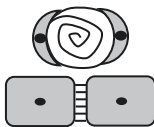




# Chapter 14. Potpourri of Quick Morphologic References

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## Tumor Differentials 101

### Differentials 101: Generic Tumor Types<sup>1</sup> (Main tumor types seen in ALMOST any organ)

Tumor type	Key features	Key immunostains
<b>Carcinoma</b>	<p>The hallmarks of epithelial cells are cohesiveness (cells stick together), distinct cell borders, and usually abundant cytoplasm (cells resembling this description are called “epithelioid”). Main types of carcinoma are listed below:</p> <p><b>Squamous cell carcinoma (SqCC)<sup>2</sup></b> The two hallmark features are as follows:</p> <ol style="list-style-type: none"> <li><b>Keratinization</b> – manifesting as keratin pearls/squamous eddies or isolated cells with glassy salmon-pink cytoplasm/dyskeratotic cells:</li> <li><b>Intercellular bridges</b> – desmosomes seen in the prickle cell layer of the epidermis: These two features may not be appreciable in nonkeratinizing or basaloid SqCC</li> </ol> <p><b>Adenocarcinoma<sup>2</sup></b></p> <ul style="list-style-type: none"> <li>Easy! All you need is gland (or papillae, micropapillae) formation, however focal. Don’t be fooled by neuroendocrine or neuroblastic rosettes though, which can mimic glands. Also, some tumors can become discohesive and mimic true glands (e.g., acantholytic SqCC).</li> <li>Intracellular mucin is another hint. Detection of mucin may be aided by a mucicarmine stain.</li> </ul> <p><b>Papillary carcinoma</b></p> <ul style="list-style-type: none"> <li>Papillary carcinomas may be squamous, urothelial, or glandular, depending on the covering epithelium. Fibrovascular cores are a defining feature.</li> <li>Note that papillary morphology applies to both in situ and invasive lesions (curiously, in situ lesions do not usually invade as papillary carcinomas – e.g., IPMN usually invades as a colloid CA, etc.).</li> <li>DDx: ovarian serous CA, lung, kidney, thyroid.</li> <li>Beware – not all that is papillary is a carcinoma (e.g., mesothelioma, myxopapillary ependymoma, papillary meningioma).</li> <li><b>Micropapillary CA</b> is a variant of papillary CA, which is defined by the absence of fibrovascular cores in the papillae (also typical are clear halos/retraction artifact around micropapillae). DDx includes the ovary (micropapillary serous CA), bladder, breast, lung, and salivary gland. Behavior is typically aggressive. Lymphovascular invasion is nearly universal.</li> </ul>	<p>Epithelial markers (CK, EMA) +</p>   
<b>Neuroendocrine (NE) neoplasm</b>	<p>This umbrella category encompasses NE neoplasms that are either low grade (e.g., carcinoid) or high grade (e.g., small cell CA, Merkel cell CA). Both types share a set of defining “NE features”:</p> <ul style="list-style-type: none"> <li>NE cytology: overall nuclear uniformity/monotony (even when high grade), stippled evenly distributed “salt and pepper” chromatin; absence of prominent nucleoli is key (although there are exceptions); low-grade lesions may have scattered cells showing “endocrine atypia,” which manifests as large bizarre nuclei with smudged/hyperchromatic chromatin.</li> <li>NE architecture: nests, trabeculae, ribbons, rosettes (subtle in high-grade lesions).</li> </ul>	<p>NE markers (SYN, CHR, CD56, INSM1); CK expression is type-dependent</p>
<b>Small cell carcinoma</b>	<p>Unless otherwise specified, this term implies small cell NE carcinoma, formerly known as “oat cell carcinoma” (note that there are small cell variants of melanoma and some carcinomas).</p> <ul style="list-style-type: none"> <li>Despite the name, small size (&lt;3 lymphocytes) is not the only defining feature.</li> <li>Other key defining features are nuclear molding, very high N/C ratio, lots of mitoses, apoptotic bodies, and geographic necrosis.</li> <li>Crush artifact with DNA streaming and DNA deposition in vessels (“Azzopardi phenomenon”) are characteristic.</li> <li>NE nature is evidenced by uniform distribution of chromatin (despite high grade), inconspicuous nucleoli, and occasional presence of trabeculae and rosettes.</li> </ul>	<p>NE markers (sometimes focal), CK+ (frequently focal), TTF1 often positive (regardless of organ of origin)</p>
<b>Melanoma</b>	<ul style="list-style-type: none"> <li>Can look like anything: epithelioid, spindle cell, small cell, pink cell, clear cell, etc. (remember – melanoma is a “great mimicker” in pathology!)</li> <li>Prominent cherry-red nucleoli and nuclear pseudoinclusions are characteristic.</li> <li>Presence of melanin pigment is diagnostic, but make sure to distinguish it from hemosiderin and tattoo pigment; also some melanomas are amelanotic.</li> </ul>	<p>S100+, SOX10+, Melan-A+, HMB45+, MITF</p>
<b>Lymphoma</b>	<ul style="list-style-type: none"> <li>Sheets of cells, ranging from normal lymphocyte-like (e.g., chronic lymphocytic leukemia) to large epithelioid cells (e.g., diffuse large B-cell lymphoma).</li> <li>Large-cell lymphoma may be histologically indistinguishable from poorly differentiated carcinoma or melanoma.</li> <li>General signature of lymphomas is cellular discohesion (cells fall apart) and clefted (indented) nuclei.</li> <li>On high power (especially cytology), look for lymphoglandular bodies (cytoplasmic fragments).</li> </ul>	<p>CD45+, many other markers</p>
<b>Sarcoma</b>	<ul style="list-style-type: none"> <li>Most commonly composed of spindle or stellate cells but can also look epithelioid, small round blue cell, clear/oncocytic, or highly pleomorphic.</li> <li>Look for specific features of muscle, vascular, neural, or adipocytic differentiation (see below).</li> </ul>	<p>Highly variable, vimentin+ (but not specific)</p>
<p>1. In general, these generic tumors have similar morphology irrespective of the organ of origin, and the origin of metastasis cannot be determined without immunostains or clinical history. However, carcinomas of some organs do have a distinctive morphology. Here are a few notable examples:</p> <ul style="list-style-type: none"> <li>Colon: tall pseudostratified nuclei and “dirty, garland necrosis”</li> <li>Prostate (acinar type): low-grade and usually monomorphic nuclei with prominent nucleoli forming acini</li> <li>Breast: relatively bland cytology, nests and glands (ductal), or solid sheet/single-file plasmacytoid cells (lobular)</li> <li>Endometrioid: complex cribriform structures with pseudostratified columnar cells (some resemblance to colon), variable squamous differentiation</li> <li>Endocervical: resembles endometrioid with prominent apoptotic bodies and apical mitoses</li> <li>Clear cell RCC: nests of cells with clear or eosinophilic cytoplasm, blood-filled follicles, and complete vascular network surrounding each nest</li> </ul> <p>2. Some tumors can have BOTH glandular and squamous differentiation (known in some organs as adenosquamous CAs); adenoCAs that commonly have a squamous component include pancreatic, endometrioid, lung (this is uncommon in the prostate, colon, breast)</p>		

