Chapter 10 Adenosine Receptors as a Paradigm to Identify Dimer/Oligomers of G-Protein-Coupled Receptors and as Targets in Parkinson's Disease and Schizophrenia



Gemma Navarro, Dasiel O. Borroto-Escuela, Kiell Fuxe, and Rafael Franco

Abstract While adrenergic receptors were instrumental to start to understand the role of GPCRs, other receptors are taking the lead to understand why GPCR homo-/ heteromers are needed and to address their physiological consequences in both healthy/homeostatic conditions and disease. Adenosine and dopamine receptors in the CNS are instrumental to understand pathogenic mechanisms in Parkinson's disease and to know the role of receptor heteromers. We here provide the account of the heteroreceptor complexes formed by adenosine receptors (A₁, A_{2A}, A_{2B}, and A₃), and their potential as therapeutic targets. Both adenosine (A₁ or A_{2A})-dopamine (D₁ or D₂) and adenosine A₁A_{2A} heteroreceptor complexes are therapeutic targets in Parkinson's disease and may be altered after chronic levodopa treatment. A short account on the potential of adenosine receptors as targets in schizophrenia is also provided. Apart from potential in combating symptoms, adenosine receptors have potential as targets for neuroprotection. However, the design of neuroprotective drugs requires to understand how adenosine affects microglia and which adenosine receptor-containing heteromers may be targeted.

R. Franco (🖂)

G. Navarro

Department of Biochemistry and Physiology, Faculty of Pharmacy, University of Barcelona, Barcelona, Spain

Centro de Investigación en Red sobre Enfermedades Neurodegenerativas. CIBERNED. Instituto de Salud Carlos III, Madrid, Spain

D. O. Borroto-Escuela · K. Fuxe Department of Neuroscience, Karolinska Institutet, Stockholm, Sweden

Centro de Investigación en Red sobre Enfermedades Neurodegenerativas. CIBERNED. Instituto de Salud Carlos III, Madrid, Spain

Department of Biochemistry and Molecular Biomedicine, Faculty of Biology, University of Barcelona, Barcelona, Spain

[©] Springer Nature Switzerland AG 2018

P. A. Borea et al. (eds.), *The Adenosine Receptors*, The Receptors 34, https://doi.org/10.1007/978-3-319-90808-3_10

Keywords Adenosine receptors · Heteroreceptor complexes · Dopamine receptors · Parkinson's disease · Schizophrenia

10.1 Introduction

Adenosine and dopamine receptors have been instrumental in identifying complexes with other members of the Class A G-protein-coupled receptor (GPCR) superfamily. For review on dopamine receptor homo-/heteromerization and its relevance, see Rashid et al. (2007), Fuxe et al. (2014a, b), George et al. (2014), Perreault et al. (2014), Borroto-Escuela et al. (2016), Borroto-Escuela and Fuxe (2017) and references therein. Dimers were first identified using coimmunoprecipitation and other biochemical approaches. Later, biophysical techniques were implemented to detect dimers (even trimers) in heterologous expression systems. The existence of receptor-receptor interactions between different GPCRs in the plasma membrane in brain tissue was first indicated in biochemical binding studies on neuropeptide modulation of the affinity and density of monoamine receptor subtypes using monoamine radioligands and membrane preparations from different brain regions (Fuxe et al. 1981, 1983, 1987; Agnati et al. 1982; Fuxe and Agnati 1985). The results gave rise to the concept of direct interactions in the plasma membrane of subtype-specific neuropeptide receptor and monoamine receptors. In 1993, it was proposed that the molecular mechanism for these GPCR receptor-receptor interactions was represented by the formation of a heterodimer in balance with the corresponding homodimers/monomers (Zoli et al. 1993).

Franco et al. (2016) reviewed the strategies that may lead to demonstrate that heteroreceptor complexes formed by GPCR are present in natural sources; in particular, the two that have provided more benefit in our experience are (i) the heteromer print (something that is particular to the complex and does not happen in individually expressed receptors) and (ii) in situ proximity ligation assays, a technique developed for assessing cancer types in samples from patients and that allows to detect GPCR clusters in cells, in samples from animal models, in samples from patients, or in samples from necropsies. The central nervous system (CNS) has been by far the substrate for identifying the complexes formed by adenosine receptors. Actually, the periphery lacks behind the CNS in identifying and addressing the physiological role of GPCRs. Exceptions do occur, and the most straightforward example in the periphery is likely provided by chemokine receptors, which may form homo- and heterodimers that provide pharmacological and signaling diversity to cells of the immunological system (see (Springael et al. 2005; Muñoz et al. 2009, 2011, 2012) and references therein). Indeed, there is consensus in that a receptor heteromer (Het) cannot be considered as such in the absence of any particular property, i.e., a given complex in a natural context should display a particular heteromer print (Het) (Ferré et al. 2009a).

10.2 Adenosine Receptors in the Formation of Heteromers with Non-purinergic GPCRs

Except for error, omission, or very recent discovery, the direct interactions reported for adenosine receptors with other members of the GPCR superfamily are those described below.

The first Het identified for receptors having different endogenous agonists was that constituted by adenosine $A_1(A_1R)$ and dopamine D_1 (Gines et al. 2000; Torvinen et al. 2002; Cao et al. 2006). In parallel, the Het for two different subtypes of receptors for the same endogenous agonist (mu and delta opioid receptors) was discovered by Gomes et al. (2000). In brain regions related to motor control, functional adenosine-dopamine receptor interactions were known. Also known was the segregation of striatal D_1 and D_2 receptors in, respectively, the so-called direct and indirect pathways of motor control. It turns out that whereas D_1 and A_1 colocalize in striatonigral GABAergic neurons, adenosine A_{2A} ($A_{2A}R$) and dopamine D_2 receptors colocalize in striatopallidal GABAergic neurons. Accordingly, we hypothesized, and later demonstrated, that A_{2A}-D₂ heteromerization in the indirect pathway paralleled the A_1 - D_1 heterometrization in the direct pathway (Hillion et al. 2002; Canals et al. 2003, 2004; Fuxe et al. 2003, 2007; Ciruela et al. 2004). The highest $A_{2A}R$ expression in a mammalian body is found in the striatum, a fact whose extent is not fully known. Hence, interactions with other dopamine receptors, which are also expressed in motor control brain areas or with receptors widely distributed in the CNS, have been reported. On the one hand, the effect of activation of $A_{2A}R$ on in vivo actions mediated by dopamine D_3 (Hillefors et al. 1999) prompted us to investigate and identify A_{2A}-D₃ Hets (Torvinen et al. 2005). In vivo activation of A_{2A}Rs in the basal ganglia causes alterations in the pharmacological characteristics of dopamine D₃ receptors that may underlie the atypical neuroleptic-like effect of A_{2A}R receptor agonists (Rimondini et al. 1997; Hillefors et al. 1999); as a matter of speculation, those in vivo effects may be a consequence of the particular pharmacological and functional properties of A2A-D3Hets. On the other hand, striatal adenosine A_{2A}Rs form functional heteromeric complexes with cannabinoid CB₁ receptors (Carriba et al. 2007) or with histamine H₃ (Márquez-Gómez et al. 2018) receptors; these Hets may, respectively, mediate the motor effects of cannabinoids and deserve attention on assessing the potential of antihistamines in the therapy of CNS diseases. Due to the intrinsic structural and conformational properties of the Class C GPCR subfamily, they can form a myriad of homo- and heteroreceptor complexes (Doumazane et al. 2011; Borroto-Escuela et al. 2014). Interestingly, the A_{2A}R may form functional but also molecular complexes with Class C metabotropic mGlu₅ receptors (Ferré et al. 2002, 2003; Nishi et al. 2003; Kachroo 2005; Borroto-Escuela et al. 2017b). Ultrastructural studies have shown that the two receptors colocalize in the nonhuman primate striatum (Bogenpohl et al. 2012). Finally, the adenosine receptor is also able to interact with the orphan GPR37 receptor (Dunham et al. 2009). Pioneering evidence on functional interactions in rat caudate putamen

suggests that the adenosine receptors may also interact with some of the opioid receptor subtypes (Noble and Cox 1995; Borroto-Escuela et al. 2014).

