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## 9.1 Apoptosis as a Tumor Suppressor Mechanism

Mesothelioma remains an incurable cancer due to the ineffectiveness of conventional cytotoxic chemotherapy. This is reflected in the preponderance of mostly negative phase II clinical trials over the last 30 years [32]. Resistance to apoptosis is a hallmark of cancer in general [48], accounts for multidrug resistance [58], and is a signature of mesothelioma [31]. During tumorigenesis, it is now understood that as in common with other solid cancers, somatic genetic alteration is a frequent event predisposing to apoptosis resistance. These changes include the activation of oncogenic cell survival pathways, and the inactivation of tumor suppressors.

This chapter will focus on how apoptosis susceptibility in mesothelioma is, in general, inhibited by the acquisition of multiple somatic alterations in oncogenic and tumor suppressor

protein expression. Growing knowledge of these key genetic changes and their requirement for sustaining the malignant mesothelioma phenotype provide insights into potential vulnerabilities that may be successfully exploited using new therapeutic strategies. I will first of all, summarize our understanding of how the core death machinery is altered in mesothelioma (summarized in Fig. 9.1). This will be followed by a summary of the most frequent genetic alterations driving oncogenic pathways or leading to dysfunction of tumor suppressors (summarized in Fig. 9.2). Translational research opportunities arising from this knowledge of mesothelioma pathobiology will then be highlighted.

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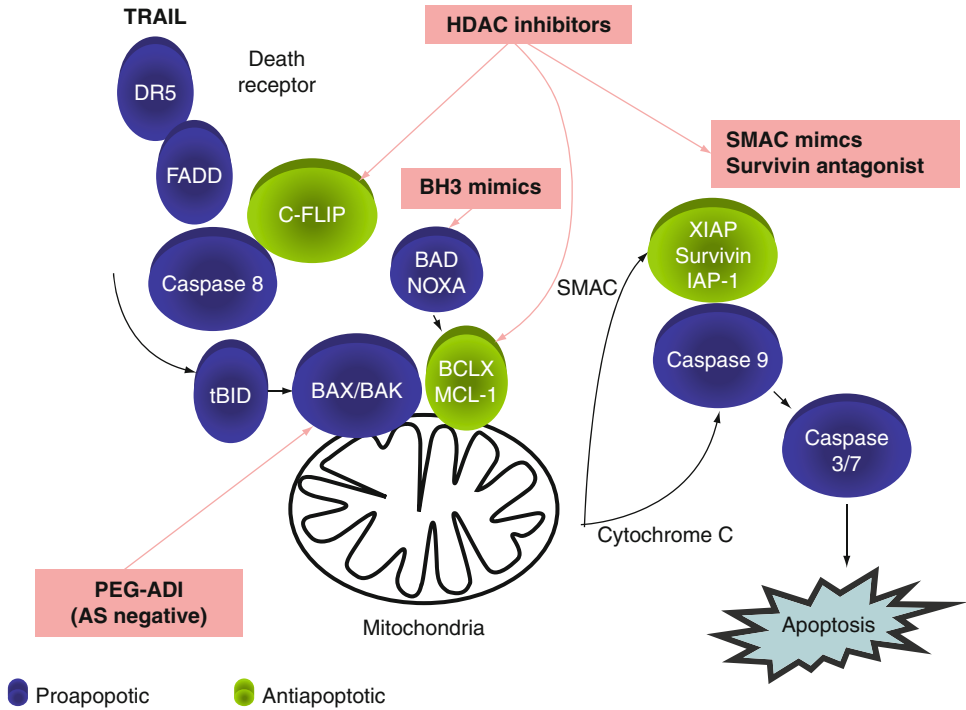
## 9.2 Key Alterations in the Core Apoptosis Signaling in Mesothelioma

### 9.2.1 Regulation of the Intrinsic (Mitochondrial) Apoptosis Pathway in Mesothelioma

The BCL-2 family of proteins constitutes the pivotal molecular regulators of the core cell death machinery. This family is subdivided into proapoptotic and antiapoptotic proteins. BCL-2, the prototypical member of the BCL-2 family

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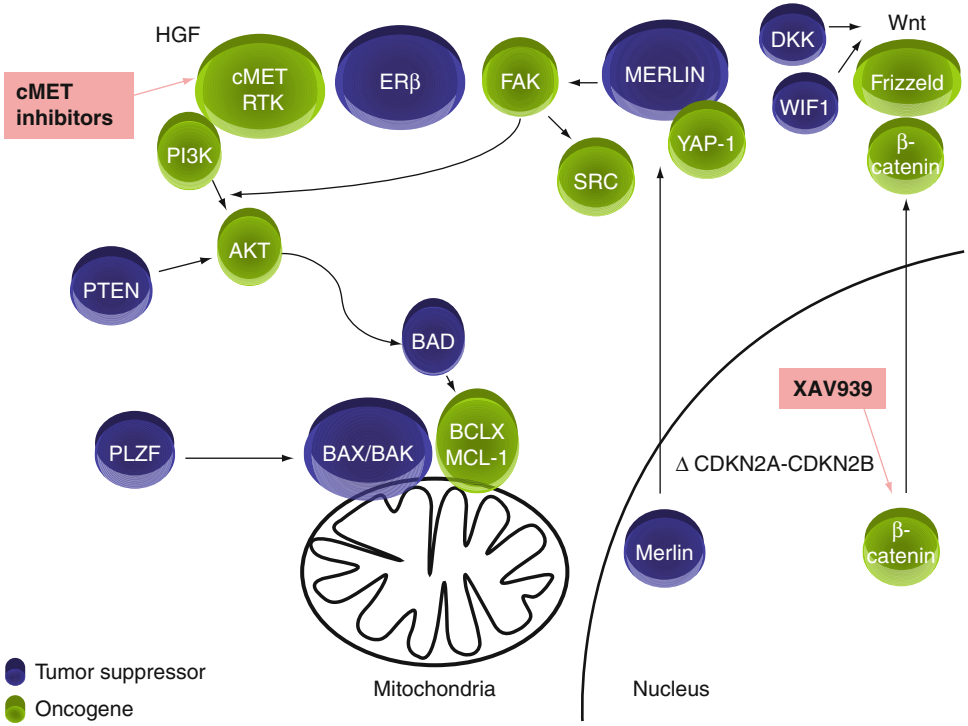
**Fig. 9.1** Altered regulation of the core apoptosis pathway in mesothelioma. Upregulated antiapoptotic proteins are highlighted (*green*)

was identified as a proto-oncogene associated with the t(14;18) translocation in follicular lymphoma [129]. The antiapoptotic protein subgroup now includes five additional proteins, MCL-1, BCL-X, BCL-W, A1 and BCL-B. Prosurvival BCL-2 family proteins regulate apoptosis at the level of the mitochondrial and endoplasmic reticulum outer membranes. The canonical cell death pathway involves mitochondria; organelles responsible for generating ATP, the cell's energy currency, through oxidative phosphorylation.

Prosurvival BCL-2 family proteins function to block a critical death switch which is responsible for making the all-or-none decision to commit a cell irreversibly to death [4,24]. This switch is the permeabilization of the outer mitochondrial membrane, induced by oligomerization and pore formation by the tumor suppressors and multidomain proapoptotic proteins BAK and BAX [99,115,146,147]. Mitochondrial outer membrane permeabilization or MOMP is a rapid,

kinetically invariant event that results in the release several proteins from the mitochondria into the cytosol. These proteins include cytochrome C [77], SMAC [27], OMI/HtrA2 [82] and apoptosis-inducing factor [122]. Cytochrome C in conjunction with APAF-1 [149] and dATP, triggers the activation of a family of zymogens called caspases, which cooperate in mediating cellular demolition by cleaving hundreds of substrates. Bax and Bak are genetically redundant tumor suppressors [140]; prosurvival BCL-2 proteins heterodimerize to prevent BAX and BAK activation, functioning as a rheostat that is dependent on the ratio of pro- to antiapoptotic proteins.

In common with other tumor suppressors, BAX deficiency has been identified in primary malignancies [84]. However, low *bcl-2/bax* ratio has been reported in mesothelioma cells despite their apoptosis resistance, implicating a mechanism other than BCL-2 in regulating apoptosis. In vivo, MCL-1 is more commonly



**Fig. 9.2** Key proteins involved in survival pathway signaling in mesothelioma (green). Tumor suppressors are shown in blue

expressed whereas BCL-2 expression is less frequent [96,119]. BAX and BAK require a subset of proapoptotic BCL-2 family proteins for activation which share homology in a death-inducing BCL-2 homology 3 (BH3) domain, but do not contain other BH domains. Two such BH3 domain-only proteins, BID [136] and BIM [95] can directly induce the oligomerization and activation of BAX. Interestingly, in mesothelioma, loss of expression of BH3-only proteins has been reported in vivo, namely, BID (37%) and BIM (18%). In addition, loss of BAX expression has been reported in one series in 42% of primary mesotheliomas [96].

Prosurvival BCL-2 family proteins are inhibited by a subset of BH3-only proteins, which are incapable of direct BAX/BAK activation, but bind directly to prosurvival counterparts. These so-called dissociator BH3-only proteins reflect a growing family and include BAD,

NOXA, PUMA, BMF, BIK and HRK. Because dissociator BH3-only proteins are endogenous prosurvival BCL-2 family inhibitors, they represent a prototype for small molecule drug development, most notably ABT737 [76,98]. BH3 mimetics are a promising class of mitochondria targeted therapy with potential for treating mesothelioma. This is suggested by studies in which silencing BCL-2 and BCL-XL was sufficient to induce apoptosis and chemosensitization [52]. However, target specificity is likely to be important for therapeutic efficacy. MCL-1 is highly expressed in mesothelioma and is one of the most commonly amplified oncogenes in human cancer [9]. It is also a resistance biomarker for ABT737 [68,132]. Nevertheless, other prosurvival BCL-2 family targeted agents such as obatoclax [92] are currently in clinical development, and may exhibit efficacy in mesothelioma.

