

# Chapter 11

## Technical Rebuilding of Movement Function Using Functional Electrical Stimulation

Margit Gföhler

**Abstract** To rebuild lost movement functions, neuroprostheses based on functional electrical stimulation (FES) artificially activate skeletal muscles in corresponding sequences, using both residual body functions and artificial signals for control. Besides the functional gain, FES training also brings physiological and psychological benefits for spinal cord-injured subjects. In this chapter, current stimulation technology and the main components of FES-based neuroprostheses including enhanced control systems are presented. Technology and application of FES cycling and rowing, both approaches that enable spinal cord-injured subjects to participate in mainstream activities and improve their health and fitness by exercising like able-bodied subjects, are discussed in detail, and an overview of neuroprostheses that aim at restoring movement functions for daily life as walking or grasping is given.

### 11.1 Introduction

Injuries or diseases can interrupt the conduction of action potentials in the neural system. Depending on the kind and severeness, this may lead to complete or partial loss of control of the muscles of the lower and/or upper extremities.

The application of electrical stimulation in a rehabilitative setting was initiated in 1961, when W.T. Liberson, a physical rehabilitation specialist and medical researcher, developed a heel switch-triggered personal electronic stimulator device to correct foot drop [1]. Functional electrical stimulation (FES) aims to generate movements or functions which mimic normal voluntary movements and so restore

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M. Gföhler (✉)

Research Group for Machine Elements and Rehabilitation Engineering, Institute of Engineering Design and Logistics Engineering, Vienna University of Technology, Karlsplatz 13, 1040 Vienna, Austria

e-mail: [margit.gfoehler@tuwien.ac.at](mailto:margit.gfoehler@tuwien.ac.at)

the functions which these movements serve. Devices that are delivering FES are a type of neuroprosthesis.

Reactivation of skeletal muscles and hence movement functions by FES may have impact on general life, reduce secondary health problems, and increase overall quality of life. Specific training with FES can cause significant improvements of the cardiovascular and pulmonary systems, reduce atrophy of skeletal muscle, increase bone density, and also lead to mental benefits ([2, 3]; Faghri et al. 1992).

## 11.2 Principle

Naturally, the signal for muscle contraction is generated in the central nervous system (CNS). This signal is propagated to and along the peripheral nerve and via the synapsis transferred to the muscle, where it induces the contraction. If this natural muscle activation process is interrupted by a lesion, the activation signals from the CNS cannot reach the muscles and consequently the muscles are paralyzed. FES is a method to artificially generate an activation potential in the peripheral nerve. A stimulator sends a stimulation pulse to the electrodes, and an activation potential is generated in the peripheral nerve and propagated to the muscle in the same way as in the physiologically intact body.

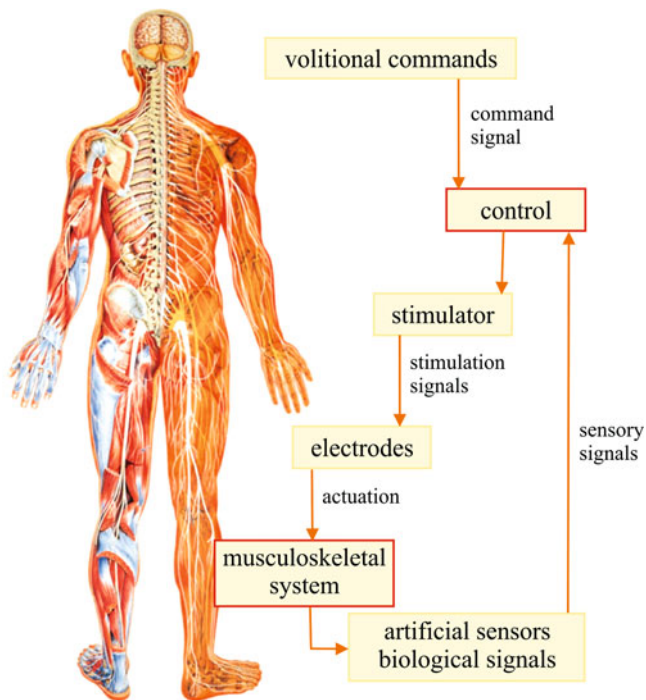
Figure 11.1 shows the main components required for a neuroprosthesis based on FES. Central element of an FES-based neuroprosthesis is the FES controller, which receives command signals and sensory input from artificial and natural sensors and controls the electronic stimulator. The stimulator generates stimulation pulses and induces muscle actuation via electrodes.

## 11.3 Actuation

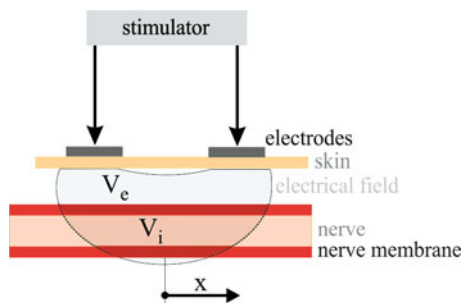
The goal of FES is to stimulate the paralyzed muscles in as natural manner as possible. This requires that the muscles are activated selectively and produce reproducible graded forces. However, many poorly controllable factors related to neuromuscular anatomy and electrode placement make these goals difficult to achieve.

Muscle activation by means of electrical stimulation usually aims at generating an activation potential in the peripheral motor nerves that innervate the muscle, presuming that the peripheral motor nerves are not damaged. Also reflexes can be elicited by electrically stimulating the afferent nerves. Muscle fibers themselves are in principle electrically excitable but require very high stimulation intensities.

The stimulation signal for FES is generated by a programmable stimulator and transferred to the electrodes which transduce electron current into ionic current in the tissue. If the depolarization is strong enough, an action potential is induced in the nerve and propagated along the nerve fiber. This activation potential is then chemically transferred to the muscle fibers via the synapsis and induces muscle contraction and consequently the tendon force. The activating function  $f(x, t)$  is



**Fig. 11.1** Main components of an FES-based neuroprosthesis (figure of the human ©2010 3B Scientific GmbH, Germany)



**Fig. 11.2** Schematic of electrical field distribution in the tissue under surface electrodes

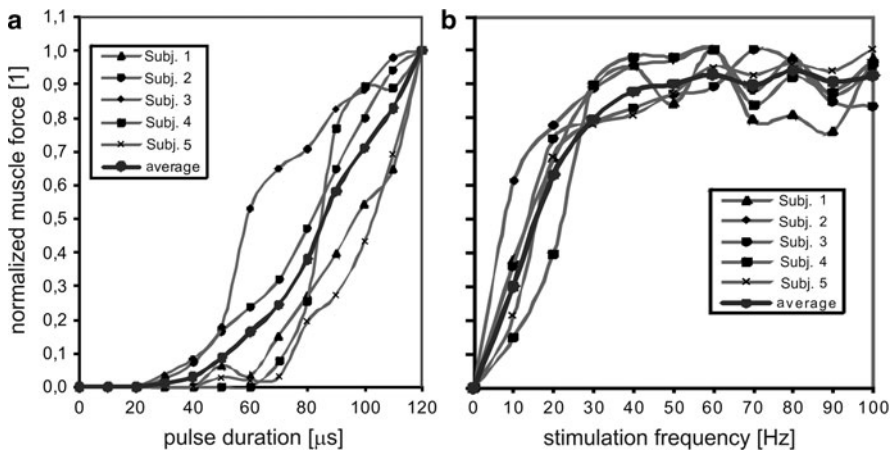
defined at each instant of time  $t$  as the second derivative of the extracellular potential  $V_e$  in direction  $x$  along the nerve fiber (Fig. 11.2) [4]:

$$f(x, t) = \frac{\partial^2 V_e(x, t)}{\partial x^2}.$$

### 11.3.1 Stimulation Signal

The stimulation signal usually consists of a train of biphasic rectangular current pulses with a frequency of between 0 and 100 Hz. Too low frequencies below the critical fusion frequency may lead to rippled muscle force output, and too high frequencies may increase fatigue [5]. For some applications, voltage-controlled pulses are used instead of current-controlled pulses; these are more easy to control but the current is the critical parameter that has to be above threshold level to depolarize the tissue and generate an action potential. The advantage of current-controlled stimulation is that if the resistance of the skin increases due to electrode drying out or sweating, the constant current stimulator will adjust automatically, whereas in a voltage-controlled stimulator the current flowing through the electrode has to be measured and the voltage adapted accordingly to achieve constant stimulation conditions. On the other hand, if an electrode loosens from the skin, the current density flowing through the remaining small contact area may increase to the level where skin damage can occur in a constant current stimulator. Generally, charge balanced pulse types are used so that no net charge is introduced to the body.