Differentials 101: Tumors by Cell Type	
Tumor type	Differential
<b>Epithelioid tumors</b>	<p>Defined as round, plump cells with some cytoplasm (may be of different quality which generates other patterns as follows) resembling epithelium. Ddx includes “carcinoma-melanoma-lymphoma-sarcoma” (the “Big 4”):</p> <ol style="list-style-type: none"> <li><b>1. Carcinoma:</b> look for evidence of glandular or squamous differentiation, however focal. Associated CIS/dysplasia seals the deal.</li> <li><b>2. Melanoma:</b> look for melanin pigment, nuclear pseudo-inclusions, and cherry-red nucleoli. Associated melanoma in situ clinches the diagnosis.</li> <li><b>3. Lymphoma:</b> DLBCL, ALCL, plasmablastic lymphoma, etc. (these can even stain for cytokeratin and ALCL stains for EMA! Always keep these in Ddx when encountering a poorly differentiated epithelioid tumor).</li> <li><b>4. Sarcoma:</b> think of epithelioid sarcoma, epithelioid hemangioendothelioma and angiosarcoma, myoepithelial tumor, epithelioid GIST, and others.</li> </ol> <p><b>Other:</b> histiocytic neoplasms, germ cell tumors, epithelioid mesothelioma.</p> <p>Diagnosis usually requires immunostains. Typical initial panel includes CK for carcinoma, SOX10/S100 for melanoma, and CD45 for lymphoma (although some lymphomas may be negative for CD45).</p>
<b>Pink cell tumors (oncocytic/eosinophilic cytoplasm)</b>	<p>Defined as epithelioid cells with abundant pink cytoplasm. Ddx has significant overlap with epithelioid pattern. Rhabdoid falls under this category but is specifically coined for cells with dense eosinophilic cytoplasmic inclusions that push the nucleus aside.</p> <ol style="list-style-type: none"> <li><b>1. Many carcinomas:</b> most notably HCC (look for bile pigment and lipid vacuoles), RCC (chromophobe or high-grade clear cell, look for nested pattern and prominent vascularity), adrenocortical CAs, thyroid Hurthle cell CA, parathyroid.</li> <li><b>2. Neuroendocrine tumors:</b> such as paraganglioma and oncocytic carcinoid/NETs</li> <li><b>3. Melanoma:</b> commonly has abundant pink cytoplasm (always think of melanoma in “pink cell tumor” Ddx).</li> <li><b>4. Mesenchymal:</b> ASPS, PEComa, CCS, pleomorphic rhabdomyosarcoma, rhabdoid tumors, epithelioid angiomyolipoma.</li> <li><b>5. Other:</b> oncocytomas of various organs, granular cell tumor</li> </ol> <p>Cytoplasmic granularity in pink cell tumors may be due to the following:</p> <ul style="list-style-type: none"> <li>• Mitochondria – oncocytic neoplasms of the salivary gland or kidney, Hurthle cell neoplasms of the thyroid</li> <li>• Lysosomes – granular cell tumor</li> <li>• Zymogen granules – acinar cell CA of the salivary gland, acinar cell CA of the pancreas</li> <li>• NE granules – carcinoid tumor</li> </ul>
<b>Clear cell (CC) tumors</b>	<p>Defined as epithelioid cells with clear cytoplasm (glycogen, lipid, etc.). Again Ddx has significant overlap with epithelioid pattern. First think carcinoma, but certain types of soft tissue tumors (e.g., CCS, PEComa) and very rarely melanoma and lymphoma can be clear. Ironically, many tumors with “clear cell” in their name can be very eosinophilic and have entirely oncocytic pattern rather than clear cell pattern and vice versa.</p> <p>The first-line differential is ccRCC (most common), adrenocortical CA, and ccHCC (uncommon).</p> <p>Complete list of CC neoplasms is vast (almost any carcinoma can have clear cell change, at least focally). Classic examples are:</p> <ul style="list-style-type: none"> <li>• <i>Head and neck:</i> oncocytic and Hurthle cell neoplasms (these are particularly prone to clear cell change), parathyroid, salivary gland neoplasms (e.g., CC carcinoma of salivary gland, myoepithelial tumors, oncocytic tumors, acinar cell, and mucoepidermoid CAs)</li> <li>• <i>Lung:</i> CC sugar tumor (PEComa), CC SqCC, or less commonly adenoCA</li> <li>• <i>GYN tract:</i> ovarian CC CA</li> <li>• <i>Soft tissue:</i> CCS, PEComa</li> </ul>
<b>Spindle cell tumors</b>	<p>Defined as cells with a long axis (like a spindle) in contrast to the round appearance of epithelioid cells. Remember: spindle cell melanoma and carcinoma are more common than sarcoma in general! Sarcoma diagnosis always comes after exclusion of carcinoma and melanoma (also mesothelioma in some locations)!</p> <p>Cytologic clues to differentiation in mesenchymal spindle cell neoplasms:</p> <ul style="list-style-type: none"> <li>• <b>Smooth muscle:</b> “cigar”-shaped (blunt-ended) nucleus with bubbly eosinophilic cytoplasm (fascicles intersect at right angles)</li> <li>• <b>Skeletal muscle:</b> pink cytoplasmic inclusions with cross-striations and “strap cells” (rhabdomyoblasts)</li> <li>• <b>Fibroblast/myofibroblast:</b> bipolar or stellate nucleus with pointy ends and wispy scant lightly eosinophilic cytoplasm myofibroblasts have more plump appearance, small nucleoli, and amphophilic cytoplasm indicating activated state</li> <li>• <b>Schwannian:</b> “club-” or “bullet”-shaped nuclei (pointed at one end), typically wavy (look for nuclear palisading)</li> <li>• <b>Perineurial:</b> cells are slender and spindle with delicate cytoplasmic processes (storiform pattern)</li> <li>• <b>GIST (pericyte):</b> nucleus is intermediate between smooth muscle (box car) and Schwann cell (pointed). Nuclei can be ridiculously long</li> </ul> <p>Other morphologic clues to differentiation are (highly selected examples):</p> <ul style="list-style-type: none"> <li>• <b>Vascular:</b> channels or slit-like spaces with prominent hemorrhage or cytoplasmic vacuoles with RBCs = vascular differentiation (e.g., angiosarcoma or EHE)</li> <li>• <b>Adipocytic:</b> presence of lipoblasts. Ironically, some non-adipocytic tumors can have lots of fat (SFT/myofibroblastoma), while some adipocytic tumors (myxoid LPS/dedifferentiated LPS) may not show any fat; lipoblast is only required for diagnosis of pleomorphic LPS</li> </ul>
<b>Small round blue cell tumors</b>	<p>Defined as sheets of small round blue cells (duh!). Cells are blue because they have very little cytoplasm. Subtle morphologic hints may be present, but generally diagnosis requires immunostains +/- molecular studies and cytogenetics. Ddx is age-dependent (see below).</p>
<b>Tumors with cytoplasmic vacuoles/inclusions</b>	<p>Can be split into several types:</p> <ul style="list-style-type: none"> <li>• Signet ring cell carcinoma (cytoplasmic mucin vacuole indenting the nucleus): gastric, some lobular breast, urothelial, lung CAs</li> <li>• Rhabdoid tumors (eosinophilic inclusion indenting the nucleus, cells typically discohesive): malignant rhabdoid tumors, carcinomas with rhabdoid component/de-differentiation</li> <li>• Lipoblastic/lipocytic differentiation: lipoblastoma, myxoid liposarcoma, pleomorphic liposarcoma</li> <li>• Intracytoplasmic lumina: vascular tumors such as epithelioid hemangioendothelioma, epithelioid hemangioma, and angiosarcoma may see RBCs in vacuoles, also called “blister” cells</li> <li>• Paranuclear vacuoles: GIST, smooth muscle tumors (bubbly cytoplasm)</li> </ul>

<b>Tumor Differentials 101: Tumors by Architectural Pattern</b> (See Glossary in Chapter 16 for definitions)	
<i>Pattern</i>	<i>Differential diagnosis</i>
<b>Hemangiopericytoma (HPC)-like pattern</b> (branching staghorn-like vessels)	SFT/hemangiopericytoma (prototype), synovial sarcoma (particularly monophasic), myofibroma/myopericytoma, mesenchymal chondrosarcoma, nasal glomangiopericytoma, nasopharyngeal angiofibroma, MPNST, endometrial stromal sarcoma; may be seen in many other soft tissue lesions (not very specific)
<b>Storiform pattern</b> (cartwheel-like arrangement of cells)	DFSP (prototype), dermatofibroma, perineurioma, some undifferentiated pleomorphic sarcomas
<b>Whorling pattern</b> (concentric growth of tumor cells)	Meningioma (prototype), follicular dendritic cell sarcoma, rarely seen in angiomatoid fibrous histiocytoma, dedifferentiated liposarcoma, and inflammatory myofibroblastic tumor
<b>Nested pattern</b> (packets of cells with intervening stroma)	Neuroendocrine tumors such as pheochromocytoma/paraganglioma (prototype; nested pattern referred to as Zellballen in this setting), ccRCC, urothelial CA, melanoma, PEComa, CCS, and others
<b>Alveolar pattern</b> (nests with central discohesion)	ASPS and alveolar rhabdomyosarcoma (prototypes), nested neoplasms may appear alveolar (e.g., RCC)
<b>Herringbone pattern</b> (fascicles alternating at acute angles)	Fibrosarcoma (infantile or adult type), fibrosarcomatous transformation of DFSP, MPNST, synovial sarcoma, biphenotypic sinonasal sarcoma, spindle cell rhabdomyosarcoma, spindle cell pattern of adamantinoma (rare)
<b>Basaloid tumors</b> (resembling basal cell carcinoma)	Basal cell CA (prototype), basaloid SqCC, HPV-related SqCC, adnexal tumors, adenoid cystic CA, and others
<b>Nuclear palisading in spindle cell tumors</b>	Schwannoma (prototype), smooth muscle tumors, GIST
<b>Biphasic tumors</b> (epithelial and stromal components)	Malignant (both components): sarcomatoid CAs/carcinosarcomas (including the bladder, lung, uterus, etc. – any epithelial organ), biphasic synovial sarcoma, biphasic malignant mesothelioma, pulmonary blastoma, biphasic Wilms tumor, and others Malignant (stroma): malignant phyllodes tumor, Müllerian adenosarcoma Benign: fibroadenoma (breast), adenofibroma/adenomyoma (GYN tract), cystic neoplasms with ovarian-type stroma (mucinous cystic neoplasm of the pancreas, mixed epithelial stromal tumor of the kidney, and others) – stromal cells are ER+, benign mixed tumor (skin/soft tissue), or pleomorphic adenoma (salivary gland)