### 9.2.2

#### Extrinsic Apoptosis Pathway Regulation in Mesothelioma

Apoptosis can be efficiently induced in mesothelioma cell lines by ligation of cell surface death receptors. Activation of death receptors by their ligands (which include TNF and FAS) leads to recruitment of FADD through a conserved death domain [22], followed by recruitment of caspase 8 [10] activating complex known as the death-inducing signaling complex or DISC [127]. Caspase 8 cleaves BID, leading to activation of BAX/BAK, mitochondrial apoptosis, and therefore signal amplification [78]. Tumor necrosis factor-related apoptosis inducing ligand (TRAIL) or TRAIL receptor agonists are currently in clinical development but have yet to be evaluated in mesothelioma. Upon interaction, its receptors (TRAIL R1 or R2) can induce apoptosis *in vitro*. TRAIL also synergizes with DNA damage induced by etoposide in a manner that requires c-jun N terminal kinase [135].

FLIP is an inhibitor of TRAIL-induced apoptosis and is recruited to the DISC [90], where it inhibits caspase 8 recruitment and activation. Mesothelioma cells overexpress FLIP resulting in inhibition of death receptor-induced apoptosis [112]. Silencing of FLIP in mesothelioma and other cancer models re-establishes sensitivity to TRAIL [112,141]. Novel, clinically applicable approaches for downregulating FLIP in the clinical setting will be highlighted later in this chapter.

### 9.2.3

#### Inhibitors of Apoptosis in Mesothelioma

Inhibitors of apoptosis (IAPs) comprise a family of structurally related proteins, which share a common 70 amino acid baculovirus IAP (BIR) repeat. IAPs interact with and inhibit caspases 9, 3 and 7. Mesotheliomas have been shown to overexpress the IAPs survivin, XIAP and IAP-1

*in vivo* using immunohistochemistry [43,65]. IAP-1 has been shown to be associated with shorter survival [44]. RNAi-mediated silencing of IAP-1 is sufficient to reduce mesothelioma cell viability and induce apoptosis by activating the mitochondrial pathway [43]. Conversely, IAP-1, IAP-2 and XIAP are upregulated by tumor necrosis factor alpha, whereas survivin and livin are not [45]. Survivin is overexpressed in mesothelioma and its silencing *in vitro* is associated with induction of apoptosis suggesting that it might be a potential molecular target [29,144,152].

IAP proteins are inhibited by Smac, which is released from the mitochondria following outer membrane permeabilization by BAX/BAK. Small molecule smac mimetics offer one way of targeting IAPs and are currently in early development, for example, AT406 and TL32711; these compounds also downregulate IAP-1 and IAP-2 [137]. Selective inhibitors of survivin, for example, YM155 are currently in clinical development in other cancers. Other approaches capable of modulating IAP proteins include histone deacetylase inhibition, which is discussed in more detail later in this chapter.

## 9.3

### Tumor Suppressor Loss in Mesothelioma

#### 9.3.1

##### Loss of nf2 Is Frequent in Mesothelioma

The short arm of chromosome 9 (9p) is a region associated with frequent cytogenetic abnormalities in mesothelioma [19,21,91,100,125]. Loss of the CDKN2b-CDKN2a locus on chromosome 9p21 in humans is a common event in cancer, in general, including mesothelioma. This locus includes the tumor suppressor p16ink4a, which is encoded by CDKN2A and is one of the most frequently silenced tumor suppressors in mesothelioma [53]. This tumor suppressor is an inhibitor of the Rb1 pathway

involved in cell cycle progression, and its loss whether by deletion (75–85%) or methylation is associated with poor prognosis [67,72]. There is frequently co-deletion of p16ink4a and p15ink4b, occurring in 75% of mesotheliomas [145]. It has been recently shown that p15ink4b, which is encoded by CDKN2b, can substitute for loss of p16ink4a, and that this back-up function could account for the frequently observed loss of the complete CDKN2b-CDKN2a locus [70].

Loss of expression has been shown to be associated with homozygous deletion of exons 1–3 [91,102], and this is more frequently associated with exposure to asbestos even in non-small cell lung cancer, compared with tobacco exposure (which is associated with hypermethylation) [3]. In mesothelioma, hypermethylation occurs in the first exon [142]. Re-expression in mesothelioma cells is sufficient to induce cell cycle arrest, as well as reduced tumor growth and spread in vivo [40,41]. Because hypermethylation silences p16ink4a in approximately 20% of mesotheliomas [142], re-expression can be achieved using demethylating agents such as cytidine analog dihydro-5-azacytidine (DHAC). Analysis of tissue samples from CALGB 8833 and 9031 clinical trials employing DHAC-based therapy identified 4/20 tumors with methylation of p16ink4a. Although there was a trend to improved survival in this clinical trial associated with p16ink4a methylation, this was not statistically significant, probably as a result of the small sample size [69].

Around 40% mesotheliomas harbor somatic mutations in the neurofibromatosis type 2 gene (NF2) located at chromosome 22q12 [116]. Treatment of Nf2 ( $\pm$ ) knockout mice with asbestos causes accelerated development of mesothelioma, with biallelic inactivation of the wild-type Nf2 allele, and loss of the CDKN2A locus [1]. Conditional knockout of nf2/p16ink4a in a murine model has been shown to exhibit more invasive, aggressive mesothelioma compared with conditional nf2/p53 knockout, with shorter survival [59]. Together, this implicates an

important role in mesothelioma [59]. Mutation of NF2 is frequent in mesothelioma but not observed in non-small cell lung cancer [116]. Somatic mutation of NF2 is conserved across mesothelioma in different species, being frequently detected in murine mesothelioma [73].

### 9.3.2

#### NF2 Encodes the Tumor Suppressor Merlin

Merlin, the gene product of NF2 is a FERM domain protein that functions at the plasma membrane where it inhibits mitogenic signaling. It functions as a growth inhibitor, and accumulates in the nucleus where it interacts with and inhibits the E3 ligase CRL4 (DCAF1) [79]. Loss of merlin has a pro-mitogenic effect, and this is lost when DCAF1 is depleted, or if a merlin insensitive mutant is expressed. Mutations of merlin disrupt the direct interaction with CRL4(DCAF1).

When Merlin expression is restored in NF2 deficient mesothelioma cells, there is a marked inhibition of cell motility, spreading and invasiveness. Focal adhesion kinases (FAK) play a critical role in regulating invasive phenotype, and are negatively targeted by merlin. This mechanism of inhibition involves merlin dependent FAK phosphorylation at a critical residue on tyrosine 397, resulting in a block of its interaction with binding partners src and the PI3kinase regulatory subunit p85 [106].

The transcriptional coactivator YAP1 [88] is an oncogene that is commonly amplified at the 11q22 locus in mesotheliomas, and physically interacts with merlin, contributing to the promitogenic effects of NF2 deletion [148]. RNAi-mediated suppression of YAP1 suppresses growth of mesothelioma cells with NF2 homozygous deletion through induction of apoptosis and cell cycle arrest. Conversely, overexpression of YAP1 in immortalized mesothelioma cells is mitogenic. Merlin inhibits YAP1 through the induction of its phosphorylation and cytoplasmic retention.

### 9.3.3

#### PLZF Is a Novel Tumor Suppressor in Mesothelioma

Focal deletion of 11q23 has been identified in mesothelioma, and involves a locus encompassing promyelocytic leukemia zinc finger (PLZF), a transcriptional repressor gene. Loss of PLZF confirmed by analysis of transcript levels, and loss of protein expression has been observed in mesothelioma compared with mesothelial cells. Ectopic expression of PLZF causes reduced clonogenicity and initiation of apoptosis involving caspase activation; together, with the loss of PLZF implicates a potentially important role in regulating mesothelioma cell survival.

## 9.4

### Therapeutic Inhibition of Survival Pathways

#### 9.4.1

##### PI3K/AKT/mTOR Axis in Mesothelioma

Mesothelioma cells, which have been grown in three dimensions to more closely resemble solid tumors, acquire multidrug resistance, including resistance to TRAIL and chemotherapy [6,62]. The molecular basis underlying acquisition of multidrug resistance has not been fully delineated, but involves activation of the phosphatidylinositol-3-kinase (PI3K)/Akt/mammalian target of rapamycin (mTOR) pathway since rapamycin or RNAi silencing of the mTOR target, S6K, can restore TRAIL sensitivity. This effect requires BID, since silencing using RNAi implicates mTOR/S6K as a major contributor of resistance to TRAIL in three-dimensional but not two-dimensional tumors. TRAIL sensitivity is also enhanced by inhibition of the PI3K/AKT pathway following heat stress, supporting a role for this pathway in blocking apoptosis [104]. Mesotheliomas exhibit an elevated level of activity in the PI3K/AKT/mTOR pathway both in mouse and human models, and its inhibition

is associated with potentiation of cisplatin-induced apoptosis [2].

AKT is antagonized by the endogenous inhibitor, phosphatase and tensin analog (PTEN), which acts to inhibit phosphorylation. When overexpressed in mesothelioma, PTEN induces loss of viability [87]. Not surprisingly, given the survival function of PTEN in regulating PI3K/AKT/mTOR signaling, the expression is lost in a significant proportion of mesotheliomas [101]. Recently it has been shown that PTEN is required for maintaining the integrity of chromosomes [117]. Loss of PTEN confers a defect in homologous repair which can be exploited by inhibition of poly ADP ribose polymerase (PARP) [85]. Given the recent evidence that PARP inhibitors are very effective in inducing tumor responses under conditions of defective DNA double strand break repair due to BRCA1 mutation [30,37], the possibility exists that a subset of PTEN deficient mesotheliomas may be sensitive to PARP inhibitors.

