

Chapter 5

MicroRNAs as Regulators of Prostate Cancer Metastasis



Divya Bhagirath, Thao Ly Yang, Rajvir Dahiya, and Sharanjot Saini

Abstract Prostate cancer causes significant morbidity in men and metastatic disease is a major cause of cancer related deaths. Prostate metastasis is controlled by various cellular intrinsic and extrinsic factors, which are often under the regulatory control of various metastasis-associated genes. Given the dynamic nature of metastatic cancer cells, the various factors controlling this process are themselves regulated by microRNAs which are small non-coding RNAs. Significant research work has shown differential microRNA expression in primary and metastatic prostate cancer suggesting their importance in prostate pathogenesis. We will review the roles of different microRNAs in controlling the various steps in prostate metastasis.

Keywords Metastasis · Prostate cancer · microRNAs · ECM · EMT

5.1 Introduction

Prostate cancer (PCa) is the second most commonly diagnosed cancer among men in the United States and is the third leading cause of cancer related deaths. Metastatic disease accounts for ~16.5% of deaths from PCa [76]. Despite improvement in early screening methods and development of effective therapies, the rates at which aggressive prostate cancers are diagnosed are showing an increasing trend. Recent data analysis from patients with PCa within the United States has shown an increased rate of metastatic disease particularly in men of age group 55–69 years [21]. The rate of metastasis incidence has significantly increased at 2.74% per year from 2012 for all recorded cases. Moreover, there has been a steady increase in the incidence of metastatic PCa among white men as opposed to other races. Furthermore, these rates are expected to increase at 0.38% per year accounting for almost 42% of

D. Bhagirath · T. L. Yang · R. Dahiya · S. Saini (✉)
Department of Urology, Veterans Affairs Medical Center, San Francisco and University of California San Francisco, California, USA
e-mail: Sharanjot.Saini@ucsf.edu

metastatic PCa cases by 2025 [51], which is an alarming increase for the most prevalent male cancer.

Tumor metastasis is a multistep process that involves dissemination of cancer cells from the primary site, their survival in the circulatory system, extravasation to the metastatic sites and subsequent colonization [14, 38]. In addition to their own genetic susceptibilities, cancer cells disseminated from primary sites depend on several growth regulatory signals such as those from chemokines and cytokines in the metastatic niche in order to survive and proliferate at secondary locations [14]. Given the inherent heterogeneity in primary tumors, one can expect emergence of clones that are fit to survive the adversities encountered during the entire process of metastasis. These tumor cells evolve both genetically and epigenetically to surmount the barriers of survival outside their primary niche [38]. There is activation of genes that facilitate metastatic progression, including those that control the epithelial-to-mesenchymal (EMT) pathways, invasion-migratory pathways and allow proteolysis and degradation of the extracellular matrix (ECM). TWIST1, SNAI1, SNAI2 and ZEB1 are some of the important transcription factors that regulate this important step of EMT activation [62]. These essential initiators of metastasis are often regulated in most tumors including PCa by small specialized RNAs called microRNAs [36]. *ANGPTL4*, *MMP1*, *PTGS2*, *EREG* are some of the genes that allow invasion and survival in the circulation and *IL11*, *IL6*, *PTHRP* are genes that facilitate subsequent colonization [62]. Further, these cells cooperate with the microenvironment and its constituent cell types such as fibroblasts and immune cells so as to attain aggressive phenotypes that entail metastatic progression [38]. MicroRNAs (miRNAs) are small non-coding regulatory RNAs which are often deregulated in tumors. In this book chapter, we will provide an overview on PCa metastasis following which we will review the roles of different miRNAs and their contribution towards different steps in PCa metastasis.

5.2 Prostate Cancer Metastasis

Prostate tumors are clinically defined as either indolent or aggressive. Majority of these tumors are localized and treated according to their stage or Gleason score which is determined primarily by biopsy sampling. Most tumors that are identified as aggressive or advanced at the time of first diagnosis and are often accompanied by micrometastasis at secondary locations. Of the different sites in the body, the skeletal system has especially high propensity to develop metastatic lesions in patients with prostate cancer [14, 30, 60]. The presence of bone metastasis in PCa patients in addition to visceral metastasis is associated with a significantly lower overall survival of 14 months [30]. Thus, signifying the importance of metastatic locations in determining disease prognosis. There are different hypotheses to explain the genetics of metastatic tumors. Primary prostate tumors are heterogeneous and are often comprised of different clonal populations that may give rise to metastasis [70]. Liu *et al* studied the association of primary tumors with their corresponding

metastatic tumors and suggested a monoclonal origin for metastatic tumors. They analyzed the copy number alterations and genome wide nucleotide polymorphisms across tumors derived from different metastatic locations in the same patients and observed a genomically stable pattern among these different tumors [57]. However, a more recent study from Hong *et al.* suggests that metastatic tumors are different from primary tumors and that acquisition of favorable mutation such as those in tumor suppressor *TP53* contributes to metastatic tumor heterogeneity and confers metastatic potential to these cells. These mutations or variation in metastatic tumors may arise in order to provide better survival advantage against therapeutic interventions [41, 70]. Furthermore, they also suggest a cross-metastatic seeding pattern in patients where metastatic tumor subclones themselves seed new tumors in more secondary locations [41].

5.2.1 Factors Regulating PCa Metastasis

Several cell intrinsic and extrinsic factors play an important role in execution of a successful metastatic process. While factors intrinsic to a cell refer to the nature of cell type and its genetic composition, microenvironmental cues correspond to the extrinsic factors controlling metastasis. Tumors cells with extensive genomic instability are considered to be the potential candidates for metastatic initiation [62]. Prostate tumor cells are well known to harbor copy number alterations, chromosomal deletions, DNA rearrangements such as chromoplexy, chromothripsis, gene fusions and certain mutations [2, 3, 85]. Presence of *TMPRSS-ERG* gene fusions is a common event among prostate cancer cells. Fluorescence *in situ* hybridization in primary and corresponding metastatic tumors derived from patients undergoing radical prostatectomies have shown association of *TMPRSS2-ERG* fusion among the tumors suggesting a common origin [35]. More recently, it has been shown that over-expression of *TMPRSS2 ERG* fusions in metastatic PCa cell line PC3M-Luc leads to increased bone metastasis in mice [25]. Epigenetic modifications also play an important role in imparting plasticity to the cell that facilitates initiation of metastatic events. EZH2, a histone lysine methyltransferase enzyme is over-expressed in prostate tumors from advanced stage prostate cancer patients and its expression directly correlates with metastatic progression of the disease [89]. Germline mutations in DNA repair genes such as *BRCA1*, *BRCA2*, *CHEK2*, *ATM*, *RAD51D* and *PALB2* is a significant prognostic factor for development of metastatic prostate cancer [66]. Stankiewicz *et al* recently identified *FBXL4* gene which is located on chromosome 6q to be deleted in metastatic bone tumors as well as the primary tumors suggesting its role as a possible tumor suppressor that is lost during metastatic progression [78]. Deletions/additions in parts of chromosomes is a commonly observed phenomenon in PCa tumors [70]. Chromosomal gain (1q, 3q, 7q, 8q, 17q and Xq) and loss (1q, 6q, 8p, 10q, 13q and 16q) are frequently observed chromosomal alterations in the prostate tumor genome [7]. Homeobox tumor suppressor gene *NKX3.1* that is involved in differentiation of the prostate gland is located at chromosome 8p

and its loss has been linked to disease progression [4, 6, 40]. Many tumor-suppressor miRNA genes are located on these chromosomes. Studies from our lab have shown that chromosome 8p is frequently lost during PCa progression, harbors miRNAs that have tumor-suppressor functions and plays an essential role in regressing EMT transition in tumor cell [8–10].

