42 Fanconi Syndrome

Takashi Igarashi

Fanconi syndrome (FS) is a generalized dysfunction of the renal proximal tubules leading to excessive urinary wasting of amino acids, glucose, phosphate, uric acid, bicarbonate, and other solutes. The patients develop failure to thrive, polyuria, polydipsia, dehydration, and rickets in children, and osteoporosis and osteomalacia in adults. The patients also manifest renal salt wasting, hypokalemia, metabolic acidosis, hypercalciuria, and lowmolecular-weight (LMW) proteinuria. De Toni, Debré, and Fanconi described children with renal rickets and glucosuria in the 1930s [\(1,](#page-22-0) [2,](#page-22-0) [3\)](#page-22-0). FS is named after Guido Fanconi, a Swiss pediatrician or it is called as de Toni-Debré-Fanconi syndrome.

Pathophysiology

The renal proximal tubules reabsorb almost all of the physiologically filtered load of proteins including of albumin, LMW proteins, amino acids, glucose, bicarbonate, sodium, chloride, phosphate, and uric acid. The transport processes in the proximal tubule can be characterized broadly as megalin/cubilin-mediated endocytic pathways and sodium (Na⁺) gradient-dependent transport systems.

The proximal tubule cells have extensive luminal receptors and endocytic apparatus such as megalin and cubilin that are critical for the reabsorption and degradation of proteins that traverse the glomerular filtration barrier ([4](#page-22-0)) (\bullet [Fig. 42-1](#page-1-0)) as well as for the extensive recycling of many functionally important membrane proteins [\(5\)](#page-22-0). Numerous filtered proteins including albumin, LMW proteins, polypeptide hormones, vitamin-binding proteins, and polybasic drugs such as aminoglycosides from glomerulus are bound to megalin and cubilin in the luminal membrane of proximal tubules. Then, the protein-receptor complex is incorporated into the endosome. The ligand and receptor are disassociated in the endosome; the receptor is recycled back to the luminal membrane and the reabsorbed proteins go into lysosome for further processing (\odot [Fig. 42-2](#page-2-0)). This disassociation is dependent on acidification of the endosome by increased concentration of H^+ and Cl^- due to the function of H^+ -ATPase (proton pump) and ClC-5 chloride channel. An abnormal

endocytosis pathway may affect the recycling of transport proteins (megalin and cubilin), the back to the luminal membrane, and the expression of megalin and cubilin in the luminal membrane, leading to decreased solute reabsorption. Perturbation of endosomal acidification in proximal tubule cells leads to diminished reabsorption and increased urinary wasting of albumin, LMW proteins, electrolytes, and solutes. Cadmium inhibits H⁺-ATPase and mitochondria, which results in a Fanconi-like syndrome (6) . Folimycin, a H⁺-ATPase inhibitor, abolishes albumin uptake by proximal tubules [\(7](#page-22-0)). Moreover, a defect of ClC-5 chloride channel in patients with Dent disease manifests Fanconi syndrome ([8](#page-22-0)). Acidification defect in the endosome in Dent disease leads to recycling from intracellular endosome into luminal membrane resulting in megalin deficiency in the luminal membrane of the proximal tubule. Analysis of normal human urine samples identified megalin as a physiologically excreted protein. The presence of megalin in normal human urine is due to shedding from the proximal tubule cells into the lumen. Patients with Dent disease demonstrate an almost complete absence of urinary megalin ([9](#page-22-0)). This megalinshedding deficiency in the urine is also observed in patients with Lowe syndrome ([9](#page-22-0)).

Reabsorption of filtered solutes including glucose, phosphate, amino acids, and bicarbonate by proximal tubule cells is accomplished by transport system at the brush border membrane that are directly or indirectly coupled to $Na⁺$ movement, by energy production and transport from the mitochondria, and by the $Na⁺, K⁺-ATPase$ at the basolateral membrane. The Na⁺, K⁺-ATPase lowers intracellular Na⁺ concentration and provides the electrochemical gradient that allows Na⁺-coupled solute entry into the cell. Disturbances in energy generation could impair net transepithelial transport in the proximal tubule. Energy is necessary for the operation of Na⁺, K⁺-ATPase and other membrane carriers that are involved with solute reabsorption of amino acid, glucose, phosphate, uric acid, and bicarbonate. Although the weight of bilateral kidneys is less than 1% of total body weight, kidneys consume about 10% of the total energy consumed by the whole body in a static condition. Moreover, most of the energy is consumed in the proximal tubule cells to operate multiple

Structure of megalin and cubilin. (Veroust PJ, Birn H, Nielsen R et al. The tandem endocytic receptors megalin and cubilin are important proteins in renal pathology. Kidney Int 2002;62:745–756).

membrane and intracellular transporter proteins. A defect in energy generation in the proximal tubule cells produces multiple transport anomalies of the proximal tubule that characterize the FS. The ATP production is severely compromised in cystine-loaded tubule in cystinosis [\(10](#page-22-0)). Thiol-containing enzymes are critical for renal energy metabolism. Cystine inhibits in vivo and in vitro the thiol-containing enzyme activity resulting in multiple renal transport anomalies ([11\)](#page-22-0). The mitochondrial

respiratory chain has a major role in ATP production during aerobic respiration. Genetic defects of enzyme complexes of the oxidative phosphorylation system or toxic substances including of drugs in the proximal tubule cells can produce mitochondrial respiratory chain defect leading to multiple renal transport anomalies [\(12](#page-22-0), [13](#page-22-0)). In contrast, isolated dysfunction of transporter proteins in proximal tubule cells results in the selective wasting of amino acids, glucose, phosphate, bicarbonate, or uric

ıCl[.]

CIC-5

D Figure 42-2

Schematic model of endocytosis in the proximal tubule cells. Albumin and low molecular weight proteins are filtered into the primary urine and endocytosed by proximal tubule cells via the megalin-cubilin receptor pathway. (a) The receptorligand complexes progress along the endocytic pathway. The endosomes undergo a progressive, ATP-dependent acidification that results in the dissociation of the receptor-ligand complexes, with megalin and cubilin being recycled in the luminal membrane, whereas the ligand is directed to lysosomes for degradation. (b) Vesicular acidification is mediated by the vacuolar H⁺-ATPase, which requires a net Cl $^-$ conductance to functions as an electrogenic nCl $^-$ /H $^+$ exchanger, which is predicted to facilitate acidification and to play a role in keeping high vesicular Cl⁻ concentration. (Devuyst O, Pirson Y. Genetics of hypercalciuric stone forming disease. Kidney Int 2007:72:1065–1092).

acids. However, glucosuria and aminoaciduria are seen in some patients with defective isolated proteins. They are familial renal glucosuria resulting from the mutations in the kidney-specific low affinity/high capacity Na⁺/glucose cotransporter gene (SLC5A2) and maturity-onset diabetes of young age type 3 (MODY3) resulting from the mutations in hepatocyte nuclear factor-1 alpha gene that acts as a regulator of transcription for SLC5A2 ([14,](#page-22-0) [15](#page-22-0)). Generalized aminoaciduria seen in these patients is considered as a consequence of the impairment in tubular glucose reabsorption, whereas the precise mechanism is not known.

Signs and Symptoms

Growth Retardation (Failure to Thrive)

Growth retardation (failure to thrive) is a common feature of FS in children [\(16](#page-22-0)). Patients with FS present severe growth failure at the time of diagnosis that persists into adult life. The pathomechanism of growth failure in FS is complex. Malnutrition, hypokalemia, hypophosphatemia, and metabolic acidosis can lead to growth retardation in patients with FS ([17\)](#page-22-0). Potassium deficiency induces growth retardation through reduced circulating levels of growth hormone (GH) and insulin-like growth factor I (IGF-I) [\(18,](#page-22-0) [19\)](#page-22-0). Hypokalemia can induce appetite decrease leading to malnutrition and extracellular volume contraction. Metabolic acidosis inhibits growth hormone secretion, the expression of IGF-I and GH receptor [\(20](#page-22-0)). Hypophosphatemia is related to severe bone changes leading to rickets and growth retardation in children with FS ([21\)](#page-22-0). In patients with adult onset FS, osteomalacia is thought to result from hypophosphatemia due to renal phosphate loss and relative 1,25-dihydroxyvitamin D3 deficiency ([22,](#page-22-0) [23\)](#page-22-0). Metabolic acidosis impairs the conversion of 25-vitamin D3 to 1,25-dihydroxy vitamin D3. The patients present bone and joint pain in the hips,

shoulders, and trunk and difficulty of walking due to multiple bone fracture. Hypomineralization of dentin structure and immature formation of craniofacial bones are seen in patients with FS ([24\)](#page-22-0). Specific forms of FS are associated with endocytotosis pathway dysfunction; disruption of megalin-mediated uptake vitamin D-binding protein/25-vitamin D3 complex produces metabolic bone disease in affected individuals ([25\)](#page-22-0).

Earlier diagnosis and efficient correction of acidosis and electrolyte balances by supportive therapy can contribute to improve growth and final height in patients with FS [\(17](#page-22-0), [21\)](#page-22-0). However, supportive therapy is frequently unable to prevent further loss of relative height in patients with FS, especially those with cystinosis.

Polyuria, Polydipsia, and Dehydration

Polyuria, polydipsia, and dehydration are frequently seen in patients with FS. Polyuria is secondary to the osmotic diuresis from the excessive urinary solute losses and urine concentration defect in the collecting ducts due to chronic hypokalemia. Recurrent acute fever due to dehydration is a frequent manifestation in infants with FS. In the most common type of cystinosis, Fanconi syndrome occurs at 6–12 months of age. Recurrent febrile episodes are often the first sign of FS in infantile patients with cystinosis [\(26](#page-22-0)).

Generalized Aminoaciduria (Generalized Hyperaminoaciduria)

Molecular weight (MW) of 20 different amino acids is small; the largest amino acid is tryptophan $[MW = 204 D]$ (daltons)]. Amino acids are not bound to proteins in the plasma. Thus, amino acids are freely filtered from glomerulus. Then, 95–99% of filtered load of amino acids are reabsorbed in the proximal tubules. More than one transporter in the proximal tubule cells absorbs amino acids. Fractional excretion of amino acid is usually less than 3% in the controls except for neonate or premature babies. However, only histidine has a fractional excretion of 5% in the controls.

> Fractional excretion of amino acid $(\%) =$ $[(Uaa/Paa)/(Ucr/Per)] \times 100$

(aa; amino acid, cr; creatinine, U; urine, P; plasma)

Therefore, the excretion more than 5% of the filtered load of amino acid is termed aminoaciduria or hyperaminoaciduria. Every amino acid is highly excreted in patients with FS, and this phenomenon is called as generalized aminoaciduria.

Glucosuria

Filtered load of glucose (D-glucose, $MW = 180$ D) is almost completely absorbed by a sodium-coupled active transport located in the brush border membrane of the proximal tubule in the normal condition. Glucose reabsorption involves a couple of transporters at the luminal and basolateral membranes of the proximal tubules. The driving force for glucose reabsorption is provided by Na^+ , K+ -ATPase in the plasma membrane. Thus, very small amount of glucose are present in the urine in the normal condition. Glucosuria is a common manifestation in FS. It is derived from impaired reabsorption of glucose when serum glucose is normal. Renal threshold of glucose is reduced in FS. Glucosuria is one of the originally described clinical features of FS [\(1,](#page-22-0) [2,](#page-22-0) [3\)](#page-22-0). 0.5–20 g of glucose a day is lost in the urine in patients with FS.

Hypophosphatemia

Most of the patients with FS manifest a low tubular reabsorption of phosphate (percent tubular reabsorption of phosphate: %TRP, >80–85% in the control) and decreased serum phosphate. Rickets and osteomalacia are produced by the increased urinary wasting of phosphate as well as by impaired 1a-hydroxylation of 25-hydroxy vitamin D3 by proximal tubule cells ([27\)](#page-22-0).

$$
\%TRP=[1\!-\!(Up/Sp)/(Ucr/Scr)]\times 100
$$

(p; phosphate, cr; creatinine, U; urine, S; serum)

The maximal threshold of phosphate (TmP/GFR) is a very sensitive indicator that reflects the reabsorption of phosphate in the renal tubules.

$$
TmP/GFR = TRP \times Sp
$$

(GFR; glomerular filtration rate, p; phosphate, S; serum)

The Tm/GFR is usually very low (2.3–4.3 in the control) in patients with FS. Rickets manifests bowing deformity of the lower limbs, distal femur, the ulna, and the radius.

Phosphate handling in the kidney is affected by a couple of factors including parathyroid hormone (PTH) and vitamin D. PTH level is normal or elevated in patients with FS. Serum 1, 25-dihydroxy vitamin D3 is variable in patients with FS [\(28,](#page-22-0) [29](#page-22-0)).

Metabolic Acidosis

More than 85% of filtered load of bicarbonate $(HCO3^{-})$ is reabsorbed by the proximal tubule cells. This is accomplished by the coordinated function of luminal membrane Na⁺/H⁺ exchanger, luminal membrane carbonic anhydrase IV and XIV, and basolateral membrane $\mathrm{Na^+}/\mathrm{HCO3}^$ cotransporter [\(30](#page-22-0)). Hyperchloremic metabolic acidosis is a common feature of FS resulting from defective bicarbonate reabsorption in the proximal tubules. Anion gap is normal. More than 30% of filtered load of $HCO3^-$ is not reabsorbed in patients with FS, and they manifest low plasma HCO3⁻ levels between 12–18 mEq L^{-1} . Fractional excretion of $HCO3^-$ (FEHCO3⁻) under the alkali treatment to increase plasma $HCO3$ ⁻ to the normal ranges is >15% in patients with FS.

> Fractional excretion of $HCO₃$ -% = $[(UHCO_{3^-}/PHCO_{3^-})/(Ucr/Per)] \times 100$

(HCO3⁻; bicarbonate, cr; creatinine, U; urine, P; plasma)

Acidification in the distal tubule is usually normal or impaired in association with chronic hypokalemia or toxic effect on distal tubules due to the original disorder in patients with FS.

Sodium and Potassium Losses

60–80% of filtered load of $Na⁺$ is reabsorbed in the proximal tubules in the normal condition. Renal Na⁺ reabsorption in the proximal tubules decreased in patients with FS. It leads to hyponatremia, hypotension, and dehydration. Hypokalemia is a secondary phenomenon. Increased delivery of $Na⁺$ into the distal tubules and activation of the renin-angiotensin system secondary to hypovolemia cause potassium (K^+) wasting in the distal tubules. Severe hypokalemia can cause sudden death.

Hypercalciuria

Hypercalciuria is a common finding in patients with FS due to several original diseases. Defective endocytosis of parathyroid hormone (PTH) in patients with Dent disease resulting in its persistence in the lumen of the proximal tubule stimulates 25-hyroxyvitamin D3 1-hydroxylase to produce more 1,25-dihydroxyvitamin D3, raising serum levels of this vitamin. 25-hydroxyvitamin D3 is presented to 25-hydroxyvitamin D3 1-hydroxylase in the form of a

complex with the vitamin D3-binding protein. As this complex is lost in the urine as a result of defective endocytosis leading to LMW proteinuria, the precursor 25-hydroxyvitamin D3 could be in short supply. The overall outcome of increased 1, 25-dihydroxyvitamin D3 levels may depend on the delicate balance between these processes. The slightly elevated serum levels of 1, 25-dihydroxyvitamin D3 in patients with FS can lead to increased intestinal Ca^{2+} reabsorption which will lead to hypercalciuria (absorptive hypercalciuria) ([29\)](#page-22-0). Hypercalciuria is rarely associated with nephrolithiasis in FS, possibly because of the polyuria and alkalized urine. However, patients with Dent disease manifest hypercalciuria and nephrolithiasis.

Hyperuricosuria (Uricosuria)

Uric acid (urate) is the end product of purine metabolism in humans. Because of its small molecular size ($MW =$ 126 D), uric acid is freely filtered from the glomerulus. Then, 90–95% of filtered load of uric acid is eventually reabsorbed in the proximal tubules. A four-component hypothesis has been proposed to explain the renal uric acid transport mechanism; it includes glomerular filtration, presecretory reabsorption, secretion, and postsecretory reabsorption [\(31](#page-22-0)). Hyperuricosuria is often present in FS, leading to secondary hypouricemia ($\langle 2 \rangle$ mg dL⁻¹) [\(32](#page-22-0)). A voltage-sensitive uric acid pathway and uric acid exchangers are located at both luminal and basolateral membranes of proximal tubule cells ([33\)](#page-23-0). Uric acidanion exchanger (URAT1) that reabsorbs uric acid from the lumen of the proximal tubules in the luminal membrane of proximal tubules regulates serum uric acid levels. This uric acid-anion exchanger can be disturbed in patients with FS. Defective URAT1 is a predominant cause of the patients with renal hypouricemia who manifest acute renal failure after exercise ([34,](#page-23-0) [35\)](#page-23-0). Hexose transporter gene (SLC2A9) is identified as a cause of gout and hyperuricemia [\(36](#page-23-0)). This transporter transports both fructose and uric acid. SLC2A9 produces two isoforms by alternative splicing; the long isoform is expressed in basolateral membrane of proximal tubular cells and the short isoform is expressed in apical membrane of proximal tubular cells. This hexose transporter can be affected in patients with FS. Uric acid is a selective antioxidant, capable especially of reaction with hydroxyradicals and hypochlorous acid, itself being converted to innocuous products such as allantoin, allantoate, glyoxylate, urea, and oxalate ([37\)](#page-23-0).

