

John A. Abraham, Brian Neuman, and Francis J. Hornicek

Contents

17.1	Introduction and Historical Perspective	277
17.2	Epidemiology	278
17.3	Clinical Features	278
17.4	Tumor Imaging	280
17.5	Histopathology and Immunohistochemistry	281
17.6	Differential Diagnosis	282
17.7	Tumor Staging	282
17.8	Management	282
17.8.1	Surgery	282
17.8.2	Radiation	283
17.8.3	Systemic Therapy and Future Directions	284
17.9	Summary of Critical Concepts Discussed in This Chapter	286
	References	286

J.A. Abraham, MD (✉)
Division of Orthopedic Oncology, Rothman Institute,
Philadelphia, PA, USA

Department of Orthopedic Surgery and Radiation Oncology,
Thomas Jefferson University,
925 Chestnut Street, Philadelphia, PA 19107, USA
e-mail: john.abraham@rothmaninstitute.com

B. Neuman, MD
Department of Orthopedic Surgery,
Thomas Jefferson University,
925 Chestnut Street, Philadelphia, PA 19107, USA
e-mail: bneumy@gmail.com

F.J. Hornicek, MD, PhD
Division of Orthopedic Oncology,
Center for Sarcoma and Connective Tissue Oncology,
Massachusetts General Hospital Cancer Center,
Massachusetts General Hospital,
Boston, MA, USA

Department of Orthopedic Surgery,
Harvard Medical School,
55 Fruit Street, Boston, MA 02114, USA
e-mail: fhornicek@partners.org

17.1 Introduction and Historical Perspective

Chordoma is the most common primary malignant bone tumor found in the spine and sacrum. While these tumors are relatively slow growing, they do have the potential to recur and metastasize. Chordoma was described histologically long before it was realized that they were probably derived from notochordal precursor cells. Early reports dating back to Virchow in 1857 describe a vacuolated cell type seen in these tumors. These cells were described as physaliferous, from Greek for “having bubbles.” It was thought at that early time that they were a cartilaginous tumor, which may have been a result of evaluation of a chondroid variant tumor. By 1923, Burrow and Stewart recognized that chordomas were a “lowly malignant tumor of slow growth, locally invasive and destructive, and only rarely giving rise to metastases.” By then, the location at either end of the spine correlated well with contemporary descriptions of the location of vestigial notochordal remnants by Muller. These observations led to the hypothesis that chordomas were not tumors of the intervertebral disc but rather malignant transformation of these notochordal remnants. In 1858, Muller coined the current name by proposing the following hypothesis: “A direct relation of these growths to the chorda dorsalis cannot be overlooked and I consider them to be excessively growing remnants of the chorda. Whosoever likes the name may designate these masses as chordoid tumors, or chordomas.” Since these early descriptions, this concept has been supported by significant indirect evidence, although there is a paucity of direct proof.

The early treatment of chordoma focused on surgical removal, but the difficulty in completely resecting these tumors was very quickly recognized. Investigation of the addition of radiation to the treatment regimen with or without surgery was described in the 1970s and continues to be investigated (Pearlman and Friedman 1970; Pearlman et al. 1972). Medical therapies failed to provide any significant benefit, and the search for an effective medical

agent continues today. This chapter provides a review of the clinical features of chordoma and the current molecular understanding of its pathobiology.

17.2 Epidemiology

Chordoma is the most frequent primary bone tumor found in the spine. Nevertheless, these tumors are rare with the age-adjusted incidence rate of 0.08 per 100,000 (McMaster et al. 2001). Chordomas make up 1–4 % of all primary bone tumors (Healey and Lane 1989; Unni 1996; Papagelopoulos et al. 2004). Approximately 50 % of these tumors are located within the sacrum (Fig. 17.1). The remaining anatomic locations are the skull-base, spheno-occipital region (35 %), and mobile spine (15 %) (Bohlman et al. 1986; Bjornsson et al. 1993; Bergh et al. 2000). The distribution of these tumors within the mobile spine was evaluated by Boriani et al. (2006) in a consecutive series over 50 years. This group demonstrated the highest frequency of involvement in the lumbar spine (57.5 %), followed by the cervical region (29 %), and least frequently in the thoracic spine (13.5 %). Chordomas comprise greater than 50 % of the primary bone tumors found in the sacrum (Boriani et al. 2006). In a recent analysis of 409 patients identified utilizing the California Cancer Registry, the racial distribution was 65 % Caucasian, 23 % Hispanic, 10 % Asian or “other,” and 1.7 % African. In this study, in evaluating chordoma-specific survival, there was a significantly decreased risk of death in Hispanics (Lee et al. 2012). In this group, chordomas were also found to be associated with younger age at diagnosis, cranial disease, and a higher rate of surgical intervention, which were all associated

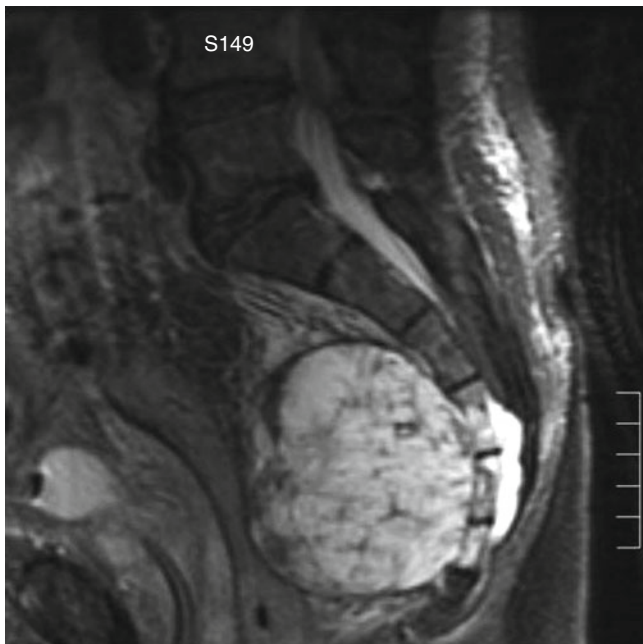


Fig. 17.1 Typical MRI appearance of large sacral chordoma

with better survival. This observation was surprising as the Hispanic population was associated with low socioeconomic status, and this factor alone should have increased the risk of death. Consistently, studies on gender distribution of chordomas show a 2:1 male predominance (Ashwood et al. 1994; Forsyth et al. 1993). The median age at the time of diagnosis is 58.5 years, while diagnosis is rare in patients younger than 30 years of age (McMaster et al. 2001; Weber and Sim 2002). These tumors are very uncommon in pediatric populations comprising less than 5 % of all chordomas, with the majority of these being skull based (McMaster et al. 2001).

17.3 Clinical Features

Patients diagnosed with a chordoma most commonly present with pain, regardless of location (Bergh et al. 2000; Boriani et al. 1996, 2006). The second most common presentation is development of neurological symptoms and least commonly a palpable mass (Bergh et al. 2000, Soo 2001). Symptom duration averages 2 years prior to diagnosis, highlighting the slow-growing nature of these tumors (Bergh et al. 2000). If untreated, pain can progress to the point of incapacitation, which in one study was found to be at approximately 50 months from the onset of symptoms (Boriani et al. 2006).

Up to 60 % of the time, chordomas can extend into the spinal canal and in some of these cases can cause significant neurologic symptoms, such as compressive myelopathy or cauda equina (Meyer et al. 1984) (Fig 17.2). Neurological symptoms are most commonly associated with chordomas of



Fig. 17.2 Large cervical spine chordoma at C2 with compression of spinal cord

the mobile spine. The spectrum of neurological symptoms widely ranges depending on the location of the tumor. Severe compression of the spinal cord is a late complication leading to paralysis. Tumors which invade the neural foramina can cause a radicular pattern of symptoms, which includes weakness and sensory deficits in the distribution of a particular nerve root (Sundaresan et al. 1990; Mindell 1981).

