Mycotoxins and Human Disease

Aleksandra Barac

Abbreviations

AFB1	Aflatoxins B1
IARC	International Agency for Research on
	Cancer
HBV	Hepatitis B
AFM1	Aflatoxin M1
DON	Deoxynivalenol
OTA	Ochratoxin A
BEN	Balkan endemic nephropathy
GI	Gastrointestinal
WDB	Water-damaged buildings
NAC	N-Acetylcysteine

14.1 History

Mycotoxins are secondary fungal metabolites that can be produced in crops and other food commodities. When ingested, mycotoxins may cause a mycotoxicosis, the term first used by Forgacs and Carll [1], which can result in an acute or chronic disease episode [2]. The oldest recognized mycotoxicosis of humans is ergotism caused by the plant parasitic fungus, *Claviceps purpurea*. After periodic outbreaks in central Europe, the disease became epidemic in the Middle Ages, where it was

A. Barac

known as St. Anthony's fire [3, 4]. Gangrenous symptoms were described in medieval episodes of ergotism in humans, where early symptoms were hallucinations and swollen limbs with burning sensations, with subsequent necrosis leading to loss of appendages [4].

Today, the word *mycotoxin* simply means a toxin produced by a fungus. Modern mycotoxicology began with the discovery of the aflatoxins in the early 1960s [3]. Since that time, numerous other mycotoxins have been discovered [5].

14.2 Introduction

Mycotoxins are a relatively large, diverse group of naturally occurring, fungal toxins, many of which have been strongly implicated as chemical agents of toxic disease in humans and animals. The history and use of mycotoxins are very long and go back to antiquity, e.g., the philosopher Socrates was executed by drinking of a mixture containing poison hemlock. Today, there are potentially 20,000 to 300,000 known mycotoxins of micro- and macrofungi. Microfungi are the surrogate for the fungus itself that produces mycotoxins on the cellular level. Macrofungi refer to the fruiting bodies (mushrooms) that are appreciated deli food but may be poisonous and deadly. The diversity of toxic mechanisms will be equally as great. The number of mycotoxins actually known to be involved in disease is considerably less, but even this number is difficult to



14

[©] Springer International Publishing AG, part of Springer Nature 2019 E. Presterl (ed.), *Clinically Relevant Mycoses*, https://doi.org/10.1007/978-3-319-92300-0_14

Clinic for Infectious and Tropical Diseases, Clinical Centre of Serbia, Faculty of Medicine, University of Belgrade, Belgrade, Serbia

assess due to the diversity of effects of these unique compounds on humans [6-9].

The usual route of mycotoxin exposure is ingestion as food or feed contaminants. However, dermal and inhalation also may be important routes of exposure. Direct effects of mycotoxins range from acute disease where severe conditions of altered health may exist prior to death as a result of exposure to the toxin. These conditions are more likely following exposure to high levels of a mycotoxin. Other, more insidious or occult conditions (e.g., growth retardation, impaired immunity, decreased disease resistance) or more chronic disease manifestations (e.g., tumor formation) may result from prolonged exposure to small quantities of toxin [7, 10, 11]. Awareness is growing regarding the hazards of mycotoxins as contaminants of food and feed.

14.3 Microfungi

14.3.1 Classes of Mycotoxins

The mycotoxins that pose the greatest potential risk to human and animal health as food and feed aflatoxins. trichothecenes. contaminants are fumonisins, zearalenone, ochratoxin A (OTA), and ergot alkaloids. However, other mycotoxins should be included because of their frequency of occurrence in commodities, their products, or their cooccurrence with other important mycotoxins. This expanded list includes cyclopiazonic acid, sterigmatocystin, gliotoxin, citrinin, penitrems (perhaps other tremorogenic mycotoxins), patulin, and miscellaneous mycotoxins such as fusarin C, fusaric acid, penicillic acid, mycophenolic acid, roquefortine, Penicillium roqueforti (PR) toxin, and isofumigaclavines A and B [7, 9–11].

14.3.1.1 Major Classes of Mycotoxins

Aflatoxins are naturally occurring mycotoxins that are produced by various species of *Aspergillus*. The major aflatoxins commonly isolated from foods and feeds are aflatoxins B1 (AFB1), B2, G1, and G2 [12]. Aflatoxin is considered as hepatotoxic, carcinogenic, immunosuppressive, and

Table 14.1 Classification of food mycotoxins as potential human carcinogens

Group	Classification of food mycotoxins
1	Aflatoxin B1, B2, G1, G2
2A	-
2B	Aflatoxin M1, ochratoxin A, sterigmatocystin
3	Citrinin, patulin, luteoskyrin, cyclochlorotine, deoxynivalenol
4	-

Group 1, carcinogenic to humans; Group 2A, probably carcinogenic to humans; Group 2B, possibly carcinogenic to humans; Group 3, not classifiable as to its carcinogenicity to humans; Group 4, probably not carcinogenic to humans (classified by the International Agency for Research on Cancer)

antinutritional contaminant of many staple food commodities. Mutation of the P53 gene is considered as a key event in aflatoxin-induced carcinogenesis [10, 13, 14]. The endo-epoxide primarily binds to cellular proteins and is associated with direct cytotoxicity and the impairment of liver function. AFB1 is classified as a group 1 carcinogen by the International Agency for Research on Cancer (IARC) (Table 14.1) [15]. Epidemiological evidence suggest a synergistic effect of AFB1 and chronic hepatitis B (HBV) infections in the prevalence of liver cancer in humans [11, 13, 16]. Animal studies show that AFB1 also interferes with vitamins A, D, B12, iron, selenium, and zinc nutrition [10, 13, 17].

Another important hepatic metabolite is aflatoxin M1 (AFM1), which is excreted into milk both in animals and in humans. This results in an undesirable exposure of infants. AFM1 is less biologically active than AFB1 but can also be converted into epoxide forms that lead to hepatotoxicity and hepatocarcinogenicity [9, 12, 13]. IARC has classified AFM1 as a group 2B carcinogen (possibly carcinogenic to humans) (Table 14.1) [15].

Trichothecenes are a family of nearly 150 structurally related compounds produced by several fungal genera: *Fusarium, Cephalosporium, Myrothecium, Stachybotrys, Trichoderma*, and others [18]. The most common trichothecene mycotoxin is deoxynivalenol (DON, sometimes referred to as vomitoxin), which is common contaminant of corn and wheat in Europe [18, 19].

