

Teresa Cerchiara, Federica Bigucci,
and Barbara Luppi

Contents

20.1	Introduction	285
20.1.1	Physical and Chemical Properties of Hydrogels	286
20.2	Applications of Hydrogels in Transdermal Drug Delivery	289
20.2.1	Semisolid Vehicles	291
20.2.2	Film-Based Vehicles	292
	Conclusions	295
	References	295

20.1 Introduction

Hydrogels date back to 1960 when Wichterle and Lim first proposed the use of hydrophilic networks of poly(2-hydroxyethylmethacrylate) (HEMA) in contact lenses (Wichterle and Lim 1960). Since then, the use of hydrogels has extended to various biomedical (Hoffman 2002; Peppas et al. 2006; Kopeceka 2007) and pharmaceutical (Peppas et al. 2000) applications. In particular, due to their physical properties, similar to those of human tissues (water content, soft and pliable consistence), hydrogels have been used for different administration routes such as oral, rectal, ocular, epidermal, and subcutaneous (Peppas et al. 2000; Guy 1996; Jatav et al. 2011).

Hydrogels are composed of hydrophilic macromolecules forming three-dimensional insoluble networks able to imbibe large amounts of water or biological fluids (Peppas and Mikos 1986). Commonly, the polymers utilized to make hydrogels are insoluble due to the presence of permanent or reversible cross-links (Berger et al. 2004). Permanent cross-linked hydrogels (Wichterle and Lim 1960; Xiao and Zhou 2003; Brasch and Burchard 1996) are characterized by covalent bonds forming tie points or junctions, whereas reversible cross-linked hydrogels (Watanabe et al. 1996; Wang et al. 1999; Qu et al. 1999) present ionic, hydrophobic, or coiled-coil physical interactions. These kinds of cross-links

T. Cerchiara (✉) • F. Bigucci • B. Luppi
Department of Pharmacy and Biotechnology-FaBiT,
Bologna University, Via San Donato 19/2,
Bologna 40127, Italy
e-mail: teresa.cerchiara2@unibo.it;
federica.bigucci@unibo.it; barbara.luppi@unibo.it

in the polymer structure yield insoluble materials able to swell in aqueous environments retaining a significant fraction of water in their structure, up to thousands of times their dry weight in water.

Hydrogels can be divided into homopolymer or copolymers based on the preparative method, but they can also be natural polymers, synthetic polymers, or derivatives. In nature hydrogels can be found in plants (pectin, pullulan), various species of brown seaweed (alginic acid, agar, carrageenan), crustaceans (chitin), and animal tissue (hyaluronic acid, collagen, fibrin). Typical simple synthetic materials applied for general-purpose hydrogels are poly(ethylene oxide), poly(vinyl alcohol), poly(vinyl pyrrolidone), poly(hydroxyethyl methacrylate), and poly(N-isopropyl acrylamide). Moreover, the synthetic pathway offers more possibilities to create hydrogels with modified functional properties. In fact, several physiologically responsive hydrogels are obtained from chemical or physical modifications of natural and synthetic polymers and tested for use in the so-called intelligent biomaterials (Hoffman 1991; Miyata et al. 2002; Murdan 2003; Chen et al. 2004) because they are capable of reacting to various environmental stimuli (temperature, pH, ionic strength, solute concentration, electric radiation, light, sound, etc.).

Hydrogels can be homogeneous, when the pores between polymer chains are the only spaces available for mass transfer and the pore size is within the range of molecular dimensions (a few nanometers or less) or porous when the effective pore size is over 10 nm. In homogeneous hydrogels the transfer of water or other solutes is achieved by a pure diffusional mechanism, which restricts the rate of absorption and to some extent the size of species that are absorbed.

Porous hydrogels can be made by different polymerization methods in the presence of dispersed porosigens (ice crystals, oil, sucrose crystals) which can be removed later to leave an interconnected meshwork, where the pore size depends on the size of the porosigens (Hickey and Peppas 1995). The introduction of a porosigen reduces mechanical strength significantly making porous hydrogels weaker than homogeneous hydrogels.

In medical, engineering, and pharmaceutical technology, hydrogel degradation is considerable

important. In fact, investigators have focused on controlling degradation behavior of hydrogels to design polymers able to be cleared from the body once they complete their roles (Anderson and Shive 1997; Timmer et al. 2002): for this reason labile bonds are frequently introduced in the gels. These bonds can be present either in the polymer backbone or in the cross-links used to prepare the gel. The labile bonds can be broken under physiological conditions either enzymatically or chemically, in most cases by hydrolysis (Damink et al. 1996; Eliaz and Kost 2000; Lee et al. 2004).

20.1.1 Physical and Chemical Properties of Hydrogels

An important property of hydrogels is their swelling behavior: it depends upon the polymer, extent of cross-linking, temperature, polymer-solvent interactions, and extent of ionization (Khare et al. 1992). In particular the extent of cross-linking can be changed to achieve a relatively strong and yet elastic hydrogel. Long-chain cross-linkers and low cross-linking ratios (the ratio of moles of cross-linking agent to the moles of polymer repeating units) produce extremely weak hydrogels, while short-chain cross-linkers and high cross-linking ratios produce extremely tight hydrogels. Tightly cross-linked hydrogels will swell less than the same hydrogels with low cross-linking ratios or long cross-linkers chains.

The presence of hydrophilic or hydrophobic groups in the chemical structure of the polymer affects the swelling behavior of hydrogels. When a dry hydrogel begins to absorb water, the first water molecules entering the matrix will hydrate polar hydrophilic groups. As the polar groups are hydrated, the network swells exposing hydrophobic groups which also interact with water molecules, leading to hydrophobically bound water. Finally an equilibrium swelling level is reached when the network imbibes additional water ("free water") which fills the space between the polymer chains. As hydrophobic groups minimize their exposure to the water molecule, hydrogels containing hydrophobic groups will swell much less than hydrogels containing hydrophilic groups.