Abbreviations: *ALCL* anaplastic large-cell lymphoma, *ASPS* alveolar soft part sarcoma, *CCS* clear cell sarcoma of soft tissue, *DFSP* dermatofibrosarcoma protuberans, *DLBCL* diffuse large B-cell lymphoma, *EHE* epithelioid hemangioendothelioma, *GIST* gastrointestinal stromal tumor, *LPS* liposarcoma, *MCC* Merkel cell carcinoma, *MPNST* malignant peripheral nerve sheath tumor, *SFT* solitary fibrous tumor

Potpourri of Differentials	
<b>Tumors with prominent lymphocytes</b>	Carcinoma associated with microsatellite instability (GI, endometrial), seminoma, lymphoepithelioma (LE) and LE-like carcinomas, thymoma, inflammatory myofibroblastic tumor, follicular dendritic cell sarcoma, clear cell CA of ovary, HPV+ oropharyngeal CA
<b>Tumors with prominent neutrophils</b>	Hodgkin lymphoma, neutrophil-rich anaplastic large-cell lymphoma, inflammatory leiomyosarcoma, anaplastic CA of the thyroid, anaplastic CA of the pancreas and lung, sarcomatoid renal cell CA, medullary CA of the kidney, NUT carcinoma
<b>Tumors with prominent eosinophils</b>	Hodgkin lymphoma, Langerhans cell histiocytosis, mast cell tumors, myeloid sarcoma (chloroma), glassy cell carcinoma of the cervix, thyroid sclerosing mucoepidermoid CA with eosinophilia, epithelioid hemangioma (i.e., angiolymphoid hyperplasia with eosinophilia)
<b>Tumors with prominent mast cells</b>	Very non-specific but can be seen in synovial sarcoma, neurofibroma, spindle cell lipoma, myxoid liposarcoma, hemangiopericytoma, hairy cell leukemia (particularly in bone marrow), and others (anything myxoid often has accompanying mast cells)
<b>Tumors with extravasated erythrocytes</b>	Kaposi sarcoma, angiosarcoma, nodular fasciitis, inflammatory myofibroblastic tumor, sinonasal glomangiopericytoma
<b>Tumors associated with granulomas</b>	Classic associations – seminoma, Hodgkin lymphoma, lymphomatoid granulomatosis, Lennert lymphoma, also some carcinomas (e.g., reaction to keratin in SqCC or endometrioid CA with squamous differentiation)
<b>Intranuclear pseudoinclusions</b>	Papillary thyroid CA, hyalinizing trabecular tumor of thyroid, medullary thyroid CA (50%), melanoma, meningioma, pheochromocytoma, lung adenoCA, usual ductal hyperplasia, and others
<b>Hyaline globules</b>	Non-specific but classic associations are yolk sac tumor, Kaposi sarcoma, solid-pseudopapillary tumor of the pancreas, HCC, clear cell CA of GYN tract
<b>Psammoma bodies</b>	Any papillary carcinoma (papillary thyroid CA, serous ovarian CA, papillary CA of the lung, papillary RCC), metanephric adenoma, meningioma (and normal meninges), mesothelioma (and benign mesothelial proliferations in peritoneum), duodenal somatostatinoma
<b>Tumors with melanin pigment</b>	Melanoma (#1, 2, and 3 in the differential), clear cell sarcoma of soft tissue (melanoma of soft parts), Bednar tumor (pigmented dermatofibrosarcoma protuberans; produced by intermixed dendritic cells, not tumor cells). Other neural crest-derived tumors occasionally produce melanin (e.g., melanotic schwannoma, melanotic medulloblastoma)
<b>Mucinous and myxoid tumors</b>	Mucin production is a common feature of carcinomas and soft tissue tumors. Soft tissue mucins are referred to as “myxoid material” to distinguish them from biochemically distinct epithelial mucin. Mucin production is vanishingly rare in melanoma and lymphoma. Colloid CA refers to tumors composed of mainly mucin with only few scattered tumor cells (usually as rows lining colloid at the periphery and floating in mucin as small, inconspicuous clusters). <b>Common sites of mucinous CA:</b> the bowel, appendix, ovary, pancreas, lung, breast. Colloid CAs generally considered clinically indolent (except the colon and ovary, mixed data on the lung) <b>DDx of myxoid soft tissue and bone tumors:</b> almost ANY soft tissue tumor can be myxoid, at least focally. Major players are intramuscular myxoma, myxofibrosarcoma, myxoid liposarcoma, low-grade fibromyxoid sarcoma, extraskeletal myxoid chondrosarcoma, neurofibroma (myxoid change common), spindle cell lipoma, nodular fasciitis and abdominal fibromatosis (sometimes myxoid), chondromyxoid fibroma, chordoma <b>Other tumors that may have myxoid change:</b> malignant mesothelioma, pleomorphic adenoma, myxopapillary ependymoma (filum terminale)
<b>Tumors with squamoid morules (have nuclear <math>\beta</math>-catenin staining)</b>	Endometrioid CA (low grade), craniopharyngioma (adamantinomatous type), cribriform-morular variant of papillary thyroid CA, pancreatoblastoma, pulmonary blastoma/well-differentiated fetal adenoCA of the lung, basal cell adenoma/adenoCA of salivary glands

Benign Mimics of Malignancy 101 – Watch Out!	
<b>Pleomorphic tumors that are actually NOT high grade (degenerative-type atypia)</b>	Classically, endocrine and NE neoplasms (“NE atypia”), schwannomas (“ancient change”), renal oncocytoma, pleomorphic xantroastrocytoma (PXA), atypical fibroxanthoma (AFX), uterine leiomyomas (“symplastic” change), pleomorphic hyalinizing angiectatic tumor (PHAT). A clue to degenerative nature of atypia is smudgy chromatin and absence of atypical mitoses.
<b>Benign proliferations which may have perineural invasion</b>	Breast sclerosing adenosis, endometriosis, vasitis nodosa, Leydig cell tumors and normal Leydig cells, prostate “benign perineural involvement” (tumor apposed but not surrounding a nerve), pyloric gland metaplasia in the gallbladder, pancreatic islet cells, granular cell tumor
<b>Benign tumors which may have isolated vascular invasion</b>	Pleomorphic adenoma [1, 2], pheochromocytoma/paraganglioma, granular cell tumor [3], giant cell tumor of bone, giant cell tumor of tendon sheath, renal oncocytoma [4]
<b>Benign tumors which may invade the bone (bone invasion <math>\neq</math> malignancy)</b>	Meningioma, pituitary adenoma, inverted Schneiderian papilloma (extension into the bone occurs as a result of pressure erosion and by itself is not an indication of malignancy)
<b>Benign inclusions in lymph nodes</b>	Müllerian (endometriosis, endosalpingiosis, endocervicosis), nevus (intracapsular location), salivary gland, thyroid (somewhat controversial), mesothelial, breast (heterotopic tissue or benign mechanical transport due to procedure or massage; usually from papillary lesions), renal tubules in patients with large Wilms tumor, lymphangioliomyomatosis
<b>Benign tumors that can metastasize (!)</b>	Pleomorphic adenoma, uterine benign metastasizing leiomyoma, ameloblastoma, benign fibrous histiocytoma, chondroblastoma, giant cell tumor of the bone, meningioma, pulmonary sclerosing pneumocytoma (formerly sclerosing hemangioma) [5]

