controls. Phosphocreatine levels in subjects with bipolar depression were negatively correlated with the Hamilton Rating Scale for Depression score [96]. Decreased ATP values have also been found in the frontal lobe of subjects with unipolar depression [94]. It is interesting that creatine levels were significantly lower in the right frontal lobe of the female subjects with bipolar disorder than male subjects [97]. These results suggest that bipolar depression and unipolar depression are associated with abnormal metabolisms of high energy phosphate. Abnormal energy metabolism in brain may be closely related to mood state of mood disorders and is also gender dependent. Frye et al. [98] used proton magnetic resonance spectroscopy to scan the anterior cingulate/medial prefrontal cortex and found that levels of glutamate + glutamine, glutamate and creatine + phosphocreatine were significantly higher in subjects with bipolar depression than healthy controls. Dager et al. [99], using twodimensional proton echo-planar spectroscopic imaging, analyzed 32 medication-free outpatients with bipolar disorder predominantly in a depressed or mixed-mood state, and found that subjects with bipolar disorder have shown increased lactate levels and increased levels of total glutamate, glutamine and gamma-aminobutyric acid in gray matter (Tab. 1). Abnormal metabolism of high-energy phosphates and increased gray matter lactate suggest that bipolar disorder may be associated with mitochondrial dysfunction that induces an energy production shift from more efficient oxidative phosphorylation in mitochondrial electron transport chain to less efficient anaerobic glycolysis.

Mitochondrial energy production is produced via the process of oxidative phosphorylation coupled to the electron transport chain complex I–V (complex I, NADH coenxyme Q reductase; complex II, succinate dehydrogenase; complex III, coenzyme Q cytochrome c reductase; complex IV, cytochrome *c* oxidase; and complex V, ATP synthase). Studies indicate increased deletion and mutation in genes coding for complex I subunits in subjects with bipolar [100–102]. Recent DNA microarray analysis in *post mortem* frontal cortex and hippocampus revealed that expression of many mRNAs, coding for subunits of complex I–V, is decreased in subjects with bipolar disorder [103, 104]. Recent genotyping studies also suggest that polymorphisms of complex I subunit NDUFV2 are associated with bipolar disorder [105, 106]. Complex I is one of the main sites where electrons are leaked to oxygen, resulting in ROS production [107]. Abnormal function of ETC complex I in BD suggests ROS overproduction that may induce oxidative stress in this disease. We recently found that oxidative damage to lipids is increased in *post mortem* cingulate cortex of



Table 1 Brain chemical concentrations by tissue type\* [99]

Abbreviations: Cho, choline-containing compounds; Cre, creatine and phosphocreatine; Glx, glutamate, glutamine, and γ-aminobutyric acid; mI, *myo*-inositol; NAA, *N*-acetyl aspartate \* Values are given as mean (SD)

 $\uparrow$  *P* < .01 vs control

BD subjects [108]. Increased oxidative damage to lipids has also been found in blood samples of BD patients [109, 110]. Additionally, Benes et al. [111] reported that expression of antioxidant enzyme GST A4 and M3 subtypes are reduced in *post mortem* hippocampus from BD subjects. These findings suggest that impairment of energy production and mitochondrial dysfunction in bipolar disorder may induce oxidative stress.

#### **Summary**

Converging lines of study suggest cell loss and damage in key brain regions which may lead to decreased volume of prefrontal and limbic brain regions of subjects with bipolar disorder. Persistent changes in these brain regions may cause disturbance in the function of neuronal circuitry involved in mood regulation, subsequently resulting in impairment of neuroplasticity. These changes may have consequences for monoaminergic neurotransmitters including serotonin, a key intermediary in major depression and a target of antidepressant drugs. Many studies of bipolar depression have shown a decrease in the main metabolite of serotonin 5-HIAA in cerebrospinal fluid and a decrease in serotonin transporter binding activity in brain and platelets.

Mitochondria are enriched in the neuronal terminals and dendrites that form synaptic connections, and its major role is generation of ATP. Mitochondria are important in regulating synthesis, release, uptake and other functions of neurotransmitters, including serotonin, via energy production. Recent evidence has shown decrease of glucose, cerebral blood flow and energy metabolism in bipolar depression, indicating deficiency of energy production in this disease.