Proteinuria (LMW Proteinuria)

The proximal tubules have a high capacity for uptake of filtered proteins from the glomerulus. The cut off molecular weight for filtration of plasma proteins is assumed to be in the range of 65 KD (kilodaltons) that corresponds to the molecular weight of serum albumin. However, small amount of larger weight proteins including gammaglobulin are filtered from glomerulus in the normal condition. Albumin and LMW proteins (MW<45,000 D) filtered in the glomerulus are considered to be the major source of urinary albumin and LMW proteins. Filtration of albumin and LMW proteins are followed by tubular reabsorption, and thus the resulting albuminuria and LMW proteinuria reflect the combined contribution of these two processes. Dysfunction of both these processes may result in increased urinary excretion of albumin and LMW proteins, and both glomerular injury and tubular impairment have been implicated in the initial events leading to proteinuria. In FS, proteinuria is predominantly caused by the dysfunction of reabsorption in the proximal tubules. Adolescent patients with FS due to Dent disease or Lowe syndrome excrete greatly increased amounts of proteins (1,740 \pm 660 mg/day) and peptide $(446 \pm 145 \text{ mg/day})$. LMW proteins ranging from 2 to 5 KD were present in 12.9 \pm 3.9-fold excess in FS compared with normal urine [\(38](#page-23-0)). The micropuncture technique in dogs revealed that the filtered load of albumin was 50 μ g m⁻² suggesting that a filtered load of albumin is 9 g/day in normal humans [\(39](#page-23-0)). However, urinary excretion of protein is less than 0.1–0.15 g/day in the normal condition. Numerous filtered proteins including albumin and LMW proteins from glomerulus are bound to megalin and cubilin in the luminal membrane of proximal tubules. Then, the protein-receptor complex is incorporated into the endosome. The ligand and receptor are disassociated in the endosome; the receptor is recycled back to the luminal membrane and the reabsorbed proteins go into lysosome for further processing. Megalin is a 600 KD glycoprotein and a member of the low-density lipoprotein receptor family. Megalin is expressed in the proximal tubule brush-border and luminal endocytic apparatus. Megalin binds to a number of structurally very different proteins. It contains a large amino-terminal, extracellular domain, a single transmembrane domain and a short carboxy-terminal cytoplasmic tail (\bullet [Fig. 42-1](#page-1-0)). Cubilin is a multiligand, endocytic receptor. It is a 460 KD protein with little structural homology to known, endocytic receptor $(① Fig. 42-1)$ $(① Fig. 42-1)$ $(① Fig. 42-1)$. Cubilin is expressed in the proximal tubule brush-border and luminal endocytic apparatus. Megalin involved in

albumin reabsorption directly as a receptor for albumin, and/or indirectly by affecting the expression and/or endocytic function of cubilin ([40,](#page-23-0) [41](#page-23-0)). Megalin's expression was decreased in patients with Dent disease. Acidification defect due to endosomal defective ClC-5 in patients with Dent disease disturbs the recycling from intracellular endosome into luminal membrane of the proximal tubule resulting in megalin and cubilin deficiency in the luminal membrane of the proximal tubule.

Etiologies

The causes of FS are divided into three main categories; hereditary, acquired, and exogenous substances ([>](#page-6-0) [Table 42-1](#page-6-0)). Most of the hereditary FS occurs as one of the manifestations of congenital metabolic disorders or as sporadic or familial disorders. Acquired forms are derived from immunological reactions, nephrotic syndrome or accumulated abnormal proteins. Exogenous substances are composed of drugs, chemical compounds, and heavy metals.

Hereditary Fanconi Syndrome

Dent Disease

Dent disease is an X-linked proximal tubulopathy characterized by LMW protinuria, hypercalciuria, nephrocalcinosis, nephrolithiasis, and eventual end stage renal failure. Hypophosphatemic rickets and metabolic acidosis are sometimes seen [\(42](#page-23-0), [43\)](#page-23-0). Almost all of the patients are males. Adult patients with Dent disease manifest FS. However, children with Dent disease often manifest LMW proteinuria and one or two of the manifestations due to proximal tubular dysfunction and this is called partial FS [\(44](#page-23-0)). They usually fall into end stage renal failure by the age of 40s. However, this is highly variable, and one third of patients with Dent disease will not develop end stage renal failure. Patients with Dent disease never manifest extrarenal manifestations, except for rickets, which may itself be a consequence of phosphaturia. School children with Dent disease manifest proteinuria. A lot of school children with Dent disease are detected as proteinuria by school urine mass screening program in Japan, and it was called as idiopathic low molecular weight proteinuria [\(45](#page-23-0), [46](#page-23-0)). Carrier females are often manifest less severe LMW proteinuria and hypercalciuria, depending on X-chromosome inactivation, but they rarely develop clinically significant problems.

Causes of Fanconi syndrome

Dent disease is associated with inactivating mutations in CLCN5 gene, which encodes 746 amino acids renal specific chloride channel-5 (ClC-5) [\(8,](#page-22-0) 47-49). ClC-5 belongs to the family of voltage-dependent chloride channels, which function as homodimeric proteins. ClC-5 is $\text{co-expressed with the vacuolar } H^+\text{-ATPase}$ and plays a key role in endosomal acidification that is a crucial function in the receptor-mediated endocytic pathway ([50\)](#page-23-0). More than 80 distinct CLCN5 mutations are reported in patients with Dent disease. They are nonsense, missense, frameshift, splice-site, insertional, and deletional mutations, which result in total or partial loss of function. There are no genotype-phenotype correlations as various mutations are associated with different clinical phenotypes, even within the same family.

Numerous filtered proteins are bound to megalin and cubilin in the luminal membrane of proximal tubules, and the protein-receptor complex is incorporated into the endosome. The ligand and receptor are disassociated in the endosome; the receptor is recycled back to the luminal membrane and the reabsorbed proteins go into lysosome for further processing. This disassociation is dependent on acidification of the lumen of endosome by increased concentration of H^+ and Cl^- due to the function of H^+ -ATPase and ClC-5 chloride channel. An abnormal endocytosis pathway due to ClC-5 dysfunction disturbs the recycling of megalin and cubilin, the back to the luminal membrane, and the expression of megalin and cubilin in the luminal membrane of proximal tubules, leading to LMW proteinuria, hypercalciuria, hyperphosphaturia, and nephrolithiasis. Proper acidification is also important for protein degradation in the endosome. Immunohistochemical analysis of proximal tubule cells in patients with Dent disease revealed an inverted polarity of the H⁺-ATPase, with redistribution to basolateral regions, suggesting that the loss of ClC-5 channel alters the function of components that co-distribute and physically interact with it (51) (51) .

Total urine protein ranges from 0.5–2.5 g a day, but may reach 4 g or higher in patients with Dent disease ([45,](#page-23-0) [52](#page-23-0)). More than 60% of the filtered proteins are LMW proteins with molecular weight less than 45 KD (\bigcirc [Fig. 42-3](#page-7-0)). Nephrotic syndrome does not occur. LMW proteinuria is the most consistent and one of the earliest presenting abnormalities. Urinary beta 2-microglobulin, a LMW protein ($MW = 11.6$ KD), is excreted in amounts 100–300 times the upper limit of the normal. Albumin is also excreted in the urine. The pattern of proteins representing the increased excretion of several LMW proteins as well as albumin ($MW = 65 KD$) is termed as tubular proteinuria [\(52](#page-23-0), [53](#page-23-0)). The terms of LMW proteinuria and tubular proteinuria have usually been used interchangeably ([53\)](#page-23-0).

Patients with Dent disease manifest hypercalciuria in the range of 4–10 mg kg^{-1} of body weight a day in children and $4-6$ mg kg^{-1} of body weight a day in adults [\(54](#page-23-0)). Nephrocalcinosis is also found in children $(\odot$ [Fig. 42-4](#page-8-0)). Defective endocytosis of parathyroid hormone (PTH) in patients with Dent disease resulting in its persistence in the lumen of the proximal tubule stimulates 25-hydroxyvitamin D3 1-hydroxylase to produce more 1,25-dihydroxyvitamin D3 resulting in the increased serum levels of this vitamin. 25-hydroxyvitamin D3 is presented to 25-hydroxyvitamin D3 1-hydroxylase in the form of a complex with the vitamin D3-binding protein. As this complex is lost in the urine as a result of defective endocytosis leading to LMW proteinuria, the precursor

Electrophoresis of urine from the family members of Dent disease on the polyacrylamide gel and stained by sliver representing LMW proteinuria. Lanes 1 and 2, molecular markers; Lane 3, 12-year-old boy with Dent disease; Lane 4, his mother; Lane 5, his father.

25-hydroxyvitamin D3 could be in short supply. The overall outcome of increased 1, 25-dihydroxyvitamin D3 levels may depend on the delicate balance between these processes. The slightly elevated serum levels of 1, 25-dihydroxyvitamin D3 in patients with Dent disease can lead to increased intestinal Ca^{2+} reabsorption resulting in hypercalciuria (absorptive hypercalciuria). In fact, 70% of the patients with CLCN5 mutations manifest hypercalciuria, even though some of these do exhibit nephrocalcinosis ([55\)](#page-23-0). This may be explained by the CLCN5 knock-out mice model; ClC-5 disruption promotes calcium crystal agglomeration, as well as a redistribution of the crystal-binding molecule annexin A2, in collecting duct epithelial cells [\(56](#page-23-0)).

The onset of renal insufficiency and progression to end-stage renal failure are quite variable. Significant decrease of glomerular filtration rate is seen in children with Dent disease, and the patients fall into end-stage renal failure by the end of the age of 40s. Renal biopsy demonstrates normal or focal global glomerulosclerosis with tubular atrophy, tubular dilatation, and interstitial infiltration of monocytes. Medullary nephrocalcinosis is a significant feature in patients with Dent disease. Patients less than 5 years of age manifest medullary nephrocalcinosis. The precise mechanism of progressive renal failure

is not known in patients with Dent disease. Nephrocalcinosis can be a candidate to disturb the glomerular filtration rate. High urinary concentrations of potentially bioactive proteins including insulin, insulin like growth factor-1 (IGF-1), and the chemokine monocyte chemoattractant protein-1 (MCP-1) may contribute to interstitial fibrosis that will lead to progressive renal failure in patients with Dent disease [\(57](#page-23-0)). Generalized proximal tubule dysfunction is associated with increased cell proliferation, dedifferentiation, and oxidative stress resulting in interstitial fibrosis and eventual renal failure ([58\)](#page-23-0). Patients and carrier females often manifest nephrolithiasis, and renal stone is calcium phosphate stone that is also seen in patients with distal renal tubular acidosis.

Dent disease is genetically heterogeneous. Mutations in the OCRL1 gene are identified in a subset of patients with the Dent disease phenotype [\(59](#page-23-0)). Unlike patients with typical Lowe syndrome, typical facial features, mental retardation, metabolic acidosis, and ocular abnormalities are usually absent in patients with Dent disease who have OCRL1 mutation. The phosphoinositol 4,5-bisphosphate phosphatase (PIP_2 5-phosphatase) activity is markedly reduced in skin fibroblasts cultured from patients with Dent disease due to OCRL1 mutations, and protein expression, measured by Western blotting, is reduced or absent. PIP_2 5-phosphatase participates the trafficking and recycling of endosome in the proximal tubules. Defective PIP_2 5-phosphatase activity can lead to endosomal dysfunction leading to LMW proteinuria. Unlike the patients with typical Lowe syndrome, none of patients have metabolic acidosis. These observations and findings suggest that OCRL1 mutations can cause the isolated renal phenotype of Dent disease and affected individuals lack the cataracts, typical facial features, renal tubular acidosis, and neurologic abnormalities that are characteristic to Lowe syndrome. It is difficult to explain that OCRL1 mutations can cause the isolated renal phenotype of Dent disease. However, it is possible that another phosphatase, INPP5B, which shares amino acid homology with OCRL1, can compensate phosphatase activity in patients with Dent disease due to OCRL1 mutations.

There are no specific interventions at present that will change the natural course of renal manifestations and progressive renal failure in patients with Dent disease. Hypercalciuria is corrected by thiazide diuretics therapy in doses similar to effective doses for idiopathic hypercalciuria, presumably by stimulating the reabsorption of calcium in the distal convoluted tubule, where ClC-5 channel is not expressed (60) (60) . However, this is not a long-term study which provides the evidence that it is Abdominal CT demonstrating bilateral medullary nephrocalcinosis in a 12-year-old boy with Dent disease.

effective to prevent or delay the progression of end stage renal failure. In animal experiment using *clcn5* (mouse chloride channel 5 gene) knock-out mice, high citrate diets can delay the progression of nephrocalcinosis and end stage renal failure ([61](#page-23-0), [62\)](#page-23-0). Treatment with an angiotensin-converting enzyme (ACE) inhibitor or angiotensin receptor blocker (ARB) may delay progression of end stage renal failure ([63\)](#page-23-0).

Lowe Syndrome

The oculocerebrorenal syndrome of Lowe (OCRL) is a rare X-linked disorder that is characterized by a complex phenotype that involves major abnormalities of the eyes (particularly congenital cataracts), central nervous system abnormality and FS ([64\)](#page-23-0). Cataracts are recognized in all the male patients. Lens opacities are present in all carriers of females by slit-lamp examination and this is a reliable screen for carriers ([65\)](#page-23-0). Other ocular abnormalities include glaucoma, microphthalmos, and corneal keloid formation. Visual acuity is frequently disturbed. Central nervous system abnormalities include infantile hypotonia (floppy infant), areflexia, and mental retardation. The patients manifest mild ventriculomegaly and cysts in the periventricular regions in the brain. Status epilepticus is sometimes recognized. FS is a major clinical feature and occurs in the 1st year of life, but the severity and age of onset vary. The patients manifest LMW proteinuria, glucosuria, aminoaciduria, hyperphosphaturia, hypercalciuria, hypophosphatemic rickets, hypercholoremic

metabolic acidosis, and progressive renal insufficiency [\(66](#page-23-0)). Renal insufficiency progresses at variable rate among patients, and it progresses to end-stage renal failure by 30s or 40s.

Like in patients with Dent disease, megalin is nearly absent from the urine in patients with Lowe syndrome suggesting of the decreased expression of megalin in the luminal membrane of the proximal tubule cells. Thus, urinary excretion of retinol-binding protein and the lysosomal enzyme N-acetyl-glucosaminidase are significantly increased in young boys with OCRL [\(67](#page-23-0)).

The gene (OCRL1) that is responsible for OCRL encodes a 105 KD Golgi protein with phosphoinositol 4,5-bisphosphate phosphatase (PIP₂ 5-phosphatase) activity (68) (68) . PIP₂ 5-phosphatase is mainly a lipid phosphatase that may control cellular levels of a critical metabolite, phosphatidylinositol 4,5-bisphosphate, and is involved in the inositol phosphatase signaling pathway (69) (69) . PIP₂ 5-phosphatase is present in cultured skin fibroblast and it is not present in peripheral blood cells. $PIP₂$ 5-phosphatase activity is markedly reduced in fibroblasts from patients with Lowe syndrome [\(70](#page-23-0)). However, this biochemical test for carrier diagnosis is not reliable; lyonization produces a highly variable pattern of tissue expression in females.

Deficiency of PIP_2 5-phosphatase leads to cellular accumulation of its substrate PIP_2 . PIP_2 accumulates in lysosomal membrane (71) (71) . PIP₂ is involved in signal transduction, vesicle trafficking and actin polymerization. Absence of PIP_2 5-phosphatase activity leads to a reduction in the number and length of actin stress fibers, a tendency of actin fibers to depolymerize when provoked, and an abnormal distribution of two actin-binding protein gelsolin and alpha-actinin. This disruption of actin function has significant effects on epithelial function through disrupting cell–cell contacts such as tight junctions or adherent junctions or by altering membrane trafficking such as transport proteins [\(72](#page-23-0)). Trans-Golgi dysfunction or altered actin polymerization can explain FS in patients with Lowe syndrome. PIP_2 5-phosphatase is localized to endosome and Golgi membranes along with clathrin, giantin, the mannose 6-phosphate receptor, transferrin, and the early endosomal antigen 1 marker. PIP₂ 5-phosphatase interacts with clathrin terminal domain and the clathrin adaptor protein AP-2. This suggests a role for PIP_2 5-phosphatase in endosomal receptor trafficking and sorting ([73,](#page-23-0) [74\)](#page-23-0). OCRL1 is present throughout the early endocytic pathway, including in endocytic clathirin-coated pits, and demonstrate a connection between OCRL1 and adaptor molecules implicated in the endocytic trafficking of receptor in the kidney ([75\)](#page-24-0).

OCRL1 has a C-terminal RhoGAP domain. OCRL1 encodes a PIP_2 5-phosphatase activity that binds to Rac GTPase. Activated Rac GTPase stably associates with the OCRL1 RhoGAP domain. In this sense, the protein encoded in OCRL1 can play a bifunctional role. Loss of OCRL1 RhoGAP domain and the resulting alteration in Rho pathways may contribute to mental retardation in Lowe syndrome, as observed in other forms of X-linked mental retardation ([76\)](#page-24-0).

OCRL1 mutations can cause the isolated renal phenotype of Dent disease and affected individuals lack the cataracts, typical facial features, renal tubular acidosis, and neurologic abnormalities that are characteristic to Lowe syndrome. It is difficult to explain that OCRL1 mutations can cause the isolated renal phenotype of Dent disease. However, another phosphatase, INPP5B, which shares amino acid homology with OCRL1, can compensate phosphatase activity in patients with Dent disease due to OCRL mutations.