Fumonisins are mycotoxins produced by *Fusarium verticillioides* [19–21]. There is evidence linking *F. verticillioides* infected corn to the high incidence of human esophageal cancer in South Africa and China [19, 21–23]. *F. verticillioides* is an almost universal inhabitant of corn [19, 21, 22, 24].

Ochratoxins are a group of structurally similar metabolites commonly produced by *Aspergillus ochraceus* and *Penicillium verrucosum*. The major mycotoxin in this group is OTA. OTA has been suggested as the etiological factor of the Balkan endemic nephropathy (BEN) [18].

Ergot alkaloids are produced by several species of *Claviceps* that infect grains or grain products. These mycotoxins are involved in either nervous or gangrenous syndromes in humans. Ergotism is one of the oldest mycotoxicoses known, although occurrence of the disease has declined over time [2–4].

14.3.1.2 Minor Classes of Mycotoxins

Cyclopiazonic acid is mycotoxin isolated from molds commonly found on agricultural commodities or used in fermented food production (*Aspergillus flavus*, *Aspergillus versicolor*, *Aspergillus tamarii*, and several *Penicillium* species used in the production of fermented sausages in Europe). This includes *Penicillium camemberti*, used in the production of Camembert cheese, and *Aspergillus oryzae*, used in the production of soy sauce in the Far East. This mycotoxin has been shown to occur naturally in corn, cheese, peanuts, and sunflower seeds [2, 7].

Sterigmatocystin is produced by several species of *Aspergillus* and *Bipolaris* and by *Penicillium luteum*. Sterigmatocystin has been detected at low concentrations in green coffee, moldy wheat, and in the rind of hard cheese [2, 7].

Citrinin is a yellow mycotoxin produced by several *Penicillium* and *Aspergillus* species, including *Penicillium verrucosum* strains that produce OTA. Like OTA, citrinin causes kidney damage in laboratory animals [2, 7].

Patulin is produced primarily by species of *Penicillium* and *Aspergillus*, and its limited toxic properties are of major concern because it occurs

in apples and, subsequently, applesauce and juice [2, 7].

Citreoviridin originally was isolated from molds obtained from rice associated with a disease called "cardiac beriberi" that has occurred for three centuries in Japan (2). This mycotoxin also occurs in corn and other foods and feeds infected by some species of *Penicillium* and *Aspergillus* [2, 7].

Fusaproliferin has been detected in corn samples infected by *Fusarium proliferatum*, and *Fusarium subglutinans* that have been tested are capable of fusaproliferin production in culture [19, 21]. Although the significance of this compound in animal and human health remains to be determined, its acute toxicity to brine shrimp exceeds that of more familiar compounds such as the fumonisins.

14.3.2 Mycotoxin-Producing Fungi

Mycotoxins are produced by a wide array of diverse fungal species that generally are not aggressive pathogens. Most of the mycotoxins that are considered to be important are produced primarily by three genera of fungi: *Aspergillus*, *Penicillium*, and *Fusarium*. *Claviceps* and *Stachybotrys* also are important producers of mycotoxins [7, 10, 12, 22, 24].

Within the genus *Aspergillus*, the major class of mycotoxins is the aflatoxins. The crops most usually affected are corn, cotton, peanuts, and certain tree nuts. Although not conclusive in all crops, high temperatures seem to play a role in aflatoxin contamination [10, 14].

Within the genus *Fusarium*, there are a number of important mycotoxin-producing species. The major causative agents are *Fusarium graminearum*, *F. verticillioides*, *F. proliferatum*, and *F. subglutinans*. These latter agents may produce fumonisins during the pathogenic state in corn. *Fusarium graminearum* is a significant pathogen on wheat, barley, corn, and oats and is a major producer of DON in these grains [19, 21, 24].

Penicillium spp. are more typically associated with storage of crops and the production of

Disease	Substrate	Etiologic agent (spp.)	Ref.
Akakabio-byo	Wheat, barley, oats, rice	Fusarium	[8]
Alimentary toxic aleukia (septic angina)	Cereal grains (toxic bread)	Fusarium	[19]
Balkan endemic nephropathy	Cereal grains	Penicillium	[41]
Cardiac beriberi	Rice	Aspergillus, Penicillium	
Dendrodochiotoxicosis	Fodder (skin, inhalation)	Dendrodochium toxicum	
Ergotism	Rye, cereal grains	Claviceps purpurea	[4]
Esophageal tumors	Corn	Fusarium moniliforme	[20]
Hepatocarcinoma (acute aflatoxicosis)	Cereal grains, peanuts	Aspergillus flavus, A. parasiticus	[37–39]
Kashin-Beck disease, "Urov disease"	Cereal grains	Fusarium spp.	[8]
Kwashiorkor	Cereal grains	Aspergillus flavus, A. parasiticus	[32]
Onyalai	Millet	Phoma sorghina	[7]
Reye's syndrome	Cereal grains	Aspergillus spp.	[60]
Stachybotryotoxicosis	Hay, cereal grains, fodder (skin contact, inhalation)	Stachybotrys atra	[25]

Table 14.2 Human diseases in which analytic and/or epidemiologic data suggest or implicate mycotoxin involvement

mycotoxins such as OTA. OTA usually is formed in storage or during drying of certain commodities for processing [22, 23].

A number of fungi are capable of producing toxic alkaloids, and *Claviceps* spp. are the most notable in this regard. Toxic alkaloids also are produced by the genera *Epichloe* and *Neotyphodium*, both of which can be endophytic in certain plant species such as fescue and rye-grass [2].

Stachybotrys is a cellulolytic saprophyte that can be found in a variety of commodities, and the trichothecene metabolites of this organism can produce disease similar to some of those produced by *Fusarium* spp. Recently, this organism seemed to be involved in human disease where building materials were contaminated with the organism and possibly its toxic metabolites [25].