Differentials 101: Small Round Blue Cell Tumors (SRBCT)					
Diagnosis	Age/clinical	Location	Histologic clues	Key immunostains	Cytogenetics
<b>SRBCTs of Adults</b>					
<b>Lymphoma</b>	Any age (type-dependent)	Lymph nodes and any extra-nodal site	No molding (cells are discohesive)	CD45+, CD20 (B cell) or CD3 (T cell), other	Various translocations
<b>Small cell NE carcinoma</b>	Older adults, ectopic hormones, early mets	Any organ	Molding, no nucleoli, "salt and pepper" chromatin, prominent necrosis NE architecture: rosettes, trabeculae (usually subtle)	CK+, NE markers+, TTF-1+ (lung and some non-lung), Neurofilament-, CK20- (opposite to MCC)	
<b>Merkel cell carcinoma (MCC)</b>	60–70 yo	Dermis Head and extremities	Molding, "dusty" vesicular chromatin Rosettes and trabeculae (occasionally)	CK+, NE markers+, always TTF-1-, neurofilament+, CK20+ (punctate), Merkel cell polyomavirus antigen+	
<b>SRBCTs of Young Adults (and Some Children)</b>					
<b>Desmoplastic small round cell tumor (DSRCT)</b>	Mean age 21, M:F = 4:1; rare tumor	Serosal cavities (peritoneum, pleura)	Angulated nests of SRBCs in desmoplastic stroma	WT1+ (C-terminus antibody), CK+, EMA+, NSE+, desmin+/actin-	t(11;22) <i>EWSR1-WT1</i>
<b>Synovial sarcoma, poorly differentiated</b>	Any age, typically young adults; mean age 26; 20% <20 yo	Most commonly extremities but can occur almost in any site	High-grade SRBCT, distinction from Ewing sarcoma or other round cell sarcoma usually requires cytogenetics or molecular study	TLE1+, CK/EMA-/+ (patchy at best), poorly-differentiated cases usually CD99+	t(X;18) <i>SS18-SSX1/2</i> A small subset can be <i>SS18</i> FISH negative but would be positive by other methods such as RT-PCR
<b>Olfactory neuroblastoma (esthesioneuroblastoma)</b>	Bimodal peaks: ages 15 and 55	Roof of nasal fossa (cribriform plate)	Similar to abdominal neuroblastoma: fibrillar rosettes and fibrillar stroma (neuropil), ganglion cells generally absent	NE markers+ (SYN most sensitive), CK can be focal but EMA always -, sustentacular cells S100+, calretinin+	
<b>Small cell osteosarcoma</b>	Bimodal age peaks: 20s and 50s	Around knee (distal femur, proximal tibia)	Tumor osteoid required for diagnosis	SATB2+	
<b>Mesenchymal chondrosarcoma</b>	Typical range 10–40yo (peak 20s and 30s)	Axial skeleton, also in soft tissue	Difficult to recognize if chondroid area is not present	SOX9+ (non-specific), S100+ in chondrocytes	<i>HEY1-NCOA2</i>
<b>High-grade myxoid (round cell) liposarcoma</b>	Peak incidence in the 30s	Deep soft tissue of extremities (thigh)	Subtle chicken-wire vessels; variable number of lipoblasts (not required for diagnosis but helpful in recognition)	S100+ in adipocytes (immunostains not helpful for the diagnosis; diagnosis requires cytogenetic or molecular tests)	t(12;16) or t(12;22) <i>FUS</i> (or <i>EWSR1-DDIT3</i> )

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SRBCTs of Children (and Some Adults) – 2 <sup>1</sup>					
Diagnosis	Age/clinical	Location	Histologic clues	Key immunostains	Cytogenetics
<b>Lymphoblastic lymphoma (LBL)</b> (>80% are T cells)	Peaks in adolescence, rare in adults Boys>>girls #1 pediatric malignancy (together with leukemia)	Thymus (>50%), nodes, spleen, and others	Dense medium-size lymphocytes, blastic (“fine lacey”) chromatin, inconspicuous nucleoli No molding (cells are discohesive) Many mitoses, sometimes “starry sky” pattern (similar to Burkitt lymphoma)	CD45 variable, TdT+, CD34+ CD3+ (if T cell) frequently CD99+	Various translocations
<b>Myeloid sarcoma (leukemic infiltrate outside of marrow, aka chloroma)</b>	Wide age range, may be de novo or have history or concurrent leukemia or myeloid disorders	Skin, lymph nodes, and bone but many more organs can be involved	Diffuse, monotonous mononuclear cells that often infiltrate background structures; cells have blastic chromatin with variable nucleoli and scant cytoplasm	CD43+, myeloperoxidase+, lysozyme+, can be CD99+ (pitfall)	Various translocations
<b>Neuroblastoma</b>	Peak age 2 yrs, 90% by age 8, rare in young adults #1 solid extracranial malignancy and #3 overall malignancy (after leukemia/lymphoma and CNS) in kids	Adrenal medulla, sympathetic ganglia	Fibrillar stroma (neuropil) and fibrillar (Homer Wright) rosettes Ganglion cells and Schwannian stroma in better differentiated tumors No molding (cells are evenly spaced apart)	NE markers+, PHOX2B (new marker)	Poor prognosis: N-myc amplification, -1p, +17q Good prognosis: age < 1 year, hyperdiploidy
<b>Ewing sarcoma (ES)/PNET (primitive neuroectodermal tumor)<sup>2</sup></b>	Mean age 11–15 yo, but can occur at any age; rare in ages <5 and > 30 Presents as rapidly growing painful mass. Skeletal form clinically mimics osteomyelitis.	(1) Skeletal: lower extremities and pelvis (2) Soft tissue: paravertebral, extremities, retroperitoneum	Monomorphic uniform cells Vesicular (open) chromatin +/- Homer Wright rosettes Cytoplasmic vacuoles (glycogen/PAS+) No neuropil outside rosettes and no ganglion cells (unlike neuroblastoma)	CD99+, NKX2.2+, NE markers +/-, some may be + for ERG (do not confuse for vascular lesion)	t(11;22) EWSR1-FLI1 – 90% t(21;22) EWSR1-ERG 5% Many more rare fusions
<b>CIC-rearranged sarcoma</b>	Mean age, 24; range 6–62	Trunk and extremities, viscera, rarely the bone	Cytology slightly more atypical than ES. Most have geographic necrosis; some have myxoid changes or spindling of cells	ETV4+ (negative in other SRBCTs), WT1+, variable CD99 (diffuse in only 20% of cases in contrast to ES)	t(4;19) or t(10;19) CIC-DUX4
<b>BCOR-rearranged sarcoma</b>	Mean age 15; range 2–44 M>>F Reference: [6]	Bone>>soft tissue>viscera	Round and spindle cells. Resembles poorly different synovial sarcoma	BCOR+, CCNB3+, SATB2+, Cyclin D1+, TLE1+	inv(X) BCOR-CCNB3 t(X;4) BCOR-MAML3
<b>Alveolar rhabdomyosarcoma, solid variant</b>	Peak age 9, can occur up to age 30 (older than embryonal) #1 pediatric sarcoma	Deep muscles of extremities; trunk (distinct from embryonal)	Look for hints of myogenic differentiation: pink cytoplasmic inclusions (cross-striations are rarely evident) and multinucleated wreath-like giant cells Dense chromatin (unlike ES) Cells are discohesive	Desmin+ (can highlight cross-striations), MyoD+, myogenin+ (usually diffuse)	t(2;13) PAX3-FOXO1 t(1;13) PAX7-FOXO1
<b>Wilms tumor (nephroblastoma), blastema predominant</b>	Peak age, 3.5 yo; range 3 mo–6 yrs.; always >3 mo and <16 yrs. of age #1 pediatric renal tumor	Kidney	May see areas with classic triphasic histology Molding present (unlike lymphoma, neuroblastoma) [7]	WT1+	11p13 (WT1 gene) deletion/mutation, Trisomy 12
<b>Medulloblastoma</b>	Peak age 7 yo; usually <20 yo (70% under age 16)	Cerebellum	High-grade SRBCT Homer Wright rosettes Sometimes nodular architecture	SYN+	Isochromosome 17q
<b>Retinoblastoma</b>	Young children	Retina	Flexner-Wintersteiner rosettes	CRX/OTX3 (new marker)	13q14 (RB gene) deletion/mutation
<b>Hepatoblastoma, small-cell variant</b>	90% in kids under age 5	Liver	Diagnosis requires areas of better-differentiated hepatoblastoma		

1. Ddx also includes small cell osteosarcoma and mesenchymal chondrosarcoma (see SRBCTs of Young Adults). Note that not all “blastomas” are pediatric small round cell tumors: for example, pulmonary blastoma and hemangioblastoma are tumors of adulthood that are non-SRBCT.

2. Ewing sarcoma (ES) and PNET (peripheral primitive neuroectodermal tumor) are now regarded as morphological manifestations of one tumor type; both are characterized by t(11;22) translocation. In general, there are usually more neuroendocrine features in PNET, whereas ES is thought to be a more undifferentiated tumor. However, there is a considerable overlap in clinical presentation, morphology, and prognosis, and most pathologists no longer separate them. In fact, the term PNET was retired in the most recent soft tissue WHO.