Studies also suggest defects of mitochondrial electron transport chain complexes and oxidative damage in bipolar disorder. Mitochondrial electron transport chain is not only a highly efficient way to store energy, but it is also the major resource for generating reactive oxygen species. Therefore mitochondrial dysfunction in bipolar disorder not only reduces energy production, but also causes overproduction of the reactive oxygen species that induce oxidative damage, resulting in impairment of neuroplasticity in specific brain regions related to mood regulation.

The neurobiology of bipolar disorder continues to be unraveled which will help us further understand the multidimensional nature of the illness and the predisposition to both manic and depressive symptoms. It is hard to draw conclusions about the neurobiology of bipolar depression, but in this chapter we have outlined several potential processes which may be relevant to understanding the basis of this phase and illness. Given the limited success of treatment for all phases of bipolar disorder, it seems very likely that further elucidation of the neurobiological mechanisms underlying bipolar depression holds great promise for the development of safe and more effective treatment for this disease.

### **References**

- 1 Isometsa ET, Henriksson MM, Aro HM, Lonnqvist JK (1994) Suicide in bipolar disorder in Finland. *Am J Psychiatry* 151: 1020–1024
- 2 Ferrier IN (1999) Treatment of major depression: is improvement enough? *J Clin Psychiatry* 60 (Suppl 6): 10–14
- 3 López P, Mosquera F, de León J, Gutiérrez M, Ezcurra J, Ramírez F, González-Pinto A (2001) Suicide attempts in bipolar patients. *J Clin Psychiatry* 62: 963–966
- 4 Judd LL, Akiskal HS, Schettler PJ, Endicott J, Maser J, Solomon DA, Leon AC, Rice JA, Keller MB (2002) The long-term natural history of the weekly symptomatic status of bipolar I disorder. *Arch Gen Psychiatry* 59: 530–537
- 5 Post RM, Denicoff KD, Leverich GS, Altshuler LL, Frye MA, Suppes TM, Rush AJ, Keck PE Jr, McElroy SL, Luckenbaugh DA et al (2003) Morbidity in 258 bipolar outpatients followed for 1 year with daily prospective ratings on the NIMH life chart method. *J Clin Psychiatry* 64: 680–690
- 6 Mitchell PB, Malhi GS (2004) Bipolar depression: phenomenological overview and clinical characteristics. *Bipolar Disord* 6: 530–539
- 7 Allman JM, Hakeem A, Erwin JM, Nimchinsky E, Hof P (2001) The anterior cingulate cortex. The evolution of an interface between emotion and cognition. *Ann N Y Acad Sci* 935: 107–117
- 8 Cardinal RN, Parkinson JA, Hall J, Everitt BJ (2002) Emotion and motivation: the role of the amygdala, ventral striatum, and prefrontal cortex. *Neurosci Biobehav Rev* 26: 321–352
- 9 Iidaka T, Terashima S, Yamashita K, Okada T, Sadato N, Yonekura Y (2003) Dissociable neural responses in the hippocampus to the retrieval of facial identity and emotion: an event-related fMRI study. *Hippocampus* 13: 429–436
- 10 Phelps EA (2006) Emotion and cognition: insights from studies of the human amygdala. *Annu Rev Psychol* 57: 27–53
- 11 Drevets WC, Price JL, Simpson Jr, JR, Todd RD, Reich T, Vannier M, Raichle ME (1997) Subgenual prefrontal cortex abnormalities in mood disorders. *Nature* 386: 824–827
- 12 Hirayasu Y, Shenton ME, Salisbury DF, Kwon JS, Wible CG, Fischer IA, Yurgelun-Todd D, Zarate C, Kikinis R, Jolesz FA et al (1999) Subgenual cingulate cortex volume in firstepisode psychosis. *Am J Psychiatry* 156: 1091–1093
