

Biofilm Inhibition by Nanoparticles

D. Bakkiyaraj and S.K. Pandian

Abstract Infectious diseases are of immediate concern due to their high rate of morbidity and mortality. Infectious diseases are life threatening in the current scenario as the causative agents are resistant to almost all the drugs in use. Apart from well-known factors like efflux pumps, receptor modifications, and drug inactivation, formation of biofilms attributes to broad-spectrum resistance toward antimicrobials. This necessitates the search for novel therapeutics that effectively control drug-resistant pathogens. Targeting biofilm formation is one such strategy to combat infectious diseases much more effectively. For over a decade diverse sources of synthetic to semisynthetic agents derived from microbes to plants have been tested for their antibiofilm potential with limited success. The birth of nanotechnology provided new insights into antibiofilm research as these nanoparticles are highly reactive and effective in penetrating the biofilm matrix. This chapter comprehensively summarizes the synthesis, application, weakness, and antibiofilm potential of nanoparticles.

1 Introduction

Infectious diseases are of major concern as they can result in high mortality and morbidity. Bacteria and fungi are the major pathogens that cause infections in humans and development of resistance by these organisms contributes to the severity of infections. Diseases caused by multidrug-resistant (MDR) pathogens are extremely difficult to treat and have a major impact on the economy. Even though numerous mechanisms like activation of efflux pumps, decreased permeability to antagonists, production of enzymes (e.g., beta-lactamase to inactivate antimicrobials), and mutation in target proteins facilitate resistance, biofilm

D. Bakkiyaraj • S.K. Pandian (✉)
Department of Biotechnology, Alagappa University, Science Campus, Karaikudi 630004,
TN, India
e-mail: sk_pandian@rediffmail.com

formation has been the predominant mechanism of broad-spectrum tolerance. Biofilm are complexes made up of microbes surrounded by a hydrated matrix that is secreted by these indwelling microbes to protect or facilitate their growth in hostile environments. Biofilms are characterized by their extracellular polymeric substances (EPS) which contain polysaccharides, proteins, lipids, and nucleic acids. Apart from conferring resistance to the inhabitants, biofilms also facilitate various other functions like aggregation, retention of water and nutrients, absorption of nutrients, protection against host immune responses, and horizontal gene transfer. Studies have even suggested that the multicellular behavior of biofilm inhabitants is similar to higher multicellular organisms.

In theory, antibiofilm agents are less likely to cause selective pressure for the evolution of resistance because they do not kill pathogens as do antibiotics. Successful antibiofilm agents can either inhibit the formation of biofilms or disrupt mature biofilms. Antibiofilm agents are preferred over antibiotics in some instances as they prevent or disrupt biofilms, facilitating their clearance by the host immune system. Numerous sources from soil to sea and herbs to plants have been screened for antibiofilm activity. Even, some antibiotics have been shown to possess antibiofilm activity at sublethal concentrations. In addition, synthetic agents with antibiofilm properties are of interest because of their feasibility and availability for application.

Various agents such as synthetic chemicals, microbial secondary metabolites, phenolic compounds and other phytochemicals from plants, antibiotics at their sublethal concentrations, nucleases, proteases and other enzymes, peptides, etc., were shown to have the potential to inhibit biofilm formation and/or disrupt mature biofilms. Numerous synthetic chemicals like thiazolidinone derivatives (Pan et al. 2010; Rane et al. 2012), aminoimidazoles (Furlani et al. 2012), diazopyrazole derivatives (Raimondi et al. 2012), bromopyrrole alkaloids (Rane et al. 2013), etc., were shown to have antibiofilm potential against Gram-positive bacterial pathogens like *Staphylococcus aureus*, *Streptococcus epidermidis*, and *Enterococcus faecalis*. Other synthetics like niclosamide (Imperi et al. 2013), esomeprazole (Singh et al. 2012), chlorogenic acid (Karunanidhi et al. 2013), and zinc (Wu et al. 2013) showed promising results against various Gram negative bacteria, especially *Pseudomonas aeruginosa*. Other chemicals such as caspofungin (Bink et al. 2012) and farnesol (Ramage et al. 2002) were shown to be active against the biofilms of *Candida albicans*. Lastly, antibiotics at sublethal concentrations were shown to inhibit biofilms, which is of interest, as these sublethal concentrations are less likely to induce the development of resistance (Balaji et al. 2013; Gilbert et al. 2002; Latimer et al. 2012).

Various microbial extracts (Bakkiyaraj and Pandian 2010; Bakkiyaraj et al. 2012; Nithya et al. 2010b, 2011; Nithya and Pandian 2010) and their secondary metabolites like usnic acid and atranorin (Pompilio et al. 2013), glycolipid biosurfactants (Kiran et al. 2010), phenylacetic acid (Musthafa et al. 2012b), ophiobolins (Arai et al. 2013), and piperazinedione (Musthafa et al. 2012a) were reported to have antibiofilm properties against bacterial and fungal pathogens. Apart from the microbial metabolites, enzymes were also shown to inhibit the

formation of biofilms. Alpha amylase produced by *Bacillus subtilis* (Kalpana et al. 2012), acylase produced by *B. pumilus* (Nithya et al. 2010a), alginate lyase (Lamppa and Griswold 2013), and protease produced by *P. aeruginosa* and actinomycetes (Park et al. 2012a, b) were also shown to have antibiofilm potential against human bacterial pathogens.

Numerous plants have been reported to display antibiofilm activities against bacterial and fungal pathogens. Cinnamaldehyde (Brackman et al. 2008), methyl eugenol (Packiavathy et al. 2012), casbane diterpene (Cardoso Sa et al. 2012), curcumin (Packiavathy et al. 2013), taxodione derivatives (Kuzma et al. 2012), gallic acid and ferulic acid (Borges et al. 2012), and ellagic acid (Sarabhai et al. 2013) are the notable plant products with potential antibiofilm activity.

The latest developments in the field of antibiofilm research employ novel agents like peptides (Amer et al. 2010; Choi and Lee 2012; Reymond et al. 2013; Zhang et al. 2010) and nanoparticles (Anghel et al. 2012; Hernandez-Delgadillo et al. 2012, 2013; Lellouche et al. 2012b; Durmus and Webster 2013; Martinez-Gutierrez et al. 2013; Sawant et al. 2013) as antibiofilm agents. The synthesis, properties, and the application of nanoparticles as antibiofilm agents will be discussed in detail in this chapter.

2 Properties and Synthesis of Nanoparticles

Among the antibiofilm technologies that have recently emerged, nanotechnology is one of the most promising. Nanotechnology can be defined as “a technology of engineering functional systems at molecular scale.” Nanotechnology can also be defined as technology involving design, synthesis, and application of materials and devices whose size and shape have been engineered at nanoscale. Particles produced through nanotechnology are called “nanoparticles” and are typically sized less than 100 nm. Nanoparticles are highly reactive and preferred over other bioactive agents because of their higher surface area in contrast to their size. For example, 1 μg of particles of 1 nm^3 size have the same surface area as 1 g of particles of size 1 mm^3 . Huge surface area of these nanoparticles facilitates their use as drug carriers.

Even though diverse chemicals like chitosan (Du et al. 2008), carboxymethyl chitosan (Zhao et al. 2013b), poly-gamma-glutamic acid (Liu et al. 2013b), cellulose (Raghavendra et al. 2013), zinc oxide (ZnO) (Dutta et al. 2013; Jones et al. 2008), magnesium fluoride (Lellouche et al. 2009, 2012b, c), polyethyleneimine (Beyth et al. 2010), hydroxyapatite (Evliyaoglu et al. 2011), fullerene (Patel et al. 2013), lipids (terpinen-4-ol) (Sun et al. 2012), and silica (Besinis et al. 2014; Li and Wang 2013) were shown to be useful, metals are the prime component of most nanoparticles. Derivatives of metals, like their oxides, form the base material for synthesis of many nanoparticles. Silver (Antony et al. 2013; Apte et al. 2013b; Besinis et al. 2014; Chernousova and Epple 2013; Jain and Pradeep 2005; Mohanty et al. 2012), gold (Annamalai et al. 2013; Geethalakshmi and

Sarada 2012; Khan et al. 2012; Naz et al. 2013; Pender et al. 2013; Ramamurthy et al. 2013), copper (Eshed et al. 2012; Kim et al. 2006; Pandiyarajan et al. 2013; Pramanik et al. 2012; Singh et al. 2013; Thekkae Padil and Cernik 2013), titanium (Besinis et al. 2014; Jayaseelan et al. 2013; Li et al. 2013), and iron (Das et al. 2013; Grumezescu et al. 2011; Leuba et al. 2013) are the predominant members of metal oxide nanoparticles and other metals like bismuth (Hernandez-Delgado et al. 2012, 2013) and cerium oxide (Shah et al. 2012) are other metal nanoparticles shown to possess bioactive potential.

Nanoparticles are synthesized either through a scale-up process, where atoms are grouped together, or a scale-down process, where larger molecules are minced to nanoscale. Irrespective of the methods used, synthesis of nanoparticles involves evaporation/dissolution, nucleation, and growth.

The synthesis of nanoparticles by scale-down or sizing-down processes can be achieved either by attrition or milling, followed by size-dependent grouping and selection. Scale-up processes can be broadly classified into three groups: gas phase fabrication; liquid phase fabrication; and biosynthesis or green synthesis of nanoparticles.

