Rhizoremediation: A Pragmatic Approach for Remediation of Heavy Metal-Contaminated Soil

9

Velmurugan Ganesan

Abstract

Soil pollution is the primary source that transmits pollutants like heavy metals from environment to living organisms. From soil, plants adsorb and accumulate heavy metals. Through the food chain, heavy metals enter the animal kingdom including humans and cause health risks. Few physicochemical and phytoremediation approaches have been proved effective in removing heavy metals from contaminated soils. However, soil characteristics and recycling of soil constituents have made their practicability questionable. One pragmatic way to reduce the deleterious effect of heavy metals in soil is rhizoremediation, in which plant–microbe interaction is explored for remediation purposes. In this strategy, the plant growth-promoting rhizobacteria (PGPR) either accumulate or detoxify the heavy metals and thereby prevent the uptake and accumulation of heavy metals in plants. In addition, PGPRs act as biofertilizer that enhance the crop yields in different ecological niches. In this chapter, rhizoremediation strategy is described and portrayed as the pragmatic way for remediation of heavy metals in soil.

9.1 Introduction

Heavy metal, the poorly defined term, is the subset of 40 elements including transition metals, metalloids, lanthanides, and actinides (Appenroth [2010](#page-10-0)) that have specific density of more than 5 g/cm^3 . They possess metallic characteristics such as ductility, conductivity, stability as cations, ligand specificity, etc. Though the term heavy metal is announced as a meaningless term by IUPAC, it is widely

V. Ganesan (\boxtimes)

Biocentre, University of Cologne, Cologne 50674, Germany

Current Address: Department of Molecular Biology, School of Biological Sciences, Madurai Kamaraj University, Madurai 625021, India e-mail: oomvel@gmail.com

used in biology (Duffus [2002\)](#page-11-0). Biologically, they are classified as essential and nonessential heavy metals. Some of the heavy metals like cobalt (Co), nickel (Ni), copper (Cu) , zinc (Zn) , selenium (Se) , and molybdenum (Mo) are essential elements whose role in metabolism is known in both prokaryotes and eukaryotes. In contrast, no nutritional function is known for heavy metals like silver (Ag) , cadmium (Cd), arsenic (As), lead (Pb), mercury (Hg), and uranium (U) (Appenroth [2010\)](#page-10-0). While in case of chromium (Cr), the role of Cr^{3+} ions in sugar and lipid metabolism is known, but the role of Cr^{6+} ions is unknown (Vincent [2000\)](#page-13-0).

The discharges of heavy metals from natural and anthropogenic activities cause the accumulation of metals into the environment. Natural input includes withering and erosion of parent rocks that transfer large quantities of metals to water bodies and lands (Gadd [2010](#page-11-0)). As per the World Health Organization (WHO) report (1992), 15,000 metric tons (mt) of Cd is added to the oceans every year. A higher level of heavy metal accumulation is reported in the marine sedimentary rocks, marine phosphates, and phosphorites. Volcanic eruptions (Hong et al. [1996](#page-11-0)) and forest fires (Shcherbov et al. [2008](#page-13-0)) also contribute to natural inputs. In addition, the use of heavy metals by humans is known for years (Nriagu [1996\)](#page-13-0). The uses include mining, smelting, fuel combustion, synthetic fertilizers, metal alloys, electroplating, and Ni–Cd batteries, as pigment in plastics and as stabilizer in PVC, in electronic goods, and in solar cells (Gadd [2010\)](#page-11-0). Because of these activities, the contents of Pb, Hg, and Cd in the pedosphere (earth's outermost soil layer) are about 10, 6, and 5 times, respectively, higher than in the lithosphere (earth's crust and outermost mantle layer) (Han et al. [2002](#page-11-0)). Here, we summarize the toxicity of heavy metals, different approaches employed for soil remediation, and the distinctive properties of rhizoremediation approach, which have made this technology user-, eco-, and economic friendly.

9.2 Soil Pollution

Soil is the major reservoir for most of the metals and nonmetals including the nutrients and is the prime site of biogeochemical cycling of elements. Over the years, continuous cropping and other agricultural activities have resulted in depletion of nutrients in soil (Zhang et al. [2006](#page-14-0)), requiring application of plant nutrients from external sources. In this context, synthetic agrochemicals especially phosphate fertilizers are excessively applied, which have resulted in heavy metal pollution of soil (Mortvedt [1996](#page-12-0); McGrath and Tunney [2010\)](#page-12-0). Besides these, sewage sludge application and industrial activities also add heavy metals to agricultural soil (Kelly et al. [1999;](#page-12-0) Han et al. [2002\)](#page-11-0). The major factors influencing metals speciation, adsorption, and distribution in soils include pH, soluble organic matter, hydrous metal oxide, clay content and type, organic and inorganic ligands, and competition from other metal ions (Dube et al. [2001](#page-11-0)).