5.2.2 *Microenvironment and PCa Metastasis*

The extracellular microenvironment is another important determinant in the process of metastatic cancer progression. The extracellular matrix (ECM) is formed of diverse matrix proteins including laminin, fibronectin, vitronectin, collagen, osteopontin and others in which stromal cells such as adipocytes, fibroblasts, immune cells and many tissue-specific cells are embedded. Together with blood and lymphatic vessels, they form a niche necessary for sustenance and proliferation of tumor cells. The stromal compartment itself is a strong prognostic indicator of progressing disease. Gene expression analysis of the stromal and epithelial compartment of tumor tissue has revealed a distinct stroma signature that significantly varies with a patient's Gleason score, thereby serving as an independent indicator of high grade PCa [87].

Expression of many basement membrane and matrix proteins are altered during PCa progression [79]. In addition, PCa cells also demonstrate altered patterns of cell adhesion molecules (CAMs) including cadherins and integrins that allow interaction with the surrounding matrix and facilitate cancer progression. The expression of integrins varies with the metastatic stage that in turn, allows protection and easier passage of tumor cells from the external barriers in the primary site, blood vessels and metastatic locations [79, 84]. Signaling through these surface molecules mediated by their interaction with ECM proteins or endothelial cells control motility, growth and proliferation of the disseminated cancer cell [84]. Proteases such as matrix metalloproteases (MMPs), urokinase plasminogen activator (uPA), and cathepsins are upregulated in the invasive disease, thus allowing for degradation of the basement membrane and dissemination of cancer cells through the matrix [79].

Soluble growth promoting signaling molecules such as growth factors, chemokines and chemo-attractants are other important players in mediating successful colonization at metastatic sites. As mentioned earlier, most PCa patients often develop bone metastasis [30]. Stephan Paget's "seed and soil" hypothesis and James Ewing's circulatory connection theory help explain this preferential metastatic spread of prostate tumor cells to bone [14, 84]. Insulin-like growth factor 1 (IGF-1), transforming Growth Factor- β (TGF- β), uPA, bone morphogenetic proteins (BMPs) and parathyroid hormone-related protein (PTHrP) are some of the soluble factors that facilitate the growth and survival of tumor cells in the bone microenvironment [60, 84]. Runt-related transcription factor (Runx2) is found to be overexpressed in metastatic PCa cell lines and has been shown to mediate bone-specific metastatic behavior in tumor cells [1]. In addition to these biochemical mediators, physical

forces from the microenvironment also influence metastatic progression [45]. Pressure changes in the bone microenvironment have been shown to enhance the migratory behavior of PCa cell lines via induction of Chemokine (C-C motif) ligand 5 (CCL5) from the osteocytes [77].

5.2.3 Exosomes in Metastasis

Extracellular vesicles (EVs) are other important tumor cell messengers that facilitate cell-to-cell communication between primary tumor cells and distant metastatic locations. Exosomes are small EVs of size ranging from 30-100 nm that are secreted by several cell types [20]. These bilayer vesicles carry different biomolecules including: RNA, DNA, protein and lipids. A very recent study by Hoshino et al. have demonstrated that organ-specific metastatic patterns are governed by exosomes released by tumor cells. They injected tagged exosomes from breast and pancreatic cancer cells that specifically metastasize to lungs or bone to study patterns of their distribution *in vivo*. They observed organ-specific deposition of the exosomes that allows for colonization of the disseminated or injected tumor cells to the exact location of exosome deposition. This study suggests that exosomes functionally educate the metastatic locations beforehand to facilitate colonization by the tumor cells [42]. Further, these small vesicular particles are abundantly secreted in most biological fluids under normal and diseased conditions, thus profiling the exosomal particles offers a great opportunity to detect cancer and other pathological conditions at various stages of the disease. Specialized small RNAs such as microRNAs are often enriched in these EVs and are thought to mediate post-transcriptional gene controls in the receptor cells [88]. Thus, they not only function as biological mediators of metastatic progression but can also serve as markers for disease severity.

In conclusion, tumor metastasis is a dynamic process that is governed by several cell intrinsic and extrinsic factors and requires continuous changes in the tumor and surrounding cells for successful execution. Stable genetic changes in the tumors confer genetic fitness to initiate this process, however different steps in the process often require post-transcriptional regulation of several genes controlling the EMT pathways, proteolysis and secretion of factors that allow for cell survival and subsequent colonization. Small non-coding RNA molecules such as miRNAs are important mediators of such posttranscriptional gene controls. We will discuss in detail in the following sections how these specialized small RNAs influence prostate tumorigenesis with a focus on their roles in PCa metastasis.

5.3 MicroRNAs: Small Non-Coding Gene Regulators

MicroRNAs are 22–23 nt long, single stranded non-coding RNA molecules that play an important role in regulating gene transcription post-transcriptionally. They were first discovered in *Caenorhabditis elegans* in 1993 and since then many regulatory miRNAs have been identified. Several studies have demonstrated their role in tumor pathogenesis by regulation of cell growth, proliferation, apoptosis, cell cycle and cell differentiation [26]. Almost 60% of the human genome is regulated by these small RNAs [12]. They are synthesized as a long chain of polycistronic RNA that is cleaved by ribonucleases DROSHA and DICER into 22–23 nt long single stranded RNAs. They bind through complete or partial complementarity to the 3'-UTRs of target mRNAs and together with Argonaute protein form a RISC complex at the mRNA strand that catalyzes its degradation [26]. Their expression is often deregulated in cancer, where miRNAs that lose their expression in tumors are called as “tumor-suppressors” and those that are up-regulated are called “oncomirs”. Studies from our and many other laboratories have provided insights into miRNAs and the important role they play in prostate tumorigenesis. In this chapter, we will be highlighting their contribution in the process of PCa metastasis.

5.3.1 MicroRNAs in Prostate Cancer Metastasis

Tumor cells undergo many gene expression changes to acquire metastatic ability. Gene expression analysis of metastatic tumors have revealed molecular signatures corresponding to cell cycle, transcription factors, signal transduction pathways that can be therapeutically targeted and can predict prognosis for disease recurrence [34, 52]. miRNAs also share this deregulatory behavior in metastatic tumors. Next generation sequencing of metastatic tumors have shown that a large number of small RNAs are differentially expressed and are believe to be important players in the metastatic process [71, 86, 90]. More recently, Xue *et al.* designed a computer algorithm to analyze different PCa metastasis datasets. Based on their observations they identified transcriptional factors AR, HOX6 and NKX2–2 that were altered in metastatic tumors and are believed to regulate the expression of various metastasis-related miRNAs. These TFs were then validated *in vitro* for their functional significance in controlling metastasis by miRNA regulation from prostate epithelial cell line RWPE1 [92]. Deregulation of miRNA expression is often accompanied by metastatic disease and miRNAs are essentially required for the process. We will be discussing roles of various miRNAs in different metastatic steps including: acquisition of EMT, regulation of factors responsible for tumor cell metastasis/colonization and regulation of the microenvironment to facilitate tumor progression.

