Chapter 6 Biofunctionalization of Metals with Polymers

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Abstract If the advantages of metals and polymers are mixed and disadvantages are eliminated by manufacturing metal—polymer composite materials, humankind will obtain ideal materials having excellent mechanical properties and biofunctions. Two kinds of metal—polymer composite materials are feasible to design: one is a combination of bulk polymeric materials and bulk metallic materials and the other is immobilization of polymers to metal surfaces. In this chapter, the above metal—polymer composite materials for biomedical use are demonstrated, and the corresponding researches are reviewed, including chemical immobilization and electrodeposition of biofunctional polymer; immobilization of biomolecules such as peptide, protein, collagen, hydrogel, and gelatin; bonding of polymers with metal through silane-coupling agent; and polymers condensed in porous titanium.

Keywords Biofunctional polymer • Immobilization • Electrodeposition • Antibacterial property • Platelet adhesion • Bone formation • Soft tissue compatibility

6.1 Introduction

Metals show high strength, elongation, fracture toughness, and durability, which are specific advantages against ceramics and polymers. Therefore, metals are widely used for medical devices especially implants (devices inserted in the human tissues) to prevent the fracture or degradation of devices during use. However, metals are typically artificial materials and have no biofunction, which makes them fairly unattractive as biomaterials.

On the other hand, polymers are widely used as biomaterials because of their high degree of flexibility, biocompatibility, and technologic properties [1]. Also, it is relatively easy to design biofunctional polymers based on biomimic techniques, because biofunctional polymers exist in the human body as parts of biomolecules, cells, tissues, and organs.

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Therefore, some of polymeric materials are widely known as biocompatible and biofunctional materials. The fast technological evolution of polymers has made it possible to apply these materials to medical devices over the last three decades; in fact, many devices made from metals have been replaced by others made from polymers. In spite of this fact, about 80 % of implant devices are still made from metals, and this percentage remains unchanged because of their high strength, toughness, and durability. Medical devices consisting of metals cannot be replaced by polymers at present.

If the above advantages of metals and polymers are mixed and disadvantages are eliminated by manufacturing metal–polymer composite materials, humankind will obtain ideal materials having excellent mechanical properties and biofunctions. Two kinds of metal–polymer composite materials are feasible to design: one is a combination of bulk polymeric materials and bulk metallic materials and the other is immobilization of polymers to metal surfaces. In this chapter, the above metal– polymer composite materials for biomedical use are demonstrated, and the corresponding researches are reviewed.

6.2 Immobilization of Poly(ethylene glycol)

6.2.1 Chemical Immobilization

The immobilization of biofunctional polymers on noble metals such as Au is usually conducted using the bonding –SH or –SS– group; however, this technique can only be used for noble metals. The adhesion of platelets and adsorption of proteins, peptides, antibodies, and DNA are controlled by modifications of the above technique.

On the other hand, poly(ethylene glycol) (PEG) is a biofunctional molecule on which adsorption of proteins is inhibited [2]. Therefore, immobilization of PEG to metal surface is an important event to biofunctionalize the metal surface. A class of copolymers based on poly(L-lysine)-g-poly(ethylene glycol), PLL-g-PEG, has been found to spontaneously adsorb from aqueous solutions onto TiO₂, Si_{0.4}Ti_{0.6}O₂, and Nb₂O₅ to develop blood-contacting materials and biosensors [3, 4]. In another case, TiO₂ and Au surfaces are functionalized by the attachment of poly(ethylene glycol)-poly(DL-lactic acid), PEG-PLA, and copolymeric micelles. The micelle layer can enhance the resistance to protein adsorption to the surfaces up to 70 % [5]. Surface modification of stainless steel by grafting of poly(ethylene glycol) was reported by Zhang et al. [6]. In their method, the surface of stainless steel was first modified by a silane-coupling agent (SCA), (3-mercaptopropyl) trimethoxysilane. The surface of the silanized stainless steel (SCA-SS) was subsequently activated by argon plasma and then subjected to UV-induced graft polymerization of poly(ethylene glycol)methacrylate (PEGMA). The PEGMA graftpolymerized stainless-steel coupon (PEGMA-g-SCA-SS) has a high graft concentration, and the high PEG content was found to be very effective to prevent the absorption of bovine serum albumin and γ -globulin. These processes require several steps but are effective for immobilization; however, no promising technique for the immobilization of PEG to a metal surface has been so far developed. Photoreactive PEG is photoimmobilized on titanium (Ti) [7].