Non-neurologic symptoms from chordoma are most commonly due to local effects of the tumor mass. For example, chordomas which develop in the cervical spine may cause

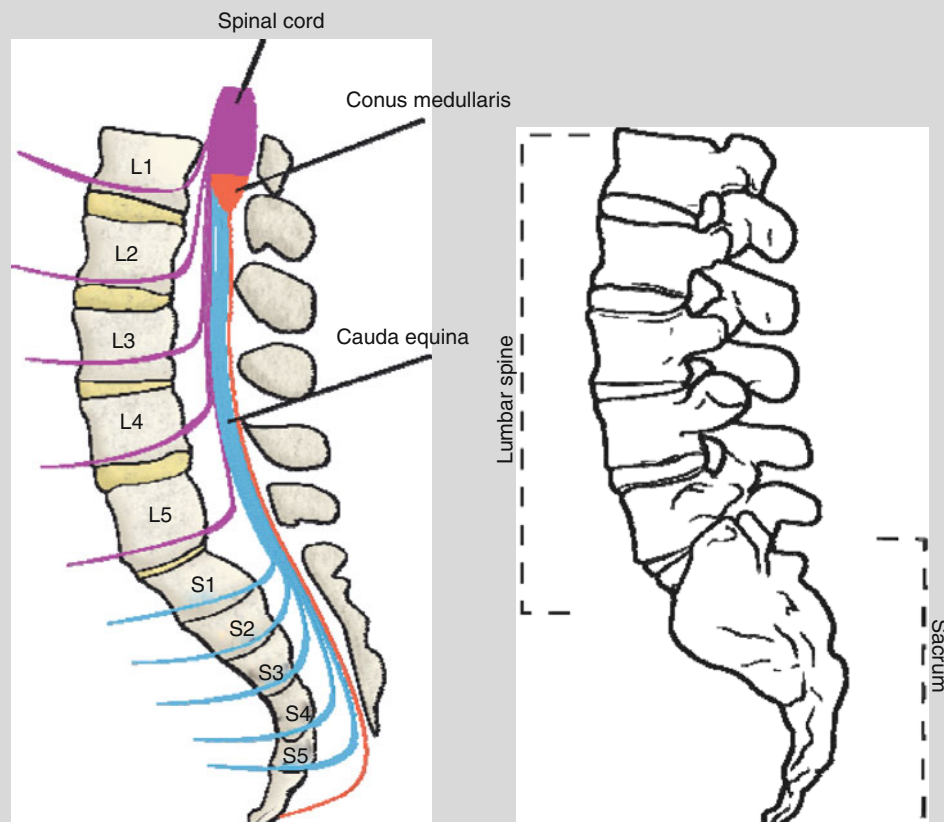
throat irritation, dysphagia, esophageal compression, dysphonia, or airway obstruction from compression of critical local structures (Singh et al. 2007; Nicoucar et al. 2008). Horner syndrome has been reported from lower cervical spine chordomas (Leone et al. 2002). Mass effect from of sacral chordomas can cause compression and displacement of the bladder or rectum leading to urinary stress incontinence, constipation, or obstruction. Sacral chordoma of sufficiently large size can be palpated on rectal exam (Fourney and Gokaslan 2003; Atalar et al. 2006).

Box 17.1 The Cauda Equina

Anatomically, the cauda equina, or “horse’s tail,” is the collection of nerve roots that travels through the spinal canal beyond the termination of the spinal cord. The spinal cord ends at approximately the L1 level in humans, at which point the lumbar (L1–5) and sacral (S1–5) roots have already branched off the spinal cord. However, because these nerve roots exit the spinal canal at successively more inferior levels in the lumbar and sacral bony spine, they travel together for a distance within the spinal canal as a bundle of nerve roots, which is called the cauda equina. Injuries or other conditions which damage the cauda equina cause a specific constellation of symptoms related to dysfunction of

these critical nerve roots and is considered a surgical emergency.

Symptoms of cauda equina syndrome from any cause include weakness in the lower extremities, urinary retention due to detrusor muscle weakness, loss of rectal tone due to anal sphincter weakness, and subsequent fecal incontinence. Sexual dysfunction and saddle anesthesia and lower extremity pain may also be present. Lower extremity reflexes are reduced or absent. The causes of cauda equina syndrome are numerous and essentially can be any inciting agent or problem which causes pressure on the cauda equina. Commonly, acutely herniated discs can cause cauda equina syndrome. Other degenerative spinal conditions such as spinal stenosis



The cauda equina is shown schematically on the *left*, and the osseous anatomy of the lumbar spine and sacrum is shown on the *right*

or spondylolisthesis can contribute as well. Trauma is another common cause, either by direct damage to nerve roots by fracture, dislocation, or penetrating trauma or by hematoma secondary to the initial traumatic injury. Tumors, such as chordoma or more commonly metastatic disease, can also cause a cauda equina syndrome, although in this situation the presenting symptoms are usually more chronic and develop over a longer period of time as the tumor grows.

Treatment of cauda equina syndrome is primarily surgical decompression of the involved nerve roots as

well as removal or correction of the inciting mechanical problem. In the case of degenerative and traumatic conditions, every effort is made to preserve the involved nerve roots. In the case of metastatic tumors, intralesional procedures may be performed in conjunction with adjuvant therapies such as radiation. Sacral chordoma requires wide resection, which usually involved resection of some or all of the sacral nerve roots, so the expectation is that the nerve deficits will not recover. It is important to discuss the specific expected deficits with any patient undergoing sacrectomy for chordoma.

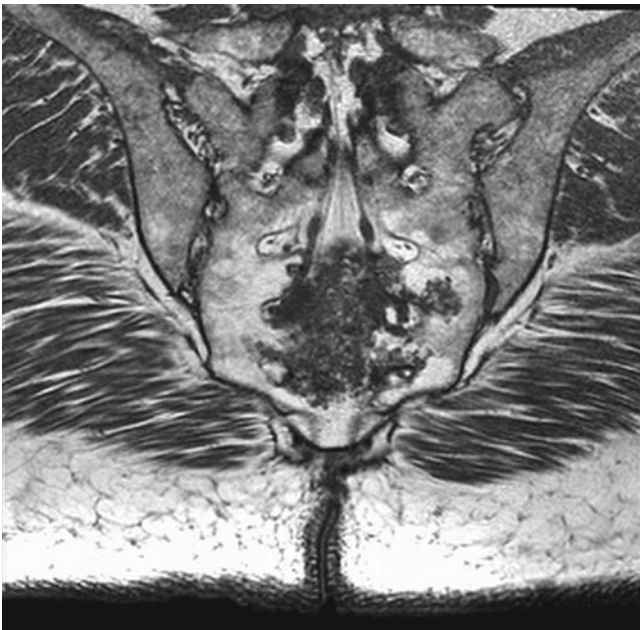


Fig. 17.3 Coronal MRI image of sacral chordoma showing extensive calcifications, sometimes seen in chordoma

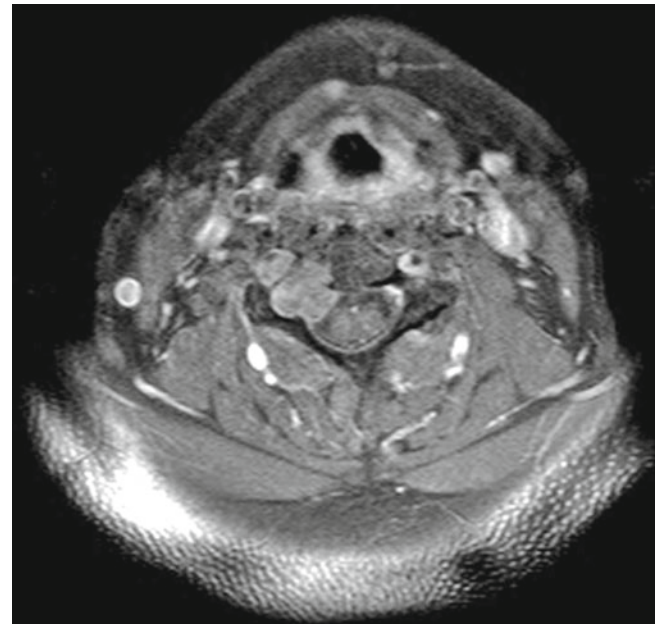


Fig. 17.4 Typical MRI appearance of cervical spine chordoma, in this case involving right-sided pedicle

17.4 Tumor Imaging

Plain radiographs are often the initial image taken for a complaint of back pain. However, detailed examination of the spine with plain imaging alone can be difficult. In particular, the obliquity of the sacrum and the overlying shadows from bowel gas and contents can often limit its utility (Manaster and Graham 2003). Nonetheless, when evaluating the sacrum on plain radiographs, there are particular features that should be scrutinized: the paired sacral foramen should appear similar with distinct sacral boundaries outlining the foramen, the anterior and posterior aspects of the sacroiliac joint should be distinct, and the posterior contour of the iliac wing should be seen underlying the sacral ala. Lack of any of these findings on pelvic radiograph could suggest a lesion of the

sacrum. Intratumoral amorphous calcifications are seen on plain radiographs within a chordoma in 50–70 % of cases and in 90 % of cases on CT scan, but have no known prognostic significance (Fig 17.3). An associated large soft tissue mass can also be seen on CT scan or MRI.