#### 9.4.2

##### HGF/cMET Pathway Is Activated in Mesothelioma

C-met receptor tyrosine kinase is overexpressed in mesothelioma by 82% compared with normal tissues, and in 90% of serous effusions [153]. It is associated with high circulating levels of its ligand scatter factor/HGF [57], which in turn is overexpressed in 40–85% of mesotheliomas. HGF stimulates mesothelioma cell motility *in vitro* via the c-met receptor [49,50,66,128], and has been shown to mediate cell survival by upregulating BCL-XL. The mechanism involves mitogen-activated protein kinase-dependent phosphorylation and activation of the ETS family of transcription factors, which bind to the promoter of BCL-XL [17]. Because phosphorylated c-met and BCL-XL expression are correlated *in vivo*, it has been proposed that the HGF/met axis mediates survival in part through this interaction at the transcriptional level [17].

The early-response proto-oncogene, *fos*-related antigen or *fra-1* transcriptionally regulates *c-met* and is upregulated in preclinical models of mesothelioma, as evidenced by expression microarray analysis [109]. Accordingly, HGF-dependent phosphorylation is inhibited by *Fra-1* silencing [111]. *Fra-1* is a component of the dimeric transcription factor, activator protein-1 or AP-1 and is regulated by phosphatidylinositol-3-kinase, extracellular signal-regulated kinases ERK1 and 2, and Src-associated pathways [110]. In addition to *c-met* being a target of *Fra-1*, it also directly regulates expression of CD44, the predominant hyaluronic receptor in mesothelioma expression, and thus potentially contributes to control of migration and invasive behavior.

The small molecule *c-met* inhibitors SU 11274 or PHA-665752, as well as RNAi silencing of *c-met*, inhibits migration of mesothelioma cells. Susceptibility to *c-met* inhibition has been reported to depend on the presence of a Met/HGF autocrine loop as evidenced by PHA-665752 [89]. Specific *c-met* mutations have been identified in two domains; N375S, M431V, and N454I mutations in the semaphorin domain; T1010I and G1085X in the juxtamembrane domain. Interestingly, two mesothelioma cell lines H513 and H2596, which harbor the T1010I mutation, are highly sensitive to SU11274. In addition to *c-met* mutations, deletion of exon 10 resulting in a splice variant of *c-met* has been identified in some mesothelioma specimens.

Although activation of the epidermal growth receptor family is observed in mesothelioma, activating mutations of the epidermal growth factor receptor (EGFR) have not been identified in patients with mesothelioma [134]. Targeting EGFR alone in mesothelioma cells has little effect, whereas simultaneous targeting of *c-met* and EGFR is associated with strong inhibition of proliferation and invasion, suggesting that blocking the coactivation of these two pathways may be more effective than targeting *c-met* alone [60].

### 9.4.3

#### WNT Pathway Activation in Mesothelioma

The Wnt signaling pathways play an important role in homeostasis and development [39]. It suppresses apoptosis through activation of beta-catenin/Tcf-mediated transcription, and is constitutively activated in mesothelioma cells [130]. The canonical Wnt signaling pathway cooperates with loss of NF2 to promote the loss of contact inhibition during proliferation [12]. Gene expression analysis of rat peritoneal mesothelioma induced by *o*-nitrotoluene or bromochloroacetic acid demonstrates an upregulation of the Wnt/beta-catenin pathway compared with non-transformed mesothelial cells [63]. Using Wnt specific microarray analysis of normal pleura versus mesothelioma, Wnt2 upregulation has been found to be the most common event in mesothelioma [83]. Knockdown of Wnt using RNAi or anti-Wnt2 antibody is sufficient to induce apoptosis, suggesting that Wnt2 could be a potential molecular target [83].

The beta-catenin gene is deleted at 3p21.3 in NCI-H28 cell line [14,118], and this model has been useful in determining the role of beta-catenin-independent Wnt signaling in mesothelioma, via the so-called noncanonical pathway. Wnt inhibitory factor (WIF-1) is a secreted protein that inhibits Wnt signaling and is downregulated in mesotheliomas compared with adjacent pleura [8]. The mechanism of downregulation involves promoter hypermethylation which is seen in malignant, but not adjacent normal pleural tissue. This suggests that epigenetic silencing of WIF-1 could be an important mechanism driving Wnt activation [8]. Similarly, RNAi-mediated knockdown has been shown to suppress cell growth, and colony formation [131]. Secreted Frizzled-related proteins (SFRPs) and the secreted protein dickopf-1 (*Dkk-1*) are negative regulators of Wnt signaling. SFRPs are silenced by promoter hypermethylation in mesothelioma [74] and re-expression of SFRP4 or *Dkk-1* is sufficient to block Wnt signaling in

beta-catenin deficient mesothelioma cells. This implicates a beta-catenin-independent, noncanonical Wnt pathway as a key regulator of cell survival in mesothelioma [51,75,150].

Given the potential importance of Wnt in maintaining mesothelioma cell survival, as well as other cancers (e.g., 80% of colorectal cancers are driven by Wnt mutations [42]), targeting Wnt is a promising strategy. However, no agents have yet entered clinical development. This is because drugging the Wnt pathway has proved difficult. Nevertheless, some small molecules have been identified with the potential to become experimental agents for future clinical studies [20,81]. One promising, but alternative strategy has been to target beta-catenin-mediated transcription. The small molecule XAV939 has been identified by genetic screening. It induces degradation of beta catenin via mechanism involving inhibition of the poly-ADP ribosylating enzymes tankyrase 1 and 2 [54]. This approach might provide a novel strategy for targeting the Wnt pathway in mesothelioma and other cancers.

#### 9.4.4

#### Estrogen Receptor Beta

Female gender is associated with a favorable prognosis and estrogen receptor beta (ER beta) has been previously shown to be lost in other cancers. This loss is associated with poor prognosis, implicating ER beta as a putative tumor suppressor [7,120]. In mesothelioma, ER beta is downregulated in tumor tissues compared with normal pleura, whereas ER alpha is not expressed [105]. ER beta was recently shown to be an independent prognostic factor for better survival. Activation of ER beta in vitro with 17 beta-estradiol reduces cell proliferation associated with G2/M cell cycle arrest, downregulation of p27, p21, and survivin. These findings suggest that selective estrogen receptor modulators may have a potential role in controlling mesotheliomas.

## 9.5

### Therapeutic Reactivation of Tumor Suppressors

#### 9.5.1

#### Epigenomic Dysregulation in Mesothelioma

Transformation of normal mesothelium into mesothelioma involves changes to the epigenome. In a study interrogating 1505 CpG loci associated with 803 cancer-associated genes in 158 mesothelioma specimens and 18 normal pleura, the methylation profile was able to effectively discriminate normal pleura from mesothelioma, and was an independent predictor of shorter survival [23]. In an independent study that examined 6157 CpG islands in 20 mesotheliomas in parallel with comparative genomic hybridization and chromatin immunoprecipitation arrays [47], 6.3% of genes were found to be hypermethylated in mesothelioma including MAPK13, KAZALD1, and TMEM30B; 11% of heterozygously deleted genes were affected by DNA methylation and/or H3K27me3. Furthermore, a group of genes silenced by histone H3 lysine 27 methylation (H3K27me3) could be reactivated by histone deacetylation.

Combined epigenetic alterations in mesothelioma are linked with poor prognosis, and these epigenetic alterations may interact cooperatively. In a study, which used nested methylation specific PCR to interrogate the promoter methylation status of nine genes from serum DNA, high incidence of methylation of E-cadherin (71.4%) and FHIT (78%) [36] was measured, whereas intermediate methylation is associated with p16(INK4a) (28.2%), APC1B (32.5%), p14(ARF) (44.2%), and RARbeta (55.8%). Low methylation frequencies were seen for ACP1A (14.3%), RASSF1A (19.5%), and DARK (20%). Interestingly, although no single gene alone predicted survival, combination of RARbeta with either RASSF1A or DARK was associated with significantly shorter survival. This implicates



that silencing of multiple genes can cooperate to influence prognosis in contrast to the effects of these single genes alone.

MicroRNAs are associated with epigenetic regulation. In a study in which 98 mesothelioma specimens were studied using a custom microRNA platform, a training set of 44 tumors and a test set of 98 tumors were analyzed [103]. The microRNA, hsa-miR-29c was shown to be a favorable independent predictor of time to progression and survival after surgical cytoreduction, and was selectively overexpressed in the epithelioid histological subtype. Overexpression of hsa-miR-29c in cell lines was associated with a reduction in clonogenicity associated with reduced proliferation, as well as invasiveness and motility. Epigenetic regulation by hsa-miR-29c was evidenced by its downregulation of DNA methyltransferases and upregulation of demethylating genes, suggesting its role as a prognostic biomarker could relate to its ability to depress transcription of tumor suppressors.

### 9.5.2

#### Targeting the Mesothelioma Epigenome via Inhibition of Histone Deacetylases

Histone deacetylases (HDACs) are a class of enzymes that repress genes by inhibiting transcription. As such, they function opposite to histone acetyltransferase which promotes transcription. HDACs remove acetyl groups from  $\epsilon$ -*N*-acetyl lysine amino acid on a histone; the effect is to remove the positive charge required for electrostatic interaction with the negatively charged phosphate/DNA backbone, leading to remodeling of chromatin (also termed chromatin expansion), resulting in increased transcription.

HDACs can be selectively inhibited by small molecules [35], and are an active molecular target for clinical development. Mesothelioma cells are sensitive to HDAC inhibition, which can directly modify signaling through the core apoptosis pathway; HDAC inhibition, for example, by sodium

butyrate [15,114], causes the downregulation of BCL-XL and induces apoptosis [16]. XIAP is downregulated by HDAC inhibition, and results in increased apoptosis when mesothelioma cells are treated with TRAIL [123]. The HDAC inhibitor Panobinostat (LBH589) is active against mesothelioma cell lines and xenografts [25]. Using a mouse model of B cell lymphoma to explore the proapoptotic pharmacodynamics of vorinostat (suberoylanilide hydroxamic acid or SAHA), the BH3- only proteins BID and BIM were identified as key regulators of intrinsic apoptosis signaling [80]. HDAC inhibition directly downregulates FLIP [18,86,126], with potential to synergize with death receptor agonists [18].