14.3.3 Impact of Mycotoxins on Human Diseases

Mycotoxicosis, the disease resulting from exposure to a mycotoxin, may be manifested as acute to chronic and ranges from rapid death to tumor formation [6, 9]. More occult disease may occur when the mycotoxin interferes with immune processes, rendering the patient more susceptible to infectious diseases. Humans likely are exposed to mycotoxins through several routes such as ingestion by contaminated food (the most prominent means of exposure), contact, and inhalation [2, 6,9]. There are many diseases that have been described in humans for which either analytic or epidemiologic evidence implicates a mycotoxin etiology (Table 14.2) [9–11, 14]. The aflatoxins are known causes of acute aflatoxicosis in humans, as well as potential cofactor of hepatic carcinoma, together with HBV [11, 16]. OTA has been conjecturally associated with the Balkan endemic nephropathy (BEN). The rural populations in the Balkans have a high incidence of chronic kidney problems and tumors of the excretory organ system [18, 42–44]. The most likely toxic products of the Fusarium spp., the trichothecenes, are conjecturally associated with the most prominent disease described alimentary toxic aleukia. The trichothecene, DON, is capable of producing a disease in mice that is similar in histological descriptions to human glomerulonephropathy [18, 22, 23]. Stachybotryotoxicosis is a disease that occurs in humans and is suspected to be caused by toxins of the organism Stachybotrys chartarum [25]. The fumonisins have been associated with esophageal cancer in

certain human populations [22]. Of interest to many investigators is that a number of mycotoxins are immunosuppressive and likely could be involved in human disease. Today, there is an apparent growing concern within the medical community regarding mycotoxin involvement in human diseases [6, 9]. The following section discusses mycotoxicoses for which there is considerable evidence for involvement of a specific mycotoxin(s). These and other human diseases where mycotoxin involvement is likely are presented in Table 14.2.

14.3.3.1 Acute Illness and Death

The symptoms of severe mycotoxicosis include hemorrhagic necrosis of the liver, bile duct proliferation, edema, and lethargy. Adult humans usually have a high tolerance of aflatoxin, and, in the reported acute poisonings, the children are those who die [26]. Acute poisonings, about 25% of which result in death, occur as a result of high levels of exposure [26, 27]. Reports of death and serious illness usually originate from developing countries within the zone of risk [28, 29].

14.3.3.2 Chronic Effects of Mycotoxins in Human Populations

In many regions of the world, dietary staples, especially cereal grains, contain low levels of mycotoxins. The impact of regular low-level intake of mycotoxins on human health is likely to be significant with a number of possible consequences including impaired growth and development, immune dysfunction, and the disease consequences of alterations in DNA metabolism [19].

14.3.3.3 Growth and Development

Numerous animal studies have shown that one of the first effects of mycotoxin ingestion is reduced feed intake and growth [22, 30]. There are facts revealing a very strong association between exposure to aflatoxin in the children and both stunting and being underweight. Both conditions reflect significant malnutrition and exposure of the children to aflatoxin in utero and subsequently after birth [30]. In West Africa, the aflatoxin exposure of children between 9 months and 5 years of age examined their growth, development, and height [30]. The children were also coexposed to a number of infectious diseases, and it is likely that the exposure to disease and aflatoxin would significantly compromise growth and development through reduced food intake and also the repartitioning of nutrients to maintain an upregulated immune system and away from growth and development [31, 32].

14.3.3.4 Immunosuppression

Immunosuppression is a likely major economic effect of mycotoxins. Mycotoxins known to have this effect are aflatoxins, certain trichothecenes, OTA, and gliotoxin. These mycotoxins can be immunotoxic and exert effects on cellular responses, humoral factors, and cytokine mediators of the immune system and can cause a variety of immune-related changes, including thymic aplasia and inhibition of phagocytosis by macrophages, delayed cutaneous hypersensitivity, lymphocyte proliferation, and leukocyte migration [23, 33, 34]. The effects on immunity and resistance are often difficult to recognize in the field because signs of disease are associated with the infection rather than the toxin that predisposed the individual to infection through decreased resistance and/or reduced vaccine or drug efficacy. There are evidences who reveal strong association between aflatoxin exposure and reduced immunocompetence in children [35] and adults [36], suggesting that aflatoxin ingestion decreases resistance to infection in human populations. DON can both stimulate and suppress the immune system, with dysregulation of IgA and the development of kidney disease in animal models that closely resembles human glomerulonephritis IgA nephropathy [33, 34].

14.3.3.5 Carcinogenicity, Mutagenicity, and Teratogenicity

Several food-contaminating mycotoxins have been defined as harmful carcinogens by the IARC (Table 14.1), that is, DON/nivalenol, zearalenone, ochratoxin, fumonisins, and aflatoxins [2, 15]. There is significant body of evidence demonstrating human exposure in utero to a number of mycotoxins, but the relevance of this exposure to birth defects or impaired embryonic development has received relatively little attention [19]. Recent epidemiological investigations of human populations in Texas, China, Guatemala, and Southern Africa that rely on foods prepared from maize, which is often contaminated with fumonisins, found a significantly higher incidence of neural tube defects in babies [19].

14.3.3.6 Hepatocellular Cancer and Mycotoxins

Two major factors, aflatoxin and HBV, which commonly occur in the same populations, influence the risk of liver cancer. Independently, each factor significantly increases the relative risk of cancer, and most studies report them, together, to be synergistic [37, 38]. The suggested mechanism for this synergy is that aflatoxin suppresses DNA repair mechanisms that help limit the development of hepatocellular cancer from HBV, and HBV prevents detoxification, but it is also possible that the immunotoxicity of aflatoxin interferes with the suppression of cancer. AFB1 is the most wellknown bioaccumulative toxin with strong mutagenic effect, involved in the development of hepatocellular cancer [37–39]. When individuals are exposed to AFB1 for a long time, monooxygenases produce reactive epoxide in the liver, leading to formation of toxic derivatives with nucleic acids and proteins [39, 40].

14.3.3.7 Mycotoxins and Balkan Endemic Nephropathy

There are convincing evidence that chronic poisoning with OTA is possible causative agents of BEN [41–44]. OTA, produced primarily by *Aspergillus ochraceous* or *Penicillium verrucosum*, occurs on several commodities prevalent in human diets, including barley and green coffee beans [42, 44]. OTA-mediated nephropathy is endemic, and outbreaks have been associated with weather conditions [43, 44]. This mycotoxin is considered a possibly carcinogenic to humans particular in the role of developing BEN-associated cancer (Table 14.1). However, the mechanism of OTA-derived tumor formation is unknown, and conflicting results regarding the potential of OTA to react with DNA to form covalent DNA adducts have been reported [45, 46].