Parameters of the stimulation signal that influence the muscle force output are stimulation intensity and pulse frequency. The stimulation intensity can be varied by pulse amplitude and pulse duration, which is limited by the signal's frequency. Figure 11.3a shows a measured isometric recruitment curve (IRC), the relation between stimulation intensity (the pulse duration is varied at constant pulse amplitude) and isometric muscle force. If the stimulation intensity is higher than a threshold value, the force increases almost linearly until saturation is reached. Figure 11.3b points out that the isometric muscle force increases with stimulation



**Fig. 11.3** (a) Isometric recruitment curve IRC, (b) relation between stimulation frequency and isometric muscle force, results of measurements on the Quadriceps muscles of five male paraplegic subjects and average [6]

frequency. The maximum force is reached at about 30 and 100 Hz for slow and fast contracting motor units, respectively. For frequencies above 50 Hz, muscle fatigue, which is a severe problem in FES applications, increases rapidly [5].

Muscle composition is changed in paralyzed muscle due to inactivity. The percentage of fast contracting, fast fatiguing muscle fibers increases, which is one reason for quick fatigue in paralyzed muscle. These inactivity-associated muscle changes can at least partially be reversed by FES training (Mohr et al., 1997).

In the case of physiological activation, first the thin, slow contracting motor units of the muscle are activated, and when higher forces are needed, bigger, fast contracting motor units are subsequently added. Similarly, the stimulation frequency is low at the beginning and raised for higher forces. The activated motor units are distributed over the muscle. In the case of artificial stimulation, the recruitment order is reversed to so-called inverse recruitment [7]. This means that big, fast contracting and fast fatiguing motor units are activated first. Additionally, motor units in the region of the muscle where the electrical field is stronger are activated first. Therefore, some parts of the muscle might be active while other parts are totally inactive.

### **11.3.2 Electrodes**

Electrodes build the interface between the neural system and the technical device of the neuroprosthesis. A variety of interface concepts have been developed, ranging from simple wires to complex microsystems with integrated electronics. In general, selectivity increases with invasiveness.

#### **11.3.2.1 Surface Electrodes**

Surface electrodes are attached to the skin above a nerve or motor end plate. Their advantage is that they are noninvasive and easy to use. Disadvantages are the high influence of the electrical resistance of the skin and other tissues between electrode and nerve with respect to the distribution of the electrical field and the geometrical restrictions. It is impossible to reach deep-lying muscles without also stimulating overlying superficial muscles. High stimulation intensities are necessary, and it is difficult to predict which portions of the muscle are reached by the electrical field. One way to improve selective activation is to dynamically switch the cathode between sets of small transcutaneous electrode elements. Recently, novel embroidered electrodes have been used [8].

#### **11.3.2.2 Subcutaneous Electrodes**

*Intramuscular electrodes* are fine wire electrodes that are either inserted directly through the skin as percutaneous electrodes or tunneled subcutaneously. Percutaneous electrodes are less invasive than fully implanted electrodes, but positioning

is difficult and relative movements can occur during movement. In addition, stress points occur where the wires cross the skin and at fascial planes between muscles. Frequent bending at these points can cause the wire to break, and the wire from the electrode comes through the skin providing a path for infection. Therefore, percutaneous electrodes are rarely used for long-term systems.

*Epimysial electrodes* are surgically placed on the muscle near the motor point. They consist of disk-shaped metals with a polymer shielding the surface away from the muscle.

### 11.3.2.3 Nerve Electrodes

Nerve electrodes are placed directly at the nerve – either adjacent, encircling, or intraneural.

*Extraneural cuff electrodes* consist of an insulating tubular sheath that encircles the nerve and contains two or more electrode contacts at their inner surface that are connected to insulated lead wires. The electrodes distributed around the circumference of a peripheral nerve are intended to activate different populations of axons. Cuff electrodes are easy to implant but may lead to nerve damage if their size is not well adjusted to the nerve diameter.

*Intraneural electrodes* are placed either longitudinally (Longitudinal Intra-Fascicular Electrode, LIFE [9]) or transversally (Utah Slanted Electrode Array, USEA [10]) in the peripheral nerve endoneurium and have higher recording selectivity and signal-to-noise ratio than extraneural electrodes. The LIFE is used for neural recording or stimulation small subsets of axons within a nerve fascicle. Typical records from LIFEs show multiunit activity where it is sometimes possible to resolve single units. LIFEs with 10  $\mu\text{m}$  thickness and 50 mm in length have been realized using thin-film microfabrication techniques on polymer substrates, which also makes them more flexible and mechanically compatible [11]. The USEA is a silicon-based, three-dimensional structure consisting of a  $10 \times 10$  array of tapered silicon electrodes that project out from a  $4 \text{ mm} \times 4 \text{ mm}$  substrate that is transversally inserted into the peripheral nerve for neural recording or stimulation. The lengths of the electrodes are graded from 0.5 to 1.5 mm along the length of the array to ensure that when it is inserted into a peripheral nerve, the electrode tips uniformly populate the nerve.

*Sieve electrodes* consist of a matrix of holes that is positioned at the end of a nerve. Ideally, the axons of the nerve will grow through the holes and build electronic contacts. Sieve electrodes have not yet been tested in human applications [11].

## 11.4 Stimulators

*External stimulators*: several multichannel programmable devices with analog and digital input and output lines are commercially available.

A commercialized *implantable* device is the eight-channel receiver-stimulator IRS-8, which receives power and control via an external close-coupled radio

frequency signal. It is used in the Freehand system<sup>®</sup> for active grasp and release. Based on the IRS-8, two implantable stimulator–telemetry systems (IST) have been developed which have additional input lines for sensory signals.

Loeb et al. [12] developed a fully *implantable wireless microstimulator*, the BION (“bionic neuron,” 2 mm diameter  $\times$  16 mm long). Multiple BIONs can be injected through the barrel of a hypodermic needle near the nerve or neuromuscular junction of interest. Each BION receives power and digital commands from a telemetry link and delivers current pulses of the requested duration and amplitude via electrodes that are mechanically fixed on either end of its elongated capsule.

## 11.5 Control

### 11.5.1 Modeling/Simulation

For simple tasks, muscle stimulation patterns are developed by combining clinical experience with trial and error, but it is difficult to find smooth and energy efficient movements with trial and error because of the dynamic interactions between the segments. For more complex movements, muscle stimulation patterns have to be determined mathematically by establishing a dynamic model of the musculoskeletal system. This model usually consists of rigid body segments that are linked by joints and the musculotendon actuators. Due to the complexity and variety of the biological system parameter, identification is a main problem. Many parameters are difficult to access in vivo, and there are big differences between subjects. Additionally, in the case of physically disabled subjects, changes in muscle structure occur depending on the injury. Recently, MRI techniques have brought advancements in estimating musculoskeletal data of individual subjects.

To determine the impact of electrical stimulation on a movement, the resulting muscle force has to be determined. Muscle models with varying complexity are available [13]. Generally, the muscle force generation is divided into two processes, activation and contraction dynamics, as shown in Fig. 11.4. Both activation and contraction dynamics act as a low pass filter with the output responding slower and more smoothly than the input [14]. Muscle activation corresponds to the

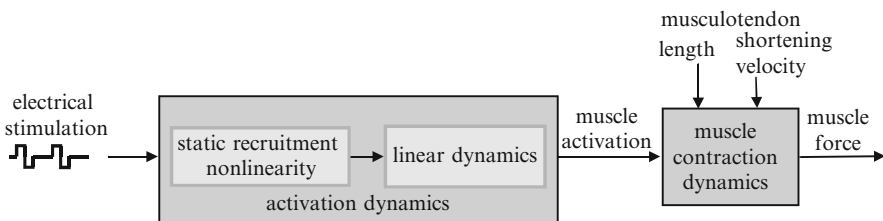


Fig. 11.4 Schematic of muscle activation and contraction dynamics

Ca-concentration and is described by the linear activation dynamics in the case of artificial activation by electrical stimulation. The static recruitment nonlinearity additionally accounts for the impact of stimulation intensity and stimulation frequency on muscle force according to the relations shown in Fig. 11.3. Muscle activation is slower and deactivation is faster in electrically stimulated muscle in comparison to physiological activation. From measurements on paralyzed leg muscles, a rise time of 108 ms was determined for 0–70% of maximal activation, and a fall time of 65 ms for 100–30% [15]. Muscle contraction dynamics describe the generation of force by activated contractile elements and basically shows the same behavior in healthy and artificially activated muscle. A muscle’s force at each instant of time is a function of the instantaneous musculotendon length and shortening velocity, the tetanic muscle force, and muscle activation.