- 13 Lopez-Larson MP, DelBello MP, Zimmerman ME, Schwiers ML, Strakowski SM (2002) Regional prefrontal gray and white matter abnormalities in bipolar disorder. *Biol Psychiatry* 52: 93–100
- 14 Frangou S, Hadjulis M, Chitnis X, Baxter D, Donaldson S, Raymont V (2002) The Maudsley Bipolar Disorder Project: brain structural changes in bipolar 1 disorder. *Bipolar Disord* 4: 123–124
- 15 Swayze VW 2nd, Andreasen NC, Alliger RJ, Yuh WT, Ehrhardt JC (1992) Subcortical and temporal structures in affective disorder and schizophrenia: a magnetic resonance imaging study. *Biol Psychiatry* 31: 221–240
- 16 Altshuler LL, Bartzokis G, Grieder T, Curran J, Mintz J (1998) Amygdala enlargement in bipolar disorder and hippocampal reduction in schizophrenia: an MRI study demonstrating neuroanatomic specificity. *Arch Gen Psychiatry* 55: 663–664
- 17 Ali SO, Denicoff KD, Altshuler LL, Hauser P, Li X, Conrad AJ, Smith-Jackson EE, Leverich GS, Post RM (2001) Relationship between prior course of illness and neuroanatomic structures in bipolar disorder: a preliminary study. *Neuropsychiatry Neuropsychol Behav Neurol* 14: 227–232
- 18 Strakowski SM, DelBello MP, Sax KW, Zimmerman ME, Shear PK, Hawkins JM, Larson ER (1999) Brain magnetic resonance imaging of structural abnormalities in bipolar disorder. *Arch Gen Psychiatry* 56: 254–260
- 19 Strakowski SM, Adler CM, Holland SK, Mills NP, DelBello MP, Eliassen JC (2005) Abnormal FMRI brain activation in euthymic bipolar disorder patients during a counting Stroop interference task. *Am J Psychiatry* 162: 1697–1705
- 20 Hauser P, Matochik J, Altshuler LL, Denicoff KD, Conrad A, Li X, Post RM (2000) MRI-based measurements of temporal lobe and ventricular structures in patients with bipolar I and bipolar II disorders. *J Affect Disord* 60: 25–32
- 21 Brambilla P, Harenski K, Nicoletti M, Sassi RB, Mallinger AG, Frank E, Kupfer DJ, Keshavan MS, Soares JC (2003) MRI investigation of temporal lobe structures in bipolar patients. *J Psychiatr Res* 37: 287–295
- 22 Blumberg HP, Kaufman J, Martin A, Whiteman R, Zhang JH, Gore JC, Charney DS, Krystal JH, Peterson BS (2003) Amygdala and hippocampal volumes in adolescents and adults with bipolar disorder. *Arch Gen Psychiatry* 60: 1201–1208
- 23 DelBello MP, Zimmerman ME, Mills NP, Getz GE, Strakowski SM (2004) Magnetic resonance imaging analysis of amygdala and other subcortical brain regions in adolescents with bipolar disorder. *Bipolar Disord* 6: 43–52
- 24 Chen BK, Sassi R, Axelson D, Hatch JP, Sanches M, Nicoletti M, Brambilla P, Keshavan MS, Ryan ND, Birmaher B et al (2004) Cross-sectional study of abnormal amygdala development in adolescents and young adults with bipolar disorder. *Biol Psychiatry* 56: 399–405
- 25 Chang K, Karchemskiy A, Barnea-Goraly N, Garrett A, Simeonova DI, Reiss A (2005) Reduced amygdalar gray matter volume in familial pediatric bipolar disorder. *J Am Acad Child Adolesc Psychiatry* 44: 565–573
- 26 Altshuler LL, Bartzokis G, Grieder T, Curran J, Jimenez T, Leight K, Wilkins J, Gerner R, Mintz J (2000) An MRI study of temporal lobe structures in men with bipolar disorder or schizophrenia. *Biol Psychiatry* 48: 147–162
- 27 Rajkowska G, Halaris A, Selemon LD (2001) Reductions in neuronal and glial density characterize the dorsolateral prefrontal cortex in bipolar disorder. *Biol Psychiatry* 49: 741–752
- 28 Benes FM, Kwok EW, Vincent SL, Todtenkopf MS (1998) A reduction of nonpyramidal cells in sector CA2 of schizophrenics and manic depressives. *Biol Psychiatry* 44: 88–97