As stated earlier, the dissolution of polymer chains and consequently hydrogel swelling ability is prevented by the presence of cross-linking in the three-dimensional network. Different chemical and physical cross-linking methods have been employed to prepare hydrogels (Hennink and van Nostrum 2002). In chemically and physically cross-linked gels, dissolution is prevented by covalent bonds and physical interactions between different polymer chains, respectively. Chemically cross-linked gels can be obtained by radical polymerization of low molecular weight monomers in the presence of cross-linking agents, chemical reaction of complementary groups, and high-energy irradiation. Physically cross-linked gels can be obtained by ionic interactions, hydrogen bonds, crystallization, and aggregation of the hydrophobic segments of multiblock copolymers or graft copolymers. An example of cross-linking by radical polymerization is the synthesis of hydrogels of Wichterle and Lim (1960), a copolymerization of HEMA with the cross-linker EGDMA

(ethylene glycol dimethacrylate) in the presence of AIBN (2,2'-azo-bis-isobutyronitrile), the radical initiator (Fig. 20.1). Chemical cross-linking agents, such as acyl dichlorides (Fig. 20.2), can establish covalent linkages with functional groups of polymers, such as activated hydroxylic groups of poly(vinyl alcohol) (Orienti et al. 2000). Polyaldehydes are utilized to cross-link proteins such as albumin (Sahin et al. 2002) and gelatin (Draye et al. 1998) or natural polysaccharides such as hyaluronic acid (Luo et al. 2000). However, a significant disadvantage of chemical cross-linking agents is their toxicity. Among various methods applied for the production of hydrogels, the radiation technique (Safrany 1997) is a simple, efficient, clean, and environment-friendly process (Fig. 20.3). Hydrogels can be obtained by radiation technique in a few ways, including irradiation of solid polymer (Nedkov and Tsvetkova 1994), monomer (in bulk or in solution) (Rosiak 1991), or aqueous solution of polymer (Kabanov 1998). For irradiation technologies, the main irradiating sources include gamma rays from

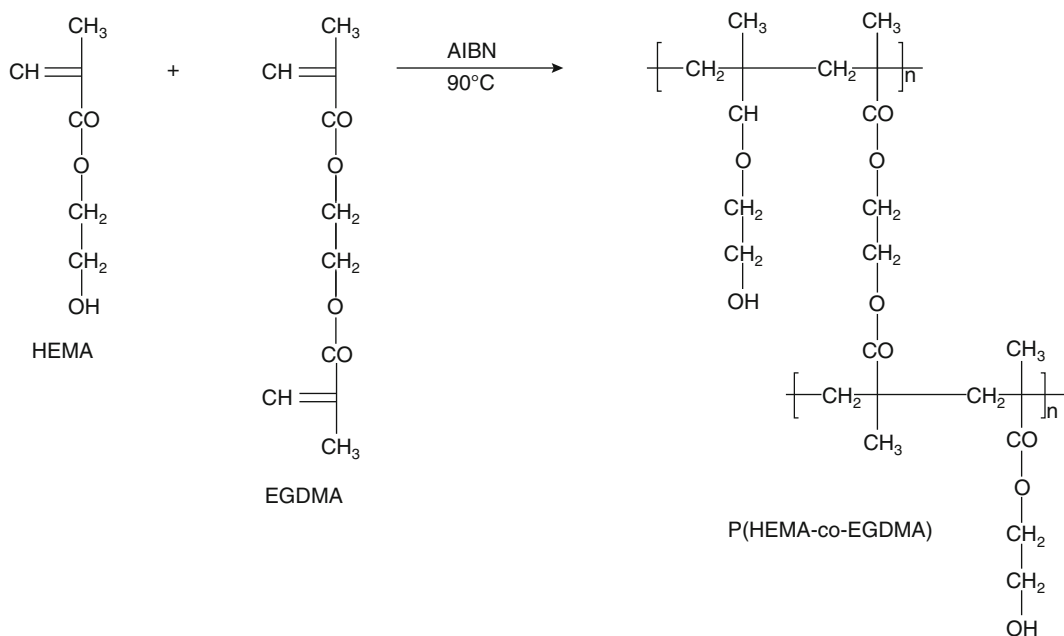


Fig. 20.1 Schematic representation of radical polymerization. Hydrogels are formed by the copolymerization of HEMA with EGDMA using AIBN as the radical initiator

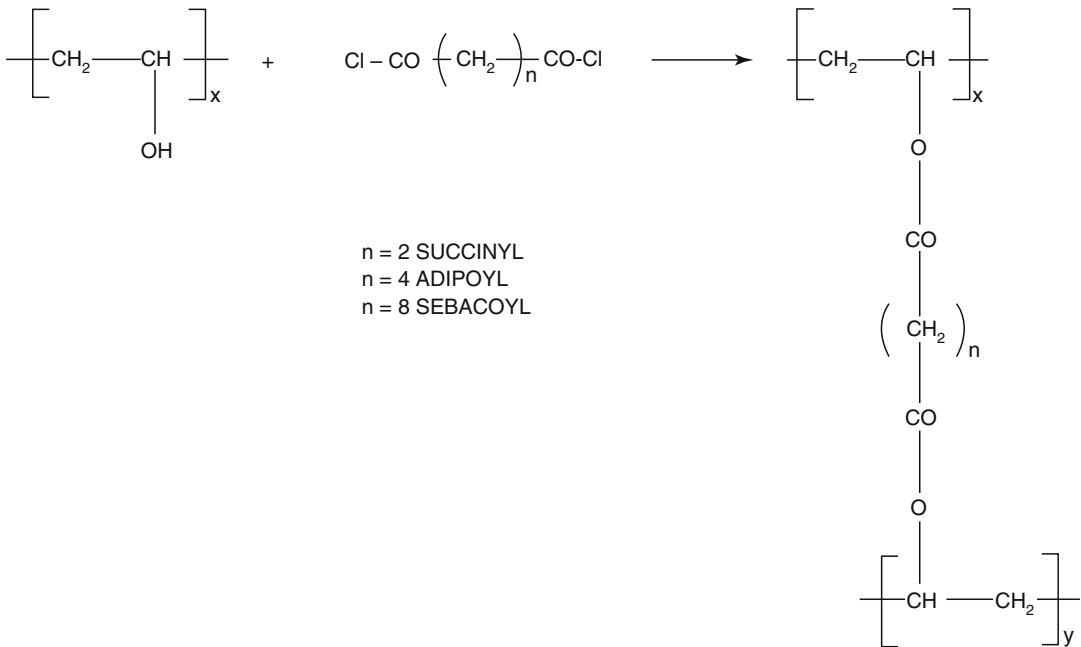


Fig. 20.2 Preparation of PVA hydrogels cross-linked by acyl dichlorides

radioactive isotopes such as cobalt 60, electron beams from electron accelerators, and X-rays converted from electron beams.

Physical cross-linking of hydrogels also avoids the use of chemical cross-linking agents. Such agents can potentially inactivate the active principle and covalently link it to the hydrogel network. Examples of ionically cross-linked alginate hydrogels have been reported (Grant et al. 1973). Alginate is a family of linear polysaccharides composed of mannuronic acid (MA) and guluronic acid (GA). The chemical composition and sequence of MA and GA residues depend on the source from which the alginate has been extracted. The gelation of alginate is mainly achieved by the exchange of sodium ions with divalent cations such as Ca^{2+} , Cu^{2+} , Zn^{2+} , or Mn^{2+} , which can form cation bridges between adjacent molecules. The “egg-box” model of Grant et al. (1973) is generally taken into consideration to explain the formation of a rodlike cross-linked complex due to the bounding of the divalent cations in the interchain cavities. Some polymeric complexes can be held together by hydrogen bonds: poly(acrylic acid) and poly(methacrylic acid) provide physically cross-linked hydrogels with poly(ethylene glycol)

due to the formation of hydrogen bonds between the oxygen of poly(ethylene glycol) and the carboxylic groups of the acrylic polymers (Eagland et al. 1994). Another physical method for producing physically cross-linked hydrogels is the formation of crystalline regions in the polymer network, obtained by casting dilute, aqueous solutions of poly(vinyl alcohol), then cooling to -20°C , and thawing back to room temperature several times (Stauffer and Peppas 1992). These freeze/thawed gels have demonstrated enhanced physical properties, such as high mechanical strength and high elasticity, that make them suitable for biomedical applications.