5.3.1.1 miRNAs in Regulating Acquisition of EMT

Dissemination from the primary site is the first step in metastasis, which requires a tumor cell to undergo transition from an epithelial to a mesenchymal state. Epidermal Growth Factor Receptor (*EGFR*) is well known oncogene that is often overexpressed in many tumors and its expression is thought to promote bone metastasis in breast and prostate cancer. A very recent study by Day *et al.* has shown that circulating tumor cells in PCa patients with bone metastasis overexpress *EGFR* while tumor cells depend on *Her2* overexpression for their growth in the bone microenvironment [24]. Additionally, *EGFR* has been also shown to regulate the expression of miR-1 in PCa cell lines which in turn, controls EMT transcription factor *TWIST1*. Together, they form a mechanistic loop where *EGFR* expression increases with progression of disease and miR-1 expression decreases that leading to an increase in *TWIST1*, thus facilitating EMT progression [15]. It has been previously shown that miR-1 is implicated in metastatic disease and its expression is lost both in primary and metastatic tumors. It also functions as a tumor-suppressor that regulates genes related to the cell-cycle, apoptosis, DNA damage and inhibits the invasive phenotype of PCa cells lines [44]. miR-143 and miR145 are downregulated in bone metastatic tumors and their overexpression in PCa cell lines decreases invasion-migration and bone metastasis forming ability of cancer cell lines *in vivo* [65]. miR-145, which is negatively correlated with *HEF1* expression directly targets the 3'-UTR of *HEF1* mRNA and ablates its EMT conferring properties in PCa cell lines [37]. miR-29b when over-expressed in PC3 cancer cells reduces the invasiveness and lung and liver metastasis forming ability *in vivo*. Furthermore, it leads to up-regulation of E-Cadherin and down-regulation of EMT markers *TWIST1*, N-Cadherin and *SNAI1* [69]. miR-182 and miR-203 target *SNAI2* and induce epithelial phenotypes and self-sufficient growth ability in prostate cells *EPT1* [68]. Human enhancer of filamentation-1 (*HEF-1*) is highly expressed in bone metastatic specimens from PCa patients and it regulates EMT and aggressiveness in PC3 cells. miR-409-3p/5p are members of the delta like 1 homolog-deiodinase, iodothyronine 3 (*DLK1-DIO3*) cluster which are known to be involved in prostate metastasis [47]. miR-409-3p and 5p have been shown to be overexpressed in PCa patient serum and are involved in mediating tumorigenicity, EMT and bone metastasis *in vivo* in PCa cell lines upon its overexpression [47]. miR-154 and miR-379 are other miRNAs found in the same *DLK1-DIO3* cluster that are overexpressed in metastatic bone lesions from PCa patients. Inhibition of these miRNAs in bone metastatic PCa cell lines leads to acquisition of *MET* and reduced bone and soft tissue metastasis from these cell lines [39]. Furthermore, combined overexpression of all 4 miRNAs found in the *DLK1-DIO3* cluster including miR-409-3p/5p, miR-154 and miR-379 was shown to promote EMT in PCa cell lines. They are believed to function in activating oncogenic pathways including Ras, hypoxia-inducible factor (*HIF*), *WNT* and *TGF β* signaling by targeting tumor-suppressors cohesion sub-unit SA-2 (*STAG2*), *SMAD7*, Von Hippel-Lindau tumor-suppressor (*VHL*) and polyhomeotic-like protein 3 (*PHC3*) [39]. miR-195 located on chromosome 17p13.1 functions as a tumor-suppressor and is found to be downregulated in high grade prostate tumors. Ribosomal protein

S6 kinase (RPS6KB-1) was found to be a miR-195 target gene. RPS6KB-1 knock-down restored cell migration, invasion and increased apoptosis observed in PCa cells as a result of miR-195 overexpression. Alterations in the miR-195-RPS6KB-1 axis were shown to regulate expression of MMP-9, BAD, E-Cadherin and VEGF that are involved in PCa progression. This study established the role of miR-195 in preventing PCa metastasis [11]. More recently, it has been shown that increased expression of miR-301a is associated with PCa recurrence in patients undergoing radical prostatectomy. Ectopic overexpression of miR-301a in PCa cell lines PC3 and LNCaP led to increased cell growth, invasion and migration. miR-301a directly targets p63, a member of the p53 family, that in turn alters the expression of EMT proteins E-Cadherin and transcription factor ZEB1 [61]. Recent studies from our lab have demonstrated that genomic loss of chromosome 8p21 is associated with PCa progression. We have identified that miRNAs, miR-3622a and miR-3622b, which are located within this genomic region play an important role in regulating PCa progression [8, 9]. We found that miR-3622a is widely downregulated in PCa and that miR-3622a represses PCa EMT by directly targeting *ZEB1* and *SNAI2*. miR-3622a loss allows tumor cells to acquire a mesenchymal phenotype, promoting invasion and metastasis [8]. Ectopic overexpression of miR-3622b in PCa cell lines led to reduced cellular viability, proliferation, invasiveness, migration and increased apoptosis. miR-3622b overexpression *in vivo* induced regression of established prostate tumor xenografts and miR-3622b was found to directly target EGFR [9].

5.3.1.2 Regulatory Effect of miRNAs on PCa Metastasis–Associated Signaling Pathways and Other Factors

miR-1 has been shown to be downregulated in PCa tumors, more so in the metastatic samples. It directly targets SRC which is known to be a promoter of PCa metastasis [59]. miR-30 is commonly downregulated in PCa tumors and increases with SRC inhibitors. Overexpression of miR-30 in VCaP cells leads to reduction in expression of EMT genes and downregulation of Ets-related genes (ERG). Thus, miR-30 plays an important role in modulating the SRC/EGF and ERG pathways in tumor cells [50]. In Ras-activated xenograft tumors, miR-34a negatively regulates the expression of WNT signaling protein transcription factor 7 (TCF7) and anti-apoptotic protein baculoviral inhibitor of apoptosis repeat containing 5 (BIRC5), both of which are required for successful PCa metastasis in Ras-driven tumors [17]. Levels of miR-194 are often elevated in serum as well as tissues of PCa patients and can serve as a marker for disease recurrence. Forced overexpression in PCa cell lines have shown a pro-metastatic and invasive role for miR-194 in PCa tumorigenesis and it has been shown to directly target ubiquitin ligase suppressor of cytokine signaling 2 (SOCS2) protein. SOCS2 further regulates ubiquitination of two important kinases JAK and FLT3 that in turn, deregulates STAT3-mediated expression of pro-metastatic genes [23]. Loss of miR-15,16 and a concomitant increase in miR-21 activates TGF β signaling pathways and plays an important role in bone colonization of PCa cells [5]. Studies from our group have identified many miRNAs that exert an anti-metastatic

effect by targeting key metastatic genes. miR-203 is frequently lost in metastatic tumors and bone metastatic PCa cell lines. miR-203 over-expression in PCa cells alters the EMT markers, reduces invasion-migration and targets key metastatic genes including survivin, *ZEB2*, *SMAD4*, *DLX5* and *RUNX2* which is a known bone metastasis promoting transcription factor [73]. More recently, we identified miR-466 as another anti-metastasis miRNA that is downregulated in PCa tumors and its overexpression reduces PCa tumor and metastasis growth *in vivo*, targets *RUNX2* and alters the expression of *RUNX2* target genes including *MMP11*, *Angiopoietin (ANGPT1)*, *ANGPT4*, *Osteopontin (OPN)* and *Osteocalcein (BGLAP)* [19].

Cancer stem cells (CSCs) are considered to be a precursor cell population for metastatic tumors and are defined by expression of cell surface markers such as CD44 [83]. These highly tumorigenic and metastatic cell populations have also been identified and characterized in PCa tumors [63, 64]. miR-34a is shown to be downregulated in CD44+ cell populations in PCa xenografts as well as purified CD44+ cells from PCa cell lines. Ectopic overexpression in PCa cell lines have shown that miR-34a directly represses CD44 expression and reduces the migratory and invasive phenotype of CD44+ cells, thus diminishing their metastatic potential [54]. Studies from our lab have demonstrated the role of miR-708 in reducing the tumorigenic potential of CD44+ PCa cells. It was shown to target CD44 and Ser/Thr kinase AKT2, thus altering tumor progression [72]. More recently, we identified that miR-383 located on chromosome 8p is lost during PCa progression and has an inhibitory effect on the CD44+ PCa cell population [10]. miR-128, miR-199-3p, miR-320 and miR141 are some of the other miRNAs that have been shown to regulate prostate metastasis by directly regulating the tumor-initiating stem populations in PCa [43, 46, 55, 56].