- 29 Benes FM, Vincent SL, Todtenkopf M (2001) The density of pyramidal and nonpyramidal neurons in anterior cingulate cortex of schizophrenic and bipolar subjects. *Biol Psychiatry* 50: 395–406
- 30 Bezchlibnyk YB, Sun X, Wang JF, MacQueen GM, McEwen BS, Young LT (2007) Neuron somal size is decreased in the lateral amygdalar nucleus of subjects with bipolar disorder. *J Psychiatry Neurosci* 32: 203–210
- 31 Berretta S, Pantazopoulos H, Lange N (2007) Neuron numbers and volume of the amygdala in subjects diagnosed with bipolar disorder or schizophrenia. *Biol Psychiatry* 62: 884–893
- 32 Dowlatshahi D, MacQueen G, Wang JF, Chen B, Young LT (2000) Increased hippocampal supragranular Timm staining in subjects with bipolar disorder. *Neuroreport* 11: 3775–3778
- 33 Sassi RB, Stanley JA, Axelson D, Brambilla P, Nicoletti MA, Keshavan MS, Ramos RT, Ryan N, Birmaher B, Soares JC (2005) Reduced NAA levels in the dorsolateral prefrontal cortex of young bipolar patients. *Am J Psychiatry* 162: 2109–2115
- 34 Winsberg ME, Sachs N, Tate DL, Adalsteinsson E, Spielman D, Ketter TA (2005) Decreased dorsolateral prefrontal N-acetyl aspartate in bipolar disorder. *Biol Psychiatry* 47: 475–481
- 35 Ongur D, Drevets WC, Price JL (1998) Glial reduction in the subgenual prefrontal cortex in mood disorders. *Proc Natl Acad Sci USA* 95: 13290–13295
- 36 Johnston-Wilson NL, Sims CD, Hofmann JP, Anderson L, Shore AD, Torrey EF, Yolken RH (2000) Disease-specific alterations in frontal cortex brain proteins in schizophrenia, bipolar disorder, and major depressive disorder. The Stanley Neuropathology Consortium. *Mol Psychiatry* 5: 142–149
- 37 Orlovskaya DD, Vikhreva OV, Zimina IS, Denisov DV, Uranova NA (1999) Ultrastructural dystrophic changes of oligodendroglial cells in autopsied prefrontal cortex and striatum in schizophrenia: a morphometric study. *Schizophr Res* 36: 82–83
- 38 Orlovskaya DD, Vostrikov VM, Rachmanova NA, Uranova NA (2000) Decreased numerical density of oligodendroglial cells in postmortem prefrontal cortex in schizophrenia, bipolar affective disorder and major depression. *Schizophr Res* 41: 105–106
- 39 Uranova N, Orlovskaya D, Vikhreva O, Zimina I, Kolomeets N, Vostrikov V, Rachmanova V (2001) Electron microscopy of oligodendroglia in severe mental illness. *Brain Res Bull* 55: 597–610
- 40 Knable MB, Barci BM, Webster MJ, Meador-Woodruff J, Torrey EF (2004) Stanley Neuropathology Consortium. Molecular abnormalities of the hippocampus in severe psychiatric illness: postmortem findings from the Stanley Neuropathology Consortium. *Mol Psychiatry* 9: 609–620
- 41 Neves-Pereira M, Mundo E, Muglia P, King N, Macciardi F, Kennedy JL (2002) The brain-derived neurotrophic factor gene confers susceptibility to bipolar disorder: evidence from a family-based association study. *Am J Hum Genet* 71: 651–655
- 42 Sklar P, Gabriel SB, McInnis MG, Bennett P, Lim YM, Tsan G, Schaffner S, Kirov G, Jones I, Owen M et al (2002) Family-based association study of 76 candidate genes in bipolar disorder: BDNF is a potential risk locus. *Mol Psychiatry* 7: 579–593
- 43 Geller B, Badner JA, Tillman R, Christian SL, Bolhofner K, Cook EH Jr (2004) Linkage disequilibrium of the brain-derived neurotrophic factor Val66Met polymorphism in children with a prepubertal and early adolescent bipolar disorder phenotype. *Am J Psychiatry* 161: 1698–1700
- 44 Lohoff FW, Sander T, Ferraro TN, Dahl JP, Gallinat J, Berrettini WH (2005) Confirmation of association between the Val66Met polymorphism in the brain-derived neurotrophic factor (BDNF) gene and bipolar I disorder *Am J Med Genet B Neuropsychiatr Genet* 139: 51–53