2.1 Gas or Vapor Phase Nanoparticle Fabrication

This process involves the evaporation of solid and liquid precursors to gaseous precursors followed by supersaturation, producing an intermediate product. Nucleation or condensation of these intermediate products results in primary particles. These primary particles, upon grain growth and agglomeration, produce nanoparticles and nanoclusters, respectively. Methods that employ gas phase fabrication are as follows:

1. Methods using solid precursors (Iskandar 2009)
 - Inert gas condensation
 - Pulsed laser ablation
 - Spark discharge generation
 - Ion sputtering
2. Methods using liquid or vapor precursors (Suciu et al. 2003)
 - Chemical vapor synthesis
 - Spray pyrolysis
 - Laser pyrolysis/photochemical synthesis
 - Thermal plasma synthesis
 - Flame synthesis
 - Flame spray pyrolysis
 - Low-temperature reactive synthesis

2.2 *Liquid Phase Nanoparticle Fabrication*

Liquid phase fabrication involves wet chemistry and the general process includes the surface reaction of solid and liquid precursors to produce corresponding intermediate products. Such intermediate products are converted to primary particles either by nucleation or condensation similar to gas phase fabrication, followed by growth or agglomeration to produce nanoparticles or nanoclusters, respectively. Methods that employ liquid phase fabrication are:

1. Co-precipitation (Murray et al. 2000)
2. Solvothermal methods (Yang et al. 2006)
3. Sol-gel methods (Yu et al. 2004)
4. Synthesis in structure media (e.g., Microemulsion) (Capek 2004)
5. Microwave synthesis (Tsuji et al. 2005)
6. Sonochemical synthesis (Zhang and Yu 2003)

2.3 *Biological Synthesis of Nanoparticles*

Synthesis of nanoparticles catalyzed by bacteria or fungi or their products is of considerable interest as it employs cleaner and greener technology. Numerous fungi and bacteria have been utilized for the bioconversion of raw chemicals into nanoparticles. For instance, the ability of the marine yeast *Yarrowia lipolytica* to catalyze the synthesis of gold nanoparticles has been reported (Agnihotri et al. 2009; Apte et al. 2013a, b). Biosynthesis of silver, gold, and bimetallic nanoparticles by fungi like *Phanerochaete chrysosporium*, *Penicillium* sp., and *Neurospora crassa* has also been reported (Castro-Longoria et al. 2011; Du et al. 2010; Vigneshwaran et al. 2006). Similarly, the synthesis of silver nanoparticles with antimicrobial potential by psychrophilic bacteria such as *Pseudomonas antarctica* and *Arthrobacter kerguelensis* has also been reported (Shivaji et al. 2011). *Lactobacillus fermentum* (Sintubin et al. 2009) and *Shewanella oneidensis* (Suresh et al. 2010) were also shown to catalyze the production of silver nanoparticles with antimicrobial potential.

Though there are numerous reports on the microbe-mediated synthesis of nanoparticles, very few studies have described the biomolecules involved in this synthesis. For example, nitrate reductase along with a protein from *Aspergillus niger* and nitrate reductase along with rhamnolipids from *P. aeruginosa* were shown to be indispensable for the synthesis of nanoparticles (Gade et al. 2008; Kumar and Mamidyala 2011). Similarly, the role of cell-bound melanin produced by the yeast *Y. lipolytica* and certain proteins produced by marine fungi *A. tubingensis* and *Bionectria ochroleuca* in the synthesis of silver nanoparticles with antibiofilm activity have been reported recently (Apte et al. 2013a, b; Rodrigues et al. 2013).

3 Diverse Applications of Nanoparticles

Nanoparticles or nanomaterials in general have diverse applications in various fields.

3.1 Industrial Applications

Many microelectronic instruments such as transistors have adapted nanotechnology (Thompson and Parthasarathy 2006). Carbon nanotubes are reported to be the nanoscale alternatives to conventional semiconductor crystals because of their diverse electronic properties from metallic to semiconducting (Jacoby 2002) or superconducting (Cristina and Kevin 2005). Carbon nanotubes have been shown to be useful in making low-voltage field-emission displays (Carey 2003). Nanomaterials like aerogel intercalation electrode materials, nanocrystalline alloys, nanosized composite materials, carbon nanotubes, and nanosized transition metal oxides have shown promise in the development of lithium-ion batteries with increased capacity and lifecycle over their conventional counterparts (Liu et al. 2006a; Scott et al. 2011).

Nanocrystalline materials synthesized by the sol-gel technique exhibit foam-like structures called “aerogel” which find application as insulation material in industries because of their negligible thermal conductivity (Hrubesh and Poco 1995). Paints that have incorporated nanoparticles (Titanium oxide) demonstrate enhanced mechanical properties, such as scratch resistance. For example, the wear resistance of paint-nanocomposite coatings is claimed to be ten times higher than that of conventional acrylic paints (Mochizuki et al. 2013).

In the automobile industry, nanoparticles of carbon black act as filler in the polymer matrix of tires and are used for mechanical reinforcement. Nanocomposites containing the flakes of clay and plastics and nanosized clay are used in manufacturing the exteriors of cars with superior properties like scratch resistance compared to traditional materials.

Nanoparticles have found their way into the food industry due to their antimicrobial properties. For example, silver-montmorillonite (Ag-MMT) nanoparticles were used in the prevention of food spoilage (Costa et al. 2011). In addition to preventing the growth of food-spoiling microbes, Ag-MMT nanoparticles also preserved color, odor, and firmness of the food (Costa et al. 2011).

Nanoparticles also have potential in controlling pollution because of their ability to catalyze the conversion of toxic gases (carbon monoxide and nitrogen oxide) from the exhaust of vehicles and power generators. Iron nanoparticles, along with palladium, converted detrimental products in groundwater to inert or less harmful products (He and Zhao 2005). The nanoparticles were also shown to be effective in removing organic chlorine (a carcinogen) from water contaminated with the chlorine-based organic solvents (used in dry cleaners).

3.2 *Nanoparticles in Biotechnology and Medicine*

Carbon nanotubes have been used as probe tips in atomic force microscopy (AFM) which is used for high-resolution imaging of nucleic acids, immunoglobulins, etc. (Hafner et al. 2001). Molecular recognition and the chemical forces between the interacting molecules can be studied by attaching AFM tips bearing these biomolecules (Hafner et al. 2001).

Nanofiber scaffolds have been employed in the regeneration of cells and organs. Experiments on a hamster with a detached optic tract demonstrated that a peptide nanofiber scaffold could facilitate the regeneration of axonal tissue (Ellis-Behnke et al. 2006). Titanium dioxide and zinc oxide are used in sunscreens and cosmetics to absorb and reflect UV light.

Nanotube membranes can act as channels for highly selective transport of molecules and ions between solutions that are present on both sides of the membrane (Jirage et al. 1997). For instance, membranes containing nanotubes with small inner dimensions (less than 1 nm) were useful for the separation of small molecules on the basis of molecular size, while the nanotubes with larger inner diameters (20–60 nm) were used to separate proteins (Martin and Kohli 2003).

The ability of nanoparticles to target and penetrate specific cells and organs has also been explored in nanomedicine. Nanospheres made of biodegradable (facilitating timely release) polymers and drugs have potential applications in acidic microenvironments as in the case of tumor tissues or sites of inflammation (Kamaly et al. 2012). Nanoparticles acted as drug carriers for the targeted release of a conjugate containing chlorotoxin (a peptide that selectively binds to glioblastoma cells) and liposomes encapsulating antisense oligonucleotides or small interfering RNAs for effective treatment of glioblastoma (Costa et al. 2013). Similarly, numerous other studies have independently demonstrated the utility of nanoparticles as drug carriers in different tumor types (Amoozgar et al. 2013; Leifert et al. 2013; Liu et al. 2013a; Shi et al. 2013; Vivek et al. 2013).

In addition, surface-functionalized nanoparticles can be used to infuse cell membranes at a much higher level than nanoparticles without a functionalized surface, which can be employed for transfer of genetic material into living cells (Lewin et al. 2000). Silica nanospheres coated with ammonium groups (cation) can bind to DNA (anion) through electrostatic interactions, which could be used to deliver the latter into the cells (Kneuer et al. 2000).

Nanospheres can act as carriers for antigens and toxoids for potential use in vaccination. Studies involving antigen-coated polystyrene nanospheres as vaccine carriers targeting human dendritic cells have been under trial for nasal vaccination (Matsusaki et al. 2005). Studies have also unveiled the potential of nanoparticles in the diagnosis and treatment of various cancers. For instance, a study by Yin et al. (2013) showed enhanced anticancer action of curcumin upon coupling it with nanoparticles made from methoxy poly(ethylene glycol)-polycaprolactone (PCL) block copolymers (Yin et al. 2013). Similarly, the silver nanoparticles were shown to inhibit lung cancer cells in a concentration-dependent manner

(Sankar et al. 2013). Iron nanoparticles coupled with high-resolution MRI detected lymph node metastases in patients with prostate cancer at a stage undetectable by any other method (Harisinghani et al. 2003) and the gold nanoparticles were employed for the accurate detection of matriptase—a cancer biomarker protein overexpressed in all types of cancer (Deng et al. 2013). Lastly, nanoparticles made of compounds with oxygen vacancies (CeO_2 and Y_2O_3) (Schubert et al. 2006) have been demonstrated to possess neuroprotective and anti-apoptotic properties.

3.3 Antimicrobial Activity of Nanoparticles

Nanoparticles have been considered to be some of the most effectual bioactive agents mainly because of their large surface area to volume ratio (Hamouda 2012). Nanopowders possess antimicrobial properties against various bacterial, fungal and viral human pathogens (Koper et al. 2002; Bosi et al. 2003) and can rapidly kill bacterial cells (90 % in 1 h). The antibacterial properties of silver and titanium dioxide nanoparticles have been assessed as coatings for surgical masks (Li et al. 2006), in addition to many other clinical uses.

Nanoparticles shown to have antimicrobial effects include silver (Lara et al. 2010; Lok et al. 2006), titanium dioxide (Li et al. 2006), fullerenes (Bosi et al. 2003), zinc oxide (Brayner et al. 2006), and magnesium fluoride (Lellouche et al. 2012c). The antibacterial activity of fullerenes was reported against *Escherichia coli*, Salmonella and *Streptococcus* spp. (Bosi et al. 2003). The ability of zinc oxide nanoparticles to disturb the membrane permeability of *E. coli* has also been reported (Brayner et al. 2006). The wide spectrum antimicrobial activity of silver nanoparticles has been attributed to their ability to destabilize the bacterial outer membrane and deplete adenosine triphosphate (principal form of energy) in bacteria (Lara et al. 2010; Lok et al. 2006).

