The Application of Support Vector Machine and Behavior Knowledge Space in the Disulfide Connectivity Prediction Problem

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Abstract. In this paper, we apply support vector machine (SVM) and behavior knowledge space (BKS) to the disulfide connectivity prediction problem. The problem aims to establish the disulfide connectivity pattern of the target protein. It is an important problem since a disulfide bond, formed by two oxidized cysteines, plays an important role in the protein folding and structure stability. The disulfide connectivity prediction problem is difficult because the number of possible patterns grows rapidly with respect to the number of cysteines. We discover some rules to discriminate the patterns with high accuracy in various methods. Then, the pattern-wise and pair-wise BKS methods to fuse multiple classifiers constructed by the SVM methods are proposed. Finally, the CSP (cysteine separation profile) method is also applied to form our hybrid method. We perform some simulation experiments with the 4-fold crossvalidation on SP39 dataset. The prediction accuracy of our method is increased to 69.1 %, which is better than the best previous result 65.9 %.

Keywords: Disulfide bond · Cysteine · Connectivity pattern · Support vector machine · Behavior knowledge space

1 Introduction

A *disulfide bond*, also called *SS-bond* or *SS-bridge*, is a single covalent bond which is usually formed from the oxidation of two thiol groups (-SH). The transformation is described as

$$
2RSH \to RS-SR + 2H^+ + 2e^- \tag{1}
$$

where R represents the carbon-containing group of atoms.

In proteins, only the thiol groups of cysteine residues can form the disulfide bonds by oxidation. The goal of the *disulfide connectivity prediction* (DCP) problem is to figure out which cysteine pair would be cross-link from all possible

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candidates. It may be conducive to the solution of protein structure prediction problem if precise disulfide connectivity information is available.

There are two main ways for connectivity pattern prediction in previous works, pair-wise and pattern-wise. The pair-wise method focuses on the bonding potential of each cysteine pair, and encodes the target based on cysteine pairs. The pattern-wise method makes a comprehensive survey of whole connectivity pattern and usually ranks the connectivity patterns by their possibilities, so the prediction ability may be limited to the diversity of patterns in a training set.

The prediction task is difficult because the number of possible connectivity patterns grows rapidly with respect to the number of cysteines. Most previous studies are limited by the number of disulfide bonds from two to five. It is well known that the number of possible patterns is given as follows:

$$
N = \frac{C_2^{2B} \times C_2^{2B-2} \times \ldots \times C_2^2}{B!} = (2B - 1)!!
$$
 (2)

where B denotes the number of disulfide bonds in the protein. For instance, if we have known which cysteines are oxidized in advance, $N = 945$ when $B = 5$, and N is up to 10395 when $B = 6$.

Some statistical analyses $[1-4]$ $[1-4]$ have been applied to the disulfide connectivity prediction problem. Many researchers tried to solve the problem with machine learning methods, such as neural network (NN) [\[5](#page-12-1)[–10](#page-12-2)] and support vector machine (SVM) [\[2](#page-12-3)[,11](#page-12-4)[–17](#page-12-5)].

Before 2005, many studies $[5,8]$ $[5,8]$ $[5,8]$ were devoted to the connectivity prediction, but most of their accuracies are below 50 %. In 2005, Zhao *et al.* [\[18\]](#page-12-7) utilized the global information of a protein, called *cysteine separation profile* (CSP), which represents the separations among all oxidized cysteines in a protein sequence. In 2007, Lu et al. $[2]$ proposed a novel concept of the CP_2 representation, which uses every two cysteine pairs (four cysteines) as one sample, and applied the genetic algorithm (GA) to the optimization of feature selection.

In 2012, Wang *et al.* [\[19](#page-12-8)] built a hybrid model based on SVM and the weighted graph matching, with accuracy 65.9 %. They extracted different feature sets depending on whether the number of disulfide bonds in a protein is odd or even. The main difference of the feature sets for the two submodels is the secondary structure information around the oxidized cysteines.