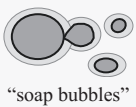





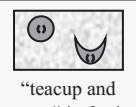
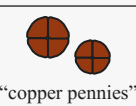
General comment: While not entirely specific (like most stains or morphologic features), many of the soft tissue sarcomas that are associated with specific molecular abnormalities have a distinct cytologic monotony that can serve as a subtle tip. Rare exceptions do occur: some translocation-associated tumors can be pleomorphic (such as myxoinflammatory fibroblastic sarcoma); rarely synovial sarcoma or Ewing sarcoma may show more pleomorphism if they harbor TP53 mutations.

Abbreviations: SRBCT small round blue cell tumor


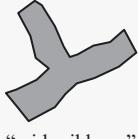

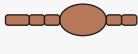
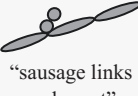
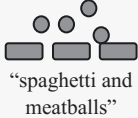


## There's a Fungus Among Us! Quick Reference for Histologic Identification of Fungi

Fungi are encountered in two major settings: (1) surface dwellers and colonizers (e.g., mucocutaneous candidiasis and fungus ball, respectively) and (2) infections involving visceral organs/soft tissue. The latter category includes two types of fungi: (1) **opportunistic** (occurring in immunocompromised host) vs. **pathogenic** (able to infect immunocompetent host). Classic opportunistic fungi include *Pneumocystis* and *Zygomycetes*. Classic pathogenic fungi include dimorphic fungi, which exist as molds in nature and yeast in tissue: *Histoplasma* (*Histo*), *Blastomyces* (*Blasto*), *Coccidioides* (*Cocci*), and *Paracoccidioides*. *Cryptococcus* (*Crypto*) is predominantly opportunistic. When you encounter a fungus in visceral organs, your DDX should vary according to the patient's immune status.

Fungi are generally inconspicuous in H&E sections and are best visualized by "pan-fungal" stains – GMS and PAS – although some larger fungi (*Blasto*, *Cocci*, *Zygomycetes*) are readily visible in H&E. *Crypto* can also be at least suspected on H&E. The sizes of a RBC and a lymphocyte nucleus are ~7 µm; these may be used as a handy-size reference. The most common histologic response to fungi is granulomatous inflammation, but some may manifest with other features, such as granulomas with neutrophils (*Blasto*), granulomas with eosinophils (*Cocci*), or frothy intra-alveolar exudate (*Pneumocystis*).

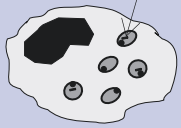
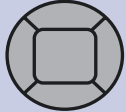
Organism or disease	Appearance (GMS or PAS)	Organism – key features	Typical tissue reaction	Comment
<b>Budding Yeast in Tissue</b>				
<i>Histoplasma capsulatum</i>	 "tiny critters in a macrophage (MF) and in tissue"	<ul style="list-style-type: none"> <li>– 2–5 µm</li> <li>– Narrow-based "teardrop" budding (difficult to see in tissue)</li> <li>– Pseudocapsule (faint halo due to retraction artifact) on Giemsa – no true capsule</li> <li>– Mostly oval shapes</li> <li>– Predominantly intracellular but usually spill into surrounding tissue, where organisms tend to remain in clusters</li> <li>– DDX: <i>Crypto</i>, <i>Pneumocystis</i>, <i>Candida</i>, <i>Penicillium</i> (rare), and small intracellular protozoa (<i>Leishmania</i>)</li> </ul>	<ul style="list-style-type: none"> <li>– Granulomas with fibrocaseous "infarct-like" necrosis</li> <li>– Old lesions typically hyalinize/calcify</li> </ul>	<ul style="list-style-type: none"> <li>– Ohio-Mississippi river valley</li> <li>– Carrier: birds and bats ("cave fever")</li> <li>– Sites: lung, GI, disseminated</li> <li>– Organisms can stain extremely pale with GMS – look closely!</li> </ul>
<i>Cryptococcus neoformans</i>	 "soap bubbles"	<ul style="list-style-type: none"> <li>– 2–15 µm</li> <li>– Narrow-based budding</li> <li>– Highly variable size (unlike <i>Histo</i> or <i>Blasto</i>, which are uniform)</li> <li>– Variable shape: spherical and elongated (football-shaped) forms</li> <li>– Polysaccharide capsule (mucicarmine+, PAS+, alcian blue+), but some organisms are capsule-deficient. India ink + (historic use only)</li> <li>– Cell wall contains melanin pigment (Fontana-Masson+; pigment not apparent on H&amp;E). Note that positive melanin stain is not entirely specific for <i>Crypto</i> since <i>Cocci</i>, <i>Blasto</i>, and <i>Sporothrix</i> can also be positive</li> </ul>	<ul style="list-style-type: none"> <li>– Granulomatous inflammation (+/- necrosis)</li> <li>– Histiocytes with bubbly cytoplasm (where organisms are usually visible by H&amp;E)</li> </ul>	<ul style="list-style-type: none"> <li>– Carrier: pigeons (droppings)</li> <li>– Sites: meningitis, the lung, other deep infections</li> </ul>
<i>Blastomyces dermatitidis</i>	 "snowman"	<ul style="list-style-type: none"> <li>– 8–15 µm</li> <li>– Broad-based budding</li> <li>– Thick double walls ("double contour"), multinucleation</li> <li>– Cell walls can be weakly positive for mucin stains</li> </ul>	<ul style="list-style-type: none"> <li>– Granulomas with neutrophils</li> </ul>	<ul style="list-style-type: none"> <li>– Ohio-Mississippi river valley</li> <li>– Sites: lung, skin, bone, disseminated</li> </ul>
<i>Paracoccidioides brasiliensis</i> (aka South American blastomycosis)	 "mariner's wheel"	<ul style="list-style-type: none"> <li>– 5–30 µm (wide size variation is characteristic)</li> <li>– Large spherule with multiple peripheral narrow-based buds (although diagnostic, the multiple budding cells are usually inconspicuous)</li> </ul>		<ul style="list-style-type: none"> <li>– Africa, Central and South America</li> <li>– Sites: skin, bone, mucous membranes (mimics <i>Blasto</i>)</li> </ul>
<i>Sporothrix schenckii</i>	 "cigar bodies"	<ul style="list-style-type: none"> <li>– 2–6 µm</li> <li>– Round or elongated "cigar-shaped" budding yeast, usually rare and difficult to find in tissue</li> <li>– "Asteroid bodies" (Splendore-Hoeppli phenomenon) – crystalline structures representing antigen-antibody complexes. Classic for <i>Sporo</i> but not specific</li> </ul>		<ul style="list-style-type: none"> <li>– "Rose-gardener's disease"</li> <li>– Sites: SubQ</li> </ul>
<b>Non-budding Spherical Fungi in Tissue</b>				
<i>Coccidioides immitis</i>	 "bag of marbles"	<ul style="list-style-type: none"> <li>– Thick-walled spherule (50–200 µm) packed with endospores (2–5 µm)</li> <li>– Endospores frequently spill into the surrounding tissue and may resemble <i>Histoplasma</i> (but there is no budding)</li> <li>– <b>DDX:</b> <ul style="list-style-type: none"> <li>• <i>Rhinosporidium</i>: nasal fungus, much larger than <i>Cocci</i>. GMS+</li> <li>• Myospherulosis: surgical packing material with entrapped RBCs in the nose/sinus. GMS-, PAS-, hemoglobin+</li> </ul> </li> </ul>	<ul style="list-style-type: none"> <li>– Granulomas with eosinophils</li> </ul>	<ul style="list-style-type: none"> <li>– Southwest American deserts ("valley fever")</li> <li>– Sites: lung, skin, disseminated</li> </ul>
<i>Penicillium marneffei</i>	 "tiny critters in a MF"	<ul style="list-style-type: none"> <li>– 2–4 µm</li> <li>– Elongated cells with septae (divides by fission, not budding)</li> <li>– predominantly intracellular (like <i>Histo</i>)</li> <li>– Mimics <i>Histo</i> (but <i>Penicillium</i> is non-budding and has no pseudocapsule)</li> </ul>		<ul style="list-style-type: none"> <li>– Southeast Asia</li> <li>– AIDS patients</li> </ul>
<i>Pneumocystis jiroveci</i> (formerly <i>P. carinii</i> )	 "teacup and saucer" in frothy exudate	<ul style="list-style-type: none"> <li>– 5–8 µm cyst (seen by GMS), 1–3 µm trophozoites (seen by Giemsa)</li> <li>– Non-budding organisms</li> <li>– GMS: round and crescent (sickle)-shaped cysts (described as a "cup-and-saucer" or "crushed ping-pong balls") with two parenthesis-shaped dots (these are part of the cyst wall)</li> <li>– Giemsa or Diff-Quik: intracyclic (up to eight) and free-roaming trophozoites</li> </ul>	<ul style="list-style-type: none"> <li>– Frothy alveolar exudate (but ~10% have a granulomatous response)</li> </ul>	<ul style="list-style-type: none"> <li>– Sites: lung</li> <li>– AIDS patients</li> </ul>
<i>Chromoblastomycosis</i>	 "copper pennies"	<ul style="list-style-type: none"> <li>– 6–12 µm</li> <li>– Brown (melanin-containing) organisms; Fontana-Masson+</li> <li>– Thick-walled spheres with horizontal and vertical septae ("copper pennies," "medlar bodies," "sclerotic bodies")</li> </ul>	<ul style="list-style-type: none"> <li>– Overlying pseudoepitheliomatous hyperplasia is typical</li> </ul>	<ul style="list-style-type: none"> <li>– Sites: SubQ</li> </ul>