Evaluation of the central spinal canal and cord or nerve root involvement is best done with MR imaging, and as such this modality is critical for evaluation of chordomas. MRI features of a chordoma on T1-weighted sequences consist of an isointense or hypointense mass as compared to muscle and on a T2-weighted sequences as a high-signal-intensity mass (Fig 17.4). If calcifications are present, they can appear as areas of low signal intensity on T1- and T2-weighted images. Chordomas enhance with gadolinium (Manaster and Graham 2003). Primary tumors and metastatic lesions both

show very high signal intensities on diffusion weighted images, which may help distinguish metastatic nodules from nodules of other unrelated etiologies (Kishimoto et al. 2012). Both MRI and CT scan can show bone destruction and extension of the tumor into the canal (Fig. 17.5). MR imaging can be used to differentiate benign notochordal cell tumors (BNCTs) from chordomas based on the lack of gadolinium uptake, bone sclerosis, and completely intraosseous location seen with BNCTs (Nishiguchi et al. 2011). Chordomas also demonstrate fluorodeoxyglucose avidity on F-18 PET scans (Lin et al. 2006; Miyazaway et al. 2008; Park and Kim 2008). Carbon-11-methionine positron emission tomography (MET-PET), used to evaluate the effectiveness of carbon-ion radiotherapy for assessment of rectal cancer and other tumors, has

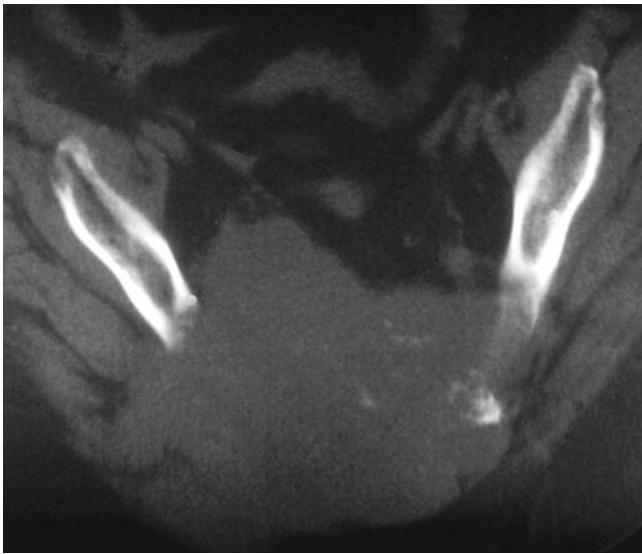


Fig. 17.5 Axial CT scan image showing extensive sacral bone destruction due to chordoma. Note that the tumor is a midline tumor, which can differentiate it from other sacral tumors

been used to study chordoma treatment. This technology has shown some promise in pre- and posttreatment imaging of chordoma (Zhang et al. 2004).

17.5 Histopathology and Immunohistochemistry

Virchow's original description of the physaliphorous cell, a vacuolated cell clustered in sheets giving the appearance of "soap bubbles" and demonstrating a lobular pattern of growth, describes the classic histologic features of chordomas (Fig. 17.6). These cells have small round dark staining nuclei with few mitotic figures but demonstrate significant atypia. The sheets of cells are separated by a fibrous septae with areas of calcification or hemorrhage representing necrotic areas (Weber and Sim 2002). These tumors can be separated into three classes: classical or conventional, chondroid, and dedifferentiated (Chugh et al. 2007). The classical form most commonly displays the typical physaliferous features. Chondroid type tumors may exhibit areas of chondrosarcoma-like cartilage as well as features of classic chordoma. Dedifferentiated tumors may have a more highly aggressive sarcomatous histological appearance. Dedifferentiated tumors may display nuclear inclusions, bi- or multinucleation, and sometimes mitotic figures (Crapanzano et al. 2001). Immunohistochemical staining is useful in the evaluation of chordomas. Significant immunoreactivity for S-100, membrane antigen (MUC-1), and cytokeratin is seen in these tumors.

Chondroid tumors are difficult to distinguish from chondrosarcomas, emphasizing the need for specific immunohistochemical markers. Brachyury is a notochordal transcription factor that is expressed in most sporadic chordomas, but not in chondrosarcomas (Vujovic et al. 2006). Interestingly, the

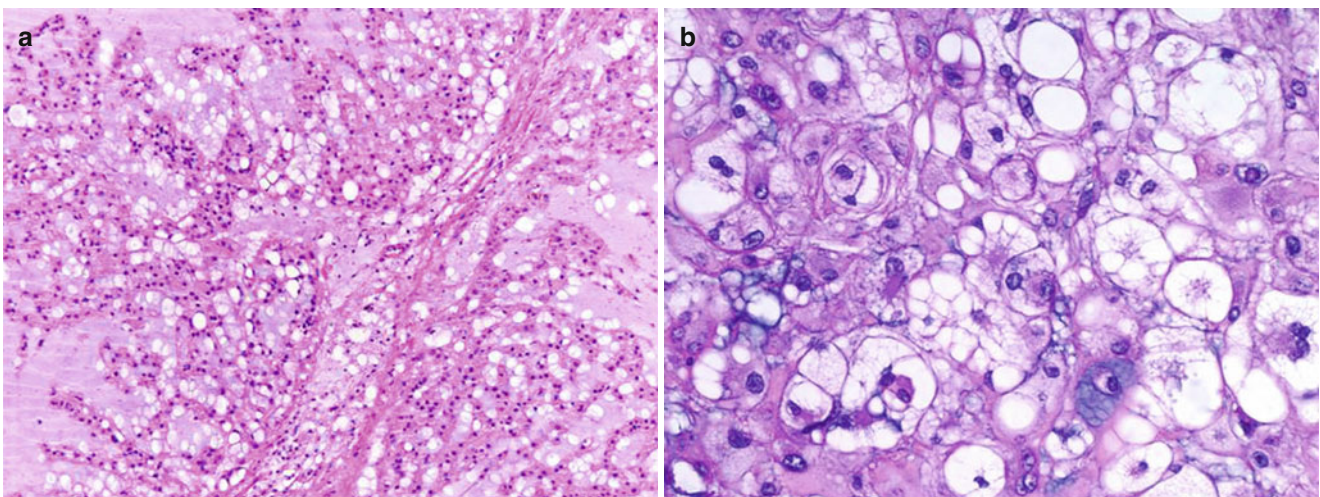


Fig. 17.6 Photomicrograph of chordoma histology demonstrating vacuolated physaliferous cells. (a) Low power, 100x; (b) high power, 400x

T gene locus containing the brachyury gene has been found to be duplicated in rare familial instances of chordoma, suggesting a critical role in the pathogenesis of this tumor (Yang et al. 2009). The addition of brachyury to the panel of biomarkers used to identify this cell type has improved the sensitivity and specificity for chordoma to 98 and 100 % (Oakley et al. 2008). Other important biomarkers such as ezrin, MMP-9, and COX-2 have also been recently investigated and hold diagnostic promise (Froehlich et al. 2012). The next step in the investigation of these markers is to study their value as potential therapeutic targets.