5.3.1.3 Microenvironmental Control of miRNAs in Regulation of PCa Metastasis

Metastasis is often marked by loss of BM protein that facilitates invasion of disseminated tumor cells. miR-205 plays an important role in deposition of the major BM protein laminin in prostate tissues. miR-205, along with TP63, regulates the deposition of BM protein and its expression is often lost with PCa tumor progression [31]. Expression of miR-25 is reduced in prostate cancer stem cells (PCSCs) when compared to differentiated luminal cells. Its overexpression has been shown to target expression of integrins αv and $\alpha 6$ in metastatic PCa cell lines and it leads to reduced migration and decreased metastasis *in vivo* [93]. miR-1207-3p is also lost during PCa progression. It has been shown to directly target fibronectin type III domain containing protein (FNDC1) that in turn, regulates fibronectin (FN1) and Androgen receptor (AR) in PCa cell lines. Loss of miR1207-3p is marked by increased expression of FNDC1/FN1/AR that is associated with PCa aggressiveness [22]. In order to understand the role of miRNAs with increasing Gleason grade, when tumors from Gleason grades 3, 4 and 5 were subjected to miRNA gene expression analysis, the results demonstrated miR-29c, miR-34a and mir-141 as

differentially expressed miRNAs that had reduced expression with increasing grade. miRNAs function as tumor-suppressors and their overexpression reduces tumor cell migration and downregulation of ECM, focal adhesion kinase and MAPK13 pathways [53]. Syndecan-1 is another ECM protein that positively regulates levels of miR-331-3p which in turn, contributes towards increased EMT and aggressiveness in prostate tumors [29].

Tumor-associated stroma also undergoes changes in response to progressing tumors. Mesenchymal stem cells (MSCs) have been shown to migrate to tumors and promote prostate tumorigenicity [49]. Co-culture of MSCs with PCa cell lines *in vitro* have been shown to induce more adipogenic differentiation in these cells that is mediated through IL6. Expression of IL6 in MSCs is further regulated by let-7 miRNA, which is downregulated in tumor cells co-cultured with MSCs, thus signifying an important regulatory role of miRNA let-7 in determining the reactivity of tumor stroma [80]. Pre-adipocytes have been shown to be associated with the prostate tumor microenvironment as opposed to normal prostate tissues. They enhance the invasiveness and metastasis of PCa cell lines via upregulation of miR-301a which targets AR expression in tumors and in turn, affects expression of metastasis associated genes *MMP9*, *SMAD3* and TGF- β 3 in PCa cells [91]. Cancer-associated fibroblasts (CAFs) are reactive fibroblasts that are often found in the tumor microenvironment. miRNA analysis of CAFs derived from patients with PCa tumors revealed miR-133a as highly expressed miRNA in these cells. miR-133a released from CAFs functions as a soluble paracrine factor that activates adjacent normal fibroblast to attain a reactive phenotype [27]. In the bone microenvironment, osteoblasts are the main effector cells that allow metastatic colonization by tumor cells under the influence of various factors. It has been shown that osteoblasts secrete Wnt1-induced secreted protein 1 (WISP1) that is released in conditioned media and acts on PCa cell lines to increase their invasion/migration abilities as well increasing the expression of vascular cell adhesion molecule 1 (VCAM1). This effect is mediated by miR-126 downregulation driven by osteoblast-derived WISP1 which further regulates α v β 1/p38 and FAK pathways in PCa cell lines [81]. miR-409-3p/5p are other miRNAs that are found elevated in CAFs in prostate tumors. They are released by EVs from CAFs and upon their uptake by PCa cells mediate tumor cell EMT and aggressiveness [48].

Prolyl 4-hydroxylase alpha polypeptide 1 (P4HA1) enzyme is involved in proper folding of pro-collagen chains. It has been shown to be overexpressed during aggressive PCa and is regulated directly by miR-124. miR-124 is downregulated in high grade PCa and is transcriptionally regulated by EZH2 and transcriptional co-repressor C-terminal binding protein 1 (CtBP1) both of which are increased in aggressive PCa tumors [13].

Comparisons of prostate epithelial cell line P69 with respect to its metastatic subline M12 have shown altered miRNA expression among the two cell lines [16]. miR-130b is down-regulated in the metastatic M12 cell line, PC3, DU145 as well as in prostate tumors. It functions as a tumor suppressor and reduces invasion-migration ability of tumor cells. MMP2 is a direct target of miR-130b and exerts its invasive effect on metastasis as a result of miR-130b down-regulation [16]. miR-296-3p is

over-expressed in M12, thereby down-regulating expression of intercellular cellular adhesion molecule 1 (ICAM1) which in turn, confers a protective effect to circulating tumor cells against natural killer (NK) cells [58].

miR-323 is upregulated in PCa cell lines and is shown to directly target adiponectin receptor 1 (AdipoR1) which in turn, negatively regulates vascular endothelial growth factor-A (VEGF-A)-mediated neovascularization. Thus, mir-323 mediated down-regulation of AdipoR1 facilitates formation of new blood vessels for the growing tumor [32]. Chemokine receptor CXCR4 is elevated in metastatic cell lines and is negatively regulated by miR-494-3p. Ectopic over-expression of miR-494-3p in PCa cell lines inhibits cell invasion and migration [75]. Circulating serum levels of miR-375 have been putatively linked to circulating tumor cells (CTCs) in metastatic CRPC patients. However, miR-375 have been shown to negatively regulate EMT and invasion in PCa cells and targets oncogene YAP1 which is often elevated in invasive PCa tumors. Furthermore, miR-375 is under negative regulation of ZEB1 which enables EMT in PCa cells, thus forming an axis of ZEB1, mir-375 and YAP1 that controls epithelial cell EMT and MET transitions [74].

5.4 Conclusions and Future Directions

Evidences from the literature suggest an essential role for miRNAs in the metastatic process (Fig. 5.1). In most PCa metastases, regulatory small RNAs are lost during tumor progression and function mostly as tumor suppressors. Thus, they inhibit

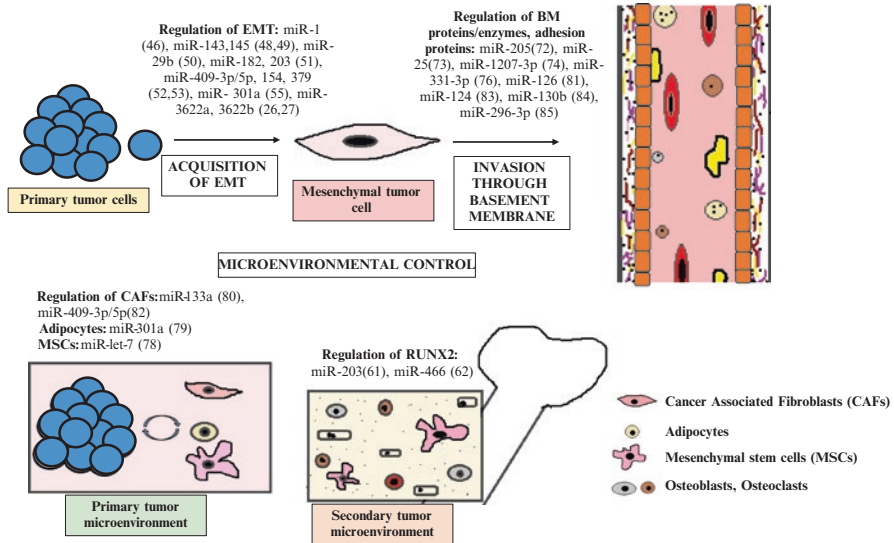


Fig. 5.1 Schematic representation of PCa metastasis and regulation of different steps by various miRNAs

metastatic initiation via EMT or regulate growth of primary tumor cells in the primary or metastatic microenvironment via control over important growth factors, chemokines, ECM and stromal components. In addition to their functional effects in mediating PCa metastasis, expression in tissues and circulatory are often important indicators of disease severity. Non-invasive sampling of cell free or EV-derived RNAs in the serum, plasma or urine offers a great opportunity for sensitive detection of metastatic disease [28]. Moreover, much research these days is focused on utilizing the therapeutic potential of miRNA in cancers. Different ways for delivering miRNA to their target cells i.e. via nanoparticles, liposomes, viral particle-mediated transfer or polyethylene glycol (PEG)-based particles, have been developed for systemic and local delivery of miRNAs [18, 66]. Atelocollagen particle-mediated delivery of miR-16 and chitosan nanoparticle-derived delivery of miR-34a to PCa xenografts *in vivo* have highlighted the promising effect of miRNA delivery in inhibiting advanced prostate cancer [33, 82]. Given the multifaceted roles of miRNAs, more research efforts are needed to improve PCa detection and the efficacy of disease therapeutics utilizing small regulatory miRNAs.