6.2.2 Electrodeposition

Both terminals of PEG (MW: 1,000) are terminated with -NH2 (NH2-PEG-NH2). The cathodic potential is charged to Ti from the open-circuit potential to -0.5 V vs. a saturated calomel electrode (SCE) and is maintained at this potential for 300 s. During charging, the terminated PEGs electrically migrate to and are immobilized on the Ti cathode, as shown in Fig. 6.1a. Not only electrodeposition but also immersion leads to the immobilization of PEG onto a Ti surface. However, more terminated amines combine with Ti oxide via an NH-O bond by electrodeposition, while more amines randomly exist as NH₃⁺ in the PEG molecule by immersion (Fig. 6.1b) [8, 9]. An atomic force microscopic image is also shown in Fig. 6.1c. The amounts of the PEG layer immobilized onto the metals are governed by the concentrations of the active hydroxyl groups on each surface oxide in the case of electrodeposition, which is governed by the relative permittivity of the surface oxide in the case of immersion [10]. The PEG-immobilized surface inhibits the adsorption of proteins and cells, as well as the adhesion of platelets [11] and bacteria [12] (Fig. 6.2), indicating that this electrodeposition technique is useful for the biofunctionalization of metal surfaces. It is also useful for all electroconductive materials and materials even having complex surface topography.

6.3 Immobilization of Biomolecules

6.3.1 Immobilization of Biomolecules

Immobilization of biomolecules is a natural approach to obtain cell and tissue compatibility and biofunctions. It has been found that these coatings act as local mediators of cell adhesion and, in consequence, as a stimulating factor for the growth and proliferation of the cells normally found around the substituted tissue. The tight attachment at the surface of the metallic implant and the conservation of the biological function of the proteins involved are prerequisites for obtaining these highly desirable properties. On the other hand, organic coatings have been scarcely used on metallic implants, because manufacturers hesitate to commercialize the



Fig. 6.1 Immobilization of NH₂–PEG–NH₂ by electrodeposition. The terminated PEGs electrically migrate to and are immobilized on the Ti cathode (**a**). More terminated amines combine with Ti oxide via an NH–O bond by electrodeposition, while more amines randomly exist as NH_3^+ in the PEG molecule by immersion (**b**). An atomic force microscopic image of the PEG-immobilized Ti surface (**c**)

biomolecules immobilized by metallic implant due to the problems on storage, safety, sterilization, etc.

The immobilization of biomolecules on the metallic surface can be achieved using self-assembled monolayers as cross-linkers. Self-assembled monolayers provide chemically and structurally well-defined surfaces that can often be manipulated using standard synthetic methodologies [13]. Thiol on self-assembled



Fig. 6.2 Inhibition of platelet adhesion and biofilm formation on Ti through immobilization of $NH_2-PEG-NH_2$ by electrodeposition. (a) Platelet adhesion and fibrin network on untreated Ti, (b) inhibition of platelet adhesion on PEG-immobilized Ti by electrodeposition, (c) biofilm formation on untreated Ti, and (d) inhibition of biofilm formation on PEG-immobilized Ti by electrodeposition

monolayers [14, 15] and siloxane-anchored self-assembled monolayers [16] has been particularly well studied. A problem related to the application of immobilized biomolecules via silanization techniques is the hydrolysis of siloxane films when exposed to aqueous (physiological) conditions [17]. More recently, alkyl phosphate films that retain robust under physiological conditions [18] have been used to provide an ordered monolayer on tantalum oxide surfaces [19, 20], and alkalphosphonic acids have been used to coat the native oxide surfaces of metals and their alloys inducing iron [21], steel [22], and Ti [23].

6.3.2 Peptide

In a living tissue, the most important role played by the ECM has been highlighted to favor cell adhesion [24]. Studies have shown that interactions occur between cell

membrane receptors and adhesion proteins derived from the bone matrix, such as type I collagen or fibronectin [25]. These proteins are characterized by a RGD (Arg-Gly-Asp) motif which specially transmembrane connections between the actin cytoskeleton and the RGD motif, and the whole system can activate several intracellular signaling pathways modulating cell behavior (e.g., proliferation, apoptosis, shape, mobility, gene expression, and differentiation) [26].

Due to the main role of the RGD sequence in cell adhesion, several research groups have developed biofunctionalized surfaces by immobilization of RGD peptides. Grafting RGD peptides has been performed on different biomaterials. such as Ti [27-29], and has been shown to improve osteoconduction in vitro. Methodologies differ by the conformation of RGD and by the technique used for peptide immobilization [24, 25, 28–30]. Since the graft of an RGD peptide is known to be efficient in bone reconstruction [31], the challenge is to develop simple and cheap methods to favor cell anchorage on biomaterial surfaces [29, 30]. Selfassembled molecular monolayers bearing RGD moieties have been grafted to numerous surfaces, using either silanes [32], phosphonates on oxidized surfaces [29], or thiols on Au [30], but have encountered some application problems for large-scale production. Phosphonates are known to adsorb on Ti. To be mechanically and physiologically stable, phosphonate layers have to be covalently bound to the material surface using drastic conditions [33, 34] which are not compatible with biomolecule stability. Monolayers of RGD phosphonates have been achieved using a complex multistep process which necessitates to tether a primer onto Ti surface, then a linker, and finally the peptide [35]. To immobilize RGD to the electrodeposited PEG on Ti, PEG with an -NH₂ group and a -COOH group (NH₂-PEG-COOH) must be employed. One terminal group, -NH₂, is required to bind stably with a surface oxide on a metal. On the other hand, the other terminal group, -COOH, is useful to bind biofunctional molecules such as RGD as shown in Fig. 6.3 [36]. This RGD/PEG/Ti surface accelerates the calcification by MC3T3-E1 cell [37]. The calcification level is the highest on the RGD/PEG/Ti surface



Fig. 6.3 Immobilization of RGD peptide on Ti through electrodeposited NH₂-PEG-COOH



Fig. 6.4 Immobilization mode of RGD peptide on Ti and calcification by MC3T3-E1 cell on each specimen

(Fig. 6.4), and the bone formation on the RGD/PEG/Ti surface is accelerated compared to RGD/Ti surface in rabbit [38].