The muscle forces act on the body segments. The behavior of the musculoskeletal system is described by the equations of motion which are derived from the Newton–Euler equations. A system with  $n$  degrees of freedom (joint angles) has  $n$  equations of motion, which can be represented in vector form:

$$[\mathbf{A}]\cdot\ddot{\theta} + [\mathbf{B}]\cdot\dot{\theta} + [\mathbf{C}]\cdot g = \underline{\mathbf{M}},$$

where  $[\mathbf{A}]$  is the  $n \times n$  mass matrix,  $\underline{\theta}$  is the vector of the system’s  $n$  degrees of freedom,  $[\mathbf{B}]$  is the gyroscopic matrix including centrifugal and coriolis terms,  $\underline{\mathbf{M}}$  is the vector of joint torques directly due to muscle forces, and the term  $[\mathbf{C}]\cdot g$  represents the torques due to gravity. As  $[\mathbf{A}]$  is generally a full matrix, a muscle acting on one joint can accelerate all other joints of the system, this is called dynamic coupling.

To generate a defined movement by FES, it has to be determined which of the stimulated muscles have to be active in which phase of the movement and at which level. A forward dynamic model (Fig. 11.5) is used to calculate the resulting movement trajectories from muscle stimulation. Muscle forces and musculoskeletal geometry give joint torques, then the equations of motion are used to determine the joint angular accelerations, double integration finally gives the joint angles.

If the desired kinematics is known, inverse dynamics (Fig. 11.6) can be used to determine the optimal timing of the muscle forces. The joint torques are calculated with the system’s equations of motion. As each joint is usually spanned by more than one muscle, there is a distribution problem when calculating muscle forces from

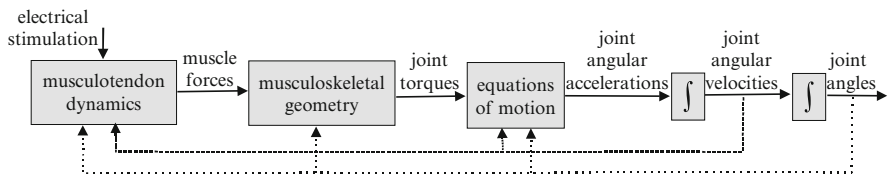
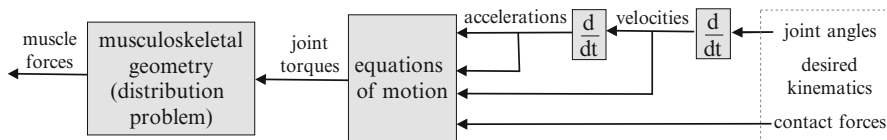


Fig. 11.5 Forward dynamic model





**Fig. 11.6** Inverse dynamic model

joint torques. To split up the total torque to the single muscles, static optimization has to be applied. A commonly used performance criterion is to minimize muscle stress, squared, summed across all muscles [16]. As the static optimization does not consider musculotendon dynamics, the resulting muscle force trajectories may be unrealistic because a muscle cannot develop a force instantaneously.

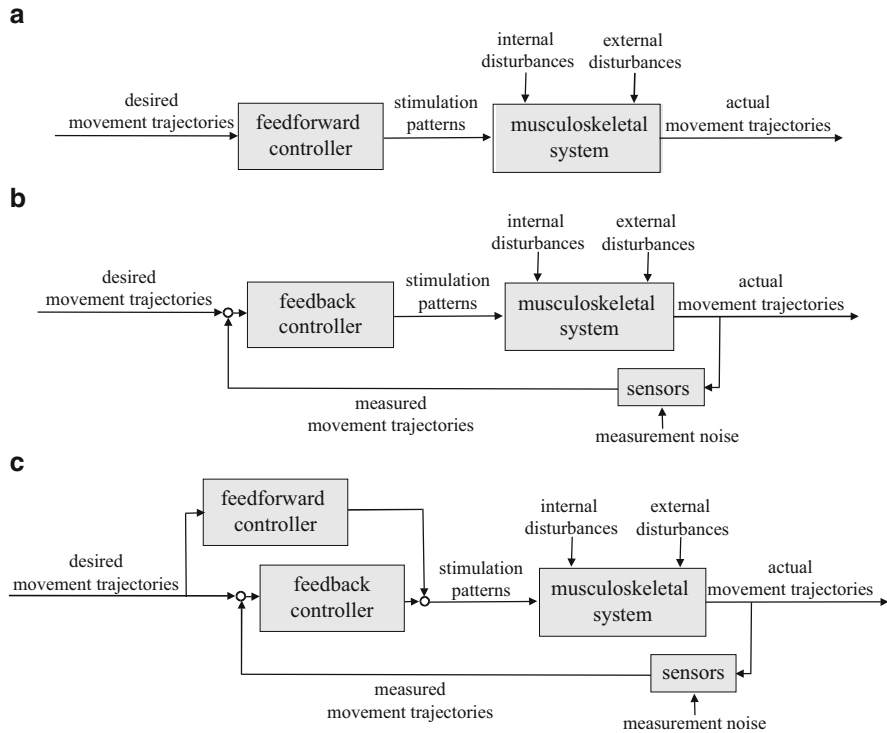
To determine optimal stimulation patterns, muscle activation dynamics have to be considered. This is possible using a forward dynamic model where stimulation patterns are the input and the resulting movement is the output. A performance criterion applicable to the entire task has to be defined and forward dynamics combined with dynamic optimization methods to determine the optimal stimulation patterns. Parameter optimization methods as described in [17] are suitable to solve this nonlinear optimization problem, though these methods cannot differentiate between local and global maxima. Statistical methods as simulated annealing or genetic algorithms are likely to find global maxima but have the disadvantage of high computational expense. An alternative for estimating muscle forces and muscle excitations that requires three orders of magnitude less CPU time than parameter optimization is neuromuscular tracking [18].

### 11.5.2 Control Systems

Designing a control system that tracks a joint trajectory or torque profile by regulating the timing and levels of the electrical stimulation delivered to the muscles is a challenging problem due to the nonlinear response of the muscles to electrical stimulation and the complexity and redundancy in the musculoskeletal system.

Electrical stimulation may be delivered through either open or closed loop control systems. The FES controller attempts at taking over control tasks from the natural sensorimotor system and interfaces with the natural system at stimulation output and depending on the control approach also at command input.

In *open loop* control systems (Fig. 11.7a), the controller determines the muscle stimulation according to the desired movement trajectory, and no information about the actual trajectory is fed back to the controller. The performance of open loop systems was found unsatisfactory for the generation of accurate movements, because external disturbances as obstacles or internal disturbances as variations in muscle force generation (e.g., due to fatigue) or inaccuracies in the model of the musculoskeletal system have impact on the actual movement trajectory.



**Fig. 11.7** Schematic of (a) open loop control, (b) closed loop control, and (c) hybrid control with both a feedforward and a feedback controller

For more complex or accurate movements, *closed loop* control (Fig. 11.7b) has to be established. During normal voluntary movements, the CNS receives sensory information on muscle-, tendon-, and cutaneous forces for neurophysiological control. In closed loop control systems, electrical stimulation is being initiated by the user's command and then modified based on some feedback measurement such as force or position. With closed loop control, the delivery of electrical stimulation is continuously modulated to control the parameter being measured by the sensors. The benefits of closed loop control are obvious, but closed loop control systems are more complex to design and implement. Furthermore, measurement noise or errors in the feedback signals can lead to unexpected behavior of the system.

For dynamical systems that are subject to both disturbances and measurement noise, hybrid control systems (Fig. 11.7c) combining feedforward and feedback control can improve the total performance [19]. Hybrid control systems have frequently been applied for control tasks of the musculoskeletal system [20–22].

In model-based control, a musculoskeletal model is directly used as the controller. An inverse dynamics model can be used as a forward controller with the desired trajectories as input and optimal stimulation patterns as output. Muscle

activation dynamics have to be linearized. Ferrarin et al. [21] designed a model-based feedforward control of the knee joint angle and combined it with a PID feedback controller. Jezernik et al. (2004) developed a sliding mode closed loop controller based on a musculoskeletal model for controlling the shank movement by FES.

The internal parameters of the musculoskeletal system may change because of internal disturbances such as fatigue-induced changes in muscle force generation. *Adaptive controllers* in which the controller parameters are allowed to adapt to changing plant parameters have been used to cope with such phenomena [23].

Due to their ability to map arbitrarily complex nonlinear input/output relationships from a given data set, *artificial neural networks* have been successfully applied to predict patterns of muscle stimulation needed to produce complex movements with FES-based neuroprostheses [20, 24–26]. EMG recordings from muscles under voluntary control and/or kinematic data have been used as input for the training of the neural controller [27].

## 11.6 Sensors

### 11.6.1 Artificial Sensors

Artificial sensors that are suitable for closed loop control of FES are force or pressure sensors. They are mostly placed at the point of contact, like ground contact in walking or grip force. Magnetic goniometers based on the Hall effect are used for measuring joint angles. DC accelerometers can be placed on the limb segments. Ambulatory position and orientation of human body segments can be measured accurately by combining an inertial measurement unit consisting of miniature gyroscopes, accelerometers, and magnetometers [28]. MEMS technology even allows incorporating accelerometers into injectable stimulation devices [29]. Most of the artificial sensors are placed externally on the moved limb, imposing further limitations on size, shape, and weight.