Even though heavy metals in soil seem to be immobile, they interact with biotic components especially the plant roots. Thereafter, they are transported to all parts of the plants including the fruits, vegetables, and seeds. Consequently, heavy metals enter the food chain of animals including humans (Fig. 9.1). It is reported that terrestrial foods account for 98% of the ingested toxic heavy metals, while 1% each of aquatic foods and drinking water (Van Assche [1998\)](#page-13-0). The Joint Food and Agricultural Organization (FAO)/WHO Expert Committee on Food Additives (JECFA) determined the provisional tolerable weekly intake (PTWI) for the toxic heavy metals which is listed in Table 9.1. Among the toxic heavy metals, Hg and Cd have PTWI values less than 10 μ g/kg body weight showing their high toxicity, while Cu and Zn being the essential metal ions have PTWI values of 3,500 and 7,000 mg/kg body weight, respectively. Various studies reported the presence of heavy metals higher than the PTWI values in vegetables, canned foods, and other eatables in all parts of the world. The technical report of Imperial College of London prepared in collaboration with Indian universities has revealed the heavy metal contamination of vegetables in Delhi, India (Marshall et al. [2003](#page-12-0)). According to the Indian Council of Medical Research ([2003\)](#page-12-0) report, nearly 50% of the tested mother's milk samples had Cd eight times more than stringent limits. Since heavy metals are not quickly eliminated from the human system, they bioaccumulate to

Fig. 9.1 Food chain exhibiting the transport of heavy metals

Reports of Joint Food and Agricultural Organization (FAO)/WHO expert committee on food additives (JECFA)

toxic levels. The biological half-life of toxic heavy metals ranges between 20 and 30 years (Sugita [1978](#page-13-0)). The WHO ([1987\)](#page-14-0) estimated that the daily intake of 200 μ g Cd for longer period can be connected with a 10% prevalence of adverse health effects suggesting threat to food security.

9.3 Heavy Metal Toxicity

Most of the heavy metals cause changes in both the environment and living organisms. Besides the nonessential heavy metals, even the essential heavy metals become toxic, when their level exceeds the physiological value. The prime reason for their toxicity is due to their ability to bind strongly to oxygen, nitrogen, and sulfur atoms because of free enthalpy of the metal–ligand product (Weast [1984\)](#page-14-0). As a result, heavy metals inactivate the enzymes by binding to –SH group, leading to changes in metabolism (Fuhrer [1982\)](#page-11-0). Many enzymes and proteins need essential divalent cations like Ca^{2+} , Mg^{2+} , Ni^{2+} , Co^{2+} , and Zn^{2+} , which are displaced by the toxic divalent ions. For instance, in case of calmodulin, the protein that is important in cell signaling, the Ca^{2+} ions are displaced by Cd^{2+} ion, leading to loss of activity (Rivetta et al. [1997\)](#page-13-0). The binding ability of heavy metals with nucleic acids allows them to act as mutagens, which lead to misreading of the genetic profile (Wong [1988\)](#page-14-0). In addition, they cause lipid peroxidation and oxidative stress, leading to membrane damage (Howlett and Avery [1997](#page-12-0)).

Sources of heavy metals for plants are the soil, irrigation water, and air emissions. Plants take up heavy metals primarily from the soil and accumulate in the plant tissues (Fig. 9.2a). Following accumulation, heavy metals cause changes in the metabolic pathways like photosynthesis (Clijsters and Van Assche [1985](#page-11-0); Somasundaram et al. [1994](#page-13-0); Pandey and Tripathi [2011](#page-13-0)), protein and nitrogen metabolism (Hemalatha et al. [1997](#page-11-0); Llorens et al. [2001](#page-12-0); Manios et al. [2002;](#page-12-0) Priti et al. [2009](#page-13-0)), uptake of nutrients

Fig. 9.2 Schematic representation of rhizoremediation

(Veselov et al. [2003\)](#page-13-0), and sugar and water metabolism (Pandey and Tripathi [2011;](#page-13-0) Stobrawa and Lorenc-Plucintska [2007;](#page-13-0) Babula et al. [2008\)](#page-11-0). Heavy metals also lead to hormonal imbalances and especially elevate ethylene synthesis (Arteca and Arteca [2007](#page-10-0)) and decrease cytokinin level due to oxidation (Hare et al. [1997](#page-11-0)). These effects lead to poor crop productivity. For humans, the heavy metals enter through polluted food, water, air, and occupational exposure. Heavy metals are carcinogens that alter the gene expression, leading to cell proliferation by the induction of proto-oncogenes or by interference with genes involved in cell growth (Beyersmann [2002\)](#page-11-0). International Agency for Research on Cancer (IARC) grouped Cd, Cr, and As in group 1 (proven human carcinogens) and Ni, Co, Hg, and Pb in group 2B (possibly carcinogenic to humans). The teratogenic effects of heavy metals like Cd, Pb, Hg, and U are also reported (Emmanouil-Nikolussi [2007\)](#page-11-0). The other health risks include renal tubular damage, bone demineralization, cardiac failure, nervous, respiratory disorders, and loss of fertility (Duruibe et al. [2007\)](#page-11-0). Even low levels of Cd, Pb, and Hg exposure are reported to diminish intellectual capacity and brain development of children (Drum [2009](#page-11-0)).