14.3.3.8 Mycotoxins and Gastrointestinal Infections

The gastrointestinal (GI) tract, as the primary targeting organ, is exposed directly to mycotoxins with a higher concentration than other tissues and organs, which can affect the regeneration, proliferation, differentiation, and repair of intestinal epithelial cells. Some mycotoxins, such as AFB1, OTA, and DON, could reduce the expression of zonula occludens protein and increase intestinal mucosal permeability, thus damaging the barrier functions of intestinal epithelial cells and inducing bacterial translocation [21, 47]. Moreover, mycotoxins can influence the immune system of the GI tract [21, 48]. Mycotoxins could trigger mucosal immunoregulatory mechanisms such as the secretion of mucus, antimicrobial peptides, and immunoglobulins and directly induce inflammation by inducing intestinal epithelial cells to secrete chemotactic factors and pro-inflammatory cytokines [49]. Mycotoxins may be associated with chronic inflammation of the intestine in genetically susceptible patients with inflammatory bowel disease or coeliac disease [5, 47]. The long-term exposure of trichothecene toxins can increase intestinal colonization by aerobes [48]. The definite disturbance of human GI fungal and bacterial microbiota induced by mycotoxins still remains largely unknown.

14.3.3.9 Mycotoxins and Airway Diseases

Inhalation of mycotoxins is especially hazardous to those living inside damp, wet, and moldy buildings [50]. These toxins, mostly produced by fungi *Stachybotrys*, elicit recruitment of alveolar macrophages and neutrophils, pulmonary hemorrhage, and cytokine production and could trigger chronic obstructive pulmonary disease, while inhalation of a toxic dose of mycotoxin leads to systemic effects exclusive of lung injury [51, 52]. The most frequently recovered mycotoxins in nasal washings of individuals with respiratory diseases living in water-damaged buildings (WDB) are macrocyclic trichothecenes, found in 44% of the nasal washing specimens, whereas aflatoxin is present in 17% of these cases. On the other hand, mycotoxins were not found in nasal washings of a healthy control population. Other positive findings for the presence of mycotoxins in various tissues include trichothecenes in sera, breast milk, placenta, umbilical cord, and tissues (sinus) of individuals exposed in WDB [53, 54]. Clinical symptoms expressed by individuals living in WDB contaminated by Stachybotrys, mostly S. atra and S. chartarum, are pulmonary irritation and headaches, fatigue, malaise, diarrhea, inflammation of the nose, chest pain, or leukopenia [54, 55]. Strategy for prevention of this mycotoxicosis is using of polycarbonate membrane filters that could retain airborne particles of trichothecenes contaminated materials [50].

14.3.3.10 Mycotoxins and Deficiency of Vitamin B12

There are several pieces of evidence to show that mycotoxins affect cellular activities of the brain for which vitamin B12 plays a major role. Vitamin B12 possibly interacts with mycotoxins to effect some of the biochemical and neurological changes [56, 57]. It is believed that the biochemical consequences of fumonisin disruption of sphingolipid metabolism are increased free sphingoid bases and their 1-phosphates, alterations in complex sphingolipids, and decreased ceramide biosynthesis [24]. It is not yet clear whether vitamin B12 deficiency precludes fungal infection or vice versa. However, the factors commonly believed to predispose to recurrent chronic fungal infections included deficiency in whole blood folate, iron, and vitamin B complex [57]. These diseases occur as result of metabolic disorders and are due to inactivity of enzymes that are characteristic of vitamin B complex deficiencies (Table 14.3) [58].

Table	14.3	Signs,	symptoms,	and	clinical	neurologic
indicat	tions o	f vitami	n B12 defici	iency		

Signs and		
symptoms	Clinical neurological indications	
Headache,	Nerve damage and demyelination	
fatigue, loss of		
appetite		
Pinky-red sore	Degeneration of perip heral	
or smooth	nervous system leading to	
tongue	paralysis	
Growth failure	Progressive peripheral neuropathy	
in children		
Psychosis,	Spinal degeneration and	
confusion	macrocytic cells	
Depression,	Alzheimer's disease, allergies,	
memory loss	asthma	
Anxiety,	Crohn's disease, multiple	
insomnia	sclerosis, insomnia, sciatic	
	neuritis, trigeminal neuralgia,	
	osteoarthritis	

14.4 Macrofungi

14.4.1 Overview

Macrofungi, well known as mushrooms, are diverse group of the visible fruit of fungi, with known toxic effect on human [59, 61-63]. Approximately 100 of the known species of mushrooms are poisonous to humans, with new toxic species continually being identified [65, 66]. The geographical distribution of toxic mushrooms as well as toxicity of different mushrooms within the same genus may vary greatly (Table 14.4). Some mushrooms initially classified as edible have recently been reclassified as toxic [59]. Mushroom poisoning, termed mycetism or mycetismus, most commonly ensues after mushrooms are foraged, misidentified, and then consumed [64-66]. Worldwide, hundreds of mushroom poisonings are fatal each year [59, 65-67]. Most fatalities are secondary to Amanita phalloides ingestions [63, 65–67].

Some toxic mushroom ingestions will produce self-limited toxicity. Others will prove fatal. Generally, most toxic mushrooms produce some GI distress, and commonly GI symptoms are the first symptoms reported [59, 65]. A latency of <6 h to development of GI symptoms was utilized to predict that a mushroom ingestion should produce limited toxicity. This practice has limitations, and these limitations have become more apparent as new toxic mushrooms are recognized and more cases are reported. A study from Turkey reported 317 cases of mycetism, without delineating which mushrooms produced toxicity, and found that common symptoms include nausea vomiting (79.8%), and diarrhea (86.6%), (21.1%). More than 20% of patients who eventually developed hepatic failure had a latent phase of <6 h [62–65]. Similarly, there are reports about severe mushroom-induced hepatitis, 33% with initial symptoms within <6 h of ingestion [59, 63]. Not all patients presenting with symptoms within 6 h of ingestion have a benign course [59, 62, 63, 65]. Interestingly, if A. phalloides ingestion is known, or strongly suspected, to have occurred and diarrhea develops in <8 h, the prognosis is poorer [63–65]. Conversely, GI symptoms occurring >6 h after ingestion remain concerning for a possible serious clinical course and possible fatality [59, 62, 64-67].

14.4.2 Neurotoxins

Neurotoxic mushrooms may produce cholinergic, epileptogenic, inebriating, encephalopathic, or hallucinogenic syndromes [59, 63]. Many of these syndromes have associated visual disturbances. Some contain toxins that affect vasculature and are classified under neurovascular toxins (Table 14.4).