More than 70 OCRL1 mutations have been described in patients with Lowe syndrome; nearly all are clustered in exons 10–23, especially exon 15, and almost none are found in exons $1-9$ [\(70](#page-23-0)).

Treatment is supportive and includes taking care of ocular manifestations, anticonvulsants, speech therapy, and dental complications. The eye abnormalities usually require therapy early in life. Bicarbonate therapy is usually necessary at a dose of 2–3 mmol kg^{-1} of body weight a day every 6–8 h. Sodium or potassium phosphate can be given in amounts of 1–4 g a day for phosphate depletion and if unsuccessful, vitamin D can be given.

Mitochondriopathies

The mitochondria (mt) have a major role in fatty acid oxidation, tricarboxylic acid cycle, urea cycle, and ATP production through the process of oxidative phosphorylation. Oxidative phosphorylation occurs at the level of the respiratory chain in the inner membrane of the mt [\(77](#page-24-0)). The respiratory chain comprises five components ([>](#page-10-0) [Fig. 42-5](#page-10-0)). Complex I (NADH-coenzyme Q reductase) carries reducing equivalents from NADH to coenzyme Q and consists of different polypeptides, seven of which are encoded by mitochondrial DNA (mtDNA). Complex II (succinate-coenzyme Q reductase) carries reducing equivalents from $FADH₂$ to coenzyme Q and contains five polypeptides that are all encoded only by mtDNA. Complex III (reduced coenzyme Q-cytochrome c reductase) carries reducing equivalents from coenzyme Q to cytochrome c and contains 11 subunits, one of which is encoded by mtDNA. Complex IV (cytochrome c oxidase) transfers reducing equivalents from cytochrome c to oxygen. This complex is composed of cytochromes a and a3, and 13 protein subunits, three of which are encoded by mtDNA. The fifth complex is ATP synthetase.

Oxidative phosphorylation consists of oxidative reactions that lead to oxygen consumption and phosphorylation of ADP to ATP. Each mt have its own 2–10 mtDNA. MtDNA genome is a 16.5 kb circular double stranded DNA with an asymmetrical base composition. The heavy strand contains more guanine residues while the light strand contains more cytosine residues. The mtDNA contains 37 genes; 13 encode for polypeptides of the respiratory chain, 2 for the ribosomal RNAs and 22 for transfer RNAs [\(78](#page-24-0)). Mitochondrial injury is due to congenital insults or may be the result of secondary events. Genetic defects of one or several polypeptide enzyme complexes of the oxidative phosphorylation system in the mt DNA or nuclear DNA which encodes structural or functional mitochondrial proteins can give rise to mitochondriopathies (mitochondrial cytopathies, mitochondrial diseases) ([79\)](#page-24-0).

Mitochondriopathies are multi-systemic disease and may begin at any age. Affected organs are diverse including central nervous system, muscle, liver, heart, kidney, gut, endocrine system, bone marrow, ear, eye, and skin ([>](#page-11-0) [Table 42-2](#page-11-0)). They display extreme heterogeneity, and make unpredictable the extent and manifestations of disease presentation ([80\)](#page-24-0). With the course of the disease, the numbers of organs involved are increased.

The screening for mitochondriopathies includes the determination of plasma lactate, pyruvate, keton bodies and their ratios in fasted and fed patients, polarographic, and spectrometric studies to evaluate the different

Map of mitochondria genome. Regions encoding cytochrome b (ctb b), various subunits of NADH-coenzyme Q reductase (ND), cytochrome c oxidase (COX), ATPase, and ribosomal RNAs (rRNA) are indicated (Niaudet P, Rötig A. The kidney in mitochondrial cytopathies. Kidney Int 1997;51:1000–1007).

enzymatic complexes of the respiratory chain, muscle histologic studies, and genetic analysis [\(4\)](#page-22-0).

Renal disease may be the first sign of mitochondriopathies, or it may appear simultaneously with neurological and neuromuscular signs ([81\)](#page-24-0). FS is particularly frequent in newborns, infants, and young children ([82–84\)](#page-24-0), whereas tubulointerstitial nephropathy is more frequent in children and young adults [\(85](#page-24-0), [86](#page-24-0)), and can be associated with hereditary focal segmental glomerulosclerosis due to the mitochondrial transfer RNA gene mutation [\(87–89](#page-24-0)) and collapsing glomerulopathy due to the mutations in the gene COQ2 encoding the parahydroxybenzoate-polyprenyl-transferase enzyme of the CoQ10 synthesis pathway (CoQ2 nephropathy) and in the gene PDSS2 encoding for decaprenyl diphosphate synthase ([90,](#page-24-0) [91\)](#page-24-0).

Most patients with FS due to mitochondriopathies manifest moderate FS including failure to thrive, dehydration, aminoaciduria, glucosuria, proteinuria, LMW proteinuria, phoshaturia, uricosuria, hypercalciuria, and bicarbonaturia. Many patients manifest FS by the age of 2 years. Extra-renal manifestations including neurological symptoms, myopathy, hepatic dysfunction, clinical features of Pearson syndrome, partial adrenal insufficiency, cardiac involvement, diabetes mellitus, deafness, and ophthalmoplegia often manifest in the patients ([12,](#page-22-0) [92](#page-24-0), [93](#page-24-0)). Patients who manifested proximal tubular acidosis with hypercalciuria or Bartter syndrome were described ([94,](#page-24-0) [95\)](#page-24-0). Some patients manifest progressive renal failure ([96\)](#page-24-0). Histological analysis reveals tubular dilatations, tubular atrophy and cytoplasmic vacuolization of the tubules. Bizarre giant mitochondria are frequently observed ([97\)](#page-24-0).

No satisfactory treatment is presently available to alter the course of mitochondriopathies. The treatment is mainly symptomatic: supplements of sodium bicarbonate, potassium, vitamin D3, phosphate, and water are necessary. Carnitine is given in case of secondary carnitine deficiency. It includes avoidance of drugs that interfere with the respiratory chain such as valproate and barbiturates, or that inhibit mitochondrial protein synthesis such as tetracyclines and chloramphenicol. Dietary recommendations include a high lipid and low carbohydrate diet in patients with complex I deficiency. Hypercaloric diet and parenteral nutrition should be avoided in these patients.

D Table 42-2

Clinical symptoms in patients with mitochondriopathies

Cystinosis

Cystinosis is reviewed in detail in Chapter 41 of this textbook, and therefore only a short description is included here. Cystinosis is an autosomal recessive lysosomal storage disorder characterized by an accumulation of cystine, the disulfide of the amino acid cysteine, in the systemic organs, notably kidney, cornea, bone marrow, thyroid, lymph nodes, liver, and spleen [\(98](#page-24-0)). Renal manifestations dominate the clinical presentation and course in infantile cystinosis. Cystinosis is the most common familial form of the FS in Western countries. Many patients, particularly in North America, have blonde or reddish-blonde hair. Other organs frequently affected include the cornea and thyroid, causing painful photophobia and hypothyroidism respectively.

Various clinical forms of the disease exist and are based on age at onset and severity of the symptoms (\odot [Table 42-3](#page-12-0)). The most severe form, infantile cystinosis, is manifested by failure to thrive, polyuria, polydypsia, dehydration, fluid and electrolyte loss, aminoaciduria, glucosuria, phosphaturia, renal tubular acidosis between 6 and 12 months of age. Some of the patients may develop vitamin D-resistant rickets due to phosphaturia and manifest severe growth failure. Renal function is generally normal at presentation. However, subsequent glomerular impairment leads to focal segmental glomerulosclerosis and eventually to end stage renal failure by 10 years of age without treatment [\(99](#page-24-0)). Patients with infantile cystinosis manifest FS, including hyperchloremic metabolic acidosis, aminoaciduria, hypokalemia, hypophosphatemia, glucosuria, and phosphaturia. There have been several patients of nephropathic cystinosis presenting with features of secondary Bartter syndrome (hypokalemia, hyperchloremic metabolic alkalosis, hyperreninemia, and hyperaldosteronism), suggest-ing abnormalities of Na⁺ and Cl⁻ reabsorption [\(100](#page-24-0), [101\)](#page-24-0). Patients with cystinosis often manifest medullary nephrocalcinosis ([102\)](#page-24-0).

Renal histopathologic changes in infantile cystinosis include severe lesions of proximal tubules; typical alterations to the glomerular podocytes, which become multinucleated giant cells; and the presence of cystine crystals, mostly in interstitial cells and podocytes [\(103](#page-24-0)). The proximal tubule is the first clinical target of the disease, but cystine crystals are rarely found in the tubular cells of patients with cystinosis. Cystine crystal deposition in the cornea leads to photophobia. Continuous widespread cystine accumulation eventually leads to rickets and retinal, endocrinologic (hypothyroidism and impaired glucose tolerance), hepatic, gastrointestinal, muscular, and neurological abnormalities.

Two less severe and less common form of cystinosis are juvenile (or late-onset) and ocular cystinosis. Patients with juvenile cystinosis manifest glomerular impairment between 12 and 15 years of age but do not suffer from severe tubulopathy or growth failure. Progression to end stage renal failure is slow and occurs at

Clinical manifestations of infantile cystinosis

Adapted from Gahl et al. [\(114\)](#page-24-0)

^amay be transient and coexist with common manifestations

variable ages ([104](#page-24-0)). Patients with ocular cystinosis do not involve kidney.

Infantile cystinosis is caused by mutations of the CNTS gene encoding cystinosin, a lysosomal transport protein, leading to complete abolition of cystine transport ([105](#page-24-0)). Cystinosin has 367 amino acids and seven transmembrane domains. Cystine transport is dependent on the pH gradient, and the transport of cystine out of the lysosome is driven by the high H^+ content within the lysosomal lumen that is produced by the activity of the H⁺-ATPase. A range of mutations in CNTS gene has been described, but a single mutation, a 57-kb intragenic deletion, accounts for as many as three quarters of all European cases (105) . The adolescent and ocular forms have one severe and one mild CTNS mutation, leading to reduced transport activity. The sparing of the kidney in patients with ocular cystinosis reflects tissue-specific expression of splicing factors, or the increased endogenous level of CTNS mRNA normally seen in the kidney [\(106\)](#page-24-0). Individuals who are heterozygous for severe CTNS mutations reveal elevated levels of leukocyte cystine but are completely asymptomatic.

The pathophysiology of tubular cystine transport defects in patients with cystinosis is poorly understood, reflecting of an animal model for the disease. Knock-out mice model lacking cystinosin gene do not manifest signs of FS, despite accumulation of lysosomal cystine in the proximal tubules ([107\)](#page-24-0). Cystine-loaded proximal tubular cells demonstrate loss of free phosphate and defective ATP production and inhibition of Na⁺-dependent transporters [\(108\)](#page-24-0). ATP depletion can reduce proximal tubular Na⁺ , K⁺-ATPase activity leading to increased Na⁺ delivery into the distal tubules and Bartter syndrome ([101](#page-24-0)). A cell culture demonstrated that cells accumulated with intracellular cystine undergo apoptosis at a rate two- to fourfold higher than controls ([109\)](#page-24-0). Another works suggests that increased oxidative stress and altered redox status in proximal tubule cells cultured from the urine of patients with cystinosis are associated with proximal tubule dysfunction [\(110\)](#page-24-0).

The diagnosis of cystinosis is confirmed by demonstrating elevated cystine levels in peripheral leukocytes [\(97](#page-24-0)). Corneal crystals detected by slit-lamp examination are diagnostic in childhood cystinosis because these crystals are not seen in patients with other hereditary FS. However, this finding is not sensitive for early diagnosis. The renal pathologic findings in infantile cystinosis consist of a chronic tubulointerstitial nephropathy, with characteristic multinucleated podocytes and intracellular crystalline inclusions in interstitial histiocytes ([111](#page-24-0)). Although numerous multinucleated podocytes are the most characteristic pathologic findings, they are not found in the sclerotic glomeruli and detected only in low frequency $(4%). The cystine crystals are birefrin$ gent under polarized light in only alcohol-fixed tissue or in unfixed frozen tissue, because they are water-soluble and not retained in the tissue after routine histologic preparation with aqueous solutions ([112](#page-24-0)).

The management and treatment for infantile cystinosis involve supportive therapy to maintain fluid balance and replace electrolyte losses at initial presentation. Early diagnosis and oral cysteamine, a cystine-depleting agent, can delay the progression of end stage renal failure and other organ involvement. Oral cysteamine therapy given at doses of 60–90 mg kg^{-1} of body weight (or between 1.3 and 1.95 g m^{-2}) a day divided every 6 h generally achieves approximately 90% depletion of cellular cystine, as measured in circulating leukocytes (<1.0 nmol halfcystine/mg protein) ([113](#page-24-0)). The dosage recommended for adults is 500 mg every 6 h, but higher dosages are often required to achieve satisfactory cystine depletion. On the basis of its beneficial effects in maintaining thyroid function and depleting muscle of cystine, oral cysteamine

therapy should continue in patients after renal transplantation to help preserve other organs. Administration of 0.55% cysteamine eye-drops, given 6–12 times a day, can dissolve corneal cystine crystals and lessen visual symptoms ([114](#page-24-0)). Other therapies to supply potassium, alkalinizing agents including citrate or bicarbonate, phosphate, and vitamin D3 are required. When the growth velocity has not improved and the patient remains below the 3rd percentile for height after one year of therapy, growth hormone therapy may be considered.

Galactosemia

Galactosemia is an autosomal recessive disease of galactose metabolism. Nursing infants must move large amounts of galactose through Leloir pathway in order to utilize the carbon skeletons for energy (\bullet [Fig. 42-6](#page-14-0)). Galactose is the preferred carbon source in mammalian neonates, since it is incorporated into glycogen more efficiently than is glucose [\(115\)](#page-24-0).

The most frequent form is classic galactosemia that is due to the deficient activity of galactose-1-phosphate uridyl-transferase (GALT) encoded by GALT1 ([116](#page-24-0)). GALT catalyzes the reaction of galactose-1-phosphate (gal-1-p) plus uridine diphosphate glucose to uridine diphosphate galactose plus glucose-phosphate. Uridine diphophate galactose can be further metabolized to either glucose or $CO₂$ and $H₂O$ via glycolysis. Milk is a major source of galactose. Accumulated gal-1-p due to defective GALT and exposure to galactose lead to acute deterioration of multiple organ systems, including liver, kidney, ovary, brain, and eye. Affected infant patients manifest vomiting, diarrhea, failure to thrive, developmental delay, liver dysfunction, coagulopathy, renal tubular dysfunction, cerebral edema, vitreous hemorrhage, and Escherichia coli sepsis. They sometimes manifest jaundice and unconjugated hyperbilirubinemia and may have severe hemolysis. Liver damage leads to hepatomegaly and cirrhosis that is potentially lethal. Neonatal screening program includes galactosemia, anticipating that early detection and intervention would prevent long-term complications such as mental retardation, premature ovarian failure, and speech delay. Although a galactoserestricted diet prevents the neonatal death, many well-treated patients continue to develop debilitating complications [\(117](#page-24-0), [118](#page-24-0)). Clinically evident speech delay and cerebellar signs are more frequent than other findings. Premature ovarian failure is nearly universal in females with galactosemia. The predominant manifestation due to kidney damage is FS including hyperaminoaciduria, LMW proteinuria, hyperphosphaturia, and bicarbonaturia ([118](#page-24-0)). Patients placed on a galactose-restricted diet are never truly free of galactose insult, as a significant amount of galactose is found in non-dairy foodstuffs such as vegetables and fruits [\(119,](#page-25-0) [120](#page-25-0)). More importantly, galctose moieties can be produced endogenously from UDP-glucose via the UDP-4-galactose epimerase reaction, and natural turnover of glycoproteins/glycolipid; the rate of endogenous galactose synthesis ranges from 0.53–1.05 mg kg^{-1} of body weight a day [\(121,](#page-25-0) [122\)](#page-25-0). Once the lactose is formed intracellularly, it will be converted to gal-1-p by GALT. The less common form of galactosemia is a deficiency of galactose kinase (GALK), which forms gal-1-p from galactose. These patients do not manifest either the acute toxicity syndrome or chronic complications seen in patients with classic galactosemia. They manifest cataracts. Since GALK-deficient patients do not accumulate gal-1-p in their tissues, gal-1-p is considered to play a significant role in the pathogenesis of classic galactosemia [\(123](#page-25-0), [124\)](#page-25-0). GALT deficiency results in accumulation of toxic galactose leading to the unfolded proteins, altered calcium homeostasis and subsequently endoplasmic reticulum (ER) stress [\(125\)](#page-25-0). ER stress caused by GALT-deficiency might contribute to accelerated apoptosis seen in the granulose cells maturing follicles in galactosemic females, leading to premature ovarian failure [\(126\)](#page-25-0). Formation of galactitol from galactose by aldose reductase has been proposed as a pathogenetic mechanism and is at least responsible for cataract formation.

The diagnosis is suggested by galactose or galactose 1-phosphate in serum, or in the urine. The diagnosis is confirmed by demonstrating deficient GALT activity in red blood cells, fibroblasts, leukocytes, or hepatocytes.

Treatment of this disorder is elimination of galactose from the diet. Acute symptoms and signs resolve within a few days after starting the diet therapy. However, developmental delay, speech disturbance, ovarian dysfunction, and growth retardation are common outcomes in this disorder [\(127\)](#page-25-0).