Fullerenes have also been shown to have neuroprotective, anti-apoptotic, and anti-HIV activities (Bosi et al. 2003). Size-dependent interactions of silver nanoparticles and HIV-1 virus were reported, which resulted in the inhibition of host–viral interactions (Elechiguerra et al. 2005). Numerous other studies have demonstrated the antimicrobial potential of various nanoparticles and drug–nanoparticle conjugates against bacterial, fungal, and viral pathogens (Zheng et al. 2013; Zhao et al. 2013a; Zhang et al. 2013c; Xiong et al. 2013; Westendorf 2013; Wang and Lim 2013; Wang et al. 2013; Vidic et al. 2013; Tavassoli Hojati et al. 2013; Su et al. 2013; Shimizu et al. 2013; Mohanty et al. 2012; Mallick et al. 2012; Lellouche et al. 2012a; Costa et al. 2011; Mukhopadhyay et al. 2010; Huda et al. 2010; Sanpui et al. 2008; Pinto et al. 2013; Hernandez-Delgadillo et al. 2013; Monteiro et al. 2012; Khan et al. 2012; Lara et al. 2010).

4 Nanoparticles as Antibiofilm Agents

The application of nanoparticles is an emerging area of antibiofilm or antipathogenic research. Nanoparticles are preferred over other agents due to their acute ability to penetrate EPS and cell membranes. Nanoparticles were found to be efficient drug carriers, effectively transporting drugs across the biofilm matrix. Silver, iron and zinc nanoparticles have received the most attention as antibiofilm agents. Silver nanoparticles are the predominant ones with antibiofilm activity against Gram-positive bacteria like *S. aureus*, *S. epidermidis*, *E. faecalis*, and *Streptococcus mutans*; Gram-negative bacteria like *P. aeruginosa*, *Salmonella paratyphi*, *E. coli*, and *Acinetobacter baumannii*; and fungal pathogens like *C. albicans* and *C. glabrata*. Details of nanoparticles with antibiofilm activities, along with their target pathogens, are given in Table 1.

The use of nanoparticles in combination with other antibiotics or drugs was found to have superior action than when alone. Chitosan nanoparticles loaded with Tamoxifen were effective in controlling tumor development in breast cancer cell lines (Vivek et al. 2013). Similarly, the side effects of daunorubicin were reduced significantly when combined with titanium oxide nanoparticles, which increased the target specificity and anticancer activity in leukemia cells (Zhang et al. 2012). Silver nanoparticles in combination with conventional antibiotics like ampicillin, chloramphenicol, and kanamycin have shown antibiofilm activity against Gram-positive and negative bacterial pathogens including *E. faecium*, *S. aureus*, *E. coli*, and *P. aeruginosa* (Hwang et al. 2012).

5 Demerits of Nanoparticles

Even though nanoparticles have historically been considered inert, they are actually highly reactive. The large surface area of nanoparticles can be both a pro and a con to their application in biology. Nanoparticles are commonly found in dust and aerosols. Inhaled nanoparticles deposited in the lungs are cleared through host processes such as mucociliary escalation into the gastrointestinal tract (from where they are eliminated through the feces) (Semmler et al. 2004), lymphatic system (Liu et al. 2006b), and circulatory systems (Oberdorster et al. 2005). Failure to clear these nanoparticles results in accumulation in lungs, subsequently increasing the risk of lung cancer (Borm et al. 2004). Accumulation of nanoparticles in lungs also elicits an inflammatory response that damages host tissues (Oberdorster et al. 1994). Adverse effects to nanoparticles include impaired phagocytosis, inflammation, epithelial cell proliferation followed by fibrosis, emphysema, and the initiation of tumors (Ferlin 1994; Oberdorster et al. 1994; Nikula et al. 1995; Dasenbrock et al. 1996; Driscoll et al. 1996; Borm et al. 2004).

Inhalation of nanoparticles can also result in immune suppression and reduction in the ability of the immune system to combat infections (Lucarelli et al. 2004).

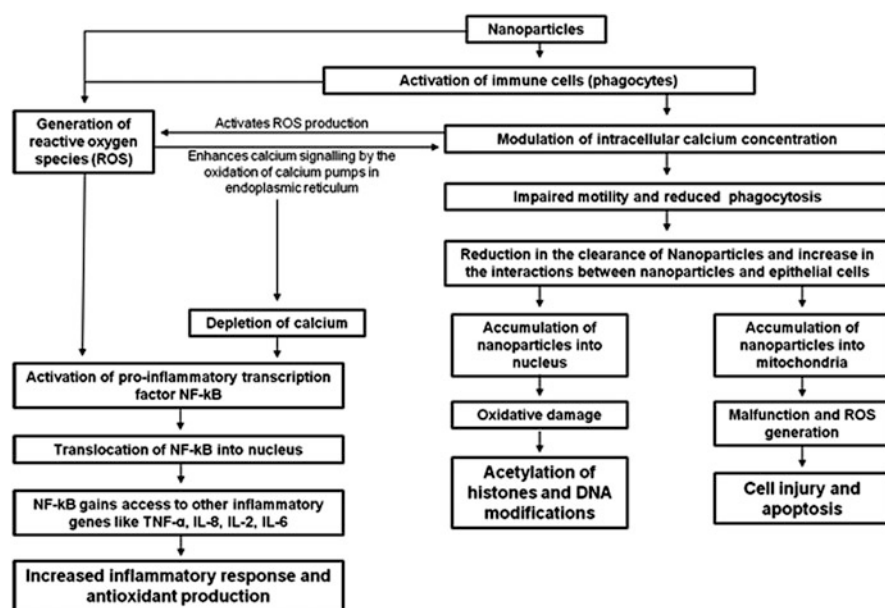
Table 1 Antibiofilm activity of nanoparticles and their target pathogens

Nanoparticle	Target organism	References
Silver nanoparticles	<i>S. paratyphi</i> , <i>P. aeruginosa</i> , <i>S. epidermidis</i>	Apte et al. (2013b), Kalishwaralal et al. (2010)
Bismuth oxide aqueous colloidal nanoparticles	<i>C. albicans</i> , <i>S. mutans</i>	Hernandez- Delgadillo et al. (2012, 2013)
Nano-oil formulation from <i>Mentha piperita</i> L.	<i>Staphylococcus</i> sp.	Anghel and Grumezescu (2013)
Nanoemulsion (detergent, oil, and water) in combination with cetylpyridinium chloride	<i>A. baumannii</i>	Hwang et al. (2013)
Silver and gold incorporated polyurethane, polycaprolactam, polycarbonate, and polymethylmethacrylate	<i>E. coli</i>	Sawant et al. (2013)
Silver nanoparticles in combination with nystatin and chlorhexidine	<i>C. albicans</i> , <i>C. glabrata</i>	Monteiro et al. (2012, 2013)
Silver nanoparticle and 12-methacryloyloxydodecylpyridinium bromide (MDPB)	Dental plaque micro-cosm biofilms	Zhang et al. (2013a, b)
Zinc	<i>Actinobacillus pleuropneumoniae</i> , <i>S. typhimurium</i> , <i>Haemophilus parasuis</i> , <i>E. coli</i> , <i>S. aureus</i> , <i>S. suis</i>	Wu et al. (2013)
Magnetite nanoparticles	<i>C. albicans</i>	Anghel et al. (2012)
<i>Eugenia carryophyllata</i> essential oil stabilized by iron oxide/oleic acid core/shell nanostructures	<i>S. aureus</i>	Grumezescu et al. (2011, 2012)
Zinc and copper oxide nanoparticles	<i>S. mutans</i>	Eshed et al. (2012)
Zerovalent bismuth nanoparticle	<i>S. mutans</i>	Hernandez- Delgadillo et al. (2012)
Silver nanoparticles in combination with ampicillin, chloramphenicol, and kanamycin	<i>Enterococcus faecium</i> , <i>S. aureus</i> , <i>E. coli</i> , <i>P. aeruginosa</i>	Hwang et al. (2012)
Dextran sulfate nanoparticle complex containing ofloxacin and levofloxacin	<i>P. aeruginosa</i>	Cheow and Hadinoto (2012)
PEG-stabilized lipid nanoparticles loaded with terpinen-4-ol	<i>C. albicans</i>	Sun et al. (2012)
Magnesium fluoride nanoparticles	<i>S. aureus</i> , <i>E. coli</i>	Lellouche et al. (2009, 2012b, c)
Yttrium fluoride nanoparticles	<i>S. aureus</i> , <i>E. coli</i>	Lellouche et al. (2012a)
Iron oxide/oleic acid in combination with essential oil from <i>Rosmarinus officinalis</i>	<i>C. albicans</i> , <i>C. tropicalis</i>	Chifiriuc et al. (2012)
Gold nanoparticles and methylene blue	<i>C. albicans</i>	Khan et al. (2012)
Starch-stabilized silver nanoparticles	<i>S. aureus</i> , <i>P. aeruginosa</i>	Mohanty et al. (2012)

(continued)

Table 1 (continued)

Nanoparticle	Target organism	References
Iron oxide–oleic acid nanofluid	<i>S. aureus</i>	Grumezescu et al. (2011)
Chitosan, zinc oxide, nitric oxide nanoparticles	<i>E. faecalis</i>	Shrestha et al. (2010)
Quaternary ammonium polyethylenimine nanoparticles	Oral biofilms	Beyth et al. (2010)
Zinc oxide nanoparticles, chitosan nanoparticles, and combination of both	<i>E. faecalis</i>	Kishen et al. (2008)
Polyurethane nanocomposite	<i>S. epidermidis</i>	Styan et al. (2007)

**Fig. 1** Molecular mechanisms involved in nanoparticle-induced cellular toxicity (adopted and modified from Buzea et al. 2007)

Exposure to nanoparticles like zirconium dioxide (ZrO_2) induces overexpression of viral receptors and in turn results in hyper-reaction of the immune system and subsequent unwarranted inflammation (Lucarelli et al. 2004). In vivo and in vitro studies have shown the ability of nanoparticles (fullerenes, carbon nanotubes, quantum dots, and automobile exhaust) to initiate the production of reactive oxygen species (ROS) (Oberdorster et al. 2005), which has been shown to play a key role in cell damage by peroxidizing lipids, damaging proteins and nucleic acids, interfering with signaling functions, and modulating gene expression (Brown et al. 2004; Risom et al. 2005; Peters et al. 2006; Mehta et al. 2008). Malfunction of mitochondria has also been observed upon nanoparticle treatment as they effectively enter these organelles and contribute to oxidative stress and damage (Li et al. 2003; Xia et al. 2006; Sioutas et al. 2005). There is also evidence of the adverse effects

(appearance of thrombi) of nanoparticles on the cardiovascular system (Schulz et al. 2005; Nemmar et al. 2002; Vermylen et al. 2005; Hoet et al. 2004). Uptake of nanoparticles through skin results in their accumulation in the lymphatic system causing pododermatitis (Corachan 1988; Blundell et al. 1989) and Kaposi's sarcoma (Montella et al. 1997; Mott et al. 2002). Molecular mechanisms involved in nanoparticle-mediated cellular toxicity are schematically represented (Fig. 1).