Glycine (G)–arginine (R)–glycine (G)–asparaginic acid (D)–serine (S) sequence peptide (GRGDS peptide) is coated through the chloride activation technique to enhance adhesion and migration of osteoblastic cells [39]. The expression levels of many genes in MC3T3-E1 cells are altered.

6.3.3 Proteins and Collagen

Among the relevant molecules involved in biochemical modification of bonecontacting surfaces, growth factor, such as bone morphogenetic protein-2 (BMP-2), is of primary interest. BMP-2 has been known to play an important role in bone healing processes and to enhance therapeutic efficiency. Ectopic bone formation by BMP-2 in animals has been well established following the first reports of BMP-2 by the Urist research group [40–42]. Synthetic receptor binding motif mimicking BMP-2 is covalently linked to Ti surfaces through a chemical conjunction process [43]. A complete and homogeneous peptide overlayer on the Ti surfaces; the content is further measured by gamma counting. Biological evaluations show that the biochemically modified Ti is active in terms of cell attachment behavior. Ti surfaces can enhance the rate of bone healing as compared with untreated Ti surface. Bone morphogenetic protein-4 (BMP-4) is immobilized on a Ti–6Al–4V alloy through lysozyme to improve the hard tissue response [44]. Proteins are silane-coupled to the oxidized surfaces of the Co–Cr–Mo alloy, the Ti– 6Al–4V alloy, Ti, and the Ni–Ti alloy to improve tissue compatibility [45].

Biomolecules are also used to accelerate bone formation and soft tissue adhesion on a material. Type I collagen is immobilized by immersion in the collagen solution [46]. Type I collagen production increases with modification by ethane-1,1,2-



Fig. 6.5 Atomic force microscopic images of Ti surfaces with electrodeposited collagen using DC potential or AC potential

triphosphonic acid and methylenediphosphonic acid grafted onto Ti [47]. Type I collagen is grafted with glutaraldehyde as a cross-linking agent [48]. For the electrodeposition, it is found that an alternating current (AC) between -1 and +1 V vs. SCE with 1 Hz is more effective than direct current (DC) to immobilize type I collagen to Ti and durability in water is high [49] as shown in Fig. 6.5.

Fibronectin is immobilized directly on Ti using a tresyl chloride activation technique [50]. L-threonine and O-phospho-L-threonine are immobilized on acidetched Ti surface [51].

6.3.4 Hydrogel and Gelatin

Immobilization or coating of hydrogel to metal surface is currently being attempted to add a drug delivery ability to orthopedic implant and stents or fluorescent sensing ability to microchips. Currently, synthetic polymeric hydrogels like poly-(hydroxyethylmethacrylate) (pHEMA) and poly(hydroxyethylacrylate) (pHEA) are widely used as compliant materials particularly in the case of contact with blood or other biological fluids [52]. Despite having good flexibility in the swollen state, hydrogels usually lack suitable mechanical properties, and this could greatly impair their use as coating materials for surgical procedure. Moreover, in case of inadequate adhesion between the hydrogel coating and the metal surface, a breakage at the coating-steel interface might occur [53]. A spray-coated method has been set up with the aim to control the coating of pHEMA onto the complex surface of a 316L steel stent for percutaneous coronary intervention (PCI) [54]. The pHEMA coating evaluation of roughness wettability together with its morphological and chemical stability after three cycles of expansion-crimping along with preliminary results after 6 months demonstrates the suitability of the coating for surgical implantation of stent.

An alternative and very promising synthetic route is represented by electrochemical polymerization, which leads to direct thin film coatings on the metal substrates with interesting applications either for corrosion protection or for the development of bioactive films [55–58]. As far as orthopedic field is concerned, in recent years, many procedures based on surface modification have been suggested to improve the biocompatibility and biofunctionality of Ti-based implant [59]. 2-hydroxy-ethyl-methacrylate (HEMA), a macromer poly(ethylene-glycol diacrylate) (PEGDE), and PEGDE copolymerized with acrylic acid were used to obtain hydrogels. A model protein and a model drug were entrapped in the hydrogel and released according to pH change [60].

6.4 Other Polymer Coatings

6.4.1 Bonding of Polymers with Metals Through Silane-Coupling Agent

The interfacial chemical structure governing the bonding strength, especially at the nanometer level, is one of the most challenging aspects to the development of composite materials. The combination of a Ti alloy with a resin for crown facings has been attempted [61]. In particular, silane-coupling agents containing S–H groups and Si–O–CH₃ groups are widely used to combine dental alloys with resins [62]. The S–H group works as a bonding agent with polymers, while the Si–O–CH₃ group works as a bonding agent with metals. The mechanical properties and durability of composite resin increase with the silanized filler [63–66]. However, in most studies on materials using silane-coupling agents in the field of dentistry, only the bonding strength is evaluated and discussed, and there are few reports that examine and discuss the chemical structures at the bonding interface and how they influence the bonding strength.