17.6 Differential Diagnosis

In the differential diagnosis of tumors of the spine, myeloma, plasmacytoma, benign notochordal cell tumor, lymphoma, osteomyelitis, giant cell tumor, and chondrosarcoma need to be considered (Sciubba et al. 2009). Key clinical, radiographic, and histologic findings help distinguish each type of tumor from a chordoma. Plasmacytoma may have a similar radiographic appearance as chordoma. A positive scintigraphy seen with a chordoma may differentiate these tumors (Greenspan et al. 2006). Radiographically, osteomyelitis and lymphoma can be difficult to distinguish from a chordoma; however, their clinical course and laboratory data can usually be used to distinguish these conditions (Sciubba et al. 2009). Benign notochordal cell tumors are usually asymptomatic, may demonstrate a more sclerotic appearance, and have no associated soft tissue mass. Giant cell tumors are benign but locally aggressive tumors that are often seen in the sacrum and can have an appearance similar to that of a chordoma. Chordomas have more of a midline predilection, however, and giant cell tumors often demonstrate a thin rim of peripheral bone encompassing the soft tissue mass which is not typically seen in a chordoma. Chondrosarcoma and chondroid chordomas can have similar appearances, and the biomarkers discussed above therefore play a critical role in distinguishing these tumors.

17.7 Tumor Staging

Staging is the process of defining the local and distant extension of a cancerous disease process. In the case of chordoma, MRI is the primary imaging modality used to evaluate the location and extent of the tumor. The presence of metastatic disease is evaluated using CT scan of the chest, abdomen, and pelvis with intravenous and oral contrast agents. Nuclear medicine bone scintigraphy can show other skeletal lesions. PET metabolic imaging can be used to demonstrate fluorodeoxyglucose avidity at sites of disease, but has some limitations based on lesion size, and is

not approved as a first-line modality for this purpose. F-18 PET or MET-PET are potentially promising techniques as described earlier.

17.8 Management

17.8.1 Surgery

The primary treatment of chordoma is wide surgical resection (Fig. 17.7) and the goal of surgery is wide excision with negative margins. This is sometimes an unrealistic goal due to the difficult locations of this tumor, especially clival or skull-base tumors, or high-level sacral tumors. Nonetheless, multiple studies have demonstrated a correlation between recurrence rate and positive margins (Boriani et al. 2006; Bilsky et al. 2004; Fuchs et al. 2005). Boriani et al. (2006) demonstrated that in the absence of negative margins, the recurrence rate was approximately 70 and 100 % following radiation treatment alone, palliative care, or intralesional excision. When wide excision with adequate margins was obtained, there was a recurrence rate of 20 % diagnosed at 56–94 months post surgery. In a group of patients all treated with en bloc excision for sacrococcygeal chordoma, Fuchs et al. (2005) demonstrated an overall survival rate of 74 % at 5 years, 52 % at 10 years, and 47 % at 15 years. Interestingly, the survival rate in this group of patients was significantly higher when negative margins were obtained, and the most significant predictor of survival was a wide margin. The size of the tumor, level of resection, and surgical approach did not significantly impact the survival rate.

Sacral resection can be achieved using either a combined anterior-posterior approach or a posterior-only approach. Generally, for tumors with significant anterior soft tissue

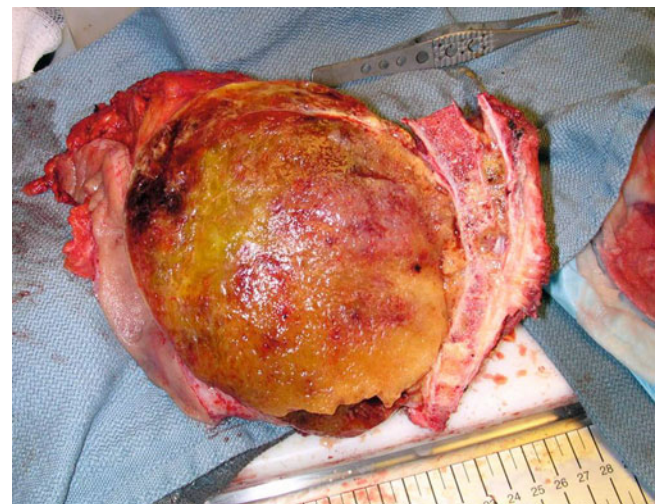


Fig. 17.7 Intraoperative photograph showing sacral chordoma after resection

mass, or high sacral resections, there is a benefit to first performing an anterior dissection prior to resection from a posterior approach. This was best shown by Fuchs et al. (2005) in a series from the Mayo Clinic in which 81 % of patients with combined anterior-posterior approaches had negative margins. Based on these results, it was recommended that a combined dual approach should be used for any patients with tumors above S3.

After chordoma resection, functional deficit is dependent on the extent of the surgery. To maximize survival rates and to prepare patients for postoperative deficits, careful preoperative planning and discussion of possible neurological problems is important. Whether partial or total sacrectomy is performed, usually one or more sacral nerve roots will need to be sacrificed, leading to a motor deficit, sensory impairment, sphincter loss, and/or sexual dysfunction. Fourney et al. (2005) developed a classification system describing the type of sacrectomy based on the level of nerve root sacrificed, as opposed to the site of the osteotomy. The type of resection was defined as low, middle, high sacral amputation, total sacrectomy, or hemisacrectomy. Sacral amputations are considered “low” if at least one S4 nerve root is sacrificed, “middle” if at least one S3 nerve root is sacrificed, and “high” if at least one S2 nerve root is sacrificed. If neither S1 nerve root can be spared, then the required amputation is a total sacrectomy. A hemisacrectomy (translumbar amputation) is performed when the tumor extends to the lumbar spine. Functionally, if both S2 roots can be spared, half of the patients or more will have normal bowel and bladder function. If an S3 root is preserved as well, these odds improve. However, perineal numbness and sexual dysfunction are still commonly observed, the latter more common in the elderly. If one S2 root is sacrificed, typically some amount of voluntary control is compromised. Sacrifice of one S1 or S2 nerve root and all lower roots, as seen in high sacral amputations, commonly leads to urinary or fecal incontinence leading to the possibility for the need of indwelling urinary catheters, intermittent straight catheterization, colostomy, or digital stimulation to defecate (Hulen et al. 2006). Loss of ankle plantar flexion is also seen after the loss of the S1 nerve root.

If resection of the lumbar spine or pelvis is required at the time of sacral amputation, spinopelvic instability must be assessed. If instability is observed, then instrumentation is warranted (Fig. 17.8). In general, about 50 % of the sacroiliac joint can be removed before instability is seen, provided the ligamentous structure of the remainder of the joint is preserved (Gunterberg et al. 1976; Stener and Gunteberg 1978). Chordomas of the mobile spine are also best treated with resection with negative margins. To achieve this goal, a total spondylectomy is the surgery of choice and is generally done through a combined anterior and posterior approach (Boriani et al. 2006).

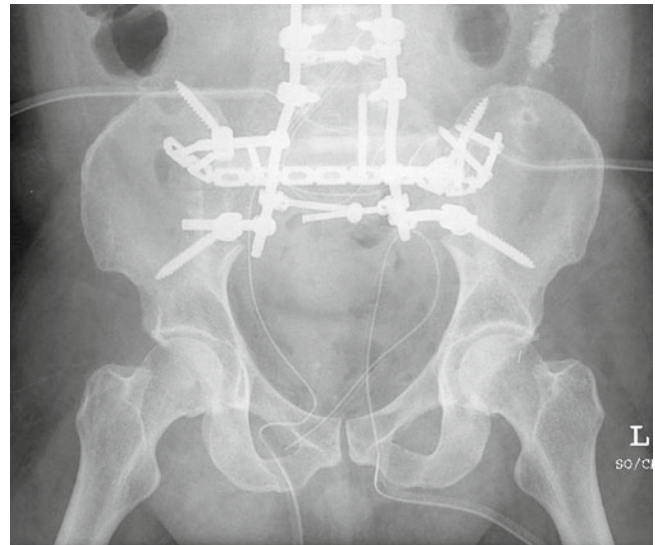


Fig. 17.8 An example of a postoperative reconstruction after resection of entire sacrum

In the skull base, despite multiple surgical approaches, wide resection surgery is rarely possible (Singh et al. 2010; Holzmann et al. 2010). Nonetheless, radiation therapy is effective in managing microscopic or limited gross amounts of residual tumor (Potluri et al. 2011). For this reason the recommended approach in these difficult anatomic areas is aggressive near-total intralesional excision that maximally preserves neurologic function.