 $A_{2A}R$ may form homodimers (Canals et al. 2004) that likely interact with other GPCRs to form high-order heteroreceptor complexes. One example is the Het formed by A_{2A} , cannabinoid CB₁, and dopamine D₂ (Carriba et al. 2007; Navarro et al. 2008; Bonaventura et al. 2014; Pinna et al. 2014a, b). Another is the complex formed by A_{2A} , D₂, and mGlu₅ receptors (Cabello et al. 2009).

Consistent with the intense research on potential heteromerization of adenosine receptors, it has been shown that β_1 - and β_2 -adrenergic receptors may directly interact with the A₁R and that the resulting Het displays particular properties in terms of differential pharmacology and coupling to the signaling machinery (Chandrasekera et al. 2013). Finally, it has been confirmed that prostanoid receptors, namely, the thromboxane A₂ TP receptor, may form hetero-oligomers with the A₁R whose functional properties are conditioned by the presence and concentration of the endogenous agonist of the two receptors (Mizuno et al. 2012, 2013a). Heteromerization has been also reported for A₁R and class C metabotropic glutamate 1 alpha (Ciruela et al. 2001; Franco et al. 2001).

For reasons that are out of the scope of the present chapter, the two most studied adenosine receptors, in terms of receptor-receptor interaction research, are the A_1 and the A_{2A} . The other two types of adenosine receptors (A_3 and A_{2B}) are lacking behind, but, interestingly, the first identified Hets containing A_3 or A_{2B} are between adenosine receptors themselves (see next Sect. 10.3).

10.3 Adenosine Receptors May Interact with Other P1 (to Form Adenosine Isoreceptor Complexes) and with P2 Purinergic Receptors

Soon after the experimental confirmation of GPCR heteromerization and the extensive work made with A_1 and A_{2A} receptors, it was tempting to search for interaction between adenosine, i.e., P1 purinergic receptors, and "ATP" P2 purinergic receptors that are also GPCR members (metabotropic P2Y receptors). Pioneering studies to prove the hypothesis led to the discovery of interactions between A_1 and $P2Y_1$ receptors to form a functional unit with a particular pharmacological print (Yoshioka et al. 2001). Interestingly, A_1 and D_2 receptors were used as negative controls thus confirming previous results and the specificity of the interactions. Discovery of more P1-P2 receptor complexes (e.g., A_1 -P2Y₂Hets), and/or their physiological roles (especially in the brain), were further reported (Yoshioka et al. 2001, 2002a, b; Suzuki et al. 2006; Tonazzini et al. 2007). The interplay between P1 and P2 receptors opens interesting avenues due, *inter alia*, to the fact that extracellular ATP acting on P2 receptors is degraded into adenosine, which activates P1 receptors (homoreceptors/monomers or forming Hets).

The interest of the P1/P2 receptor interplay prompted (Schicker et al. 2009) the performance of an ambitious project to discover mixed P1/P2 receptor-receptor interactions. The authors tested A₁, A_{2A}, P2Y₁, P2Y₂, P2Y₁₂, and P2Y₁₃ receptors and the P2X₂ (ligand-gated ion channel) ionotropic receptors. They provided evidence for the formation of *heterooligomers among each other*. P2Y₁, P2Y₁₂, P2Y₁₃, A₁, A_{2A}, and P2X₂ receptors are also able to exist as homomers (Schicker et al. 2009). Reviews on the role of P1/P2 receptor-receptor interactions may be found in Nakata et al. (2010) and Suzuki et al. (2013). Of further interest for the present article, these results confirmed the occurrence of A₁R homodimers (Ciruela et al. 1995), A_{2A}R homodimers (Canals et al. 2004), and of A₁A_{2A}Hets for which a structural basis has recently been provided (see next Sect. 10.4).

After prediction by computational means of homodimerization of A_3 receptors (Kim and Jacobson 2006), Hill and colleagues detected both A_3 homodimers and heterodimers with A_1 receptors (May et al. 2011; Hill et al. 2014). We also have evidence of A_1A_3 Het expression in the CNS (data in preparation). It is likely that more A_3 -receptor-containing Hets exist, but they have not yet (to our knowledge) been identified.

Although some indirect evidence suggested that A_{2B} receptors ($A_{2B}R$) could be interacting with other GPCRs (Moriyama and Sitkovsky 2010), the direct proof is given in a recent publication (Hinz et al. 2018). As a matter of fact, the A_{2B} is an atypical receptor as the affinity for adenosine is very low, but its activation in lymphocytes may lead to calcium mobilization (Mirabet et al. 1997). In summary, it is assumed that A_{2B} receptors are activated is reservoirs with elevated adenosine levels or when hypoxic conditions lead to very high concentrations of the nucleoside. Also, there has been a lack of pharmacological tools that has been progressively solved. Intriguingly the A_{2B} protein has also been described in the CNS as a receptor for netrin-1, involved in axon guidance (Corset et al. 2000; Shewan et al. 2002). The discovery of heteromers formed by A_{2B} and A_{2A} receptors has led to a significant finding, namely, that the activation of the first alters the pharmacology and signaling of the latter. In a heterometric context, the affinity of $A_{2A}R$ selective ligands is markedly reduced, i.e., the activation of the receptor demands higher concentrations of A_{2A} receptor agonists. Accordingly, the efficacy of ligands targeting the $A_{2A}R$ would be dependent on the heteromeric context, especially in the case of A_{2A}A_{2B}Het occurrence (Hinz et al. 2018).

10.4 The A₁-A_{2A} Receptor Heteromer (A₁A_{2A} Het): A Unique Functional Unit

 A_1A_{2A} Het is in itself a paradigm to understand a fact that was inscrutable for decades, namely, the co-expression of one receptor for adenosine coupled to G_s and another receptor for adenosine coupled to G_i . In such cells, adenosine would lead to a "contradictory" output as, on the one hand, it would increase adenylate cyclase

activity (via G_s), and, on the other hand, it would decrease adenylate cyclase activity (via G_i). Co-expression of different receptors for a given neurotransmitter in the same cell is quite common, for instance for serotonin receptors (Santana et al. 2004). One of the possibilities (especially in neurons) was to assume that one of the receptors was expressed in a specific location of the cell, whereas the second receptor was located in a different location (always in the cell membrane but far away in spatial terms). In the case of the A_1 and A_{2A} receptors, the explanation is totally different, and, furthermore, it constitutes a clear paradigm of the need of GPCR Hets. In brief, the A_1A_{2A} Het is a device to sense the adenosine concentration to act accordingly, i.e., decreasing cAMP levels when [adenosine] is low and to increase cAMP levels when [adenosine] is high. Adenosine not only increases in hypoxia but its level varies with the metabolic status. Again, this is especially important in regions where neurons are very active and the adenosine/ATP ratio is high. The A_1A_{2A} Het was discovered by Ciruela et al. (2006), and the results on heteromerization of those receptors were later validated by Schicker et al. (2009).

There are different cell types in which the two receptors are co-expressed and where they may likely form A_1A_{2A} Hets. The physiological role of the A_1A_{2A} Het has however shown in the CNS and in relationship to control of neurotransmitter transport by adenosine and, importantly, in both neurons and glial cells. Our results centered in the striatum showed that the levels of co-expression of the two receptors in glutamatergic terminals reaching the striatum were markedly high and that low or high concentrations of adenosine led to opposite effects on glutamate release (Ciruela et al. 2006). This finding in 2006 did not provide any molecular mechanism but suggested that the coupling was different, to either G_s or G_i , depending on the concentration of the nucleoside. Furthermore, it seemed that the heteromeric context was the substrate to block A_1R -mediated signaling when the $A_{2A}R$ was activated. In summary at relatively low adenosine concentrations, the Het was providing $A_{2A}R$ -dependent signaling.