### 11.6.2 Natural Sensors in the Peripheral Nervous System

Natural sensors in the peripheral nervous system such as those found in the skin, muscles, tendons, and joints present an attractive alternative to artificial sensors for FES systems. Most of the peripheral sensory apparatus is still viable after injuries in the brain or spinal cord, yet not connected to the CNS.

Nerve cuff electrodes similar to the cuff electrodes for nerve stimulation have been used for chronic recording of *ENG signals* from sensory nerves. The ENG has a very small amplitude and a major source of interference is the myoelectric activity

of nearby muscles. The EMG amplitude is approximately three orders of magnitude larger than the  $\mu\text{V}$  ENG and their spectra overlap. Implantable amplifiers have been designed, which, placed close to the recording electrode, remove EMG overlap. Control of FES thumb force using slip information obtained from the cutaneous electroneurogram has been used to control FES grip force [30].

Multicontact nerve cuff electrodes can work bidirectional by stimulating individual fascicles of nerve trunks and recording multiunit afferent activity from peripheral nerves [29].

The Utah Slanted Electrode Array USEA is inserted into the peripheral nerve for neural recording or stimulation.

### ***11.6.3 Volitional Biological Signals***

#### **11.6.3.1 EMG**

EMG is used to assess residual volitional motor activities. In so-called EMG-triggered stimulation, movement phases are initiated by volitionally activating the muscle whose EMG is measured. More sophisticated approaches establish closed loop control by modulating the stimulation intensity proportional to the measured EMG signal. EMG signals can be recorded by transcutaneous electrodes giving a noninvasive and relatively robust method for sensory input to the FES control. Any muscle the user can volitionally activate can be used for EMG recording. In the case of incomplete paralysis, it is also possible to record voluntary EMG from the same muscle that is stimulated. Bidirectional electrodes are available that can both record EMG and stimulate the muscle.

#### **11.6.3.2 Brain Computer Interfaces**

Brain computer interfaces (BCI) systems extract commands directly from the brain. The user imagines to perform a movement and brain activity signals are gained directly from the neuronal activity patterns in the corresponding motor areas of the brain. An advantage for the control of neuroprostheses is the fact that the imagined movement need not necessarily be the desired movement. Any type of command signal that is convenient for the user to generate can be used by the FES system. For example, foot movement can be imagined to trigger the FES system to open/close the hand. Due to the high inter- and intrasubject variability motor learning strategies have to be applied.

*Noninvasive systems* record the electroencephalogram (EEG) from the scalp or use functional magnetic resonance imaging (fMRI). The acquisition of high levels of control usually requires extensive user training. EEG-based BCIs are frequently used to trigger preprogrammed movements by FES-like hand grasp. Classifier functions are used to choose between two or more different brain states. These signals are then used like switches between different phases of a movement pattern.

*Invasive* methods use local activity from multiple neurons recorded within the brain. They show higher selectivity and are more successfully applied for complex control tasks but have the disadvantage of significant clinical risks and limited stability.

*Electrocorticographic* (ECoG) recording from the cortical surface has been tested as an alternative to current noninvasive and invasive recording methods [31].

First human pilot trials with both invasive and noninvasive systems suggest that BCIs could be a future option for the control of neuroprosthesis in patients with high-level SCI [32–34]. Still, there is little information available on the changes in the neural circuits in the brain after spinal cord injury, and optimal signal processing techniques have to be found to convert the existing brain signals efficiently and accurately into operative control commands.

## 11.7 Applications for the Lower Limb

Lower limb FES systems are used to restore walking [35], standing [36], sit-to-stand [37], cycling [38], and rowing [39]. Balance and the risk of falling are main problems in all upright body positions. As relatively high muscle forces are necessary for carrying the body weight, muscle fatigue is a serious problem in lower limb systems because it can cause falls and possible injury.

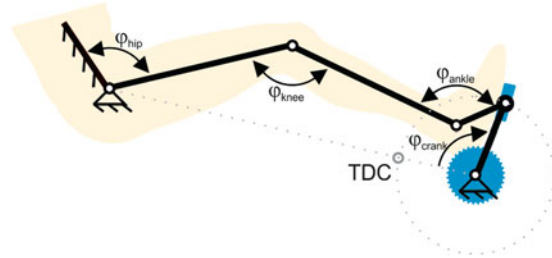
### 11.7.1 Cycling

Mobile FES cycling outdoors is attractive for paraplegics because they can use a standard bike (tricycle) with only a few modifications and move independently, powered by their own muscle force, over relatively long distances. Problems with balance are avoided by the seated body position, and compared to other types of movement, cycling has the advantage that the force applied to the pedal is converted into motion with very high efficiency.

FES leg cycling ergometry is frequently applied for muscle training in rehabilitation. The first commercialized leg cycling exercising system was ERGYS (Therapeutic Alliances Inc.) in 1984. So far, only external FES systems with surface electrodes have been used for FES cycling. For paraplegics, a number of leisure and sport activities are available, like basketball or hand-cycling, where only the intact upper extremities are activated. But the muscle mass of the upper extremities alone is not big enough to achieve oxygen consumption and heart rates above threshold, where the training is effective for reducing risk factors for cardiovascular and metabolic diseases. In comparison, the physiological benefits of the FES cycling training are relatively high [40].

Research on cycling by means of FES has been a focus of rehabilitation engineering at the Vienna University of Technology for several years. An instrumented

**Fig. 11.8** Two-dimensional skeletal model [41]



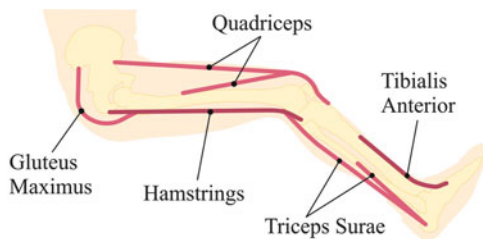
FES cycling system has been developed [38] that serves as both a stationary cycle ergometer and a mobile tricycle for paraplegics.

### 11.7.1.1 Simulation

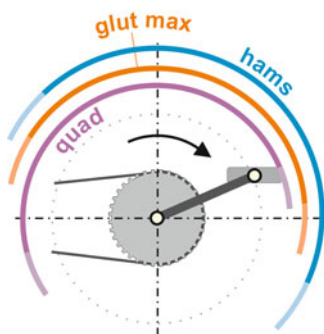
A forward dynamic simulation was established to optimize the stimulation patterns for FES cycling and to determine the influence of parameter changes.

A musculoskeletal model of paraplegic isokinetic cycling on a recumbent cycle was established [41]. The skeletal model is two-dimensional and effectively consists of five rigid segments connected in frictionless hinge joints (Fig. 11.8). These segments represent the crank, foot, lower leg, upper leg, and head-arms-trunk (HAT). The point of contact between foot and pedal is under the metatarsophalangeal (MTP) joint by default. Usually, in FES cycling the ankle joint is fixed by an orthosis that also stabilizes the leg. This means that the skeletal system has only one degree of freedom and consequently the leg kinematics are entirely determined by the imposed crank kinematics, and the muscle stimulation affects the forces but does not affect the kinematics. But as the power output in FES cycling is usually quite low and overcoming the dead center is sometimes problematic, it was investigated what effect releasing the ankle joint and additionally stimulating the muscles spanning the ankle joint has on the power output and overcoming the dead center. Releasing the ankle joint adds a second degree of freedom to the linkage and thus aggravates a control problem. Not only the force applied to the pedal but also the movement has to be controlled by the muscle stimulation. On the other hand, releasing the ankle joint and additionally stimulating the muscles spanning the ankle joint brings additional physiological benefits. The equations of motion of the skeletal system were derived from the Newton–Euler equations.