Hereditary Fructose Intolerance

Hereditary fructose intolerance (HFI) is an autosomal recessive disorder caused by a deficiency of aldolase B, an enzyme of liver, intestine, and renal cortex catalyzing the metabolism of fructose of exogenous origin [\(128\)](#page-25-0). Frequency of HFI is estimated at 1 in 20,000 live births. Aldolase B catalyses the specific and reversible cleavage of

Composite diagram of the Leloir pathway and uridine diphosphate (UDP)-glucose pyrophosphorylase pathway. Galk, galactokinase; galt, galactose-1-phosphate uridyltransferase; Gale, UDP-galactose 4-epimerase; Hex, hexokinase; Pgm, phosphoglucomutase; Ugp, UDP-glucose pyrophosphorylase (Leslie ND. Insights into pathogenesis of galatosemia. Annu Rev Nutr 2003;23:59–80).

fructose-1,6-bisphophate (FBP) and fructose-1-phosphate (F1P) into dihydroxyacetone phosphate and D-glyceraldehyde-3-phophate, or D-glyceraldehyde, respectively. Aldolase B is equally active with FBP and F1P, whereas aldolase A and aldolase C, the other two vertebrate isozymes, are more active with FBP than with F1P. Aldolase B is encoded in Aldolase B gene (ALDOB) mapped to chromosome 9q21.3–q22.2 [\(129,](#page-25-0) [130](#page-25-0)). Missense and nonsense mutations, large and small gene deletions and mutations in the splicing region have been identified in ALDOB of HFI patients ([131](#page-25-0)).

Affected individuals manifest symptomatic hypoglycemia, vomiting and life-threatening episodes shortly after the intake of fructose or related sugars including sucrose and sorbitol [\(132](#page-25-0)). Prolonged ingestion leads to failure to thrive, hepatomegaly, jaundice, hepatic cirrhosis, and nephrocalcinosis, and may lead to convulsions, coma, and death from severe liver and kidney failure. Symptoms of HFI appear during infancy when infants with HFI are fed a formula or foods including fruits, vegetables, and sweetened cereals that contain sucrose. Patients with HFI may develop a protective aversion to sweets and fruits, which is a reason that diagnosis is frequently missed, and which also explains that reliable prevalence numbers for different populations do not exist.

HFI is associated with proximal tubule dysfunction leading to aminoaciduria, bicarbonaturia, phosphaturia, and lactic acidosis. These manifestations appear rapidly after the ingestion of fructose [\(133,](#page-25-0) [134\)](#page-25-0). The development of lactic acidosis adds significantly to the metabolic acidosis [\(135\)](#page-25-0). Chronic fructose ingestion leads to nephrocalcinosis and impaired distal tubular function. In contrast, resolution of proximal tubule dysfunction can take days or weeks with strict restriction of fructose and sucrose [\(136\)](#page-25-0).

Aldolase B coexists abundantly in endocytosis zones of the proximal tubule cells with H^+ -ATPase [\(137\)](#page-25-0). Nonfunctional aldolase B impairs the coupling of H⁺-ATPase to glycolysis and endosomal acidification that will lead to FS.

Diagnosis includes the metabolic response to an intravenous fructose load or an enzymatic assay of liver or intestinal biopsy samples. However, both of them are bothering and invasive ([138](#page-25-0)). Fructose breath hydrogen test is one of the standard procedures for the diagnosis. However, it can develop life-threatening adverse effects during the test [\(139\)](#page-25-0). Molecular analysis is available for the diagnosis.

Strict avoidance of foods or drugs containing fructose, sucrose, and sorbitol is the predominant treatment.

Glycogen Storage Disease Type I (von Gierke Disease)

Glycogen storage disease type I (GSD-I) is a group of autosomal recessive disorders with an incidence of 1 in 100,000. There are two major subtypes. Glycogen storage disease type Ia (GSD-Ia, von Gierke disease) is common and is caused by s deficiency in glucose-6-phosphatasealpha (G6Pase-alpha), a key enzyme in glucose homeostasis that catalyzes the hydrolysis of glucose-6-phosphate (G6P) to glucose and phosphate in the terminal step of gluconeogenesis and glycogenolysis [\(140](#page-25-0)). G6Pase-alpha is a hydrophobic endoplasmic reticulum-associated transmembrane protein. Glycogen storage disease type Ib (GSD-Ib) is caused by a deficiency of microsomal glucose-6-phosphatase transporter (G6PT). G6PT translocates G6P from cytoplasm to the lumen of the endoplasmic reticulum. Therefore, G6PT and G6Pase-alpha work in concert to maintain glucose homeostasis. Whereas G6Pase is exclusively expressed in gluconeogenic cells, G6PT is ubiquitously expressed and its deficiency generally causes a more severe phenotype.

Patients with GSD-Ia manifest a phenotype of disturbed glucose homeostasis characterized by fast lifethreatening hypoglycemia, hepatomegaly, nephromegaly, hypercholesterolemia, hypertriglyceridemia, hyperuricemia, lactic acidemia, neutrophilia, and growth retardation [\(141,](#page-25-0) [142](#page-25-0)). Infants with GSD-Ia typically present with seizures and hepatomegaly at 6–8 months of age. Approximately 75% of adolescent and adult patients develop hepatocellular adenoma (HCA), which can lead to considerable morbidity and mortality [\(143\)](#page-25-0). The incidence of HCA to hepatocellular carcinoma is recently increasing because the patients can live longer than before [\(144\)](#page-25-0). The presence of GSD-Ia and GSD-Ib are associated with reduced quality of life, independent functioning, and elevated levels of internalizing distress, and parental stress relative to healthy peers.

Renal complications include renal enlargement, gout nephropathy, renal stones, nephrocalcinosis, Fanconi-like syndrome, and chronic renal disease leading to renal insufficiency [\(145\)](#page-25-0). Hepatomegaly is a common finding in GSD-Ia. Hyperuricemia and uric acid stone in GSD-Ia are explained by a combination of increased synthesis of purine and a competitive inhibition of renal tubular excretion of uric acid (urate) by lactate ([146](#page-25-0)). Proximal tubular dysfunction has been observed in patients with GSD-Ia. Patients manifest proximal renal tubular acidosis due to loss of bicarbonate in the urine, hyperphosphaturia, generalized aminoaciduria and increased excretion of beta 2-microglobulin which are ameliorated by intensive diet therapy [\(147,](#page-25-0) [148\)](#page-25-0). This finding suggests that good metabolic control can prevent proximal tubular dysfunction. Chronic renal disease is a long-term complication. Renal biopsies reveal interstitial fibrosis, tubular atrophy, and focal segmental glomerulosclerosis with marked glomerular basement membrane (GBM) thickening and lamellation in patients with GSD-Ia [\(149–151\)](#page-25-0). Glycogen granules are present in the areas of abnormal GBM. The glycogen content in the mesangium and in the epithelial, mesangial and endothelial cells is increased. Recent treatment has significantly alleviated the metabolic abnormalities and delayed the clinical manifestation of chronic renal disease and renal insufficiency in patients with GSD-Ia. However, glomerular hyperfiltration, hypercalciuria, hypocitraturia that worsens with age, and urinary albumin excretion still occur in metabolically compensated patients with GSD-Ia ([152](#page-25-0), [153](#page-25-0)). Although the molecular mechanism responsible for chronic renal disease is still poorly understood, activation of the angiotensin system is suggested to have an important role for the disease progression [\(154\)](#page-25-0). The expression of TGF-beta 1 in kidney tissue is increased in a patient with GSD-Ia manifesting proteinuria, interstitial fibrosis, and tubular atrophy ([155](#page-25-0)).

The objective of treatment is to maintain normoglycemia to avoid metabolic complications and lactic acidosis. Normoglycemia is accomplished at night with nasogastric feeding of glucose or with orally administered

uncooked cornstarch ([156](#page-25-0)). A single dose (1.75–2.5 g kg⁻¹ of body weight) of uncooked cornstarch will maintain serum glucose concentration higher than 3.9 mmol L^{-1} for 7 h in most young adults ([157](#page-25-0), [158\)](#page-25-0).

Liver transplantation is indicated in the patients when medical treatment fails to control the metabolic problems or when HCA or hepatocellular carcinoma develops. Living-donor liver transplantation is a viable option to restore normal metabolic balance and keeping normal renal function [\(159\)](#page-25-0). Hepatocyte transplantation can be a potential therapeutic intervention to prevent hypoglycemia despite the discontinuation of cornstarch meals [\(160\)](#page-25-0).

Fanconi-Bickel Syndrome

Fanconi-Bickel syndrome (FBS) is an autosomal recessive disorder characterized by failure to thrive, "doll-like" face, hepatomegaly, nephromegaly, and severe rickets. Patients with FBS manifest glycogen accumulation in hepatocytes and proximal tubular cells, fasting hypoglycemia, galactose intolerance, and FS including glucosuria, aminoaciduria, hyperuricosuria, hyperphosphaturia, proteinuria, and sodium and potassium wastage ([161,](#page-25-0) [162](#page-25-0)). Some patients manifest cataracts in neonatal period [\(163\)](#page-26-0). Overall prognosis of FGS is considered as favorable [\(164\)](#page-26-0). However, some patients manifest neonatal diabetes mellitus and galactosemia and die of hepatic failure during infancy [\(165\)](#page-26-0).

FBS is caused by the mutations in facilitative glucose transporter gene (SLC2A2, also referred to as GLUT2) expressed in liver, kidney, intestine, and pancreatic islet cells ([166\)](#page-26-0). Over 60 mutations in SLC2A2 were reported [\(167\)](#page-26-0). This facilitative glucose transporter is expressed in hepatocytes, pancreatic beta cells, and renal and intestinal epithelial cells and is important for the exchange of glucose between these cell types and the bloodstream [\(168\)](#page-26-0). Renal histology reveals an increase in mesangial cellularity, glomerulosclerosia, and patchy swelling of epithelial foot process and irregularly thickened lamina rara interna in the glumeruli, and vacuolization of epithelial nuclei in the proximal tubule cells suggesting the presence of glycogen in a 7 year-old patient with FBS ([169](#page-26-0)).

The therapy for FBS is directed at the renal solute losses including sodium bicarbonate and potassiumsodium phosphate; treatment of rickets including active vitamin D3; and frequent feeding including night-time supplementation to prevent ketosis. Uncooked cornstarch has been shown to lessen hypoglycemia and improved growth [\(170\)](#page-26-0). Galactose–free milk is also used for infant patients ([165](#page-26-0), [171\)](#page-26-0).

Tyrosinemia I

Hereditary tyrosinemia type I (TI) is an autosomal recessive disorder of an amino acid metabolism. TI is due to the defect in the fumarylacetoacetate hydrolase (FAH) gene ([172](#page-26-0), [173\)](#page-26-0). FAH is the last enzyme in the tyrosine catabolic pathway.

Patients with TI display a variety of clinical symptoms, such as liver damage from infancy that advances to cirrhosis, reduced coagulation factors, hypoglycemia, high plasma concentrations of methionine, phenylalanine, and aminolevulinic acid, high risk of hepatocellular carcinoma, and tubular and glomerular renal dysfunction [\(174\)](#page-26-0).

Progressive renal damage begins from early infancy in severe form. Chronic liver damage with a high incidence of hepatoma (hepatocellular carcinoma) is characteristic in milder form ([175](#page-26-0)). Even a patient without clinical manifestations of TI can manifest hepatoma during childhood ([176](#page-26-0)). Accumulated fumarylacetoacetate in the patients with TI is pathogenic for hepatoma. Patients with milder form of TI are at risk for acute exacerbation of liver dysfunction. A common presentation mode is the "acute hepatic crisis" in which ascites, jaundice, and gastrointestinal bleeding are precipitated by an acute event such as an infection. Acute hepatic crises usually resolve spontaneously but on occasion progress to complete liver failure and encephalopathy. Acute, painful peripheral neuropathy may appear and can lead to transient paralysis. Autonomic dysfunction with hypertension and tachycardia can be associated with this acute neuropathy ([177](#page-26-0)). Plasma tyrosine and methionine levels usually are elevated in untreated patients. The presence of succinylacetone in plasma and urine is diagnostic of TI. A rapid ultra performance liquid chromatography tandem mass spectrometric method is used for mass screening of tyrosinemia ([178](#page-26-0)).

FS and developmental hypophosphatemic rickets are features of the kidney involvement. Generalized aminoaciduria, renal tubular acidosis, and mild proteinuria are also often seen, whereas glucosuria is less common because plasma glucose levels are low. Kidney enlargement is common, and nephrocalcinosis can be seen [\(179\)](#page-26-0). FS leads to carnitine deficiency ([180](#page-26-0)). Glomerulosclerosis and impaired GFR may be seen with time.

Disturbances in tyrosine metabolism lead to increased levels of succinylacetone and succinylacetoacetate. However, the mechanisms causing liver failure, cirrhosis, renal tubular dysfunction, and hepatocarcinoma are still unknown. Apoptosis of hepatocytes and renal tubular epithelial cells are characteristic features of this disease and the apoptotic signal in this disease seems to be initiated by fumarylacetoacetate [\(181](#page-26-0), [182](#page-26-0)). Accumulated maleylacetoacetate and fumarylacetoacetate in affected tissues can react with free sulfhydryl groups and reduce intracellular levels of glutathione. They may be capable of acting as alkylating agents. Maleylacetoacetate and fumarylacetoacetate are not detectable in plasma or urine but are converted to succinylacetoacetate. Succinylacetone, a metabolite of succinylacetoacetate, is structurally similar to maleic acid, which is known to induce FS and may be the cause of tubular dysfunction of HI. Experimentally, succinylacetone administration to rats leads to FS [\(183,](#page-26-0) [184](#page-26-0)).

Treatment with a low-phenylalanine and low-tyrosine diet dramatically improves the renal tubular dysfunction [\(185\)](#page-26-0). However, this treatment cannot necessarily improve the hepatic involvement. Moreover, there is a risk of inducing deficiencies of phenylalanine or tyrosine. The formation of pathogenic fumarylacetoacetae is prevented by 2- (2-nitro-4-trifluoromethylbenzoyl)-1,3-cyclohexanedione (NTBC). NTBC is used for the patients with TI during the first 6 months of life in addition to a diet low in tyrosine and phenylalanine. NTPC clearly improves the vital prognosis and quality of life in the patients ([186](#page-26-0)). However, some patients with NTBC treatment develop hepatoma. A rise of alpha-fetoprotein (AFP), a slow AFP decrease, and never normalizing levels of AFP are important predictors of hepatoma development ([187](#page-26-0)). Liver transplantation has been used for patients with liver failure and to prevent the development of hepatoma [\(174\)](#page-26-0). Liver transplantation leads to rapid correction of FS [\(188\)](#page-26-0).

Wilson Disease

Wilson disease (WD, progressive hepatolenticular degeneration) is an autosomal recessive inborn error of copper (Cu) metabolism that affects numerous organ systems [\(189\)](#page-26-0). Biliary excretion of Cu and incorporation into ceruloplasmin is impaired, leading to liver damage, neuronal degeneration, and impairment of other organs from accumulation of Cu in patients with WD.

The majority of patients with WD presents with either predominantly hepatic or neuropsychiatric symptoms, and with either clinically asymptomatic or symptomatic liver involvement. Approximately 40% of patients presents with liver disease, 40% with extrapyramidal symptoms, and 20% with psychiatric or behavioral abnormalities. Symptoms rarely occur before 6 years of age. Hepatic involvement includes acute hepatitis, fulminant hepatic failure, or progressive chronic liver disease in the form of either chronic active hepatitis or cirrhosis of the macronodular type. Neuropsychiatric involvements are variable. The most common initial presentation is bulbar symptoms characterized by difficulties with speech and swallowing, and drooling. They frequently manifest dysarthria and coordination defects of voluntary movements accompanied by involuntary movements. One third of the patients with WD manifest psychiatric disturbances. The remaining patients with WD may present with symptoms including hemolytic anemia, bone fracture, arrhythmias, FS, hyperpigmentation, Kayser-Fleisher ring, cataract, and gynecological problems that are attributable to the involvement of the organs.

WD is caused by a mutation in the gene ATP7B that encodes a P-type Cu transporting ATPase beta polypeptide enzyme (ATP7B) [\(190](#page-26-0)). This ATPase is targeted to the mitochondria, suggesting that its role in Cu dependent processes takes place in this organelle. The disease frequency is estimated to be between 1 in 5,000 and 1 in 30,000, and the carrier frequency is approximately 1 in 90 ([191](#page-26-0)).

Cu is absorbed by the intestinal cells and stored with metallothionein in a non-toxic form. The Cu is later delivered into the circulation by a Cu transporter 15,000 amino acid protein, Cu-transporting P-type ATPase 1 (ATP7A), which is located on the membrane of enterocytes (\bullet [Fig. 42-7](#page-18-0)) ([192\)](#page-26-0). It is then transported to the liver tagged with albumin, from where it is accepted by hepatocytes. The ATOX1 chaperone protein directs Cu to its binding targets in the hepatocytes. Some of Cu bounds to metallothionein for storage, and the remainders are excreted into ATP7B-regulated biliary canaliculi. ATP7B also mediates the transfer of Cu to apoceruloplasmin to form a six-Cu binding protein, ceruloplasmin ([193](#page-26-0)). Ceruloplasmin is released into the blood, carries 90% of the Cu present in the plasma, and acts as a source for peripheral organs. Mutations of the ATP7A gene result in the storage of Cu in enterocytes, preventing entry of Cu into the circulation and thus causing a complete Cu deficiency (Menkes kinky hair disease) ([192](#page-26-0)).