9.4 Soil Remediation Approaches

Many different physical, chemical, and biological methods are proposed for the remediation of metal-contaminated soil.

9.4.1 Physicochemical Methods

The conventional ex situ methods like land filling, incineration, leaching, and chemical methods are adopted to remediate metal-contaminated soils, but they are not effective (Lambert et al. [2000](#page-12-0)). Other in situ approaches like vitrification and electrokinetics that involve the application of electrical voltages are efficient, but labor safety and cost factors are of major concern (Mulligan et al. [2001\)](#page-12-0). All these techniques just transfer the contaminants from soils to some other material, which needs to be transported and recycled. Hence, the low efficiency, high cost, safety problems, recycling, transfer, and need to analyze the nature of the contaminants and type of soil are the major setbacks for these approaches. In addition, the ex situ modes and transfer of absorbed material in other techniques pose possibilities of spread of pollutants during transport. Furthermore, the nonbiological methods disrupt the soil characteristics and ecology that make the land unsuitable for agriculture and other purposes.

9.4.2 Bioremediation

9.4.2.1 Phytoremediation

Remediation of soil contaminated with hazardous substances utilizing the innate capabilities of plants is generally termed phytoremediation, an in situ eco-friendly and perpetual approach which does not require any specialized equipments. In addition, application of plants for abatement/rehabilitation of heavy metal-stressed soil is indeed a promising but emerging area of interest because it is an ecologically sound and environmentally safe method for restoration of degraded lands. In this context, about 0.2% angiosperms (Baker and Brooks [1989](#page-11-0)) are reported to tolerate and accumulate excessively high concentrations of metals and are often termed hyperaccumulators. Plants like Alyssum species, Brassica juncea, Arabidopsis halleri, Noccaea sp. (formerly Thlaspi sp.), Viola calaminaria, and Astragalus racemosus are hyperaccumulators. The molecular mechanism underlying hyperaccumulation is attributed to the involvement of metal-specific transporters, chelators such as phytochelatins (PC), metallothioneins (MT), and organic acids (OA) like citrate and antioxidants like glutathione (Kramer [2010](#page-12-0)). Phytoremediation as a technique broadly involves (1) phytoextraction: uptake and accumulation of metals in plants; (2) rhizofiltration: roots absorb, concentrate, or precipitate the metals; (3) phytostabilization: plant reduces the heavy metal mobility by precipitation; (4) phytodegradation: the pollutants are taken up by plants and degraded by the plant enzymes; and (5) phytovolatilization: uptake and release of metals into air as volatile compounds (Raskin and Ensley [2002](#page-13-0)). These characteristics are commonly associated with only hyperaccumulators, and hence, the normal plants are genetically engineered by introducing the genes involved in metal chelation, transport, and stress responses (Susan and D'Souza [2005\)](#page-13-0). For example, engineering of human MT and mouse MT genes in different plants like Arabidopsis, tobacco, and rapeseed plants has shown enhanced Cd uptake and accumulation (Misra and Gedamu [1989](#page-12-0)). Thus, phytoremediation is a promising option, but longer time, climatic conditions, recycling of accumulated plants, and soil characteristics are some of the major constraints. In addition, the use of transgenic plants poses many unanswered ecological questions. Therefore, the better strategy that answers the problems with the above strategies is achieved by rhizoremediation that combines the advantages of plant–microbe symbiosis.

9.4.2.2 Rhizoremediation

Rhizosphere is defined as the soil zone of biological activity around the plant roots, which is the sink of nutrients. In this region, an intense interaction between plant roots and microbes takes place. Microbes inhabiting this area are generally termed rhizobacteria and regulate the biogeochemical cycles, degrade organic materials, and preserve the soil chemistry (Haferburg and Kothe [2007\)](#page-11-0). The proven traits for effective root colonization include the synthesis of the O-antigen of lipopolysaccharide and cellulose, thiamine and biotin production, amino acid synthesis, an isoflavonoid inducible efflux pump, and a nine-polar flagellar arrangement (Lugtenberg et al. [2001\)](#page-12-0). Using in vivo expression technology (IVET), about 20 genes were demonstrated to be induced in root-colonizing pseudomonads (Silby and Levy [2004](#page-13-0)). Around the roots, they form microcolonies, often called biofilms which are covered by mucoid layer (Lugtenberg and Kamilova [2009](#page-12-0)).