The skeleton is actuated by muscles/muscle groups of the lower extremity that are stimulated during FES cycling. For fixed ankle, these are Quadriceps (Vastii and Rectus Femoris receiving identical stimulation), Gluteus Maximus, and Hamstrings. For released ankle, in addition Soleus and Gastrocnemius (receiving identical stimulation) and Tibialis Anterior are included (Fig. 11.9). A Hill-type muscle model [42] is used to represent these muscles. It consists of a contractile element, a series elastic element, and a parallel elastic element. The latter element is present in the model but has no effect in the optimal solutions. According to measurements on



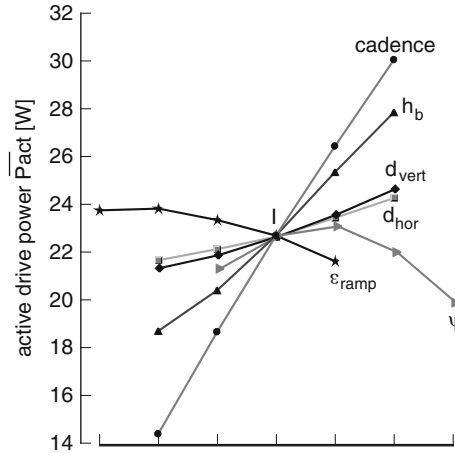
**Fig. 11.9** Electrically stimulated leg muscles during FES cycling



**Fig. 11.10** Optimal stimulation pattern for fixed ankle joint for isokinetic FES cycling at 45 rpm. The light regions at the beginning and end of each stimulation interval indicate that the stimulation is switched on and off gradually along a ramp to avoid spasms and after-twitches

paralyzed muscles [15], the muscle activation and deactivation constants were set to 0.108 s and 0.065 s, respectively, based on the 0–70% rise time and 100–30% fall time for muscle force during isometric contraction and maximum isometric forces were set to 17% of the mean values for able-bodied subjects.

A forward dynamic simulation of isokinetic FES cycling at 30/45/60 rpm was performed. As optimization method, a parallel genetic algorithm [43] was applied. Input is muscle stimulation. The optimization criterion was to maximize mean mechanical power output over one full rotation of the crank. Figure 11.10 shows the optimal stimulation patterns for isokinetic cycling at 45 rpm, and the generated drive power is 90 W. For released ankle, the optimization results with the described model show that the ankle plantar flexors are unable to resist the torque of the pedal reaction force. To avoid this problem, it is either necessary to shorten the effective foot length by moving the position of the contact point of the foot sole and the pedal or to increase the maximal isometric force of the ankle musculature. Higher maximal isometric force of the ankle musculature might be realistic in many patients because spastic contractions reduce muscle atrophy. Shortening the effective foot length from 0.165 to 0.055 m resulted in a 10% power increase. Double maximal isometric force in the ankle musculature plus shortening the effective foot length



**Fig. 11.11** Active power output of all stimulated muscles summed up and influence of variation of the single parameters. All curves pass through the point I where the optimization is performed with an initial average set of parameters (details in [44]).  $h_b$ ... body height;  $d_{vert}$ ,  $d_{hor}$ ... vertical and horizontal distances between crank axis and hip joint, respectively;  $\psi$ ... backrest angle (from vertical),  $\epsilon_{ramp}$ ... length of ramp at begin and end of stimulation

to 0.11 m shows a 29% power increase and the generated drive torque is positive over the full crank rotation what means that overcoming the dead center might be facilitated.

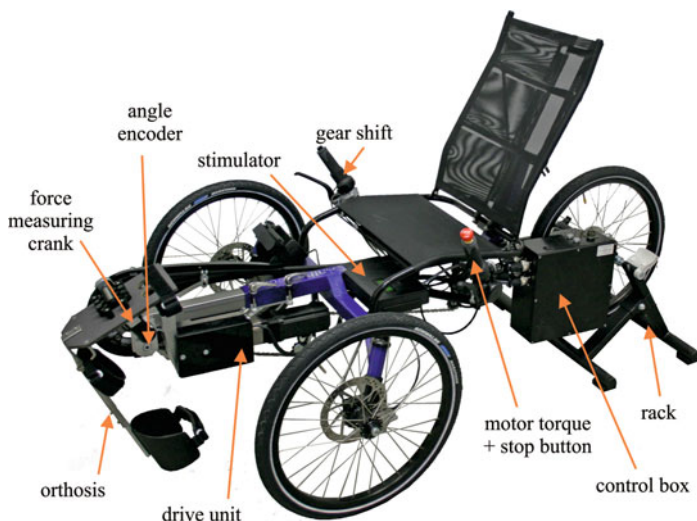
FES-cycling performance is influenced by a number of parameters such as seating position, physiological parameters, conditions of surface stimulation, and pedaling rate. A sensitivity analysis was performed to determine the influence of the most important parameters on optimal muscle stimulation patterns and power output of FES cycling (Fig. 11.11) [44].

The results of the simulation show in which regions the individual leg muscles should be stimulated and what influence some parameters have on stimulation and power output. Still there is a number of parameters which cannot be considered adequately in the simulation. These include unpredictable spasm activity which also strongly depends on daily condition of the patient, co-stimulation of antagonists, muscle fatigue, and muscle force hysteresis, as the muscle force depends on both activation and movement history. Also muscle condition is very different among patients due to factors such as training status, spasms, type of and time since injury. It is important to test the relation between stimulation parameters and generated muscle forces for each patient individually.

### 11.7.1.2 Instrumented FES Cycling System

For details see [38].





**Fig. 11.12** Instrumented test- and training system and main components. The control box contains the motor control and the accumulators

### Mechanical Design

A commercially available tricycle was adapted as the basic frame for the FES-cycling system (see Fig. 11.12). The horizontal distance between the crank bearing and the seat can easily be adjusted to different leg lengths. For easier transfer from a wheelchair to the tricycle, the right steering handle can be dismantled by opening a quick clamp and a transfer board hugged on to the frame.

### Orthoses

Figure 11.13 shows the orthoses which are mounted on the pedals. Their function is to stabilize the legs in the parasagittal plane during pedaling. Due to the telescopic shaft, the orthoses are adaptable to the length of the users shank.

### Force Measurement Cranks

The force measurement cranks are based on strain gauge technology. Strain gauges are arranged in three full Wheatstone bridges on the aluminum corpus of the cranks. The arrangement of the strain gauges allows measurement of the radial force (in the direction of the crank), the tangential force (rectangular to the crank in the pedaling plane), and the torque around the longitudinal axis of the crank. The signals of each crank are amplified, digitized, and sent to a laptop computer. A specialized time



**Fig. 11.13** Orthoses for leg stabilization in the parasagittal plane. *Left*: lightweight orthoses fixing the ankle joint; *right*: orthosis with a ball bearing and adjustable movement range at the ankle and a force measuring unit for measurements on the generated ankle torque

synchronized telemetry network is used to ensure time-correlated measurements of left and right leg.

### Electrical Stimulation

Figure 11.14 shows the current-controlled 10-channel stimulator that was developed for FES cycling and rowing applications. The stimulator induces biphasic rectangular pulses with a stimulation current from 0 to 150 mA (at 1 k $\Omega$ ), frequency from 0 to 100 Hz, and pulse width from 0 to 700  $\mu$ s. In “time mode” the stimulation is delivered as a function of time, in “angle mode” as a function of an angle signal that is processed in the stimulator. The stimulator automatically shifts the stimulation pattern backward as a function of the actual cadence to consider the dynamic characteristics (activation and deactivation times) of the muscles. Up to three sets of individual stimulation patterns can be stored in the stimulator. During FES cycling, the preset stimulation currents can be scaled from 0 to 100% by turning an adjusting knob on the right steering bar of the FES cycle.

### Drive Train

The main component of the drive train is the motor unit consisting of a servo-motor and a planetary gear. An electromagnetic coupling connects the motor unit to a bevel gear on which the pinion is mounted. The pinion is then connected to the cranks by a chain.



The motor generates a constant drive torque which is controlled by a turning handle on the left steering bar of the cycle by the user. If no motor support is necessary, the motor can be decoupled by switching off the electromagnetic coupling.

The gears of the gear hub in the back wheel are changed by turning the outer part of the right steering bar.

- Stationary cycling and measurement mode

For stationary cycling, the cycle is hooked up on a rack.

Enhanced control for both stimulator and motor is available by connecting a computer to the system via an RS 232 Interface. A LabView-based control program then communicates with the stimulator and the motor control, and reads in motor current and crank angle data and additionally force data from the force measuring cranks. The control program offers two modes of operation: an expert mode with unlimited access to all parameters of the muscle stimulation and motor control, and a wizard mode with limited access which guides the user step by step through a predefined series of training and measurement units. The data processing is automated and the measurement data of all the training units of one person are stored together with the personal log file.

To allow reproducible force measurements, the motor can either move the cranks at constant angular velocity for isokinetic measurements or hold the cranks on a defined position for isometric measurements. Predefined measurement routines allow to determine individual sets of optimal stimulation parameters based on the results of the mathematical simulation. At first, isometric measurements determine the muscle's force response and adequate stimulation intensity for cycling, then the crank angle interval is determined, in which the muscle applies positive crank torque. Isokinetic measurements then adapt the optimal stimulation interval to higher cadences.