The rest of this paper is organized as follows. We introduce some preliminary knowledge, including support vector machine and behavior knowledge space in Sect. [2.](#page-1-0) In Sect. [3,](#page-3-0) we present our hybrid method for solving the DCP problem. The experimental results are given in Sect. [4,](#page-9-0) and we also describe the performance comparison between our method and some previous works. Finally, our conclusion is given in Sect. [5.](#page-11-1)

2 Preliminary

In this section, we introduce some background knowledge used in this paper, including support vector machine and behavior knowledge space.

2.1 Support Vector Machine

Support Vector Machine (SVM) is a machine learning method for classification and regression. It was first introduced by Vapnik [\[20\]](#page-12-9) in 1999. SVM seeks to create a hyperplane to discriminate different labels of the vectors in the training set and utilizes the model to predict the labels of other data. To discover the discriminative features is the key point for applying SVM. Figure [1](#page-2-0) shows an example of the SVM solution with maximum *margin* which means the distance between the hyperplane and the given objects.

Fig. 1. An example of the SVM solution with maximum margin.

For SVM implementation, we use the LIBSVM package [\[21\]](#page-12-10) which is an easyto-use tool for *support vector classification* (SVC) and *support vector regression* (SVR). The SVC function classifies the data with their probabilities, and the SVR function generates the regression value of each target data element.

2.2 Behavior Knowledge Space

Behavior knowledge space (BKS) [\[22\]](#page-13-0) is a kind of method for fusing multiple classifiers. It is a table look-up approach for estimating the probability of every vote combination. Assume there are m classifiers composing an ensemble for a classification task of n labels. The BKS table contains n^m entries, the number of all possible combinations of m classifiers' outputs. And each entry records the distribution of n true labels in the training set.

Table [1](#page-3-1) illustrates an example of the BKS table for the 3-label classification problem with two classifiers. The 'C1' and 'C2' represent the predicted outputs from the two classifiers, and the entries below them are all possible prediction combinations. Cells below 'Real label', 'L1', 'L2', and 'L3', are the distribution of the true labels associated with the predicted label vectors. For example, when 'C1'='L1' and 'C2'='L3', the fused answer should be 'L2' since it is the most possible label. And, if we have 'C1'='L3', 'C2'='L2', the fused answer should go to 'L3'.

Predicted label Real label				
C1	\mid C2	L1	L2	L ₃
L1	T.1	23	8	$\overline{2}$
L1	L ₂	5	Ω	4
L1	L3	$\overline{2}$	7	1
L3	L ₂	1	1	5
L3	L ₃	1	3	12

Table 1. An example of the BKS table.

3 Algorithms for Connectivity Prediction

We observe that the prediction accuracies of Chung *et al.* [\[23\]](#page-13-1) and Wang *et al.* [\[19](#page-12-8)] are 63.5 % and 65.9 %, respectively. It may be hard to find more features with good discrimination capability for a single SVM method in the connectivity prediction. However, we may get better accuracies if we fuse the advantages of the multiple models.

Our method utilizes BKS to fuse the results obtained from SVM models. The features and cysteine-pair representation we adopted are inspired by Wang *et al.* [\[19](#page-12-8)] and Lu *et al.* [\[2](#page-12-3)]. In addition, we also combine the CSP method [\[18\]](#page-12-7) to our hybrid method.

3.1 Feature Extraction

In the past, the bonding states of each cysteine pair are usually used to describe the disulfide pattern and used as the samples of SVM. Lu *et al.* [\[2\]](#page-12-3) call it as the CP_1 representation. Lu *et al.* further proposed a novel concept of the CP_2 representation which use every two cysteine pairs (four cysteines) as the samples. In our method, we adopt the features used by Wang *et al.* [\[19](#page-12-8)]. In addition, we encode the CP₂ representation as the *permutation order*, which is also included in our feature set. The definition of the permutation order is given as follows.

Permutation Order: This feature implies the order of feature extraction in each cysteine window. For every cysteine-pair combination in the $\rm CP_2$ representation, we encode the samples in three permutations illustrated in Table [2.](#page-4-0) For example, C_1 - C_3 - C_2 - C_4 means that the first and third cysteines form a disulfide bond in these four cysteines, and the second and fourth form the other bond. This bond pattern is represented by the feature vector $(0.25, 0.75, 0.5, 1)$.