Acknowledgements We thank Dr. Roger Erickson for his support with preparation of the manuscript. Research in authors' lab is supported by *the National Cancer Institute at the NIH* (Grant Number RO1CA177984).

Conflict of Interest None.

References

1. Akech J, Wixted JJ, Bedard K, Van der Deen M, Hussain S, Guise TA, Van Wijnen AJ, Stein JL, Languino LR, Altieri DC, Pratap J, Keller E, Stein GS, Lian JB (2010) Runx2 association with progression of prostate cancer in patients: mechanisms mediating bone osteolysis and osteoblastic metastatic lesions. *Oncogene* 29:811–821
2. Baca SC, Prandi D, Lawrence MS, Mosquera JM, Romanel A, Drier Y, Park K, Kitabayashi N, Macdonald TY, Ghandi M, Van Allen E, Kryukov GV, Sboner A, Theurillat JP, Soong TD, Nickerson E, Auclair D, Tewari A, Beltran H, Onofrio RC, Boysen G, Guiducci C, Barbieri CE, Cibulskis K, Sivachenko A, Carter SL, Saksena G, Voet D, Ramos AH, Winckler W, Cipicchio M, Ardlie K, Kantoff PW, Berger MF, Gabriel SB, Golub TR, Meyerson M, Lander ES, Elemento O, Getz G, Demichelis F, Rubin MA, Garraway LA (2013) Punctuated evolution of prostate cancer genomes. *Cell* 153:666–677
3. Berger MF, Lawrence MS, Demichelis F, Drier Y, Cibulskis K, Sivachenko AY, Sboner A, Esgueva R, Pflueger D, Sougnez C, Onofrio R, Carter SL, Park K, Habegger L, Ambrogio L, Fennell T, Parkin M, Saksena G, Voet D, Ramos AH, Pugh TJ, Wilkinson J, Fisher S, Winckler W, Mahan S, Ardlie K, Baldwin J, Simons JW, Kitabayashi N, Macdonald TY, Kantoff PW, Chin L, Gabriel SB, Gerstein MB, Golub TR, Meyerson M, Tewari A, Lander ES, Getz G, Rubin MA, Garraway LA (2011) The genomic complexity of primary human prostate cancer. *Nature* 470:214–220
4. Bhatia-Gaur R, Donjacour AA, Scivolino PJ, Kim M, Desai N, Young P, Norton CR, Gridley T, Cardiff RD, Cunha GR, Abate-Shen C, Shen MM (1999) Roles for Nkx3.1 in prostate development and cancer. *Genes Dev* 13:966–977

5. Bonci D, Coppola V, Patrizii M, Addario A, Cannistraci A, Francescangeli F, Pecci R, Muto G, Collura D, Bedini R, Zeuner A, Valtieri M, Sentinelli S, Benassi MS, Gallucci M, Carlini P, Piccolo S, de Maria R (2016) A microRNA code for prostate cancer metastasis. *Oncogene* 35:1180–1192
6. Bowen C, Bubendorf L, Voeller HJ, Slack R, Willi N, Sauter G, Gasser TC, Koivisto P, Lack EE, Kononen J, Kallioniemi OP, Gelmann EP (2000) Loss of NKX3.1 expression in human prostate cancers correlates with tumor progression. *Cancer Res* 60:6111–6115
7. Boyd LK, Mao X, Lu YJ (2012) The complexity of prostate cancer: genomic alterations and heterogeneity. *Nat Rev Urol* 9:652–664
8. Bucay N, Bhagirath D, Sekhon K, Yang T, Fukuhara S, Majid S, Shahryari V, Tabatabai Z, Greene KL, Hashimoto Y, Shiina M, Yamamura S, Tanaka Y, Deng G, Dahiya R, Saini S (2017) A novel microRNA regulator of prostate cancer epithelial-mesenchymal transition. *Cell Death Differ* 24:1263–1274
9. Bucay N, Sekhon K, Majid S, Yamamura S, Shahryari V, Tabatabai ZL, Greene K, Tanaka Y, Dahiya R, Deng G, Saini S (2016a) Novel tumor suppressor microRNA at frequently deleted chromosomal region 8p21 regulates epidermal growth factor receptor in prostate cancer. *Oncotarget* 7:70388–70403
10. Bucay N, Sekhon K, Yang T, Majid S, Shahryari V, Hsieh C, Mitsui Y, Deng G, Tabatabai ZL, Yamamura S, Calin GA, Dahiya R, Tanaka Y, Saini S (2016b) MicroRNA-383 located in frequently deleted chromosomal locus 8p22 regulates CD44 in prostate cancer. *Oncogene*
11. Cai C, Chen QB, Han ZD, Zhang YQ, He HC, Chen JH, Chen YR, Yang SB, Wu YD, Zeng YR, Qin GQ, Liang YX, Dai QS, Jiang FN, Wu SL, Zeng GH, Zhong WD, Wu CL (2015) miR-195 inhibits tumor progression by targeting RPS6KB1 in human prostate Cancer. *Clin Cancer Res* 21:4922–4934
12. Calin GA, Croce CM (2006) MicroRNA signatures in human cancers. *Nat Rev Cancer* 6:857–866
13. Chakravarthi BV, Pathi SS, Goswami MT, Cieslik M, Zheng H, Nallasivam S, Arekapudi SR, Jing X, Siddiqui J, Athanikar J, Carskadon SL, Lonigro RJ, Kunju LP, Chinnaiyan AM, Palanisamy N, Varambally S (2014) The miR-124-prolyl hydroxylase P4HA1-MMP1 axis plays a critical role in prostate cancer progression. *Oncotarget* 5:6654–6669
14. Chambers AF, Groom AC, Macdonald IC (2002) Dissemination and growth of cancer cells in metastatic sites. *Nat Rev Cancer* 2:563–572
15. Chang YS, Chen WY, Yin JJ, Sheppard-Tillman H, Huang J, Liu YN (2015) EGF receptor promotes prostate Cancer bone metastasis by downregulating miR-1 and activating TWIST1. *Cancer Res* 75:3077–3086
16. Chen Q, Zhao X, Zhang H, Yuan H, Zhu M, Sun Q, Lai X, Wang Y, Huang J, Yan J, Yu J (2015a) MiR-130b suppresses prostate cancer metastasis through down-regulation of MMP2. *Mol Carcinog* 54:1292–1300
17. Chen WY, Liu SY, Chang YS, Yin JJ, Yeh HL, Mouhieddine TH, Hadadeh O, Abou-Kheir W, Liu YN (2015b) MicroRNA-34a regulates WNT/TCF7 signaling and inhibits bone metastasis in Ras-activated prostate cancer. *Oncotarget* 6:441–457
18. Chen Y, Gao DY, Huang L (2015c) In vivo delivery of miRNAs for cancer therapy: challenges and strategies. *Adv Drug Deliv Rev* 81:128–141
19. Colden M, Dar AA, Saini S, Dahiya PV, Shahryari V, Yamamura S, Tanaka Y, Stein G, Dahiya R, Majid S (2017) MicroRNA-466 inhibits tumor growth and bone metastasis in prostate cancer by direct regulation of osteogenic transcription factor RUNX2. *Cell Death Dis* 8:e2572
20. Colombo M, Raposo G, Thery C (2014) Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. *Annu Rev Cell Dev Biol* 30:255–289
21. Dalela D, Sun M, Diaz M, Karabon P, Seisen T, Trinh QD, Menon M, Abdollah F (2017) Contemporary trends in the incidence of metastatic prostate Cancer among US men: Results from Nationwide analyses. *Eur Urol Focus*