### 11.7.1.3 Clinical Application

The FES cycling system offers multifunctional equipment for FES-cycling training and therapy. Currently, the system is being tested as a rehabilitation tool in clinical rehabilitation for spinal cord-injured subjects in a clinical study in cooperation with the AUVA Rehabilitation Center Weisser Hof in Austria. Patients are doing an FES training session three times a week over 2 months as part of their rehabilitation program and also do outdoor cycling. Figure 11.16 shows a paraplegic subject performing stationary training on the FES tricycle. Due to the force measurements and the automated data processing, the therapy progress can be well monitored. Also spasticity is assessed before and after each FES session, and it has been shown in accordance with earlier studies that the FES training reduces spasticity at least temporarily [45].

**Fig. 11.16** Paraplegic subject performing stationary training on the FES tricycle



## 11.7.2 Rowing

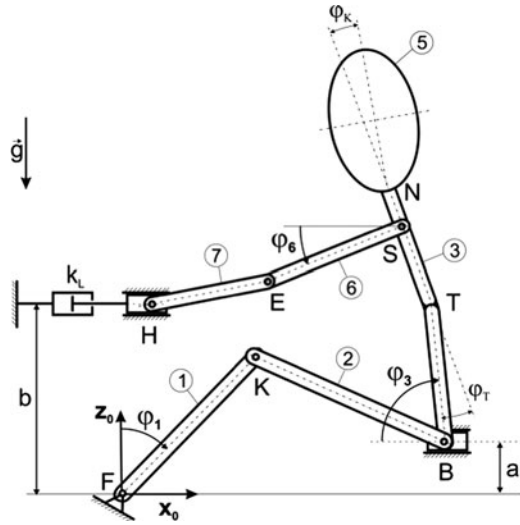
Ergometer rowing with FES of the muscles in the lower extremity enables paraplegics to participate in this mainstream activity for health, leisure, and sport. As muscle mass of both upper and lower extremities is metabolically active during the rowing motion, the cardiovascular training is higher than in exercises where only the muscle mass of the lower extremities is activated (Hettinga et al. 2004).

### 11.7.2.1 Simulation

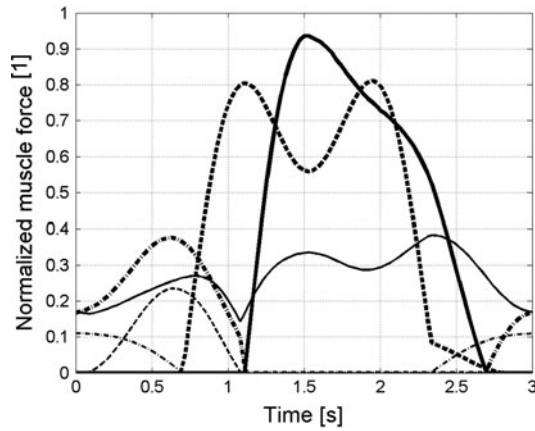
For determination of the optimal stimulation patterns for ergometer rowing by means of FES, a musculoskeletal model was established in Matlab Simulink (Kuchler and Gföhler, 2004). The model of the user-rowing machine system represents a planar 8-link kinematic chain with three degrees of freedom (Fig. 11.17). The resistance mechanism was modeled by Euler's principal equation for flow machinery (damping element  $k_L$  in Fig. 11.17). The equations of motion were derived using the Newton–Euler equations.

Seventeen muscle groups of the upper and lower extremities were considered. The muscles of the lower extremities are activated by surface electrodes. Maximum isometric forces were scaled according to measurements [15]. The three degrees of freedom represented by the angles  $\varphi_1$ ,  $\varphi_3$ , and  $\varphi_6$  and the vertical contact force at the seat were used as inputs to solve the inverse dynamic problem [46]. The muscle forces were determined using mathematical optimization [16]. Figure 11.18 shows the normalized muscle forces in the lower extremities over one complete rowing cycle. The results show that a high muscle force from Iliopsoas is necessary for hip flexion at the beginning of the recovery phase. But the deep-lying Iliopsoas muscle

**Fig. 11.17** Model of the paraplegic-rowing machine system



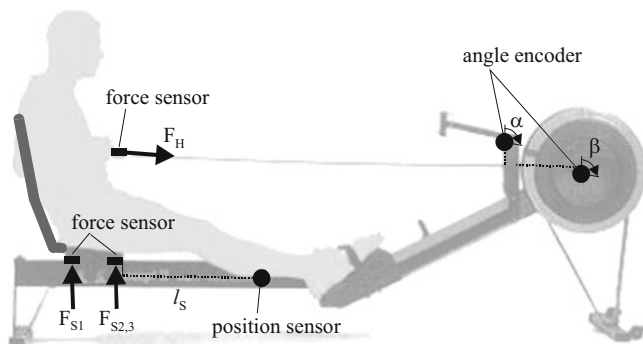
**Fig. 11.18** Forces of Tibialis anterior (*thick dashed*), Soleus and Gastrocnemius (*thin dash-dot*), Vastii and Rectus femoris (*thin dashed*), Hamstrings (*thin solid*), Gluteus maximus (*thick dash-dot*), and Iliopsoas (*thick solid*) normalized by the corresponding maximum isometric force during one full rowing stroke



cannot be stimulated with surface electrodes; consequently, there will be difficulties with generating a sufficient hip flexion torque, and this has to be considered in ergometer design.

**11.7.2.2 Instrumented FES Rowing Ergometer**

For experimental investigations on FES rowing, an instrumented rowing ergometer was designed based on a standard Concept II (Concept2 Deutschland GmbH, Hamburg, DE) ergometer [47]. The sliding seat was replaced by a construction with soft seating and adjustable backrest for stability of the upper body. Orthoses are fixed on the foot rest for side-to-side stability of the legs. Five muscles/muscle



**Fig. 11.19** Instrumented FES rowing ergometer

groups are stimulated in each leg by surface electrodes: Tibialis anterior, Soleus and Gastrocnemius, Vastii and Rectus femoris, Hamstrings, and Gluteus maximus.

Instrumentation of the ergometer allows the determination of actual kinematics and kinetics for further investigations and individual optimization of the rowing motion. The vertical seat force is measured by a combination of three force measuring cells so that the torques can be eliminated. A force measuring cell which is placed between handle and chain gives the pull force at the handle. The horizontal position of the seat is measured by a position sensor. Two angle decoders are used for measuring the length and angular position of the chain from the handle to the resistance mechanism and hence define the two-dimensional position of the handle. The digital signals from the angle decoders and the position sensor are read in the control via a microprocessor. The analog signals from the force measuring cells are read into a data acquisition card and from there also to the control. The joint trajectories of all three degrees of freedom can be calculated from the measured position data. Together with the measured vertical seat force, all necessary input data are available to solve the inverse dynamic problem with subject-specific input data (Fig. 11.19).

The control for the FES rowing motion is based on the same LabView program as for FES cycling; the 10-channel stimulator which was developed for FES cycling was extended with a rowing mode where the stimulation of the leg muscles is controlled by the horizontal position of the seat which is detected by a position sensor. The range of horizontal movement of both handle and seat are defined as a function of the leg length for each individual subject and the stimulation pattern is scaled accordingly.

### 11.7.3 Gait

*Foot drop* is a gait-limiting factor in patients with stroke or other disorders of the CNS. Due to weakness of the ankle dorsiflexors, the foot drops and may drag on

the ground during the swing phase of gait. The first system to correct foot drop was proposed in 1961 by Liberson. In this device, the common peroneal nerve was stimulated when the heel came off the ground to flex the ankle and thereby lift the foot. A few systems based on this principle are commercially available. For the stimulation either surface electrodes on the anterior leg muscles or implanted electrodes near the peroneal nerve are used, the stimulation is synchronized to the gait phase by either a switch under the heel or neural signals recorded from the sural nerve with a cuff electrode or inertia sensors on the foot (WalkAide<sup>®</sup>, Innovative Neurotronics; Veltink 2003; Weber et al. 2005).

*Walking* by means of FES is an attractive and desirable goal for many patients because it gives them the ability to move as people do naturally. However, mainly due to the problem of keeping balance and weight carrying, equipment for FES walking is cumbersome to don and operate and only safe to operate under highly controlled conditions. During FES walking, patients usually have to use a rollator or crutches what makes it less applicable in daily life. The first systems enabling walking were external devices that were developed by Kralj et al. [48]. Currently mainly systems with surface electrodes are used, but also fully implanted systems with stimulation of up to 48 muscles are available. The implantation frees the patient from cabling, but still a walking frame or crutches are necessary. All available systems are open loop controlled. The user has to initiate the step, and then preprogrammed stimulation sequences are carried out for the whole step cycle. Hybrid systems combine electrical stimulation with mechanical bracing and can potentially combine the best features of mechanical bracing and FES into new systems for walking after SCI that offer more advantages than the individual components acting alone.

