Fast-Acting Topical Hydrophilic Drug Delivery via a Natural Nano-Injection System

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21.1 Introduction

Transdermal drug delivery systems provide attractive solutions for local and systemic drug delivery, with advantages over drug delivery via conventional oral or hypodermic administration. The benefits include avoidance of first pass metabolism, elimination of drastic drug fluctuations, prevention of adverse side effects, and a general increase in patient compliance. Nevertheless, after three decades of extensive research only a relatively small number of drugs can be introduced transdermally (Prausnitz and Langer [2008](#page-6-0); Subedi et al. [2010\)](#page-7-0). The main barrier to topically applied medications is the outermost layer of the skin, the stratum corneum (SC), which permits penetration and delivery of small lipophilic drugs, but not of compounds that are large or hydrophilic (Prausnitz and Langer [2008;](#page-6-0) Cevc and Vierl [2010;](#page-6-1) Barry [2004\)](#page-6-2). A variety of vehicles and technologies have been developed to penetrate the skin barrier, allowing transcutaneous drug delivery (Brown et al. [2006](#page-6-3); Cevc and Vierl [2010\)](#page-6-1). The two principal approaches adopted to enhance skin permeability and penetration of actives are based either on chemical passive enhancers or on physical active or energetic devices. Passive technologies have the advantage of high patient compliance but usually require a long application time because of slow absorption (Baroli [2010;](#page-6-4) Elsayed et al. [2007\)](#page-6-5), whereas active methods rapidly permeate the

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skin barrier but necessitate professional administration and are expensive (Ogura et al. [2008;](#page-6-6) Prausnitz [2004;](#page-6-7) Charoo et al. [2010\)](#page-6-8). In this chapter we describe biologically derived microdevices that are applied in topical formulations and which, when activated, inject the desired compound into the epidermis. These natural micro-devices integrate the benefits of the chemical passive and physical active approaches by conferring the dual advantages of ease of application and rapid delivery. Furthermore, the system is specifically designed to deliver hydrophilic compounds.

21.2 The Biological Micro-device System

The micro-devices used for drug delivery are isolated from sea anemones that belong to the phylum Cnidaria, which includes hydra, corals, sea anemones, and jellyfish, and dates back about 700 million years (Park et al. [2012](#page-6-9)). Cnidarians have a simple body structure, containing a large population of stem cells that provide them with almost unlimited ability for regeneration, reminiscent of the Greek mythological hydra with constantly growing heads (Bosch et al. [2010\)](#page-6-10). The most characteristic features of the cnidarians are their stinging mechanisms, which are located within stinging cells and are constantly renewed. The stinging mechanisms are composed of microcapsules incorporating tightly folded submicron- to nano-sized injectors. The microcapsular wall and the nano-injector, which is a continuation of the microcapsule, are made of fibrils composed mainly of two proteins. One, known as mini-collagen, is similar to collagen but is much shorter, and the other is highly enriched in cysteine (Engel et al. [2002\)](#page-6-11). These two proteins are cross-linked with disulfide bonds to create a condensed micro-device structure with a tensile strength almost as high as that of steel (Holstein et al. [1994;](#page-6-12) Ozbek et al. [2002\)](#page-6-13). Owing to its fibrillary structure, the microcapsule wall is permeable to solutions and essentially performs like a porous net. Upon activation of the microcapsule, water flows through it, and a high

internal pressure of 150 bars is rapidly developed, resulting in discharge of the long, thin, folded injector at an ultrafast acceleration of 5*106 G (Nüchter et al. [2006](#page-6-14)) (Fig. [21.1](#page-2-0)). The discharge is controlled by osmotic balance. The water flowing through the porous wall of the activated microcapsule causes dissociation of the main matrix of the microcapsules consisting of a large aggregated of poly-*γ*-glutamate trapped with cationic metal inside the microcapsule (Ayalon et al. [2011](#page-6-15); Weber [1990\)](#page-7-1). This in turn increases the osmotic pressure, resulting in ejection of the nano-injector and continuous injection of the microcapsule content till the poly-*γ*glutamate is fully swept out from the microcapsule (Szczepanek et al. [2002](#page-7-2)). The system is capable of puncturing keratinous tissue such as skin, hair, and nail plate and immediately delivering the microcapsule content (Lotan [2008\)](#page-6-16). About 30 subtypes of microcapsules are known; they differ in size and shape, but all function according to the same basic principle (Tardent [1995\)](#page-7-3). In the following section, we elaborate on the microcapsule type that is derived from the sea anemone *Aiptasia diaphana* and can be used for drug delivery.