22. Das DK, Naidoo M, Ilboudo A, Park JY, Ali T, Krampis K, Robinson BD, Osborne JR, Ogunwobi OO (2016) miR-1207-3p regulates the androgen receptor in prostate cancer via FNDC1/fibronectin. *Exp Cell Res* 348:190–200
23. Das R, Gregory PA, Fernandes RC, Denis I, Wang Q, Townley SL, Zhao SG, Hanson AR, Pickering MA, Armstrong HK, Lokman NA, Ebrahimie E, Davicioni E, Jenkins RB, Karnes RJ, Ross AE, Den RB, Klein EA, Chi KN, Ramshaw HS, Williams ED, Zoubeidi A, Goodall GJ, Feng FY, Butler LM, Tilley WD, Selth LA (2017) MicroRNA-194 promotes prostate Cancer metastasis by inhibiting SOCS2. *Cancer Res* 77:1021–1034
24. Day KC, Lorenzatti Hiles G, Kozminsky M, Dawsey SJ, Paul A, Brose LJ, Shah R, Kunja LP, Hall C, Palanisamy N, Daignault-Newton S, El-Sawy L, Wilson SJ, Chou A, Ignatowski KW, Keller E, Thomas D, Nagrath S, Morgan T, Day ML (2017) HER2 and EGFR overexpression support metastatic progression of prostate Cancer to bone. *Cancer Res* 77:74–85
25. Deplus R, Delliaux C, Marchand N, Flourens A, Vanpouille N, Leroy X, de Launoit Y, Duterque-Coquillaud M (2017) TMPRSS2-ERG fusion promotes prostate cancer metastases in bone. *Oncotarget* 8:11827–11840
26. di Leva G, Garofalo M, Croce CM (2014) MicroRNAs in cancer. *Annu Rev Pathol* 9:287–314
27. Doldi V, Callari M, Giannoni E, D'Aiuto F, Maffezzini M, Valdagni R, Chiarugi P, Gandellini P, Zaffaroni N (2015) Integrated gene and miRNA expression analysis of prostate cancer associated fibroblasts supports a prominent role for interleukin-6 in fibroblast activation. *Oncotarget* 6:31441–31460
28. Fendler A, Stephan C, Yousef GM, Kristiansen G, Jung K (2016) The translational potential of microRNAs as biofluid markers of urological tumours. *Nat Rev Urol* 13:734–752
29. Fujii T, Shimada K, Tatsumi Y, Tanaka N, Fujimoto K, Konishi N (2016) Syndecan-1 up-regulates microRNA-331-3p and mediates epithelial-to-mesenchymal transition in prostate cancer. *Mol Carcinog* 55:1378–1386
30. Gandaglia G, Karakiewicz PI, Briganti A, Passoni NM, Schiffmann J, Trudeau V, Graefen M, Montorsi F, Sun M (2015) Impact of the site of metastases on survival in patients with metastatic prostate Cancer. *Eur Urol* 68:325–334
31. Gandellini P, Profumo V, Casamichele A, Fenderico N, Borrelli S, Petrovich G, Santilli G, Callari M, Colecchia M, Pozzi S, de Cesare M, Folini M, Valdagni R, Mantovani R, Zaffaroni N (2012) miR-205 regulates basement membrane deposition in human prostate: implications for cancer development. *Cell Death Differ* 19:1750–1760
32. Gao Q, Yao X, Zheng J (2015) MiR-323 inhibits prostate Cancer vascularization through adiponectin receptor. *Cell Physiol Biochem* 36:1491–1498
33. Gaur S, Wen Y, Song JH, Parikh NU, Mangala LS, Blessing AM, Ivan C, Wu SY, Varkaris A, Shi Y, Lopez-Berestein G, Frigo DE, Sood AK, Gallick GE (2015) Chitosan nanoparticle-mediated delivery of miRNA-34a decreases prostate tumor growth in the bone and its expression induces non-canonical autophagy. *Oncotarget* 6:29161–29177
34. Glinksy GV, Glinksii AB, Stephenson AJ, Hoffman RM, Gerald WL (2004) Gene expression profiling predicts clinical outcome of prostate cancer. *J Clin Invest* 113:913–923
35. Guo CC, Wang Y, Xiao L, Troncoso P, Czerniak BA (2012) The relationship of TMPRSS2-ERG gene fusion between primary and metastatic prostate cancers. *Hum Pathol* 43:644–649
36. Guo F, Parker Kerrigan BC, Yang D, Hu L, Shmulevich I, Sood AK, Xue F, Zhang W (2014) Post-transcriptional regulatory network of epithelial-to-mesenchymal and mesenchymal-to-epithelial transitions. *J Hematol Oncol* 7:19
37. Guo W, Ren D, Chen X, Tu X, Huang S, Wang M, Song L, Zou X, Peng X (2013) HEF1 promotes epithelial mesenchymal transition and bone invasion in prostate cancer under the regulation of microRNA-145. *J Cell Biochem* 114:1606–1615
38. Gupta GP, Massague J (2006) Cancer metastasis: building a framework. *Cell* 127:679–695
39. Gururajan M, Jossan S, Chu GC, Lu CL, Lu YT, Haga CL, Zhau HE, Liu C, Lichterman J, Duan P, Posadas EM, Chung LW (2014) miR-154* and miR-379 in the DLK1-DIO3 microRNA mega-cluster regulate epithelial to mesenchymal transition and bone metastasis of prostate cancer. *Clin Cancer Res* 20:6559–6569