14.4.2.1 Cholinergic Syndromes

Some mushrooms contain muscarine that stimulates peripheral muscarinic receptors. Muscarine acts like acetylcholine, but is not degraded by cholinesterase, and therefore has a longer duration of action [63]. The amount of muscarine in *Amanita muscaria* is very low, and muscarinic symptoms are rarely seen after its consumption [59, 65]. *Inocybe*, *Clitocybe*, *Boletus*, and *Rubinoboletus* species can contain sufficient muscarine to produce muscarinic toxicity. Onset of toxicity is 15 min to 5 h after ingestion. Flushing, vasodilation, diaphoresis, lacrimation,

Table 14.4 Geographical distribution of recently described toxic mushroom

Primary system affected by toxins	Geographical location of	
(effects)	mushroom harvest ^a	Mushrooms
Neurological (encephalopathy)	Germany	Hapalopilus rutilans (purple dye polypore)
Neurological (convulsive encephalopathy)	Japan	Pleurocybella porrigens (angel's wing)
Neurovascular (red, swollen, painful extremities)	Japan, South Korea	Clitocybe acromelalgia
Neurovascular (red, swollen,	France, Italy, Morocco	Clitocybe amoenolens (poison dwarf
painful extremities)		bamboo or burn mushroom)
Cardiac (sudden death)	Yunnan Province, China	Trogia venenata (little white)
Cardiac (sudden death)	Jiangxi Province, China	Amanita franchetii, Ramaria rufescens
Renal	Canada (South-West), USA	Amanita smithiana (toxic lepidella or North
	(Pacific Coast to North-West)	American lepidella)
Renal	France (South), Spain, Italy	Amanita proxima (Mediterranean Amidella)
Renal	Japan	Amanita pseudoporphyria Hongo
Muscular (rhabdomyolysis, myocarditis)	France, Poland	<i>Tricholoma equestre</i> (yellow trich or yellow knight or man on horseback)
Muscular (rhabdomyolysis, myocarditis)	Taiwan, Japan, Korea, China, Nepal	Russula subnigricans (blackening russula)
Immune/heme (hemolysis,	Northern Hemisphere	Paxillus involutus (poison pax or brown roll
hepatorenal failure)		rim)
Immune/heme (pancytopenia)	Northeastern Asia	Ganoderma neojaponicum Imazeki ^b
Immune/heme (pancytopenia)	Japan, China, Korea, Java	Podostroma cornu-damae

^aCopyright: J. Med. Toxicol. (2014) 10;173-189; Reference: [59]

^bNewly reported and association not as well established as other causes of mycetism

miosis, blurred vision, hypersalivation, bronchorrhea, bronchospasm, vomiting, diarrhea, abdominal pain, tremor, restlessness, and bladder contraction can follow ingestion. Hallucinations, bradycardia or tachycardia, hypotension or hypertension, syncope, shock, and confusion are common [66, 67]. Treatment is supportive: fluid and electrolyte replacement, vasopressors, and atropine in case of bronchorrhea and bradycardia. Toxicity generally resolves within 12 h of ingestion [59, 63].

14.4.2.2 Epileptogenic Syndromes

The classic epileptogenic mushroom is Gyromitra esculenta (false morel), which may be confused with Morchella sp. (true morel, which is deemed edible but which can produce neurological symptoms if poorly cooked) or *Verpa bohemica* (early morel) [68]. Symptoms begin 4-12 h after ingestion. Clinical progression is vomiting and diarrhea followed by neurological symptoms (vertigo, ataxia, nystagmus, tremor, convulsions, and coma), sometimes followed by hepatic necrosis, jaundice, methemoglobinemia, hemolysis, and rhabdomyolysis [66–69]. Approximately 10% of poisoned patients die [70]. Treatment is supportive, while pyridoxine is recommended for neurologic symptoms including seizures with dosing regimen of 70 mg/kg up to 5 g, intravenously [66-**69**, **71**]. Seizures may be resistant to benzodiazepines and barbiturates prior to GABA repletion aided by pyridoxine treatment [69]. In Japan, 2004, convulsive encephalopathy occurred epidemically, with latency of 1-31 days, when patients with hemodialysisdependent renal failure ingested Pleurocybella porrigens (also called Nothopanus porrigens, Sugihiratake, and angel's wing). Patients exhibited convulsions (78%), myoclonus (47%), dysarthria (31%), ataxia (25%), and paresis or paralysis (22%) [72]. Intractable status epilepticus has also been reported [73–76]. Magnetic resonance imaging may reveal intracranial lesions involving the subcortical white matter of the insular cortex, claustrum, external capsule, putamen, and globus pallidus [76]. The mortality rate is approximately 30% [72, 73].

14.4.2.3 Encephalopathic Syndromes

In Germany, encephalopathy and hepatorenal insufficiency associated with purple urine have been reported following ingestion of Hapalopilus rutilans (Purple Dye Polypore) [59, 63, 65, 74]. The syndrome begins with nausea, vomiting, and abdominal pain approximately 12 h after ingestion. Hepatorenal laboratory abnormalities and neurological symptoms, such as vertigo, ataxia, drowsiness, hypotonia, and visual complaints, follow [66, 74]. P. porrigens, Grifola frondosa, and Pleurotus eryngii poisonings may be also presented with encephalopathy. Any mushroom that produces fulminant hepatic failure (e.g., A. phalloides) or that is epileptogenic (e.g., G. escu*lenta*) may be associated with encephalopathy [63, 69].

14.4.2.4 Hallucinogenic Syndromes

Hallucinogenic mushrooms generally contain psilocybin and include Psilocybe spp., Panaeolus spp., and Stropharia aeruginosa [64]. Psilocybin-containing species may turn bluish (bruise) upon handling especially the stalks; however, this is nonspecific, as some other species that do not contain psilocybin bruise similarly [64]. Symptoms reflected in altered time-space perceptions, auditory, and visual hallucinations begin 15-30 min after ingestion and generally last up to 6 h and rarely last 12 h [66]. Mydriasis, hypertension, tachycardia, dysrhythmias, and myocardial infarction have also been reported [59, 63, 75]. Seizures and hyperthermia have been reported in children [75]. Treatment is supportive. Benzodiazepine and barbiturates have been used to control agitation and seizures [69, 75].