- 45 Cunha AB, Frey BN, Andreazza AC, Goi JD, Rosa AR, Gonçalves CA, Santin A, Kapczinski F (2006) Serum brain-derived neurotrophic factor is decreased in bipolar disorder during depressive and manic episodes. *Neurosci Lett* 398: 215–219
- 46 Monteleone P, Serritella C, Martiadis V, Maj M (2008) Decreased levels of serum brain-derived neurotrophic factor in both depressed and euthymic patients with unipolar depression and in euthymic patients with bipolar I and II disorders. *Bipolar Disord* 10: 95–100
- 47 Tseng M, Alda M, Xu L, Sun X, Wang JF, Grof P, Turecki G, Rouleau G, Young LT (2008) BDNF protein levels are decreased in transformed lymphoblasts from lithium-responsive bipolar disorder subjects. *J Psychiatry Neurosci* 33: 449–453
- 48 Asberg M, Bertillsson L, Martensson B, Scalia-Tomba GP, Thoren P, Traskman-Bendz L (1984) CSF monoamine metabolites in melancholia. *Acta Psychiatr Scand* 69: 201–219
- 49 Träskman L, Asberg M, Bertilsson L, Sjöstrand L (1981) Monoamine metabolites in CSF and suicidal behaviour. *Arch Gen Psychiatry* 38: 631–636
- 50 Swann AC, Secunda S, Davis JM, Robins E, Hanin I, Koslow SH, Maas JW (1983) CSF monoamine metabolites in mania. *Am J Psychiatry* 140: 396–400
- 51 Sher L, Carballo JJ, Grunebaum MF, Burke AK, Zalsman G, Huang YY, Mann JJ,
- Oquendo MA (2006) A prospective study of the association of cerebrospinal fluid monoamine metabolite levels with lethality of suicide attempts in patients with bipolar disorder. *Bipolar Disord* 8: 543–550
- 52 Young LT, Warsh JJ, Kish SJ, Shannak K, Hornykeiwicz O (1994) Reduced brain 5-HT and elevated NE turnover and metabolites in bipolar affective disorder. *Biol Psychiatry* 35: 121–127
- 53 Rudnick G (2006) Serotonin transporters structure and function. *J Membr Biol* 213: 101–110
- 54 White KJ, Walline CC, Barker EL (2005) Serotonin transporters: implications for antidepressant drug development. *AAPS J* 7: E421–433
- 55 Leake A, Fairbairn AF, McKeith IG, Ferrier IN (1991) Studies on the serotonin uptake binding site in major depressive disorder and control *post mortem* brain: neurochemical and clinical correlates. *Psychiatry Res* 39: 155–165
- 56 Sneddon JM (1973) Blood platelets as a model for monoamine containing neurons. *Prog Neurobiol* 1: 151–198
- 57 Stahl SM, Woo DJ, Mefford IN, Berger PA, Ciaranello RD (1983) Hyperserotonemia and platelet serotonin uptake and release in schizophrenia and affective disorders. *Am J Psychiatry* 140: 26–30
- 58 Scott M, Reading HW, Loudon JB (1979) Studies on human blood platelets in affective disorder. *Psychopharmacology (Berl)* 60: 131–135
- 59 Meltzer HY, Arora RC, Baber R, Tricou BJ (1981) Serotonin uptake in blood platelets of psychiatric patients. *Arch Gen Psychiatry* 38: 1322–1326
- 60 Marazziti D, Lenzi A, Cassano GB (1991) Serotonergic dysfunction in bipolar disorder. *Pharmacopsychiatry* 24: 164–166
- 61 Muscettola SA, Dilauro A, Giannini CP (1986) Platelet <sup>3</sup>H-imipramine binding in bipolar patients. *Psychiatry Res* 18: 343–353
- 62 Lewis DA, McChesney C (1985) Tritiated imipramine binding distinguishes among subtypes of depression. *Arch Gen Psychiatry* 42: 485–488
- 63 Oquendo MA, Hastings RS, Huang YY, Simpson N, Ogden RT, Hu XZ, Goldman D, Arango V, Van Heertum RL, Mann JJ et al (2007) Brain serotonin transporter binding in depressed patients with bipolar disorder using positron emission tomography. *Arch Gen Psychiatry* 64: 201–208