Permutations Feature vector
C_1 -C ₂ -C ₃ -C ₄ (0.25, 0.5, 0.75, 1)
C_1 -C ₃ -C ₂ -C ₄ (0.25, 0.75, 0.5, 1)
C_1 -C ₄ -C ₂ -C ₃ (0.25, 1, 0.5, 0.75)

Table 2. The feature vector of the permutation order.

3.2 SVM Method

We implement three SVM models with different feature sets, $\rm CP_1F_{521}, \rm CP_1F_{623}$ and $\text{CP}_2\text{Label}_2$, as shown in Table [3.](#page-4-1) These features are encoded by the segments of every cysteine pair. The cysteine segment is a window centering at a target cysteine. Many previous works $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ $[2,4,8,9,12,14,16,17,24-26]$ also adopted the similar idea of the window approach. Here we set the window size to 13. In other words, $2k + 1 = 13$. So there are 521 features in $\text{CP}_1\text{F}_{521}$ and 623 features in $\rm CP_1F_{623}.$

Table 3. The feature sets used in our three models.

Feature	size	M^a	M ^b	M ^c
Distance of cysteines		Y	Y	Y
Cysteine order	$\overline{2}$		Y	
Protein weight	1		Y	
Protein length	1		Y	
Amino acid composition	20		Y	
PSSM around cysteine	$(2k+1)\times 20\times 2$ Y		Y	
Secondary structure around cysteine	$(2k+1)\times 3\times 2$		Y	
Permutation order	4			

^a $\text{CP}_1\text{F}_{521}$ model.

 b CP₁F₆₂₃ model.

 $\rm ^c$ CP₂Label₂ model.

Table 4. The details of the probability intervals for the BKS method, where *B* denotes the number of bonds in a target protein.

		B Type of BKS Probability intervals
$\overline{2}$		Pattern-wise $(0, 0.15, 0.2, 0.25, 0.3, 0.35, 0.5, 1)$
3	Pattern-wise	(0, 0.25, 0.5, 1)
	Pair-wise	(0, 0.1, 0.2, 0.3, 0.4, 0.5, 1)
	5 Pair-wise	(0, 0.1, 0.2, 0.3, 0.4, 0.5, 1)

3.3 BKS Method

We adopt the concept of the behavior knowledge space (BKS) to fuse the above SVM classifiers. We design two BKS models, pattern-wise BKS and pairwise BKS, combined with the probability intervals, where the probabilities are obtained from the prediction of SVM classifiers. The details of the probability intervals for the proteins with various number of disulfide bonds are shown in Table [4.](#page-4-2)

Pattern-Wise BKS Method. After the two classifiers $\text{CP}_1\text{F}_{521}$ and $\text{CP}_1\text{F}_{623}$ finish the pattern prediction, the probability of each bonding pattern is obtained. Then, the pattern-wise BKS is constructed according to the prediction probabilities. We adopt the pattern-wise BKS method for the prediction of proteins with two or three bonds. Table [5](#page-5-0) illustrates an example of the partial pattern-wise BKS table for 2-bond proteins. For example, in the second row, the probabilities of the predicted pattern 1-1-2-2 for the two classifiers locate in $(0.15, 0.2)$. In this case, 5, 3 and 1 proteins have the true patterns 1-1-2-2, 1-2-1-2 and 1-2-2-1, respectively. Thus, the fused answer is decided to be 1-1-2-2.

We set the threshold of the patterns supported in the pattern-wise BKS table to 2, and reject to give an answer in the case below the threshold. Table [6](#page-5-1) shows some examples for 3-bond proteins whose prediction can be corrected by the pattern-wise BKS method.

CP_1F_{521}	Interval	CP_1F_{623}	Interval		$1-1-2-2$ $1-2-1-2$ $1-2-2-1$	
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		(0, 0.15)	θ	1	0
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		$(0.15, 0.2)$ 5		3	
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		$(0.2, 0.25)$ 4		Ω	0
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		$(0.25, 0.3)$ 0		Ω	0
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		$(0.3, 0.35)$ 0		Ω	θ
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		$(0.35, 0.5)$ 1		Ω	0
$1 - 1 - 2 - 2$	$(0.15, 0.2)$ 1-1-2-2		(0.5, 1)	$\overline{0}$	Ω	0

Table 5. An example of the partial pattern-wise BKS table for 2-bond proteins.