Valproate is an HDAC inhibitor, and has been shown to synergistically interact with cisplatin and pemetrexed in both cell lines, and a xenograft model of mesothelioma [133]. In cells, its cytotoxic activity is associated with activation of both the extrinsic apoptosis pathway, and the intrinsic pathway. Hyperacetylation of histone H3 is induced by valproate consistent with its pharmacodynamics as an HDAC inhibitor. Induction of cell death involves the generation of reactive oxygen species; accordingly, cells can be rescued by the antioxidant *N*-acetylcysteine.

HDAC inhibition may be a promising new development in the treatment of mesothelioma. Although a phase II trial of belinostat (PXD101) which targets class I and II HDACs was shown to be inactive [108], vorinostat exhibited significant activity in a phase I trial, in which monotherapy achieved partial responses [71]. A randomized phase II/III comparing oral vorinostat versus placebo is currently enrolling patients who have relapsed following first line therapy [143]. Given the lack of standard therapy in this clinical setting, this large randomized trial has potential to change practice if it is positive. Recent evidence implicates HR23B as a resistance biomarker of HDAC inhibitors, albeit in cutaneous T cell lymphoma, an indication for which vorinostat has received FDA approval. HR23B shuttles ubiquitinated proteins to the

proteasome. Loss of expression confers resistance to HDAC inhibitors as originally identified by genome-wide RNAi screen. As such, HR23B may represent a potential biomarker for vorinostat in other indications such as treatment of mesothelioma [38,61,113,121,138,151].

### 9.5.3

#### Targeting the Ubiquitin Proteasome Pathway

Protein degradation is an essential cellular process which involves tagging with ubiquitin by enzymes called ubiquitin ligases. Proteins are then ferried to the proteasome where degradation to peptides occurs. Small molecule proteasome inhibitors such as bortezomib (velcade) activates BCL-2 family tumor suppressors, leading to induction of apoptosis [33]. These include myc-dependent upregulation of the MCL-1 inhibitor NOXA [34,93,107,139], and other BH3 only proteins such as BIK and BIM [94]. Gene expression studies have implicated dysregulation of the ubiquitin proteasome pathway in mesothelioma [11], and preclinical studies have demonstrated proapoptotic efficacy of proteasome inhibitors in vitro and in vivo [46,113,121,138,151]. This promising activity has led to completion of phase II trials of bortezomib in mesothelioma; EORTC 08052 exploring combination with cisplatin in the first-line setting, and bortezomib monotherapy in the relapsed setting. Mutation and overexpression of proteasome subunit B5 (PSMB5) has been previously identified as a cause of resistance to bortezomib. However, the existence of such mutations in mesothelioma has not yet been established [97].

### 9.6

#### Synthetic Lethal Strategies

Mutation of a putative tumor suppressor gene may expose vulnerabilities in a cancer that can be exploited therapeutically. This has been most

dramatically demonstrated in the case of somatic BRCA1/BRCA2 mutations, which through inactivation of DNA repair render cancers vulnerable to DNA damage resulting from PARP inhibition [30,85]. Two examples of synthetic lethality associated with dysfunctions in tumor metabolism in mesothelioma will now be considered, where loss of function due to genetic or epigenetic alterations may be exploited, with translation into the clinical setting.

Homozygous codeletion of CDKN2A is frequently associated (90%) with loss of methylthioadenosine phosphorylase (MTAP) [55]. MTAP deficient tumors are responsive to inhibitors of de novo AMP synthesis in the preclinical setting, suggesting a strategy for mediating synthetic lethality. In a multicenter phase II trial to test this concept, patients with MTAP deficient tumors including mesothelioma (as well as non-small cell lung cancer, soft tissue sarcoma, osteosarcoma or pancreatic cancer) were treated with L-alanosine at a dose of 180 mg/m<sup>2</sup> by continuous intravenous infusion daily for 5 out of 21 days. However, no objective responses to therapy were observed leading the investigators to conclude a lack of efficacy [64].

The gene encoding argininosuccinate synthetase (AS), a rate-limiting enzyme involved in arginine metabolism is epigenetically silenced in mesotheliomas, implicating it as a tumor suppressor and highlighting a potential vulnerability which may be exploited therapeutically [26]. AS was shown to be downregulated both in mesothelioma cell lines and a high proportion (63%) of primary mesothelioma specimens [124]. Cell lines lacking AS were unable to synthesize arginine following depletion of arginine from the medium, and underwent apoptosis associated with activation of BAX and mitochondrial depolarization. Silencing of AS was associated with gene methylation.

Induction of apoptosis in AS negative cells following withdrawal of arginine is selective, and not observed in AS positive cell lines, reflecting arginine auxotrophy of AS deficient

cells. Accordingly, lack of AS presents a potential metabolic Achilles' heel in mesothelioma. This phenotype can be targeted pharmacologically, by removing arginine from the circulation using pegylated arginine deiminase, an agent that has received orphan drug status from the FDA for the treatment of hepatocellular carcinoma, and has shown efficacy in melanoma [5,13,28,56]. Because of the high frequency of AS deficiency in mesothelioma, a phase II trial will be evaluating this strategy in patients, tailoring treatment to patients with AS negative mesothelioma [26,124].

## 9.7

### Summary

In recent years, it has become clear that mesothelioma is characterized by frequent activation of survival pathways and inactivation of tumor suppressors. This has opened the door to a growing number of new, rational treatment strategies for targeting vulnerabilities in mesothelioma, that for the first time have real potential for significantly improving treatment response in this chemoresistant cancer, and improving survival outcomes, particularly in the relapsed setting where it is still an unmet clinical need.

### References

1. Altomare DA, Vaslet CA, Skele KL, De Rienzo A, Devarajan K, Jhanwar SC, McClatchey AI, Kane AB, Testa JR (2005) A mouse model recapitulating molecular features of human mesothelioma. *Cancer Res* 65(18):8090–8095
2. Altomare DA, You H, Xiao GH, Ramos-Nino ME, Skele KL, De Rienzo A, Jhanwar SC, Mossman BT, Kane AB, Testa JR (2005) Human and mouse mesotheliomas exhibit elevated AKT/PKB activity, which can be targeted pharmacologically to inhibit tumor cell growth. *Oncogene* 24(40):6080–6089
3. Andujar P, Wang J, Descatha A, Galateau-Salle F, Abd-alsamad I, Billon-Galland MA, Blons H, Clin B, Danel C, Housset B et al (2010) p16INK4A inactivation mechanisms in non-small-cell lung cancer patients occupationally exposed to asbestos. *Lung Cancer (Amsterdam, Netherlands)* 67(1):23–30
4. Antonsson B, Conti F, Ciavatta A, Montessuit S, Lewis S, Martinou I, Bernasconi L, Bernard A, Mermod JJ, Mazzei G et al (1997) Inhibition of Bax channel-forming activity by Bcl-2. *Science (New York)* 277(5324):370–372
5. Ascierto PA, Scala S, Castello G, Daponte A, Simeone E, Ottaviano A, Beneduce G, De Rosa V, Izzo F, Melucci MT et al (2005) Pegylated arginine deiminase treatment of patients with metastatic melanoma: results from phase I and II studies. *J Clin Oncol* 23(30):7660–7668
6. Barbone D, Yang TM, Morgan JR, Gaudino G, Broaddus VC (2008) Mammalian target of rapamycin contributes to the acquired apoptotic resistance of human mesothelioma multicellular spheroids. *J Biol Chem* 283(19):13021–13030
7. Batistatou A, Kyzas PA, Goussia A, Arkoumani E, Voulgaris S, Polyzoidis K, Agnantis NJ, Stefanou D (2006) Estrogen receptor beta (ERbeta) protein expression correlates with BAG-1 and prognosis in brain glial tumours. *J Neurooncol* 77(1):17–23
8. Batra S, Shi Y, Kuchenbecker KM, He B, Reguart N, Mikami I, You L, Xu Z, Lin YC, Clement G et al (2006) Wnt inhibitory factor-1, a Wnt antagonist, is silenced by promoter hypermethylation in malignant pleural mesothelioma. *Biochem Biophys Res Commun* 342(4):1228–1232
9. Beroukhim R, Mermel CH, Porter D, Wei G, Raychaudhuri S, Donovan J, Barretina J, Boehm JS, Dobson J, Urashima M et al (2010) The landscape of somatic copy-number alteration across human cancers. *Nature* 463(7283):899–905
10. Boldin MP, Goncharov TM, Goltsev YV, Wallach D (1996) Involvement of MACH, a novel MORT1/FADD-interacting protease, in Fas/APO-1- and TNF receptor-induced cell death. *Cell* 85(6):803–815
11. Borczuk AC, Cappellini GC, Kim HK, Hesdorffer M, Taub RN, Powell CA (2007) Molecular profiling of malignant peritoneal mesothelioma identifies the ubiquitin-proteasome pathway as a therapeutic target in poor prognosis tumors. *Oncogene* 26(4):610–617
12. Bosco EE, Nakai Y, Hennigan RF, Ratner N, Zheng Y (2010) NF2-deficient cells depend on the Rac1-canonical Wnt signaling pathway to

promote the loss of contact inhibition of proliferation. *Oncogene* 29(17):2540–2549