- 64 Malison RT, Price LH, Berman R, van Dyck CH, Pelton GH, Carpenter L, Sanacora
- G, Owens MJ, Nemeroff CB, Rajeevan N et al (1998) Reduced brain serotonin transporter availability in major depression as measured by [123I]-2 beta-carbomethoxy-3 beta-(4-iodophenyl)tropane and single photon emission computed tomography. *Biol Psychiatry* 44: 1090–1098
- 65 Newberg AB, Plossl K, Mozley PD, Stubbs JB, Wintering N, Udeshi M, Alavi A, Kauppinen T, Kung HF (2004) Biodistribution and imaging with (123)I-ADAM: a serotonin transporter imaging agent. *J Nucl Med* 45: 834–841
- 66 Parsey RV, Hastings RS, Oquendo MA, Huang YY, Simpson N, Arcement J, Huang Y, Ogden RT, Van Heertum RL, Arango V et al (2006) Lower serotonin transporter binding potential in the human brain during major depressive episodes. *Am J Psychiatry* 163: 48–57
- 67 Ramamoorthy S, Blakely RD (1999) Phosphorylation and sequestration of serotonin transporters differentially modulated by psychostimulants. *Science* 285: 763–766
- 68 Soucy J-P, Lafaille F, Lemoine P, Mrini A, Descarries L (1994) Validation of the transporter ligand cyanoimipramine as a marker of serotonin innervation density in brain. *J Nucl Med* 35: 1822–1830
- 69 Cannon DM, Ichise M, Fromm SJ, Nugent AC, Rollis D, Gandhi SK, Klaver JM, Charney DS, Manji HK, Drevets WC (2006) Serotonin transporter binding in bipolar disorder assessed using [ 11C]DASB and positron emission tomography. *Biol Psychiatry* 60: 207–217
- 70 Cannon DM, Ichise M, Rollis D, Klaver JM, Gandhi SK, Charney DS, Manji HK, Drevets WC (2007) Elevated serotonin transporter binding in major depressive disorder assessed using positron emission tomography and  $\lbrack$ <sup>11</sup>C<sub>J</sub>DASB; comparison with bipolar disorder. *Biol Psychiatry* 62: 870–877
- 71 Stockmeier CA (2003) Involvement of serotonin in depression: evidence from postmortem and imaging studies of serotonin receptors and the serotonin transporter. *J Psychiatr Res* 37: 357–373
- 72 Madhav TR, Pei Q, Zetterstrom TS (2001) Serotonergic cells of the rat raphe nuclei express mRNA of tyrosine kinase B (trkB), the high-affinity receptor for brain derived neurotrophic factor (BDNF). *Brain Res* 93: 56–63
- 73 Djalali S, Höltje M, Grosse G, Rothe T, Stroh T, Grosse J, Deng DR, Hellweg R, Grantyn R, Hörtnagl H et al (2005) Effects of brain-derived neurotrophic factor (BDNF) on glial cells and serotonergic neurones during development. *J Neurochem* 92: 616–627
- 74 Rumajogee P, Madeira A, Verge D, Hamon M, Miquel MC (2002) Up-regulation of the neuronal serotoninergic phenotype *in vitro*: BDNF and cAMP share Trk B-dependent mechanisms. *J Neurochem* 83: 1525–1528
- 75 Hensler JG, Advani T, Monteggia LM (2007) Regulation of serotonin-1A receptor function in inducible brain-derived neurotrophic factor knockout mice after administration of corticosterone. *Biol Psychiatry* 62: 521–529
- 76 Nibuya M, Morinobu S, Duman RS (1995) Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. *J Neurosci* 15: 7539–7547
- 77 Nibuya M, Nestler EJ, Duman RS (1996) Chronic antidepressant administration increases the expression of cAMP response element binding protein (CREB) in rat hippocampus. *J Neurosci* 16: 2365–2372
- 78 Chen B, Dowlatshahi D, MacQueen GM, Wang JF, Young LT (2001) Increased hippocampal BDNF immunoreactivity in subjects treated with antidepressant medication. *Biol Psychiatry* 50: 260–265