17.8.2 Radiation

Radiation therapy has become increasingly important in the management of chordomas. As a result of advances in radiation techniques and modalities, it is now possible to deliver higher doses to the tumor while sparing critical surrounding structures. The limiting factor is the tolerance of the spinal cord, in particular at its upper end, which is lower than the dose needed to effectively treat the tumor. In general, conventional external beam radiation alone, at doses of 40–60 Gy, is suboptimal for treatment of chordoma, resulting in 5-year local control of 10–40 % (Catton et al. 1996; Cummings et al. 1983). Doses of up to 80 Gy have a high rate of associated radiation-induced myelopathy.

Use of high-dose protons and charged particles such as carbon, helium, or neon ions (collectively categorized as hadrons) enables higher doses to be delivered to tumors with limited radiation damage to critical surrounding structures. Indeed, as proton therapy has no measurable exit dose, peripheral structures are spared. Proton beam treatment of chordomas, either alone or in combination with photons, when used in conjunction with primary resection surgery, has proven to be an excellent method for local control

(Hug et al. 1999; Noël et al. 2001; Fuji et al. 2011). This is particularly relevant to skull-base tumors since the tolerance is lower than in the peri-sacral regions. In the sacrum, primary surgery and radiation provide better results when compared with treating chordoma recurrence, thereby supporting a role for this approach as an effective first-line treatment option (Park et al. 2006).

Carbon-ion radiotherapy has also been studied in chordoma management. Since carbon ions are heavier than protons, this approach is thought to provide a higher biological effectiveness. Interestingly the effectiveness increases with depth, reaching its peak at the end of the beam's range. This is an extremely attractive property for local control of cancer and as such has led to significant use of carbon-ion therapy in the management of chordoma. 5-year local control rates of 70–88 % and 10-year local control rates of 80–82 % have been reported in skull-base tumors using carbon-ion therapy (Schulz-Ertner et al. 2007; Mizoe et al. 2009; Tsujii and Kamada 2012).

An alternate approach to using hadrons in treatment of chordomas has been the use of highly conformal delivery techniques, such as intensity-modulated radiation therapy (IMRT) or stereotactic radiosurgery (SRS). In a recent study of the North American Gamma Knife Consortium, SRS was found to be an excellent option for small-sized chordomas, especially for young patients, and when combined with surgery, provides an overall 80 % 5-year local control rate (Kano et al. 2011).

17.8.3 Systemic Therapy and Future Directions

Chordomas are generally considered to be insensitive to conventional chemotherapy. Many conventional agents have been tried, with varying levels of response, including anthracycline, cisplatin, alkylating agents, and camptothecin analogues, but no single drug has emerged as a reliable first-line agent. Some small-scale sporadic reports suggest that dedifferentiated chordomas may have an increased

sensitivity to aggressive chemotherapy (Fleming et al. 1993), but overall there is no role for chemotherapy in treatment of localized disease. Currently, for metastatic disease, both the timing of initiating treatment and the choice of chemotherapy regimen are generally made on an individualized basis with significant consideration given to limiting the side effect profile.

Based on the observation that chordomas have a high expression of platelet-derived growth factor receptors (PDGFRB and PDGFRA) and KIT receptors (Tamborini et al. 2006), tyrosine kinase inhibitors have been used for the treatment of metastatic chordoma. Imatinib, a tyrosine kinase inhibitor with specificity for PDGFRB and KIT receptors, was initially studied in a small group of chordoma patients with advanced disease (Casali et al. 2005); the promising results of this study led to a larger phase II study with 50 patients (Stacchiotti et al. 2012). This recent study showed only one partial response obtained at 6 months; however, there were 35 patients with stable disease and a 64 % clinical benefit rate, confirming the findings of smaller-scale studies, and certainly warranting further investigation. The EGFR pathway has also been implicated in the pathogenesis of chordoma (Dewaele et al. 2011), leading to study of inhibitors of this pathway, including cetuximab, gefitinib, and erlotinib (Hof et al. 2006; Singhal et al. 2009). A multi-center trial of sunitinib which chordomas, making up 19 % of the study group, demonstrated a 44 % stable disease rate for 16 weeks (George et al. 2009). Current ongoing systemic trials include nilotinib, dasatinib, lapatinib, and everolimus.

Preclinical studies are also accelerating, primarily due to the development of several cell lines which have been characterized, including CH8, GP60, and U-CH-1 (Yang et al. 2010) and more recently CH22 (Liu et al. 2012). The development and characterization of these cell lines may help identify as yet unknown targets and help clarify the role of suspected targets such as brachyury (Hsu et al. 2011), leading to the development of further future clinical trials.

Box 17.2 RTK Inhibition in Chordoma

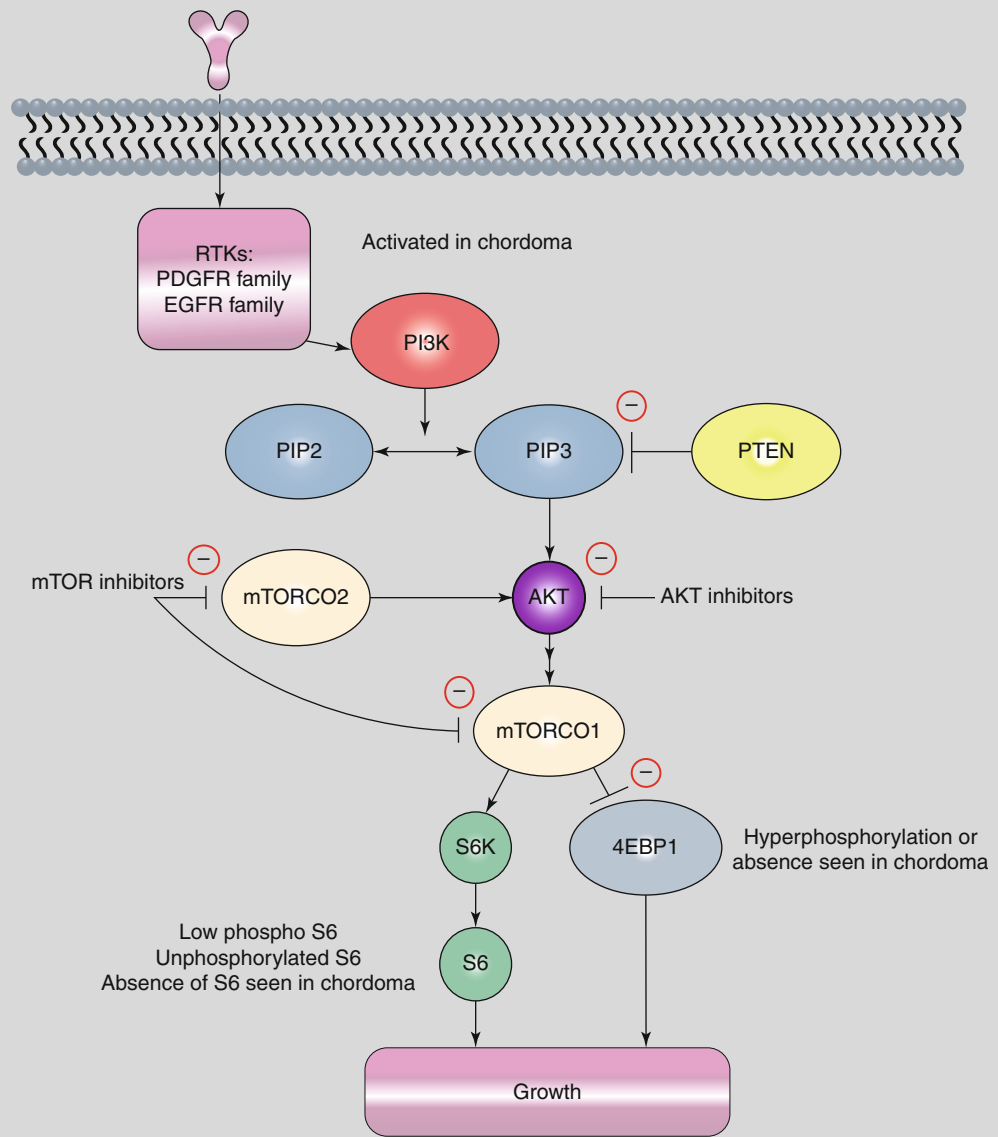
There is significant interest in the role of receptor tyrosine kinase (RTK) inhibition in the treatment of many sarcomas. Specifically, it has been demonstrated that chordomas express activated platelet-derived growth factor receptors (PDGFRB) (Tamborini et al. 2006). Furthermore, a subset of chordomas are known to express EGFR and c-MET, both of which signal through a tyrosine kinase pathway (Weinberger et al. 2005). mTOR is downstream of the receptor tyrosine kinases and is activated via