40. He WW, Sciavolino PJ, Wing J, Augustus M, Hudson P, Meissner PS, Curtis RT, Shell BK, Bostwick DG, Tindall DJ, Gelmann EP, Abate-Shen C, Carter KC (1997) A novel human prostate-specific, androgen-regulated homeobox gene (NKX3.1) that maps to 8p21, a region frequently deleted in prostate cancer. *Genomics* 43:69–77
41. Hong MK, Macintyre G, Wedge DC, Van Loo P, Patel K, Lunke S, Alexandrov LB, Sloggett C, Cmero M, Marass F, Tsui D, Mangiola S, Lonie A, Naeem H, Sapre N, Phal PM, Kurganovs N, Chin X, Kerger M, Warren AY, Neal D, Gnanapragasam V, Rosenfeld N, Pedersen JS, Ryan A, Haviv I, Costello AJ, Corcoran NM, Hovens CM (2015) Tracking the origins and drivers of subclonal metastatic expansion in prostate cancer. *Nat Commun* 6:6605
42. Hoshino A, Costa-Silva B, Shen TL, Rodrigues G, Hashimoto A, Tesic Mark M, Molina H, Kohsaka S, di Giannatale A, Ceder S, Singh S, Williams C, Soplop N, Uryu K, Pharmed L, King T, Bojmar L, Davies AE, Ararso Y, Zhang T, Zhang H, Hernandez J, Weiss JM, Dumont-Cole VD, Kramer K, Wexler LH, Narendran A, Schwartz GK, Healey JH, Sandstrom P, Labori KJ, Kure EH, Grandgenett PM, Hollingsworth MA, de Sousa M, Kaur S, Jain M, Mallya K, Batra SK, Jarnagin WR, Brady MS, Fodstad O, Muller V, Pantel K, Minn AJ, Bissell MJ, Garcia BA, Kang Y, Rajasekhar VK, Ghajar CM, Matei I, Peinado H, Bromberg J, Lyden D (2015) Tumour exosome integrins determine organotropic metastasis. *Nature* 527:329–335
43. Hsieh IS, Chang KC, Tsai YT, Ke JY, Lu PJ, Lee KH, Yeh SD, Hong TM, Chen YL (2013) MicroRNA-320 suppresses the stem cell-like characteristics of prostate cancer cells by down-regulating the Wnt/beta-catenin signaling pathway. *Carcinogenesis* 34:530–538
44. Hudson RS, Yi M, Esposito D, Watkins SK, Hurwitz AA, Yfantis HG, Lee DH, Borin JF, Naslund MJ, Alexander RB, Dorsey TH, Stephens RM, Croce CM, Ambs S (2012) MicroRNA-1 is a candidate tumor suppressor and prognostic marker in human prostate cancer. *Nucleic Acids Res* 40:3689–3703
45. Jaalouk DE, Lammerding J (2009) Mechanotransduction gone awry. *Nat Rev Mol Cell Biol* 10:63–73
46. Jin M, Zhang T, Liu C, Badeaux MA, Liu B, Liu R, Jeter C, Chen X, Vlassov AV, Tang DG (2014) miRNA-128 suppresses prostate cancer by inhibiting BMI-1 to inhibit tumor-initiating cells. *Cancer Res* 74:4183–4195
47. Josson S, Gururajan M, Hu P, Shao C, Chu GY, Zhou HE, Liu C, Lao K, Lu CL, Lu YT, Lichterman J, Nandana S, Li Q, Rogatko A, Berel D, Posadas EM, Fazli L, Sareen D, Chung LW (2014) miR-409-3p/–5p promotes tumorigenesis, epithelial-to-mesenchymal transition, and bone metastasis of human prostate cancer. *Clin Cancer Res* 20:4636–4646
48. Josson S, Gururajan M, Sung SY, Hu P, Shao C, Zhou HE, Liu C, Lichterman J, Duan P, Li Q, Rogatko A, Posadas EM, Haga CL, Chung LW (2015) Stromal fibroblast-derived miR-409 promotes epithelial-to-mesenchymal transition and prostate tumorigenesis. *Oncogene* 34:2690–2699
49. Jung Y, Kim JK, Shiozawa Y, Wang J, Mishra A, Joseph J, Berry JE, Mcgee S, Lee E, Sun H, Wang J, Jin T, Zhang H, Dai J, Krebsbach PH, Keller ET, Pienta KJ, Taichman RS (2013) Recruitment of mesenchymal stem cells into prostate tumours promotes metastasis. *Nat Commun* 4:1795
50. Kao CJ, Martinez A, Shi XB, Yang J, Evans CP, Dobi A, Devere White RW, Kung HJ (2014) miR-30 as a tumor suppressor connects EGF/Src signal to ERG and EMT. *Oncogene* 33:2495–2503
51. Kelly SP, Anderson WF, Rosenberg PS, Cook MB (2017) Past, current, and future incidence rates and burden of metastatic prostate Cancer in the United States. *Eur Urol Focus*
52. Latulippe E, Satagopan J, Smith A, Scher H, Scardino P, Reuter V, Gerald WL (2002) Comprehensive gene expression analysis of prostate cancer reveals distinct transcriptional programs associated with metastatic disease. *Cancer Res* 62:4499–4506
53. Lichner Z, Ding Q, Samaan S, Saleh C, Nasser A, Al-Haddad S, Samuel JN, Fleshner NE, Stephan C, Jung K, Yousef GM (2015) miRNAs dysregulated in association with Gleason grade regulate extracellular matrix, cytoskeleton and androgen receptor pathways. *J Pathol* 237:226–237

54. Liu C, Kelnar K, Liu B, Chen X, Calhoun-Davis T, Li H, Patrawala L, YAN H, Jeter C, Honorio S, Wiggins JF, Bader AG, Fagin R, Brown D, Tang DG (2011) The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. *Nat Med* 17:211–215
55. Liu C, Liu R, Zhang D, Deng Q, Liu B, Chao HP, Rycaj K, Takata Y, Lin K, Lu Y, Zhong Y, Krolewski J, Shen J, Tang DG (2017) MicroRNA-141 suppresses prostate cancer stem cells and metastasis by targeting a cohort of pro-metastasis genes. *Nat Commun* 8:14270
56. Liu R, Liu C, Zhang D, Liu B, Chen X, Rycaj K, Jeter C, Calhoun-Davis T, Li Y, Yang T, Wang J, Tang DG (2016) miR-199a-3p targets stemness-related and mitogenic signaling pathways to suppress the expansion and tumorigenic capabilities of prostate cancer stem cells. *Oncotarget* 7:56628–56642
57. Liu W, Laitinen S, Khan S, Vihinen M, Kowalski J, Yu G, Chen L, Ewing CM, Eisenberger MA, Carducci MA, Nelson WG, Yegnasubramanian S, Luo J, Wang Y, Xu J, Isaacs WB, Visakorpi T, Bova GS (2009) Copy number analysis indicates monoclonal origin of lethal metastatic prostate cancer. *Nat Med* 15:559–565
58. Liu X, Chen Q, Yan J, Wang Y, Zhu C, Chen C, Zhao X, Xu M, Sun Q, Deng R, Zhang H, Qu Y, Huang J, Jiang B, Yu J (2013) MiRNA-296-3p-ICAM-1 axis promotes metastasis of prostate cancer by possible enhancing survival of natural killer cell-resistant circulating tumour cells. *Cell Death Dis* 4:e928
59. Liu YN, Yin J, Barrett B, Sheppard-Tillman H, Li D, Casey OM, Fang L, Hynes PG, Ameri AH, Kelly K (2015) Loss of androgen-regulated MicroRNA 1 activates SRC and promotes prostate Cancer bone metastasis. *Mol Cell Biol* 35:1940–1951
60. Msaouel P, Pissimissis N, Halapas A, Koutsilieris M (2008) Mechanisms of bone metastasis in prostate cancer: clinical implications. *Best Pract Res Clin Endocrinol Metab* 22:341–355
61. Nam RK, Benatar T, Wallis CJ, Amemiya Y, Yang W, Garbens A, Naeim M, Sherman C, Sugar L, Seth A (2016) MiR-301a regulates E-cadherin expression and is predictive of prostate cancer recurrence. *Prostate* 76:869–884
62. Nguyen DX, Bos PD, Massague J (2009) Metastasis: from dissemination to organ-specific colonization. *Nat Rev Cancer* 9:274–284
63. Patrawala L, Calhoun-Davis T, Schneider-Broussard R, Tang DG (2007) Hierarchical organization of prostate cancer cells in xenograft tumors: the CD44+alpha2beta1+ cell population is enriched in tumor-initiating cells. *Cancer Res* 67:6796–6805
64. Patrawala L, Calhoun T, Schneider-Broussard R, Li H, Bhatia B, Tang S, Reilly JG, Chandra D, Zhou J, Claypool K, Coghlan L, Tang DG (2006) Highly purified CD44+ prostate cancer cells from xenograft human tumors are enriched in tumorigenic and metastatic progenitor cells. *Oncogene* 25:1696–1708
65. Peng X, Guo W, Liu T, Wang X, Tu X, Xiong D, Chen S, Lai Y, Du H, Chen G, Liu G, Tang Y, Huang S, Zou X (2011) Identification of miRs-143 and -145 that is associated with bone metastasis of prostate cancer and involved in the regulation of EMT. *PLoS One* 6:e20341
66. Pereira DM, Rodrigues PM, Borralho PM, Rodrigues CM (2013) Delivering the promise of miRNA cancer therapeutics. *Drug Discov Today* 18:282–289
67. Pritchard CC, Mateo J, Walsh MF, de Sarkar N, Abida W, Beltran H, Garofalo A, Gulati R, Carreira S, Eeles R, Elemento O, Rubin MA, Robinson D, Lonigro R, Hussain M, Chinnaiyan A, Vinson J, Filipenko J, Garraway L, Taplin ME, Aldubayan S, Han GC, Beightol M, Morrissey C, Nghiem B, Cheng HH, Montgomery B, Walsh T, Casadei S, Berger M, Zhang L, Zehir A, Vijai J, Scher HI, Sawyers C, Schultz N, Kantoff PW, Solit D, Robson M, van Allen EM, Offit K, de Bono J, Nelson PS (2016) Inherited DNA-repair gene mutations in men with metastatic prostate Cancer. *N Engl J Med* 375:443–453
68. Qu Y, Li WC, Hellem MR, Rostad K, Popa M, McCormack E, Oyan AM, Kalland KH, Ke XS (2013) MiR-182 and miR-203 induce mesenchymal to epithelial transition and self-sufficiency of growth signals via repressing SNAI2 in prostate cells. *Int J Cancer* 133:544–555