Fairly similar results were obtained in astrocytes and the control of the transport of one of the main inhibitory neurotransmitters in the CNS, gamma-aminobutyric acid (GABA). First, colocalization of the two receptors and occurrence of A_1A_{2A} Het was demonstrated. Second, the regulation of GABA uptake by cultures of astroglia depended on the concentration of adenosine. Indeed, the regulation of GAT-1 and GAT-3 transporters was via G_i or via G_s depending on whether the receptor activated within the A_1A_{2A} Het was, respectively, A_1R or $A_{2A}R$ (Cristóvão-Ferreira et al. 2013). The molecular basis of such a phenomenon was recently elucidated and described in the next Sect. 10.5.

10.5 The A₁-A_{2A} Receptor Heterotetramer: A Reliable Structural Model

On the one hand, the quaternary structure is crucial for Het function (Navarro et al. 2010). On the other hand, three-dimensional structures of GPCRs are difficult to decipher due to the technical difficulties in obtaining crystals of membrane proteins. Protein engineering and complementary technological advances have led to the elucidation of several GPCR structures and, also, to key structural elements of the GPCR-G protein interactions (see (Cordomí et al. 2015) and references therein). Those advances have served to understand that the most abundant G-proteincoupled signaling unit in the plasma membrane is a GPCR dimer. Exceptions may occur, i.e., a monomer GPCR may eventually couple to a G protein and be able to convey signal toward the inside of the cell. Indeed, this is not the case of the A_1A_{2A} Het whose minimal structure is likely constituted by one A_1R homodimer and one A2AR homodimer, i.e., a heterotetramer. Identification of GPCR oligomers combined with structural data and with modeling and other in silico approaches has provided relevant information concerning the structure of heteroreceptor complexes and their coupled G proteins. Also relevant is the fact that a substantial movement occurs within a GPCR and a G protein when the receptor becomes activated by agonists. Overall, membrane-attached GPCRs, which contain seven transmembrane domains and a tightly coupled alpha G protein subunit, likely form homodimers in a head-to-head fashion. Even allowing to increase the size of the heteroreceptor complex by considering four GPCRs and two coupled G proteins, the number of possible structures for the macromolecular complex is very few, as reported in the quite revealing work by Cordomí et al. (2015)

Using such in silico information, interfering peptides containing transmembrane sequences and, also, data from resonance energy transfer (using both receptors and G protein subunits as probes) and complementation assays, the first reliable structure for a GPCR Hets in complex with one G_s and one G_i protein was provided (Navarro et al. 2016b). The rhombus-shaped structure that contains alpha subunits of G_s or G_i bound to the outer protomers (in both A_1R and $A_{2A}R$ homodimers) would allow signaling via A_1R and via $A_{2A}R$. We mean that such a symmetrical structure cannot provide an asymmetrical signaling as that involved in the control (by adenosine) of glutamate or GABA transport regulation. The clue that explains the uniqueness and the functional properties of the A_1A_{2A} Het is a recent finding involving the C-terminal tail of the A2AR (Navarro et al. 2018). Different GPCRs display a wide range of lengths in their C-terminal domain. Whereas A1R has a short C-terminal end, the tail of the A_{2A}R is quite long. Then we hypothesized that a long C-terminal tail would, upon activation of the receptor, block the G-protein-mediated signaling arising from a closely located receptor. Exhaustive experimental and in silico work has provided reliable data showing that removal of the C-terminal domain of the A2AR leads to the disappearance of the Het fingerprint, i.e., activation of a truncated A2AR does not result in impairment of A₁R activation and G₁-mediated signaling. The huge diversity

in the length and structure of C-terminal domains deserves a closer look and is a challenge in future work in the GPCR field.

10.6 Adenosine-Receptor-Containing Heteromers and Schizophrenia

Although the evidence is higher in Parkinson's disease and the success is already evident by the approval of an $A_{2A}R$ antagonist in the therapy of Parkinson's (see below), we would like to make a brief account of data showing that adenosine receptors have also potential in the therapy of schizophrenia. Fuxe et al. (2005) reviewed possibilities of the heteromer as target for schizophrenia. Moreover, the dopamine D₃ receptor is one of the proposed therapeutic targets for treatment of the disease and, accordingly, the discovery of the $A_{2A}D_3$ Het receptor (Torvinen et al. 2005) place $A_{2A}R$ ligands as potential therapeutic drugs. Also along this line of reasoning is the above-described occurrence of occurrence of A_{2A} mGlu₅Hets. Reviews on the cumulative data that, based on adenosine-receptor-containing heteromers, open new perspectives in antischizophrenia therapy were provided by Fuxe et al. (2008, 2010) and Wardas (2008).

10.7 Adenosine-Receptor-Containing Heteromers and Parkinson's Disease (PD) and Levodopa-Induced Dyskinesia

In this section, we will first focus on the antiparkinsonian efficacy of adenosine receptor ligands to then take into consideration that any drug used by patients is – mostly – targeting Hets. Afterward, we will focus on the adenosine-receptor-containing heteromers that have been studied in both healthy and parkinsonian conditions. Hets constituted by adenosine receptor themselves and by adenosine and dopamine receptors fulfill these rules. The main objective in translational research is to identify suitable targets and efficacious drugs. To this respect dual adenosine-dopamine receptor ligands and bivalent compounds have been developed. The former (dual compounds) (Vendrell et al. 2007) may constitute the basis for the development of novel antiparkinsonian drugs. Instead, the latter (bivalent ligands), being unable to cross the blood-brain barrier and susceptible of being hydrolyzed soon after intake, have been instrumental to confirm the occurrence of adenosine-dopamine Hets in the striatum (A_{2A} - D_2 bivalents in Soriano et al. (2009) and A_1 - D_1 bivalents in Shen et al. (2013). Hence, such heteromers are demonstrable targets of antiparkinsonian drugs.

10.7.1 Efficacious Antiparkinsonian A_{2A}R Antagonists

Levodopa-based dopamine replacement therapy started decades ago and is still regarded as being of highest benefit for today's patients (Birkmayer and Hornykiewicz 1962, 1964; Olanow et al. 2004; Hornykiewicz 2006). Based on the early work of Fuxe and Ungerstedt (1974), on translational research and on data from clinical trials (Mizuno et al. 2013b; Saki et al. 2013; Kondo et al. 2015), a selective $A_{2A}R$ antagonist, istradefylline (NouriastTM), was approved in Japan for adjunctive antiparkinsonian therapy. The underlying idea was to reduce the dose of levodopa (or the dopamine-receptor-related medication) to diminish the side effects. In fact, long-term treatment with levodopa may lead to uncontrolled movements.

Cumulative evidence along decades, in different laboratories and under a variety of experimental setups, led to find a dopamine-adenosine antagonism in striatum. Even assuming that receptors are expressed individually (and not as heteromers), activation of adenosine A_1 and dopamine D_1 receptors in the direct pathway (or A_{2A} and D_2 in the indirect pathway) would lead to opposite effects as one of the receptors is coupled to G_i and another to G_s . Solid reviews describing the molecular basis of the antagonism may be found in the literature. As the complete list of reviews is quite notable, we here suggest the following ones that arise from different laboratories and/or present different but complementary perspectives (Bibbiani et al. 2003; Tanganelli et al. 2004; Schwarzschild et al. 2006; Ferré et al. 2007a, 2009b, 2010a, Fuxe et al. 2007, 2010, 2015; Simola et al. 2008; Armentero et al. 2011; Beggiato et al. 2014; Navarro et al. 2016a; Borroto-Escuela et al. 2017b).

In vivo experimental data on the potential of $A_{2A}R$ ligands to i) affect striatal dopaminergic neurotransmission and striatal plasticity and ii) to be efficacious in the unilateral 6-hydroxydopamine rat model of Parkinson's disease were provided by *inter alia* Pinna et al. (1997, 2007), Strömberg et al. (2000), Agnati et al. (2004). $A_{2A}R$ knockout (KO) mice have been used to ensure that a lack of $A_{2A}R$ -mediated signaling (and of any A_{2A} Het-mediated signaling) provides data that reinforces the antiparkinsonian potential of receptor blockade (Kachroo 2005).