21.3 Harnessing a Biological System for Drug Delivery

More than 10,000 species of Cnidaria contain nano-injection systems. We were interested in microcapsules with relatively short nanoinjectors that would be able to penetrate the SC and enter only the upper part of the viable epidermis. We therefore chose sea anemone microcapsules, which contain smooth nano-injectors 50 μm in length. These microcapsules are isolated, purified, and desiccated, all under sterile conditions, yielding a powder of intact microdevices whose potential activation is unimpaired (Ayalon et al. [2011\)](#page-6-15), and which, when formulated as an anhydrous gel, can be spread over the skin. Activation occurs only when the gel formulation is hydrated by the addition of a hydrophilic formulation containing the drug (Fig. [21.1\)](#page-2-0). Thus, administration of the drug comprises two steps:

first, spreading of the anhydrous gel formulation containing the micro-device system over the skin and, second, application of the hydrophilic drug formulation. Because activation is immediate, the drug is delivered to the epidermis in less than 5 min, and owing to its short and thin nano-sized dimensions, the delivery is virtually noninvasive (see movie in (Ayalon et al. [2011\)](#page-6-15)).

21.4 Delivery of Hydrophilic Compounds

Owing to the characteristics of the skin, hydrophilic compounds are the most difficult molecules to deliver by passive chemical application. The nano-injection system is specifically designed to address this obstacle and hence to facilitate the delivery mainly of hydrophilic compounds. We have previously shown that various compounds, including organic compounds and peptides, can be delivered, via our micro-devices, locally into the skin (Ayalon et al. [2011;](#page-6-15) Lotan [2005,](#page-6-17) [2008\)](#page-6-16), or systemically (Shaoul et al. [2012\)](#page-6-18). The unique delivery apparatus is described and illustrated in the next section.

21.4.1 Kinetics of Drug Release

Activation of the anhydrous gel containing the nano-injection system is immediate when combined with a hydrophilic drug. The applied soluble drug is pumped into the microcapsule and

Fig. 21.2 (**a**) Kinetics of 5% lidocaine hydrochloride, 5% trolamine salicylate, and 5% nicotinamide through the full-thickness skin of nude mice, assessed in a Franz diffusion cell. The microcapsules were applied in gel formulation containing 2% hydroxypropyl cellulose in absolute ethanol over the skin, and the test drugs were placed over the microcapsule-containing gel for 5 min. The skin

was then thoroughly washed and samples were taken between 0.5 and 24 h. (**b**) Accumulation of delivered compounds indicated in (**a**), measured 24 h after application of the microcapsule-containing gel (*black bar*) or of the gel formulation without microcapsules (*gray bar*). Error bars represent means±SD

through the nano-injector to its target (Fig. [21.1\)](#page-2-0). This process continues until the poly-*γ*-glutamate driving force of the system is washed out through the nano-injector. As the entire process takes less than a second, we studied the kinetics of the system over a short period (5 min) of exposure to a test drug delivery, after which the drug was removed from the skin by thorough washing. This brief period, although considerably longer than the time required for activation of the system, was chosen because of technical limitations. Using a Franz diffusion cell system, we carried out in vitro measurements of the permeability and kinetics of various test compounds in a sample of full-thickness skin from nude mice. Figure [21.2a](#page-3-0) demonstrates the delivery kinetics of three hydrophilic drugs (lidocaine hydrocloride, triethanolamine HCl, trolamine salicylate and nicotinamide), all of which are used in topical formulations for various therapeutic purposes such as local anesthesia and arthritic pain as well as in skin cosmetics but which usually necessitate prolonged application to permeate the skin. Exposure of the skin for 5 min to these drugs after applying the gel formulation containing the biological micro-devices resulted, in each case, in the successful permeation through the full-thickness skin of 40% of the delivered drug as early as 30 min after application and of more than 90% at 5 h. Control experiments carried out under identical conditions but without the