6 Conclusions

Nanotechnology is a nascent field of science with promising potential in many fields including physics, chemistry, biology, pharmacology, and medicine. As discussed in this chapter, nanoparticles can be our friend or foe. Although there are reports which state nanoparticles are toxic, there is always the potential for improvement and development of safe and effective novel nanoparticles or nanocomposites. The utility of nanoparticles as drug carriers appears to be an important tool for targeted tumor therapy, and enhancing the efficacy of drugs could be another attractive application for nanomaterials. Even though the use of nanoparticles in vivo is debatable for now, their use on inanimate objects is effective. Without any doubt, the future will witness increasing use of nanoparticles in many fields, hopefully for the improvement of mankind.

References

- Agnihotri M, Joshi S, Kumar AR, Zinjarde S, Kulkarni S (2009) Biosynthesis of gold nanoparticles by the tropical marine yeast *Yarrowia lipolytica* NCIM 3589. *Mater Lett* 63 (15):1231–1234
- Amer LS, Bishop BM, van Hoek ML (2010) Antimicrobial and antibiofilm activity of cathelicidins and short, synthetic peptides against *Francisella*. *Biochem Biophys Res Commun* 396(2):246–251
- Amoozgar Z, Park J, Lin Q, Weidle JH 3rd, Yeo Y (2013) Development of quinic acid-conjugated nanoparticles as a drug carrier to solid tumors. *Biomacromolecules* 14(7):2389–2395
- Anghel I, Grumezescu AM (2013) Hybrid nanostructured coating for increased resistance of prosthetic devices to staphylococcal colonization. *Nanoscale Res Lett* 8(1):6
- Anghel I, Grumezescu AM, Andronesu E, Anghel AG, Ficai A, Saviuc C, Grumezescu V, Vasile BS, Chifiriuc MC (2012) Magnetite nanoparticles for functionalized textile dressing to prevent fungal biofilms development. *Nanoscale Res Lett* 7(1):501
- Annamalai A, Christina VL, Sudha D, Kalpana M, Lakshmi PT (2013) Green synthesis, characterization and antimicrobial activity of Au NPs using *Euphorbia hirta* L. leaf extract. *Colloids Surf B Biointerfaces* 108:60–65
- Antony JJ, Nivedheetha M, Siva D, Pradeepha G, Kokilavani P, Kalaiselvi S, Sankarganesh A, Balasundaram A, Masilamani V, Achiraman S (2013) Antimicrobial activity of *Leucas aspera* engineered silver nanoparticles against *Aeromonas hydrophila* in infected *Catla catla*. *Colloids Surf B Biointerfaces* 109:20–24

- Apte M, Girme G, Bankar A, Ravikumar A, Zinjarde S (2013a) 3, 4-dihydroxy-L-phenylalanine-derived melanin from *Yarrowia lipolytica* mediates the synthesis of silver and gold nanostructures. *J Nanobiotechnol* 11:2
- Apte M, Sambre D, Gaikawad S, Joshi S, Bankar A, Kumar AR, Zinjarde S (2013b) Psychrotrophic yeast *Yarrowia lipolytica* NCYC 789 mediates the synthesis of antimicrobial silver nanoparticles via cell-associated melanin. *AMB Express* 3(1):32
- Arai M, Niikawa H, Kobayashi M (2013) Marine-derived fungal sesterterpenes, ophiobolins, inhibit biofilm formation of *Mycobacterium* species. *J Nat Med* 67(2):271–275
- Bakkiyaraj D, Pandian SK (2010) *In vitro* and *in vivo* antibiofilm activity of a coral associated actinomycete against drug resistant *Staphylococcus aureus* biofilms. *Biofouling* 26(6):711–717
- Bakkiyaraj D, Sivasankar C, Pandian SK (2012) Inhibition of quorum sensing regulated biofilm formation in *Serratia marcescens* causing nosocomial infections. *Bioorg Med Chem Lett* 22(9):3089–3094
- Balaji K, Thenmozhi R, Pandian SK (2013) Effect of subinhibitory concentrations of fluoroquinolones on biofilm production by clinical isolates of *Streptococcus pyogenes*. *Indian J Med Res* 137(5):963–971
- Besinis A, De Peralta T, Handy RD (2014) The antibacterial effects of silver, titanium dioxide and silica dioxide nanoparticles compared to the dental disinfectant chlorhexidine on *Streptococcus mutans* using a suite of bioassays. *Nanotoxicology* 8(1):1–16
- Beyth N, Yudovin-Farber I, Perez-Davidi M, Domb AJ, Weiss EI (2010) Polyethyleneimine nanoparticles incorporated into resin composite cause cell death and trigger biofilm stress *in vivo*. *Proc Natl Acad Sci USA* 107(51):22038–22043
- Bink A, Kucharikova S, Neirinck B, Vleugels J, Van Dijck P, Cammue BP, Thevissen K (2012) The nonsteroidal antiinflammatory drug diclofenac potentiates the *in vivo* activity of caspofungin against *Candida albicans* biofilms. *J Infect Dis* 206(11):1790–1797
- Blundell G, Henderson WJ, Price EW (1989) Soil particles in the tissues of the foot in endemic elephantiasis of the lower legs. *Ann Trop Med Parasitol* 83(4):381–385
- Borges A, Saavedra MJ, Simoes M (2012) The activity of ferulic and gallic acids in biofilm prevention and control of pathogenic bacteria. *Biofouling* 28(7):755–767
- Borm PJ, Schins RP, Albrecht C (2004) Inhaled particles and lung cancer, part B: paradigms and risk assessment. *Int J Cancer* 110(1):3–14
- Bosi S, Da Ros T, Spalluto G, Prato M (2003) Fullerene derivatives: an attractive tool for biological applications. *Eur J Med Chem* 38(11–12):913–923
- Brackman G, Defoirdt T, Miyamoto C, Bossier P, Van Calenbergh S, Nelis H, Coenye T (2008) Cinnamaldehyde and cinnamaldehyde derivatives reduce virulence in *Vibrio* spp. by decreasing the DNA-binding activity of the quorum sensing response regulator LuxR. *BMC Microbiol* 8:149
- Brayner R, Ferrari-Iliou R, Brivois N, Djediat S, Benedetti MF, Fievet F (2006) Toxicological impact studies based on *Escherichia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium. *Nano Lett* 6(4):866–870
- Brown DM, Donaldson K, Borm PJ, Schins RP, Dehnhardt M, Gilmour P, Jimenez LA, Stone V (2004) Calcium and ROS-mediated activation of transcription factors and TNF-alpha cytokine gene expression in macrophages exposed to ultrafine particles. *Am J Physiol Lung Cell Mol Physiol* 286(2):L344–L353
- Buzeza C, Blandino IIP, Robbie K (2007) Nanomaterials and nanoparticles: sources and toxicity. *Biointerphases* 2(4):MR17–MR172
- Capek I (2004) Preparation of metal nanoparticles in water-in-oil (w/o) microemulsions. *Adv Colloid Interface Sci* 110(1–2):49–74
- Cardoso Sa N, Cavalcante TT, Araujo AX, dos Santos HS, Albuquerque MR, Bandeira PN, da Cunha RM, Cavada BS, Teixeira EH (2012) Antimicrobial and antibiofilm action of Casbane Diterpene from *Croton nepetaefolius* against oral bacteria. *Arch Oral Biol* 57(5):550–555