69. Ru P, Steele R, Newhall P, Phillips NJ, Toth K, Ray RB (2012) miRNA-29b suppresses prostate cancer metastasis by regulating epithelial-mesenchymal transition signaling. *Mol Cancer Ther* 11:1166–1173
70. Rycaj K, Li H, Zhou J, Chen X, Tang DG (2017) Cellular determinants and microenvironmental regulation of prostate cancer metastasis. *Semin Cancer Biol* 44:83–97
71. Sadeghi M, Ranjbar B, Ganjalikhany MR, Khan FM, Schmitz U, Wolkenhauer O, Gupta SK (2016) MicroRNA and transcription factor gene regulatory network analysis reveals key regulatory elements associated with prostate Cancer progression. *PLoS One* 11:e0168760
72. Saini S, Majid S, Shahryari V, Arora S, Yamamura S, Chang I, Zaman MS, Deng G, Tanaka Y, Dahiya R (2012) miRNA-708 control of CD44(+) prostate cancer-initiating cells. *Cancer Res* 72:3618–3630
73. Saini S, Majid S, Yamamura S, Tabatabai L, Suh SO, Shahryari V, Chen Y, Deng G, Tanaka Y, Dahiya R (2011) Regulatory role of mir-203 in prostate Cancer progression and metastasis. *Clin Cancer Res* 17:5287–5298
74. Selth LA, Das R, Townley SL, Coutinho I, Hanson AR, Centenera MM, Stylianou N, Sweeney K, Soekmadji C, Jovanovic L, Nelson CC, Zoubeidi A, Butler LM, Goodall GJ, Hollier BG, Gregory PA, Tilley WD (2017) A ZEB1-miR-375-YAP1 pathway regulates epithelial plasticity in prostate cancer. *Oncogene* 36:24–34
75. Shen PF, Chen XQ, Liao YC, Chen N, Zhou Q, Wei Q, Li X, Wang J, Zeng H (2014) MicroRNA-494-3p targets CXCR4 to suppress the proliferation, invasion, and migration of prostate cancer. *Prostate* 74:756–767
76. Siegel RL, Miller KD, Jemal A (2017) Cancer statistics, 2017. *CA Cancer J Clin* 67:7–30
77. Sottnik JL, Dai J, Zhang H, Campbell B, Keller ET (2015) Tumor-induced pressure in the bone microenvironment causes osteocytes to promote the growth of prostate cancer bone metastases. *Cancer Res* 75:2151–2158
78. Stankiewicz E, Mao X, Mangham DC, Xu L, Yeste-Velasco M, Fisher G, North B, Chaplin T, Young B, Wang Y, Kaur Bansal J, Kudahetti S, Spencer L, Foster CS, Moller H, Scardino P, Oliver RT, Shamash J, Cuzick J, Cooper CS, Berney DM, Lu YJ (2017) Identification of FBXL4 as a metastasis associated gene in prostate Cancer. *Sci Rep* 7:5124
79. Stewart DA, Cooper CR, Sikes RA (2004) Changes in extracellular matrix (ECM) and ECM-associated proteins in the metastatic progression of prostate cancer. *Reprod Biol Endocrinol* 2:2
80. Sung SY, Liao C H, Wu HP, Hsiao WC, Wu IH, Jinpu Yu, Lin SH, Hsieh CL (2013) Loss of let-7 microRNA upregulates IL-6 in bone marrow-derived mesenchymal stem cells triggering a reactive stromal response to prostate cancer. *PLoS One* 8:e71637
81. Tai HC, Chang AC, Yu HJ, Huang CY, Tsai YC, Lai YW, Sun HL, Tang CH, Wang SW (2014) Osteoblast-derived WNT-induced secreted protein 1 increases VCAM-1 expression and enhances prostate cancer metastasis by down-regulating miR-126. *Oncotarget* 5:7589–7598
82. Takeshita F, Patrawala L, Osaki M, Takahashi RU, Yamamoto Y, Kosaka N, Kawamata M, Kelnar K, Bader AG, Brown D, Ochiya T (2010) Systemic delivery of synthetic microRNA-16 inhibits the growth of metastatic prostate tumors via downregulation of multiple cell-cycle genes. *Mol Ther* 18:181–187
83. Tang DG, Patrawala L, Calhoun T, Bhatia B, Choy G, Schneider-Broussard R, JETER C (2007) Prostate cancer stem/progenitor cells: identification, characterization, and implications. *Mol Carcinog* 46:1–14
84. Tantivejkul K, Kalikin LM, Pienta KJ (2004) Dynamic process of prostate cancer metastasis to bone. *J Cell Biochem* 91:706–717
85. Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R, Sun XW, Varambally S, Cao X, Tchinda J, Kuefer R, Lee C, Montie JE, Shah RB, Pienta KJ, Rubin MA, Chinnaiyan AM (2005) Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. *Science* 310:644–648
86. Tong AW, Fulgham P, Jay C, Chen P, Khalil I, Liu S, Senzer N, Eklund AC, Han J, Nemunaitis J (2009) MicroRNA profile analysis of human prostate cancers. *Cancer Gene Ther* 16:206–216

87. Tyekuceva S, Bowden M, Bango C, Giunchi F, Huang Y, Zhou C, Bondi A, Lis R, Van Hemelrijck M, Andren O, Andersson SO, Watson RW, Pennington S, Finn SP, Martin NE, Stampfer MJ, Parmigiani G, Penney KL, Fiorentino M, Mucci LA, Loda M (2017) Stromal and epithelial transcriptional map of initiation progression and metastatic potential of human prostate cancer. *Nat Commun* 8:420
88. Valencia K, Luis-Ravelo D, Bovy N, Anton I, Martinez-Canarias S, Zanduetta C, Ormazabal C, Struman I, Tabruyn S, Rebmann V, de Las Rivas J, Guruceaga E, Bandres E, Lecanda F (2014) miRNA cargo within exosome-like vesicle transfer influences metastatic bone colonization. *Mol Oncol* 8:689–703
89. Varambally S, Dhanasekaran SM, Zhou M, Barrette TR, Kumar-Sinha C, Sanda MG, Ghosh D, Pienta KJ, Sewalt RG, Otte AP, Rubin MA, Chinnaiyan AM (2002) The polycomb group protein EZH2 is involved in progression of prostate cancer. *Nature* 419:624–629
90. Watahiki A, Wang Y, Morris J, Dennis K, O'dwyer HM, Gleave M, Gout PW, Wang Y (2011) MicroRNAs associated with metastatic prostate cancer. *PLoS One* 6:e24950
91. Xie H, Li L, Zhu G, Dang Q, Ma Z, He D, Chang L, Song W, Chang HC, Krolewski JJ, Nastiuk KL, Yeh S, Chang C (2015) Infiltrated pre-adipocytes increase prostate cancer metastasis via modulation of the miR-301a/androgen receptor (AR)/TGF-beta1/Smad/MMP9 signals. *Oncotarget* 6:12326–12339
92. Xue M, Liu H, Zhang L, Chang H, Liu Y, Du S, Yang Y, Wang P (2017) Computational identification of mutually exclusive transcriptional drivers dysregulating metastatic microRNAs in prostate cancer. *Nat Commun* 8:14917
93. Zoni E, van der Horst G, van de Merbel AF, Chen L, Rane JK, Pelger RC, Collins AT, Visakorpi T, Snaar-Jagalska BE, Maitland NJ, van der Pluijm G (2015) miR-25 modulates invasiveness and dissemination of human prostate Cancer cells via regulation of alphav- and alpha6-integrin expression. *Cancer Res* 75:2326–2336