## 11.8 Applications for the Upper Limb

FES systems restoring upper limb function are mainly suitable for tetraplegics with cervical lesions; the peripheral nerves innervating the arm muscles must be intact. There are systems based on surface electrodes and percutaneous electrodes as well as fully implanted systems available. The higher the level of the spinal cord injury, the more joints must be controlled. Also the higher the injury, the fewer biological control signals are available. FES has so far concentrated on restoring grasp in patients with C5/C6 injuries. If the injury occurs below this level, tendon transfer of functioning muscles is used to recreate the ability to grasp objects. At injury levels above C4, it is difficult to stabilize the arm. Shoulder and elbow usually retain some voluntary function in C5/6 spinal cord-injured people, but usually the arm is weak due to the paralysis of some key muscles.

The Bionic Glove [49] is a completely noninvasive FES device designed to activate hand muscles in C6/C7 spinal cord-injured persons who have some active wrist movement. It consists of self-adhesive surface electrodes which are placed above the motor points of the muscles to be stimulated and a glove which contains



a wrist position sensor and a box containing stimulator and control. Stimulation of the muscles that produce hand grasp is triggered by extending the wrist. Stimulation of the muscles that produce hand opening is triggered by flexing the wrist.

A clinically accepted example for an upper extremity neuroprosthesis for persons with C5/C6 spinal cord injury is the Freehand System [50] which consists of fully implanted electrodes and stimulator and an external control unit including an inductive coil and a shoulder position sensor: Eight epimysial electrodes control palmar and lateral hand grasp by neuromuscular stimulation of hand and arm muscles. Hand grasp is triggered through operation of an external joystick, controlled by the movement of the opposing nonparalyzed shoulder, which through a radiofrequency-powered and -controlled implanted stimulator delivers electrical stimulation. The Freehand system gives hundreds of persons the ability to feed and groom themselves; some can even operate a computer with their hand. In order to establish closed loop control of the Freehand system, nerve cuff electrodes have been used to record activity from cutaneous mechanoreceptors at the index finger to determine grip force.

## 11.9 Outlook

FES-based neuroprostheses for upper and lower extremities evoke lost movement functions in SCI subjects. Exercises such as FES cycling and rowing enable paraplegics to participate in mainstream activities and improve their health and fitness by exercising like able-bodied subjects. The completely external systems are relatively easy to don and handle, but the possibilities for control and selective muscle actuation are limited. Besides the physiological training, a main desire of the patients is to be able to move naturally as able-bodied subjects do. For use in daily living the neuroprosthesis should support the user in a cooperative way. It should respond to the moment-to-moment needs of the user without unduly distracting his attention. The user should be released of tasks that can be automatized but keep control over the movement coordination.

Recent developments in technology allow the design of miniaturized devices with sophisticated control systems. But still it is a challenge to detect what movement the patient desires to make and to transfer the signal proportionally into muscle force.

One crucial point is signal processing of biosignals that are recorded from brain and muscle activity to get a reliable and reproducible proportional command signal.

Currently, another limiting point are the electrodes because the correlation between stimulation and muscle force is difficult to predict. One major problem here is also fatigue which is a main issue in all FES applications.

At present, implanted systems use either batteries, disposable or rechargeable, or external AC power as power supply. Currently it is investigated whether biothermal power sources which use small temperature gradients in the body to create electrical power could make body powered implants possible (Biophan Technologies of West Henrietta).

Today the majority of spinal cord-injured subjects has an incomplete lesion. Consequently, there is a need for modular programmable systems that can be adapted to individual situations. FES with surface electrodes can be problematic for subjects with incomplete SCI, because when the sensory function is still intact the electrical field can evoke sensations of pain already at low stimulation intensities where only low muscle forces are generated.

More sophisticated systems will be available in the future, both external and implanted. Still besides technology also other personal factors have to be considered in the decision if a system is appropriate for an individual SCI subject or not. From a social point of view, patients should certainly not be dependent on using a neuroprosthesis for daily living. But each patient should have the freedom to choose using a neuroprosthesis for more independent daily living and self-responsible exercising for health and fitness.

Even if a cure of spinal cord injury should be possible in the future, those patients who have exercised their body and kept their musculoskeletal structures in shape will benefit first and most of it.

## References

1. W.T. Liberson, H.J. Holmquest, D. Scot, M. Dow, Stimulation of the peroneal nerve, synchronized with the swing phase of the gait of hemiplegic patients. *Arch. Phys. Med. Rehabil.* **42**, 101–105 (1961)
2. T.W.J. Janssen, R.M. Glaser, D.B. Shuster, Clinical efficacy of electrical stimulation exercise training: effects on health, fitness, and function. *Top. Spinal Cord Inj. Rehabil.* **3**, 33–49 (1998)
3. S. Malagodi, M.W. Ferguson-Pell, R.D. Masiello, A functional electrical stimulation exercise system designed to increase bone density in spinal cord injured individuals. *IEEE Trans. Rehab. Eng.* **TRE 1**, 213–219 (1993)
4. F. Rattay, Modeling the excitation of fibers under surface electrodes. *IEEE Trans. Biomed. Eng.* **BME 35**, 199–202 (1988)
5. M. Solomonow, External control of the neuromuscular system. *IEEE Trans. Biomed. Eng.* **BME 31**, 752–763 (1984)
6. W. Reichenfelser, M. Gföhler, T. Kakebeeke, P. Lugner, G. Feik, Determination of efficient stimulation patterns for FES – cycling; IFMBE Proc. EMBEC 05 Prague (2005)
7. W.K. Durfee, K.I. Palmer, Estimation of force-activation, force-length, and force-velocity properties in isolated, electrically stimulated muscle. *IEEE Trans. Biomed. Eng.* **BME 41**, 205–216 (1994)
8. M. Lawrence, G. Gross, M. Lang, A. Kuhn, T. Keller, M. Morari et al., Assessment of finger forces and wrist torques for functional grasp using new multichannel textile neuroprostheses. *Artif. Organs* **32**(8), 634–638 (2008)
9. T.G. McNaughton, K.W. Horch, Metallized polymer fibers as leadwires and intrafascicular microelectrodes. *J. Neurosci. Methods* **70**(1), 103–110 (1996)
10. A. Branner, R.B. Stein, R.A. Normann, Selective stimulation of cat sciatic nerve using an array of varying-length microelectrodes. *J. Neurophysiol.* **85**(4), 1585–1594 (2001)
11. N. Lago, K. Yoshida, K.P. Koch, X. Navarro, Assessment of biocompatibility of chronically implanted polyimide and platinum intrafascicular electrodes. *IEEE Trans. Biomed. Eng.* **54**(2), 281–290 (2007)
12. G.E. Loeb, R. Peck, W.H. Moore, K. Hood, BION system for distributed neural prosthetic interfaces. *Med. Eng. Phys.* **23**(1), 9–18 (2001)