## There's a Fungus Among Us! Quick Reference for Histologic Identification of Fungi – 2

Organism or disease	Appearance (GMS or PAS)	Key histologic features	Comment
<b>Hyphae in Tissue</b>			
<p><i>Aspergillus</i> spp. and others <b>Hyalohyphomycoses</b> (septate nonpigmented molds, e.g., <i>Fusarium</i>)</p>	 <p>“slingshots “</p>	<ul style="list-style-type: none"> <li>– Thin (2–5-µm-thick) hyphae WITH septae (“septate hyphae”)</li> <li>– Frequent dichotomous narrow-angle (45°) branching (Y-shaped)</li> <li>– When invasive, tends to be angioinvasive</li> <li>– Hyphae tend to grow in a radial “sunburst”-like fashion</li> <li>– Occasional fruiting bodies (in aerated sites)</li> <li>– Definitive diagnosis requires cultures because in tissue, <i>Aspergillus</i> is indistinguishable from other hyaline mold, including <i>Pseudallescheria boydii</i> and <i>Fusarium</i> (both resistant to amphotericin B). Definitive morphologic speciation possible only if diagnostic fruiting bodies are present, which is rare.</li> </ul>	<p>Types of <i>Aspergillus</i>-related diseases include:</p> <ol style="list-style-type: none"> <li>1. Vaso-invasive infections (sinus, lung, disseminated) in immunocompromised host.</li> <li>2. Allergic bronchopulmonary aspergillosis and allergic fungal sinusitis in atopic host (eos + tigroid mucin+Charcot Leyden crystals)</li> <li>3. Aspergilloma/mycetoma/fungus ball = colonization of cavities (such as sinuses or cavitary lung disease)</li> <li>4. Less well-defined form is “subacute” = limited tissue invasive infection in mildly immunocompromised host (like diabetic), occurs in the sinus and lung.</li> </ol>
<p><b>Zygomycetes</b> (<i>Rhizopus</i>, <i>Absidia</i>, <i>Mucor</i>) Disease: Zygomycosis = Mucormycosis = Phycomycosis</p>	 <p>“wide ribbons”</p>	<ul style="list-style-type: none"> <li>– Wide (6–50-µm-thick) hyphae with INFREQUENT or absent septae</li> <li>– Wide-angle (90°) branching (branching less frequent than <i>Aspergillus</i>)</li> <li>– Undulating, twisting (ribbonlike), often fractured, “empty-looking” hyphae</li> <li>– Angioinvasive</li> <li>– Stain weakly with GMS and PAS; organisms best visualized by H&amp;E</li> <li>– Definitive diagnosis requires culture because treated or degenerating <i>Aspergillus</i> may look like <i>Zygomycetes</i></li> </ul> <p>*Note regarding the terminology: these organisms are frequently referred to collectively as “Mucor” in pathology, but <i>Mucor</i> is only one of several organisms (and not even the most common) in this group.</p>	<p>Aggressive vaso-invasive disease (sinus, disseminated) in immunocompromised host. This is a life-threatening emergency.</p> <p>Why distinguishing <i>Zygomycetes</i> vs. <i>Aspergillus</i> is important:</p> <ol style="list-style-type: none"> <li>1. <i>Zygomycetes</i> are more aggressive</li> <li>2. <i>Zygomycetes</i> are treated with amphotericin B. They are resistant to most azoles (except posaconazole).</li> </ol>
<p><b>Dermatophytes</b> (<i>Microsporum</i> spp., <i>Epidermophyton</i> spp., <i>Trichophyton</i> spp.)</p>		<ul style="list-style-type: none"> <li>– Septate hyphae with rare branching that break into segments (arthroconidia)</li> <li>– 2–3 µm thick</li> <li>– Hyphae confined to the skin, nails, hair</li> </ul>	<p>Superficial infections of the and hair (“tinea” or “ringworm”)</p>
<p><b>Phaeohyphomycosis</b> (pigmented molds)</p>		<ul style="list-style-type: none"> <li>– Septate branching hyphae; may resemble <i>Aspergillus</i> in tissue though are often thinner with less branching, have constrictions at their frequent septae and vesicular swellings</li> <li>– Contain melanin (Fontana-Masson+)</li> <li>– Brown pigment sometimes (but not always) evident on H&amp;E</li> </ul>	<p>SubQ and deep infections</p>
<b>Yeast and Hyphae in Tissue</b>			
<p><i>Candida</i> spp.</p>	 <p>“sausage links and yeast”</p>	<ul style="list-style-type: none"> <li>– 3–5 µm budding yeast</li> <li>– 5–10 µm pseudohyphae: elongated budding yeast joined end-to-end like “sausage links”; occasionally true hyphae (no constrictions) are present</li> <li>– <i>C. glabrata</i> is unique in that it does not produce any hyphae; it may mimic <i>Histo</i> and other small yeast</li> </ul>	<p>Mucocutaneous and deep infections</p>
<p><b>Pityriasis versicolor</b> (<i>Malassezia furfur</i>)</p>	 <p>“spaghetti and meatballs”</p>	<ul style="list-style-type: none"> <li>– 3–8 µm budding yeast (meatballs) and 5–10 µm fragmented hyphae (spaghetti) often arranged end-to-end</li> <li>– Involves epidermis only, only rarely seen in tissue (skin scraping preferred method of diagnosis)</li> </ul>	<p>Site: skin only</p>
<b>Mold-Like Branching Filamentous Bacteria</b>			
<p><i>Nocardia asteroides</i></p>		<ul style="list-style-type: none"> <li>– Delicate narrow (1 µm) beaded filaments; right-angle branching</li> <li>– Gram+, modified AFB (Fite)+, GMS+</li> <li>– DDX includes <i>Streptomyces</i> (AFB-)</li> </ul>	<p>Deep infection in immunocompromised host</p>
<p><i>Actinomyces israelii</i></p>	 <p>“dust bunnies”</p>	<ul style="list-style-type: none"> <li>– Delicate narrow (&lt;1 µm) branching filaments intertwined in a dense radiating meshwork</li> <li>– “Sulfur granules” (grossly yellow flecks; do not, in fact, contain sulfur)</li> <li>– Gram+, AFB-, GMS+</li> </ul>	<p>Normal commensal inhabitant of the oral cavity</p> <p>May become pathogenic in oropharynx with local tissue damage (such as dental work), may cause draining sinus tracts IUD-related infections</p>