13. Bowles TL, Kim R, Galante J, Parsons CM, Virudachalam S, Kung HJ, Bold RJ (2008) Pancreatic cancer cell lines deficient in argininosuccinate synthetase are sensitive to arginine deprivation by arginine deiminase. *Int J Cancer* 123(8):1950–1955
14. Calvo R, West J, Franklin W, Erickson P, Bemis L, Li E, Helfrich B, Bunn P, Roche J, Brambilla E et al (2000) Altered HOX and WNT7A expression in human lung cancer. *Proc Natl Acad Sci USA* 97(23):12776–12781
15. Candido EP, Reeves R, Davie JR (1978) Sodium butyrate inhibits histone deacetylation in cultured cells. *Cell* 14(1):105–113
16. Cao XX, Mohuiddin I, Ece F, McConkey DJ, Smythe WR (2001) Histone deacetylase inhibitor downregulation of bcl-xl gene expression leads to apoptotic cell death in mesothelioma. *Am J Respir Cell Mol Biol* 25(5):562–568
17. Cao X, Littlejohn J, Rodarte C, Zhang L, Martino B, Rascoe P, Hamid K, Jupiter D, Smythe WR (2009) Up-regulation of Bcl-xl by hepatocyte growth factor in human mesothelioma cells involves ETS transcription factors. *Am J Pathol* 175(5):2207–2216
18. Carlisi D, Lauricella M, D'Anneo A, Emanuele S, Angileri L, Di Fazio P, Santulli A, Vento R, Tesoriere G (2009) The histone deacetylase inhibitor suberoylanilide hydroxamic acid sensitises human hepatocellular carcinoma cells to TRAIL-induced apoptosis by TRAIL-DISC activation. *Eur J Cancer* 45(13):2425–2438
19. Center R, Lukeis R, Dietzsch E, Gillespie M, Garson OM (1993) Molecular deletion of 9p sequences in non-small cell lung cancer and malignant mesothelioma. *Genes Chromosomes Cancer* 7(1):47–53
20. Chen B, Dodge ME, Tang W, Lu J, Ma Z, Fan CW, Wei S, Hao W, Kilgore J, Williams NS et al (2009) Small molecule-mediated disruption of Wnt-dependent signaling in tissue regeneration and cancer. *Nat Chem Biol* 5(2):100–107
21. Cheng JQ, Jhanwar SC, Klein WM, Bell DW, Lee WC, Altomare DA, Nobori T, Olopade OI, Buckler AJ, Testa JR (1994) p16 alterations and deletion mapping of 9p21-p22 in malignant mesothelioma. *Cancer Res* 54(21):5547–5551
22. Chinnaiyan AM, O'Rourke K, Tewari M, Dixit VM (1995) FADD, a novel death domain-containing protein, interacts with the death domain of Fas and initiates apoptosis. *Cell* 81(4):505–512
23. Christensen BC, Houseman EA, Godleski JJ, Marsit CJ, Longacker JL, Roelofs CR, Karagas MR, Wrensch MR, Yeh RF, Nelson HH et al (2009) Epigenetic profiles distinguish pleural mesothelioma from normal pleura and predict lung asbestos burden and clinical outcome. *Cancer Res* 69(1):227–234
24. Cory S, Adams JM (2005) Killing cancer cells by flipping the Bcl-2/Bax switch. *Cancer Cell* 8(1):5–6
25. Crisanti MC, Wallace AF, Kapoor V, Vandermeers F, Dowling ML, Pereira LP, Coleman K, Campling BG, Fridlender ZG, Kao GD et al (2009) The HDAC inhibitor panobinostat (LBH589) inhibits mesothelioma and lung cancer cells in vitro and in vivo with particular efficacy for small cell lung cancer. *Mol Cancer Ther* 8(8):2221–2231
26. Delage B, Fennell DA, Nicholson L, McNeish I, Lemoine NR, Crook T, Szlosarek PW (2010) Arginine deprivation and argininosuccinate synthetase expression in the treatment of cancer. *Int J Cancer* 126(12):2762–2772
27. Du C, Fang M, Li Y, Li L, Wang X (2000) Smac, a mitochondrial protein that promotes cytochrome c-dependent caspase activation by eliminating IAP inhibition. *Cell* 102(1):33–42
28. Ensor CM, Holtzberg FW, Bomalaski JS, Clark MA (2002) Pegylated arginine deiminase (ADI-SS PEG20, 000 mw) inhibits human melanomas and hepatocellular carcinomas in vitro and in vivo. *Cancer Res* 62(19):5443–5450
29. Falleni M, Pellegrini C, Marchetti A, Roncalli M, Nosotti M, Palleschi A, Santambrogio L, Coggi G, Bosari S (2005) Quantitative evaluation of the apoptosis regulating genes Survivin, Bcl-2 and Bax in inflammatory and malignant pleural lesions. *Lung Cancer (Amsterdam, Netherlands)* 48(2):211–216
30. Farmer H, McCabe N, Lord CJ, Tutt AN, Johnson DA, Richardson TB, Santarosa M, Dillon KJ, Hickson I, Knights C et al (2005) Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature* 434(7035):917–921
31. Fennell DA, Rudd RM (2004) Defective core-apoptosis signalling in diffuse malignant pleural mesothelioma: opportunities for effective drug development. *Lancet Oncol* 5(6):354–362
32. Fennell DA, Gaudino G, O'Byrne KJ, Mutti L, van Meerbeek J (2008) Advances in the

- systemic therapy of malignant pleural mesothelioma. *Nat Clin Pract* 5(3):136–147
33. Fennell DA, Chacko A, Mutti L (2008) BCL-2 family regulation by the 20S proteasome inhibitor bortezomib. *Oncogene* 27(9):1189–1197
  34. Fernandez Y, Verhaegen M, Miller TP, Rush JL, Steiner P, Pipari AW Jr, Lowe SW, Soengas MS (2005) Differential regulation of noxa in normal melanocytes and melanoma cells by proteasome inhibition: therapeutic implications. *Cancer Res* 65(14):6294–6304
  35. Finin MS, Donigian JR, Cohen A, Richon VM, Rifkind RA, Marks PA, Breslow R, Pavletich NP (1999) Structures of a histone deacetylase homologue bound to the TSA and SAHA inhibitors. *Nature* 401(6749):188–193
  36. Fischer JR, Ohnmacht U, Rieger N, Zemaitis M, Stoffregen C, Kostrzewa M, Buchholz E, Manegold C, Lahm H (2006) Promoter methylation of RASSF1A, RARBeta and DAPK predict poor prognosis of patients with malignant mesothelioma. *Lung Cancer* (Amsterdam, Netherlands) 54(1):109–116
  37. Fong PC, Boss DS, Yap TA, Tutt A, Wu P, Mergui-Roelvink M, Mortimer P, Swaisland H, Lau A, O'Connor MJ et al (2009) Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers. *N Engl J Med* 361(2):123–134
  38. Fotheringham S, Epping MT, Stimson L, Khan O, Wood V, Pezzella F, Bernards R, La Thangue NB (2009) Genome-wide loss-of-function screen reveals an important role for the proteasome in HDAC inhibitor-induced apoptosis. *Cancer Cell* 15(1):57–66
  39. Fox S, Dharmarajan A (2006) WNT signaling in malignant mesothelioma. *Front Biosci* 11: 2106–2112
  40. Frizelle SP, Grim J, Zhou J, Gupta P, Curiel DT, Geradts J, Kratzke RA (1998) Re-expression of p16INK4a in mesothelioma cells results in cell cycle arrest, cell death, tumor suppression and tumor regression. *Oncogene* 16(24):3087–3095
  41. Frizelle SP, Rubins JB, Zhou JX, Curiel DT, Kratzke RA (2000) Gene therapy of established mesothelioma xenografts with recombinant p16INK4a adenovirus. *Cancer Gene Ther* 7(11):1421–1425
  42. Garber K (2009) Drugging the Wnt pathway: problems and progress. *J Natl Cancer Inst* 101(8):548–550
  43. Gordon GJ, Appasani K, Parcells JP, Mukhopadhyay NK, Jaklitsch MT, Richards WG, Sugarbaker DJ, Bueno R (2002) Inhibitor of apoptosis protein-1 promotes tumor cell survival in mesothelioma. *Carcinogenesis* 23(6): 1017–1024
  44. Gordon GJ, Mani M, Mukhopadhyay L, Dong L, Edenfield HR, Glickman JN, Yeap BY, Sugarbaker DJ, Bueno R (2007) Expression patterns of inhibitor of apoptosis proteins in malignant pleural mesothelioma. *J Pathol* 211(4):447–454
  45. Gordon GJ, Mani M, Mukhopadhyay L, Dong L, Yeap BY, Sugarbaker DJ, Bueno R (2007) Inhibitor of apoptosis proteins are regulated by tumour necrosis factor-alpha in malignant pleural mesothelioma. *J Pathol* 211(4):439–446
  46. Gordon GJ, Mani M, Maulik G, Mukhopadhyay L, Yeap BY, Kindler HL, Salgia R, Sugarbaker DJ, Bueno R (2008) Preclinical studies of the proteasome inhibitor bortezomib in malignant pleural mesothelioma. *Cancer Chemother Pharmacol* 61(4):549–558
  47. Goto Y, Shinjo K, Kondo Y, Shen L, Toyota M, Suzuki H, Gao W, An B, Fujii M, Murakami H et al (2009) Epigenetic profiles distinguish malignant pleural mesothelioma from lung adenocarcinoma. *Cancer Res* 69(23):9073–9082
  48. Hanahan D, Weinberg RA (2000) The hallmarks of cancer. *Cell* 100(1):57–70
  49. Harvey P, Warn A, Newman P, Perry LJ, Ball RY, Warn RM (1996) Immunoreactivity for hepatocyte growth factor/scatter factor and its receptor, met, in human lung carcinomas and malignant mesotheliomas. *J Pathol* 180(4): 389–394
  50. Harvey P, Warn A, Dobbin S, Arakaki N, Daikuhara Y, Jaurand MC, Warn RM (1998) Expression of HGF/SF in mesothelioma cell lines and its effects on cell motility, proliferation and morphology. *Br J Cancer* 77(7):1052–1059
  51. He B, Lee AY, Dadfarmay S, You L, Xu Z, Reguart N, Mazieres J, Mikami I, McCormick F, Jablons DM (2005) Secreted frizzled-related protein 4 is silenced by hypermethylation and induces apoptosis in beta-catenin-deficient human mesothelioma cells. *Cancer Res* 65(3): 743–748
  52. Hopkins-Donaldson S, Cathomas R, Simoes-Wust AP, Kurtz S, Belyanskaya L, Stahel RA, Zangemeister-Wittke U (2003) Induction of apoptosis and chemosensitization of mesothelioma cells by Bcl-2 and Bcl-xL antisense treatment. *Int J Cancer* 106(2):160–166
  53. Hu Q, Akatsuka S, Yamashita Y, Ohara H, Nagai H, Okazaki Y, Takahashi T, Toyokuni S