- 79 Dwivedi Y, Rao JS, Rizavi HS, Kotowski J, Conley RR, Roberts RC, Tamminga CA, Pandey GN (2003) Abnormal expression and functional characteristics of cyclic adenosine monophosphate response element binding protein in postmortem brain of suicide subjects. *Arch Gen Psychiatry* 60: 273–282
- 80 Karege F, Vaudan G, Schwald M, Perroud N, La Harpe R (2005) Neurotrophin levels in postmortem brains of suicide victims and the effects of antemortem diagnosis and psychotropic drugs. *Brain Res Mol Brain Res* 136: 29–37
- 81 Ferrari R, Pedersini P, Bongrazio M, Gaia G, Bernocchi P, Di Lisa F, Visioli O (1993) Mitochondrial energy production and cation control in myocardial ischaemia and reperfusion. *Basic Res Cardiol* 88: 495–512
- 82 Hertz L, Kala G (2007) Energy metabolism in brain cells: effects of elevated ammonia concentrations. *Metab Brain Dis* 22: 199–218
- 83 Baxter L, Phelps ME, Mazziotta JC, Schwartz JM, Gerner RH, Selin CE, Sumida RM (1985) Cerebral metabolic rates for glucose in mood disorders. Studies with positron emission tomography and fluorodeoxyglucose F 18. *Arch Gen Psychiatry* 42: 441–447
- 84 Baxter LR, Schwartz JM, Phelps ME, Mazziotta JC, Guze BH, Selin CE, Gerner RH, Sumida RM (1989) Reduction of prefrontal cortex glucose metabolism common to three types of depression. *Arch Gen Psychiatry* 46: 243–250
- 85 Drevets WC, Price JL, Bardgett ME, Reich T, Todd RD, Raichle ME (2002) Glucose metabolism in the amygdala in depression: relationship to diagnostic subtype and plasma cortisol levels*. Pharmacol Biochem Behav* 71: 431–447
- 86 Ito H, Kawashima R, Awata S, Ono S, Sato K, Goto R, Koyama M, Sato M, Fukuda H (1996) Hypoperfusion in the limbic system and prefrontal cortex in depression: SPECT with anatomic standardization technique. *J Nucl Med* 37: 410–414
- 87 Rubin E, Sackeim HA, Prohovnik I, Moeller JR, Schnur DB, Mukherjee S (1995) Regional cerebral blood flow in mood disorders: IV. Comparison of mania and depression. *Psychiatry Res* 61:  $1 - 10$
- 88 Tutus A, Simsek A, Sofuoglu S, Nardali M, Kugu N, Karaaslan F, Gönül AS (1998) Changes in regional cerebral blood flow demonstrated by single photon emission computed tomography in depressive disorders: comparison of unipolar *versus* bipolar subtypes. *Psychiatry Res* 83: 169–177
- 89 Krüger S, Alda M, Young LT, Goldapple K, Parikh S, Mayberg HS (2006) Risk and resilience markers in bipolar disorder: brain responses to emotional challenge in bipolar patients and their healthy siblings. *Am J Psychiatry* 163: 257–264
- 90 Moon R, Richards JH (1973) Determination of intracellular pH by 31P magnetic resonance. *J Biol Chem* 25: 7276–7278
- 91 Petroff OAC, Prichard JW, Behar KL, Alger JR, den Hollander JA, Shulman RG (1985) Cerebral intracellular pH by 31P nuclear magnetic resonance spectroscopy. *Neurology* 35: 781–788
- 92 Kato T, Takahashi S, Shioiri T, Inubushi T (1992) Brain phosphorous metabolism in depressive disorders detected by phosphorus-31 magnetic resonance spectroscopy. *J Affect Disord* 26: 223–230
- 93 Kato T, Shioiri T, Murashita J, Hamakawa H, Inubushi T, Takahashi S (1994a) Phosphorus-31 magnetic resonance spectroscopy and ventricular enlargement in bipolar disorder. *Psychiatry Res* 55: 41–50
- 94 Volz HP, Rzanny R, Riehemann S, May S, Hegewald H, Preussler B, Hübner G, Kaiser WA, Sauer H (1998) 31P magnetic resonance spectroscopy in the frontal lobe of major depressed patients*. Eur Arch Psychiatry Clin Neurosci* 248: 289–295