Reviews on the role of Hets in the pathogenesis of Parkinson's disease and their potential as therapeutic targets of the disease appeared soon after the discovery of GPCR heteromers. Reviews with titles reflecting the relevance of purinergic signaling and/or receptor heteromerization were provided by Maggio et al. (2010), Navarro et al. (2016a, 2017), Borroto-Escuela et al. (2017a). However, the list of relevant reviews on the subject is quite broad. From such list we would recommend the following reviews (and references therein): (Schwarzschild et al. 2002; Morelli et al. 2007; Ferre et al. 2008; Fuxe et al. 2008, 2015; Ferré et al. 2009b).

We also believe that the paper by Short et al. (2006) provides a solid account on an interdependence between dopamine and adenosine receptors disclosed from characterizing receptor expression in adenosine and dopamine receptor KO mice (single KOs, i.e., only one receptor gene knocked out in each of the transgenic lines). This supports the early work of Fuxe and Ungerstedt (1974). Authors concluded that "the existence of functional interactions between dopaminergic and *purinergic systems in these reward and motor-related brain regions*" (Short et al. 2006).

Therefore, antagonists of adenosine receptors were soon proposed to increase dopamine action in Parkinson's disease, which consists of the depletion of dopamine in striatum due to nigral neurodegeneration. In summary, the conceptual approach was to use adenosine receptor antagonists to increase the dopaminergic action in striatal GABAergic neurons.

10.7.2 Heteromers as Targets of Antiparkinsonian Drugs

Soon after identification of A_1D_1 Het and of $A_{2A}D_2$ Het, these heteromers were proposed as targets or Parkinson's disease (Fuxe et al. 2003). Despite forgotten due to the usual way to develop novel drugs, i.e., by screening cells expressing individual receptors, it is evident that any antiparkinsonian medication is acting on receptors in heteromeric contexts. Thus, levodopa does not act on isolated dopaminergic receptors but on receptors forming Hets. In the case of NouriastTM, the drug is acting on those Hets identified as of today, namely, $A_{2A}D_2$ Hets with or without CB₁ or mGlu₅ receptors. The potential of cannabinoids or mGlu₅ receptor ligands has been suggested (Ferré et al. 2009b, 2010a), but the underlying reasons are out of the scope of the present article. In terms of adenosine receptors, it was suggested that A_1 receptor agonists acting on A_1R , which are expressed in the direct pathway, reduce D₁ receptor and levodopa-induced dyskinesia (see (Ferré et al. 1994; Florán et al. 2002; Franco et al. 2005; Mango et al. 2014) and references therein). It should be noted that in dyskinesia the level of the D₃ receptor and of D₁D₃Hets increase (Marcellino et al. 2008; Farré et al. 2015). These results suggest that also D₃ receptor ligands may be useful in the therapy of dyskinesia and that Hets may be considered targets for drugs able to counteract this side effect of chronic medication of levodopa and dopamine receptor agonists.

In what concerns the A_1A_{2A} Het, which is presynaptic (unlike A_1D_1 or $A_{2A}D_2$ Hets that are postsynaptic), it is not known how istradefylline (NouriastTM) is affecting its function in striatal glutamatergic terminals of patients. In healthy conditions, blockade of $A_{2A}R$ does not seem to produce any evident effect via those heteromers. In fact, $A_{2A}R$ antagonists are very safe, and this fits with a general rule (surely with exceptions) that receptors antagonists may be taken in chronic regimes by patients of diverse illnesses. A deeper look into the differential pharmacology of Hets has led to find that different drugs may have different "potencies" for the same receptor but in different heteromeric contexts. In brief, there is data showing that the affinity of an antagonist for $A_{2A}R$, or of caffeine for A_1R or $A_{2A}R$, is different when tested in different Hets. With data using different (pre- and postsynaptic) Hets and different antagonists, Orru et al. (2011) have suggested that "*on the basis of their preferential pre- versus postsynaptic actions, SCH-442416 and KW-6002 may be used as lead compounds to obtain more effective antidyskinetic and antiparkinsonian compounds,* *respectively.*" SCH-442416 is a broadly studied $A_{2A}R$ selective antagonist, whereas KW-6002 is another one (also known as istradefylline).

Interestingly, a recent report has linked early-onset Parkinson's disease cases to a point mutation in the gene of the A_1R (ADORA1). The mutation leads to the substitution of a conserved amino acid in transmembrane 7 (Jaberi et al. 2016). Based on current data and in the proposed models for receptor Hets, this mutation would not affect interacting interfaces of homo- or heteromers; then alternative explanations include altered binding of adenosine or altered signaling.

Taking into account the successful case of NouriastTM, one wonders why it is relevant to consider A_{2A} Hets as targets. On the one hand, the adenosine-dopamine antagonism is evidenced at the Het level, i.e., it is a significant print of the adenosinedopamine Hets. Therefore, the "intracellular" antagonism due to counterbalancing second messenger cAMP levels is complemented with antagonism at the receptor level within the A_{2A} -D₂Het context. The added value of having those Hets in a very precise location, the striatal spine module, also plays a role, as pointed out by Fuxe et al. (1998, 2007), Tanganelli et al. (2004), Ferré et al. (2007a, 2009b, 2010a), Beggiato et al. (2014) and as deduced by its role in controlling striatal glutamatergic neurotransmission (Ferré et al. 2007b).

Unlike for the A₁-A_{2A}Het, no detailed structural model exists for adenosinedopamine Hets. Allosteric interactions within the quaternary structure are essential for Het function, i.e., for integrating the dopamine and adenosine inputs (Fuxe et al. 2010). Remarkably, the $A_{2A}D_2$ Het has been a paradigm to detect electrostatic interactions that are key for the functional activity of the signaling unit. Apart from the consensus on the involvement of transmembrane domains in Het formation, it was demonstrated that strings of amino acid residues with opposite charges do interact, do it tightly, and are important for quaternary structure and function (Borroto-Escuela et al. 2010). One example is provided by the epitope-epitope interactions involving arginine residues in the N-terminal part of the third intracellular loop of the D₂R and acidic residues in the C-terminal end of the A_{2A}R (Ciruela et al. 2004). Complexes formed by synthetic peptides mimicking the interaction are even resistant to mass spectrometry processing thus demonstrating the strength of the epitope-epitope intraction. Finally, it should be noted that structure may be affected by phosphorylation, i.e., whereas serine would not participate on epitope-epitope interactions, a negatively charged phosphorylated serine would. One of the properties of Hets is a differential traffic respect to individually expressed receptors; apart from co-internalization and particular processing of internalized receptors that may be target to degradation or recycled back to the cell surface, there is involvement of both ß-arrestin/clathrin- and caveolin-dependent pathways (Escriche et al. 2003; Genedani et al. 2005; Franco et al. 2007; Borroto-Escuela et al. 2011)

Finally, it is worth mentioning the role of Ca^{2+} in heteromer-mediated signaling. It is likely that changes in the concentration of the ion may alter the quaternary structure of Hets in which electrostatic interactions are relevant. In fact, Ca^{2+} and/or calcium-binding proteins (e.g., calmodulin) modulate structure and function of $A_{2A}D_2$ Hets (Ferré et al. 2010b; Woods et al. 2008; Navarro et al. 2009). These results explain, at least in part, the elusive relationship between dopaminergic transmission and calcium ions.

10.7.3 How Levodopa-Induced Dyskinesia Affects Heteromerization

Parkinson's disease and adenosine-receptor-containing Hets constitute another paradigm due to the fact that their relationships have been investigated in healthy conditions and in the disease before and after chronic medication. On the one hand, the presence of A_{2A}D₂Hets and of Hets also including CB₁ receptors was demonstrated in rodent models of the disease (Pinna et al. 2014a, b). In our opinion, these results are important, as they show that these Hets are indeed targets of the dopamine replacement therapy. On the other hand, A_{2A} -CB₁-D₂ receptor heteromerization is disrupted after chronic levodopa administration (Pinna et al. 2014a, b). Remarkably, these results obtained in a rodent model were confirmed in a non-human primate model (Bonaventura et al. 2014), thus pointing to their validity for patients. Also consistent with those findings are the results showing in A2AR knockout animals a reduction in levodopa-induced dyskinesia as reported by Xiao et al. (2011). Interestingly, similar results were obtained upon deletion of the A₁R (Xiao et al. 2011). While it is not known whether heteromer disruption is cause or consequence of chronic medication, these results show that the target of the antiparkinsonian medication changes with time. To our understanding, these results may provide the basis for the design of optimal therapeutic approaches, i.e., varying the medication and/or the dose at different stages of the disease may reduce the side effects that for Parkinson's are not only dyskinesias but cognition deficits.