Table 6. Examples for 3-bond proteins corrected by the pattern-wise BKS method.

Proteins	Real patterns CP_1F_{521}	$\rm{CP_1F_{623}}$	Predicted by BKS
$CXOA$ _{-CONMA} $ 1-2-3-1-2-3 $		$1-2-1-3-2-3$ $1-2-1-3-2-3$ $1-2-3-1-2-3$	
HST1 ECOLI	$1-2-3-1-2-3$	$1-2-1-3-2-3$ 1-2-1-3-2-3 1-2-3-1-2-3	
HCYA_PANIN	$1 - 1 - 2 - 2 - 3 - 3$	$1-1-2-2-3-3$ $1-1-2-3-3-2$ $1-1-2-2-3-3$	
CXOB_CONST	$1-2-3-1-2-3$	$1-2-1-3-2-3$ $1-2-1-3-2-3$ $1-2-3-1-2-3$	

Pair-Wise BKS Method. The pattern-wise BKS method is not suitable for the prediction of all proteins. The number of all possible combinations of patterns grows rapidly with respect to the number of bonds, so the number of the training samples is relatively not enough. We propose the pair-wise BKS method for the prediction of proteins with four or five bonds. The pair-wise BKS table records the ratio of the pairs truly bonded or not in various probability intervals from the two classifiers, $\text{CP}_1\text{F}_{521}$ and $\text{CP}_2\text{Label}_2$. Table [7](#page-6-0) shows an example of the partial pair-wise BKS table for 5-bond proteins. For every cysteine pair, we advisably adjust the original probability from $\rm CP_1F_{521}$ method according to the ratio of the truly bonded pairs in the pair-wise BKS table. Table [8](#page-6-1) illustrates the adjustment rules. Eventually, the predicted pattern is derived from the top N maximum weighted graph matching by the adjusted weighted matrix until the matching pattern belongs to a real pattern in PDB dataset according to our statistics, where the probabilities obtained by SVM classifiers are input as the edge weights in graph matching.

Pairs from $\rm CP_1F_{521}$	Pairs from $\rm CP_2Label_2$ Truly bonded Not bonded		
(0.3, 0.4)	(0, 0.1)		
(0.3, 0.4)	(0.1, 0.2)	θ	
(0.3, 0.4)	(0.2, 0.3)	6	5
(0.3, 0.4)	(0.3, 0.4)	4	6
(0.3, 0.4)	(0.4, 0.5)	6	6
(0.3, 0.4)	(0.5, 1)		12

Table 7. An example of the partial pair-wise BKS table for 5-bond proteins.

Table 8. The adjustment rules for the pair-wise BKS method.

Truly bonded Not bonded		Adjustment ratio
$\overline{0}$	0	1
[1, 10)	0	4
≥ 10	0	8
$\overline{0}$	[1, 10)	0.25
$\overline{0}$	>10	0.125
$\mathbf x$	$\mathbf x$	1
(x, 2x)	X	$\overline{2}$
$\geq 2x$	X	4
$\mathbf x$	$\frac{[x, 2x)}{\geq 2x}$	0.5
X		0.25

3.4 Hybrid Method

Instead of a large amount of features used by the SVM method, Zhao *et al.* [\[18\]](#page-12-7) adopted only one feature, CSP (cysteine separations profile), to achieve nearly 50 % accuracy in the dataset with insufficient information. The CSP of protein x with $2n$ oxidized cysteines (*n* disulfide bonds) is defined as

$$
CSP_x = (\delta 1, \delta_2, \dots, \delta_{2n-1}) = (\rho_2 - \rho_1, \rho_3 - \rho_2, \dots, \rho_{2n} - \rho_{2n-1})
$$
(3)

where ρ_i denotes the sequence position of the *i*th oxidized cysteine in the protein and δ_i denotes the separation distance between oxidized cysteines i and $i + 1$.