- (2010) Homozygous deletion of CDKN2A/2B is a hallmark of iron-induced high-grade rat mesothelioma. *Lab Invest: A Journal of Technical Methods and Pathology* 90(3): 360–373
54. Huang SM, Mishina YM, Liu S, Cheung A, Stegmeier F, Michaud GA, Charlat O, Wiellette E, Zhang Y, Wiessner S et al (2009) Tankyrase inhibition stabilizes axin and antagonizes Wnt signalling. *Nature* 461(7264): 614–620
  55. Illei PB, Rusch VW, Zakowski MF, Ladanyi M (2003) Homozygous deletion of CDKN2A and codeletion of the methylthioadenosine phosphorylase gene in the majority of pleural mesotheliomas. *Clin Cancer Res* 9(6): 2108–2113
  56. Izzo F, Marra P, Beneduce G, Castello G, Vallone P, De Rosa V, Cremona F, Ensor CM, Holtzberg FW, Bomalaski JS et al (2004) Pegylated arginine deiminase treatment of patients with unresectable hepatocellular carcinoma: results from phase I/II studies. *J Clin Oncol* 22(10):1815–1822
  57. Jagadeeswaran R, Ma PC, Seiwert TY, Jagadeeswaran S, Zumba O, Nallasura V, Ahmed S, Filiberti R, Paganuzzi M, Puntoni R et al (2006) Functional analysis of c-Met/hepatocyte growth factor pathway in malignant pleural mesothelioma. *Cancer Res* 66(1):352–361
  58. Johnstone RW, Ruefli AA, Lowe SW (2002) Apoptosis: a link between cancer genetics and chemotherapy. *Cell* 108(2):153–164
  59. Jongsma J, van Montfort E, Vooijs M, Zevenhoven J, Krimpenfort P, van der Valk M, van de Vijver M, Berns A (2008) A conditional mouse model for malignant mesothelioma. *Cancer Cell* 13(3):261–271
  60. Kawaguchi K, Murakami H, Taniguchi T, Fujii M, Kawata S, Fukui T, Kondo Y, Osada H, Usami N, Yokoi K et al (2009) Combined inhibition of MET and EGFR suppresses proliferation of malignant mesothelioma cells. *Carcinogenesis* 30(7):1097–1105
  61. Khan O, Fotheringham S, Wood V, Stimson L, Zhang C, Pezzella F, Duvic M, Kerr DJ, La Thangue NB (2010) HR23B is a biomarker for tumor sensitivity to HDAC inhibitor-based therapy. *Proc Natl Acad Sci USA* 107(14): 6532–6537
  62. Kim KU, Wilson SM, Abayasiriwardana KS, Collins R, Fjellbirkeland L, Xu Z, Jablons DM, Nishimura SL, Broaddus VC (2005) A novel in vitro model of human mesothelioma for studying tumor biology and apoptotic resistance. *Am J Respir Cell Mol Biol* 33(6): 541–548
  63. Kim Y, Ton TV, DeAngelo AB, Morgan K, Devereux TR, Anna C, Collins JB, Paules RS, Crosby LM, Sills RC (2006) Major carcinogenic pathways identified by gene expression analysis of peritoneal mesotheliomas following chemical treatment in F344 rats. *Toxicol Appl Pharmacol* 214(2):144–151
  64. Kindler HL, Burris HA III, Sandler AB, Oliff IA (2009) A phase II multicenter study of L-alanosine, a potent inhibitor of adenine biosynthesis, in patients with MTAP-deficient cancer. *Invest New Drugs* 27(1):75–81
  65. Kleinberg L, Lie AK, Florenes VA, Nesland JM, Davidson B (2007) Expression of inhibitor-of-apoptosis protein family members in malignant mesothelioma. *Hum Pathol* 38(7): 986–994
  66. Klominek J, Baskin B, Liu Z, Hauzenberger D (1998) Hepatocyte growth factor/scatter factor stimulates chemotaxis and growth of malignant mesothelioma cells through c-met receptor. *Int J Cancer* 76(2):240–249
  67. Kobayashi N, Toyooka S, Yanai H, Soh J, Fujimoto N, Yamamoto H, Ichihara S, Kimura K, Ichimura K, Sano Y et al (2008) Frequent p16 inactivation by homozygous deletion or methylation is associated with a poor prognosis in Japanese patients with pleural mesothelioma. *Lung Cancer (Amsterdam, Netherlands)* 62(1):120–125
  68. Konopleva M, Contractor R, Tsao T, Samudio I, Ruvolo PP, Kitada S, Deng X, Zhai D, Shi YX, Sneed T et al (2006) Mechanisms of apoptosis sensitivity and resistance to the BH3 mimetic ABT-737 in acute myeloid leukemia. *Cancer Cell* 10(5):375–388
  69. Kratzke RA, Wang X, Wong L, Kratzke MG, Green MR, Vokes EE, Vogelzang NJ, Kindler HL, Kern JA (2008) Response to the methylation inhibitor dihydro-5-azacytidine in mesothelioma is not associated with methylation of p16INK4a: results of cancer and leukemia group B 159904. *J Thorac Oncol* 3(4):417–421
  70. Krimpenfort P, Ijpenberg A, Song JY, van der Valk M, Nawijn M, Zevenhoven J, Berns A (2007) p15Ink4b is a critical tumour suppressor in the absence of p16Ink4a. *Nature* 448(7156):943–946
  71. Krug LM, Curley T, Schwartz L, Richardson S, Marks P, Chiao J, Kelly WK (2006) Potential

- role of histone deacetylase inhibitors in mesothelioma: clinical experience with suberoylanilide hydroxamic acid. *Clin Lung Cancer* 7(4): 257–261
72. Ladanyi M (2005) Implications of P16/CDKN2A deletion in pleural mesotheliomas. *Lung Cancer (Amsterdam, Netherlands)* 49(Suppl 1):S95–S98
  73. Lecomte C, Andujar P, Renier A, Kheuang L, Abramowski V, Mellottee L, Fleury-Feith J, Zucman-Rossi J, Giovannini M, Jaurand MC (2005) Similar tumor suppressor gene alteration profiles in asbestos-induced murine and human mesothelioma. *Cell Cycle (Georgetown, TX)* 4(12):1862–1869
  74. Lee AY, He B, You L, Dadfarmay S, Xu Z, Mazieres J, Mikami I, McCormick F, Jablons DM (2004) Expression of the secreted frizzled-related protein gene family is downregulated in human mesothelioma. *Oncogene* 23(39):6672–6676
  75. Lee AY, He B, You L, Xu Z, Mazieres J, Reguart N, Mikami I, Batra S, Jablons DM (2004) Dickkopf-1 antagonizes Wnt signaling independent of beta-catenin in human mesothelioma. *Biochem Biophys Res Commun* 323(4):1246–1250
  76. Letai A, Bassik MC, Walensky LD, Sorcinelli MD, Weiler S, Korsmeyer SJ (2002) Distinct BH3 domains either sensitize or activate mitochondrial apoptosis, serving as prototype cancer therapeutics. *Cancer Cell* 2(3):183–192
  77. Li P, Nijhawan D, Budihardjo I, Srinivasula SM, Ahmad M, Alnemri ES, Wang X (1997) Cytochrome c and dATP-dependent formation of Apaf-1/caspase-9 complex initiates an apoptotic protease cascade. *Cell* 91(4):479–489
  78. Li H, Zhu H, Xu CJ, Yuan J (1998) Cleavage of BID by caspase 8 mediates the mitochondrial damage in the Fas pathway of apoptosis. *Cell* 94(4):491–501
  79. Li W, You L, Cooper J, Schiavon G, Pepe-Caprio A, Zhou L, Ishii R, Giovannini M, Hanemann CO, Long SB et al (2010) Merlin/NF2 suppresses tumorigenesis by inhibiting the E3 ubiquitin ligase CRL4(DCAF1) in the nucleus. *Cell* 140(4):477–490
  80. Lindemann RK, Newbold A, Whitecross KF, Cluse LA, Frew AJ, Ellis L, Williams S, Wiegman AP, Dear AE, Scott CL et al (2007) Analysis of the apoptotic and therapeutic activities of histone deacetylase inhibitors by using a mouse model of B cell lymphoma. *Proc Natl Acad Sci USA* 104(19):8071–8076
  81. Lu J, Ma Z, Hsieh JC, Fan CW, Chen B, Longgood JC, Williams NS, Amatruda JF, Lum L, Chen C (2009) Structure-activity relationship studies of small-molecule inhibitors of Wnt response. *Bioorg Med Chem Lett* 19(14): 3825–3827
  82. Martins LM, Iaccarino I, Tenev T, Gschmeissner S, Totty NF, Lemoine NR, Savopoulos J, Gray CW, Creasy CL, Dingwall C et al (2002) The serine protease Omi/HtrA2 regulates apoptosis by binding XIAP through a reaper-like motif. *J Biol Chem* 277(1):439–444
  83. Mazieres J, You L, He B, Xu Z, Twogood S, Lee AY, Reguart N, Batra S, Mikami I, Jablons DM (2005) Wnt2 as a new therapeutic target in malignant pleural mesothelioma. *Int J Cancer* 117(2):326–332
  84. Meijerink JP, Mensink EJ, Wang K, Sedlak TW, Sloetjes AW, de Witte T, Waksman G, Korsmeyer SJ (1998) Hematopoietic malignancies demonstrate loss-of-function mutations of BAX. *Blood* 91(8):2991–2997
  85. Mendes-Pereira AM, Martin SA, Brough R, McCarthy A, Taylor JR, Kim JS, Waldman T, Lord CJ, Ashworth A (2009) Synthetic lethal targeting of PTEN mutant cells with PARP inhibitors. *EMBO Mol Med* 1(6–7):315–322
  86. Mitsiades CS, Poulaki V, McMullan C, Negri J, Fanourakis G, Goudopoulou A, Richon VM, Marks PA, Mitsiades N (2005) Novel histone deacetylase inhibitors in the treatment of thyroid cancer. *Clin Cancer Res* 11(10):3958–3965
  87. Mohiuddin I, Cao X, Ozvaran MK, Zumstein L, Chada S, Smythe WR (2002) Phosphatase and tensin analog gene overexpression engenders cellular death in human malignant mesothelioma cells via inhibition of AKT phosphorylation. *Ann Surg Oncol* 9(3):310–316
  88. Moye-Rowley WS, Harshman KD, Parker CS (1989) Yeast YAP1 encodes a novel form of the jun family of transcriptional activator proteins. *Genes Dev* 3(3):283–292
  89. Mukohara T, Civiello G, Davis IJ, Taffaro ML, Christensen J, Fisher DE, Johnson BE, Janne PA (2005) Inhibition of the met receptor in mesothelioma. *Clin Cancer Res* 11(22):8122–8130
  90. Muzio M, Chinnaiyan AM, Kischkel FC, O'Rourke K, Shevchenko A, Ni J, Scaffidi C, Bretz JD, Zhang M, Gentz R et al (1996) FLICE, a novel FADD-homologous ICE/CED-3-like protease, is recruited to the CD95 (Fas/APO-1) death-inducing signaling complex. *Cell* 85(6): 817–827
  91. Neragi-Miandoab S, Sugarbaker DJ (2009) Chromosomal deletion in patients with malignant