10.7.4 Adenosine Receptor and Adenosine-Receptor-Containing Heteromers and Neuroprotection

The success of approval of istradefylline for the therapy of Parkinson's disease provides further hopes for other neurodegenerative diseases. In vivo assays in animal models demonstrate the usefulness of $A_{2A}R$ antagonists in acute neural damage, for instance, in hypoxia (Chen and Pedata 2008; Melani et al. 2015; Boia et al. 2017). In the case of (chronic) neurodegenerative diseases (Parkinson's, Alzheimer's, etc.), the real issue is to know whether $A_{2A}R$ antagonists are addressing symptoms or are also affecting disease progression. Animals lacking expression of $A_{2A}R$ are more resistant to neuronal death in an α -synuclein model of Parkinson's disease (see (Kachroo and Schwarzschild 2012) and references therein). Surely one of today's challenges is to demonstrate whether $A_{2A}R$ antagonists are neuroprotective, i.e., they prevent neuronal death (see Franco and Navarra 2018 and references therein). Apart from the issue of demonstrating whether a given compound is neuroprotective in humans (Kieburtz and Olanow 2015; Olanow et al. 2017), there is evidence of microglia involvement in both promoting neuroinflammation, neuronal death and the release of factors that prevent neuronal death. In fact, after an insult and microglia cell recruitment and activation, there are two possible phenotypes: M1 or proinflammatory and M2 or neuroprotective (see (Franco and Fernández-Suárez 2015) and references therein). Due to the expression of adenosine receptors in resting and reactive microglia, it is suggested that adenosine receptor ligands may be protective (Corriden and Insel 2012; Koizumi et al. 2013; Beamer et al. 2016; Pedata et al. 2016; Woods et al. 2016). However, data is missing on how ligands acting on adenosine receptors may produce M2-skewed cells and on how adenosine-receptorcontaining heteromers may contribute to the inflammatory/neuroprotective balance.

References

- Agnati LF, Fuxe K, Zoli M et al (1982) New vistas on synaptic plasticity: the receptor mosaic hypothesis of the engram. Med Biol 60:183–190
- Agnati LF, Leo G, Vergoni AV et al (2004) Neuroprotective effect of L-DOPA co-administered with the adenosine A2A receptor agonist CGS 21680 in an animal model of Parkinson's disease. Brain Res Bull 64:155–164
- Armentero MT, Pinna A, Ferré S et al (2011) Past, present and future of A2A adenosine receptor antagonists in the therapy of Parkinson's disease. Pharmacol Ther 132:280–299
- Beamer E, Gölöncsér F, Horváth G et al (2016) Purinergic mechanisms in neuroinflammation: an update from molecules to behavior. Neuropharmacology 104:94–104
- Beggiato S, Antonelli T, Tomasini MC et al (2014) Adenosine A2A-D2 receptor-receptor interactions in putative heteromers in the regulation of the striato-pallidal gaba pathway: possible relevance for parkinson's disease and its treatment. Curr Protein Pept Sci 15:673–680
- Bibbiani F, Oh JD, Petzer JP et al (2003) A2A antagonist prevents dopamine agonist-induced motor complications in animal models of Parkinson's disease. Exp Neurol 184:285–294
- Birkmayer W, Hornykiewicz O (1962) The L-dihydroxyphenylalanine (L-DOPA) effect in Parkinson's syndrome in man: on the pathogenesis and treatment of Parkinson akinesis. Arch Psychiatr Nervenkr Z Gesamte Neurol Psychiatr 203:560–574
- Birkmayer W, Hornykiewicz O (1964) Additional experimental studies on L-DOPA in Parkinson's syndrome and reserpine parkinsonism. Arch Psychiatr Nervenkr 206:367–381
- Bogenpohl JW, Ritter SL, Hall RA et al (2012) Adenosine A2A receptor in the monkey basal ganglia: ultrastructural localization and colocalization with the metabotropic glutamate receptor 5 in the striatum. J Comp Neurol 520:570–589
- Boia R, Elvas F, Madeira MH et al (2017) Treatment with A2A receptor antagonist KW6002 and caffeine intake regulate microglia reactivity and protect retina against transient ischemic damage. Cell Death Dis 8:e3065
- Bonaventura J, Rico AJ, Moreno E et al (2014) L-DOPA-treatment in primates disrupts the expression of A2A adenosine-CB1 cannabinoid-D2 dopamine receptor heteromers in the caudate nucleus. Neuropharmacology 79:90–100
- Borroto-Escuela DO, Fuxe K (2017) Diversity and bias through dopamine D2R heteroreceptor complexes. Curr Opin Pharmacol 32:16–22

- Borroto-Escuela DO, Romero-Fernandez W, Tarakanov AO et al (2010) Characterization of the A2AR-D2R interface: focus on the role of the C-terminal tail and the transmembrane helices. Biochem Biophys Res Commun 402:801–807
- Borroto-Escuela DO, Romero-Fernandez W, Tarakanov AO et al (2011) On the existence of a possible A2A-D2-β-Arrestin2 complex: A2A agonist modulation of D2 agonist-induced β-arrestin2 recruitment. J Mol Biol 406:687–699
- Borroto-Escuela DO, Brito I, Romero-Fernandez W et al (2014) The G protein-coupled receptor heterodimer network (GPCR-HetNet) and its hub components. Int J Mol Sci 15:8570–8590
- Borroto-Escuela DO, Wydra K, Pintsuk J et al (2016) Understanding the functional plasticity in neural networks of the basal ganglia in cocaine use disorder: a role for allosteric receptor-receptor interactions in A2A-D2 heteroreceptor complexes. Neural Plast 2016:1–12
- Borroto-Escuela D, Narváez M, Navarro G et al (2017a) Heteroreceptor complexes implicated in Parkinson's disease. In: G-protein-coupled receptor dimers. The Receptors, vol 33. Humana Press, Cham, pp 477–501
- Borroto-Escuela DO, Narváez M, Wydra K et al (2017b) Cocaine self-administration specifically increases A2AR-D2R and D2R-sigma1R heteroreceptor complexes in the rat nucleus accumbens shell. Relevance for cocaine use disorder. Pharmacol Biochem Behav 155:24–31
- Cabello N, Gandía J, DCG B et al (2009) Metabotropic glutamate type 5, dopamine D 2 and adenosine A 2a receptors form higher-order oligomers in living cells. J Neurochem 109:1497–1507
- Canals M, Marcellino D, Fanelli F et al (2003) Adenosine A2A-dopamine D2 receptor-receptor heteromerization: qualitative and quantitative assessment by fluorescence and bioluminescence energy transfer. J Biol Chem 278:46741–46749
- Canals M, Burgueño J, Marcellino D et al (2004) Homodimerization of adenosine A2A receptors: qualitative and quantitative assessment by fluorescence and bioluminescence energy transfer. J Neurochem 88:726–734
- Cao Y, Sun WC, Jin L et al (2006) Activation of adenosine A1 receptor modulates dopamine D1 receptor activity in stably cotransfected human embryonic kidney 293 cells. Eur J Pharmacol 548:29–35
- Carriba P, Ortiz O, Patkar K et al (2007) Striatal adenosine A2A and cannabinoid CB1 receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids. Neuropsychopharmacology 32:2249–2259
- Chandrasekera PC, Wan TC, Gizewski ET et al (2013) Adenosine A1 receptors heterodimerize with β1- and β2-adrenergic receptors creating novel receptor complexes with altered G protein coupling and signaling. Cell Signal 25:736–742
- Chen JF, Pedata F (2008) Modulation of ischemic brain injury and neuroinflammation by adenosine A2A receptors. Curr Pharm Des 14:1490–1499
- Ciruela F, Casadó V, Mallol J et al (1995) Immunological identification of A1 adenosine receptors in brain cortex. J Neurosci Res 42:818–828
- Ciruela F, Escriche M, Burgueno J et al (2001) Metabotropic glutamate 1alpha and adenosine A1 receptors assemble into functionally interacting complexes. J Biol Chem 276:18345–18351
- Ciruela F, Burgueño J, Casadó V et al (2004) Combining mass spectrometry and pull-down techniques for the study of receptor heteromerization. Direct epitope-epitope electrostatic interactions between adenosine A2A and dopamine D2receptors. Anal Chem 76:5354–5363
- Ciruela F, Casadó V, Rodrigues RJ et al (2006) Presynaptic control of striatal glutamatergic neurotransmission by adenosine A1-A2A receptor heteromers. J Neurosci 26:2080–2087
- Cordomí A, Navarro G, Aymerich MS et al (2015) Structures for G-protein-coupled receptor tetramers in complex with G proteins. Trends Biochem Sci 40:548–551
- Corriden R, Insel PA (2012) New insights regarding the regulation of chemotaxis by nucleotides, adenosine, and their receptors. Purinergic Signal 8:587–598
- Corset V, Nguyen-Ba-Charvet KT, Forcet C et al (2000) Netrin-1-mediated axon outgrowth and cAMP production requires interaction with adenosine A2b receptor. Nature 407:747–750
- Cristóvão-Ferreira S, Navarro G, Brugarolas M et al (2013) A1R-A2AR heteromers coupled to Gs and G i/o proteins modulate GABA transport into astrocytes. Purinergic Signal 9:433–449