The divergence (D) of two CSPs for two proteins x and y is defined [\[18](#page-12-7)] as follows:

$$
D = \sum_{i=1}^{i=2n-1} |\delta_{x,i} - \delta_{y,i}|.
$$
 (4)

It has been shown that the CSP is an important global feature for the disulfide connectivity prediction, so we also combine the CSP method into our hybrid method. Figure [2](#page-8-0) exhibits the flow chart of our work. Our hybrid method for predicting the disulfide connectivity pattern is described as follows.

Algorithm. The hybrid method.

Input: A protein sequence and the bonding states of its all cysteines.

Output: The predicted disulfide connectivity pattern.

Case 1: For a 2-bond or 3-bond protein.

- Step 1.1: Apply the maximum weighted graph matching algorithm to derive the pattern from the $\rm CP_1F_{521}$ method for 2-bond proteins (the $\rm CP_1F_{623}$ method for 3-bond proteins). If the normalized weight of one pattern is greater than or equal to 0.5, report this pattern as the predicted pattern.
- Step 1.2: If the condition meets a predefined threshold in the pattern-wise BKS method, report this pattern as the predicted pattern.
- Step 1.3: If the minimum divergence obtained by the CSP search is less than or equal to a predefined threshold, report this pattern as the predicted pattern.
- Step 1.4: For the remaining, take the original maximum weighted pattern from the $\text{CP}_1\text{F}_{521}$ method for 2-bond proteins (the $\text{CP}_1\text{F}_{623}$ method for 3-bond proteins) as the predicted result.

Case 2: For a 4-bond or 5-bond protein.

- Step 2.1: Apply the maximum weighted graph matching algorithm to derive the pattern from the $\rm CP_1F_{521}$ method. If the normalized weight of one pattern is greater than or equal to 0.5, report this pattern as the predicted pattern.
- Step 2.2: If the minimum divergence obtained by the CSP search is less than or equal to a predefined threshold, report this pattern as the predicted pattern.
- Step 2.3: Adjust the original weight (probability) of each pair from the $\rm CP_1F_{521}$ method according to the pair-wise BKS table and report the pattern derived from weighted graph matching algorithm as the predicted result.

Fig. 2. The system flow chart of our method.

4 Experimental Results

In this section, we introduce our testing dataset and performance evaluation criteria of the disulfide connectivity prediction. In addition, we show the experimental results of various methods.

4.1 Dataset

For the fair comparison of the prediction accuracy with previous works, we use SP39 dataset, which is the same dataset adopted in some previous works, for our training and testing. Table [9](#page-9-1) illustrates the summary of SP39 dataset. This dataset contains 446 proteins with two to five disulfide bonds, derived from the SWISS-PROT release no. 39. It was first used by Vullo and Frasconi [\[10](#page-12-2)]. We also use the same way as Wang *et al.*'s [\[19\]](#page-12-8) to divide SP39 dataset into four subsets for the 4-fold cross-validation. The sequence identity of proteins between any two subsets is less than 30 %.

Defined by Vullo and Frasconi [\[10\]](#page-12-2).

4.2 Performance Evaluation

The definition of k -fold cross-validation is given as follows. A dataset D is divided into k subsets D_1, D_2, \ldots, D_k , which are disjoint to each other. Each time, we take a subset D_i , $1 \leq i \leq k$, as the testing set and use the other $k-1$ subsets for training. Repeat this procedure k times until each subset is tested once. Here, we adopt the 4-fold cross-validation. For the measurement of the performance in connectivity pattern prediction, the accuracy is calculated as follows:

$$
Q_p = \frac{C_p}{T_p},\tag{5}
$$

where C_p denotes the number of proteins whose connectivity patterns are correctly predicted, and T_p is the total number of proteins for testing.

4.3 Results

In the $\text{CP}_1\text{F}_{521}$ method, combined by the SVM method with the maximum weighted graph matching [\[27](#page-13-4)], we discover that the prediction accuracy is very high when the normalized weight of one predicted pattern is greater than or equal to 0.5 (half). Table [10](#page-10-0) shows the ratio and accuracies of these patterns. In other words, the confidence of such prediction is very high. Thus, in Step 1.1 or Step 2.1 of our method, the answer is settled down for these predictions.