- Carey JD (2003) Engineering the next generation of large-area displays: prospects and pitfalls. *Philos Trans A Math Phys Eng Sci* 361(1813):2891–2907
- Castro-Longoria E, Vilchis-Nestor AR, Avalos-Borja M (2011) Biosynthesis of silver, gold and bimetallic nanoparticles using the filamentous fungus *Neurospora crassa*. *Colloids Surf B Biointerfaces* 83(1):42–48
- Cheow WS, Hadinoto K (2012) Green preparation of antibiotic nanoparticle complex as potential anti-biofilm therapeutics via self-assembly amphiphile-polyelectrolyte complexation with dextran sulfate. *Colloids Surf B Biointerfaces* 92:55–63
- Chemousova S, Epple M (2013) Silver as antibacterial agent: ion, nanoparticle, and metal. *Angew Chem Int Ed Engl* 52(6):1636–1653
- Chifiriuc C, Grumezescu V, Grumezescu AM, Saviuc C, Lazar V, Andronescu E (2012) Hybrid magnetite nanoparticles/*Rosmarinus officinalis* essential oil nanobiosystem with antibiofilm activity. *Nanoscale Res Lett* 7:209
- Choi H, Lee DG (2012) Antimicrobial peptide pleurocidin synergizes with antibiotics through hydroxyl radical formation and membrane damage, and exerts antibiofilm activity. *Biochim Biophys Acta* 1820(12):1831–1838
- Corachan M (1988) Endemic non-filarial elephantiasis of the lower limbs: podoconiosis. *Med Clin (Barc)* 91(3):97–100
- Costa C, Conte A, Buonocore GG, Del Nobile MA (2011) Antimicrobial silver-montmorillonite nanoparticles to prolong the shelf life of fresh fruit salad. *Int J Food Microbiol* 148(3):164–167
- Costa PM, Cardoso AL, Mendonca LS, Serani A, Custodia C, Conceicao M, Simoes S, Moreira JN, Pereira de Almeida L, Pedroso de Lima MC (2013) Tumor-targeted chlorotoxin-coupled nanoparticles for nucleic acid delivery to glioblastoma cells: a promising system for glioblastoma treatment. *Mol Ther Nucleic Acids* 2:e100
- Cristina B, Kevin R (2005) Assembling the puzzle of superconducting elements: a review. *Supercond Sci Technol* 18(1):R1
- Das B, Mandal M, Upadhyay A, Chattopadhyay P, Karak N (2013) Bio-based hyperbranched polyurethane/Fe₃O₄ nanocomposites: smart antibacterial biomaterials for biomedical devices and implants. *Biomed Mater* 8(3):035003
- Dasenbrock C, Peters L, Creutzenberg O, Heinrich U (1996) The carcinogenic potency of carbon particles with and without PAH after repeated intratracheal administration in the rat. *Toxicol Lett* 88(1–3):15–21
- Deng D, Zhang D, Li Y, Achilefu S, Gu Y (2013) Gold nanoparticles based molecular beacons for *in vitro* and in vivo detection of the matriptase expression on tumor. *Biosens Bioelectron* 49C:216–221
- Driscoll KE, Carter JM, Howard BW, Hassenbein DG, Pepelko W, Baggs RB, Oberdorster G (1996) Pulmonary inflammatory, chemokine, and mutagenic responses in rats after subchronic inhalation of carbon black. *Toxicol Appl Pharmacol* 136(2):372–380
- Du WL, Xu YL, Xu ZR, Fan CL (2008) Preparation, characterization and antibacterial properties against *E. coli* K(88) of chitosan nanoparticle loaded copper ions. *Nanotechnology* 19(8):085707
- Du L, Xian L, Feng J-X (2010) Rapid extra-/intracellular biosynthesis of gold nanoparticles by the fungus *Penicillium* sp. *J Nanopart Res* 13(3):921–930
- Durmuss NG, Webster TJ (2013) Eradicating antibiotic-resistant biofilms with silver-conjugated superparamagnetic iron oxide nanoparticles. *Adv Healthc Mater* 2(1):165–171
- Dutta RK, Nenavathu BP, Gangishetty MK, Reddy AV (2013) Antibacterial effect of chronic exposure of low concentration ZnO nanoparticles on *E. coli*. *J Environ Sci Health A Tox Hazard Subst Environ Eng* 48(8):871–878
- Elechiguerra JL, Burt JL, Morones JR, Camacho-Bragado A, Gao X, Lara HH, Yacaman MJ (2005) Interaction of silver nanoparticles with HIV-1. *J Nanobiotechnol* 3:6
- Ellis-Behnke RG, Liang YX, You SW, Tay DK, Zhang S, So KF, Schneider GE (2006) Nano neuro knitting: peptide nanofiber scaffold for brain repair and axon regeneration with functional return of vision. *Proc Natl Acad Sci USA* 103(13):5054–5059

- Eshed M, Lellouche J, Matalon S, Gedanken A, Banin E (2012) Sonochemical coatings of ZnO and CuO nanoparticles inhibit *Streptococcus mutans* biofilm formation on teeth model. *Langmuir* 28(33):12288–12295
- Evliyaoglu Y, Kobaner M, Celebi H, Yelsel K, Dogan A (2011) The efficacy of a novel antibacterial hydroxyapatite nanoparticle-coated indwelling urinary catheter in preventing biofilm formation and catheter-associated urinary tract infection in rabbits. *Urol Res* 39(6):443–449
- Ferin J (1994) Pulmonary retention and clearance of particles. *Toxicol Lett* 72(1–3):121–125
- Furlani RE, Yeagley AA, Melander C (2012) A flexible approach to 1,4-di-substituted 2-aminoimidazoles that inhibit and disperse biofilms and potentiate the effects of beta-lactams against multi-drug resistant bacteria. *Eur J Med Chem* 62C:59–70
- Gade AK, Bonde P, Ingle AP, Marcato PD, Duran N, Rai MK (2008) Exploitation of *Aspergillus niger* for synthesis of silver nanoparticles. *J Biobased Mater Bioenergy* 2:243–247
- Geethalakshmi R, Sarada DV (2012) Gold and silver nanoparticles from *Trianthema decandra*: synthesis, characterization, and antimicrobial properties. *Int J Nanomedicine* 7:5375–5384
- Gilbert P, Allison DG, McBain AJ (2002) Biofilms in vitro and in vivo: do singular mechanisms imply cross-resistance? *J Appl Microbiol* 92(Suppl):98S–110S
- Grumezescu AM, Saviuc C, Chifiriuc MC, Hristu R, Mihaiescu DE, Balaure P, Stanciu G, Lazar V (2011) Inhibitory activity of Fe(3) O(4)/oleic acid/usnic acid-core/shell/extra-shell nanofluid on *S. aureus* biofilm development. *IEEE Trans Nanobiosci* 10(4):269–274
- Grumezescu AM, Chifiriuc MC, Saviuc C, Grumezescu V, Hristu R, Mihaiescu DE, Stanciu GA, Andronesu E (2012) Hybrid nanomaterial for stabilizing the antibiofilm activity of *Eugenia caryophyllata* essential oil. *IEEE Trans Nanobiosci* 11(4):360–365
- Hafner JH, Cheung CL, Woolley AT, Lieber CM (2001) Structural and functional imaging with carbon nanotube AFM probes. *Prog Biophys Mol Biol* 77(1):73–110
- Hamouda IM (2012) Current perspectives of nanoparticles in medical and dental biomaterials. *J Biomed Res* 26(3):143–151
- Harisinghani MG, Barentsz J, Hahn PF, Deserno WM, Tabatabaei S, van de Kaa CH, de la Rosette J, Weissleder R (2003) Noninvasive detection of clinically occult lymph-node metastases in prostate cancer. *N Engl J Med* 348(25):2491–2499
- He F, Zhao D (2005) Preparation and characterization of a new class of starch-stabilized bimetallic nanoparticles for degradation of chlorinated hydrocarbons in water. *Environ Sci Technol* 39(9):3314–3320
- Hernandez-Delgado R, Velasco-Arias D, Diaz D, Arevalo-Nino K, Garza-Enriquez M, De la Garza-Ramos MA, Cabral-Romero C (2012) Zerovalent bismuth nanoparticles inhibit *Streptococcus mutans* growth and formation of biofilm. *Int J Nanomedicine* 7:2109–2113
- Hernandez-Delgado R, Velasco-Arias D, Martinez-Sanmiguel JJ, Diaz D, Zumeta-Dube I, Arevalo-Nino K, Cabral-Romero C (2013) Bismuth oxide aqueous colloidal nanoparticles inhibit *Candida albicans* growth and biofilm formation. *Int J Nanomedicine* 8:1645–1652
- Hoet PH, Bruske-Hohlfeld I, Salata OV (2004) Nanoparticles: known and unknown health risks. *J Nanobiotechnol* 2(1):12
- Hrubesh LW, Poco JF (1995) Thin aerogel films for optical, thermal, acoustic and electronic applications. *J Non Cryst Solids* 188(1–2):46–53
- Huda S, Smoukov SK, Nakanishi H, Kowalczyk B, Bishop K, Grzybowski BA (2010) Antibacterial nanoparticle monolayers prepared on chemically inert surfaces by cooperative electrostatic adsorption (CELA). *ACS Appl Mater Interfaces* 2(4):1206–1210
- Hwang IS, Hwang JH, Choi H, Kim KJ, Lee DG (2012) Synergistic effects between silver nanoparticles and antibiotics and the mechanisms involved. *J Med Microbiol* 61(Pt 12):1719–1726
- Hwang YY, Ramalingam K, Bienek DR, Lee V, You T, Alvarez R (2013) Antimicrobial activity of nanoemulsion in combination with cetylpyridinium chloride on multidrug-resistant *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 57(8):3568–3575