Finally, physically cross-linked hydrogels can be obtained by hydrophobic modification of polymers and in particular of polysaccharides such as chitosan, dextran, pullulan, and carboxymethyl curdlan (Noble et al. 1999; Sludden et al. 2000; Cerchiara et al. 2002). Glycol chitosan substituted with palmitoyl chains is an example of a hydrophobized polysaccharide. The attachment of hydrophobic groups to glycol chitosan yields an amphiphilic polymer capable of self-assembly into vesicles (Uchegbu et al. 1998). Non-covalent cross-linking is achieved by the

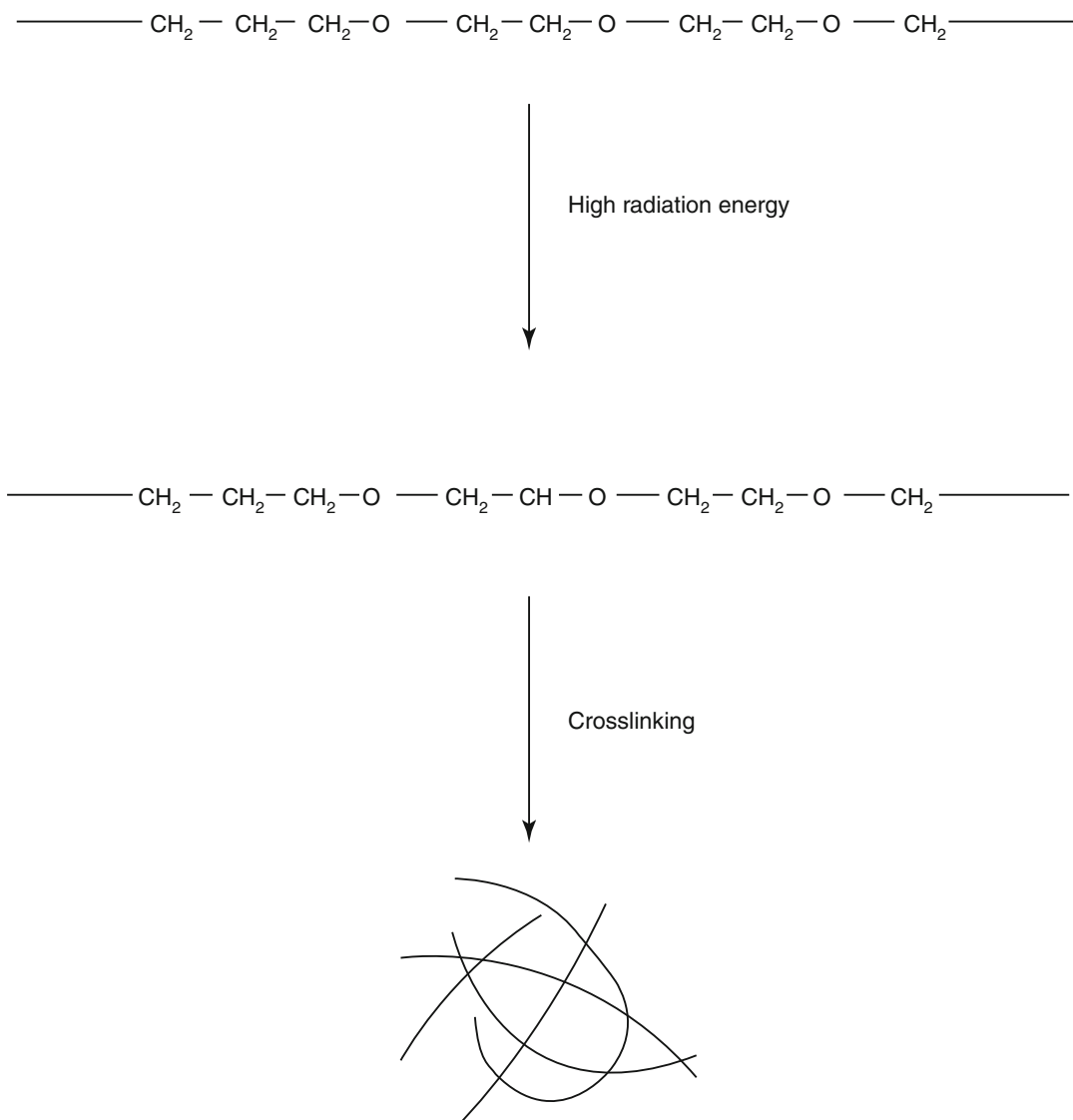


Fig. 20.3 Schematic representation of the radiation method to design hydrogels

hydrophobic interactions of the palmitoyl groups and a gel matrix is formed. Finally, our research group (Cerchiara et al. 2002) reported physically cross-linked chitosan hydrogels with lauric, myristic, palmitic, or stearic acid prepared by freeze drying and studied for transdermal use (Fig. 20.4). These polymers produce hydrogels with different functional properties related to the different acyl chains introduced in the polymer structure. In particular, the permeation of hydrophilic substances through the skin can be modulated by increased or decreased drug solubility

due to the interaction of the different acyl chains with the stratum corneum.

20.2 Applications of Hydrogels in Transdermal Drug Delivery

Transdermal delivery is an attractive and promising alternative compared to conventional administration routes (e.g., oral and injectable) for transport through the skin into the blood circulation of drugs