## There's a Fungus Among Us! Quick Reference for Histologic Identification of Fungi – 3

Yeastlike Organisms in Tissue			
	Organism	Key histologic features	Comment
Protozoa	<b><i>Leishmania</i> spp</b> 	<ul style="list-style-type: none"> <li>– 2–4 <math>\mu\text{m}</math> round to oval aflagellate amastigotes (extravascular form of organisms)</li> <li>– Amastigotes are intracellular</li> <li>– Transverse paranuclear bar-like kinetoplast</li> <li>– <i>Leishmania</i> is a close mimic of <i>Histoplasma</i> (look for kinetoplast)</li> <li>– Organisms stain lightly in H&amp;E</li> <li>– GMS–, PAS–, Giemsa+</li> </ul>	<b>Visceral leishmaniasis (kala-azar):</b> <ul style="list-style-type: none"> <li>– Middle East, Africa, India</li> <li>– Sites: reticuloendothelial system (liver, spleen, bone marrow)</li> </ul> <b>Cutaneous leishmaniasis:</b> Old World (“oriental sore”) and New World (“chiclé ulcer”) <b>Mucocutaneous leishmaniasis:</b> Central and South America
	<b><i>Trypanosoma cruzi</i></b>	<ul style="list-style-type: none"> <li>– Organisms in tissue look identical to <i>Leishmania</i> spp.</li> <li>– <i>T. gambiense</i> and <i>T. rhodesiense</i> (African trypanosomiasis) are confined to blood and do not invade tissue</li> </ul>	<b><i>T. cruzi</i> (Chagas disease):</b> <ul style="list-style-type: none"> <li>– Central and South America</li> <li>– Usual sites: heart, colon, esophagus</li> </ul>
	<b><i>Toxoplasma gondii</i></b>	<ul style="list-style-type: none"> <li>– 5–7 <math>\mu\text{m}</math> crescent-shaped tachyzoites (non-encysted organisms in tissue)</li> <li>– 10–50 <math>\mu\text{m}</math> pseudocysts packed with 2–3 <math>\mu\text{m}</math> round bradyzoites</li> <li>– Basophilic in H&amp;E (unlike yeast)</li> <li>– GMS+, PAS+, Giemsa+</li> </ul>	<ul style="list-style-type: none"> <li>– Worldwide disease, cat vector</li> <li>– Sites: disseminated disease (especially brain) in immunosuppressed patients</li> </ul>
	<b><i>Cryptosporidium</i></b>	<ul style="list-style-type: none"> <li>– 2–6 <math>\mu\text{m}</math> round organisms in the brush border of small bowel mucosa</li> <li>– Giemsa+</li> </ul>	Chronic diarrhea in immunosuppressed patients
	<b><i>Cyclospora</i></b>	<ul style="list-style-type: none"> <li>– 8–10 <math>\mu\text{m}</math> oocysts in stool</li> <li>– Modified acid fast or safranin stain (stool)+</li> <li>– Autofluorescence+</li> </ul>	
	<b><i>Cystoisospora belli</i></b>	<ul style="list-style-type: none"> <li>– 25–30 <math>\mu\text{m}</math> elliptical organisms interposed between adjacent enterocytes</li> <li>– Giemsa+</li> </ul>	
	<b><i>Microsporidium</i></b>	<ul style="list-style-type: none"> <li>– 1–3 <math>\mu\text{m}</math> round organisms in the cytoplasm of enterocytes</li> <li>– Invisible by H&amp;E</li> <li>– Gram+</li> </ul>	
Algae	<b><i>Prototheca</i> spp.</b> 	<ul style="list-style-type: none"> <li>– 2–12 <math>\mu\text{m}</math></li> <li>– Sporulating forms are sporangia with up to 20 polygonal or wedge-shaped endospores whose cell walls mold together (“morulas”)</li> <li>– GMS+, PAS+</li> </ul>	<ul style="list-style-type: none"> <li>– Two human infections: cutaneous (usually immunosuppressed) and olecranon bursitis (usually otherwise healthy with a history of trauma)</li> </ul>

References: [8–10]

## Quick Reference for Histological Identification of Viruses


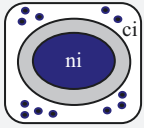
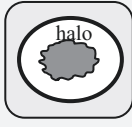

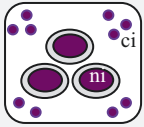
In general, nuclear inclusions are associated with DNA viruses (HSV, CMV, adenovirus, JC, and BK viruses). One major exception is CMV in that in addition to nuclear inclusions, it also forms cytoplasmic inclusions. Some DNA viruses do not have any recognizable cytopathic effects (EBV, HHV8). Note that HPV does not manifest as inclusions but has a unique cytopathic effect (see below).

RNA viruses as a rule do not have recognizable cytopathic changes; few do have cytoplasmic inclusions (RSV, Negri bodies in rabies). Measles is an exception in that it is an RNA virus that forms nuclear inclusions.

Nuclear inclusions of virally infected cells fall into two morphologic categories:

1. **Cowdry type A:** eosinophilic “owl-eye” nuclear inclusion (as in CMV)
2. **Cowdry type B:** (aka “smudge cells”) nucleus with a “homogenized, ground glass” chromatin and obliterated nuclear detail

Note that most DNA viruses (HSV, adenovirus) can have nuclear inclusion of either Cowdry A and/or Cowdry B type even within the same lesion. Exception is CMV in that it forms exclusively type A inclusions.

Virus	Appearance	Nuclear inclusions	Cytoplasmic inclusions	Specific features	Infected cell type	Clinical
<b>HSV</b>	 “eggs in a basket” or “pomegranate seeds”	+ (Cowdry A or B), pink, steel gray, or purple	–	The 3 M’s: <b>M</b> ultinucleation, <b>M</b> olding, <b>M</b> argination of chromatin (peripheral clearing or “halo effect”)	Squamous and some glandular epithelial cells (look at the periphery of an ulcer on mucosal surfaces)	Gingivostomatitis and genital lesions in immunocompetent host. Opportunistic infection of any body site (pneumonia, esophagitis, neurons in encephalitis)
<b>CMV</b>	 “owl-eye nuclear inclusion and cytoplasmic speckles”	+ (Cowdry A), blue	+ (blue speckles)	Nuclear and cytoplasmic enlargement Nuclear inclusion has a prominent halo (“owl eye”), which corresponds to marginated chromatin pushed aside by viral particles.	Stromal and endothelial cells (look at the ulcer base); rarely in epithelial cells	Opportunistic infection of any body site (lung, bowel, retina, neurons in encephalitis)
<b>Adenovirus</b>		+ (Cowdry A or B), blue	–		Epithelial cells (bronchial cells and pneumocytes in the lung)	Opportunistic infections (bladder, kidney, lung, bowel)
<b>HPV</b>	 “koilocyte”	–	–	Clear perinuclear vacuole (Greek <i>koilos</i> = hollow), wrinkled (raisin-like) nucleus, binucleation (common), condensed keratohyaline granules typical in skin	Squamous cells	Papillary lesions (warts, condyloma, laryngeal papillomas) and squamous dysplasia and intraepithelial neoplasia (genital organs and rectum)
<b>JC and BK (polyoma)</b>	 “decoy cell”	+ (Cowdry B), blue	–	Nuclear enlargement Non-haloed smudgy (type B) inclusion	JC – brain BK – urothelium (mimics CIS in urine; “decoy” cells)	JC – PML BK – cystitis in immunosuppressed
<b>Measles</b>	 “Warthin-Finkeldey giant cell”	+ (Cowdry A or B), pink	+ (pink speckles)	Giant cells with multinucleation	Depends on the site: -Lung: epithelial cells, most commonly bronchial -Lymph node: lymphoreticular cells (infected cells in lymph node are called Warthin-Finkeldey giant cells) -Brain: oligodendroglia	Pneumonitis, lymphadenitis, SSPE
<b>RSV</b>		–	+ (large pale-pink globs)	Giant cells with multinucleation No nuclear inclusion (this is an RNA virus)	Epithelial cells	Bronchiolitis and pneumonia in children (rarely biopsied)

Abbreviations: *ni* nuclear inclusion, *ci* cytoplasmic inclusion, *PML* progressive multifocal leukoencephalopathy, *SSPE* subacute sclerosing panencephalitis

## Tumor Viruses: Quick Reference for Tumor/Viral Associations

Detection of viral molecules is a very helpful adjunct in the diagnosis of the virally induced tumors. In tissue sections, viral proteins can be detected by immunohistochemistry (e.g., EBV-LMP), or viral nucleic acids may be identified by in situ hybridization/ISH (e.g., EBER ISH and HPV by DNA-ISH and more recently RNA-ISH, which is much more sensitive). p16 is NOT a viral protein but an endogenous cell cycle protein that is markedly overexpressed as a result of high-risk HPV infection. In cervical cytology, the specimens are tested for HPV by a DNA-based method (hybrid capture).