- Imperi F, Massai F, Ramachandran Pillai C, Longo F, Zennaro E, Rampioni G, Visca P, Leoni L (2013) New life for an old drug: the anthelmintic drug niclosamide inhibits *Pseudomonas aeruginosa* quorum sensing. *Antimicrob Agents Chemother* 57(2):996–1005
- Iskandar F (2009) Nanoparticle processing for optical applications: a review. *Adv Powder Technol* 20(4):283–292
- Jacoby M (2002) Nanoscale electronics. *Chem Eng News Arch* 80(39):38–43
- Jain P, Pradeep T (2005) Potential of silver nanoparticle-coated polyurethane foam as an antibacterial water filter. *Biotechnol Bioeng* 90(1):59–63
- Jayaseelan C, Rahuman AA, Roopan SM, Kirthi AV, Venkatesan J, Kim SK, Iyappan M, Siva C (2013) Biological approach to synthesize TiO₂ nanoparticles using *Aeromonas hydrophila* and its antibacterial activity. *Spectrochim Acta A Mol Biomol Spectrosc* 107:82–89
- Jirage KB, Hulteen JC, Martin CR (1997) Nanotubule-based molecular-filtration membranes. *Science* 278(5338):655–658
- Jones N, Ray B, Ranjit KT, Manna AC (2008) Antibacterial activity of ZnO nanoparticle suspensions on a broad spectrum of microorganisms. *FEMS Microbiol Lett* 279(1):71–76
- Kalishwaralal K, BarathManiKanth S, Pandian SR, Deepak V, Gurunathan S (2010) Silver nanoparticles impede the biofilm formation by *Pseudomonas aeruginosa* and *Staphylococcus epidermidis*. *Colloids Surf B Biointerfaces* 79(2):340–344
- Kalpna BJ, Aarthi S, Pandian SK (2012) Antibiofilm activity of alpha-amylase from *Bacillus subtilis* S8-18 against biofilm forming human bacterial pathogens. *Appl Biochem Biotechnol* 167(6):1778–1794
- Kamaly N, Xiao Z, Valencia PM, Radovic-Moreno AF, Farokhzad OC (2012) Targeted polymeric therapeutic nanoparticles: design, development and clinical translation. *Chem Soc Rev* 41(7):2971–3010
- Karunanidhi A, Thomas R, van Belkum A, Neela V (2013) *In vitro* antibacterial and antibiofilm activities of chlorogenic acid against clinical isolates of *Stenotrophomonas maltophilia* including the trimethoprim/sulfamethoxazole resistant strain. *Biomed Res Int* 2013:392058
- Khan S, Alam F, Azam A, Khan AU (2012) Gold nanoparticles enhance methylene blue-induced photodynamic therapy: a novel therapeutic approach to inhibit *Candida albicans* biofilm. *Int J Nanomedicine* 7:3245–3257
- Kim YH, Lee DK, Cha HG, Kim CW, Kang YC, Kang YS (2006) Preparation and characterization of the antibacterial Cu nanoparticle formed on the surface of SiO₂ nanoparticles. *J Phys Chem B* 110(49):24923–24928
- Kiran GS, Sabarathnam B, Selvin J (2010) Biofilm disruption potential of a glycolipid biosurfactant from marine *Brevibacterium casei*. *FEMS Immunol Med Microbiol* 59(3):432–438
- Kishen A, Shi Z, Shrestha A, Neoh KG (2008) An investigation on the antibacterial and antibiofilm efficacy of cationic nanoparticulates for root canal disinfection. *J Endod* 34(12):1515–1520
- Kneuer C, Sameti M, Bakowsky U, Schiestel T, Schirra H, Schmidt H, Lehr CM (2000) A nonviral DNA delivery system based on surface modified silica-nanoparticles can efficiently transfect cells in vitro. *Bioconj Chem* 11(6):926–932
- Koper OB, Klabunde JS, Marchin GL, Klabunde KJ, Stoimenov P, Bohra L (2002) Nanoscale powders and formulations with biocidal activity toward spores and vegetative cells of bacillus species, viruses, and toxins. *Curr Microbiol* 44(1):49–55
- Kumar CG, Mamidyalu SK (2011) Extracellular synthesis of silver nanoparticles using culture supernatant of *Pseudomonas aeruginosa*. *Colloids Surf B Biointerfaces* 84(2):462–466
- Kuzma L, Wysokinska H, Rozalski M, Budzynska A, Wieckowska-Szakiel M, Sadowska B, Paszkiewicz M, Kisiel W, Rozalska B (2012) Antimicrobial and anti-biofilm properties of new taxodione derivative from hairy roots of *Salvia austriaca*. *Phytomedicine* 19(14):1285–1287
- Lamppa JW, Griswold KE (2013) Alginate lyase exhibits catalysis-independent biofilm dispersion and antibiotic synergy. *Antimicrob Agents Chemother* 57(1):137–145

- Lara HH, Garza-Trevino EN, Ixtepan-Turrent L, Singh DK (2010) Silver nanoparticles are broad-spectrum bactericidal and virucidal compounds. *J Nanobiotechnol* 9:30
- Latimer J, Forbes S, McBain AJ (2012) Attenuated virulence and biofilm formation in *Staphylococcus aureus* following sublethal exposure to triclosan. *Antimicrob Agents Chemother* 56(6):3092–3100
- Leifert A, Pan-Bartnek Y, Simon U, Jahnen-Dechent W (2013) Molecularly stabilised ultrasmall gold nanoparticles: synthesis, characterization and bioactivity. *Nanoscale* 5(14):6224–6242
- Lellouche J, Kahana E, Elias S, Gedanken A, Banin E (2009) Antibiofilm activity of nanosized magnesium fluoride. *Biomaterials* 30(30):5969–5978
- Lellouche J, Friedman A, Gedanken A, Banin E (2012a) Antibacterial and antibiofilm properties of yttrium fluoride nanoparticles. *Int J Nanomedicine* 7:5611–5624
- Lellouche J, Friedman A, Lahmi R, Gedanken A, Banin E (2012b) Antibiofilm surface functionalization of catheters by magnesium fluoride nanoparticles. *Int J Nanomedicine* 7:1175–1188
- Lellouche J, Friedman A, Lellouche JP, Gedanken A, Banin E (2012c) Improved antibacterial and antibiofilm activity of magnesium fluoride nanoparticles obtained by water-based ultrasound chemistry. *Nanomedicine* 8(5):702–711
- Leuba KD, Durmus NG, Taylor EN, Webster TJ (2013) Carboxylate functionalized superparamagnetic iron oxide nanoparticles (SPION) for the reduction of *S. aureus* growth post biofilm formation. *Int J Nanomedicine* 8:731–736
- Lewin M, Carlesso N, Tung CH, Tang XW, Cory D, Scadden DT, Weissleder R (2000) Tat peptide-derivatized magnetic nanoparticles allow *in vivo* tracking and recovery of progenitor cells. *Nat Biotechnol* 18(4):410–414
- Li LL, Wang H (2013) Enzyme-coated mesoporous silica nanoparticles as efficient antibacterial agents *in vivo*. *Adv Healthc Mater* 2(10):1351–1360
- Li N, Sioutas C, Cho A, Schmitz D, Misra C, Sempf J, Wang M, Oberley T, Froines J, Nel A (2003) Ultrafine particulate pollutants induce oxidative stress and mitochondrial damage. *Environ Health Perspect* 111(4):455–460
- Li Y, Leung P, Yao L, Song QW, Newton E (2006) Antimicrobial effect of surgical masks coated with nanoparticles. *J Hosp Infect* 62(1):58–63
- Li D, Cui F, Zhao Z, Liu D, Xu Y, Li H, Yang X (2013) The impact of titanium dioxide nanoparticles on biological nitrogen removal from wastewater and bacterial community shifts in activated sludge. *Biodegradation*. doi:10.1007/s10532-013-9648-z
- Liu HK, Wang GX, Guo Z, Wang J, Konstantinov K (2006a) Nanomaterials for lithium-ion rechargeable batteries. *J Nanosci Nanotechnol* 6(1):1–15
- Liu J, Wong HL, Moselhy J, Bowen B, Wu XY, Johnston MR (2006b) Targeting colloidal particulates to thoracic lymph nodes. *Lung Cancer* 51(3):377–386
- Liu G, Mao J, Jiang Z, Sun T, Hu Y, Zhang C, Dong J, Huang Q, Lan Q (2013a) Transferrin-modified doxorubicin-loaded biodegradable nanoparticles exhibit enhanced efficacy in treating brain glioma-bearing rats. *Cancer Biother Radiopharm* 28(9):691–696
- Liu Y, Sun Y, Xu Y, Feng H, Fu S, Tang J, Liu W, Sun D, Jiang H, Xu S (2013b) Preparation and evaluation of lysozyme-loaded nanoparticles coated with poly-gamma-glutamic acid and chitosan. *Int J Biol Macromol* 59:201–207
- Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, Tam PK, Chiu JF, Che CM (2006) Proteomic analysis of the mode of antibacterial action of silver nanoparticles. *J Proteome Res* 5(4):916–924
- Lucarelli M, Gatti AM, Savarino G, Quattroni P, Martinelli L, Monari E, Boraschi D (2004) Innate defence functions of macrophages can be biased by nano-sized ceramic and metallic particles. *Eur Cytokine Netw* 15(4):339–346
- Mallick S, Sharma S, Banerjee M, Ghosh SS, Chattopadhyay A, Paul A (2012) Iodine-stabilized Cu nanoparticle chitosan composite for antibacterial applications. *ACS Appl Mater Interfaces* 4(3):1313–1323
- Martin CR, Kohli P (2003) The emerging field of nanotube biotechnology. *Nat Rev Drug Discov* 2(1):29–37

- Martinez-Gutierrez F, Boegli L, Agostinho A, Sanchez EM, Bach H, Ruiz F, James G (2013) Antibiofilm activity of silver nanoparticles against different microorganisms. *Biofouling* 29 (6):651–660
- Matsusaki M, Larsson K, Akagi T, Lindstedt M, Akashi M, Borrebaeck CA (2005) Nanosphere induced gene expression in human dendritic cells. *Nano Lett* 5(11):2168–2173
- Mehta M, Chen LC, Gordon T, Rom W, Tang MS (2008) Particulate matter inhibits DNA repair and enhances mutagenesis. *Mutat Res* 657(2):116–121
- Mochizuki D, Tamura S, Yasutake H, Kataoka T, Mitsuo K, Wada Y (2013) A photostable bi-luminophore pressure-sensitive paint measurement system developed with mesoporous silica nanoparticles. *J Nanosci Nanotechnol* 13(4):2777–2781
- Mohanty S, Mishra S, Jena P, Jacob B, Sarkar B, Sonawane A (2012) An investigation on the antibacterial, cytotoxic, and antibiofilm efficacy of starch-stabilized silver nanoparticles. *Nanomedicine* 8(6):916–924
- Monteiro DR, Silva S, Negri M, Gorup LF, de Camargo ER, Oliveira R, Barbosa DB, Henriques M (2012) Silver nanoparticles: influence of stabilizing agent and diameter on antifungal activity against *Candida albicans* and *Candida glabrata* biofilms. *Lett Appl Microbiol* 54(5):383–391
- Monteiro DR, Silva S, Negri M, Gorup LF, de Camargo ER, Oliveira R, Barbosa DB, Henriques M (2013) Antifungal activity of silver nanoparticles in combination with nystatin and chlorhexidine digluconate against *Candida albicans* and *Candida glabrata* biofilms. *Mycoses* 56 (6):672–680
- Montella M, Franceschi S, Geddes da Filicaia M, De Macro M, Arniani S, Balzi D, Delfino M, Iannuzzo M, Buonanno M, Satriano RA (1997) Classical Kaposi sarcoma and volcanic soil in southern Italy: a case-control study. *Epidemiol Prev* 21(2):114–117
- Mott JA, Meyer P, Mannino D, Redd SC, Smith EM, Gotway-Crawford C, Chase E (2002) Wildland forest fire smoke: health effects and intervention evaluation, Hoopa, California, 1999. *West J Med* 176(3):157–162
- Mukhopadhyay A, Basak S, Das JK, Medda SK, Chattopadhyay K, De G (2010) Ag-TiO₂ nanoparticle codoped SiO₂ films on ZrO₂ barrier-coated glass substrates with antibacterial activity in ambient condition. *ACS Appl Mater Interfaces* 2(9):2540–2546
- Murray CB, Kagan CR, Bawendi MG (2000) Synthesis and characterization of monodisperse nanocrystals and close-packed nanocrystal assemblies. *Annu Rev Mater Sci* 30(1):545–610
- Musthafa KS, Balamurugan K, Pandian SK, Ravi AV (2012a) 2,5-Piperazinedione inhibits quorum sensing-dependent factor production in *Pseudomonas aeruginosa* PAO1. *J Basic Microbiol* 52(6):679–686
- Musthafa KS, Sivamaruthi BS, Pandian SK, Ravi AV (2012b) Quorum sensing inhibition in *Pseudomonas aeruginosa* PAO1 by antagonistic compound phenylacetic acid. *Curr Microbiol* 65(5):475–480
- Naz SS, Islam NU, Shah MR, Alam SS, Iqbal Z, Bertino M, Franzel L, Ahmed A (2013) Enhanced biocidal activity of Au nanoparticles synthesized in one pot using 2, 4-dihydroxybenzene carbodithioic acid as a reducing and stabilizing agent. *J Nanobiotechnol* 11(1):13
- Nemmar A, Hoylaerts MF, Hoet PH, Dinsdale D, Smith T, Xu H, Vermynen J, Nemery B (2002) Ultrafine particles affect experimental thrombosis in an in vivo hamster model. *Am J Respir Crit Care Med* 166(7):998–1004
- Nikula KJ, Snipes MB, Barr EB, Griffith WC, Henderson RF, Mauderly JL (1995) Comparative pulmonary toxicities and carcinogenicities of chronically inhaled diesel exhaust and carbon black in F344 rats. *Fundam Appl Toxicol* 25(1):80–94
- Nithya C, Pandian SK (2010) The *in vitro* antibiofilm activity of selected marine bacterial culture supernatants against *Vibrio* spp. *Arch Microbiol* 192(10):843–854
- Nithya C, Aravindraja C, Pandian SK (2010a) *Bacillus pumilus* of Palk Bay origin inhibits quorum-sensing-mediated virulence factors in Gram-negative bacteria. *Res Microbiol* 161 (4):293–304