					$ B=2 B=3 B=4 B=5 B=2\cdots 5$
Accuracy (Q_n)	-100	-93.0	93.9	192.9	-96.0
Ratio in the dataset $ 37.8\rangle$		129.5	33.3	± 31.1	33.4

Table 11. The accuracy and ratio of the pattern-wise BKS method in SP39 dataset when the threshold is set to 2.

	$B = 2 B = 3$	
Accuracy (Q_p)	94.0	95.7
Ratio in the dataset 53.2		31.5

Table 12. The *Q^p* and ratio of the divergence of the CSP in SP39 dataset.

We use the BKS as a supporting role in our method. For the two kinds of the BKS, the performance of the pattern-wise BKS is more effective than the pair-wise BKS. Table [11](#page-10-1) illustrates the details of the pattern-wise BKS method when we set the threshold to 2 for 2-bond and 3-bond proteins.

Our hybrid method is combined by the SVM method and the BKS method. In addition, we also take the CSP method into consideration. Table [12](#page-10-2) shows the Q_p and ratio of the divergence of the CSP. Take 3-bond proteins as an example. There are 34% proteins with CSP search less than or equal to 5, and the Q_p of these proteins reaches up to 96 %. According to the observation, we set the applicable thresholds of CSP to pick out the patterns as results. Here, we set the threshold of CSP to 0, 5, 10, and 15 for proteins with two to five bonds, respectively.

Table 13 shows the Q_p of our methods and some previous works in SP39 dataset. The accuracies of the three SVM models are derived from the patterns with the maximum weighted graph matching. However, we find that it is hard to improve the accuracy by one single SVM model. Although the performance of $\text{CP}_2\text{Label}_2$ is not better than $\text{CP}_1\text{F}_{521}$ or $\text{CP}_1\text{F}_{623}$, $\text{CP}_2\text{Label}_2$ provides the effect for pair-wise BKS since $\text{CP}_2\text{Label}_2$ represents another concept of pair extraction. Eventually, the prediction accuracy of our hybrid method with SVM and BKS reaches 65.9 %, and up to 69.1 % combined with CSP method.

Method	$B=2$				$B = 3 B = 4 B = 5 B = 2 \cdots 5$
CSP ^a	72.4	54.1	33.3	17.8	52.2
Wang's method ^b	84.0	60.3	55.6	44.4	65.9
CP_1F_{521}	84.0	53.4	55.6	46.7	63.9
CP_1F_{623}	78.2	60.3	53.5	44.4	63.5
CP_2 Label ₂	75.0	49.3	52.5	40.0	58.1
$CP_1F_{521} + BKS$	84.0	56.8	55.6	55.6	65.9
$CP_1F_{521} + BKS + CSP$	84.0	64.4	57.6	57.8	69.1

Table 13. The *Q^p* of our methods and previous works in SP39 dataset.

^aProposed by Zhao *et al.* [\[18](#page-12-7)].

^bProposed by Wang *et al.* [\[19\]](#page-12-8).

5 Conclusion

According to the study of Wang *et al.* [\[19\]](#page-12-8), which focuses SVM models on varied features, and the concept of different cysteine-pair representations proposed by Lu *et al.* [\[2\]](#page-12-3), we do many integrated experiments. However, the improvement of the pure SVM methods is not so significant although the SVM method is still relatively better, compared with other machine learning methods. Some studies [\[28](#page-13-5)[,29](#page-13-6)] combine the SVM method with CSP or sequence alignment to raise the accuracy. The key step of the CSP method and the sequence alignment method is to search for a good template set. However, the accuracy of these two methods depends on the pattern varieties in the template set.

We think that the design of hybrid methods is the trend in the disulfide connectivity prediction problem. In this paper, we gather some statistics about the disulfide bonds, and have successfully found some rules to discriminate the patterns with high accuracy in several methods. Furthermore, we adopt the patternwise and pair-wise BKS methods to fuse multiple SVM models, and use the predicted patterns from the original SVM method for the rest of proteins.

In the future, we may examine our hybrid method to other datasets, and explore more methods for fusing multiple classifiers, such as the weighted majority vote. We may try the CSP method with the inter-bond template dataset to explore more possibilities of the development with the concept of subpatterns.

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