One way to reduce the deleterious effects of heavy metals taken up from the environment by some plants involves the use of plant growth-promoting rhizobacteria (Khan et al. [2009](#page-12-0)) and mycorrhizae (Heggo and Angle [1990;](#page-11-0) Saraswat and Rai [2011\)](#page-13-0), and this strategy is termed rhizoremediation. More precisely, the rhizoremediation is defined as the biological treatment of organic or inorganic contaminants in soils by bacterial or fungal activity in the rhizosphere (Kuiper et al. [2004\)](#page-12-0). All through rhizoremediation, low- (e.g., phenolics, organic acid) and high-molecular-weight (e.g., proteins) exudates released from growing plants stimulate the viability and functionality of the plant growth-promoting rhizobacteria (PGPR), which consequently results in a more efficient transformation/degradation of environmental pollutants. The microbial activity also prevents the uptake and accumulation of heavy metals in different organs of plants (Fig. [9.2b](#page-3-0)). In general, the heavy metals are, however, toxic to the microbes, which in turn affect the fertility of soil. As a survival strategy, some microbes have evolved resistance/avoidance mechanisms that cause change in metal speciation (White et al. [1997\)](#page-14-0). The strategies include biosorption of heavy metals by cell walls, polysaccharides, and pigments (Gadd [2009](#page-11-0)) and removal by the efflux pumps and by the synthesis of metal-binding peptides and proteins like MTs (Silver [1996\)](#page-13-0). Thus, the microbial biomass acts as sink for the toxic heavy metals (Gadd [2010](#page-11-0)). In addition, some microbes detoxify the heavy metals like Cd, Hg, and Pb by enzymatic action (Aiking et al. [1985](#page-10-0)). Various studies reported the successful rhizoremediation of heavy metals in soil using heavy metal-resistant rhizobacteria and different plant species (Table [9.2\)](#page-7-0). In heavy metal soil, the metal-resistant rhizobacteria also enhance the mycorrhizal and nodulation efficiency in plants (Vivas et al. [2006\)](#page-14-0). Besides native bacteria, transgenic approaches both at the microbial and plant levels are reported for higher efficiency of rhizoremediation. For example, the expression of metal-binding peptide (EC20) in rhizobacteria Pseudomonas putida 06909 improved both cell growth and cadmium binding in the presence of the cadmium in sunflower seedlings (Wu et al. [2006\)](#page-14-0). It was shown that the introduction of genetically modified microorganisms designed for rhizoremediation induces changes in native bacteria in the rhizosphere but not in the surrounding soil (Carcer et al. [2007\)](#page-11-0).

9.5 Plant Growth-Promoting Activities of PGPR

Besides the heavy metal accumulation or detoxification, the plant-growth-promoting activities of rhizobacteria improve the efficiency of rhizoremediation. The PGPR promote the plant growth by hormonal regulation, enhanced mineral uptake, sequestering iron by siderophore production, antagonism by antibiotics production, and root growth stimulators production (Fig. [9.3\)](#page-8-0) (Khan et al. [2009](#page-12-0); Lugtenberg and Kamilova [2009](#page-12-0); Martínez-Viveros et al. [2010;](#page-12-0) Ahemad and Khan [2011\)](#page-10-0). Studies with metal-sensitive PGPR have also been found effective in preventing the accumulation

and toxicity of heavy metals in plants (Table [9.2\)](#page-7-0). The different PGPR activities are discussed below in correlation with heavy metal toxicity in plants.

9.5.1 Heavy Metal Stress Tolerance

Primarily, heavy metal stress may cause hormonal imbalance in plants, leading to reduced root growth. Ethylene synthesis, for example, is increased upon treatment with Cd, Cu, and Zn. In the case of Cd and Cu, this increase is due to an upregulation of ACC synthase transcription and enhanced activity (Waldemar [2007\)](#page-14-0). The microbial enzyme 1-aminocyclopropane carboxylic acid (ACC) deaminase catabolizes the immediate ethylene precursor ACC (Fig. 9.3), which is released in the root exudates. Thus, rhizobacteria acts as a sink for ACC by stimulating plants to exude more ACC and thereby reducing ethylene stress in plants (Penrose and Glick [2001\)](#page-13-0). On treatment with Cd, the abscisic acid (ABA) hormone content rapidly increased in rice (Oryza sativa) seedlings (Hsu and Kao [2003\)](#page-12-0). Secretion of cytokinin by the microbes decreases the ABA content and its effects (Cowan et al. [1999](#page-11-0)). In addition, PGPR also produce antioxidants like catalases and pyrroloquinoline quinine (PQQ) that helps in degrading the reactive oxygen species (ROS) which is synthesized during stress conditions (Fig. 9.3) (Yang et al. [2009](#page-14-0)).

Fig. 9.3 Plant growth-promoting activities of rhizobacteria

9.5.2 Mineral Uptake

Various studies indicated the reduced uptake of minerals like iron, phosphate, nitrate, and other nutrients by plants grown in soils contaminated with heavy metals (Rubio et al. [1994](#page-13-0); Huang et al. [2007\)](#page-12-0). The deficiency of such elements in plants results in different types of symptoms on plants. For example, leaf chlorosis is one of the key morphological effects of heavy metals in plants due to iron starvation. Siderophores, the low-molecular iron-chelating substances produced by rhizobacteria, help in sequestration of iron by both microbes and plants (Neilands [1995\)](#page-12-0). Siderophore-overproducing mutant of Kluyvera ascorbata SUD165, for example, enhanced the plant growth, chlorophyll contents in foliage, and protein content and decreased the heavy metal accumulation in tomato plants grown in metalcontaminated soil (Burd et al. [2000](#page-11-0)). Siderophore production is reported to be induced by heavy metals like Cd, Pb, Al, and Zn in Pseudomonas and Rhizobium spp. (Roy and Chakrabartty [2000;](#page-13-0) Sinha and Mukherjee [2008;](#page-13-0) Ganesan [2008](#page-11-0)), but the molecular mechanism underlying the synthesis of siderophore is not well explained.