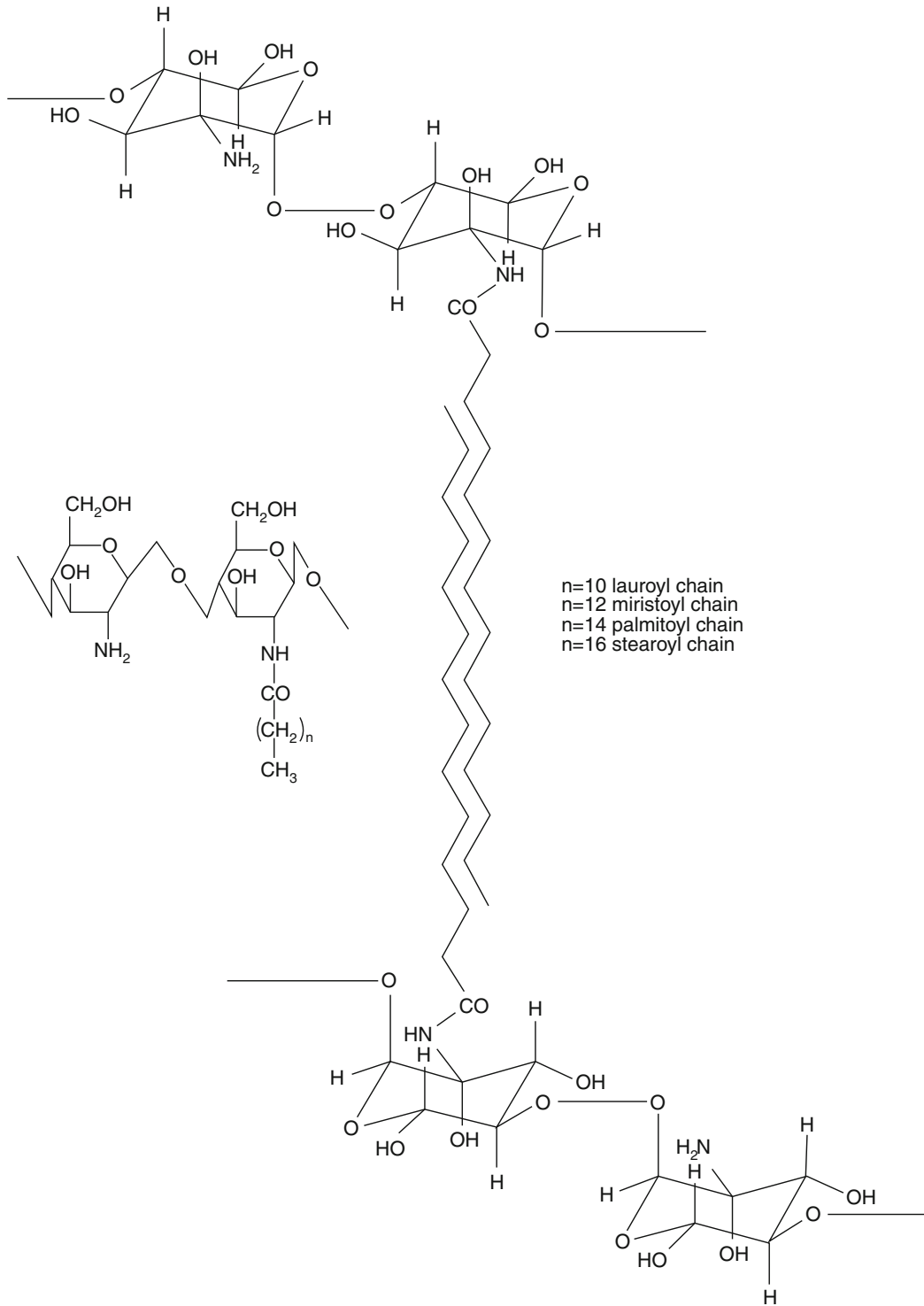


Fig. 20.4 Structural representation of physically cross-linked chitosan hydrogels

such as hydrophobic small molecules, hydrophilic molecules, and macromolecules. In fact, this route offers several advantages over conventional routes (Peppas et al. 2000; Brown et al. 2006; Prausnitz and Langer 2008):

- Is noninvasive and pain-free, with no trauma or risk of infection (Denet et al. 2004), thus resulting in an enhanced patient compliance
- Maintains constant drug levels in the blood, eliminating plasma peaks and valleys associated to oral and injectable administration
- Avoids gastrointestinal tract and circumvents hepatic first-pass metabolism, thus resulting in a lower drug amount administered and in a reduction of systemic side effects
- Is inexpensive

In spite of the advantages, major disadvantage of transdermal drug delivery is that the drug itself or the materials used to fabricate the vehicles may sometimes induce an irritation or sensitization reaction of the skin (Kurihara-Bergstrom et al. 1991; Murphy and Carmichael 2000; Ale et al. 2009; Wohlrab et al. 2011). Moreover, few molecules have been successfully delivered transdermally, mainly due to the stratum corneum that forms a barrier to the permeation of hydrophilic drugs, especially macromolecules such as proteins, peptides, and vaccines.

Consequently, research trends are focusing on approaches to overcome the barrier presented by the skin, including physical penetration enhancers such as microneedles (Henry et al. 1998), iontophoresis and electroosmosis (Pikal 2001), electroporation (Hu et al. 2000), radiofrequency energy (Sintov et al. 2003), and chemical penetration enhancers such as sulfoxides, alkanones, alcohols, polyols, amides, fatty acids, fatty acid esters, surfactants, terpenes, organic acids, and cyclodextrins (Thong et al. 2007). Recent studies have introduced a new category of transdermal penetration enhancers such as positively charged polymers. In fact, Taveira et al. (2009) described that chitosan interacts with negative charges in the skin improving drug penetration in the deeper layers, while He et al. (2008, 2009) demonstrated that chitosan and its derivatives such as N-trimethyl chitosan and mono-N-carboxymethyl chitosan are able to change the secondary structure of keratin

in stratum corneum leading to a less organized structure of this protein and enhancing transdermal permeation of drugs.

As regards the transdermal drug delivery systems, the traditional formulations as plasters, cream, and ointments have been replaced in order to minimize skin irritation, promote adhesion properties, guarantee dosage flexibility, enhance patient acceptability, and improve ease of use. In this context, particular attention has been paid to the formulations based on hydrogels such as semisolid systems and film-based systems (matrix-type systems, membrane-coated systems, film-forming solution).

20.2.1 Semisolid Vehicles

Hydrogels are three-dimensional networks based on linear hydrophilic polymers that are able to absorb large amounts of water, remaining insoluble due to the presence of chemical or physical cross-links (Peppas et al. 2000). The relatively high water content makes them a good alternative to other dosage forms such as creams, ointments, and patches, enhancing skin moisturization and elasticity and providing a better feel when applied to the skin. The most important and well-known polymers used for preparing these hydrogels are biopolymers such as polysaccharides (starch, cellulose, chitin, alginate, hyaluronate) or proteins (collagens, gelatins, caseins, albumins) and synthetic polymers such as polyvinyl alcohol, polyvinylpyrrolidone, polyethylene glycol, and polyacrylates.