micro-devices resulted in permeation of negligible amounts (Fig. [21.2b](#page-3-0)). Evidently, therefore, the rapid delivery obtained with the biological microdevices is not typical for passive topical or transdermal formulations but is rather an outcome of the active mode of penetration via nano-injection.

21.4.2 Dose Control in a Peptide Model

The quantity of drug delivered by the nanoinjection system can be controlled by regulating three basic parameters: drug concentration, size of application area, and number of microcapsules. Common to most transdermal formulations including the nano-injection system are the first two parameters, resulting in an increased drug delivery as a function of drug concentration and application area size (Ayalon et al. [2011](#page-6-15); Lotan [2008\)](#page-6-16). The unique feature of the nano-injection system is that, as with fixed fabricated microneedles, the number of penetration points can be controlled. However, unlike in the case of microneedle use, the concentration of microcapsules in the anhydrous gel formulation that is spread over the skin can be varied.

Our experimental results of topical application of a hydrophilic peptide demonstrate the effect of microcapsule number on drug delivery.

Fig. 21.3 Accumulation of desmopressin acetate delivered to the full-thickness skin of nude mice (*black squares*) or to hairless guinea pig epidermis (*gray triangles*), as a function of microcapsule number. Desmopressin (5% solution) was added to gel formulations containing different numbers of microcapsules per square centimeter of skin or, as a control, to the same gel formulation without microcapsules (0 microcapsules). After exposure to desmopressin for 5 min, the drug was removed from the skin by thorough washing, and the skin samples were left in the diffusion cell for 24 h to allow continuing subcutaneous diffusion of the delivered desmopressin. Error bars represent means±SD

Peptides and proteins are relatively limited in their delivery through the SC, and many advanced formulations and physical techniques are being developed for that purpose, only a few of which are currently on the market (Kalluri and Banga [2011](#page-6-19)). We tested delivery of desmopressin acetate, a 9-amino-acid synthetic replacement for the hormone vasopressin. This peptide is relatively stable and can be traced under the skin (Cormier et al. [2004\)](#page-6-20). Our nano-injection system, in which different numbers of microcapsules in a gel formulation were combined with 5% desmopressin solution, was applied to the full-thickness skin of nude mice for 5 min. We found that the amount of drug delivered was proportional to the number of microcapsules up to approximately 1×10^6 microcapsules per square centimeter of skin (Fig. [21.3\)](#page-4-0). In other experiments (Ayalon et al. [2011](#page-6-15)) we found that above this concentration, there was a reduction in the amount of drug delivered, suggesting that the ongoing increase in microcapsule density probably resulted in microcapsules overload, preventing their optimal contact with the skin. The nude mouse skin used in the above experiments is a convenient and readily available model for percutaneous penetration (Simon and Maibach [1998\)](#page-6-21); nevertheless, to

verify that successful application is not limited to the anatomy of the mouse skin, we also tested it on the epidermis of the rodent model that most closely resembles human skin, namely, that of the hairless guinea pig (Sueki et al. [2000](#page-7-4)). The results were similar to those obtained in nude mice, showing that the system is not restricted to a particular model and suggesting that it can be successfully applied to different skin anatomies (Fig. [21.3\)](#page-4-0). Thus, these experiments confirmed that the delivered dose can be controlled by altering microcapsule density and that the system is compatible with peptide drugs. These findings offer promising possibilities for biopharmaceutical drug delivery.