- 95 Kato T, Takahashi S, Shioiri T, Murashita J, Hamakawa H, Inubushi T (1994b) Reduction of brain phosphocreatine in bipolar II disorder detected by phosphorus-31 magnetic resonance spectroscopy. *J Affect Disord* 31: 125–133
- 96 Kato T, Shioiri T, Murashita J, Hamakawa H, Takahashi Y, Inubushi T, Takahashi S (1995) Lateralized abnormality of high energy phosphate metabolism in the frontal lobes of patients with bipolar disorder detected by phase-encoded 31P-MRS. *Psychol Med* 25: 557–566
- 97 Hamakawa H, Kato T, Shioiri T, Inubushi T, Kato N (1999) Quantitative proton magnetic resonance spectroscopy of the bilateral frontal lobes in patients with bipolar disorder. *Psychol Med* 29: 639–644
- 98 Frye MA, Watzl J, Banakar S, O'Neill J, Mintz J, Davanzo P, Fischer J, Chirichigno JW, Ventura J, Elman S et al (2007) Increased anterior cingulate/medial prefrontal cortical glutamate and creatine in bipolar depression. *Neuropsychopharmacology* 32: 2490–2499
- 99 Dager SR, Friedman SD, Parow A, Demopulos C, Stoll AL, Lyoo IK, Dunner DL, Renshaw PF (2004) Brain metabolic alterations in medication-free patients with bipolar disorder. *Arch Gen Psychiatry* 61: 450–458
- 100 Kato T, Kunugi H, Nanko S, Kato N (2001) Mitochondrial DNA polymorphisms in bipolar disorder. *J Affect Disord* 62: 151–164
- 101 Kato T, Kunugi H, Nanko S, Kato N (2000) Association of bipolar disorder with the 5178 polymorphism in mitochondrial DNA. *Am J Med Genet* 96: 182–186
- 102 Munakata K, Tanaka M, Mori K, Washizuka S, Yoneda M, Tajima O, Akiyama T, Nanko S, Kunugi H, Tadokoro K et al (2004) Mitochondrial DNA 3644 T–>C mutation associated with bipolar disorder. *Genomics* 84: 1041–1050
- 103 Sun X, Wang JF, Tseng M, Young LT (2006) Down regulation in components of mitochondrial electron transport chain in *post mortem* frontal cortex from subjects with bipolar disorder. *J Psychiatry Neurosci* 31: 189–196
- 104 Konradi C, Eaton M, MacDonald ML, Walsh J, Benes FM, Heckers S (2004) Molecular evidence for mitochondrial dysfunction in bipolar disorder. *Arch Gen Psychiatry* 61: 300–308
- 105 Washizuka S, Kakiuchi C, Mori K, Kunugi H, Tajima O, Akiyama T, Nanko S, Kato T (2003) Association of mitochondrial complex I subunit gene NDUFV2 at 18p11 with bipolar disorder. *Am J Med Genet B Neuropsychiatr Genet* 120: 72–78
- 106 Washizuka S, Iwamoto K, Kazuno AA, Kakiuchi C, Mori K, Kametani M, Yamada K, Kunugi H, Tajima O, Akiyama T et al (2004) Association of mitochondrial complex I subunit gene NDUFV2 at 18p11 with bipolar disorder in Japanese and the National Institute of Mental Health pedigrees. *Biol Psychiatry* 56: 483–489
- 107 Halliwell B (1992) Reactive oxygen species and the central nervous system. *J Neurochem* 59: 1609–1623
- 108 Wang JF, Shao L, Sun X, Young LT (2007) Increased lipid peroxidation in postmortem cingulate cortex from subjects with bipolar disorder and schizophrenia. *Soc Neurosci Abstr* 33, Program No. 707.11
- 109 Kuloglu M, Ustundag B, Atmaca M, Canatan H, Tezcan AE, Cinkilinc N (2002) Lipid peroxidation and antioxidant enzyme levels in patients with schizophrenia and bipolar disorder. *Cell Biochem Funct* 20: 171–175
- 110 Ranjekar PK, Hinge A, Hegde MV, Ghate M, Kale A, Sitasawad S, Wagh UV, Debsikdar VB, Mahadik SP (2003) Decreased antioxidant enzymes and membrane essential polyunsaturated fatty acids in schizophrenic and bipolar mood disorder patients. *Psychiatry Res* 121: 109–122
- 111 Benes FM, Matzilevich D, Burke RE, Walsh J (2006) The expression of proapoptosis genes is increased in bipolar disorder, but not in schizophrenia. *Mol Psychiatry* 11: 241–251