Chitosan, a polysaccharide comprising copolymer of glucosamine and N-acetylglucosamine, derived by the partial deacetylation of chitin, is a nontoxic and bioabsorbable polymer (Muzzarelli et al. 1988; Luppi et al. 2010a) extensively studied for the release of many drugs. Our research group (Cerchiara et al. 2002) described physically cross-linked chitosan hydrogels with lauric, myristic, palmitic, or stearic acid able to enhance the skin permeation of propranolol hydrochloride selected as a hydrophilic model drug. The aim of the work was to improve the permeation of drugs through biological membranes, using hydrogels

made of amphiphilic polymer (Noble et al. 1999). The concomitant presence of hydrophobic and hydrophilic groups in the polymer influenced the swelling properties. So, at pH 7.4 all hydrogels swelled slowly and their behavior influenced the drug release. Among the different chitosan gels, chitosan laurate and chitosan myristate enhanced drug permeation through the skin with respect to chitosan palmitate and chitosan stearate hydrogels (flux values (mg/h cm^2) of propranolol hydrochloride from physically cross-linked chitosans oversaturated hydrogels through porcine skin were 1.00 ± 0.02 for chitosan laurate, 0.87 ± 0.05 for chitosan myristate, 0.47 ± 0.03 for chitosan palmitate, and 0.37 ± 0.01 for chitosan stearate). This could be explained by the interaction of the hydrogels with the stratum corneum, increasing the solubility of the drug in the skin.

Another example of hydrophilic and biocompatible polymer used to design hydrogels able to release hydrophilic drugs through the skin is polyvinyl alcohol. Polyvinyl alcohol cross-linked with succinyl, adipoyl, or sebacoyl chloride was employed as a supporting material to release propranolol hydrochloride. In particular, these hydrogels increased the transdermal permeation of drug, and as described in the previous work (Cerchiara et al. 2002), this effect seems to be linked to an increased drug solubility in the skin, probably produced by the interaction of the polymer with the stratum corneum. Moreover, the maximum enhancement of the drug permeation has been observed in the presence of the higher degree of cross-linking and the shorter length of the cross-linker acyl chain (Orienti et al. 2000).

Luppi and coworkers (2003) used also the cross-linked poly(methyl vinyl ether-co-maleic anhydride) (GZ) as a topical vehicles for pyridoxine hydrochloride, selected as a hydrophilic model drug. In particular, poly(methyl vinyl ether-co-maleic anhydride) was cross-linked with ethylene glycol (GZ-ET), butanediol (GZ-BUT), 1,6-exandiol (GZ-EX), 1,8-octanediol (GZ-OCT), 1,10-decanediol (GZ-DEC), or 1,12-dodecanediol (GZ-DOD). In vitro permeation studies were influenced by the nature of the cross-linker: the decrease in cross-linker acyl chain length provides vehicles

accelerating drug permeation through the skin. In fact, flux values (mg/h cm^2) of pyridoxine hydrochloride from hydrogels through porcine skin were 1.29 ± 0.12 for GZ, 5.83 ± 0.22 for GZ-ET, 4.91 ± 0.14 for GZ-BUT, 4.32 ± 0.10 GZ-EX, 3.82 ± 0.09 GZ-OCT, 3.25 ± 0.11 GZ-DEC, and 2.42 ± 0.12 GZ-DOD.

20.2.2 Film-Based Vehicles

Film-based vehicles are the widest utilized and studied transdermal delivery systems based on hydrogels. They are generally distinguished in matrix-type systems, membrane-coated systems, and film-forming solution.

The matrix-type systems are essentially a polymeric layer containing the drug, eventually added with an adhesive layer to enhance the bioadhesion and an impermeable layer to force the release to the skin. In absence of the adhesive layer and in order to assure a controlled delivery of the drug, it is necessary to choose polymers able to guarantee an intimate and prolonged contact with the skin after the application and flexibility and elasticity sufficient to follow the movements of the skin. An adhesive hydrogel patch based on a hydrophilic matrix of poly(N-vinylpyrrolidone) and oligomeric short-chain poly(ethylene glycol) was reported by Feldstein and coworkers (1996). They observed that the delivery rates of drugs with various chemical structures (propranolol, glyceryl trinitrate, isosorbide dinitrate) from the hydrophilic transdermal systems were higher than from the hydrophobic ones (stirene-butadiene rubber/mineral oil, polydimethylsiloxane/ silicone oil, polyisobutylene/mineral oil), and the drug delivery from the hydrophilic matrix across human cadaver skin epidermis or skin-imitating Carbosil membrane in vitro was characterized by zero-order drug delivery kinetics up to the point of 75–85 % drug release from initial contents in matrix. Drug delivery rates from the hydrophilic matrix were controlled by the skin or skin-imitating membrane permeability and may be described by Fick's law (Iordanskii et al. 2000).

Ethylene-vinyl acetate (EVA) matrix was tested as a system for transdermal delivery of atenolol in the presence of plasticizers able to increase the rate of drug release. The effects of drug concentration, temperature, and plasticizers on drug release were investigated. The release rate from EVA matrix was enhanced increasing temperature and drug concentration. In particular, the release rate of drug increased about 1.72-fold when the temperature of release system was raised from 32 to 42 °C and about 1.24-fold when the drug loading dose was increased from 0.5 to 1.5 %. Moreover, drug release from the polymeric matrix followed a diffusion-controlled model, where the quantity released per unit area was proportional to the square root of time. Among the plasticizers used such as alkyl citrates and phthalates, tributyl citrate showed the best enhancing effects: flux values ($\mu\text{g}/\text{cm}^2/\text{h}^{1/2}$) were 8.056 and 5.327 for EVA matrix containing tributyl citrate and EVA matrix without plasticizer, respectively. The results obtained confirmed that ethylene-vinyl acetate matrix could be used for transdermal delivery of hydrophilic drug (Kim and Shin 2004).

Padula and coworkers (2003) proposed a polyvinyl alcohol film not adhesive in the dry state, but bioadhesive when applied on wet skin. The film was applied to the skin in the presence of a certain amount of water. Water swelled the film on the surface in contact with the skin, transforming a dry polymeric matrix into a jellified polymer layer. This particular film is flexible, is mechanically resistant, and can avoid skin occlusion because of its permeability to water vapor. Compared to a typical patch, the bioadhesive film has a monolayer structure which includes backing, adhesive, and drug reservoir functions. This simple delivery system composed of a smaller number of layers simplifies the preparation procedure of transdermal patches and represents a great innovation in the field of transdermal patch.

Another polymer that represents a good candidate as a starting material for film-based vehicles because of its good film-forming properties is chitosan (Mengatto et al. 2012). Frequently, chitosan is modified by cross-linking reactions to achieve films with improved mechanical properties and obtain an efficient control of drug delivery. In this

context, films based on polyelectrolyte complexes were developed for topical and transdermal administration of drugs by Silva et al. (2008). The complexes were prepared with chitosan and different polyacrylic acid polymers, cross-linked with allyl pentaerythritol (Carbopol 71G NF®) or divinylglycol (Noveon AA-1®) at different cross-linking densities. The interaction between the polymers was maximized controlling the preparative conditions of complexes, and the film properties were improved by means of different plasticizers (glycerol or polyethylene glycol 200), a moisturizing agent (Hydrovance®), and an hydrophilic pressure sensitive adhesive (polyvinylpyrrolidone/polyethylene glycol 400). Between the different vehicles obtained, the film prepared by cross-linking with Noveon, plasticized with glycerol and covered with adhesive, has shown very good flexibility, resistance, and bioadhesion, making it a good candidate for further incorporation of drugs for topical and transdermal administration. Recently, chitosan-polyvinyl alcohol blend reticulated with glutaraldehyde has been utilized to prepare transdermal film suitable for insulin release in diabetes chemotherapy (Zu et al. 2012). The hydrogel obtained had a honeycomb-like structure and showed good mechanical and thermal properties. Moreover, the *in vitro* release studies showed that insulin release is comply with Fick's first law of diffusion showing a high permeation rate ($4.421 \mu\text{g}/(\text{cm}^2\text{h})$).