14.4.2.5 Neurovascular Toxins

Clitocybe amoenolens (burn mushroom) and *Clitocybe acromelalga* are associated with erythromelalgia. Erythromelalgia involves erythema, swelling, and pain of the distal extremities. Ingestion of only a few mushrooms can produce toxicity. In humans, symptoms appear 24 h after ingestion, and patients present with paresthesia of the digits followed by paroxysmal burning pain [63, 74]. Paroxysmal dilation of the blood vessels of the skin occurs, what is associated with tactile and burning pain and red, swollen hands and feet. Heat and a position decline induced paroxysmal crisis, which generally occurs nocturnally. Cyanosis or erythema may worsen during pain paroxysms. More severe cases are associated with local diaphoresis and trophic changes of the digits. The pain can be incapacitating. Dipping the extremities in cold water often provides relief, while traditional analgesics may be ineffective. The syndrome can last for weeks to months [59, 63, 69, 74].

14.4.2.6 Cardiotoxins and Sudden Death

A seasonal illness, called "Yunnan" sudden unexplained death, has been epidemiologically traced to *Trogia venenata* (little white) consumption. The exact toxin responsible for the syndrome is not yet known [77, 78]. Patients report nausea, vomiting, diarrhea, abdominal pain, and fatigue, while recurrent syncope and sudden unexpected death may occur. Many patients report palpitations, chest discomfort, dizziness, syncope, seizures, ventricular tachycardia, fibrillation, and elevated plasma enzymes in the hours preceding death [77, 78]. Postmortem examination revealed

focal lymphocytic myocarditis, with breakage of the muscle fibers, lymphocytic infiltration of the liver, pulmonary alveolar edema, acute kidney necrosis, hepatocyte necrosis, and congestion of the liver, lung, or spleen [78].

14.4.2.7 Hepatotoxins

Amatoxins and gyromitrin are protoplasmic poisons that produce hepatotoxicity [77]. Many *Amanita* spp. contain amatoxin, in the first place *A. phalloides* (death cap), as well as some *Galerina* and *Lepiota* (e.g., *L. brunneoincarnata* and *L. helveola*). As little as 0.1 mg amatoxin/kg body weight may be lethal in adults [67, 77, 79].

In general, supportive care is the mainstay for mycetism [59, 79]. If a patient is actively vomiting, activated charcoal may not contribute to decontamination. Antiemetics may limit natural decontamination. Determining what mushrooms are likely to have been ingested, and therefore what treatments are most appropriate, will likely be based on the geographic location of mushroom harvesting (see Table 14.5) [63]. If a hepatotoxic mushroom, such as *A. phalloides*, has been ingested or is suspected, prompt treatment with an antidote seems prudent (Table 14.5) [67, 79].

Antidote	Mechanism of action ^a	Dose
Silibinin ^b	Competes with amatoxins for transmembrane transport; inhibits penetration of amanitin into hepatocytes; scavenges free radicals; produces anti-inflammatory effects; increases ribosomal RNA synthesis, increases protein synthesis	5 mg/kg IV over 1 h; then 20 mg/kg/24 h continuous infusion (diluted in 5% glucose to a concentration of 2 mg silibinin/ml) for 3 days (alternatively, 20 mg/kg/24 h can be divided into 6 doses)
<i>N</i> -acetylcysteine (NAC)	Promotes glutathione regeneration; scavenges free radicals	Follow dosing for APAP treatment: 150 mg/kg IV over 1 h, followed by 12.5 mg/kg/h for 4 h, followed by 6.25 mg/kg/h until hepatic failure resolution

Table 14.5 Antidotes for amatoxin in poisoning (recommended doses based on previously published reports)

^aRetrieved from: J. Med. Toxicol. (2014) 10;173-189; Reference: [59]

^bSilibinin is not approved as a therapeutic treatment for hepatic disease by the Food and Drug Administration in the USA; it is approved and available in Europe and Australia

References

- Forgacs J, Carll WT (1962) Mycotoxicoses. Adv Vet Sci 7:273–382
- Council of Agricultural Science & Technology (CAST) (2003) Mycotoxins: risks in plant, animal and human systems. task force report No. 139. Council of Agricultural Science & Technology, Iowa, USA
- Matossian MK (1989) Poisons of the past: molds, epidemics, and history. Yale University Press, New Haven, p 190
- Van Rensburg SJ, Altenkirk B (1974) Claviceps purpurea: ergotism. In: IFH Purchase (ed) Mycotoxins. Elsevier, New York, pp 69–96
- Alonso VA, Pereyra CM, Keller LA et al (2013) Fungi and mycotoxins in silage: an overview. J Appl Microbiol 115(3):637–643
- Paterson RR, Lima N (2010) Toxicology of mycotoxins. EXS 100:31–63
- Wambacq E, Vanhoutte I, Audenaert K, De Gelder L, Haesaert G (2016) Occurrence, prevention and remediation of toxigenic fungi and mycotoxins in silage: a review. J Sci Food Agric 96(7):2284–2302
- Bryden WL (2007) Mycotoxins in the food chain: human health implications. Asia Pac J Clin Nutr 16(1):95–101
- Marin S, Ramos AJ, Cano-Sancho G, Sanchis V (2013) Mycotoxins: occurrence, toxicology, and exposure assessment. Food Chem Toxicol 60:218–237
- Ahmad M, Ahmad MM, Hamid R, Abdin MZ, Javed S (2013) Use of response surface methodology to study the effect of media composition on aflatoxin production by Aspergillus flavus. Mycotoxin Res 29(1):39–45
- Wu HC, Santella R (2012) The role of aflatoxins in hepatocellular carcinoma. Hepat Mon 12(10):e7238
- Leong YH, Latiff AA, Ahmad NI, Rosma A (2012) Exposure measurement of aflatoxins and aflatoxin metabolites in human body fluids. A short review. Mycotoxin Res 28(2):79–87
- Bedard LL, Massey TE (2006) Aflatoxin B1-induced DNA damage and its repair. Cancer Lett 241(2):174–183
- Wild CP, Turner PC (2002) The toxicology of aflatoxins as a basis for public health decisions. Mutagenesis 17(6):471–481
- IARC Working Group on the Evaluation of Carcinogenic Risks to Humans (2002) Some traditional herbal medicines, some mycotoxins, naphthalene and styrene. IARC Monogr Eval Carcinog Risks Hum 82:1–556
- Nordenstedt H, White DL, El-Serag HB (2010) The changing pattern of epidemiology in hepatocellular carcinoma. Dig Liver Dis 42(3):S206–S214