- Doumazane E, Scholler P, Zwier JM et al (2011) A new approach to analyze cell surface protein complexes reveals specific heterodimeric metabotropic glutamate receptors. FASEB J 25:66–77
- Dunham JH, Meyer RC, Garcia EL et al (2009) GPR37 surface expression enhancement via N-terminal truncation or protein-protein interactions. Biochemistry 48:10286–10297
- Escriche M, Burgueño J, Ciruela F et al (2003) Ligand-induced caveolae-mediated internalization of A1 adenosine receptors: morphological evidence of endosomal sorting and receptor recycling. Exp Cell Res 285:72–90
- Farré D, Muñoz A, Moreno E et al (2015) Stronger dopamine D1 receptor-mediated neurotransmission in dyskinesia. Mol Neurobiol 52:1408–1420
- Ferré S, O'Connor WT, Snaprud P et al (1994) Antagonistic interaction between adenosine A2A receptors and dopamine D2 receptors in the ventral striopallidal system implications for the treatment of schizophrenia. Neuroscience 63:765–773
- Ferré S, Karcz-Kubicha M, Hope BT et al (2002) Synergistic interaction between adenosine A2A and glutamate mGlu5 receptors: implications for striatal neuronal function. Proc Natl Acad Sci U S A 99:11940–11945
- Ferré S, Ciruela F, Woods AS et al (2003) Glutamate mGluR5/adenosine A2A/dopamine D2 receptor, interactions in the striatum implications for drug therapy in neuro-psychiatric disorders and drug abuse. Curr Med Chem Cent Nerv Syst Agents 3:1–26
- Ferré S, Agnati LF, Ciruela F et al (2007a) Neurotransmitter receptor heteromers and their integrative role in 'local modules': the striatal spine module. Brain Res Rev 55:55–67
- Ferré S, Ciruela F, Woods AS et al (2007b) Functional relevance of neurotransmitter receptor heteromers in the central nervous system. Trends Neurosci 30:440–446
- Ferre S, Ciruela F, Borycz J et al (2008) Adenosine A1-A2A receptor heteromers: new targets for caffeine in the brain. Front Biosci 13:2391–2399
- Ferré S, Baler R, Bouvier M et al (2009a) Building a new conceptual framework for receptor heteromers. Nat Chem Biol 5:131–134
- Ferré S, Goldberg SR, Lluis C et al (2009b) Looking for the role of cannabinoid receptor heteromers in striatal function. Neuropharmacology 56:226–234
- Ferré S, Lluís C, Justinova Z et al (2010a) Adenosine-cannabinoid receptor interactions implications for striatal function. Br J Pharmacol 160(3):443–453
- Ferré S, Woods AS, Navarro G et al (2010b) Calcium-mediated modulation of the quaternary structure and function of adenosine A2A-dopamine D2 receptor heteromers. Curr Opin Pharmacol 10:67–72
- Florán B, Barajas C, Florán L et al (2002) Adenosine A1 receptors control dopamine D1-dependent [(3)H]GABA release in slices of substantia nigra pars reticulata and motor behavior in the rat. Neuroscience 115:743–751
- Franco R, Fernández-Suárez D (2015) Alternatively activated microglia and macrophages in the central nervous system. Prog Neurobiol 131:65–86
- Franco R, Ferré S, Torvinen M et al (2001) Adenosine/dopamine receptor-receptor interactions in the central nervous system. Drug Dev Res 52:296–302
- Franco R, Ciruela F, Casadó V et al (2005) Partners for adenosine A1receptors. J Mol Neurosci 26:221–231
- Franco R, Lluis C, Canela EI et al (2007) Receptor-receptor interactions involving adenosine A1 or dopamine D1 receptors and accessory proteins. J Neural Transm 114:93–104
- Franco R, Martínez-Pinilla E, Lanciego JL et al (2016) Basic pharmacological and structural evidence for class A G-protein-coupled receptor heteromerization. Front Pharmacol 7:76
- Franco R, Navarro G (2018) Adenosine A2A Receptor Antagonists in Neurodegenerative Diseases: Huge Potential and Huge Challenges. Front Psychiatry 9:68
- Fuxe K, Agnati LF (1985) Receptor-receptor interactions in the central nervous system A new integrative mechanism in synapses. Med Res Rev 5:441–482

- Fuxe K, Ungerstedt U (1974) Action of caffeine and theophyllamine on supersensitive dopamine receptors: considerable enhancement of receptor response to treatment with DOPA and dopamine receptor agonists. Med Biol 52:48–54
- Fuxe K, Agnati LF, Benfenati F et al (1981) Modulation by cholecystokinins of 3 H-spiroperidol binding in rat striatum: evidence for increased affinity and reduction in the number of binding sites. Acta Physiol Scand 113:567–569
- Fuxe K, Agnati LF, Benfenati F et al (1983) Evidence for the existence of receptor-receptor interactions in the central nervous system studies on the regulation of monoamine receptors by neuropeptides. J Neural Transm Suppl 18:165–179
- Fuxe K, Härfstrand A, Agnati LF et al (1987) Central catecholamine-neuropeptide Y interactions at the pre- and postsynaptic level in cardiovascular centers. J Cardiovasc Pharmacol 10(Suppl 1):1–13
- Fuxe K, Ferré S, Zoli M et al (1998) Integrated events in central dopamine transmission as analyzed at multiple levels evidence for intramembrane adenosine A2A/dopamine D2 and adenosine A1/ dopamine D1 receptor interactions in the basal ganglia. Brain Res Brain Res Rev 26:258–273
- Fuxe K, Agnati LFF, Jacobsen K et al (2003) Receptor heteromerization in adenosine A2A receptor signaling: relevance for striatal function and Parkinson's disease. Neurology 61:S19–S23
- Fuxe K, Ferré S, Canals M et al (2005) Adenosine A2A and dopamine D2 heteromeric receptor complexes and their function. J Mol Neurosci 26:209–220
- Fuxe K, Marcellino D, Genedani S et al (2007) Adenosine A2A receptors dopamine D2 receptors and their interactions in Parkinson's disease. Mov Disord 22:1990–2017
- Fuxe K, Marcellino D, Rivera A et al (2008) Receptor–receptor interactions within receptor mosaics impact on neuropsychopharmacology. Brain Res Rev 58:415–452
- Fuxe K, Marcellino D, Leo G et al (2010) Molecular integration via allosteric interactions in receptor heteromers A working hypothesis. Curr Opin Pharmacol 10:14–22
- Fuxe K, Borroto-Escuela D, Fisone G et al (2014a) Understanding the role of heteroreceptor complexes in the central nervous system. Curr Protein Pept Sci 15:647–654
- Fuxe K, Tarakanov A, Romero Fernandez W et al (2014b) Diversity and bias through receptorreceptor interactions in GPCR heteroreceptor complexes focus on examples from dopamine D2 receptor heteromerization. Front Endocrinol (Lausanne) 5:1–11
- Fuxe K, Guidolin D, Agnati LF et al (2015) Dopamine heteroreceptor complexes as therapeutic targets in Parkinson's disease. Expert Opin Ther Targets 19:377–398
- Genedani S, Guidolin D, Leo G et al (2005) Computer-assisted image analysis of caveolin-1 involvement in the internalization process of adenosine A2A-dopamine D2receptor heterodimers. J Mol Neurosci 26:177–184
- George SR, Kern A, Smith RG et al (2014) Dopamine receptor heteromeric complexes and their emerging functions. Prog Brain Res 211:183–200
- Gines S, Hillion J, Torvinen M et al (2000) Dopamine D1 and adenosine A1 receptors form functionally interacting heteromeric complexes. Proc Natl Acad Sci 97:8606–8611
- Gomes I, Jordan BA, Gupta A et al (2000) Heterodimerization of mu and delta opioid receptors: a role in opiate synergy. J Neurosci 20:RC110
- Hill SJ, May LT, Kellam B et al (2014) Allosteric interactions at adenosine A(1) and A(3) receptors: new insights into the role of small molecules and receptor dimerization. Br J Pharmacol 171:1102–1113
- Hillefors M, Hedlund PB, Euler G (1999) Effects of adenosine A(2A) receptor stimulation in vivo on dopamine D3 receptor agonist binding in the rat brain. Biochem Pharmacol 58:1961–1964
- Hillion J, Canals M, Torvinen M et al (2002) Coaggregation cointernalization and codesensitization of adenosine A2A receptors and dopamine D2 receptors. J Biol Chem 277:18091–18097
- Hinz S, Navarro G, Borroto-Escuela D et al (2018) Adenosine A2A receptor ligand recognition and signaling is blocked by A2B receptors. Oncotarget 9:13593–13611
- Hornykiewicz O (2006) The discovery of dopamine deficiency in the parkinsonian brain. J Neural Transm 9:15