Transdermal delivery represents also an important opportunity for vaccine administration (Prausnitz and Langer 2008). In fact, although vaccines are generally large entities such as macromolecules or viral particles, their small dose facilitates transdermal administration. Ishii et al. (2008) describes simple, easy-to-use, noninvasive transcutaneous vaccination system, formed by an adhesive matrix. This patch is composed of cross-linked acrylic medical adhesive, octyldodecyl lactate, glycerin, and sodium hyaluronate and delivers antigenic proteins to Langerhans cells resident in the epidermal layer without destroying or removing the stratum corneum and induces Th2 (Type 2 helper T cells)-dominant immune response (production of neutralizing immunoglobulin G1 antibodies), effectively preventing viral and bacterial infection.

Finally, Luppi et al. (2010b) formulated transdermal hydroxypropyl methylcellulose-based films containing chlorpromazine hydrochloride for the treatment of psychotic disorders. Film composition was modified by incorporating a chemical permeation enhancer or binary enhancer combinations (oleic acid or polysorbate 80, or both) and a plasticizer (propylene glycol). Both oleic acid and polysorbate 80 had significant effect on drug permeation with respect to the control formulation and films containing a mixture of oleic acid and polysorbate 80 provided the best enhancement activity for chlorpromazine. In fact, the amount of chlorpromazine hydrochloride permeated through pig ear skin from hydroxypropyl methylcellulose films after 100 h were 15.2 ± 0.7 mg/cm² for film containing oleic acid (2.15, % w/w on dry basis) and polysorbate 80 (2.15, % w/w on dry basis) and 6.5 ± 0.3 mg/cm² for control formulation (film with the same composition, but without permeation enhancers). Moreover, also the hydroxypropyl methylcellulose type and the different concentration of drug and plasticizer contributed to modulate drug permeation. A decrease of hydroxypropyl methylcellulose viscosity, as a function of its molecular weight, and an increase in propylene glycol and chlorpromazine content provided higher cumulative amounts of drug permeated. These results confirm that chlorpromazine permeation can be easily modulated by varying the composition of hydroxypropyl methylcellulose-based films.

Another type of film-based vehicles is membrane-coated systems that consist of a drug depot and an hydrogel-based membrane able to control drug release. Tacharodi and Panduranga Rao (1995) proposed a patch consisting of a chitosan membrane cross-linked with different concentrations of glutaraldehyde and a chitosan gel as propranolol hydrochloride reservoir. Drug release can be easily tailored by changing cross-link density within the membrane and strongly depends on the area of the device. In fact, devices characterized by a diameter of 2.5 cm and chitosan membrane having low cross-link density released about 12 mg of drug within 24 h, while devices with diameter of 2.5 cm and membrane

having high cross-link density released 7.5 mg within 24 h. Moreover, devices having a diameter of 2.5 cm and characterized by uncross-linked chitosan membrane released about 15 mg after 24 h, while the same device having a diameter of 1.5 cm released about 4 mg after 24 h. Finally, all the devices delivered propranolol hydrochloride in a near zero order fashion suggesting that these chitosan membranes might be used successfully for the fabrication of membrane-controlled transdermal delivery systems.

A more innovative approach in transdermal drug delivery is represented by film-forming solutions. These systems are essentially polymer solutions, containing dispersed or dissolved drug. After application onto the skin, solvent evaporation guarantees the formation of a bioadhesive and thin film. Compared with matrix-type vehicles and membrane-coated vehicles, these systems are very easy to prepare and they possess higher dosage flexibility, less irritation of the skin, better cosmetic appearance associated with high ease of use. Schroeder et al. (2007) tested a broad range of polymers (acrylates, polyurethane-acrylates, cellulose derivatives, polyvinylpyrrolidones, silicones) as film-forming materials. Polymers at different concentration were solubilized in a volatile solvent (ethanol or volatile silicone), added with different amounts of plasticizer (triethyl citrate, triacetin, dibutyl phthalate) and possibly with a cross-linking agent (succinic acid). Formulations with adequate properties for the application on the skin (low viscosity, short drying time, low outward stickiness, high cosmetic attractiveness, and integrity on the skin for a prolonged time) were evaluated in terms of mechanical properties (tensile strength and elongation at break), water vapor permeability, and transepidermal water loss. The positively evaluated preparations resulting from these experiments provide the basis for the development of film-forming polymeric solutions as a transdermal dosage form.

A novel organic-inorganic hybrid film-forming agent for transdermal drug delivery was prepared by a modified poly(vinyl alcohol) (PVA) gel using γ -(glycidyoxypropyl)trimethoxysilane (GPTMS) as a cross-linking agent, poly(N-

vinyl pyrrolidone) as a tackifier, and glycerol as a plasticizer. The obtained gels can be applied to the skin by a coating method and in situ form very thin and transparent films with good flexibility and adhesive properties. Furthermore, the skin irritation tests showed that the formulations produced no skin irritation after topical application, while the in vitro release studies revealed that these films are able to release both hydrophilic drugs such as 5-fluorouracil both lipophilic molecules such as ibuprofen. The cumulative release of hydrophilic 5-fluorouracil was much higher than hydrophobic ibuprofen and introduction of adequate GPTMS amount (GPTMS/(PVA + GPTMS) ratio=20–30 %) into PVA matrix can decrease the crystalline regions of PVA and enhance drug diffusion (Guo et al. 2011).

Conclusions

Transdermal delivery is a major administration route for drugs that are destroyed by the liver when taken orally (Langer 2004). Recently, much attention has been paid to the hydrogels as vehicles for transdermal drug delivery, and their success can be mainly attributed to the possibility to modulate drug release kinetics (Kim et al. 1992). Currently, few drugs have been successfully delivered through the skin utilizing hydrogels, and new opportunities may be envisaged.