Studies on silane-coupling agents to combine polymers with metals have been performed in other fields. An aluminum–vegetable oil composite using a silane-coupling agent has been developed [67, 68]. Rubber-to-metal bonding by a silane-coupling agent was investigated [69]. In addition, the surface modification of stainless steel by grafting poly(ethylene glycol) using a silane-coupling agent has been reported [6]. However, only the chemical structure is investigated in these studies. In other words, the relationship between the bonding strength and the interfacial chemical structure containing a silane-coupling agent layer has not been studied.

The unequivocal relationship between the shear bonding strength and the chemical structure at the bonding interface of a Ti–SPU composite through a silanecoupling agent (γ -mercaptopropyl trimethoxysilane (γ -MPS)) is investigated [70]. The bonding interface between SPU and Ti substrate is determined by the thickness of γ -MPS layer as shown in Fig. 6.6. On the other hand, the shear bond



Fig. 6.7 Inhibition of platelet adhesion and fibrin network formation on SPU-coated Ti

strength of the Ti/SPU interface increased with ultraviolet (UV) irradiation according to the increase of the cross-linkage in SPU. Platelet adhesion to Ti is inhibited by SPU, as shown in Fig. 6.7. This technique is used for the creation of a new meta-based material having high strength, high toughness, and biofunction. UV irradiation to a Ti–SPU composite is clearly a factor governing the shear bond strength of the Ti/SPU interface [71]. In addition, active hydroxyl groups on the surface oxide film are clearly factors governing the shear bond strength [72]. After good bonding between metal and polymer is produced, biofunctionalization techniques developed in the field of polymers could be applied to the composite materials.

The effects of different kinds of terminal functional groups and the thickness of the silane layers (SIL) on the adhesive strength between Ti-29Nb-13Ta-4.6Zr (TNTZ) alloy and SPU were investigated by means of shear bonding tests. The following silane-coupling agents were employed in this study: 3-methacryloxypropyltrimethoxysilane $(\gamma$ -MPTS), aminopropyltriethoxysilane (APS), and γ -MPS. Furthermore, the shear bonding strength of the TNTZ/SIL/ SPU interface was also characterized after immersion in water for 30 days. Silanecoupling treatment produces a tenfold increase in the shear bonding strength, independent of the type of terminal functional groups and the thickness of the silane layers. The TNTZ/SIL/SPU composites are partially fractured at the interfaces of the TNTZ/SIL, while the rest of the fracture occurs at the interfaces of the SIL/SPU in single sample. The shear bonding strength decreases after immersion in water for 30 days when APS and γ -MPS are used as the silane-coupling agents, because stable chemical bonding is not achieved between the silane layer and SPU, whereas the bonding of the γ -MPTS composite is not affected by exposure to water [73]. The number of hydroxyl groups increases with an increase in treatment time at a H_2O_2 concentration of 5 %. On the other hand, an increase from 5 to 30 % in H₂O₂ concentration leads to a decrease in the number of hydroxyl groups on the TNTZ surface, because at higher H_2O_2 concentrations, the reaction that consumes the hydroxyl groups is dominant. The shear bonding strength is doubled compared with the untreated TNTZ/SIL/SPU interface. Although the shear bonding strength decreases after immersion in water for 30 days when APS and γ -MPS are used, TNTZ/y-MPTS/SPU composites exhibit good durability to water and maintain an equivalent shear bonding strength before immersion in water [74]. The adhesive strength of the SPU coating on the nanotube structure formed on TNTZ by anodization for 3,600 s increases by 50 %. These improvements in the adhesive strength of SPU are the result of an anchor effect introduced by the nanostructures formed by anodization. Fracture occurs at the interface of the nanoporous structure and the SPU coating layer. In contrast, in the case that SPU coating has been performed on the nanotube structure, fracture occurs inside the nanotubes [75].

6.4.2 Polymers Condensed in Porous Titanium

The Young's modulus of metallic materials is relatively larger than that of cortical bone: about 200 GPa in stainless steels and Co–Cr–Mo alloys, about 100 GPa in α -type and (α + β)-type Ti alloys, and 15–20 GPa in cortical bone. When fractured bone is fixed with metallic bone fixator such as bone plate and bone nail, during healing a load to fixation part is mainly received by metallic fixators because of the difference in their Young's modulus. This phenomenon is well known as so-called "stress shielding" in orthopedics. This large Young's modulus generates other problems. When a metal is used as a metallic spacer in spinal fixation, the spacer is mounted in matrix bone. In the case of dental implant, occlusal pressure is not absorbed by the implant and directly conducts to jaw bone.

To solve the above problems, metals with low Young's modulus are required. Two approaches are feasible: the decrease of the Young's modulus of a metallic material itself and decrease of the apparent Young's modulus by forming a porous body. In the latter case, the pores are sometimes filled by polymers to control the apparent Young's modulus. Ultra-high-molecular-weight polyethylene (UHMWPE) [76] and poly(methyl methacrylate) (PMMA) [77] are attempted to fill the pores in porous Ti. Figure 6.8 shows porous Ti whose pores are filled by UHMWPE.