- pleural mesothelioma. *Interact Cardiovasc Thorac Surg* 9(1):42–44
92. Nguyen M, Marcellus RC, Roulston A, Watson M, Serfass L, Murthy Madiraju SR, Goulet D, Viallet J, Belec L, Billot X et al (2007) Small molecule obatoclax (GX15-070) antagonizes MCL-1 and overcomes MCL-1-mediated resistance to apoptosis. *Proc Natl Acad Sci USA* 104(49):19512–19517
  93. Nikiforov MA, Riblett M, Tang WH, Gratchouk V, Zhuang D, Fernandez Y, Verhaegen M, Varambally S, Chinnaiyan AM, Jakubowiak AJ et al (2007) Tumor cell-selective regulation of NOXA by c-MYC in response to proteasome inhibition. *Proc Natl Acad Sci USA* 104(49):19488–19493
  94. Nikrad M, Johnson T, Puthalalath H, Coultas L, Adams J, Kraft AS (2005) The proteasome inhibitor bortezomib sensitizes cells to killing by death receptor ligand TRAIL via BH3-only proteins Bik and Bim. *Mol Cancer Ther* 4(3):443–449
  95. O'Connor L, Strasser A, O'Reilly LA, Hausmann G, Adams JM, Cory S, Huang DC (1998) Bim: a novel member of the Bcl-2 family that promotes apoptosis. *EMBO J* 17(2):384–395
  96. O'Kane SL, Pound RJ, Campbell A, Chaudhuri N, Lind MJ, Cawkwell L (2006) Expression of bcl-2 family members in malignant pleural mesothelioma. *Acta Oncol (Stockholm, Sweden)* 45(4):449–453
  97. Oerlemans R, Franke NE, Assaraf YG, Cloos J, van Zantwijk I, Berkers CR, Scheffer GL, Debipersad K, Vojtekova K, Lemos C et al (2008) Molecular basis of bortezomib resistance: proteasome subunit beta5 (PSMB5) gene mutation and overexpression of PSMB5 protein. *Blood* 112(6):2489–2499
  98. Oltersdorf T, Elmore SW, Shoemaker AR, Armstrong RC, Augeri DJ, Belli BA, Bruncko M, Deckwerth TL, Dinges J, Hajduk PJ et al (2005) An inhibitor of Bcl-2 family proteins induces regression of solid tumours. *Nature* 435(7042):677–681
  99. Oltvai ZN, Milliman CL, Korsmeyer SJ (1993) Bcl-2 heterodimerizes in vivo with a conserved homolog, Bax, that accelerates programmed cell death. *Cell* 74(4):609–619
  100. Onofre FB, Onofre AS, Pomjanski N, Buckstegge B, Grote HJ, Bocking A (2008) 9p21 Deletion in the diagnosis of malignant mesothelioma in serous effusions additional to immunocytochemistry, DNA-ICM, and AgNOR analysis. *Cancer* 114(3):204–215
  101. Opitz I, Soltermann A, Abaecherli M, Hinterberger M, Probst-Hensch N, Stahel R, Moch H, Weder W (2008) PTEN expression is a strong predictor of survival in mesothelioma patients. *Eur J Cardiothorac Surg* 33(3):502–506
  102. Papp T, Schipper H, Pemsel H, Bastrop R, Muller KM, Wiethage T, Weiss DG, Dopp E, Schiffmann D, Rahman Q (2001) Mutational analysis of N-ras, p53, p16INK4a, p14ARF and CDK4 genes in primary human malignant mesotheliomas. *Int J Oncol* 18(2):425–433
  103. Pass HI, Goparaju C, Ivanov S, Donington J, Carbone M, Hoshen M, Cohen D, Chajut A, Rosenwald S, Dan H et al (2010) hsa-miR-29c\* is linked to the prognosis of malignant pleural mesothelioma. *Cancer Res* 70(5):1916–1924
  104. Pespeni MH, Hodnett M, Abayasiriwardana KS, Roux J, Howard M, Broaddus VC, Pittet JF (2007) Sensitization of mesothelioma cells to tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis by heat stress via the inhibition of the 3-phosphoinositide-dependent kinase 1/Akt pathway. *Cancer Res* 67(6):2865–2871
  105. Pinton G, Brunelli E, Murer B, Puntoni R, Puntoni M, Fennell DA, Gaudino G, Mutti L, Moro L (2009) Estrogen receptor-beta affects the prognosis of human malignant mesothelioma. *Cancer Res* 69(11):4598–4604
  106. Poulidakos PI, Xiao GH, Gallagher R, Jablonski S, Jhanwar SC, Testa JR (2006) Re-expression of the tumor suppressor NF2/merlin inhibits invasiveness in mesothelioma cells and negatively regulates FAK. *Oncogene* 25(44):5960–5968
  107. Qin JZ, Ziffra J, Stennett L, Bodner B, Bonish BK, Chaturvedi V, Bennett F, Pollock PM, Trent JM, Hendrix MJ et al (2005) Proteasome inhibitors trigger NOXA-mediated apoptosis in melanoma and myeloma cells. *Cancer Res* 65(14):6282–6293
  108. Ramalingam SS, Belani CP, Ruel C, Frankel P, Gitlitz B, Koczywas M, Espinoza-Delgado I, Gandara D (2009) Phase II study of belinostat (PXD101), a histone deacetylase inhibitor, for second line therapy of advanced malignant pleural mesothelioma. *J Thorac Oncol* 4(1):97–101
  109. Ramos-Nino ME, Scapoli L, Martinelli M, Land S, Mossman BT (2003) Microarray analysis and RNA silencing link fra-1 to cd44 and c-met expression in mesothelioma. *Cancer Res* 63(13):3539–3545