Mutations in ATP7B gene lead to a reduction in the conversion of apoceruloplasmin into ceruloplasmin, which is usually present at low levels in the patients of WD. A failure to excrete Cu into the biliary canaliculi leads to toxic effect to hepatocytes. Excess Cu damages mitochondria, which produce oxidative damage to the cells and allows spillage of Cu into the blood, thereby overloading other tissues including the brain, kidney, and red blood cells, initiating toxic effects.

Excessive accumulation of Cu in the kidney leads to renal tubular dysfunction in patients with WD. Patients show most features of FS before the onset of hepatic

failure and is characterized by intermittent glucosuria, aminoaciduria, hyperphosphaturia, hyperuricosuria, and proteinuria [\(194–196](#page-26-0)). Patients can manifest rickets or osteomalacia. hypercalciuria, urolithiasis, nephrocalcinosis, decreased urine concentrating ability and distal renal tubular acidosis are reported [\(197\)](#page-26-0). Glomerular function decreases as the disease progresses, but death from extrarenal causes occurs before the onset of renal failure.

Histological analysis of the patients with WD revealed no alteration on light microscopy or only flattened proximal tubule cells without recognizable brush border [\(198\)](#page-26-0). Electron microscopy disclosed loss of the brush borders, electron dense deposit in the subapical region of tubular cytoplasm probably representing metalloproteins, and cavitation of the mitochondria with disruption of the normal cristae pattern ([199](#page-26-0)).

All subjects presenting with symptomatic or asymptomatic liver disease with no apparent causes or with extrapyramidal features along with a past or family history of similar hepatic or neurological illnesses in other siblings should be screened for WD [\(200](#page-26-0)). Measurement of the serum ceruloplasmin is valuable for diagnosis. Any value below 200 mg L^{-1} is abnormal, and reduced levels are seen in up to 95% of the patients with WD. An estimation of 24 h urinary Cu excretion is another reliable test for the diagnosis of WD. Normal excretion is between 20 and 50 mg a day, and Cu excretion is increased to in excess of 100 µg a day in patients with WD. Serum free Cu is a measure of nonceruloplasmin toxic Cu in the blood, and normal value range from 1.3 to 1.9 μ mol L⁻¹ (8–12 μ g dL^{-1}) in parallel with the increased urinary Cu excretion, because of saturation of the hepatic storage of Cu. A hepatic biopsy and a measurement of its Cu content are helpful for diagnosis. 80% of the patients manifest increased hepatic Cu $(>250 \text{ µg/g of dry tissue weight}).$ Genetic analysis of ATP7B gene is a helpful confirmation of WD. Brain MRI is a very sensitive method for revealing abnormalities in patients with WD. Generalized brain atrophy and hyper-intensity in the basal ganglia, white matter, thalamus, or brainstem are common findings in the patients. Patients exhibit characteristic features on MRI; ''face of the giant panda'' is seen in the midbrain and "face of the miniature panda" is seen in the tegmentum region of the pons in T2-weighted images [\(201,](#page-26-0) [202\)](#page-26-0).

WD has a fatal outcome if not treated appropriately and in a timely manner. The aim of treatment for WD is to remove the toxic deposit of Cu from the body to produce a negative Cu balance, and to prevent its reaccumulation [\(190,](#page-26-0) [203](#page-26-0)). Successful therapy is measured in terms of a restoration of normal levels of free serum Cu and its excretion in the urine. The average daily diet

Schematic representation of cooper metabolism within a liver cell. ATP7b, causative gene for Wilson disease. (Das SK, Ray K. Wilson's disease: an update. Nat Clin Pract Neurol 2006;2:482–493).

contains 2–4 mg of Cu, and 0.8 mg is normally lost into the feces. Patients should avoid Cu-rich foods including chocolate, nuts, shellfish, mushrooms, and liver. D-penicillamine therapy (20 mg kg^{-1} of body weight a day) has been the most commonly used chelating agent that may reverses multiple tissue dysfunction including FS. However, use of D-penicillamine has been questioned, because of reported side effects. The side effects from D-penicillamine can occur both early and late in the treatment period. Early side effects include a hypersensitivity reaction characterized by fever, skin rash, and lymphadenopathy. Delayed side effects including Goodpasture's syndrome, polymyositis, systemic lupus erythematosus, and bone marrow suppression are caused by immunological reactions. Trientine is another effective chelating agent (750–2,000 mg a day for adults). Ammonium tetrathiomolybdate $(2-3 \text{ mg kg}^{-1})$ of body weight a day in six doses along with meals and in the interval between meals) is a potent agent in removing Cu from the body and may be the drug of choice for patients with neurologic disease to prevent the immediate worsening of symptoms that can occur with D-penicillamine therapy. Zinc acetate or sulfate induces intestinal metallothionein and helps in the prevention of Cu absorption from the gut $(3-5 \text{ mg kg}^{-1} \text{ of body weight a day in three divided doses})$ before meals). D-penicillamine should be taken 2 h after meals to avoid any interaction with the zinc. Zinc acetate or zinc sulfate has been used successfully in asymptomatic or presymptomatic affected family with WD, and is equally as effective as D-penicillamine in a group of patients predominantly with neurological disease [\(204,](#page-26-0) [205\)](#page-26-0). The use of trientine, tetrathiomolybdate, and zinc has been advocated, although results of long-term trials are awaited. The best therapeutic approach remains controversial and there is no universally accepted regimen. Liver transplantation is effective for the patients with progressive live failure or acute liver failure. Liver transplantation is also indicated for patients with WD in whom medical therapy is ineffective. Symptomatic patients with WD require lifelong treatment, because an interruption to therapy or inadequate treatment can lead to fatalities within 9 months to 3 years [\(189,](#page-26-0) [203](#page-26-0)).

Idiopathic Fanconi Syndrome

Although the significant numbers of genetic causes for the disorders leading to FS are identified, there exist patients with idiopathic FS. Idiopathic FS should be diagnosed when all other known causes have been excludes. Idiopathic FS can be inherited as an autosomal dominant ([206](#page-26-0)–[211\)](#page-27-0), autosomal recessive [\(212,](#page-27-0) [213\)](#page-27-0), and X-linked form [\(214](#page-27-0)). However, most of the familial forms are autosomal dominant inheritance.

Children with idiopathic FS manifest failure to thrive, frequent bouts of dehydration, and rickets. They often have other features of FS, including polyuria, polydipsia, hypokalemia, hypophosphatemia, proximal renal tubular acidosis, aminoaciduria, glucosuria, and proteinuria. Glomerular filtration rate is usually normal during childhood. However, some develop chronic renal insufficiency or chronic renal failure 10–30 years after the onset of symptoms ([208](#page-27-0), [209,](#page-27-0) [213\)](#page-27-0). Nephrocalcinosis and genu vulgum (knock knee) are seen in some patients ([213](#page-27-0)). Bone pain, bone fracture, and scoliosis due to osteomalacia can become serious complications in adult patients with idiopathic FS $(\bullet$ Fig. 42-8).

Renal histology reveals chronic tubulointerstitial fibrosis. The interstitium demonstrates patchy fibrosis associated wit tubular atrophy and focal collections of

D Figure 42-8

Plain X-ray showing scoliosis and osteopenia in a 35-yearold female with idiopathic Fanconi syndrome.

mononuclear inflammatory cells. Occasional cystically dilated tubules are seen containing eosinophilic proteinaceous material that stains positive for PAS ([213](#page-27-0)).

Treatment for idiopathic FS remains symptomatic. Careful follow-up of these patients is necessary to prevent recurrent bouts of dehydration, electrolyte imbalance, and metabolic bone diseases. Glomerular function and nephrocalcinosis must be checked regularly. Renal transplantation has been done in a few patients who had endstage renal failure ([209](#page-27-0)).

Acquired Fanconi Syndrome

Nephrotic syndrome is associated with FS [\(215\)](#page-27-0). The renal pathology is focal segmental glomerulosclerosis. Although the true pathogenesis is not clarified, mitochondriopathies can manifest FS, focal segmental glomerulosclerosis leading to nephrotic syndrome, and both [\(79](#page-24-0)–[84](#page-24-0)).

Immunological or hematological disorders are associated with dysproteinuria leading to FS. They are multiple myeloma, Sjögren syndrome, and amyloidosis. Almost all of the patients with theses diseases are adults. In early stages of myeloma, light chain nephrotoxicity often presents with proximal tubular functional abnormalities leading to FS. Proximal tubule dysfunction is the most common mode of renal involvement and it can manifest in a variety of ways. Endocytosis in the proximal tubules is overloaded and cell stress responses that include phosphorylation of MAPKs, prominently, p38 MAPK, and nuclear transcription factors NF-kappa B, AP-1 are activated resulting in production of inflammatory and proinflammatory cytokines, TNF-alpha, interleukin-6, 8 and monocyte chemo-attractant protein-1 [\(216\)](#page-27-0). These proximal tubule alterations often progress to a severe tubulointerstitial nephritis and end stage renal failure.

Sjögren syndrome is an autoimmune connective tissue disorder that affects exocrine glands. Renal involvement of Sjögren syndrome is mainly manifested as tubular disorders; 70% of the patients manifest distal renal tubular acidosis [\(217\)](#page-27-0). Urinary concentration defect, proteinuria and LMW proteinuria are often seen in the patients. Only 4% of the patients manifest FS [\(217](#page-27-0)). Patients with FS and Sjögren syndrome manifest osteomalacia including bony deformities of rib cage, bilateral humeral shaft fractures, and marked cortical bone thinning [\(218](#page-27-0)). Characteristic histological feature of Sjögren syndrome is chronic interstitial nephritis, with diffuse or focal plasmacytoid lymphocytic infiltration. In the late stage of the disease, tubulointerstitial fibrosis is severe. Corticosteroid or/and immunosuppressant therapy can improve the prognosis.

FS has appeared rarely after renal transplantation [\(219](#page-27-0)). Acute tubular necrosis, chronic rejection reaction, and nephrotoxic drugs can induce the progression of FS in the patients.

Acute tubulointerstitial nephritis with uveitis (TINU) syndrome is an immunological disease that leads to tubulointerstitital nephritis and anterior uveitis [\(220\)](#page-27-0). Patients with TINU syndrome manifest asthenia, malaise, weight loss, nocturia, and thirst. Patients also manifest incomplete or complete FS including proteinuria, LMW proteinuria, glucosuria, aminoaciduria, bicarbonaturia, phosphaturia, and uricosuria due to proximal tubule dys-function and acute renal failure [\(221\)](#page-27-0). Urine concentration is decreased in the patients. Corticosteroid therapy can improve renal and eye manifestations.

Autoimmune interstitial nephritis and membranous nephropathy is a distinct disorder. The patients manifest failure to thrive, multiple renal tubular disorders including FS and proteinuria [\(222](#page-27-0), [223\)](#page-27-0). Renal biopsy revealed interstitial nephritis with lymphocytic infiltration and fibrosis, and membranous nephropathy. In advanced stage, focal segmental glomerulosclerosis and tubular atrophy develop. Immunofluorescence analysis shows linear staining of IgG along the glomerular capillaries and the tubular basement membrane. These renal lesions result from an autoimmune response to the 58-kD tubular basement membrane autoantibody [\(224\)](#page-27-0). This disorder is genetically related to HLA B7 serotype [\(224\)](#page-27-0).

A patient with anorexia nervosa is described to manifest reversible FS like condition including glucosuria, phophaturia, and uricosuria, although the precise pathogenesis is not known ([225](#page-27-0)). These manifestations subside with nutritional recovery.

Untreated patients with distal renal tubular acidosis manifest LMW proteinuria, generalized aminoaciduria, phosphaturia, uricosuria, and hypercalciuria ([226](#page-27-0), [227\)](#page-27-0). These proximal tubular abnormalities are transient and disappear by the alkali and potassium therapy. Although the precise pathogenic mechanisms underling the development of proximal tubular dysfunction remains unclear, decreased pH in the cytoplasm of the proximal tubule cells resulting from the intracytoplasmic accumulation of H⁺ due to luminal membrane H⁺-ATPase dysfunction can disturb trafficking of endosome.

Exogenous Factors

Drugs

Numerous drugs and herbs are implicated in the pathogenesis of FS. Drugs and herbs are usually filtered from the glomerulus and reabsorbed in the proximal tubules. They include outdated tetracycline [\(228\)](#page-27-0), aminoglycosides [\(229,](#page-27-0) [230](#page-27-0)), salicylate [\(231](#page-27-0)), valproic acid [\(232,](#page-27-0) [233](#page-27-0)), and Chinese herbs ([234,](#page-27-0) [235\)](#page-27-0). Aminoglycoside antibiotics reduce glucose reabsorption in kidney tissue by reducing mRNA, protein expression, and function of the Na+ -dependent glucose transporter, which is located in the luminal membrane of the proximal tubule ([236](#page-27-0)). Covalent binding of salicylate or its metabolites to mitochondria in proximal tubule cells alters the function of mitochondria ([231](#page-27-0)). Valproic acid produces the defects of mitochondrial respiratory chain and lysosomal enzyme activity in the proximal tubule cells leading to multiple renal transport abnormalities ([13,](#page-22-0) [237](#page-27-0)). Chinese herbs containing aristolochic acids cause proximal tubular injury, and this is called as aristolochic acid-related nephropathy.

A number of cancer chemotherapy agents are associated with renal glomerular and tubular dysfunctions including FS. The nephrotoxicity of cancer chemotherapy agents is dose dependent and often irreversible. Ifosfamide is an alkylating agent widely used in the treatment of various solid tumors. Chloroacetaldehyde (CAA), one of the main metabolites of ifosfamide, contributes to inhibit endocytosis in the proximal tubule cells ([238](#page-27-0)). CAA decreases total glutathione and ATP levels in the proximal tubule cells. CAA also inhibits endosomal H⁺-ATPase activity, which disturbs intracellular vesicle trafficking [\(239\)](#page-27-0). Patients receiving ifosfamide who have received prior cisplatin are at significantly higher risk of developing FS than are those who have received no prior nephrotoxic therapy ([240](#page-27-0)). When the patients manifest FS, renal sonography reveals hyperechogenicity of the parenchyma with good corticomedullar differentiation ([241](#page-27-0)). Taurine can protect against ifosfamide-induced renal dysfunction without compromising its anti-tumor activity ([242](#page-27-0)). Cisplatin also reduces glucose reabsorption in kidney tissue by reducing mRNA, protein expression, and function of the Na⁺-dependent glucose transporter [\(243\)](#page-27-0). Cisplatin inhibits various types of amino acid transporters in the proximal tubule cells leading to a generalized aminoaciduria ([243](#page-27-0)).

Imanitib mesylate is a specific tyrosine kinase inhibitor that is the first line therapy for patients with chronic myeloid leukemia. This agent induces partial FS including phosphaturia and uricosuria with mild renal failure. Combined blockade of both platelet-derived growth factor receptor and c-Kit receptor tyrosine kinase in proximal tubules causes partial FS ([244](#page-27-0)).

Nucleotide reverse transcriptase inhibitors that are used as anti-human immunodeficiency virus (HIV) agents including adefovir, cidofovir, and tenofovir induce

FS, nephrogenic diabetes insipidus, and acute renal failure ([245](#page-27-0)–[248\)](#page-27-0). Adefovir and cidofovir interact with organic anion transporters (OAT); these drugs enter into proximal tubule cells by activated OAT located in the basolateral membrane. However, their efflux into the tubular lumen is decreased by inactivated multidrug-resistanceprotein 2 (MRP 2) located in the luminal membrane. Thus, these drugs are accumulated in the proximal tubule cells leading to mitochondrial damage and tubular toxicity. Cytotoxicity of adefovir and cidofovir is proportional to cellular OAT expression ([246](#page-27-0)). Histologic and ultrastructural examination reveals tubular degenerative changes of proximal tubules with swollen and dysmorphic mitochondria. In tubular cells, respiratory chain components encoded by mitochondrial DNA (cytochrome oxidase subunit I) are selectively deficient in renal tubular cells, and mitochondrial DNA is quantitatively reduced ([249](#page-27-0)). In contrast to adefovir and cidofovir, renal toxicity of tenofovir is much less frequent. Tenofovir has little mitochondrial toxicity and it does not interact with OAT ([250](#page-27-0)). Therefore, the precise mechanisms of nephrotoxicity by tenofovir remain unknown.

Chemical Compounds

Paraquat, a non-selective herbicide, and colloidal bismus subcitrate cause FS ([251](#page-27-0), [252](#page-28-0)). Large amount of these compounds are usually ingested in a suicide attempt. Patients manifest FS and acute renal failure. Treatment with the chelating agent sodium-2,3-dimercapto-1-propanesulfonate in combination with hemodialysis is highly effective in reducing the serum bismus level. Methyl-3-chromone (diachrome) ([253](#page-28-0)), 6-mercaptopurine ([254](#page-28-0)), and toluene also lead to FS [\(255\)](#page-28-0).

Heavy Metals

Heavy metals such as lead, cadmium, mercury, chromium, and platinum are a major environmental and occupational hazard. They are very toxic at very low doses. The kidney is the first target organ of heavy metal toxicity. The extent of renal damage by heavy metals depends on the nature, dose, route, and duration of exposure. Both acute and chronic intoxication have been demonstrated to cause nephropathies, with various levels of severity ranging from tubular dysfunctions like acquired FS to severe renal failure [\(256\)](#page-28-0). Lead poisoning leads to FS, predominantly in children [\(257\)](#page-28-0). As lead is non-biodegradable with a very long biological half-life, aminoaciduria and

glycosuria persist up to 13 years after childhood severe lead poisoning ([258](#page-28-0)).