- Nithya C, Begum MF, Pandian SK (2010b) Marine bacterial isolates inhibit biofilm formation and disrupt mature biofilms of *Pseudomonas aeruginosa* PAO1. *Appl Microbiol Biotechnol* 88 (1):341–358
- Nithya C, Devi MG, Pandian SK (2011) A novel compound from the marine bacterium *Bacillus pumilus* S6-15 inhibits biofilm formation in gram-positive and gram-negative species. *Biofouling* 27(5):519–528
- Oberdorster G, Ferin J, Lehnert BE (1994) Correlation between particle size, *in vivo* particle persistence, and lung injury. *Environ Health Perspect* 102(Suppl 5):173–179
- Oberdorster G, Oberdorster E, Oberdorster J (2005) Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect* 113(7):823–839
- Packiavathy IASV, Agilandeswari P, Musthafa KS, Pandian SK, Ravi AV (2012) Antibiofilm and quorum sensing inhibitory potential of *Cuminum cyminum* and its secondary metabolite methyl eugenol against Gram negative bacterial pathogens. *Food Res Int* 45(1):85–92
- Packiavathy IA, Sasikumar P, Pandian SK, Ravi A (2013) Prevention of quorum-sensing-mediated biofilm development and virulence factors production in *Vibrio* spp. by curcumin. *Appl Microbiol Biotechnol* 97(23):10177–10187
- Pan B, Huang RZ, Han SQ, Qu D, Zhu ML, Wei P, Ying HJ (2010) Design, synthesis, and antibiofilm activity of 2-arylimino-3-aryl-thiazolidine-4-ones. *Bioorg Med Chem Lett* 20 (8):2461–2464
- Pandiyarajan T, Udayabhaskar R, Vignesh S, James RA, Karthikeyan B (2013) Synthesis and concentration dependent antibacterial activities of CuO nanoflakes. *Mater Sci Eng C Mater Biol Appl* 33(4):2020–2024
- Park JH, Lee JH, Cho MH, Herzberg M, Lee J (2012a) Acceleration of protease effect on *Staphylococcus aureus* biofilm dispersal. *FEMS Microbiol Lett* 335(1):31–38
- Park JH, Lee JH, Kim CJ, Lee JC, Cho MH, Lee J (2012b) Extracellular protease in Actinomycetes culture supernatants inhibits and detaches *Staphylococcus aureus* biofilm formation. *Biotechnol Lett* 34(4):655–661
- Patel MB, Harikrishnan U, Valand NN, Modi NR, Menon SK (2013) Novel cationic quinazolin-4 (3H)-one conjugated fullerene nanoparticles as antimycobacterial and antimicrobial agents. *Arch Pharm (Weinheim)* 346(3):210–220
- Pender DS, Vangala LM, Badwaik VD, Willis CB, Aguilar ZP, Sangoi TN, Paripelly R, Dakshinamurthy R (2013) Bactericidal activity of starch-encapsulated gold nanoparticles. *Front Biosci* 18:993–1002
- Peters A, Veronesi B, Calderon-Garciduenas L, Gehr P, Chen LC, Geiser M, Reed W, Rothen-Rutishauser B, Schurch S, Schulz H (2006) Translocation and potential neurological effects of fine and ultrafine particles a critical update. *Part Fibre Toxicol* 3:13
- Pinto RJ, Almeida A, Fernandes SC, Freire CS, Silvestre AJ, Neto CP, Trindade T (2013) Antifungal activity of transparent nanocomposite thin films of pullulan and silver against *Aspergillus niger*. *Colloids Surf B Biointerfaces* 103:143–148
- Pompilio A, Pomponio S, Di Vincenzo V, Crocetta V, Nicoletti M, Piovano M, Garbarino JA, Di Bonaventura G (2013) Antimicrobial and antibiofilm activity of secondary metabolites of lichens against methicillin-resistant *Staphylococcus aureus* strains from cystic fibrosis patients. *Future Microbiol* 8(2):281–292
- Pramanik A, Laha D, Bhattacharya D, Pramanik P, Karmakar P (2012) A novel study of antibacterial activity of copper iodide nanoparticle mediated by DNA and membrane damage. *Colloids Surf B Biointerfaces* 96:50–55
- Raghavendra GM, Jayaramudu T, Varaprasad K, Sadiku R, Ray SS, Mohana Raju K (2013) Cellulose-polymer-Ag nanocomposite fibers for antibacterial fabrics/skin scaffolds. *Carbohydr Polym* 93(2):553–560
- Raimondi MV, Maggio B, Raffa D, Plescia F, Cascioferro S, Cancemi G, Schillaci D, Cusimano MG, Vitale M, Daidone G (2012) Synthesis and anti-staphylococcal activity of new 4-diazopyrazole derivatives. *Eur J Med Chem* 58:64–71

- Ramage G, Saville SP, Wickes BL, Lopez-Ribot JL (2002) Inhibition of *Candida albicans* biofilm formation by farnesol, a quorum-sensing molecule. *Appl Environ Microbiol* 68(11):5459–5463
- Ramamurthy CH, Padma M, Samadanam ID, Mareeswaran R, Suyavaran A, Kumar MS, Premkumar K, Thirunavukkarasu C (2013) The extra cellular synthesis of gold and silver nanoparticles and their free radical scavenging and antibacterial properties. *Colloids Surf B Biointerfaces* 102:808–815
- Rane RA, Sahu NU, Shah CP (2012) Synthesis and antibiofilm activity of marine natural product-based 4-thiazolidinones derivatives. *Bioorg Med Chem Lett* 22(23):7131–7134
- Rane RA, Sahu NU, Shah CP, Shah NK (2013) Design, synthesis and anti-staphylococcal activity of marine pyrrole alkaloid derivatives. *J Enzyme Inhib Med Chem*. doi:10.3109/14756366.2013.793183
- Reymond JL, Bergmann M, Darbre T (2013) Glycopeptide dendrimers as *Pseudomonas aeruginosa* biofilm inhibitors. *Chem Soc Rev* 42:4814–4822
- Risom L, Moller P, Loft S (2005) Oxidative stress-induced DNA damage by particulate air pollution. *Mutat Res* 592(1–2):119–137
- Rodrigues AG, Ping LY, Marcato PD, Alves OL, Silva MC, Ruiz RC, Melo IS, Tasic L, De Souza AO (2013) Biogenic antimicrobial silver nanoparticles produced by fungi. *Appl Microbiol Biotechnol* 97(2):775–782
- Sankar R, Karthik A, Prabu A, Karthik S, Shivashangari KS, Ravikumar V (2013) *Origanum vulgare* mediated biosynthesis of silver nanoparticles for its antibacterial and anticancer activity. *Colloids Surf B Biointerfaces* 108:80–84
- Sanpui P, Murugadoss A, Prasad PV, Ghosh SS, Chattopadhyay A (2008) The antibacterial properties of a novel chitosan-Ag-nanoparticle composite. *Int J Food Microbiol* 124(2):142–146
- Sarabhai S, Sharma P, Capalash N (2013) Ellagic acid derivatives from *Terminalia chebula* Retz. Downregulate the expression of quorum sensing genes to attenuate *Pseudomonas aeruginosa* PAO1 virulence. *PLoS ONE* 8(1):e53441
- Sawant SN, Selvaraj V, Prabhawathi V, Doble M (2013) Antibiofilm properties of silver and gold incorporated PU, PCLm, PC and PMMA nanocomposites under two shear conditions. *PLoS ONE* 8(5):e63311
- Schubert D, Dargusch R, Raitano J, Chan SW (2006) Cerium and yttrium oxide nanoparticles are neuroprotective. *Biochem Biophys Res Commun* 342(1):86–91
- Schulz H, Harder V, Ibaldo-Mulli A, Khandoga A, Koenig W, Krombach F, Radykewicz R, Stampfl A, Thorand B, Peters A (2005) Cardiovascular effects of fine and ultrafine particles. *J Aerosol Med* 18(1):1–22
- Scott ID, Jung YS, Cavanagh AS, Yan Y, Dillon AC, George SM, Lee SH (2011) Ultrathin coatings on nano-LiCoO₂ for Li-ion vehicular applications. *Nano Lett* 11(2):414–418
- Semmler M, Seitz J, Erbe F, Mayer P, Heyder J, Oberdorster G, Kreyling WG (2004) Long-term clearance kinetics of inhaled ultrafine insoluble iridium particles from the rat lung, including transient translocation into secondary organs. *Inhal Toxicol* 16(6–7):453–459
- Shah V, Shah S, Shah H, Rispoli FJ, McDonnell KT, Workeneh S, Karakoti A, Kumar A, Seal S (2012) Antibacterial activity of polymer coated cerium oxide nanoparticles. *PLoS ONE* 7(10):e47827
- Shi P, Aluri S, Lin YA, Shah M, Edman M, Dhandhukia J, Cui H, Mackay JA (2013) Elastin-based protein polymer nanoparticles carrying drug at both corona and core suppress tumor growth in vivo. *J Control Release* 171(3):330–338
- Shimizu N, Otsuka K, Sawada H, Maejima T, Shirotake S (2013) Bacteriolysis by vancomycin-conjugated acryl nanoparticles and morphological component analysis. *Drug Dev Ind Pharm*. doi:10.3109/03639045.2013.788012
- Shivaji S, Madhu S, Singh S (2011) Extracellular synthesis of antibacterial silver nanoparticles using psychrophilic bacteria. *Process Biochem* 46(9):1800–1807
- Shrestha A, Shi Z, Neoh KG, Kishen A (2010) Nanoparticulates for antibiofilm treatment and effect of aging on its antibacterial activity. *J Endod* 36(6):1030–1035