110. Ramos-Nino ME, Blumen SR, Pass H, Mossman BT (2007) Fra-1 governs cell migration via modulation of CD44 expression in human mesotheliomas. *Mol Cancer* 6:81
111. Ramos-Nino ME, Blumen SR, Sabo-Attwood T, Pass H, Carbone M, Testa JR, Altomare DA, Mossman BT (2008) HGF mediates cell proliferation of human mesothelioma cells through a PI3K/MEK5/Fra-1 pathway. *Am J Respir Cell Mol Biol* 38(2): 209–217
112. Rippo MR, Moretti S, Vescovi S, Tomasetti M, Orecchia S, Amici G, Catalano A, Procopio A (2004) FLIP overexpression inhibits death receptor-induced apoptosis in malignant mesothelial cells. *Oncogene* 23(47): 7753–7760
113. Sartore-Bianchi A, Gasparri F, Galvani A, Nici L, Darnowski JW, Barbone D, Fennell DA, Gaudino G, Porta C, Mutti L (2007) Bortezomib inhibits nuclear factor-kappaB dependent survival and has potent in vivo activity in mesothelioma. *Clin Cancer Res* 13(19):5942–5951
114. Sealy L, Chalkley R (1978) The effect of sodium butyrate on histone modification. *Cell* 14(1):115–121
115. Sedlak TW, Oltvai ZN, Yang E, Wang K, Boise LH, Thompson CB, Korsmeyer SJ (1995) Multiple Bcl-2 family members demonstrate selective dimerizations with Bax. *Proc Natl Acad Sci USA* 92(17):7834–7838
116. Sekido Y, Pass HI, Bader S, Mew DJ, Christman MF, Gazdar AF, Minna JD (1995) Neurofibromatosis type 2 (NF2) gene is somatically mutated in mesothelioma but not in lung cancer. *Cancer Res* 55(6):1227–1231
117. Shen WH, Balajee AS, Wang J, Wu H, Eng C, Pandolfi PP, Yin Y (2007) Essential role for nuclear PTEN in maintaining chromosomal integrity. *Cell* 128(1):157–170
118. Shigemitsu K, Sekido Y, Usami N, Mori S, Sato M, Horio Y, Hasegawa Y, Bader SA, Gazdar AF, Minna JD et al (2001) Genetic alteration of the beta-catenin gene (CTNNB1) in human lung cancer and malignant mesothelioma and identification of a new 3p21.3 homozygous deletion. *Oncogene* 20(31):4249–4257
119. Soini Y, Kinnula V, Kaarteenaho-Wiik R, Kurttila E, Linnainmaa K, Paakko P (1999) Apoptosis and expression of apoptosis regulating proteins bcl-2, mcl-1, bcl-X, and bax in malignant mesothelioma. *Clin Cancer Res* 5(11):3508–3515
120. Sumi K, Matsuyama S, Kitajima Y, Miyazaki K (2004) Loss of estrogen receptor beta expression at cancer front correlates with tumor progression and poor prognosis of gallbladder cancer. *Oncol Rep* 12(5):979–984
121. Sun X, Gulyas M, Hjerpe A, Dobra K (2006) Proteasome inhibitor PSI induces apoptosis in human mesothelioma cells. *Cancer Lett* 232(2):161–169
122. Susin SA, Lorenzo HK, Zamzami N, Marzo I, Snow BE, Brothers GM, Mangion J, Jacotot E, Costantini P, Loeffler M et al (1999) Molecular characterization of mitochondrial apoptosis-inducing factor. *Nature* 397(6718):441–446
123. Symanowski J, Vogelzang N, Zawel L, Atadja P, Pass H, Sharma S (2009) A histone deacetylase inhibitor LBH589 downregulates XIAP in mesothelioma cell lines which is likely responsible for increased apoptosis with TRAIL. *J Thorac Oncol* 4(2):149–160
124. Szlosarek PW, Klabatsa A, Pallaska A, Sheaff M, Smith P, Crook T, Grimshaw MJ, Steele JP, Rudd RM, Balkwill FR et al (2006) In vivo loss of expression of argininosuccinate synthetase in malignant pleural mesothelioma is a biomarker for susceptibility to arginine depletion. *Clin Cancer Res* 12(23):7126–7131
125. Taguchi T, Jhanwar SC, Siegfried JM, Keller SM, Testa JR (1993) Recurrent deletions of specific chromosomal sites in 1p, 3p, 6q, and 9p in human malignant mesothelioma. *Cancer Res* 53(18):4349–4355
126. Takada Y, Gillenwater A, Ichikawa H, Aggarwal BB (2006) Suberoylanilide hydroxamic acid potentiates apoptosis, inhibits invasion, and abolishes osteoclastogenesis by suppressing nuclear factor-kappaB activation. *J Biol Chem* 281(9):5612–5622
127. Tartaglia LA, Ayres TM, Wong GH, Goeddel DV (1993) A novel domain within the 55 kd TNF receptor signals cell death. *Cell* 74(5): 845–853
128. Thirkettle I, Harvey P, Hasleton PS, Ball RY, Warn RM (2000) Immunoreactivity for cadherins, HGF/SF, met, and erbB-2 in pleural malignant mesotheliomas. *Histopathology* 36(6):522–528
129. Tsujimoto Y, Finger LR, Yunis J, Nowell PC, Croce CM (1984) Cloning of the chromosome breakpoint of neoplastic B cells with the t(14;18) chromosome translocation. *Science* (New York) 226(4678):1097–1099
130. Uematsu K, Kanazawa S, You L, He B, Xu Z, Li K, Peterlin BM, McCormick F, Jablons DM

- (2003) Wnt pathway activation in mesothelioma: evidence of Dishevelled overexpression and transcriptional activity of beta-catenin. *Cancer Res* 63(15):4547–4551
131. Uematsu K, Seki N, Seto T, Isoe C, Tsukamoto H, Mikami I, You L, He B, Xu Z, Jablons DM et al (2007) Targeting the Wnt signaling pathway with dishevelled and cisplatin synergistically suppresses mesothelioma cell growth. *Anticancer Res* 27(6B):4239–4242
  132. van Delft MF, Wei AH, Mason KD, Vandenberg CJ, Chen L, Czabotar PE, Willis SN, Scott CL, Day CL, Cory S et al (2006) The BH3 mimetic ABT-737 targets selective Bcl-2 proteins and efficiently induces apoptosis via Bak/Bax if Mcl-1 is neutralized. *Cancer Cell* 10(5):389–399
  133. Vandermeers F, Hubert P, Delvenne P, Mascaux C, Grigoriu B, Burny A, Scherpereel A, Willems L (2009) Valproate, in combination with pemetrexed and cisplatin, provides additional efficacy to the treatment of malignant mesothelioma. *Clin Cancer Res* 15(8):2818–2828
  134. Velcheti V, Kasai Y, Viswanathan AK, Ritter J, Govindan R (2009) Absence of mutations in the epidermal growth factor receptor (EGFR) kinase domain in patients with mesothelioma. *J Thorac Oncol* 4(4):559
  135. Vivo C, Liu W, Broaddus VC (2003) c-Jun N-terminal kinase contributes to apoptotic synergy induced by tumor necrosis factor-related apoptosis-inducing ligand plus DNA damage in chemoresistant, p53 inactive mesothelioma cells. *J Biol Chem* 278(28):25461–25467
  136. Wang K, Yin XM, Chao DT, Milliman CL, Korsmeyer SJ (1996) BID: a novel BH3 domain-only death agonist. *Genes Dev* 10(22):2859–2869
  137. Wang L, Du F, Wang X (2008) TNF-alpha induces two distinct caspase-8 activation pathways. *Cell* 133(4):693–703
  138. Wang Y, Rishi AK, Puliappadamba VT, Sharma S, Yang H, Tarca A, Ping Dou Q, Lonardo F, Ruckdeschel JC, Pass HI et al (2009) Targeted proteasome inhibition by Velcade induces apoptosis in human mesothelioma and breast cancer cell lines. *Cancer Chemother Pharmacol* 10:1235–1244
  139. Wang Q, Mora-Jensen H, Weniger MA, Perez-Galan P, Wolford C, Hai T, Ron D, Chen W, Trenkle W, Wiestner A et al (2009) ERAD inhibitors integrate ER stress with an epigenetic mechanism to activate BH3-only protein NOXA in cancer cells. *Proc Natl Acad Sci USA* 106(7):2200–2205
  140. Wei MC, Zong WX, Cheng EH, Lindsten T, Panoutsakopoulou V, Ross AJ, Roth KA, MacGregor GR, Thompson CB, Korsmeyer SJ (2001) Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. *Science (New York)* 292(5517):727–730
  141. Wilson TR, Redmond KM, McLaughlin KM, Crawford N, Gately K, O'Byrne K, Le-Clorrene C, Holohan C, Fennell DA, Johnston PG et al (2009) Procaspase 8 overexpression in non-small-cell lung cancer promotes apoptosis induced by FLIP silencing. *Cell Death Differ* 16(10):1352–1361
  142. Wong L, Zhou J, Anderson D, Kratzke RA (2002) Inactivation of p16INK4a expression in malignant mesothelioma by methylation. *Lung Cancer (Amsterdam, Netherlands)* 38(2):131–136
  143. [www.cancer.gov](http://www.cancer.gov): Suberoylanilide hydroxamic acid (Vorinostat, MK0683) versus placebo in advanced malignant pleural mesothelioma. In [www.http://clinicaltrials.gov/ct2/show/NCT00128102](http://clinicaltrials.gov/ct2/show/NCT00128102).
  144. Xia C, Xu Z, Yuan X, Uematsu K, You L, Li K, Li L, McCormick F, Jablons DM (2002) Induction of apoptosis in mesothelioma cells by antisurvivin oligonucleotides. *Mol Cancer Ther* 1(9):687–694
  145. Xio S, Li D, Vijg J, Sugarbaker DJ, Corson JM, Fletcher JA (1995) Codeletion of p15 and p16 in primary malignant mesothelioma. *Oncogene* 11(3):511–515
  146. Yin XM, Oltvai ZN, Korsmeyer SJ (1994) BH1 and BH2 domains of Bcl-2 are required for inhibition of apoptosis and heterodimerization with Bax. *Nature* 369(6478):321–323
  147. Yin C, Knudson CM, Korsmeyer SJ, Van Dyke T (1997) Bax suppresses tumorigenesis and stimulates apoptosis in vivo. *Nature* 385(6617):637–640
  148. Yokoyama T, Osada H, Murakami H, Tatematsu Y, Taniguchi T, Kondo Y, Yatabe Y, Hasegawa Y, Shimokata K, Horio Y et al (2008) YAP1 is involved in mesothelioma development and negatively regulated by Merlin through phosphorylation. *Carcinogenesis* 29(11):2139–2146
  149. Yoshida H, Kong YY, Yoshida R, Elia AJ, Hakem A, Hakem R, Penninger JM, Mak TW (1998) Apaf1 is required for mitochondrial pathways of apoptosis and brain development. *Cell* 94(6):739–750

150. You L, He B, Uematsu K, Xu Z, Mazieres J, Lee A, McCormick F, Jablons DM (2004) Inhibition of Wnt-1 signaling induces apoptosis in beta-catenin-deficient mesothelioma cells. *Cancer Res* 64(10):3474–3478
151. Yuan BZ, Chapman JA, Reynolds SH (2008) Proteasome inhibitor MG132 induces apoptosis and inhibits invasion of human malignant pleural mesothelioma cells. *Transl Oncol* 1(3): 129–140
152. Zaffaroni N, Costa A, Pennati M, De Marco C, Affini E, Madeo M, Erdas R, Cabras A, Kusamura S, Baratti D et al (2007) Survivin is highly expressed and promotes cell survival in malignant peritoneal mesothelioma. *Cell Oncol* 29(6):453–466
153. Zimmerman RL, Fogt F (2001) Evaluation of the c-Met immunostain to detect malignant cells in body cavity effusions. *Oncol Rep* 8(6):1347–1350