Cadmium intoxication leads to FS after a long exposure [\(259\)](#page-28-0). The industrial waste contaminating cadmium in the Jinzu River basin in Toyama prefecture in Japan produced a lot of patients with Itai-Itai (ouch-ouch) disease that is compatible to FS with severe osteomalacia. Patients complained severe bone pains that are derived from advanced non-traumatic multiple bone fractures. Cadmium produces free radicals that alter mitochondrial activity or induce mitochondrial gene deletion in the proximal tubules ([260](#page-28-0), [261\)](#page-28-0). Cadmium inhibits H^+ -ATPase, which results in a Fanconi-like syndrome ([6](#page-22-0)).

Therapy

Identification of the underling cause for FS is a first step and is critical to direct specific therapy. Avoidance of offending nutrients in galactosemia, HFI, and tyrosinemia and avoidance of Cu-rich foods in WD are therapeutically critical. Specific treatments with Cu-chelating agents including D-penicillamine, trientine, and ammonium tetrathiomolybdate, and zinc are effective for WD. Imuunosuppressive drugs are used for immunologically induced disorders including Sjögren syndrome, TINU syndrome and autoimmune interstitial nephritis and membranous nephropathy. These treatments can completely resolve FS.

When specific therapy does not exist, therapy is directed at the biochemical abnormalities secondary to renal solute and fluid losses and the metabolic bone diseases. Proximal renal tubular acidosis usually requires large amount of alkali $(2-15 \text{ mEq} \text{ kg}^{-1} \text{ of body weight})$ a day) divided into four to six daily doses. High dose of alkali can produce volume expansion, further bicarbonate wasting and potassium loss in the patients with FS. 1–3 mg kg^{-1} of body weight a day of hydrochlorothiazide can reduce the dose of alkali by preventing the volume expansion. Administration of potassium salt of citrate, bicarbonate, or acetate fulfills the dual purpose of treating acidosis and preventing hypokalemia. Sodium wasting and dehydration are treated with combination of sodium bicarbonate, citrate, and chloride, depending on the degree of acidosis. Ensuring adequate fluid and electrolyte intake is essential, especially in the case of infants or gastrointestinal diseases. Early intervention with intravenous replacement therapy is required for the patients with FS who manifest vomiting and diarrhea.

Hypophosphatemia and impaired renal vitamin D3 metabolism in patients with FS lead to rickets and other metabolic bone diseases. 1–3 g of phosphate supplementation is necessary as neutral phosphate (the mixture of sodium phosphate dibasic 1.94 g and potassium phosphate monobasic 0.34 g contains 0.5 g of phosphate) divided into four to six daily doses. Supplementation of 1,25 dihydroxyvitamin D3 or dihydrotachysterol is effective to treat or prevent rickets and osteomalacia. Vitamin D3 therapy improves the hypophosphatemia and lessens the risk of hyperparathyroidism. Hypercalcemia and hypercalciuria are toxic side effects of vitamin D3 therapy. An adequate amount of physical activity, as well as appropriate diet with calcium, phosphate, and vitamin D3, is necessary to prevent bone deformations, non-traumatic fractures leading to bone pain, deterioration of motor development and disability [\(262\)](#page-28-0).

Aminoaciduria, glucosuria, proteinuria, LMW proteinuria, and uricosuria usually do not induce clinical symptoms and do not require specific treatments.

Growth failure is a major complication in FS. Despite correction of electrolyte abnormalities, some patients manifest severe growth retardation, especially those with cystinosis and Fanconi-Bickel syndrome. A patient with FS was reported to have growth hormone deficiency ([263](#page-28-0)). Supplemental growth hormone has been used successfully in a few patients with FS.