- Anyanwu EC, Kanu I (2007) Biochemical impedance on intracellular functions of vitamin B12 in chronic toxigenic mold exposures. Sci World J 7:1649–1657
- Staneva R, Rukova B, Hadjidekova S et al (2013) Whole genome methylation array analysis reveals new aspects in Balkan endemic nephropathy etiology. BMC Nephrol 14:225
- Voss KA, Gelineau-vanWaes JB, Riley RT (2006) Fumonisins: current research trends in developmental toxicology. Mycotoxin Res 22:61–69
- Yoshizawa T, Yamashita A, Luo Y (1994) Fumonisin occurrence in corn from high- and low-risk areas for human esophageal cancer in China. Appl Environ Microbiol 60(5):1626–1629
- Bouhet S, Hourcade E, Loiseau N et al (2004) The mycotoxin fumonisin B1 alters the proliferation and the barrier function of porcine intestinal epithelial cells. Toxicol Sci 77:165–171
- 22. Dersjant-Li Y, Verstegen MWA, Gerrits WJJ (2003) The impact of low concentrations of aflatoxin, deoxynivalenol or fumonisins in diets on growing pigs and poultry. Nutr Res Rev 16:223–239
- Pestka JJ, Zhou RR, Moon Y, Chung YJ (2004) Cellular and molecular mechanisms for immune modulation by deoxynivalenol and other tricothecenes; unraveling a paradox. Toxicol Lett 153:61–73
- Riley RT, Enongene E, Voss KA et al (2001) Sphingolipid perturbations as mechanisms for fumonisin carcinogenesis. Environ Health Perspect 109(2):301–308
- Kuhn DM, Ghannoum MA (2003) Indoor mold, toxigenic fungi, and Stachybotrys chartarum: infectious disease perspective. Clin Microbiol Rev 16(1):144–172
- 26. Cullen JM, Newberne PM (1993) Acute hepatotoxicity of aflatoxins. In: Eaton DL, Groopman JD (eds) The toxicology of aflatoxins: human health, veterinary, and agricultural significance. Academic, London, pp 1–26
- Eaton D, Ramsdell HS, Neal G (1993) Biotransformation of aflatoxins. In: Eaton D, Groopman JD (eds) The toxicology of aflatoxins: human health, veterinary, and agricultural significance. Academic, London, pp 45–72
- Vargas EA, Preis RA, Castro L, Silva CM (2001) Co-occurrence of aflatoxins B1, B2, G1, G2, zearalenone and fumonisin B1 in Brazilian corn. Food Addit Contam 18:981–986
- Henry SH, Bosch FX, Bowers JC (2002) Aflatoxin, hepatitis and worldwide liver cancer risks. Adv Exp Med Biol 504:229–233
- Gong Y, Cardwell K, Hounsa A et al (2002) Dietary aflatoxin exposure and impaired growth in children from Beninand Togo: Cross sectional study. Br Med J 325:20–21

- 31. Elsasser TJ, Klasing KC, Filipov N, Thompson F (2000) The metabolic consequences of stress: targets for stress and priorities of nutrient use. In: Moberg GP, Mench JA (eds) The biology of animal stress. CABI Publishing, New York, pp 77–110
- Hendrickse RC (1991) Kwashiokor: the hypothesis that incriminates aflatoxin. Paediatrics 88:376–379
- Oswald IP, Marin DE, Bouhet S, Pinton P, Taranu I, Accensi F (2005) Immunotoxicological risk of mycotoxins for domestic animals. Food Addit Contam 22:354–360
- Bondy G, Pestka JJ (2000) Immunomodulation by fungal toxins. J Toxicol Environ Health 3:109–143
- 35. Turner PC, Moore SE, Hall AJ, Prentice AM, Wild CP (2003) Modification of immune function through exposure to dietary aflatoxin in Gambian children. Environ Health Perspect 111:217–220
- 36. Jiang YI, Jolly PE, Ellis WO, Wang JS, Phillips TD, Williams JH (2005) AflatoxinB1 albumin adduct levels and cellular immune status in Ghanaians. Int Immunol 17:807–814
- Chuang SC, La Vecchia C, Boffetta P (2009) Liver cancer: descriptive epidemiology and risk factors other than HBV and HCV infection. Cancer Lett 286:9–14
- Llovet JM, Burroughs A, Bruix J (2003) Hepatocellular carcinoma. Lancet 362:1907–1917
- Gomaa AI, Khan SA, Toledano MB, Waked I, Taylor-Robinson SD (2008) Hepatocellular carcinoma: epidemiology, risk factors and pathogenesis. World J Gastroenterol 14:4300–4308
- Matsuda Y, Ichida T, Fukumoto M (2011) Hepatocellular carcinoma and liver transplantation: clinical perspective on molecular targeted strategies. Med Mol Morphol 44:117–124
- Stiborova M, Arlt VM, Schmeiser HN (2016) Balkan endemic nephropathy: an update on its aetiology. Arch Toxicol 90(11):2595–2615
- Anandagoda N, Lord GM (2015) Preventing aristolochic acid nephropathy. Clin J Am Soc Nephrol 10:167–168
- 43. Schmeiser HH, Kucab JE, Arlt VM et al (2012) Evidence of exposure to aristolochic acid in patients with urothelial cancer from a Balkan endemic nephropathy region of Romania. Environ Mol Mutagen 53:636–641
- 44. Stefanovic V, Toncheva D, Polenakovic M (2015) Balkan nephropathy. Clin Nephrol 83(1):64–69
- 45. Pfohl-Leszkowicz A (2009) Ochratoxin A and aristolochic acid involvement in nephropathies and associated urothelial tract tumours. Arh Hig Rada Toksikol 60:465–483
- 46. Mantle PG, Faucet-Marquis V, Manderville RA, Squillaci B, Pfohl-Leszkowicz A (2010) Structures of covalent adducts between DNA and ochratoxin A: a new factor in debate about genotoxicity and human risk assessment. Chem Res Toxicol 23:89–98
- 47. Pinton P, Braicu C, Nougayrede JP, Laffitte J, Taranu I, Oswald IP (2010) Deoxynivalenol impairs porcine intestinal barrier function and decreases the protein expression of claudin-4 through a mitogen