Fig. 6.8 Fracture by bending of porous Ti whose pores are filled by UHMWPE (Provided by Dr. Naoyuki Nomura, Tohoku University)

6.5 Conclusions

Metallic materials are widely used in medicine for not only orthopedic and dental implants but also cardiovascular devices and other purposes. The metal surface may be biofunctionalized by various techniques such as the immobilization of biofunctional molecules and the creation of metal–polymer composites. These techniques make it possible to apply metals to a scaffold in tissue engineering. Artificial materials such as metal–polymer composites will continue to be used as biomaterials in the future, because of their excellent tissue compatibility and biofunctions. Some of metal–polymer composites are reviewed in other textbooks [78–80].

References

- 1. Ratner BD, Hoffman AS, Schoen FJ, Lemons JE (eds) (2004) Biomaterials science: an introduction to materials in medicine. Elsevier, Amsterdam
- 2. Mahato RI (2005) Biomaterials for delivery and targeting of proteins and nucleic acids. CRC Press, Roca Raton
- Kenausis GL, Vörös J, Elbert DL, Huang N, Hofer R, Ruiz-Taylor L, Textor M, Hubbell JA, Spencer ND (2000) Poly(L-lysine)-g-poly(ethylene glycol) layers on metal oxide surfaces: attachment mechanism and effects of polymer architecture on resistance to protein adsorption. J Phys Chem B104:3298–3309
- Huang NP, Michel R, Vörös J, Textor M, Hofer R, Rossi AD, Dlbert L, Hubbell JA, Spencer ND (2001) Poly(L-lysine)-g-poly(ethylene glycol) layers on metal oxide surfaces: surfaceanalytical characterization and resistance to serum and fibrinogen adsorption. Langmuir 17:489–498

- Huang NP, Csucs G, Emoto K, Nagasaki Y, Kataoka K, Textor M, Spencer ND (2002) Covalent attachment of novel poly(ethylene glycol)-poly(DL-lactic acid) copolymeric micelles to TiO₂ surfaces. Langmuir 18:252–258
- Zhang F, Kang ET, Neoh KG, Wang P, Tan KL (2001) Surface modification of stainless steel by grafting of poly(ethylene glycol) for reduction in protein adsorption. Biomaterials 22:1541–1548
- 7. To Y, Hasuda H, Sakuragi M, Tsuzuki S (2007) Surface modification of plastic, glass and titanium by photoimmobilization of polyethylene glycol for antibiofouling. Acta Biomater 3:1024–1032
- Tanaka Y, Doi H, Iwasaki Y, Hiromoto S, Yoneyama T, Asami K, Imai H, Hanawa T (2007) Electrodeposition of amine-terminated-poly(ethylene glycol) to Ti surface. Mater Sci Eng C27:206–212
- Tanaka Y, Doi H, Kobayashi E, Yoneyama T, Hanawa T (2007) Determination of the immobilization manner of amine-terminated poly(ethylene glycol) electrodeposited on a Ti surface with XPS and GD-OES. Mater Trans 48:287–292
- Tanaka Y, Saito H, Tsutsumi Y, Doi H, Imai H, Hanawa T (2008) Active hydroxyl groups on surface oxide film of Ti, 316l stainless steel, and cobalt-chromium-molybdenum alloy and its effect on the immobilization of poly(ethylene glycol). Mater Trans 49:805–811
- 11. Tanaka Y, Matsuo Y, Komiya T, Tsutsumi Y, Doi H, Yoneyama T, Hanawa T (2010) Characterization of the spatial immobilization manner of poly(ethylene glycol) to a titanium surface with immersion and electrodeposition and its effects on platelet adhesion. J Biomed Mater Res 92A:350–358
- Tanaka Y, Matin K, Gyo M, Okada A, Tsutsumi Y, Doi H, Nomura N, Tagami J, Hanawa T (2010) Effects of electrodeposited poly(ethylene glycol) on biofilm adherence to titanium. J Biomed Mater Res 95A:1105–1113
- 13. Balachnder N, Sukenik SN (1990) Monolayer transformation by nucleophilic substitution: applications to the creation of new monolayer assemblies. Langmuir 6:1621–1627
- 14. Bain C, Troughton Y, Tao YT, Evall J, Whitesides GM, Nuzzo RG (1989) Formation of monolayer films by the spontaneous assembly of organic thiols from solution onto gold. J Am Chem Soc 111:331–335
- Dubois LH, Nuzzo RG (1992) Synthesis, structure, and properties of model organic surfaces. Ann Rev Phys Chem 43:437–463
- 16. UlMan A (1996) Formation and structure of self-assembled monolayers. Chem Rev 96:1533-1554
- Xiao SJ, Textor M, Spencer ND (1998) Covalent attachment of cell-adhesive, (Arg-Gly-Asp)containing peptides to titanium surfaces. Langmuir 14:5507–5516
- Gawalt ES, Avaltroni MJ, Danahy MP, Silverman BM, Hanson EL, Midwood KS, Schwarzbauer JE, Schwartz J (2003) Bonding organics to Ti alloys: facilitating human osteoblast attachment and spreading on surgical implant materials. Langmuir 19:200–204
- Brovelli D, Hahner G, Ruis L, Hofer R, Kraus G, Waldner A, Schlosser J, Oroszlan P, Ehart M, Spencer ND (1999) Highly oriented, self-assembled alkanephosphate monolayers on tantalum (V) oxide surfaces. Langmuir 15:4324–4327
- Textor M, Ruiz L, Hofer R, Rossi K, Feldman K, Hahner G, Spencer ND (2000) Structural chemistry of self-assembled monolayers of octadecylphosphoric acid on tantalum oxide surfaces. Langmuir 16:3257–3271
- Fang JL, Wu NJ, Wang ZW, Li Y (1991) XPS, AES and Raman studies of an antitarnish film on tin. Corrosion 47:169–173
- 22. Van Alsten JG (1999) Self-assembled monolayers on engineering metals: structure, derivatization, and utility. Langmuir 15:7605–7614
- 23. Gawalt ES, Avaltroni MJ, Koch N, Schwartz J (2001) Self-assembly and bonding of alkanephosphonic acids on the native oxide surface of titanium. Langmuir 17:5736–5737