References

- 1. De Toni G. Remarks on the relations between renal and rickets (renal dwarfism) and renal diabetes. Acta Paediatr 1933;16:479–484.
- 2. Debré R, Marie J, Cléret F et al. Rachitisme tradif coexistànt avec une nephrite chronique et une glycosurie. Arch Med Enf 1934;37: 597–606.
- 3. Fanconi G. Die nichit diabeteishen glykosurien und hyperglykamien des altern kinds. Jahrb Kinderheilkd 193;133:257–300.
- 4. Marshansky V, Bourgoin S, Londino I et al. Receptor-mediated endocytosis in kidney proximal tubules; recent advances and hypothesis. Electrophoresis 1997;18:2661–2676.
- 5. Brown D, Stow JL. Protein trafficking and polarity in kidney epithelium; from cell biology to physiology. Physiol Rev 1996; 76:245–297.
- 6. Herak-Kramberger CM, Stow JL. Protein trafficking and polarity in kidney vacuolar H⁺-ATPase and endocytosis in rat cortex. Kidney Int 1998;53:1713–1726.
- 7. Marshansky V, Richard M, Bartle J et al. Regulation of renal albumin reabsorption by endosomal proton transport [Abstract]. J Am Soc Nephrol 1996;7:1311.
- 8. Lloyd SE, Pearce SH, Fisher SE et al. A common molecular basis for three inherited kidney stone diseases. Nature 1996;379:3445–3449.
- 9. Norden AGW, Lapsley M, Igarashi Tet al. Urinary megalin deficiency implicates abnormal tubular endocytotic function in Fanconi syndrome. J Am Soc Nephrol 2002;13:123–133.
- 10. Sakarcan A. The Fanconi syndrome of cystinosis: insights into the pathophysiology. Tur J Pediatr 2002;44:279–282.
- 11. Rech VC, Athaydes GA, Feksa LR et al. Inhibition of creatine kinase activity by cysine in the kidney of young rats. Pediatr Res 2006;60:190–195.
- 12. Niaudet P, Rötig A. The kidney in mitochondrial cytopathies. Kidney Int 1997;51:1000–1007.
- 13. Hawkins E, Brewer E. Renal toxicity induced by valproic acid (Depaken). Pediatr Pathol 1993;13:863–868.
- 14. Magen D, Sprecher E, Zelikovic I et al. A novel missense mutation in SLC5A2 encoding SGLT2 underlies autosomal-recessive renal glucosuria and aminoaciduria. Kideny Int 2005;67:34–41.
- 15. Bingham C, Ellard S, Cheret C et al. The generalized amonoaciduria seen in patients with hepatocyte nuclear factor-1 alpha mutation is a feature of all patients with diabetes and is associated with glucosuria. Diabetes 2001;50:2047–2052.
- 16. Tokaymat A, Sakarcan A, Neiberger R. Idiopathic Fanconi syndrome in a family. I. Clinical aspects. J Am Soc Nephrol 1992;2: 1310–1317.
- 17. Haffner D, Weinfurth A, Seidel C et al. Body growth in primary de Toni-Debré-Fanconi syndrome. Pediatr Nephrol 1997;11:40-45.
- 18. Flyvbjerg A, Dørup I, Everts ME et al. Evidence that potassium deficiency induces growth retardation through reduced circulating levels of growth hormone and insulin-like growth factor I. Metabolism 1991;40:769–775.
- 19. Tsao T, Fawcett J, Fervenzas FC et al. Expression of insulin-like growth factor-I and transforming growth factor-beta in hypokalemic nephropathy in the rat. Kidney Int 2001;59:96–105.
- 20. Brünger M, Hutler HN, Krapf R. Effect of chronic metabolic acidosis on the growth hormone/IGF-I endocrine axis: new cause of growth hormone insensitivity in humans. Kidney Int 1997;51:216–221.
- 21. Hsu SY, Tsai IJ, Tsau YK. Comparison of growth in primary Fanconi syndrome and proximal renal tubular acidosis. Pediatr Nephrol 2005;20:460–464.
- 22. Tsilchorozidou T, Yovos JG. Hypophosphataemic osteomalacia due to de Toni-Debré-Fanconi syndrome in a 42-year old girl. Hormones (Athens) 2005;4:171–176.
- 23. Urabe Y, Tagami T, Suwabe T et al. A patient with symptomatic osteomalacia associated with Fanconi syndrome. Mod Rheumatol 2005;15:207–212.
- 24. Morisaki I, Abe K, Sobue S. Orofacial manifestations in a child with Fanconi's syndrome. Oral Surg Oral Med Oral Pathol 1989;68: 171–174.
- 25. Armando N. Proximal tubule endocytic apparatus as the specific renal uptake mechanism for vitamin D binding protein/25-(OH) D3 complex. Nephrology 2006;11:510–515.
- 26. Gahl WA. Cysitinosis coming of age. Adv Pediatr 1986;33:95–126.
- 27. Deshpande P, Ali U. Primary Fanconi syndrome. Ind Pediatr 1997;34:547–549.
- 28. Brewer ED, Tsai HC, Norris RC. Evidence for impairment of metabolism of 25-hydroxyvitamin D3, in children with Fanconi syndrome. Clin Res 1976;24:154A.
- 29. Scheinman SJ. X-linked hypercalciuric nephrolithiasis: clinical syndromes and chloride channel mutation. Kidney Int 1998;53:2–17.
- 30. Kaunisto K, Parkkila S, Rajaniemi H et al. Carbonic anhydrase XIV: Luminal expression suggests key role in renal acidification. Kidney Int 2002;61:2111–2118.
- 31. Levinson DJ, Sorensen LB. Renal handling of uric acid in normal and gouty subject: Evidence for a 4-component system. Ann Rheum Dis 1980;39:173–179.
- 32. Meisel AD, Diamond HS. Hyperuricosuria in the Fanconi syndrome. Am J Med Sci 1977;273:109–115.
- 33. Roch-Ramel F, Guisan B, Diezi J. Effects of uricosuric and antiuricosuric agents on urate transport in human brush-border membrane vesicles. J Pharmacol Exp Ther 1997;280:839–845.
- 34. Enomoto A, Kimura H, Chairoungdua A et al. Molecular identification of a renal urate anion exchanger that regulates blood urate levels. Nature 2002;417:447–452.
- 35. Ohta T, Sakano T, Igarashi Tet al. Exercise-induced acute renal failure associated with renal hypouricemia: Results of a questionnaire-based survey in Japan. Nephrol Dial Transplant 2004;19:1447–1453.
- 36. Viart V, Rudan I, Hayward C et al. SLC2A9 is a newly identified urate transporter influencing serum urate concentration, urate excretion and gout. Nature Genet 2008;40:437–442.
- 37. Becker BF. Towards the physiological function of uric acid. Free Rad Biol Med 1993;14:615–631.
- 38. Norden AGW, Sharratt P, Cutillas PR et al. Quantitative amino acid and proteomic analysis: Very low excretion of polypeptides >750 Da in normal urine. Kidney Int 2004;66:1994–2003.
- 39. Maack T. Renal handling of proteins and polypeptides. In Handbook of Physiology. Renal Physiology, Windhager EE (ed.). New York, Oxford University Press, 1992, pp. 2039–2082.
- 40. Birn H, Fyfe JC, Jacobsen C et al. Cubilin is an albumin binding protein important for renal tubular albumin reabsorption. J Clin Invest 2000;105:1353–1361.
- 41. Birn H, Christensen EI. Renal albumin absorption in physiology and pathology. Kidney Int 2006;69:440–449.
- 42. Dent CE, Friedman M. Hypercalciuric rickets associated with renal tubular change. Arch Dis Child 1964;39:240–249.
- 43. Wrong OM, Norden AG, Freest TG et al. Dent's disease; a familial renal tubular syndrome with low-molecular weight proteinuria, hypercalciuria, nephroclcinosis, metabolic bone disease, progressive renal failure and a marked male predominance. QJM 1994;87:473–493.
- 44. Hodgin JB, Corey HE, Kaplan BS et al. Dent disease presenting as partial Fanconi syndrome and hypercalciuria. Kidney Int 2008; 73:1320–1323.
- 45. Suzuki Y, Okada T, Higuchi A et al. The low molecular weight of protein components in children urine. Acta Peadiatr Jpn 1980;22:1–5.
- 46. Igarashi T, Hayakawa H, Shiraga H et al. Hypercalciuria and nephrocalcinosis in patients with idiopathic low-molecular-weight proteinuria in Japan: Is the disease identical to Dent's disease in United Kingdom? Nephron 1995;69:242–247.
- 47. Lloyd SE, Pearce SHS, Gunter H et al. Idiopathic low molecular weight proteinuria associated with hypercalciuria, nephrocalcinosis in Japanese children is due to mutations of the renal chloride channel (CLCN5). J Clin Invest 1997;99:967–974.
- 48. Akuta N, Lloyd SE, Igarashi T et al. Mutations of CLCN5 in Japanese children with idiopathic low molecular weight proteinuria, hypercalciuria and nephrocalcinosis. Kidney Int 1997;52:911–916.
- 49. Igarashi T, Inatomi J, Ohara T et al. Clinical and genetic studies of CLCN5 mutations in Japanese families with Dent's disease. Kidney Int 2000;58:520–527.
- 50. Jentsch TJ, Poet M, Furhmann JK et al. Physiological functions of ClC Cl channels gleaned from human genetic disease and mouse models. Annu Rev Physiol 2005;67:779–807.
- 51. Moulin P, Igarashi T, van der Smissen P et al. Altered polarity and expression of H⁺-ATPase without ultrastructural changes in kidneys of Dent's disease patients. Kidney Int 2003;63:1285–1295.
- 52. Frymoyer SC, Scheinman SJ, Dunham PB et al. X-linked recessive nephrolithiasis with renal failure. N Engl J Med 1991;325:681–686.
- 53. Norden AGW, Scheinman SJ, Deschodt-Lanckman MM et al. Tubular proteinuria defined by a study of Dent's (CLCN5 mutation) and other tubular diseases. Kidney Int 2000;57:240–249.
- 54. Scheinman SJ. X-linked hypercalciuric nephrolithiasis: Clinical syndromes and chloride channel mutations. Kidney Int 1998;53:3–17.
- 55. Ludwig M, Utsch B, Balluch B et al. Hypercalciuria in patients with CLCN5 mutations. Pediatr Nephrol 2006;21:1241–1250.
- 56. Gailly P, Jouret F, Martin D et al. A novel renal carbonic anhydrase type III plays a role in proximal tubule dysfunction. Kidney Int 2008; 74:52–61.
- 57. Carr G, Simmons NL, Sayer JA et al. Disruption of clc-5 leads to redistribution of annexin A2 and promotes calcium crystal agglomeration in collecting duct epithelial cells. Cell Mol Life Sci 2006;63:367–377.
- 58. Norden AGW, Lapsley M, Lee PJ et al. Glomerular protein sieving and implications for renal failure in Fanconi syndrome. Kidney Int 2001;60:1885–1892.
- 59. Hoopes RR Jr., Raja KM, Koich A et al. Evidence for genetic heterogeneity in Dent's disease. Kidney Int 2004;65:1615–1620.
- 60. Raja KA, Schurman S, D'Mello et al. Responsiveness of hypercalciuria to thiazide in Dent's disease. J Am Soc Nephrol 2002; 13:2938–2944.
- 61. Cebotaru V, Kaul S, Devuyst O et al. High citrate diet delays progression of renal insufficiency in the ClC-5 knockout mouse model of Dent's disease. Kidney Int 2005;68:642–652.
- 62. Guggino SE. Mechanism of disease: What can mouse models tell us about the molecular process underlying Dent disease? Nat Clin Pract Nephrol 2007;3:449–455.
- 63. Copelvitch L, Nash MA, Kaplan BS. Hypothesis: Dent disease is an underrecognized cause of focal glomerulosclerosis. Clin J Am Soc Nephrol 2007;2:914–918.
- 64. Lowe CU, Terrey M, MacLachlan EA. Organic aciduria, decreased renal ammonia production, hydrophthalmos and mental retardation: A clinical entity. Am J Dis Child 1952;83:164–184.
- 65. Lin T, Lewis RA, Nussbaum RI. Molecular confirmation of carriers of Lowe syndrome. Ophthalmology 1999;106:119–122.
- 66. Charnas LR, Bernardini I, Rader D et al. Clinical and laboratory findings in the oculocerebrorenal syndrome of Lowe, with special reference to growth and renal function. N Engl J Med 1991;324: 1318–1325.
- 67. Laube G, Russel-Egitt I, van't Hoff W. Early proximal tubular dysfunction in Lowe's syndrome. Arch Dis Child 2004;89:479–480.
- 68. Attree O, Olivos IM, Okabe I et al. The Lowe's oculocerebrorenal syndrome gene encodes a protein highly homologous to inositol polyphosphate-5-phosphatase. Nature 1992;358:239–242.
- 69. Zhang X, Jefferson AB, Auethavekiat V et al. The protein deficient in Lowe syndrome is a phosphatidylinositol 4,5-bisphosphate 5-Phosphatase. Proc Natl Acad Sci USA 1995;92:4853–4856.
- 70. Lin T, Orrison BM, Leahey AM et al. Spectrum of mutations in the OCRL1 gene in the Lowe oculocerebrorenal syndrome. Am J Hum Genet 1997;60:1384–1388.
- 71. Zhang X, Hartz PA, Philip E et al. Cell lines from kidney proximal tubules of a patient with Lowe syndrome lacks OCRL inositol polyphosphate 5-phosphatase and accumulate phosphatidylinositol 4,5-bisphosphate. J Biol Chem 1998;273:1574–1582.
- 72. Suchy SF, Nussbaum RL. The deficiency of PIP₂ 5-phosphatased in Lowe syndrome affects actin polymerization. Am J Hum Genet 2002;71:1420–1427.
- 73. Ungewickell A, Ward M, Ungewickell E et al. The inositol polyphosphate 5-phosphatase Ocrl associates with endosome that are partially coated with clathrin. Proc Natl Acad Sci USA 2004; 101:13501–13506.
- 74. Lowe M. Structure and function of Lowe syndrome protein. Traffic 2005;6:711–719.
- 75. Erdmann KS, Mao Y, McCrea HJ et al. A role of Lowe syndrome protein OCRL in early steps of the endocytotic pathway. Dev Cell 2007;13:377–390.
- 76. Faucherre A, Desbois P, Satre V et al. Lowe syndrome protein OCRL interacts with Rac GTPase in the trans-Golgi network. Hum Mol Genet 2003;12:2449–2456.
- 77. Hatefi Y. The mitochondrial electron transport and oxidative phosphorylation system. Annu Rev Biochem 1985;54:1015–1069.
- 78. Clayton DA. Structure and function of the mitochondrial genome. J Inherit Metab Dis 1992;15:439–447.
- 79. DiMauro S, Bonilla E, Lombes A et al. Mitochondrial encephalomyopathies. Neurol Clin 1990;8:483–506.
- 80. Niaudet P. Mitochondrial disorders and the kidney. Arch Dis Child 1998;78:387–390.
- 81. Ueda Y, Ando A, Nagata T et al. A boy with mitochondrial disease: Asymptomatic proteinuria without neuromyopathy. Pediatr Nephrol 2004;19:107–110.
- 82. Morris AA, Taylor RW, Birchi-Marchin MA et al. Neonatal Fanconi syndrome due to deficiency of complex III of the respiratory chain. Pediatr Nephrol 1995;9:407–411.
- 83. Kuwertz-Broking E, Koch HG, Marquardt T et al. Renal Fanconi syndrome: First sign of partial respiratory chain complex IV deficiency. Pediatr Nephrol 2000;14:495–498.
- 84. Au KM, Lau SC, Mak YF et al. Mitochondrial DNA deletion in a girl with Fanconi syndrome. Pediatr Nephrol 2007;22:136–140.
- 85. Tzen CY, Tsai JD, Wu TY et al. Tubulointerstitial nephritis associated with a novel mitochondrial point mutation. Kidney Int 2001; 59:846–854.
- 86. Szabolcs MJ, Seigle R, Shanske S et al. Mitochondrial DNA deletion: A cause of chronic tubulointerstitial nephropathy. Kidney Int 1994;45:1388–1396.
- 87. Mochizuki H, Joh K, Kawame H et al. Mitochondrial encephalomyopathies preceded by de Toni-Debré-Fanconi syndrome or focal segmental glomerulosclerosis. Clin Nephrol 1996;46:347–352.
- 88. Gucer S, Talim B, Asan E et al. Focal segmental glomerulosclerosis associated with mitochondrial cytopathy: Report of two cases with special emphasis on podocytes. Pediatr Dev Pathol 2005;8:710–717.
- 89. Hotta O, Inoue CN, Miyabayashi S et al. Clinical and pathologic features of focal segmental glomerulosclerosis with mitochondrial tRNA^{Leu(UUR)} gene mutation. Kidney Int 2001;59:1236-1243.
- 90. Barisoni L, Diomedi-Camassei F, Santorelli FM et al. Collapsing glomerulopathy associated with inherited mitochondrial injury. Kindey Inter 2008;74:237–243.
- 91. Lopez LC, Schuelke M, Quinzii CM et al. Leigh syndrome with nephropathy and CoQ10 deficiency due to decaprenyl diphophate synthase subunit 2 (PDSS2) mutations. Am J Hum Genet 2006;79:1125–1129.
- 92. Niaudet P, Heidet L, Munnich A et al. Deletion of the mitochondrial DNA in a case of de Toni-Debré-Fanconi syndrome and Pearson syndrome. Pediatr Nephrol 1994;8:164–168.
- 93. Zaffanello M, Zamboni G. Therapeutic approach in a case of Pearson's syndrome. Minerva Pediatr 2005;57:143–146.
- 94. Matsutani H, Mizusawa Y, Shimoda M et al. Partial deficiency of cytochrome c oxidase with isolated proximal renal tubular acidosis and hypercalciuria. Clin Nephrol Urol 1992;12:221–224.
- 95. Goto Y, Itami N, Kajii N et al. Renal tubular involvement mimicking Bartter syndrome in a patient with Kearn-Sayre syndrome. J Pediatr 1990;116:904–910.
- 96. Moraes CT, Shanske S, Trischler HJ et al. Mitochondrial DNA depletion with variable tissue expression: A novel genetic abnormality in mitochondrial disease. Am J Hum Genet 1991;48:492–501.
- 97. Gilber RD, Emms M. Pearson's syndrome presenting with Fanconi syndrome. Ultrastruct Pathol 1996;20:473–475.
- 98. Gahl WA, Thoene JG, Schneidel JA. Cystinosis. N Engl J Med 2003;347:111–121.
- 99. van't Hoff WG, Ledermann SE, Waldron M et al. Early-onset chronic renal failure as a presentation of infantile nephropathy cystinosis. Pediatr Nephrol 1995;9:483–484.
- 100. Pennesi M, Marchetti F, Crovella S et al. A new mutation in two siblings with cystionosis presenting with Bartter syndrome. Pediatr Nephrol 2005;20:217–219.
- 101. Yildiz B, Durmus-Aydogdu S, Kural N et al. A patient with cystinosis presenting transient features of Bartter syndrome. Turk J Pediatr 2006;48:260–262.
- 102. Theodoropolos DS, Shawker TH, Heinrichs C et al. Medullary nephrocalcinosis in nephropathic cystinosis. Pediatr Nephrol 1995;9:412–418.
- 103. Gubler MC, Lacoste M, Sich M et al. The pathology of the Kidney in Cystinosis. Paris, France, Elsevier, 1999.
- 104. Servais A, Moriniere V, Grünfeld JP et al. Late onset nephropathic cystinosis: Clinical presentation, outcome, and genotyping. Clin J Am Soc Nephrol 2008;3:27–35.
- 105. Town M, Jean G, Cherqui S et al. A novel gene encoding an integral membrane protein is mutated in nephropathic cystinosis. Nature Get 1998;18:319–324.
- 106. Anikster Y, Lucero C, Guo J et al. Ocular nonnephropathic cystinosis: Clinical, biochemical, and molecular correlations. Pediatr Res 2000;47:17–23.
- 107. Cherqui S, Sevin C, Hamard G et al. Intralysosomal cystine accumulation in mice lacking cystinosis, the protein defective in cystinosis. Mol Cell Biol 2002;22:7622–7632.
- 108. Cerinkys I, Schlatter E, Hirsch JR et al. Inhibition of Na⁺-dependent transporters in cystine-loaded human renal cells: Electrophysiological studies on the Fanconi syndrome. J Am Soc Nephrol 2002;13:2085–2093.
- 109. Park MA, Thoene JG. Potential role of apoptosis in development of the cystinotic phenotype. Pediatr Nephrol 2005;20:441–446.
- 110. Wilmer MJ, de Graaf-Hess A, Blom HJ et al. Elevated oxidative glutathione in cystinotic proximal tubular epithelial cells. Biochem biophys Res Commun 2005;337:610–614.
- 111. Bonsib SM, Horvth F Jr. Multinucleated podocytes in a child with nephrotic syndrome and Fanconi's syndrome: A unique clue to the diagnosis. Am J Kidney Dis 1999;34:966–971.
- 112. Spear GS, Slusser RJ, Tousimis AJ et al. Cystinosis. An ultrastructural and electron-probe study of the kidney with unusual findings. Arch Pathol 1971;91:206–221.
- 113. Kleta R, Gahl WA. Pharmacological treatment of nephropathic cystinosis with cysteamine. Expet Opin Pharmacother 2004;5: 2255–2262.
- 114. Gahl WA, Kuehl EM, Iwata F et al. Corneal crystals in nephropathic cystinosis: natural history and treatment with cysteamine eye drops. Mol Genet Metab 2000;71:100–120.
- 115. Kleigman RM, Sparks JW. Perinatal galctose metabolism. J Pediatr 1985;107:831–841.
- 116. Tyfield L, Reichardt J, Fridovich-Keil J et al. Classical galactosemia and mutation at the galactose-1-uridyl transferase (GALT) gene. Human Mutat 1999;13:417–430.
- 117. Waggoner DD, Buist NRM, Donnel GN et al. Long-term prognosis in galactosemia: Results in a survey of 350 cases. J Inherit Metab Dis 1990;13:802–818.
- 118. Waggoner DD, Buist NRM. Long-term complications in treated galactsemia-175 U.S. cases. Int Pediatr 1993;8:97–199.
- ment. EurJ Pediatr 1995;154:S87–S92. 120. Berry GT, Palmieri M, Gross KC et al. The effect of dietary fruits and
- vegetables on urinary galactitol excretion in galactose-1-phosphate uridyltransferase deficiency. J Inherit Metab Dis 1993;16:91–100.
- 121. Berry GT, Mate PJ, Reynold RA. The rate of de novo galactose synthesis in patients with galactose-1-phosphate uridyltransferase deficiency. Mol Genet Metab 2004;81:22–30.
- 122. Berry GT, Nissim I, Lin Z et al. Endogenous synthesis of galactose in normal men and patients with hereditary galactosemia. Lancet 1995;346:1073–1074.
- 123. Gitzelmann R, Wells HJ, Segal S. Galactose metabolism in a patient with hereditary galactokinase deficiency. Eur J Clin Invest 1974;4: 79–84.
- 124. Gitzelmann R. Additional findings in galactokinase deficiency. J Pediatr 1975;87:1007–1008.
- 125. Slepak TI, Tang M, Slepak VZ et al. Involvement of endoplasmic reticulum stress in a novel classic galactosemia model. Mol Genet Met 2007;92:78–87.
- 126. Lai KW, Cheng LY, Choung AL et al. Inhibitor of apoptosis proteins and ovarian dysfunction in galactosemic rats. Cell Tissue Res 2003;311:417–425.
- 127. Chung MA. Galactosemia in infancy: Diagnosis, management, and prognosis. Pediatr Nurs 1997;23:563–469.
- 128. Ali M, Rellos P, Cox TM. Hereditary fructose intolerance. J Med Genet 1998;35:353–365.
- 129. Rottmann WH, Tolan DR, Penhoet EE. Complete amino acid sequence for human aldolase B derived from cDNA and genomic clones. Proc Natl Acad Sci USA 1984;81:2738–2742.
- 130. Mukai T, Yatsuki H, Joh K et al. Human aldolase b gene: Characterization of the genomic aldolase B gene and analysis of sequences required for multiple polyadenylations. J Biochem 1987;102:1043–1051.
- 131. Esposito G, Vitagliano L, Santamaria R et al. Structural and functional analysis of aldolase B mutants related to hereditary fructose intolerance. FEBS Lett 2002;531:152–156.
- 132. Cross NC, Cox TM. Hereditary fructose intolerance. Int J Biochem 1990;22:685–689.
- 133. Morris RC Jr. An experimental renal acidification defect in patients with hereditary fructose intolerance: I. Its resemblance to renal tubular acidosis. J Clin Invest 1967;47:1389–1398.
- 134. Morris RC Jr. An experimental renal acidification defect in patients with hereditary fructose intolerance: II. Its distinction from classic renal acidosis and its resemblance to the renal acidification defect associated with the Fanconi syndrome of children with cystinosis. J Clin Invest 1968;47:1648–1663.
- 135. Richardson RMA, Little JA, Pattern RL et al. Pathogenesis of acidosis in hereditary fructose intolerance. Metabolism 1979;28:1133–1138.
- 136. Levin B, Snodgrass GLAI, Oberholzer VG et al. Fructosemia. Observations in seven cases. Am J Med 1968;45:826–838.
- 137. Lu M, Holliday LS, Zhang L et al. Interaction between aldolase and vacuolar H⁺-ATPase: Evidence for direct coupling of glycolysis to the ATP-hydrolyzing proton pump. J Biol Chem 2001;276:30407–30413.
- 138. Steinmann B, Gitzelmann R. The diagnosis of hereditary fructose intolerance. Helv Paediatr Acta 1981;36:297–316.
- 139. Müller P, Meier C, Böhme HJ et al. Fructose breath hydrogen test- is it really a harmless diagnostic procedure? Dig Dis 2003;21:276–278.
- 140. Chou JY, Matern D, Mansfield BC et al. Type I glycogen storage diseases: disorders of the glucose-6-phosphatase complex. Curr Mol Med 2002;2:121–143.
- 141. von Gierke E. Hepato-nephro-megalia glycogenica (Glykogenespecicher-krankheit der Lber und Nieren). Beitr Pathol Anat 1929;82: 497–513.
- 142. Kim Sy, Vhen Ly, Yiu WH et al. Neutrophilia and elevated serum cytokines are implicated in glycogen storage disease type Ia. FEBS Lett 2007;581:3833–3838.
- 143. Rocco Di, Calevo MG, Taro's M et al. Hepatocellular adenoma and metabolic balance in patients with type Ia glycogen storage disease. Mol Genet Metab 2008;93:398–401.
- 144. Reddy SK, Kishnani PS, Sullivan JA et al. Resection of hepatocellular adenoma in patients with glycogen storage disease type Ia. J Hepatol 2007;47:658–663.
- 145. Reitsma-Bierens WCC. Renal complications in glycogen storage disease type I. Eur J Pediatr 1993;152:S60–S62.
- 146. Hers HG, van Hoof F, de Barsy T. Glycogen storage disease. In The Metabolic Basis of Inherited Disease, 6th edn. Scriver CR, Beaudet Al, Sly WS et al. (eds.) New York, McGraw-Hill Inc, 1989, pp. 425–437.
- 147. Matsuo N, Tsuchiya M, Cho H et al. Proximal renal tubular acidosis in a child with type I glycogen storage disease. Acta Pediatr Scand 1986;75:332–335.
- 148. Chen YT, Scheinman JI, Park HK et al. Amelioration of proximal renal tubular dysfunction in type I glycogen storage disease with dietary therapy. N Engl J Med 1990;323:590–593.
- 149. Chen YT, Coleman RA, Scheinman JI et al. Renal disease in type I glycogen storage disease. N Engl J Med 1988;318:7–11.
- 150. Verani R, Bernstein J. Renal glomerular and tubular abnormalities in glycogen storage disease type I. Arch Pathol Lab Med 1988;112: 271–274.
- 151. Baker L, Dahlem S, Goldfarb S et al. Hyperfiltration and renal disease in glycogen storage disease. Kidney Int 1989;35:1345–1350.
- 152. Weinstein DA, Somers MJ, Wolfsdorf JI. Decreased urinary citrate excretion in type 1a glycogen storage disease. J Pediatr 2001;138: 378–382.
- 153. Rake JP, Visser G, Labrune P et al. Glycogen storage disease type I: Diagnosis, management, clinical course and outcome. Results of the European study on glycogen storage disease type I (ESGSD I). Eur J Pediatr 2002;161:S20–S34.
- 154. Yiu WH, Pan C-J, Ruef RA et al. Angiotensin mediates renal fibrosis in the nephropathy of glycogen storage disease type I. Kidney Int 2008;73:716–723.
- 155. Urushihara M, Kagami S, Ito M et al. Transforming growth factorbeta in renal disease with glycogen storage disease I. Pediatr Nephrol 2004;19:676–678.
- 156. Greene HL, Slonim AE, O'Neill JA Jr. et al. Continuous nocturnal intragastric feeding for management of type 1 glycogen storage disease. N Engl J Med 1976;294:423–425.
- 157. Wolfsdorf JI, Crigler JF Jr. Cornstarch regimens for nocturnal treatment of young adults with type I glycogen storage disease. Am J Clin Nutr 1997;65:1507–1511.
- 158. Chen YT, Cornblath M, Sidbury JB et al. Cornstarch therapy in type I glycogen storage disease. N Engl J Med 1984;310:171–175.
- 159. Iyer SG, Chen CL, Wang CC et al. Long-term results of living donor liver transplantation for glycogen storage disorders in children. Liver Transpl 2007;13:848–852.
- 160. Lee KW, Lee JH, Shin SW et al. Hepatocyte transplantation for glycogen storage type Ib. Cell Transplant 2007;16:629–637.
- 161. Fanconi G, Bickel H. Die chronishe aminoaidurie (aminosäurendiabetes oder nehrotishßglukosurisher zwergwuchs) bei der glykogenose und der cystinkrankhein. Helv Paediatr Acta 1949;4:359–396.
- 162. Manz F, Bickel H, Brodehl J et al. Fanconi-Bickel syndrome. Pediatr Nephrol 1987;1:509–519.