the MAPK or PI3K/AKT pathways. As the schematic indicates downstream of mTOR, the 40S ribosomal protein S6 kinase (p70^{S6k}) and the eukaryotic initiation factor 4E-binding protein-1 (4E-BP1) initiate protein synthesis and promote cell growth and proliferation. eIF4E in an initiation factor that binds to the mRNA cap. When hyperphosphorylated, 4E-BP1 binds to eIF4E repressing its translational initiation.

A recent study investigated the role of RTK inhibition in chordomas (Tamborini et al. 2010). It was noted that

activated PDGFR, FLT3, CSF1-R, all components of the PDGFR family, and EGFR family members EGFR, Her2/neu, and HER4 were present in chordoma tissue samples. These findings are in strong support of the concept that the PDGFR and EGFR pathways are activated in chordoma. Another observation was that EGFR and PDGFRB were co-immunoprecipitated, suggesting heterodimer formation. This information could explain why some chordomas are resistant to imatinib treatment. It is possible that a bimodal approach using anti-PDGFR and anti-EGFR agents may be required to fully silence the activation of mTOR in chordomas. Interestingly, in two chordoma patients, there was a clinical response to cetuximab, an anti-EGFR monoclonal antibody (Hof et al. 2006).

With respect to downstream effectors of mTOR, Western blot analysis showed that 14 out of 22 chordomas cases demonstrated eIF4E release from translational repression of 4E-BP1. In 13 of these cases, this was due to hyperphosphorylation of 4E-BP1, and in one case there was an absence of 4E-BP1. At the same time, phospho S6 was only present at low or very low levels in 11 cases; unphosphorylated S6 was found at low levels in 3 cases and not expressed at all in 8 cases. It is unclear if this suggests a tumor suppressor role for S6 or that the majority of downstream effect of mTOR in chordoma is mediated via 4E-BP1/eIF4E, but this discrepancy certainly warrants further study.



Schematic demonstrating signaling through RTKs via mTOR and critical effectors PTEN, PI3K, and AKT, and downstream effectors S6K and 4EBP1

Box 17.3 Genetics of Chordoma

Molecular studies of tumors from families with familial chordoma syndromes have shed some light on the genetics of chordoma. A recent study by Yang et al. (2009) details the identification of T (brachyury) gene duplication and its role in conferring susceptibility to familial chordoma. These studies were performed using combined genetic linkage and high-resolution array CGH (comparative genomic hybridization) analyses to identify unique duplications of a region on 6q27 in four families with more than three cases of chordoma in each. This locus was found to contain the T (brachyury) gene. Brachyury is a tissue-specific transcription factor expressed in the nucleus of notochord cells. Chordomas express brachyury, but its expression has been studied, and it is not found to be expressed in nonneoplastic tissues and in 42 other types of neoplasm. Its exact role in the pathogenesis of chordoma is unclear, but the finding represents a significant advancement in the understanding of chordoma biology.

The majority of chordomas, however, are sporadic. Sporadic chordomas do not display duplication or amplification of the brachyury gene. Karyotype analyses of sporadic chordomas demonstrate several abnormalities that identify this disease as one of significant genetic instability, as demonstrated in a study by Le et al. (2011). Copy number variations involving copy number losses are seen more frequently than copy number gains. The chromosomes with relevant losses include 1p,3,4,9,10,13,14, and 18. PTEN, which is an important tumor suppressor gene located on 10q23.3, was found to have hemizygous deletion in 80 % of the sporadic chordomas studied. Interestingly, hyperactivation of Akt/mTORC1 signaling in sporadic sacral chordomas has

been described and is consistent with loss of PTEN (Han et al. 2009, also see Box 17.2 for a review of RTK signaling in chordoma). CDKN2A is a tumor suppressor gene that inhibits the function of cdk4- and cdk6-cyclin D complexes. These cdk-cyclin complexes regulate the retinoblastoma protein, thereby controlling the G1-S checkpoint of cell cycle progression. This gene was found to be deficient in 80 % of sporadic tumors studied as well.

Although the precise genetic mechanism for the development of chordoma is unclear, these data taken together suggest that T/brachyury is a key figure in the mechanism, at least in the case of familial chordoma. It is possible that T/brachyury is important in sporadic chordoma as well, but if this is the case, then the mechanism must be one other than copy number variation based on these findings.

References

- Han S, Polizzano C, Neilsen GP, Hornicek FJ, Rosenberg AE, Ramesh V (2009) Aberrant hyperactivation of akt and Mammalian target of rapamycin complex 1 signaling in sporadic chordomas. *Clin Cancer Res* 15:1940–1946
- Le LP, Nielsen GP, Rosenberg AE, Thomas D, Batten JM, Deshpande V, Schwab J, Duan Z, Xavier RJ, Hornicek FJ, Iafrate AJ (2011) Recurrent chromosomal copy number alterations in sporadic chordomas. *PLOS One* 6:e18846
- Yang XR, Ng D, Alcorta DA, Liebsch NJ, Sheridan E, Li S, Goldstein AM, Parry DM, Kelley MJ (2009) T (brachyury) gene duplication confers major susceptibility to familial chordoma. *Nat Genet* 41:1176–1178

17.9 Summary of Critical Concepts Discussed in This Chapter

- Chordomas are rare tumors of notochordal origin.
- Surgical treatment with wide resection is the mainstay of therapy.
- Local recurrence and survival rates are dependent on margins.
- Radiation is an important modality of treatment, generally as an adjuvant to surgery.
- Alternate forms of radiation such as proton therapy, hadron therapy, or stereotactic radiation are all playing increasingly important roles.
- Platelet-derived growth factor receptor signaling is important in the oncogenesis of chordoma.

- The role for chemotherapy is limited but advancements in systemic therapy using agents targeting tyrosine kinase pathways show significant promise.

References

- Ashwood N, Hoskin PJ, Saunders MI (1994) Metastatic chordoma: pattern of spread and response to chemotherapy. *Clin Oncol (R Coll Radiol)* 6:341–342
- Atalar H, Selek H, Yildiz Y, Sağlık Y (2006) Management of sacrococcygeal chordomas. *Int Orthop* 30(6):514–518
- Bergh P, Kindblom LG, Gunterberg B, Remotti F, Ryd W, Meis-Kindblom JM (2000) Prognostic factors in chordoma of the sacrum and mobile spine: a study of 39 patients. *Cancer* 88:2122–2134
- Bilsky MH, Yamada Y, Yenice KM, Lovelock M, Hunt M, Gutin PH, Leibel SA (2004) Intensity-modulated stereotactic radiother-