- Verrier S, Pallu S, Bareille R, Jonczyk A, Meyer J, Dard M, Amedee J (2002) Function of linear and cyclic RGD-containing peptides in osteoprogenitor cells adhesion process. Biomaterials 23:585–596
- Reyes CD, Petrie TA, Burns KL, Schwartz Z, Garcia AJ (2007) Biomolecular surface coating to enhance orthopaedic tissue healing and integration. Biomaterials 28:3228–3235
- 26. Hynes RO (2002) Integrins: bidirectional, allosteric signaling machines. Cell 110:673-687
- 27. Bagno A, Piovan A, Dettin M, Chiarion A, Brun P, Gambaretto R, Fontana G, Di Bello C, Palu G, Castagliuolo I (2007) Human osteoblast-like cell adhesion on titanium substrates covalently functionalized with synthetic peptides. Bone 40:693–699
- Elmengaard B, Bechtold JE, Soballe K (2005) In vivo study of the effect of RGD treatment on bone on growth on press-fit titanium alloy implants. Biomaterials 26:3521–3526
- Rammelt S, Illert T, Bierbaum S, Scharnweber D, Zwipp H, Schneiders W (2006) Coating of titanium implants with collagen. RGD peptide and chondroitin sulfate. Biomaterials 27:5561–5571
- 30. Auernheimer J, Zukowski D, Dahmen C, Kantlehner M, Enderle A, Goodman SL, Kessker H (2005) Titanium implant materials with improved biocompatibility through coating with phosphate-anchored cyclic RGD peptides. ChemBioChem 6:2034–2040
- Ferris DM, Moodie GD, Dimond PM, Gioranni CW, Ehrlich MG, Valentini RF (1999) RGD-coated titanium implants stimulate increased bone formation in vivo. Biomaterials 20:2323–2331
- 32. Xiao SJ, Textor M, Spencer ND, Wieland M, Keller B, Sigrist H (1997) Immobilization of the cell-adhesive peptide Arg-Gly-Asp-Cys (RGDC) on titanium surfaces by covalent chemical attachment. J Mater Sci Mater Med 8:867–872
- 33. Gawalt ES, Avaltroni MJ, Koch N, Schwartz J (2001) Self-assembly and bonding of alkanephosphonic acids on the native oxide surface of titanium. Langmuir 17:5736–5738
- 34. Silverman BM, Wieghaus KA, Schwartz J (2005) Comparative properties of siloxane vs phosphonate monolayers on a key titanium alloy. ACS J Surf Colloid 21:225–228
- 35. Schwartz J, Avaltroni MJ, Danahy MP, Silverman BM, Hanson EL, Schwarzbauer JE, Midwood KS, Gawalt ES (2003) Cell attachment and spreading on metal implant materials. Mater Sci Eng C23:395–400
- 36. Tanaka Y, Saito H, Tsutsumi Y, Doi H, Nomura N, Imai H, Hanawa T (2009) Effect of pH on the interaction between zwitterion and titanium oxide. J Colloid Interface Sci 330:138–143
- 37. Oya K, Tanaka Y, Saito H, Kurashima K, Nogi K, Tsutsumi H, Tsutsumi Y, Doi H, Nomura N, Hanawa T (2009) Calcification by MC3T3-E1 cells on RGD peptide immobilized on titanium through electrodeposited PEG. Biomaterials 30:1281–1286
- 38. Park JW, Kurashima K, Tustusmi Y, An CH, Suh YJ, Doi H, Nomura N, Noda K, Hanawa T (2011) Bone healing of commercial oral implants with RGD immobilization through electrodeposited poly(ethylene glycol) in rabbit cancellous bone. Acta Biomater 7:3222–3229
- 39. Yamanouchi N, Pugdee K, Chang WJ, Lee SY, Yoshinari M, Hayakawa T, Abiko Y (2008) Gene expression monitoring in osteoblasts on titanium coated with fibronectin-derived peptide. Dent Mater J 27:744–750
- 40. Urist MR (1965) Bone: formation by autoinduction. Science 150:893-899
- 41. Lee YM, Nam SH, Seol YJ, Kim TI, Lee SJ, Ku Y, Rhyu IC, Chung CP, Han SB, Choi SM (2003) Enhanced bone augmentation by controlled release of recombinant human bone morphogenetic protein-2 from bioabsorbable membranes. J Periodontol 74:865–872
- 42. Wikesjo UM, Lim WH, Thomson RC, Cook AD, Wozney JM, Hardwick WR (2003) Periodontal repair in dogs: evaluation of a bioabsorbable space-providing macroporous membrane with recombinant human bone morphogenetic protein-2. J Periodontol 74:635–647
- 43. Seol YJ, Park YJ, Lee SC, Kim KH, Lee JY, Kim TI, Lee YM, Ku Y, Rhyu IC, Han SB, Chung CP (2006) Enhanced osteogenic promotion around dental implants with synthetic binding motif mimicking bone morphogenetic protein (BMP)-2. J Biomed Mater Res 77A:599–607