activated protein kinase-dependent mechanism. J Nutr 140:1956–1962

- Grenier B, Applegate TJ (2013) Modulation of intestinal functions following mycotoxin ingestion: metaanalysis of published experiments in animals. Toxins 5:396–430
- Maresca M, Fantini J (2010) Some food associated mycotoxins as potential risk factors in humans predisposed to chronic intestinal inflammatory diseases. Toxicon 56:282–294
- Eduard W (2009) Fungal spores: a critical review of the toxicological and epidemiological evidence as a basis for occupational exposure limit setting. Crit Rev Toxicol 39(10):799–864
- 51. Lichtenstein JH, Molina RM, Donaghey TC et al (2010) Pulmonary responses to Stachybotrys chartarum and its toxins: mouse strain affects clearance and macrophage cytotoxicity. Toxicol Sci 116(1):113–121
- 52. Liu C, Shen H, Yi L et al (2015) Oral administration of aflatoxin G(1) induces chronic alveolar inflammation associated with lung tumorigenesis. Toxicol Lett 232(3):547–556
- 53. Thrasher JD, Gray MR, Kilburn KH, Dennis D, Yu A (2012) A water-damaged home and health of occupants: A case study. J Environ Public Health 2012:312836
- 54. Brasel TL, Campbell AW, Demers RE et al (2004) Detection of trichothecene mycotoxins in sera from individuals exposed to Stachybotrys chartarum in indoor environments. Arch Environ Health 59:317–323
- 55. Layton RC, Purdy CW, Jumper CA, Straus DC (2009) Detection of macrocyclic trichothecene mycotoxins in a caprine (goat) tracheal instillation model. Toxicol Ind Health 25:693–701
- 56. Enongene EN, Sharma RP, Bhandari N, Voss KA, Riley RT (2000) Disruption of sphingolipid metabolism in small intestines, liver and kidney of mice dosed subcutaneously with fumonisin B (1). Food Chem Toxicol 38(9):793–799
- Samaranayake LP (1986) Nutritional factors and oral candidosis. J Oral Pathol 15(2):61–65
- Hitzig WH (1983) Protean appearances of immunodeficiencies: syndromes and inborn errors involving other systems, which express associated primary immunodeficiency. Birth Defects 19(3):307–312
- Graeme KA (2014) Mycetism: a review of the recent literature. J Med Toxicol 10:173–189
- Rogan WJ, Yang GC, Kimbrough RD (1985) Aflatoxin and Reye's syndrome: a study of livers from deceased cases. Arch Environ Health 40(2):91–95
- 61. Kirchmair M, Carrilho P, Pfab R et al (2012) Amanita poisoning resulting in acute, reversible renal failure: new cases, new toxic Amanita mushrooms. Nephrol Dial Transplant 27:1380–1386
- 62. Mendez-Navarro J, Ortiz-Olivera NX, Villegas-Rios M et al (2011) Hepatotoxicity from ingestion of wild mushrooms of the genus Amanita section Phalloideae collected Mexico City: two case reports. Ann Hepatol 10(4):568–574

- French LK, Hendrickson RG, Horowitz BZ (2011) Amanita phalloides poisoning. Clin Toxicol 49:128–129
- Hawksworth DL, Wiltshire PEJ (2011) Forensic mycology: the use of fungi in criminal investigations. Forensic Sci Int 206:1–11
- Eren SH, Demirel Y, Ugurlu S, Korkmaz I, Aktas C, Guven FM (2010) Mushroom poisoning: retrospective analysis of 294 cases. Clinics 65(5):491–496
- 66. Lima ADL, Costa Fortes R, Carvalho Garbi Novaes MR, Percario S (2012) Poisonous mushrooms: a review of the most common intoxications. Nutr Hosp 27(2):402–408
- Lukasik-Glebocka M, Druzdz A, Naskret M (2011) Clinical symptoms and circumstances of acute poisoning with fly agaric (Amanita muscaria) and panther cap (Amanita pantherina). Przegl Lek 68(8):449–452
- Pauli JL, Foot CL (2005) Fatal muscarinic syndrome after eating wild mushrooms. Med J Aust 182:294–295
- Karlson-Stiber C, Persson H (2003) Cytotoxic fungian overview. Toxicon 2(4):339–349
- Michelot D, Toth B (1991) Poisoning by Gyromitra esculenta-a review. J Appl Toxicol 11(4):235–243
- Lheureux P, Penaloza A, Gris M (2005) Pyridoxine in clinical toxicology: a review. Eur J Emerg Med 12:78–85
- 72. Gejyo F, Homma N, Higuchi N et al (2005) A novel type of encephalopathy associated with mushroom Sugihiratake ingestion in patients with chronic kidney diseases. Kidney Int 68(1):188–192

- 73. Kuwabara T, Arai A, Honma N, Nishizawa M (2005) Acute encephalopathy among patients with renal dysfunction after ingestion of "sugihiratake", angel's wing mushroom—study on the incipient cases in the northern area of Niigata Prefecture. Rinsho Shinkeigaku 45(3):239–245
- Saviuc P, Danel V (2006) New syndromes in mushroom poisoning. Toxicol Rev 25(3):199–209
- Satora L, Goszcz H, Ciszowski K (2005) Poisonings resulting from the ingestion of magic mushrooms in Krakow. Przegl Lek 62(6):394–396
- 76. Kurokawa K, Sato H, Nakajima K, Kawanami T, Kato T (2005) Clinical, neuroimaging and electroencephalographic findings of encephalopathy occurring after the ingestion of "sugihiratake" (Pleurocybella porrigens), an autumn mushroom: a report of two cases. Rinsho Shinkeigaku 45(2):111–116
- 77. Zhang Y, Li Y, Wu G et al (2012) Evidence against barium in the mushroom Trogia venenata as a cause of sudden unexpected death in Yunnan, China. Appl Environ Microbiol 78(24):8834
- Shi GQ, Huang WL, Zhang J et al (2012) Clusters of sudden unexplained death associated with the mushroom, Trogia venenata, in rural Yunnan. PLoS ONE 7(5):e35894
- 79. Santi L, Maggioli C, Mastroroberto M, Tufoni M, Napoli L, Caraceni P (2012) Acute liver failure caused by Amanita phalloides poisoning. Int J Hepatol 2012:48748