21.4.3 Systemic Delivery

To demonstrate systemic delivery, we used the potent muscarinic receptor antagonist, scopolamine. This naturally occurring alkaloid is one of the most effective single agents used to prevent motion sickness (Spinks and Wasiak [2007](#page-7-5)) and is also commonly used for the prevention of postoperative nausea and vomiting (Apfel et al. [2010](#page-6-22)). It was one of the first drugs to be incorporated, into patches for transdermal delivery. However, given the slow incremental diffusion of this drug, the patch has to be applied up to 8 h in advance of need, a major treatment disadvantage (Renner et al. [2005\)](#page-6-23). Because the activity of the micro-injection system is immediate, we expected that its use would enable faster drug delivery resulting in rapidly detectable plasma drug levels. To test this hypothesis, we used a porcine model, known to be particularly reliable for transdermal research in vivo, as comprehensively reviewed by Simon and Maibach (Simon and Maibach [2000](#page-7-6)). In vivo exposure of the porcine skin to a topical gel consisting of microdevices combined with 5% scopolamine hydrobromide was limited to 5 min, after which the sites of application were thoroughly washed, and blood samples were collected. The results showed that the nano-injection system had facilitated rapid accumulation of scopolamine in the plasma, with a time to peak concentration

(Tmax) of 30 min and a peak plasma concentration (Cmax) up to five times higher than that of control pigs exposed to similar topical treatment but without micro-devices (Fig. [21.4](#page-5-0)). Thus, despite its topical formulation, the pharmacokinetic characteristics of the drug in the presence of the micro-devices were comparable to those reported for its subcutaneous injection (Renner et al. [2005\)](#page-6-23).

Fig. 21.4 Plasma levels of scopolamine in pigs after brief topical exposure. Test $(n=4)$ and control $(n=3)$ groups were exposed for 5 min to the same solution of 5% scopolamine hydrobromide. The graph shows averaged values (means+SD) of the test group (*black squares*) treated with gel formulation containing microcapsules and of the control group (*open triangles*) treated with gel formulation only (Modified from Shaoul et al. ([2012\)](#page-6-18))

21.5 Discussion

The skin is the most accessible site for targeted topical and systemic delivery of drugs. Nevertheless, the intrinsic structure and lipophilicity of the SC serve as an almost impenetrable barrier for the transdermal route to hydrophilic compounds. The biological micro-device system can overcome this barrier because the delivery through the SC is not dependent on passive diffusion, but rather on active penetration via the hollow nano-injectors directly into the epidermis. The penetrating nano-injectors are relatively short, up to 50 μm long, and can therefore only transverse the SC, delivering the drug to the upper viable epidermis (Fig. [21.5](#page-5-1)) creating submicron penetration points. These physical characteristics make the system almost noninvasive in comparison to other devices such as sono/electroporation, which creates entry points of more than 200 μm in pore diameter, or minimally invasive microneedles with varied diameter of $25-200 \mu m$ and length of up to 1 mm, which can reach to the dermis (Donnelly et al. [2010](#page-6-24); Cevc and Vierl [2010](#page-6-1); van der Maaden et al. [2012\)](#page-7-7). Safety trials on more than 100 human volunteers have demonstrated no irritation or allergic potential caused by the nano-injection system, and

Fig. 21.5 Application of the micro-device system in comparison to application by a regular syringe. The microcapsules are topically activated to inject their nano-injectors through the SC. Because of the dimensions of the nano-injectors, their skin penetration is limited mainly to the SC and the upper viable epidermis, whereas the regular syringe penetrates the skin layers and reaches deep into the subcutaneous layer

studies in pigs showed that the nano-injectors can be wiped off the skin after application (Lotan [2005](#page-6-17), [2008\)](#page-6-16). The use of this biological system as a drug delivery platform allows rapid topical application that is self-activated via internal osmotic pressure, without the need to apply any external energy device. After application the system can be removed by wiping, as the delivery process is by then already completed. The system can be used not only for local treatments but also, depending on the characteristics of the drug, for systemic delivery. As it is specially designed for delivery of hydrophilic organic compounds and peptides, further studies will be required to investigate its potential use for delivery of large proteins. The findings to date suggest that this 700-million-year-old natural system can be exploited as a novel technique for the fast-acting delivery of therapeutic drugs.

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