Fanconi Syndrome 1065

- 163. Furlan F, Santer R, Vismara E et al. Bilateral nuclear cataracts as the first neonatal sign of Fanconi-Bickel syndrome. J Inherit Metab Dis 2006;29:685.
- 164. Santer R, Schneppenheim R, Dombrowski A et al. Fanconi-Bickel syndrome- a congenital defect of the liver-type facilitative glucose transporter. J Inherit Metab Dis 1998;21:191–194.
- 165. Yoo H-W, Shin Y-K, Seo E-J et al. Identification of a novel mutation in then GLUT2 gene in a patient with Fanconi-Bickel syndrome presenting with neonatal diabetes mellitus and galactosaemia. Eur J Pediatr 2002;161:351–353.
- 166. Santer R, Schneppenheim R, Dombrowski A et al. Mutations in GLUT2, the gene for the liver-type glucose transporter, in patients with Fanconi-Bickel syndrome. Nat Genet 1997;17:324–326.
- 167. Santer R, Groth S, Kinner M et al. The mutation spectrum of the facilitative glucose transporter gene SLC2A2 (GLUT2) in patients with Fanconi-Bickel syndrome. Hum Genet 2002;110:21–29.
- 168. Bell GI, Burnant CF, Takeda J et al. Structure and function of mammalian facilitative sugar transporters. J Biol Chem 1993;268: 19161–19164.
- 169. Berry GT, Baker L, Kaplan FS et al. Diabetes-like renal glomerular disease in Fanconi-Bickel syndrome. Pediatr Nephrol 1995;9: 287–291.
- 170. Lee PJ, van't Hoff WG, Leonard JV. Catch-up growth in Fanconi-Bickel syndrome with uncooked cornstarch. J Inherit Metab Dis 1995;18:153–156.
- 171. Riva S, Ghisalberti C, Parini R et al. The Fanconi-Bickel syndrome: a case of neonatal onset. J Perinatol 2004;24:322–323.
- 172. Berfer R, Smit GP, Stoker, de Varies SA et al. Deficiency of fumarylacetoacetase in a patient with hereditary tyrosinemia. Clin Chim Acta 1981;114:37–44.
- 173. Kvittingen EA, Jellum E, Stokke O et al. Assay of fumarylacetoacetate fumarylhydrolase in human liver: Deficient activity in a case of hereditary tyrosinemia. Clin Chim Acta 1981;115:311–319.
- 174. Holme E, Lindstedt S. Diagnosis and management of tyrosinemia type I. Curr Opin Pediatr 1995;6:726–732.
- 175. Weinberg AG, Mize CE, Worthen HG. The occurrence of hepatoma in the chronic form of hereditary tyrosinemia. J Pediatr 1976;88: 434–438.
- 176. Castilloux J, laberge AM, Martin SR et al. "Silent" tyrosinemia presenting as hepatocellular carcinoma in a 10-year-old girl. J Pediatr Gastroenterol Nurs 2007;44:375–377.
- 177. Mitchell G, Larochell J, Lambert M et al. Neurologic crises in hereditary tyrosinemia. N Eng J Med 1990;322:432–437.
- 178. Freeto S, Mason D, Chen J et al. A rapid ultra performance liquid chromatography tandem mass spectrometric method for measuring amino acids associated with maple syrup urine disease, tyrosinemia and phenylketonuria. Ann Clin Biochem 2007; 44:474–481.
- 179. Pardis K, Weber A, Seidman EG et al. Liver transplantation for hereditary tyrosinemia: The Quebec experience. Am J Hum Genet 1990;47:338–342.
- 180. Nissenkorn A, korman SH, Vardi O et al. Carnitine-deficient myopathy as a presentation of tyrosinemia type I. J Child Neurol 2001;16:642–644.
- 181. Endo F, Sun MS. Tyrosinemia type I and apoptosis of hepatocytes and renal tubular cells. J Inhert Metab Dis 2002;25:227–234.
- 182. Nakamura K, Tanaka Y, Mitsubishi H et al. Animal models of tyrosinemia. J Nutr 2007;137:1556S–1560S.
- 183. Spencer PD, Medow MS, Moses LC et al. Effects of succinylacetone on the uptake of sugars and amino acids by brush border vesicles. Kidney Int 1988;34:671–677.
- 184. Roth KS, Carter BE, Higgins ES. Succinylacetone effects on renal tubular phosphate metabolism: A new model for experimental Fanconi syndrome. Proc Soc Exp Biol Med 1991; 196:428–431.
- 185. Fairney A, Francis D, Ersser RS et al. Diagnosis and treatment of tyrosinosis. Arch Dis Child 1968;43:540–547.
- 186. Masurl-Paulet A, Poggi-Bach J, Rolland MO et al. NTBC treatment in tyrosinemia type I: long-term outcome in French patients. J Inherit Meteb Dis 2008;31:81–87.
- 187. Koelink CJ, van Hasselt P, van der Ploeg A et al. Tyrosinemia type I treated by NTBC: how does AFP predict liver cancer? Mol Genet Metab 2006;89:310–315.
- 188. Shoemaker LR, Strife CF, Balisteri WF et al. Rapid improvement of the renal tubular dysfunction associated with tyrosinemia after hepatic replacement. Pediatrics 1992;89:251–255.
- 189. Das SK, Ray K. Wilson's disease: an update. Nat Clin Pract Neurol 2006;2:482–493.
- 190. Bull PC, Thomas GR, Rommens JM et al. The Wilson disease is a putative copper transporting P-type ATPase similar to the Menkes gene. Nat Genet 1993;5:327–337.
- 191. Figus A, Angius A, Loudianos G et al. Molecular pathology and haplotype analysis of Wilson disease in Mediterranean population. Am J Hum Genet 1995;57:1318–1324.
- 192. Vulpe C, Levinson B, Whitney S et al. Isolation of a candidate gene for Menkes disease and evidence that it encodes a coppertransporting ATPase. Nat Genet 1993;3:7–13.
- 193. Yang XL, Miura N, Kawarada Y et al. Two forms of Wilson disease protein produced by alternative splicing are localized in distinct cellular compartments. Biochem J 1997;326:897–902.
- 194. Reynolds ES, Tannen RL, Tyler HR. The renal lesion in Wilson's disease. Am J Med 1966;40:518–537.
- 195. Sozeri E, Feist D, Ruder H et al. Proteinuria and other renal functions in Wilson's disease. Pediatr Nephrol 1997;11:307–311.
- 196. Kalra V, Mahjan S, Kesarwani PK et al. Rare presentation of Wilson's disease: A case report. Int Urol Nephrol 2004;36:289–291.
- 197. Fulop M, Sternlieb I, Scheinberg IM. Defective urinary acidification in Wilson's disease. Ann Intern Med 1968;68:770–777.
- 198. Leu ML, Strickland GT, Gutman RA. The renal lesion on Wilson's disease: Response to penicilamine therapy. Am J Med Sci 1970;260: 381–398.
- 199. Elasas LG, Hayslett Jp, Sprgo BH et al. Wilson's disease with reversible renal tubular dysfunction. Correlation with proximal tubular ultrastructure. Ann Intern Med 1971;75:427–433.
- 200. Ala A, Borjigin J, Rochwarger A et al. Wilson disease in septuagenarian siblings: Raising the bar for diagnosis. Hepatology 2005;41: 668–670.
- 201. Page RA, Davie CA, McManus D et al. Clinical correlation of brain MRI and MRS abnormalities in patients with Wilson disease. Neurology 2004;63:638–643.
- 202. Kuruvilla A, Joseh S. ''Face of the giant panda'' sighn in Wilson's disease; revisited. Neurol India 2000;48:395–396.
- 203. Ala A, Walker A, Ashkan K et al. Wilson's disease. Lancet 2007;369:397–408.
- 204. Brewer GJ, Dick RD, Johnson V et al. Treatment of Wilson's disease with zinc: XV. Long-term follow-up. J Lab Clin Med 1998;132: 264–278.
- 205. Czlonkowska A, Gajda J, Rodo M. Effects of long-term treatment in Wilson's disease with D-penicillamine and zinc sulphate. J Neurol 1996;243:269–273.
- 206. Ben-Ishay D, Dreyfuss F, Ylmann TD. Fanconi syndrome with hypouricemia in an adult. Am J Med 1961;31:793–800.
- 207. Sheldon W, Luder J, Webb B. A familial tubular absorption defect of glucose and amino acids. Arch Dis Child 1961;36:90–95.
- 208. Friedman AL, Trygstad CW, Chesney RW. Autosomal dominant Fanconi syndrome with early renal failure. Am J Clin Genet 1978;2:225–232.
- 209. Patrick A, Vameron JS, ogg CS. A family with a dominant form of idiopathic Fanconi syndrome leading to renal failure in adult life. Clin Nephrol 1981;16:289–292.
- 210. Wen SF, Friedman AL, Oberley TD. Two case studies from a family with primary Fanconi syndrome. Am J Kidney Dis 1989;13:240–246.
- 211. Tolaymat A, Sakarcan A, Neiberger R. idiopathic Fanconi syndrome in a family. Part I. Clinical aspects. J Am Soc Nephrol 1992;2: 1310–1317.
- 212. Tieder M, Sakarcan A, Neiberger R. Elevated serum 1,25-dihydroxyvitamin D concentrations in siblings with primary Fanconi's syndrome. N Engl J Med 1988;319:845–849.
- 213. Wornell P, Crocker J, Wade A et al. An Acadian variant of Fanconi syndrome. Pediatr Nephrol 2007;22:1711–1715.
- 214. Nieman N, Pierson M, Marchal C et al. Nephropathie familiale glomerulotubulaire avec syndrome de Toni-Debré-Fanconi. Arch Fr Pediatr 1968;25:43–69.
- 215. McVicar M, Exeni R, Susin M. Nephrotic syndrome and multiple tubular defects in children: an early sign of focal segmental glomerulosclerosis. J Pediatr 1980;97:918–922.
- 216. Batuman V. Proximal tubular injury in myeloma. Contrib Nephrol 2007;153:87–104.
- 217. Ren H, Wang W-M, Chen X-N et al. Renal involvement and follow up of 130 patients with primary Sjögren syndrome. J Rheumatol 2008;35:278–284.
- 218. Yang Y-S, Peng C-H, Sia S-K et al. Acquired hypophosphatemia osteomalacia associated with Fanconi's syndrome in Sjögren syndrome. Rheumatol Int 2007;27:593–597.
- 219. Friedman AL, Chesney R. Fanconi's syndrome in renal transplantation. Am J Nephrol 1981;1:145–147.
- 220. Dobrin RS, Vernier RL, Fish AJ. Acute eosinophilic interstitial nephritis and renal failure with bone marrow-lymph node granuloma and anterior uveitis. Am J Med 1975;59:325–333.
- 221. Igarashi T, Kawato H, Kamoshita S et al. Acute tubulointersitial nephritis with uveitis sydnorme presenting as multiple tubular dysfunction including Fanconi's syndrome. Pediatr Nephrol 1992;6:547–549.
- 222. Tung KS, Black WC. Association of renal glomerular and tubular immune complex disease and autoimmune basement membrane antibody. Lab Invest 1975;32:696–700.
- 223. Griswold WR, Krous HF, Reznik V et al. The syndrome of autoimmune interstitial nephritis and membranous nephropathy. Pediatr Nephrol 1997;11:699–702.
- 224. Makker SP, Widstrom R, Huang J. Membranous nephropathy, interstitial nephritis, and Fanconi syndrome – glomerular antigen. Pediatr Nephrol 1996;10:7–13.
- 225. Alexandridis G, Liamis G, Elisaf M. Reversible tubular dysfunction that mimicked Fanconi's syndrome in a patient with anorexia nervosa. Int J Eat Disord 2001;30:227–230.
- 226. Igarashi T, Kawato H, Kamoshita S. Reversible low-molecularweight proteinuria in patients with distal renal tubular acidosis. Pediatr Nephrol 1990;4:593–596.
- 227. Watanabe T. Proximal renal tubular dysfunction in primary distal renal tubular acidosis. Pediatr Nephrol 2005;20:86–88.
- 228. Cleveland WW, Adams WC, Mann JC et al. Acquired Fanconi syndrome following degraded tetracycline. J Pediatr 1965;66: 333–342.
- 229. Gainza FJ, Minguela JI, Lampreabe I. Aminoglycoside-associated Fanconi's syndrome: an underrecognized entity. Nephron 1997;77: 205–211.
- 230. Ghiculescu R, Kubler P. Aminoglycoside-associated Fanconi syndrome. Am J Kidney Dis 2006;48:E89–E93.
- 231. Tsimihodiomos V, Psychogios N, Kakaidi V et al. Salicylate-induced proximal tubular dysfunction. Am J Kidney Dis 2007;50:463–467.
- 232. Watanabe T, Yoshikawa H, Yamazaki S et al. Secondary renal Fanconi syndrome caused by valproate therapy. Pediatr Nephrol 2005;20:814–817.
- 233. Zaki EL, Springate JE. Renal injury from valproic acid: Case report and literature review. Pediatr Neurol 2002;27:318–319.
- 234. Bagnis CI, Deray G, Baumelou A et al. Herbs and the kidney. Am J Kidney Dis 2004;44:1–11.
- 235. Hong Y-T, Fu L-S, Chung L-H et al. Fanconi's syndrome, interstitial fibrosis and renal failure by aristolochic acid in Chinese herbs. Pediatr Nephrol 2006;21:577–579.
- 236. Takamoto K, Kawada M, Usui T et al. Aminoglycoside antibiotics reduce glucose reabsrption in kidney through down-regulation of SGLT1. Biochem Biophys Res Commun 2003;308:866–871.
- 237. Humes HD. Aminoglycoside nephrotoxicity. Kidney Int 1988; 900–911.
- 238. Zamialuski-Tucker MJ, Morris ME, Springate JE. Ifosfamide metabolite chloroacetaldehyde causes Fanconi syndrome in the perfused rat kidney. Toxicol Appl Pharmacol 1994;129:170–175.
- 239. Yaseen X, Michoudet C, Baverel G et al. Mechanisms of the ifosfamide-induced inhibition of endocytosis in the rat proximal kidney tubule. Arch Toxicol 2008; On line.
- 240. Pratt CB, Meyer WH, Jenkins JJ et al. Ifosfamide, Fanconi's syndrome, and rickets. J Clin Oncol 1991;9:1495–1499.
- 241. Hanquinet S, Wouters M, Devalck C et al. Increased renal parenchymal echogenicity in ifosfamide-induced renal Fanconi syndrome. Med Pediatr Oncol 1995;24:116–118.
- 242. Badary OA. Taurine attenuates Fanconi syndrome induced by ifosfamide without compromising its antitumor activity. Oncol Res 1998;10:355–360.
- 243. Portill D, Nagothu KK, Megyesi J et al. Metabolomic study of cisplatin-induced nephrotoxiciy. Kidney Int 2006;69:2194–2204.
- 244. François H, Coppo P, Hayman J-P et al. Partial Fanconi syndrome induced by Imanitib therapy: A novel cause of urinary phosphate loss. Am J Kidney Dis 2008;51:298–301.
- 245. Meier P, Dautheville-Gibal S, Ronco PM et al. Cidofovir-induced end-stage renal failure. Nephrol Dial Transplant 2002;17:148–149.
- 246. Ho ES, Lin DC, Mendel DB et al. Cytotoxicity of antiviral nucleotides adefovir and cidofovir is induced by the expression of human renal organic anion transporter 1. J Am Soc Nephrol 2000;11: 383–393.
- 247. Tanji N, Tanji K, Kambham N et al. Adefovir nephotoxicity: Possible role of mitochondrial DNA depletion. Hum Pathol 2001;32: 734–740.
- 248. Daugas E, Rougier J-P, Hill G. HAART-related nephropathies in HIV-infected patients. Kidney Int 2005;67:393–403.
- 249. Verheist D, Monge M, Meynard J-L et al. Fanconi syndrome and renal failure induced by tenofovir: A first case report. Am J Kidney Dis 2002;40:1331–1333.
- 250. Malik A, Abraham P, Malik N. Acute renal failure and Fanconi syndrome in an AIDS patient on tenofovir treatment-case report and review of literature. J Infect 2005;51:e61–e65.
- 251. Gil HW, Yang JO, Lee EY et al. Paraquat-induced Fanconi syndrome. Nephrology (Carlton) 2005;10:430–432.
- 252. Hruz P, Mayr M, Löw R et al. Fanconi's syndrome, acute renal failure, and tonsil ulceration after colloidal bismuth substrate intoxication. Am J Kidney DIS 2002;39:E18.
- 253. Otten J, Vis HL. Acute reversible renal tubular dysfunction following intoxication with methyl-3-choromone. J Pediatr 1968; 73:422–425.
- 254. Butler HE, Morgan JM, Smythe CM. Mercaptopurine and acquired tubular dysfunction in adult nephrosis. Arch Intern Med 1965;116:853–856.
- 255. Moss AH, Gabow PA, Kaehny WD et al. Fanconi syndrome and distal renal tubular acidosis after glue sniffing. Ann Intern Med 1980;92:69–70.
- 256. Barbier O, Jacquillet G, Tau M et al. Effect of heavy metals on, and handling by, the kidney. Nephron Physiol 2005;99:105–110.
- 257. Chisolm JJ, Harrison HC, Eberlein WE et al. Aminoaciduria, hyperphosphaturia and rickets in lead poisoning. Am J Dis Child 1955;89:159–168.
- 258. Logman-Adham M. Aminoaciduira and glycosuria following severe childhood lead poisoning. Pediatr Nephrol 1998;12:218–221.
- 259. Goyer RA, Tsuchuja K, Leonard DL et al. Aminoaciduria in Japanese workers in the lead and cadmium industries. Am J Clin Pathol 1972;57:635–642.
- 260. Uetani M, Kobayashi E, Suwazono Y et al. Investigation of renal damage in the cadmium-polluted Jinzu River basin, based on health examinations in 1967 and 1968. Int J Environ Health Res 2007;17:231–242.
- 261. Takebayashi S, Jimi S, Segawa M et al. Mitochondrial DNA deletion of proximal tubules is the result of itai-itai disease. Clin Exp Nephrol 2003;7:18–26.
- 262. Elizbieta S-J, Roman L. Metabolic bone disease in children: etiology and treatment options. Treat Endocrinol 2006:5;297–318.
- 263. Plank C, Konrad M, Dörr HG et al. Growth failure in a girl with Fanconi syndrome and growth hormone deficiency. Nephrol Dial Transplant 2004;19:1910–1912.