- 44. Puleo DA, Kissling RA, Sheu MS (2002) A technique to immobilize bioactive proteins, including bone morphogenetic protein-4 (BMP-4), on titanium alloy. Biomaterials 23:2079–2087
- 45. Nanci A, Wuest JD, Peru L, Brunet P, Sharma V, Zalzal S, McKee MD (1998) Chemical modification of titanium surfaces for covalent attachment of biological molecules. J Biomed Mater Res 40:324–335
- 46. Nagai M, Hayakawa T, Fukatsu A, Yamamoto M, Fukumoto M, Nagahama F, Mishima H, Yoshinari M, Nemoto K, Kato T (2002) In vitro study of collagen coating of titanium implants for initial cell attachment. Dent Mater J 21:250–260
- 47. Viornery C, Guenther HL, Aronsson BO, Pöchy P, Descouts P, Grötzel M (2002) Osteoblast culture on polished titanium disks modified with phosphonic acids. J Biomed Mater Res 62:149–155
- Chang WJ, Qu KL, Lee SY, Chen JY, Abiko Y, Lin CT, Huang HM (2008) Type I collagen grafting on titanium surfaces using low-temperature grow discharge. Dent Mater J 27:340–346
- 49. Kamata H, Suzuki S, Tanaka Y, Tsutsumi Y, Doi H, Nomura N, Hanawa T, Moriyama K (2011) Effects of pH, potential, and deposition time on the durability of collagen electrodeposited to titanium. Mater Trans 52:81–89
- 50. Pugdee K, Shibata Y, Yamamichi N, Tsutsumi H, Yoshinari M, Abiko Y, Hayakawa T (2007) Gene expression of MC3T3-E1 cells on fibronectin-immobilized titanium using tresyl chloride activation technique. Dent Mater J 26:647–655
- 51. Abe Y, Hiasa K, Takeuchi M, Yoshida Y, Suzuki K, Akagawa Y (2005) New surface modification of titanium implant with phosphor-amino acid. Dent Mater J 24:536–540
- 52. Cadotte AJ, DeMarse TB (2005) Poly-HEMA as a drug delivery device for in vitro neutral network on micro-electrode arrays. J Neural Eng 2:114–122
- 53. Belkasm JS, Munro CA, Shoichet MS, Johnston M, Midha R (2005) Long-term in vivo biochemical properties and biocompatibility of poly(2-hydroxyethyl methacrylate-co-methyl methacrylate) nerve conduits. Biomaterials 26:1741–1749
- 54. Indolfi L, Causa F, Netti PA (2009) Coating process and early stage adhesion evaluation of poly(2-hydroxy-ethyl-methacrylate) hydrogel coating of 316L steel surface for stent applications. J Mater Sci Mater Med 20:1541–1551
- 55. Fenelo AM, Breslin CB (2003) The electropolymerization of pyrrole at a CuNi electrode: corrosion protection properties. Corros Sci 45:2837–2850
- 56. Mengoli G (1979) Feasibility of polymer film coatings through electroinitiated polymerization in aqueous medium. Adv Polym Sci 33:1–31
- 57. De Giglio E, Guascito MR, Sabbatini L, Zambonin G (2001) Electropolymerization of pyrrole on titanium substrates for the future development of new biocompatible surfaces. Biomaterials 22:2609–2616
- Rammelt U, Nguyen PT, Plieth W (2003) Corrosion protection by ultrathin films of conducting polymers. Electrochem Acta 48:1257–1262
- 59. De Giglio E, Gennaro I, Sabbatini L, Zambonin G (2001) Analytical characterization of collagen- and /or hydroxyapatite-modified polypyrrole films electrosynthesized on Ti-substrates for the development of new bioactive surfaces. J Biomater Sci Polym Ed 12:63–76
- 60. De Giglio E, Cometa S, Satriano C, Sabbatini L, Zambonin G (2009) Electrosynthesis of hydrogel films on metal substrates for the development of coatings with tunable drug delivery performance. J Biomed Mater Res 88A:1048–1057
- 61. Taira Y, Imai Y (1995) Primer for bonding resin to metal. Dent Mater 11:2-6
- 62. Smith NA, Antoun GG, Ellis AB, Crone WC (2004) Improved adhesion between nickeltitanium shape memory alloy and polymer matrix via silane coupling agents. Compos Part A Apply Sci 35:1307–1312
- 63. Abboud M, Casaubieilh L, Morval F, Fontanille M, Duguet E (2000) PMMA-based composite materials with reactive ceramic fillers: IV. Radiopacifying particles embedded in PMMA beads for acrylic bone cements. J Biomed Mater Res 53:728–736