Heavy metal ions also disrupt some of the important plant enzymes like nitrate and nitrite reductases, glutamine synthetase, glutamate synthase, and glutamate dehydrogenase (Llorens et al. [2001\)](#page-12-0), leading to reduced uptake of ammonium and nitrate and low-protein content. This effect was circumvented by PGPR like Rhizobia and diazotrophs capable of producing nitrogenase. Due to these activities, legume–Rhizobia symbiosis has shown higher efficiency in rehabilitation of heavy metal-poisoned soils (Pajuelo et al. [2008;](#page-13-0) Wani et al. [2009](#page-14-0)). Similarly, phosphatesolubilizing microbes and arbuscular mycorrhizal fungi (AMF) enhance the P uptake in plants (Khan et al. [2007](#page-12-0); Zaidi and Khan [2007](#page-14-0); Zaidi et al. [2009;](#page-14-0) Lugtenberg and Kamilova [2009\)](#page-12-0). Besides these, the production of hormones like indole acetic acid (IAA), cytokinin, and other metabolites (Fig. [9.3\)](#page-8-0) promotes the root growth, modifies the root architecture, and induces the membrane transporters, which leads to the enhanced nutrient uptake by plants (Waldemar [2007\)](#page-14-0).

9.6 Eco-economics

In spite of the billions of funding and development of newer technologies and programs aimed at restoring heavy metal-polluted soils, the severity of heavy metal problems is increasing alarmingly every year around the world. This is partly due to the lack of awareness but largely due to economic constraints mostly in developing countries. However, when applied, the comparative estimates and additional factors involved in remediation of metals, for example, cadmium per ton of soil (Glass [1999\)](#page-11-0) employing various approaches, are presented in Table [9.3](#page-10-0). The cost given in this table suggests that the rhizoremediation approach when employed properly with sound understanding is inexpensive. The estimates shown here include only the remediation cost, while the other expenses like cost of transport, recycling, and monitoring may further increase the overall cost of remediation. Further, since

Remediation approaches	Estimates (US \$/ton)	Additional factors and expenses
Land filling	$100 - 500$	Transport/excavation
Vitrification	$75 - 425$	Long-term monitoring
Chemical treatment	$100 - 500$	Recycling
Electrokinetics	$20 - 200$	Monitoring
Phytoremediation	$5 - 40$	Recycling and monitoring
Rhizoremediation	$5 - 20$	Monitoring

Table 9.3 Estimates of cost for cadmium remediation per ton of the soil by different approaches

Adapted from Glass ([1999\)](#page-11-0)

rhizoremediation approach involves the use of cheap renewable resources like PGPR having multiple properties, this technology could be more profitable than other remedial technology. The biocontrol activities like antagonism and competition for nutrients and niches (CNN) (Lugtenberg and Kamilova [2009\)](#page-12-0) add further strength to the economic friendliness of rhizoremediation approach by cutting off the costs for pesticides and thereby circumventing phytopathogens naturally. Thus, rhizoremediation approach is made environmentally as well as economically more pragmatic.

Conclusion

Rhizoremediation approach is aesthetically pleasing and low cost, uses solar energy, requires minimal maintenance, presents no need for further recycling, and preserves the soil fertility and ecology. As a result, this strategy is gaining wider acceptance. Besides remediation and earning, it ensures the food security for humans and prevents them from a lot of ailments. However, large-scale field trials and its assessment are required to guarantee the practicability of rhizoremediation. However, how this technology could be useful in the rehabilitation of metal contaminated but nonagricultural soils with poor nutrients or nutrient deficient soils is indeed a challenge before scientists. Considering different facets of remediation methods, it is evident that all these methods in general provide only a temporary solution for the abatement of polluted lands and not complete destruction of metals from the contaminated sites. Hence, the practice of organic farming along with the remedial technology should be promoted in order to prevent metal pollution in agricultural soil.

References

Ahemad M, Khan MS (2011) Functional aspects of plant growth promoting rhizobacteria: recent advancements. Insight Microbiol 1:39–54

- Aiking HH, Goves H, Riet JV (1985) Detoxification of mercury, cadmium and lead in Klebsiella aerogenes NCTC418 growing in continuous culture. Appl Environ Microbiol 50:1262–1267
- Appenroth K (2010) Definition of "heavy metals" and their role in biological systems. In: Sherameti I, Varma A (eds) Soil heavy metals, vol 19, Soil biology., pp 19–29

Arteca RN, Arteca JM (2007) Heavy metal-induced ethylene production in Arabidopsis thaliana. Plant Physiol 164:1480–1488

- Babula P, Adam V, Opatrilova R, Zehnalek J, Havel L, Kizek R (2008) Uncommon heavy metals, metalloids and their plant toxicity: a review. Environ Chem Lett 6:189–213
- Baker AJM, Brooks RR (1989) Terrestrial higher plants which hyperaccumulate metallic elements–a review of their distribution, ecology and phytochemistry. Biorecovery 1:81–126