- Jaberi E, Rohani M, Shahidi GA et al (2016) Mutation in ADORA1 identified as likely cause of early-onset parkinsonism and cognitive dysfunction. Mov Disord 31:1004–1011
- Kachroo A (2005) Interactions between metabotropic glutamate 5 and adenosine A2A receptors in normal and Parkinsonian mice. J Neurosci 25:10414–10419
- Kachroo A, Schwarzschild MA (2012) Adenosine A(2A) receptor gene disruption protects in an α -synuclein model of Parkinson's disease. Ann Neurol 71:278–282
- Kieburtz K, Olanow CW (2015) Advances in clinical trials for movement disorders. Mov Disord 30:1580–1587
- Kim SK, Jacobson KA (2006) Computational prediction of homodimerization of the A3 adenosine receptor. J Mol Graph Model 25:549–561
- Koizumi S, Ohsawa K, Inoue K et al (2013) Purinergic receptors in microglia: functional modal shifts of microglia mediated by P2 and P1 receptors. Glia 61:47–54
- Kondo T, Mizuno Y, Japanese Istradefylline Study Group (2015) A long-term study of istradefylline safety and efficacy in patients with Parkinson disease. Clin Neuropharmacol 38:41–46
- Maggio R, Aloisi G, Silvano E et al (2010) Heterodimerization of dopamine receptors: new insights into functional and therapeutic significance. Parkinsonism Relat Disord 15:S2–S7
- Mango D, Bonito-Oliva A, Ledonne A et al (2014) Adenosine A1 receptor stimulation reduces D1 receptor-mediated GABAergic transmission from striato-nigral terminals and attenuates I-DOPA-induced dyskinesia in dopamine-denervated mice. Exp Neurol 261:733–743
- Marcellino D, Ferré S, Casadó V et al (2008) Identification of dopamine D1-D3 receptor heteromers: indications for a role of synergistic D1-D3 receptor interactions in the striatum. J Biol Chem 283:26016–26025
- Márquez-Gómez R, Robins MT, Gutiérrez-Rodelo C et al (2018) Functional histamine H 3 and adenosine A2A receptor heteromers in recombinant cells and rat striatum. Pharmacol Res 129:515–525
- May LT, Bridge LJ, Stoddart L et al (2011) Allosteric interactions across native adenosine-A3 receptor homodimers: quantification using single-cell ligand-binding kinetics. FASEB J 25:3465–3476
- Melani A, Dettori I, Corti F et al (2015) Time-course of protection by the selective A2A receptor antagonist SCH58261 after transient focal cerebral ischemia. Neurol Sci 36:1441–1448
- Mirabet M, Mallol J, Lluis C et al (1997) Calcium mobilization in Jurkat cells via A(2b) adenosine receptors. Br J Pharmacol 122:1075–1082
- Mizuno N, Suzuki T, Hirasawa N et al (2012) Hetero-oligomerization between adenosine A₁ and thromboxane A₂ receptors and cellular signal transduction on stimulation with high and low concentrations of agonists for both receptors. Eur J Pharmacol 677:5–14
- Mizuno N, Suzuki T, Kishimoto Y et al (2013a) Biochemical assay of G protein-coupled receptor oligomerization: adenosine A1 and thromboxane A2 receptors form the novel functional hetero-oligomer. Methods Cell Biol 117:213–227
- Mizuno Y, Kondo T, Japanese Istradefylline Study Group (2013b) Adenosine A2A receptor antagonist istradefylline reduces daily OFF time in Parkinson's disease. Mov Disord 28:1138–1141
- Morelli M, Paolo T, Di Wardas J et al (2007) Role of adenosine A2A receptors in parkinsonian motor impairment and I-DOPA-induced motor complications. Prog Neurobiol 83:293–309
- Moriyama K, Sitkovsky MV (2010) Adenosine A2A receptor is involved in cell surface expression of A2B receptor. J Biol Chem 285:39271–39288
- Muñoz LM, Lucas P, Navarro G et al (2009) Dynamic regulation of CXCR1 and CXCR2 homoand heterodimers. J Immunol 183:7337–7346
- Muñoz LM, Lucas P, Holgado BL et al (2011) Receptor oligomerization: a pivotal mechanism for regulating chemokine function. Pharmacol Ther 131:351–358
- Muñoz LM, Holgado BL, Martínez AC et al (2012) Chemokine receptor oligomerization: a further step toward chemokine function. Immunol Lett 145:23–29
- Nakata H, Suzuki T, Namba K et al (2010) Dimerization of G protein-coupled purinergic receptors: increasing the diversity of purinergic receptor signal responses and receptor functions. J Recept Signal Transduction 30:337–346