- 64. Yoshida K, Tanagawa M, Atsuta M (2001) Effects of filler composition and surface treatment on the characteristics of opaque resin composites. J Biomed Mater Res 58:525–530
- 65. Kanie T, Arikawa H, Fujii K, Inoue K (2004) Physical and mechanical properties of PMMA resins containing γ-methacryloxypropyltrimethoxysilane. J Oral Rehabil 31:161–171
- 66. Ferracane JL, Berge HX, Condon JR (1998) In vitro aging of dental composites in water-effect of degree of conversion, filler volume, and filler matrix coupling. J Biomed Mater Res 42:465–472
- 67. Bexell U, Olsson M, Jhansson M, Samuelsson J, Sundell PE (2003) A tribological study of a novel pre-treatment with linseed oil bonded to mercaptosilane-treated aluminum. Surf Coat Technol 166:141–152
- Bexell U, Olsson M, Sundell PE, Jhansson M, Carlsson P, Hellsing MA (2004) ToF-SIMS study of linseed oil bonded to mercaptosilane-treated aluminum. Appl Surf Sci 231–232:362–365
- 69. Jayaseelan SK, Ooji WJV (2001) Rubber-to-metal bonding by silanes. J Adhes Sci Technol 15:967–991
- 70. Sakamoto H, Doi H, Kobayashi E, Yoneyama T, Suzuki Y, Hanawa T (2007) Structure and strength at the bonding interface between a titanium-segmented polyurethane composite through 3-(trimethoxysilyl) propyl methacrylate for artificial organs. J Biomed Mater Res 82A:52–61
- 71. Sakamoto H, Hirohashi Y, Saito H, Doi H, Tsutsumi Y, Suzuki Y, Noda K, Hanawa T (2008) Effect of active hydroxyl groups on the interfacial bond strength of titanium with segmented polyurethane through -mercaptopropyl trimethoxysilane. Dent Mater J 27:81–92
- 72. Sakamoto H, Hirohashi Y, Doi H, Tsutsumi Y, Suzuki Y, Noda K, Hanawa T (2008) Effect of UV irradiation on the shear bond strength of titanium with segmented polyurethane through -mercapto propyl trimethoxysilane. Dent Mater J 27:124–132
- 73. Hieda J, Niinomi M, Nakai M, Kamura H, Tsutsumi H, Hanawa T (2012) Effect of terminal functional groups of silane layers on adhesive strength between biomedical Ti-29Nb-13Ta-4.6Zr alloy and segment polyurethanes. Surf Coat Technol 206:3137–3141
- 74. Hieda J, Niinomi M, Nakai M, Kamura H, Tsutsumi H, Hanawa T (2013) Improvement of adhesive strength of segmented polyurethane on Ti–29Nb–13Ta–4.6Zr alloy through H₂O₂ treatment for biomedical applications. J Biomed Mater Res Part B Appl Biomater 101B:776–783
- 75. Hieda J, Niinomi M, Nakai M, Cho K, Mohri T, Hanawa T (2014) Adhesive strength of medical polymer on anodic oxide nanostructures fabricated on biomedical β-type titanium alloy. Mater Sci Eng C36:244–252
- 76. Nomura N, Baba Y, Kawamura A, Fujinuma S, Chiba A, Masahashi N, Hanada S (2007) Mechanical properties of porous titanium compacts reinforced by UHMWPE. Mater Sci Forum 539–543:1033–1037
- 77. Nakai M, Niinomi M, Akahori T, Yamanoi H, Itsuno S, Haraguchi N, Itoh Y, Ogasawara T, Onishi T, Shindo T (2008) Effect of silane coupling treatment on mechanical properties of porous pure titanium filled with PMMA for biomedical applications. J Jpn Inst Metal 72:839–845
- Possart W (1998) Adhesion of polymers. In: Hansen JA, Breme HJ (eds) Metals as biomaterials. Wiley, New York, pp 197–218
- Worch H (1998) Special thin organic film. In: Hansen JA, Breme HJ (eds) Metals as biomaterials. Wiley, New York, pp 177–196
- Xiao SJ, Kenausis G, Textor M (2001) Biochemical modification of titanium surfaces. In: Brunrtte DM, Tenvall P, Textor M, Thomsen P (eds) Titanium in medicine. Springer, Amsterdam, pp 417–455