Beyersmann D (2002) Effects of carcinogenic metals on gene expression. Toxicol Lett 127:63–68 Burd GI, Dixon DG, Glick BR (2000) Plant growth-promoting bacteria that decrease heavy metal

- toxicity in plants. Can J Microbiol 46:237–245
- Burd GI, Dixon DG, Glick BR (1998) A plant growth promoting bacterium that decreases nickel toxicity in seedlings. Appl Environ Microbiol 64:3663–3668
- Carcer DA, Martin M, Mackova M, Macek T, Karlson U, Rivilla R (2007) The introduction of genetically modified microorganisms designed for rhizoremediation induces changes on native bacteria in the rhizosphere but not in the surrounding soil. ISME 1:205–223
- Clijsters H, Van Assche F (1985) Inhibition of photosynthesis by heavy metals. Photosynth Res 7:31–40
- Cowan A, Cairns A, Bartels-Rahm B (1999) Regulation of abscisic acid metabolism towards a metabolic basis for abscisic acid-cytokinin antagonism. J Exp Bot 50:595–603
- Drum DA (2009) Are toxic biometals destroying your children's future? Biometals 22:697–700
- Dube A, Zbytnieski R, Kowalkowski T, Cukrowska E, Buszewski B (2001) Adsorption and migration of heavy metals in soil. Pol J Environ Stud 10:1–10
- Duffus JH (2002) "Heavy metals" a meaningless term? (IUPAC Technical Report). Pure Appl Chem 74:793–807
- Duruibe JO, Ogwuegbu MOC, Egwurugwu JN (2007) Heavy metal pollution and human biotoxic effects. Int J Phys Sci 2:112–118
- Emmanouil-Nikolussi EN (2007) The role of heavy metals in environmental toxicity and pregnancy outcomes. Reprod Toxicol 24:80
- Faisal M, Hasnain S (2005) Bacterial Cr (VI) reduction concurrently improves sunflower (Helianthus annuus L.) growth. Biotechnol Lett 27:943–947
- Fuhrer J (1982) Ethylene biosynthesis and cadmium toxicity in leaf tissue of beans Phaseolus vulgaris L. Plant Physiol 70:162–167
- Gadd GM (2009) Biosorption: critical review of scientific rationale, environmental importance and significance for pollution treatment. J Chem Technol Biotechnol 84:13–28
- Gadd GM (2010) Metals, minerals and microbes: geomicrobiology and bioremediation. Microbiology 156:609–643
- Ganesan V (2008) Rhizoremediation of cadmium soil using a heavy metal resistant plant growth promoting rhizopseudomonad. Curr Microbiol 56:403–407
- Glass DJ (1999) Economic potential of phytoremediation. In: Raskin I, Ensley BD (eds) Phytoremediation of toxic metals: using plants to clean up the environment. Wiley, New York
- Gupta A, Rai V, Bagdwal N, Geol R (2005) In situ characterization of mercury-resistant growthpromoting fluorescent pseudomonads. Microbiol Res 160:385–388
- Haferburg G, Kothe E (2007) Microbes and metals: interactions in the environment. J Basic Microbiol 47:453–467
- Han FX, Banin A, Su Y, Monts LM, Plodinec MJ, Kingery WL, Triplett GE (2002) Industrial age anthropogenic inputs of heavy metals into the pedosphere. Naturwissenshaften 89:497–504
- Hare PD, Cress WA, van Staden J (1997) The involvement of cytokinins in plant responses to environmental stress. Plant Growth Regul 23:79–103
- Heggo A, Angle JS (1990) Effects of vesicular-arbuscular mycorrhizal fungi on heavy metal uptake by soybeans. Soil Biol Biochem 22:865–869
- Hemalatha S, Anburaj A, Francis K (1997) Effect of heavy metals on certain biochemical constituents and nitrate reductase activity in Orzya sativa L. seedlings. J Environ Biol 18:313–319
- Hong S, Candelone J, Boutron CF (1996) Deposition of atmospheric heavy metals to the Greenland ice sheet from the 1783–1784 volcanic eruption of Laki, Iceland. Earth Planet Sci Lett 144:605–610
- Howlett NG, Avery SV (1997) Induction of lipid peroxidation during heavy metal stress in Saccharomyces cerevisiae and influence of plasma membrane fatty acid unsaturation. Appl Environ Microbiol 63:2971–2976