- Navarro G, Carriba P, Gandía J et al (2008) Detection of heteromers formed by cannabinoid CB1 dopamine D2 and adenosine A2A G-protein-coupled receptors by combining bimolecular fluorescence complementation and bioluminescence energy transfer. Sci World J 8:1088–1097
- Navarro G, Aymerich MS, Marcellino D et al (2009) Interactions between calmodulin adenosine A2A and dopamine D2 receptors. J Biol Chem 284:28058–28068
- Navarro G, Ferre S, Cordomi A et al (2010) Interactions between intracellular domains as key determinants of the quaternary structure and function of receptor heteromers. J Biol Chem 285:27346–27359
- Navarro G, Borroto-Escuela DO, Fuxe K et al (2016a) Purinergic signaling in Parkinson's disease relevance for treatment. Neuropharmacology 104:161–168
- Navarro G, Cordomí A, Zelman-Femiak M et al (2016b) Quaternary structure of a G-proteincoupled receptor heterotetramer in complex with Gi and Gs. BMC Biol 14:26
- Navarro G, Borroto-Escuela D, Angelats E et al (2017) Receptor-heteromer mediated regulation of endocannabinoid signaling in activated microglia relevance for Alzheimer's disease and levo-dopa-induced dyskinesia. Brain Behav Immun 67:139–151
- Navarro G, Cordomí A, Brugarolas M et al (2018) Cross-communication between Gi and Gs in a G-protein-coupled receptor heterotetramer guided by a receptor C-terminal domain. BMC Biol 16:24
- Nishi A, Liu F, Matsuyama S et al (2003) Metabotropic mGlu5 receptors regulate adenosine A2A receptor signaling. Proc Natl Acad Sci U S A 100:1322–1327
- Noble F, Cox BM (1995) Differential regulation of D1 dopamine receptor and of A2A Adenosine receptor stimulated adenylyl cyclase by mu- delta 1- and delta 2 opioid agonists in rat caudate putamen. J Neurochem 65:125–133
- Olanow CW, Agid Y, Mizuno Y et al (2004) Levodopa in the treatment of Parkinson's disease: current controversies. Mov Disord 19:997–1005
- Olanow CW, Kieburtz K, Katz R (2017) Clinical approaches to the development of a neuroprotective therapy for PD. Exp Neurol 298:246–251
- Orru M, Bakešová J, Brugarolas M et al (2011) Striatal pre- and postsynaptic profile of adenosine A(2A) receptor antagonists. PLoS One 6:e16088
- Pedata F, Dettori I, Coppi E et al (2016) Purinergic signalling in brain ischemia. Neuropharmacology 104:105–130
- Perreault ML, Hasbi A, O'dowd BF et al (2014) Heteromeric dopamine receptor signaling complexes: emerging neurobiology and disease relevance. Neuropsychopharmacology 39:156–168
- Pinna A, Wardas J, Cristalli G et al (1997) Adenosine A(2A) receptor agonists increase Fos-like immunoreactivity in mesolimbic areas. Brain Res 759:41–49
- Pinna A, Pontis S, Borsini F et al (2007) Adenosine A2A receptor antagonists improve deficits in initiation of movement and sensory motor integration in the unilateral 6-hydroxydopamine rat model of Parkinson's disease. Synapse 61:606–614
- Pinna A, Bonaventura J, Farré D et al (2014a) L-DOPA disrupts adenosine A2A-cannabinoid CB-1-dopamine D-2 receptor heteromer cross-talk in the striatum of hemiparkinsonian rats: biochemical and behavioral studies. Exp Neurol 253:180–191
- Pinna A, Bonaventura J, Farré D et al (2014b) l-DOPA disrupts adenosine A2A–cannabinoid CB1–dopamine D2 receptor heteromer cross-talk in the striatum of hemiparkinsonian rats: biochemical and behavioral studies. Exp Neurol 253:180–191
- Rashid AJ, So CH, Kong MMC et al (2007) D1-D2 dopamine receptor heterooligomers with unique pharmacology are coupled to rapid activation of Gq/11 in the striatum. Proc Natl Acad Sci U S A 104:654–659
- Rimondini R, Ferré S, Ogren SO et al (1997) Adenosine A2A agonists: a potential new type of atypical antipsychotic. Neuropsychopharmacology 17:82–91
- Saki M, Yamada K, Koshimura E et al (2013) In vitro pharmacological profile of the A2A receptor antagonist istradefylline. Naunyn Schmiedeberg's Arch Pharmacol 386:963–972

- Santana N, Bortolozzi A, Serrats J et al (2004) Expression of serotonin1A and serotonin2A receptors in pyramidal and GABAergic neurons of the rat prefrontal cortex. Cereb Cortex 14:1100–1109
- Schicker K, Hussl S, Chandaka GK et al (2009) A membrane network of receptors and enzymes for adenine nucleotides and nucleosides. Biochim Biophys Acta 1793:325–334
- Schwarzschild MA, Chen JF, Ascherio A (2002) Caffeinated clues and the promise of adenosine A(2A) antagonists in PD. Neurology 58:1154–1160
- Schwarzschild MA, Agnati L, Fuxe K et al (2006) Targeting adenosine A2A receptors in Parkinson's disease. Trends Neurosci 29:647–654
- Shen JJ, Zhang L, Song W et al (2013) Design synthesis and biological evaluation of bivalent ligands against A(1)-D(1) receptor heteromers. Acta Pharmacol Sin 34:441–452
- Shewan D, Dwivedy A, Anderson R et al (2002) Age-related changes underlie switch in netrin-1 responsiveness as growth cones advance along visual pathway. Nat Neurosci 5:955–962
- Short JL, Ledent C, Borrelli E et al (2006) Genetic interdependence of adenosine and dopamine receptors: evidence from receptor knockout mice. Neuroscience 139:661–670
- Simola N, Morelli M, Pinna A (2008) Adenosine A2A receptor antagonists and Parkinson's disease: state of the art and future directions. Curr Pharm Des 14:1475–1489
- Soriano A, Ventura R, Molero A et al (2009) Adenosine A2A receptor-antagonist/dopamine D2 receptor-agonist bivalent ligands as pharmacological tools to detect A2A/ D2receptor heteromers. J Med Chem 52:5590–5602
- Springael JY, Urizar E, Parmentier M (2005) Dimerization of chemokine receptors and its functional consequences. Cytokine Growth Factor Rev 16:611–623
- Strömberg I, Popoli P, Müller CE et al (2000) Electrophysiological and behavioural evidence for an antagonistic modulatory role of adenosine A2A receptors in dopamine D2 receptor regulation in the rat dopamine-denervated striatum. Eur J Neurosci 12:4033–4037
- Suzuki T, Namba K, Tsuga H et al (2006) Regulation of pharmacology by hetero-oligomerization between A1 adenosine receptor and P2Y2 receptor. Biochem Biophys Res Commun 351:559–565
- Suzuki T, Namba K, Mizuno N et al (2013) Hetero-oligomerization and specificity changes of G protein-coupled purinergic receptors: novel insight into diversification of signal transduction. Methods Enzymol 521:239–257
- Tanganelli S, Sandager Nielsen K, Ferraro L et al (2004) Striatal plasticity at the network level focus on adenosine A2A and D2 interactions in models of Parkinson's disease. Parkinsonism Relat Disord 10:273–280
- Tonazzini I, Trincavelli ML, Storm-Mathisen J et al (2007) Co-localization and functional crosstalk between A1 and P2Y1 purine receptors in rat hippocampus. Eur J Neurosci 26:890–902
- Torvinen M, Ginés S, Hillion J et al (2002) Interactions among adenosine deaminase adenosine A1 receptors and dopamine D1 receptors in stably cotransfected fibroblast cells and neurons. Neuroscience 113:709–719
- Torvinen M, Marcellino D, Canals M et al (2005) Adenosine A2A receptor and dopamine D3 receptor interactions: evidence of functional A2A/D3 heteromeric complexes. Mol Pharmacol 67:400–407
- Vendrell M, Angulo E, Casadó V et al (2007) Novel ergopeptides as dual ligands for adenosine and dopamine receptors. J Med Chem 50:3062–3069
- Wardas J (2008) Potential role of adenosine A2A receptors in the treatment of schizophrenia. Front Biosci 13:4071–4096
- Woods AS, Marcellino D, Jackson SN et al (2008) How calmodulin interacts with the adenosine A2A and the dopamine D2 receptors. J Proteome Res 7:3428–3434
- Woods LT, Ajit D, Camden JM et al (2016) Purinergic receptors as potential therapeutic targets in Alzheimer's disease. Neuropharmacology 104:169–179
- Xiao D, Cassin JJ, Healy B et al (2011) Deletion of adenosine A1 or A2A receptors reduces I-34dihydroxyphenylalanine-induced dyskinesia in a model of Parkinson's disease. Brain Res 1367:310–318

- Yoshioka K, Saitoh O, Nakata H (2001) Heteromeric association creates a P2Y-like adenosine receptor. Proc Natl Acad Sci U S A 98:7617–7622
- Yoshioka K, Hosoda R, Kuroda Y et al (2002a) Hetero-oligomerization of adenosine A1 receptors with P2Y1 receptors in rat brains. FEBS Lett 531:299–303
- Yoshioka K, Saitoh O, Nakata H (2002b) Agonist-promoted heteromeric oligomerization between adenosine A(1) and P2Y(1) receptors in living cells. FEBS Lett 523:147–151
- Zoli M, Agnati LF, Hedlund PB et al (1993) Receptor-receptor interactions as an integrative mechanism in nerve cells. Mol Neurobiol 7:293–334