References

- Ale I, Lachapelle JM, Maibach HI (2009) Skin tolerability associated with transdermal drug delivery systems: an overview. *Adv Ther* 26(10):920–935
- Anderson JM, Shive MS (1997) Biodegradation and biocompatibility of PLA and PLGA microspheres. *Adv Drug Deliv Rev* 28:5–24
- Berger J, Reist M, Mayer MJ, Felt O, Peppas NA, Gurny R (2004) Structure and interactions in covalently and ionically crosslinked chitosan hydrogels for biomedical applications. *Eur J Pharm Biopharm* 57:19–34
- Brasch U, Burchard W (1996) Preparation and solution properties of microhydrogels from poly(vinyl alcohol). *Macromol Chem Phys* 197:223–235
- Brown MB, Martin GP, Jones SA, Akomeah FK (2006) Dermal and transdermal drug delivery systems: current and future prospects. *Drug Deliv* 13(3):175–187
- Cerchiara T, Luppi B, Bigucci F, Orienti I, Zecchi V (2002) Physically cross-linked chitosan hydrogels as topical vehicles for hydrophilic drugs. *J Pharm Pharmacol* 54:1453–1459
- Chen L, Tian Z, Du Y (2004) Synthesis and pH sensitivity of carboxymethyl chitosan-based polyampholyte hydrogels for protein carrier matrices. *Biomaterials* 25(17):3725–3732
- Damink LHHO, Dijkstra PJ, vanLuyn MJA, vanWachem PB, Nieuwenhuis P, Feijen J (1996) In vitro degradation of dermal sheep collagen cross-linked using a water-soluble carbodiimide. *Biomaterials* 17:679–684
- Denet AR, Vanbever R, Pr  at V (2004) Skin electroporation for transdermal and topical delivery. *Adv Drug Deliv Rev* 56:659–674
- Draye JP, Delaey B, van de Voorde A, van den Bulcke A, Bogdanov B, Schacht E (1998) In vitro release characteristics of bioactive molecules from dextran dialdehyde cross-linked gelatin hydrogel films. *Biomaterials* 19:99–107
- Eagland D, Crowther NJ, Butler CJ (1994) Complexation between polyoxyethylene and polymethacrylic acid – the importance of the molar mass of polyethylene. *Eur Polym J* 30:767–773
- Eliaz RE, Kost J (2000) Characterization of a polymeric PLGA-injectable implant delivery system for the controlled release of proteins. *J Biomed Mater Res* 50:388–396
- Feldstein MM, Tohmakhchi VN, Malkhazov LB, Vasiliev AE, Plate NA (1996) Hydrophilic polymeric matrices for enhanced transdermal drug delivery. *Int J Pharm* 131:229–242
- Grant G, Morris ER, Rees DA, Smith PJC, Thom D (1973) Biological interaction between polysaccharides and divalent cations: the egg-box model. *FEBS Lett* 32(1):195–198
- Guo R, Du X, Zhang R, Deng L, Dong A, Zhang J (2011) Bioadhesive film formed from a novel organic-inorganic hybrid gel for transdermal drug delivery system. *Eur J Pharm Biopharm* 79(3):574–583
- Guy RH (1996) Current status and future prospects of transdermal drug delivery. *Pharm Res* 13:1765–1769
- He W, Guo X, Zhang M (2008) Transdermal permeation enhancement of N-trimethyl chitosan for testosterone. *Int J Pharm* 356(1–2):82–87
- He W, Guo X, Xiao L, Feng M (2009) Study on the mechanisms of chitosan and its derivatives used as transdermal penetration enhancers. *Int J Pharm* 382(1–2):234–243
- Hennink WE, van Nostrum CF (2002) Novel crosslinking methods to design hydrogels. *Adv Drug Deliv Rev* 54(1):13–36
- Henry S, McAllister DV, Allen MG, Prausnitz MR (1998) Microfabricated microneedles: a novel approach to transdermal drug. *J Pharm Sci* 87:922–925
- Hickey AS, Peppas NA (1995) Mesh size and diffusive characteristics of semicrystalline poly(vinyl alcohol) membranes prepared by freezing/thawing techniques. *J Membr Sci* 107:229–237