- Hsu YT, Kao CH (2003) Role of abscisic acid in cadmium tolerance of rice (Oryza sativa L.) seedlings. Plant Cell Environ 26:867–874
- Huang YZ, Wei K, Yang J, Dai F, Zhang GP (2007) Interaction of salinity and cadmium stresses on mineral nutrients, sodium, and cadmium accumulation in four barley genotypes. J Zhejiang Univ Sci B 8:476–485
- Indian Council of Medical Research (2003) ICMR, New Delhi, India
- Kelly JJ, Haggblom M, Tatelll RL (1999) Effects of the land application of sewage sludge on soil heavy metal concentrations and soil microbial communities. Soil Biol Biochem 31:1467–1470
- Khan MS, Zaidi A, Wani PA, Ahemad M, Oves M (2009) Functional diversity among plant growth-promoting rhizobacteria. In: Khan MS, Zaidi A, Musarrat J (eds) Microbial strategies for crop improvement. Springer, Heidelberg, pp 105–132
- Khan MS, Zaidi A, Wani PA (2007) Role of phosphate solubilizing microorganisms in sustainable agriculture—a review. Agron Sustain Dev 27:29–43
- Koo SY, Cho KS (2009) Isolation and characterization of plant growth-promoting rhizobacterium, Serratia sp., SY5. J Microbiol Biotechnol 19:1431–1438
- Kramer U (2010) Metal hyperaccumulation in plants. Annu Rev Plant Biol 61:517–534
- Kuiper I, Lagendijk EL, Bloemberg GV, Lugtenberg BJJ (2004) Rhizoremediation: a beneficial plant-microbe interaction. Mol Plant Microbe Interact 17:6–15
- Lambert M, Leven BA, Green RM (2000) New methods of cleaning up heavy metal in soils and water. Environmental science and technology briefs for citizens. Kansas State University, Manhattan, KS
- Llorens N, Arola L, Blade C, Mas A (2001) Effects of copper exposure upon nitrogen metabolism in tissue cultured Vitis vinifera. Plant Physiol 160:159–163
- Lugtenberg BJJ, Dekkers L, Bloemberg GV (2001) Molecular determinants of rhizosphere colonization by Pseudomonas. Annu Rev Phytopathol 39:461–490
- Lugtenberg BJJ, Kamilova F (2009) Plant-growth-promoting rhizobacteria. Annu Rev Microbiol 20:541–546
- Madhaiyan N, Poonguzhali S, Sa T (2007) Metal tolerating methylotropic bacteria reduces nickel and cadmium toxicity and promotes plant growth of tomato (Lycopersicon esculentum L.). Chemosphere 69:220–228
- Marshall F, Agarwal R et al (2003) Heavy metal contamination of vegetables in Delhi. Executive summary of technical report. Imperial College, London
- Martínez-Viveros O, Jorquera MA, Crowley DE, Gajardo G, Mora ML (2010) Mechanisms and practical considerations involved in plant growth promotion by rhizobacteria. J Soil Sci Plant Nutr 10:293–319
- McGrath D, Tunney H (2010) Accumulation of cadmium, fluorine, magnesium, and zinc in soil after application of phosphate fertilizer for 31 years in a grazing trial. J Plant Nutr Soil Sci 173:548–553
- Misra S, Gedamu L (1989) Heavy metal tolerant transgenic Brassica napus L. and Nicotiana tobaccum L plant. Theor Appl Genet 78:16-18
- Mortvedt JJ (1996) Heavy metal contaminants in inorganic and organic fertilizers. Fertil Res 43:55–61
- Manios T, Stentiford EI, Millner P (2002) The effect of heavy metals on the total protein concentration of Typha latifolia plants, growing in a substrate containing sewage sludge compost and watered with metalliferous wastewater. J Environ Sci Health A Tox Hazard Subst Environ Eng 37:1441–1451
- Mulligan CN, Yong RN, Gibbs BF (2001) An evaluation of technologies for the heavy metal remediation of dredged sediments. J Hazard Mater 85:145–163
- Neilands JB (1995) Siderophores: structure and function of microbial iron transport compounds. J Biol Chem 270:26723–26726