13. J.M. Winters and S.L.-Y. Woo(eds), *Multiple Muscle Systems* (Springer Verlag, New York, 1990)
14. F.E. Zajac, Muscle and tendon: properties, models, scaling, and application to biomechanics and motor control. *Crit. Rev. Biomed. Eng.* **17**(4), 359–411 (1989) (Review)
15. M. Gföhler, J. Wassermann, P. Eser, T. Kakebeeke, H. Lechner, W. Reichenfeller et al., Muscle behavior in artificially activated muscle – measurements on neurologically intact and paraplegic subjects, in *International Society of Biomechanics XIXth Congress*, ed. by P. Milburn, B. Wilson, T. Yanai (International Society of Biomechanics, Dunedin, 2003), p. 5
16. R.D. Crowninshield, R.A. Brand, A physiologically based criterion of muscle force prediction in locomotion. *J. Biomech.* **14**(11), 793–801 (1981)
17. M.G. Pandy, F.E. Zajac, E. Sim, W.S. Levine, An optimal control model for maximum-height human jumping. *J. Biomech.* **23**(12), 1185–1198 (1990)
18. A. Seth, M.G. Pandy, A neuromusculoskeletal tracking method for estimating individual muscle forces in human movement. *J. Biomech.* **40**(2), 356–366 (2007)
19. A.D. Kuo, The relative roles of feedforward and feedback in the control of rhythmic movements. *Motor Control* **6**(2), 129–145 (2002)
20. G.C. Chang, J.J. Luh, G.D. Liao, J.S. Lai, C.K. Cheng, B.L. Kuo et al., A neuro-control system for the knee joint position control with quadriceps stimulation. *IEEE Trans. Rehab. Eng.* **5**(1), 2–11 (1997)
21. M. Ferrarin, F. Palazzo, R. Riener, J. Quintern, Model-based control of FES-induced single joint movements. *IEEE Trans. Neural Syst. Rehabil. Eng.* **TRE 9**(3), 245–257 (2001)
22. H. Park, D.M. Durand, Motion control of musculoskeletal systems with redundancy. *Biol. Cybern.* **99**(6), 503–516 (2008)
23. L.A. Bernotas, P.E. Crago, H.J. Chizeck, Adaptive control of electrically stimulated muscle. *IEEE Trans. Biomed. Eng.* **BME-34**(2), 140–147 (1987)
24. M. Goffredo, I. Bernabucci, M. Schmid, S. Conforto, A neural tracking and motor control approach to improve rehabilitation of upper limb movements. *J. Neuroeng. Rehabil.* **5**, 5 (2008)
25. L. Johnson, A.J. Fuglevand, Evaluation of probabilistic methods to predict muscle activity: implications for neuroprosthetics. *J. Neural Eng.* **6**(5), 55008 (2009)
26. J.L. Lujan, Crago PE automated optimal coordination of multiple-DOF neuromuscular actions in feedforward neuroprostheses. *IEEE Trans. Biomed. Eng.* **BME 56**(1), 179–187 (2009)
27. J.J.G. Hincapie, R.F. Kirsch, Feasibility of EMG-based neural network controller for an upper extremity neuroprosthesis. *IEEE Trans. Neural Syst. Rehabil. Eng.* **17**(1), 80–90 (2009)
28. D. Roetenberg, P.J. Slycke, P.H. Veltink, Ambulatory position and orientation tracking fusing magnetic and inertial sensing. *IEEE Trans. Biomed. Eng.* **54**(5), 883–890 (2007)
29. G.E. Loeb, R. Davoodi, The functional reanimation of paralyzed limbs. *IEEE Eng. Med. Biol. Mag.* **24**(5), 45–51 (2005)
30. M. Haugland, A. Lickel, J. Haase, T. Sinkjaer, Control of FES thumb force using slip information obtained from the cutaneous electroneurogram in quadriplegic man. *IEEE Trans. Rehabil. Eng.* **7**(2), 215–227 (1999)
31. K.J. Miller, M. denNijs, P. Shenoy, J.W. Miller, R.P. Rao, J.G. Ojemann et al., Real-time functional brain mapping using electrocorticography. *Neuroimage* **37**(2), 504–507 (2007)
32. G.R. Müller-Putz, R. Scherer, G. Pfurtscheller, R. Rupp, EEG-based neuroprosthesis control: a step towards clinical practice. *Neurosci. Lett.* **382**(1–2), 169–174 (2005)
33. G. Pfurtscheller, G.R. Müller, J. Pfurtscheller, H.J. Gerner, R. Rupp, ‘Thought’-control of functional electrical stimulation to restore hand grasp in a patient with tetraplegia. *Neurosci. Lett.* **351**(1), 33–36 (2003)
34. Y. Song, D. Borton, S. Park, W.R. Patterson, C.W. Bull, F. Laiwalla et al., Active microelectronic neurosensor arrays for implantable brain communication interfaces. *IEEE Trans. Neural Syst. Rehabil. Eng.* **17**(4), 339–345 (2009)
35. T.A. Thrasher, M.R. Popovic, Functional electrical stimulation of walking: function, exercise and rehabilitation. *Ann. Readapt. Med. Phys.* **51**(6), 452–460 (2008) (Epub 2008 Jun 18. Review. English, French)

36. G.P. Braz, M. Russold, R.M. Smith, G.M. Davis, Efficacy and stability performance of traditional versus motion sensor-assisted strategies for FES standing. *J. Biomech.* **42**(9), 1332–1338 (2009)
37. R. Davoodi, B.J. Andrews, Optimal control of FES-assisted standing up in paraplegia using genetic algorithms. *Med. Eng. Phys.* **21**(9), 609–617 (1999)
38. W. Reichenfelser, H. Hackl, S. Mina, S. Hanke, P. Lugner, M. Gföhler, Trainings- und measurement-system for FES-cycling, in “*IFESS 2008- from movement to mind*”, *Biomedizinische Technik*, vol. 53, Suppl.1 (Berlin, New York, 2008), pp. 265–267. ISSN: 0939–4990
39. R. Davoodi, B.J. Andrews, G.D. Wheeler, R. Lederer, Development of an indoor rowing machine with manual FES controller for total body exercise in paraplegia. *IEEE Trans. Neural Syst. Rehabil. Eng.* **TRE 10**(3), 197–203 (2002)
40. F. Ché, G.M. Davis, Cardiovascular and metabolic responses during functional electric stimulation cycling at different cadences. *Arch. Phys. Med. Rehab.* **89**(4), 719–725 (2008)
41. A.J.K. van Soest, M. Gföhler, R. Casius, Consequences of ankle joint fixation on FES cycling power output; a simulation study. *Med. Sci. Sport. Exercise* **1**, S.797–S.806 (2005)
42. A.V. Hill, The heat of shortening and dynamics constants of muscles. *Proc. R. Soc. Lond. B* (London: Royal Society) **126**(843): 136–195 (October 1938)
43. A.J. van Soest, L.J.R. Casius, The merits of a parallel genetic algorithm in solving hard optimization problems. *J. Biomech. Eng.* **125**:141–146 (2003).
44. M. Gföhler, P. Lugner, Dynamic simulation of FES-cycling: influence of individual parameters. *IEEE Trans. Neural Syst. Rehabil. Eng.* **12**(4), 398–405 (2004)
45. W. Reichenfelser, H. Hackl, J. Wiedner, J. Hufgard, K. Gstaltner, S. Mina, S. Hanke, M. Gföhler: Influence of FES cycling on spasticity in subjects with incomplete spastic paraplegia; in *Rehabilitation: Mobility, Exercise & Sports*, Assistive Technology Research Series, vol 26 (IOS Press, Amsterdam, 2010), pp. 317–319. ISBN: 978-1-60750-080-3
46. M. Kuchler, M. Gföhler, Development of a biomechanical model of the human body including the upper and the lower extremities used to simulate the motion on a rowing ergometer – the inverse dynamic problem, in *Congress Handbook and Book of Abstracts of the International Society of Biomechanics XIXth Congress, 4 pages* (2003), p. 122
47. I. Hauer, Instrumentierter Ruderergometer für Anwendung in Sport und Rehabilitation, Betreuer/in(nen), Konstruktionswissenschaften und Technische Logistik, ed. by M. Gföhler (2007) (Diploma Thesis, May 2007)
48. A.R. Kralj, T. Bajd, M. Muni, R. Turk, FES gait restoration and balance control in spinal cord-injured patients. *Prog. Brain. Res.* **97**, 387–396 (1993)
49. A. Prochazka, M. Gauthier, M. Wieler, Z. Kenwell, The bionic glove: an electrical stimulator garment that provides controlled grasp and hand opening in quadriplegia. *Arch. Phys. Med. Rehabil.* **78**, 608–614 (1997)
50. J.J. Pancrazio, P.H. Peckham, Neuroprosthetic devices: how far are we from recovering movement in paralyzed patients? *Expert Rev. Neurother.* **9**(4), 427–430 (2009)
51. M. Kuchler, M. Gföhler, Mechanical modeling and simulation of the human movement on a rowing ergometer. *Simul. News Eur.* **40**, S.10–S.19 (2004). ISSN 0929–2268
52. P.D. Faghri, R.M. Glaser, S.F. Ficoni, Functional electrical stimulation leg cycle ergometer exercise: training effects on cardiorespiratory responses of spinal cord injured subjects at rest and during submaximal exercise. *Arch. Phys. Med. Rehabil.* **73**(11), 1085–1093 (1992)
53. T. Mohr, J.L. Andersen, F. Biering-Sørensen, H. Galbo, J. Bangsbo, A. Wagner, M. Kjaer, Long-term adaptation to electrically induced cycle training in severe spinal cord injured individuals. *Spinal Cord* **35**(1), 1–16 (1997)
54. S. Jezernik, R.G. Wassink, T. Keller, Sliding mode closed-loop control of FES: controlling the shank movement. *IEEE Trans. Biomed. Eng.* **51**(2), 263–272 (2004)
55. D.M. Hettinga, B.J. Andrews, G.D. Wheeler, J.Y. Jeon, J. Verellen, J.J. Laskin, L.M. Olenik, R. Lederer, R. Burnham, R.D. Steadward, FES-rowing for persons with spinal cord injury, in *Proceedings of the 9th Annual Conference of the International FES Society September 2004*, Bournemouth, 2004

56. P.H. Veltink, P. Slycke, J. Hemssems, R. Buschman, G. Bultstra, H. Hermens, Three dimensional inertial sensing of foot movements for automatic tuning of a two-channel implantable drop-foot stimulator. *Med. Eng. Phys.* **25**(1), 21–28 (2003)
57. D.J. Weber, R.B. Stein, K.M. Chan, G. Loeb, F. Richmond, R. Rolf, K. James, S.L. Chong, BIONic WalkAide for correcting foot drop. *IEEE Trans. Neural Syst. Rehabil. Eng.* **13**(2), 242–246 (2005)