- Singh V, Arora V, Alam MJ, Garey KW (2012) Inhibition of biofilm formation by esomeprazole in *Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Antimicrob Agents Chemother* 56 (8):4360–4364
- Singh S, Ashfaq M, Singh RK, Joshi HC, Srivastava A, Sharma A, Verma N (2013) Preparation of surfactant-mediated silver and copper nanoparticles dispersed in hierarchical carbon micro-nanofibers for antibacterial applications. *New Biotechnol* 30:656–665
- Sintubin L, De Windt W, Dick J, Mast J, van der Ha D, Verstraete W, Boon N (2009) Lactic acid bacteria as reducing and capping agent for the fast and efficient production of silver nanoparticles. *Appl Microbiol Biotechnol* 84(4):741–749
- Sioutas C, Delfino RJ, Singh M (2005) Exposure assessment for atmospheric ultrafine particles (UFPs) and implications in epidemiologic research. *Environ Health Perspect* 113(8):947–955
- Styan K, Abrahamian M, Hume E, Poole-Warren LA (2007) Antibacterial polyurethane organosilicate nanocomposites. *Key Eng Mat* 342:757–760
- Su R, Jin Y, Liu Y, Tong M, Kim H (2013) Bactericidal activity of Ag-doped multi-walled carbon nanotubes and the effects of extracellular polymeric substances and natural organic matter. *Colloids Surf B Biointerfaces* 104:133–139
- Suciu CV, Iwatsubo T, Deki S (2003) Investigation of a colloidal damper. *J Colloid Interface Sci* 259(1):62–80
- Sun LM, Zhang CL, Li P (2012) Characterization, antibiofilm, and mechanism of action of novel PEG-stabilized lipid nanoparticles loaded with terpinen-4-ol. *J Agric Food Chem* 60 (24):6150–6156
- Suresh AK, Pelletier DA, Wang W, Moon JW, Gu B, Mortensen NP, Allison DP, Joy DC, Phelps TJ, Doktycz MJ (2010) Silver nanocrystallites: biofabrication using *Shewanella oneidensis*, and an evaluation of their comparative toxicity on gram-negative and gram-positive bacteria. *Environ Sci Technol* 44(13):5210–5215
- Tavassoli Hojati S, Alaghemand H, Hamze F, Ahmadian Babaki F, Rajab-Nia R, Rezvani MB, Kaviani M, Atai M (2013) Antibacterial, physical and mechanical properties of flowable resin composites containing zinc oxide nanoparticles. *Dent Mater* 29(5):495–505
- Thekkae Padil VV, Cernik M (2013) Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application. *Int J Nanomedicine* 8:889–898
- Thompson S, Parthasarathy S (2006) Moore's law: the future of Si microelectronics. *Mater Today* 9(6):20–25
- Tsuji M, Hashimoto M, Nishizawa Y, Kubokawa M, Tsuji T (2005) Microwave-assisted synthesis of metallic nanostructures in solution. *Chem Eur J* 11(2):440–452
- Vermynen J, Nemmar A, Nemery B, Hoylaerts MF (2005) Ambient air pollution and acute myocardial infarction. *J Thromb Haemost* 3(9):1955–1961
- Vidic J, Stankic S, Haque F, Ciric D, Le Goffic R, Vidy A, Jupille J, Delmas B (2013) Selective antibacterial effects of mixed ZnMgO nanoparticles. *J Nanopart Res* 15(5):1595
- Vigneshwaran N, Kathe AA, Varadarajan PV, Nachane RP, Balasubramanya RH (2006) Biometics of silver nanoparticles by white rot fungus, *Phaenerochaete chrysosporium*. *Colloids Surf B Biointerfaces* 53(1):55–59
- Vivek R, Nipun Babu V, Thangam R, Subramanian KS, Kannan S (2013) pH-responsive drug delivery of chitosan nanoparticles as Tamoxifen carriers for effective anti-tumor activity in breast cancer cells. *Colloids Surf B Biointerfaces* 111C:117–123
- Wang X, Lim TT (2013) Highly efficient and stable Ag-AgBr/TiO₂ composites for destruction of *Escherichia coli* under visible light irradiation. *Water Res* 47(12):4148–4158
- Wang H, Liu J, Wu X, Tong Z, Deng Z (2013) Tailor-made Au@Ag core-shell nanoparticle 2D arrays on protein-coated graphene oxide with assembly enhanced antibacterial activity. *Nanotechnology* 24(20):205102
- Westendorf AM (2013) Applications of nanoparticles for treating cutaneous infection. *J Invest Dermatol* 133(5):1133–1135
- Wu C, Labrie J, Tremblay YD, Haine D, Mourez M, Jacques M (2013) Zinc as an agent for the prevention of biofilm formation by pathogenic bacteria. *J Appl Microbiol* 115(1):30–40

- Xia T, Kovochich M, Brant J, Hotze M, Sempf J, Oberley T, Sioutas C, Yeh JI, Wiesner MR, Nel AE (2006) Comparison of the abilities of ambient and manufactured nanoparticles to induce cellular toxicity according to an oxidative stress paradigm. *Nano Lett* 6(8):1794–1807
- Xiong R, Lu C, Zhang W, Zhou Z, Zhang X (2013) Facile synthesis of tunable silver nanostructures for antibacterial application using cellulose nanocrystals. *Carbohydr Polym* 95(1):214–219
- Yang X, Konishi H, Xu H, Wu M (2006) Comparative sol–hydro(Solvo)thermal synthesis of TiO₂ nanocrystals. *Eur J Inorg Chem* 2006(11):2229–2235
- Yin H, Zhang H, Liu B (2013) Superior anticancer efficacy of curcumin-loaded nanoparticles against lung cancer. *Acta Biochim Biophys Sin (Shanghai)* 45(8):634–640
- Yu JC, Wang X, Fu X (2004) Pore-wall chemistry and photocatalytic activity of mesoporous titania molecular sieve films. *Chem Mater* 16(8):1523–1530
- Zhang L, Yu JC (2003) A sonochemical approach to hierarchical porous titania spheres with enhanced photocatalytic activity. *Chem Commun* 9(16):2078–2079
- Zhang R, Zhou M, Wang L, McGrath S, Chen T, Chen X, Shaw C (2010) Phylloseptin-1 (PSN-1) from *Phyllomedusa sauvagei* skin secretion: a novel broad-spectrum antimicrobial peptide with antibiofilm activity. *Mol Immunol* 47(11–12):2030–2037
- Zhang H, Wang C, Chen B, Wang X (2012) Daunorubicin-TiO₂ nanocomposites as a “smart” pH-responsive drug delivery system. *Int J Nanomedicine* 7:235–242
- Zhang K, Cheng L, Imazato S, Antonucci JM, Lin NJ, Lin-Gibson S, Bai Y, Xu HH (2013a) Effects of dual antibacterial agents MDPB and nano-silver in primer on microcosm biofilm, cytotoxicity and dentine bond properties. *J Dent* 41(5):464–474
- Zhang K, Li F, Imazato S, Cheng L, Liu H, Arola DD, Bai Y, Xu HH (2013b) Dual antibacterial agents of nano-silver and 12-methacryloyloxydodecylpyridinium bromide in dental adhesive to inhibit caries. *J Biomed Mater Res B Appl Biomater* 101(6):929–938
- Zhang X, Li Z, Yuan X, Cui Z, Bao H, Li X, Liu Y, Yang X (2013c) Cytotoxicity and antibacterial property of titanium alloy coated with silver nanoparticle-containing polyelectrolyte multilayer. *Mater Sci Eng C Mater Biol Appl* 33(5):2816–2820
- Zhao J, Wang Z, Dai Y, Xing B (2013a) Mitigation of CuO nanoparticle-induced bacterial membrane damage by dissolved organic matter. *Water Res* 47(12):4169–4178
- Zhao L, Zhu B, Jia Y, Hou W, Su C (2013b) Preparation of biocompatible carboxymethyl chitosan nanoparticles for delivery of antibiotic drug. *Biomed Res Int* 2013:236469
- Zheng F, Wang S, Wen S, Shen M, Zhu M, Shi X (2013) Characterization and antibacterial activity of amoxicillin-loaded electrospun nano-hydroxyapatite/poly(lactic-co-glycolic acid) composite nanofibers. *Biomaterials* 34(4):1402–1412