- apy of paraspinous tumors: a preliminary report. *Neurosurgery* 54: 823–830
- Bjornsson J, World LE, Ebersold MJ, Laws ER (1993) Chordoma of the mobile spine. A clinicopathological analysis of 40 patients. *Cancer* 71:735–740
- Bohman HH, Sachs BL, Carter JR, Riley L, Robinson RA (1986) Primary neoplasms of the cervical spine. Diagnosis and treatment of twenty-three patients. *J Bone Joint Surg Am* 68:483–494
- Boriani S, Chevalley F, Weinstein JN, Biagini R, Campanacci L, De Iure F, Piccilli P (1996) Chordoma of the spine above the sacrum. Treatment and outcome in 21 cases. *Spine* 21:1569–1577
- Boriani S, Bandiera S, Biagini R, Bacchini P, Boriani L, Cappuccio M, Chevalley F, Gasbarrini A, Picci P, Weinstein JN (2006) Chordoma of the mobile spine: fifty years of experience. *Spine* 31:493–503
- Burrow JF, Stewart MJ (1923) Malignant sphenoid-occipital chordoma. *J Neurol Psychopathol* 4(15):205–217
- Casali PG, Messina A, Stacchiotti S, Tamborini E, Crippa F, Gronchi A, Orlandi R, Ripamonti C, Spreafico C, Bertieri R, Bertulli R, Colecchia M, Fumagalli E, Greco A, Grosso F, Olmi P, Pierotti MA, Pilotti S (2005) Imatinib mesylate in 18 advanced chordoma patients. *J Clin Oncol* 23:9012
- Catton C, O'Sullivan B, Bell R, Laperriere N, Cummings B, Fornasier V, Wunder J (1996) Chordoma: long-term follow up after radical photon irradiation. *Radiother Oncol* 41:67–72
- Chugh R, Tawbi H, Lucas DR, Biermann JS, Schuetze SM, Baker LH (2007) Chordoma: the nonsarcoma primary bone tumor. *Oncologist* 12:1344–1350
- Crapanzano JP, Ali SZ, Ginsberg MS, Zakowski MF (2001) Chordoma: a cytologic study with histologic and radiologic correlation. *Cancer* 93:40–51
- Cummings BJ, Hodson DI, Bush RS (1983) Chordoma: the results of megavoltage radiation therapy. *Int J Radiat Oncol Biol Phys* 9: 633–642
- Dewaele B, Maggiani F, Floris G, Ampe M, Vanspauwen V, Wozniak A, Debiec-Rychter M, Sciort R (2011) Frequent activation of EGFR in advanced chordomas. *Clin Sarcoma Res* 1(1):4
- Fleming GF, Heimann PS, Stephens JK, Simon MA, Ferguson MK, Benjamin RS, Samuels BL (1993) Dedifferentiated chordoma. Response to aggressive chemotherapy in two cases. *Cancer* 72(3):714–718
- Forsyth PA, Cascino TL, Shaw EG, Scheithauer BW, O'Fallon JR, Dozier JC, Piegras DG (1993) Intracranial chordomas: a clinicopathological and prognostic study of 51 cases. *J Neurosurg* 78: 741–747
- Fourney DR, Gokaslan ZL (2003) Current management of sacral chordoma. *Neurosurg Focus* 15:E9
- Fourney DR, Rhines LD, Hentschel SJ, Skibber JM, Wolinsky JP, Weber KL, Suki D, Gallia GL, Garonzik I, Gokaslan ZL (2005) En bloc resection of primary sacral tumors: classification of surgical approaches and outcome. *J Neurosurg Spine* 3:111–122
- Froehlich EV, Scheipl S, Lazary A, Varga PP, Schmid C, Stammberger H, Beham A, Bodo K, Schroettner H, Quehenberger F, Windhager R, Liegl B, Leithner A (2012) Expression of ezrin, MMP-9, and COX-2 in 50 chordoma specimens: a clinical and immunohistochemical analysis. *Spine* 37(13):E757–E767
- Fuchs B, Dickey I, Yaszemski MJ, Inwards CY, Sim FH (2005) Operative management of sacral chordoma. *J Bone Joint Surg Am* 87:2211–2216
- Fuji H, Nakasu Y, Ishida Y, Horiguchi S, Mitsuya K, Kashiwagi H, Murayama S (2011) Feasibility of proton beam therapy for chordoma and chondrosarcoma of the skull base. *Skull Base* 21(3):201–206
- George S, Merriam P, Maki RG, Van den Abbeele AD, Yap JT, Akhurst T, Harmon DC, Bhuchar G, O'Mara MM, D'Adamo DR, Morgan J, Schwartz GK, Wagner AJ, Butrynski JE, Demetri GD, Keohan ML (2009) Multicenter phase II trial of sunitinib in the treatment of nongastrointestinal stromal tumor sarcomas. *J Clin Oncol* 27(19):3154–3160
- Greenspan A, Jundt G, Remagen W (2006) Chordoma, in differential diagnosis in orthopaedic oncology, 2nd edn. Lippincott Williams & Wilkins, Philadelphia, pp 445–446
- Gunterberg B, Stener B, Romanus B (1976) Pelvic strength after major amputation of the sacrum. An experimental study. *Acta Orthop Scand* 47:635–642
- Healey JH, Lane JM (1989) Chordoma: a critical review of diagnosis and treatment. *Orthop Clin North Am* 20:417–426
- Hof H, Welzel T, Debus J (2006) Effectiveness of cetuximab/ gefitinib in the therapy of a sacral chordoma. *Onkologie* 29(12):572–574
- Holzmann D, Reisch R, Kraysenbuhl H, Hug E, Bernays RL (2010) The transnasal transclival approach for clivus chordoma. *Minim Invasive Neurosurg* 53:211–217
- Hsu W, Mohyeldin A, Shah SR, Rhys CM, Johnson LF, Sedora-Roman NI, Kosztowski TA, Awad OA, McCarthy EF, Loeb DM, Wolinsky JP, Gokaslan ZL, Quiñones-Hinojosa A (2011) Generation of chordoma cell line JHC7 and the identification of Brachyury as a novel molecular target. *J Neurosurg* 115(4):760–769
- Hug EB, Loredò LN, Slater JD, DeVries A, Grove RI, Schaefer RA, Rosenberg AE, Slater JM (1999) Proton radiation therapy for chordomas and chondrosarcomas of the skull base. *J Neurosurg* 91(3): 432–439
- Hulen CA, Temple HT, Fox WP, Sama AA, Green BA, Eismont FJ (2006) Oncologic and functional outcome following sacrectomy for sacral chordoma. *J Bone Joint Surg Am* 88:1532–1539
- Kano H, Iqbal FO, Sheehan J, Mathieu D, Seymour ZA, Niranjan A, Flickinger JC, Kondziolka D, Pollock BE, Rosseau G, Sneed PK, McDermott MW, Lunsford LD (2011) Stereotactic radiosurgery for chordoma: a report from the North American Gamma Knife Consortium. *Neurosurgery* 68(2):379–389
- Kishimoto R, Omatsu T, Hasegawa A, Imai R, Kandatsu S, Kamada T (2012) Imaging characteristics of metastatic chordoma. *Jpn J Radiol* 30(6):509–516
- Lee J, Bhatia NN, Hoang BH, Ziogas A, Zell JA (2012) Analysis of prognostic factors for patients with chordoma with use of the California Cancer Registry. *J Bone Joint Surg Am* 94(4):356–363
- Leone A, Cerase A, Tarquini E, Mulè A (2002) Chordoma of the low cervical spine presenting with Horner's syndrome. *Eur Radiol* 12(Suppl 3):S43–S47
- Lin CY, Kao CH, Liang JA, Hsieh TC, Yen KY, Sun SS (2006) Chordoma detected on F-18 FDG PET. *Clin Nucl Med* 31(8):506–507
- Liu X, Nielsen GP, Rosenberg AE, Waterman PR, Yang W, Choy E, Sassi S, Yang S, Harmon DC, Yang C, Schwab JH, Kobayashi E, Mankin HJ, Xavier R, Weissleder R, Duan Z, Hornicek FJ (2012) Establishment and characterization of a novel chordoma cell line: CH22. *J Orthop Res*. doi:10.1002/jor.22113
- Manaster BJ, Graham T (2003) Imaging of sacral tumors. *Neurosurg Focus* 15(2):E2
- McMaster ML, Goldstein AM, Bromley CM, Ishibe N, Parry DM (2001) Chordoma: incidence and survival patterns in the United States, 1973–1995. *Cancer Causes Control* 12:1–11
- Meyer JE, Lepke RA, Lindfors KK, Pagani JJ, Hirschy JC, Hayman LA, Momose KJ, McGinnis B (1984) Chordomas: their CT appearance in the cervical, thoracic and lumbar spine. *Radiology* 153(3):693–696
- Mindell E (1981) Chordoma. *J Bone Joint Surg Am* 63:501–505
- Miyazawa N, Ishigame K, Kato S, Satoh Y, Shinohara T (2008) Thoracic chordoma: review and role of FDG-PET. *J Neurosurg Sci* 52(4): 117–121
- Mizoe JE, Hasegawa A, Takagi R, Bessho H, Onda T, Tsujii H (2009) Carbon ion radiotherapy for skull base chordoma. *Skull Base* 19(3):219–224
- Muller H (1858) Über das Vorkommen von Resten der Chorda dorsalis bei Menschen nach der Geburt und über ihr Verhältniss zu den Gallertgeschwülsten am Clivus. *Zeitschr f rat Med* ii:202–229