Nriagu JO (1996) A history of global metal pollution. Science 272:223–224

- Pandey P, Tripathi AK (2011) Effect of heavy metals on morphological and biochemical characteristics of Albizia procera (Roxb.) Benth. seedlings. Int J Environ Sci 5:1009-1018
- Pajuelo E, Dary M, Palomares AJ, Rodriguez-Llorente ID, Carrasco JA, Chamber MA (2008) Biorhizoremediation of heavy metals toxicity using rhizobium-legume symbiosis, Biological nitrogen fixation: towards poverty alleviation through sustainable agriculture. Springer, New York
- Penrose DM, Glick BR (2001) Levels of 1-aminocylcopropane carboxylic acid (ACC) in exudates and extracts of canola seeds treated with plant growth promoting rhizobacteria. Can J Microbiol 47:368–372
- Priti B, Chaturvedi AK, Prasad P (2009) Effect of enhanced lead and cadmium in soil on physiological and biochemical attributes of Phaseolus vulgaris L. Nat Sci 7:63–75
- Raskin I, Ensley BD (eds) (2002) Phytoremediation of toxic metals using plants to clean the environment. Wiley, New York
- Reichman SM (2007) The potential use of the legume-*Rhizobium* symbiosis for the remediation of arsenic contaminated sites. Soil Biol Biochem 39:2587–2593
- Rivetta A, Negrini N, Cocucci M (1997) Involvement of Ca^{2+} -calmodulin in Ca^{2+} toxicity during the early phases of radish (Raphanus sativus L.) seed germination. Plant Cell Environ 20:600–608
- Roy N, Chakrabartty K (2000) Effect of aluminium on the production of siderophore by Rhizobium sp. (Cicer arietinum). Curr Microbiol 41:5–10
- Rubio MI, Escrig I, Martinez-Cortina C, Lopez-Benet FJ, Sanz A (1994) Cadmium and nickel accumulation in rice plants. Effects on mineral nutrition and possible interactions of abscisic acid and gibberellic acids. Plant Growth Nutr 14:151–157
- Saraswat S, Rai JPN (2011) Mechanism of metal tolerance and detoxification in mycorrhizal fungi. In: Khan MS, Zaidi A, Goel R, Musarrat J (eds) Biomanagement of metal-contaminated soils. Springer, The Netherlands, pp 225–240
- Shcherbov BL, Zavgorodnyaya NV, Lazareva EV (2008) Ecogeochemical consequences of forest fires in Belt Pine forests of Altai Krai. Cont Prob Ecol 1:459–466
- Silby MW, Levy SB (2004) Use of in vivo expression technology to identify genes important in growth and survival of Pseudomonas fluorescens Pf0-1 in soil: discovery of expressed sequences with novel genetic organization. J Bacteriol 186:7411–7419
- Silver S (1996) Bacterial heavy metal resistance: new surprises. Annu Rev Microbiol 50:753–789
- Sinha S, Mukherjee SK (2008) Cadmium-induced siderophore production by a high Cd-resistant bacterial strain relieved Cd toxicity in plants through root colonization. Curr Microbiol 56:55–60
- Somasundaram R, Muthuchelian K, Murugesan S (1994) Inhibition of chlorophyll, protein, photosynthesis, nitrate reductase and nitrate content by vanadium in Oryza sativa L. J Environ Biol 15:41–48
- Stobrawa K, Lorenc-Plucin´ska G (2007) Changes in carbohydrate metabolism in fine roots of the native European black poplar (Populus nigra L.) in a heavy-metal-polluted environment. Sci Total Environ 373:157–165
- Sugita M (1978) The biological half-time of heavy metals. Int Arch Occup Environ Health 41:25–40
- Susan E, D'Souza SF (2005) Prospects of genetic engineering of plants for phytoremediation of toxic metals. Biotechnol Adv 23:97–114
- Van Assche FJ (1998) A stepwise model to quantify the relative contribution of different environmental sources to human cadmium exposure. Proceedings of the 8th international nickel-cadmium battery conference, Prague, Czech Republic, 21–22 Sept 1998
- Veselov D, Kudoyarova G, Symonyan M, Veselov St. (2003) Effect of cadmium on ion uptake, transcription and cytokinin content in what seedlings. Bulg J Plant Physiol Special Issue:353–359
- Vincent JB (2000) Elucidating a biological role for chromium at a molecular level. Acc Chem Res 33:503–510
- Vivas A, Biro B, Ruiz-Lozano JM, Barea JM, Azcon R (2006) Two bacterial strains isolated Zn-polluted soil enhance plant growth and mycorrhizal efficiency under Zn-toxicity. Chemosphere 62:1523–1533
- Waldemar M (2007) Signaling responses in plants to heavy metal stress. Acta Physiol Plant 29:177–187
- Wani PA, Khan MS (2010) Bacillus species enhance growth parameters of chickpea (Cicer arietinum L.) in chromium stressed soils. Food Chem Toxicol 48:3262–3267
- Wani PA, Zaidi A, Khan MS (2009) Chromium reducing and plant growth promoting potential of Mesorhizobium species under chromium stress. Bioremediation J 13:121–129
- Weast RC (1984) Handbook of chemistry and physics, 64th edn. CRC, Boca Raton, FL
- White C, Sayer JA, Gadd GM (1997) Microbial solubilization and immobilization of toxic metals: key biogeochemical processes for treatment of contamination. FEMS Microbiol Rev 20:503–516
- World Health Organization (1987) Air Quality Guidelines. Copenhagen: WHO Regional Office for Europe
- Wong PK (1988) Mutagenicity of heavy metals. Bull Environ Contam Toxicol 40:597–603
- Wu CH, Wood TK, Mulchandani A, Chen W (2006) Engineering Plant-microbe symbiosis for rhizoremediation of heavy metals. Appl Environ Microbiol 72:1129–1134
- Yang J, Kloepper JW, Ryu CM (2009) Rhizosphere bacteria help plants tolerate abiotic stress. Trends Plant Sci 14:1–4
- Zaidi A, Khan MS, Ahemad M, Oves M (2009) Plant growth promotion by phosphate solubilizing bacteria. Acta Microbiol Immunol Hung 56:263–284
- Zaidi A, Khan MS (2007) Stimulatory effects of dual inoculation with phosphate solubilizing microorganisms and arbuscular mycorrhizal fungus on chickpea. Aust J Exp Agric 47:1016–1022
- Zhang Q, Wang GH, Feng YK, Sun QZ, Witt C, Dobermann A (2006) Changes in soil phosphorus fractions in a calcareous paddy soil under intensive rice cropping. Plant Soil 288:141–154