Virus	Tumor associations	Detection in tissue	
<b>EBV</b>	<p><b>Epithelial lesions:</b></p> <ul style="list-style-type: none"> <li>– Nasopharyngeal carcinoma (NPC), aka lymphoepithelial carcinoma (= lymphoepithelioma [LE]) and LE-like carcinomas:               <ul style="list-style-type: none"> <li>○ EBV (+): NPC and LE-like carcinomas of upper aerodigestive tract (lung, thymus, salivary gland) and stomach</li> <li>○ EBV (–): LE-like carcinoma of non-aerodigestive tract (bladder, breast, skin, cervix)</li> </ul> </li> <li>– Oral hairy leukoplakia</li> <li>– Gastric adenocarcinoma (5%)</li> </ul> <hr/> <p><b>Lymphoid/heme lesions:</b></p> <ul style="list-style-type: none"> <li>– Infectious mononucleosis</li> <li>– Posttransplant lymphoproliferative disease (PTLD)</li> <li>– Classic Hodgkin lymphoma (Mixed cellularity – 70%, AIDS-related)</li> <li>– Non-Hodgkin lymphoma:               <ul style="list-style-type: none"> <li>○ Burkitt lymphoma (endemic 100%; sporadic 20%)</li> <li>○ Nasal-type NK/T-cell lymphoma (&gt;95%)</li> <li>○ Aggressive NK cell leukemia</li> <li>○ Angioimmunoblastic T-cell lymphoma</li> <li>○ Lymphomatoid granulomatosis (&gt;95%)</li> <li>○ CNS lymphoma in AIDS (95%)</li> <li>○ Plasmablastic lymphoma (HIV)</li> <li>○ Primary effusion lymphoma (has both EBV and HHV8)</li> <li>○ EBV+ DLBCL</li> </ul> </li> <li>– Germiotropic lymphoproliferative disorder (has both EBV and HHV8)</li> <li>– Inflammatory pseudotumorlike follicular dendritic cell sarcoma</li> <li>– EBV-positive mucocutaneous ulcer</li> </ul> <hr/> <p><b>Smooth muscle tumors</b> in immunosuppressed (AIDS, transplant)</p>	<ol style="list-style-type: none"> <li>1. EBER (EBV encoded early RNA). Most sensitive marker for EBV (in situ hybridization method). IDs all EBV-related tumors.</li> <li>2. EBV-LMP (late membrane protein). Less sensitive than EBER. IDs PTLN and AIDS-related lymphomas, variable in NPC, Hodgkin, and Burkitt lymphoma, usually negative in plasmablastic lymphoma</li> <li>3. EBNA (EBV nuclear antigen). Least sensitive marker. IDs PTLN and AIDS-related lymphomas only.</li> </ol>	
<b>HPV</b>	<p><b>Female genital tract:</b></p> <ul style="list-style-type: none"> <li>– Squamous dysplasia and carcinoma of the cervix, vagina, vulva (simplex/differentiated VIN and associated SqCC occur in the setting of lichen sclerosus and other dermatoses in older women and are HPV-unrelated)</li> </ul> <hr/> <p>– Cervical adenocarcinoma (in situ and invasive)</p> <hr/> <p><b>Penis:</b></p> <ul style="list-style-type: none"> <li>– Squamous cell carcinoma, warty and basaloid type (verrucous and papillary SqCC are HPV-unrelated)</li> </ul> <hr/> <p>– Bowenoid papulosis and Erythroplasia de Queyrat</p> <hr/> <p><b>Anus:</b> Squamous neoplasia (in situ and invasive) – analogous to cervix</p> <hr/> <p><b>Head and neck:</b></p> <ul style="list-style-type: none"> <li>– Squamous cell carcinoma of the oropharynx (tonsil and base of tongue)</li> </ul> <hr/> <p>– Laryngeal papillomatosis</p> <hr/> <p>– Focal epithelial hyperplasia (Heck disease) of oral mucosa</p> <hr/> <p>– Sinonasal HPV-related multiphenotypic CA</p> <hr/> <p><b>Mucocutaneous:</b></p> <ul style="list-style-type: none"> <li>– Warts (verruca)</li> </ul> <hr/> <p>– Condyloma acuminatum (genital sites)</p>	<p>HSIL and associated SqCC caused by high-risk HPV (16, 18, 31, 33) LSIL is caused by</p> <ul style="list-style-type: none"> <li>– low-risk HPV (6, 11) in 20%</li> <li>– high-risk HPV in 80% (therefore, high-risk HPV does not distinguish HSIL and LSIL)</li> </ul> <p>HPV 18 &gt; 16</p> <p>HPV 16</p> <p>HPV 16</p> <p>HPV 16, 18</p> <p>HPV 16, 18</p> <p>HPV 6, 11</p> <p>HPV 13, 32</p> <p>HPV33 and others (not 16 or 18)</p> <p>HPV 1, 2, 4, 7</p> <p>HPV 6, 11</p>	<ol style="list-style-type: none"> <li>1. In situ hybridization for HPV (DNA or RNA)</li> <li>2. IHC for p16 is a surrogate marker of high-risk HPV</li> </ol> <p>Detection of HPV may be used to identify anogenital or oropharyngeal origin of metastatic SqCC of unknown primary.</p> <p>HPV-related SqCC of some (but not all) sites have basaloid morphology:</p> <ul style="list-style-type: none"> <li>– Sites where HPV-related SqCC are basaloid: oropharynx, penis, vulva</li> <li>– Sites where HPV-related SqCC are either basaloid or conventional: cervix, anus</li> <li>– Sites where basaloid SqCC are unrelated to HPV: breast, lung, non-oropharyngeal head and neck</li> </ul>
<b>HHV8</b>	Kaposi sarcoma, primary effusion lymphoma (also has EBV), germiotropic lymphoproliferative disorder (also has EBV), Castleman disease (multicentric)	HHV8 can be detected by IHC	
<b>HTLV1</b>	Adult T-cell leukemia/lymphoma		
<b>Hepatitis B</b>	Hepatocellular carcinoma (Hep C causes HCC indirectly – virus is not present in tumor cells)	HBsAg, HBeAg – rarely used for tumor Dx	
<b>Merkel cell polyomavirus</b>	Nearly all Merkel cell carcinomas	Viral antigen can be detected in Merkel cell CA by IHC, (–) in small cell carcinoma	

# Quick Electron Microscopy Reference for Tumors and Select Non-tumor Diagnoses

By Marina K Baine

Morphologic  
References

General Cell Types	
<b>Epithelial</b>	Desmosomes
<b>Neuroendocrine</b>	Neurosecretory (dense-core) granules
<b>Fibroblast</b>	Abundant rough endoplasmic reticulum
<b>Muscle</b>	Actin filaments
<b>Skeletal muscle</b>	Ribosome-filament complexes Z-band
<b>Smooth muscle</b>	Dense bodies (subplasmallema) Filaments
<b>Endothelial</b>	Weibel-Palade bodies (elongated, “pear-shaped” storage granules with microtubule-like inclusions appearing striated) Pinocytotic vesicles
General Tumor Types	
<b>Carcinoma</b>	Desmosomes (tight junctions)
<b>Adenocarcinoma</b>	Short luminal (“intestinal type”) microvilli Extra- and intracellular lumina Mucin granules Tight junctions
<b>Squamous cell carcinoma</b>	Well-formed intercellular junctions Tonofilaments
<b>Melanoma</b>	Pre-melanosomes and melanosomes
<b>Mesothelioma</b>	Long and thin microvilli Tight junctions Long desmosomes Perinuclear tonofilament bundles
<b>Lymphoma</b>	Abundant polyribosomes Paucity of organelles Devoid of cell junctions
<b>Sarcoma</b>	Absence of true desmosomes and true lumens (most) (Other features vary depending on subtype)
<b>Leukemia</b>	Lineage dependent
<b>Lymphoid</b>	Scant cytoplasm Free ribosomes and polyribosomes Paucity of cell organelles
<b>Myeloid</b>	Abundant cytoplasm Prominent Golgi and rough endoplasmic reticulum +/- Azurophilic granules
Specific Tumor Types and Tumorlike Lesions with Distinctive EM Findings	
<b>Leydig cell tumor</b>	Reinke crystals
<b>Sertoli cell tumor</b>	Charcot-Bottcher crystals
<b>Alveolar soft part sarcoma</b>	Membrane-bound rhomboid crystals with a lattice pattern with a 10-nm periodicity in fibrils (crystal precursor is Golgi) Numerous electron dense vesicles near Golgi
<b>Granular cell tumor</b>	Pleomorphic secondary lysosomes Basal lamina around cell groups Angulate lysosomes (“Gaucher-like”) in stromal fibrohistiocytic cells
<b>Schwannoma</b>	Luse bodies (long-spaced collagen) Reduplicated basal lamina
<b>Langerhans cell histiocytosis</b>	Birbeck granules (“tennis racket”)
<b>Rosai-Dorfman disease</b>	Emperipolesis of nucleated and nonnucleated blood cells (also seen on light microscopy)
Selected Storage Disorders	
<b>Gaucher disease</b>	Angulated lysosomes
<b>Tay-Sachs disease</b>	Laminated (concentric structure of membranous cytoplasmic bodies)
<b>Niemann-Pick disease</b>	Zebra bodies (aka myelin figures)
<b>Fabry disease</b>	Zebra bodies (aka myelin figures)
Selected Viruses	
<b>VZV/HSV/CMV</b>	Bull’s eye appearance in cytoplasm (indistinguishable by EM)
<b>Adenovirus (and other non-enveloped viruses)</b>	Honeycomb (viral particles arranged in paracrystalline arrays)
<b>Papovavirus</b>	Spaghetti and meatballs (virions are both filamentous and spherical)