- Nicoucar K, Rausch T, Becker M, Dulguerov P (2008) Cervical chordoma with retropharyngeal extension presenting with impaired voice. *Tumori* 94:873–876
- Nishiguchi T, Mochizuki K, Ohsawa M, Inoue T, Kageyama K, Suzuki A, Takami T, Miki Y (2011) Differentiating benign notochordal cell tumors from chordomas: radiographic features on MRI, CT, and tomography. *AJR Am J Roentgenol* 196(3):644–650
- Noël G, Habrand JL, Mammar H, Pontvert D, Haie-Méder C, Hasboun D, Moisson P, Ferrand R, Beaudré A, Boisserie G, Gaboriaud G, Mazal A, Kérody K, Schlienger M, Mazon JJ (2001) Combination of photon and proton radiation therapy for chordomas and chondrosarcomas of the skull base: the centre de Protonthérapie D'Orsay experience. *Int J Radiat Oncol Biol Phys* 51(2):392–398
- Oakley GJ, Fuhrer K, Seethala RR (2008) Brachyury, SOX-9, and podoplanin, new markers in the skull base chordoma vs chondrosarcoma differential: a tissue microarray-based comparative analysis. *Mod Pathol* 21:1461–1469
- Papagelopoulos PJ, Mavrogenis AF, Currier BL, Katonis P, Galanis EC, Sapkas GS, Korres DS (2004) Primary malignant tumor of the cervical spine. *Orthopedics* 27:1067–1075
- Park SA, Kim HS (2008) F-18 FDG PET/CT evaluation of sacrococcygeal chordoma. *Clin Nucl Med* 33(12):906–908
- Park L, Delaney TF, Liebsch NJ, Hornicek FJ, Goldberg S, Mankin H, Rosenberg AE, Rosenthal DI, Suit HD (2006) Sacral chordomas: impact of high-dose proton/photon-beam radiation therapy combined with or without surgery for primary versus recurrent tumor. *Int J Radiat Oncol Biol Phys* 65(5):1514–1521
- Pearlman AW, Friedman M (1970) Radical radiation therapy of chordoma. *Am J Roentgenol Radium Ther Nucl Med* 108(2):332–341
- Pearlman AW, Singh RK, Hoppenstein R, Wilder J (1972) Chordoma: combined therapy with radiation and surgery: case report and new operative approach. *Bull Hosp Joint Dis* 33(1):47–57
- Potluri S, Jeffries SJ, Jena R, Harris F, Burton KE, Prevost AT, Burnet NG (2011) Residual postoperative tumor volume predicts outcome after high-dose radiotherapy for chordoma and chondrosarcoma of the skull base and spine. *Clin Oncol* 32:199–208
- Schulz-Ertner D, Karger CP, Feuerhake A, Nikoghosyan A, Combs SE, Jäkel O, Edler L, Scholz M, Debus J (2007) Effectiveness of carbon ion radiotherapy in the treatment of skull-base chordomas. *Int J Radiat Oncol Biol Phys* 68(2):449–457
- Sciubba D, Cheng JJ, Petteys RJ, Weber KL, Frassica DA, Gokaslan ZL (2009) Chondroma of the sacrum and vertebral bodies. *J Am Acad Orthop Surg* 17(11):708–717
- Singh N, Soo M, De Cruz M, Gomes L, Maclean F, Dandie G (2007) Cervical chordoma presenting as retropharyngeal mass and dysphonia: case report and literature review. *Australas Radiol* 51(suppl):B183–B188
- Singh H, Harrop J, Schiffmacher P, Rosen M, Evans J (2010) Ventral Surgical approaches to craniovertebral junction chordomas. *Neurosurgery* 66(suppl 3):96–103
- Singhal N, Kotasek D, Parnis FX (2009) Response to erlotinib in a patient with treatment refractory chordoma. *Anticancer Drugs* 20(10):953–955
- Soo MY (2001) Chordoma: review of clinicoradiological features and factors affecting survival. *Australas Radiol* 45:427–434
- Stacchiotti S, Longhi A, Ferraresi V, Grignani G, Comandone A, Stupp R, Bertuzzi A, Tamborini E, Pilotti S, Messina A, Spreafico C, Gronchi A, Amore P, Vinaccia V, Casali PG (2012) Phase II study of imatinib in advanced chordoma. *J Clin Oncol* 30(9):914–920
- Stener B, Gunterberg B (1978) High amputation of the sacrum for extirpation of tumors. Principles and technique. *Spine* 3:351–366
- Sundaresan N, Schmidek HH, Schiller AL, Rosenthal D (1990) Tumors of the spine: diagnosis and clinical management. W.B. Saunders, Philadelphia
- Tamborini E, Miselli F, Negri T et al (2006) Molecular and biochemical analyses of platelet-derived growth factor receptor (PDGFR) B, PDGFRA, and KIT receptors in chordomas. *Clin Cancer Res* 12:6920–6928
- Tamborini E, Virdis E, Negri T, Orsenigo M, Bricchi S, Conca E, Gronchi A, Stacchiotti S, Manenti G, Casali PG, Pierotti MA, Pilotti S (2010) Analysis of receptor tyrosine kinases (RTKs) and downstream pathways in chordomas. *Neuro Oncol* 12(8):776–789.
- Tsuji H, Kamada T (2012) A review of update clinical results of carbon ion radiotherapy. *Jpn J Clin Oncol* 42(8):670–685
- Unni KK (1996) Chordoma. In: Unni KK (ed) *Dahlin's bone tumors: general aspects and data on 11,087 cases*, 5th edn. Lippincott-Raven, Philadelphia, pp 291–305
- Virchow RL (1857) *Untersuchungen ueber die Entwicklung des Schaedelgrundes*. G Rimer, Berlin
- Vujovic S, Henderson S, Presneau N, Odell E, Jacques TS, Tirabosco R, Boshoff C, Flanagan AM (2006) Brachyury, a crucial regulator of notochordal development, is a novel biomarker for chordomas. *J Pathol* 209:157–165
- Weber K, Sim FH (2002) Chordoma. In: Bulstrode C, Buckwalter J, Carr A et al (eds) *Oxford textbook of orthopaedics and trauma*. Oxford University Press, New York, pp 294–299
- Weinberger PM, Yu Z, Kowalski D, Joe J, Manger P, Psyrrri A, Sasaki CT (2005) Differential expression of epidermal growth factor receptor, c-Met, and HER2/neu in chordoma compared with 17 other malignancies. *Arch Otolaryngol Head Neck Surg* 131(8):707–711.
- Yang XR, Ng D, Alcorta DA, Liebsch NJ, Sheridan E, Li S, Goldstein AM, Parry DM, Kelley MJ (2009) T (brachyury) gene duplication confers major susceptibility to familial chordoma. *Nat Genet* 41(11):1176–1178
- Yang C, Hornicek FJ, Wood KB, Schwab JH, Choy E, Iafrate J, Rosenberg A, Nielsen GP, Xavier RJ, Mankin H, Duan Z (2010) Characterization and analysis of human chordoma cell lines. *Spine* 35(13):1257–1264
- Zhang H, Yoshikawa K, Tamura K, Sagou K, Tian M, Suhara T, Kandatsu S, Suzuki K, Tanada S, Tsujii H (2004) Carbon-11-methionine positron emission tomography imaging of chordoma. *Skeletal Radiol* 33(9):524–530