- Hoffman AS (1991) Environmentally sensitive polymers and hydrogels – “smart” biomaterials. *MRS Bull XVI*:42–46
- Hoffman AS (2002) Hydrogels for biomedical applications. *Adv Drug Deliv Rev* 54:3–12
- Hu Q, Liang W, Bao J, Ping Q (2000) Enhanced transdermal delivery of tetracaine by electroporation. *Int J Pharm* 202:121–124
- Iordanskii AL, Feldstein MM, Markin VS, Hadgraft J, Plate NA (2000) Modeling of the drug delivery from a hydrophilic transdermal therapeutic system across polymer membrane. *Eur J Pharm Biopharm* 49:287–293
- Ishii Y, Nakae T, Sakamoto F, Matsuo K, Quan YS, Kamiyama F et al (2008) A transcutaneous vaccination system using a hydrogel patch for viral and bacterial infection. *J Control Release* 131(2):113–120
- Jatav VS, Singh H, Singh SK (2011) Recent trends on hydrogels in human body. *IJRPBS* 2:442–447
- Kabanov VY (1998) Preparation of polymeric biomaterials with the aid of radiation-chemical methods. *Russ Chem Rev* 67:783–816
- Khare AR, Peppas NA, Massimo G, Colombo P (1992) Measurement of the swelling force in ionic polymeric networks. I. Effect of pH and ionic content. *J Control Release* 22:239–244
- Kim J, Shin SC (2004) Controlled release of atenolol from the ethylene-vinyl acetate matrix. *Int J Pharm* 273: 23–27
- Kim SW, Bae YH, Okano T (1992) Hydrogels: swelling, drug loading and release. *Pharm Res* 9:283–290
- Kopecka J (2007) Hydrogel biomaterials: a smart future? *Biomaterials* 28:5185–5192
- Kurihara-Bergstrom T, Good WR, Feisuln S, Signur C (1991) Skin compatibility of transdermal drug delivery systems. *J Control Release* 15:271–278
- Langer R (2004) Transdermal drug delivery: past progress, current status and future prospects. *Adv Drug Deliv Rev* 56:557–558
- Lee KY, Bouhadir KH, Mooney DJ (2004) Controlled degradation of hydrogels using multi-functional cross-linking molecules. *Biomaterials* 25(13):2461–2466
- Luo Y, Kirker RK, Prestwich GD (2000) Crosslinked hyaluronic acid hydrogels films: new biomaterials for drug delivery. *J Control Release* 69:169–184
- Luppi B, Cerchiara T, Bigucci F, Di Pietra AM, Orienti I, Zecchi V (2003) Crosslinked poly(methyl vinyl ether-co-maleic anhydride) as topical vehicles for hydrophilic and lipophilic drugs. *Drug Deliv* 10:239–244
- Luppi B, Bigucci F, Cerchiara T, Zecchi V (2010a) Chitosan-based hydrogels for nasal drug delivery: from inserts to nanoparticles. *Expert Opin Drug Deliv* 7:811–828
- Luppi B, Bigucci F, Baldini M, Abruzzo A, Cerchiara T, Corace G et al (2010b) Hydroxypropylmethylcellulose films for prolonged delivery of the antipsychotic drug chlorpromazine. *J Pharm Pharmacol* 62:305–309
- Mengatto LN, Helbling IM, Luna JA (2012) Recent advances in chitosan films for controlled release of drugs. *Recent Pat Drug Deliv Formul* 6(2):156–170
- Miyata T, Urugami T, Nakamae K (2002) Biomolecule-sensitive hydrogels. *Adv Drug Deliv Rev* 54(1):79–98
- Murdan S (2003) Electro-responsive drug delivery from hydrogels. *J Control Release* 92(1–2):1–17
- Murphy M, Carmichael AJ (2000) Transdermal drug delivery systems and skin sensitivity reactions: incidence and management. *Am J Clin Dermatol* 1(6):361–368
- Muzzarelli R, Baldassarre V, Conti F, Ferrara P, Biagini G, Gazzanelli G et al (1988) Biological activity of chitosan: ultrastructural study. *Biomaterials* 9:247–252
- Nedkov E, Tsvetkova S (1994) Effect of γ -irradiation on the crystalline structure of ultra high molecular weight poly(ethylene oxide). *Radiat Phys Chem* 43:397–401
- Noble L, Gray AL, Sadiq L, Uchegbu IF (1999) A non-covalently cross-linked chitosan based hydrogel. *Int J Pharm* 192(2):173–182
- Orienti I, Di Pietra A, Luppi B, Zecchi V (2000) Crosslinked polyvinylalcohol hydrogels as vehicles for hydrophilic drugs. *Arch Pharm Pharm Med Chem* 333:421–424
- Padula C, Colombo G, Nicoli S, Catellani PL, Massimo G, Santi P (2003) Bioadhesive film for the transdermal delivery of lidocaine: in vitro and in vivo behaviour. *J Control Release* 88(2):277–285
- Peppas NA, Mikos AG (1986) Preparation methods and structure of hydrogels. In: Peppas NA (ed) *Hydrogels in medicine and pharmacy*. CRC press, Boca Raton, pp 1–25
- Peppas NA, Bures P, Leobandung W, Ichikawa H (2000) Hydrogels in pharmaceutical formulations. *Eur J Pharm Biopharm* 50:27–46
- Peppas NA, Hilt JZ, Khademhosseini A, Langer R (2006) Hydrogels in biology and medicine: from molecular principles to bionanotechnology. *Adv Mater* 18:1345–1360
- Pikal MJ (2001) The role of electroosmotic flow in transdermal iontophoresis. *Adv Drug Deliv Rev* 46:281–305
- Prausnitz MR, Langer R (2008) Transdermal drug delivery. *Nat Biotechnol* 26(11):1261–1268
- Qu X, Wirsén A, Albertson AC (1999) Synthesis and characterization of pH-sensitive hydrogels based on chitosan and D, L-lactic acid. *J Appl Polym Sci* 74: 3186–3192
- Rosiak JM (1991) Hydrogel dressings. In: Clough RL, Shalaby SW (eds) *Radiation effects on polymers*. ACS symposium series 475. American Chemical Society, Washington, DC, pp 271–299
- Safrany A (1997) Radiation processing: synthesis and modification of biomaterials for medical use. *Nucl Inst Methods Phys Res B* 131(1–4):376–381
- Sahin S, Selek H, Ponchel G, Ercan MT, Sargon M, Hincal AA et al (2002) Preparation, characterization and in vivo distribution of terbutaline sulfate loaded albumin microspheres. *J Control Release* 82(2–3): 345–358
- Schroeder IZ, Franke P, Schaefer UF, Lehr C (2007) Development and characterization of film forming

- polymeric solutions for skin drug delivery. *Eur J Pharm Biopharm* 65(1):111–121
- Silva CL, Pereira JC, Ramalho A, Pais AACC, Sousa JJS (2008) Films based on chitosan polyelectrolyte complexes for skin drug delivery: development and characterization. *J Membr Sci* 320:268–279
- Sintov AC, Krimberk I, Daniel D, Hannan T, Sohn Z, Levin G (2003) Radiofrequency-driven skin micro-channeling as a new way for electrically assisted transdermal delivery of hydrophilic drugs. *J Control Release* 89:311–320
- Sludden J, Uchegbu IF, Schatzlein AG (2000) The encapsulation of bleomycin within chitosan based polymeric vesicles does not alter its biodistribution. *J Pharm Pharmacol* 52:377–382
- Stauffer SR, Peppas NA (1992) Poly(vinyl alcohol) hydrogels prepared by freezing-thawing cyclic processing. *Polymers* 33(18):3932–3936
- Taveira SF, Nomizo A, Lopez RFV (2009) Effect of the iontophoresis of a chitosan gel on doxorubicin skin penetration and cytotoxicity. *J Control Release* 134:35–40
- Thacharodi D, Rao KP (1995) Development and *in vitro* evaluation of chitosan-based transdermal drug delivery systems for the controlled delivery of propranolol hydrochloride. *Biomaterials* 16:145–148
- Thong HY, Zhai H, Maibach HI (2007) Percutaneous penetration enhancers: an overview. *Skin Pharmacol Physiol* 20(6):272–282
- Timmer MD, Jo S, Wang C, Ambrose CG, Mikos AG (2002) Characterization of the cross-linked structure of fumarate-based degradable polymer networks. *Macromolecules* 35:4373–4379
- Uchegbu IF, Schatzlein AG, Tetley L, Gray AI, Sludden J, Siddique S, Mosha E (1998) Polymeric chitosan-based vesicles for drug deliver. *J Pharm Pharmacol* 50:453–458
- Wang C, Steward RJ, Kopecek J (1999) Hybrid hydrogels assembled from synthetic polymers and coiled-coil protein domains. *Nature* 397:417–420
- Watanabe T, Ohtsuka A, Murase N, Barth P, Gersonde K (1996) NMR studies on water and polymer diffusion in dextran gels. Influence of potassium ions on microstructure formation and gelation mechanism. *Magn Reson Med* 35:697
- Wichterle O, Lim D (1960) Hydrophilic gels for biological use. *Nature* 185:117–118
- Wohlrab J, Kreft B, Tamke B (2011) Skin tolerability of transdermal patches. *Expert Opin Drug Deliv* 8(7):939–948
- Xiao C, Zhou G (2003) Synthesis and properties of degradable poly(vinyl alcohol) hydrogel. *Polym Degrad Stab* 81(2):297–301
- Zu Y, Zhang Y, Zhao X, Shan C, Zu S, Wang K et al (2012) Preparation and characterization of chitosan–polyvinyl alcohol blend hydrogels for the controlled release of nano-insulin. *Int J Biol Macromol* 50:82–87