Endophytic Yeasts: Biology, Ecology and Applications



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Abstract While the study of bacteria and filamentous fungi that inhabit plants has been extensive, endophytic yeast biology remains less understood. Recent research is encouraging as to the potential of endophytic yeasts for industrial and agricultural applications, providing strong incentives for better understanding these yeasts. Endophytic yeasts could have significant advantages over bacterial and filamentous endophytes and they can easily be cultured, stored long term and applied to crops. While more research and especially field trials are required to assess their potential, it seems the use of endophytic yeasts could be a viable way of reducing fertilizer and water inputs in agriculture, and potentially increasing yields. Their application seems especially promising in the field of remediation of heavy metal pollutants, and as biocontrol agents to protect plants from pathogens. However, many aspects of endophytic yeasts are able to colonize their niche. This chapter reviews recent research on endophytic yeasts.

1 Introduction

While the study of endophytic bacteria and filamentous fungi has been extensive, endophytic yeast biology and ecology remain poorly understood. Whether this is due to bias in isolation and cultivation techniques towards bacteria and filamentous fungi, or simply a lesser prevalence of yeasts in the phytobiome, is unclear. This apparent rarity could also be exacerbated by the complex and cryptic nature of their life styles, especially in the case of basidiomycetous yeasts. However, yeasts present many advantages for agricultural use over filamentous fungi given their simpler cultivation and application techniques. Yeasts applied to plants seem to be

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distributed systemically, unlike some filamentous fungi, and can be easily cultivated in a similar way to bacteria. They also present advantages over bacteria including their ability to be freeze-dried more efficiently than bacteria and therefore be more easily distributed for agricultural use. Given their ubiquitous presence in the phytobiome and their potentially great agricultural and industrial applications, it is crucial to further study endophytic yeasts. This chapter reviews recent research on endophytic yeasts and points to the need for further research into the ecology of these valuable microorganisms.

2 Defining Endophytic Yeasts

Here we define endophytic yeasts as fungi that have a unicellular part of their lifecycle, which typically reside within plant tissues and do not cause damage to their hosts. The divide between epiphytic and endophytic yeasts has remained blurred, especially in the case of yeasts that populate fruit surfaces, and could potentially penetrate the outer layer of fruits. However, several groups have reported their confidence in surface sterilization techniques, reporting a lack of epiphytes in the washing media after surface sterilization (Prior et al. 2017; Tantirungkij et al. 2015).

Different hypotheses describing the typical composition of the endophytic yeast community published recently point to diverging views over this issue. This is likely in part due to the large variation of endophytic communities based on the host plant as well as various biotic and abiotic factors (Prior et al. 2017). Opinions differ especially over whether the phytobiome is dominated by the commonly known and studied ascomycetous yeasts or the more cryptic basidiomycetous yeasts. One school of thought claims that, given that ascomycete fungi are more common in endophytic filamentous fungi communities and in the world as a whole, endophytic yeasts must be mostly ascomycetes (Prior et al. 2017). This claim is supported by the fact that basidiomycetes are strict aerobes and therefore ascomycetes may be more successful in microaerobic environments inside plant tissues or fruits (Glushakova and Kachalkin 2017). However, there have been reports of a strong prevalence of basidiomycetous yeasts in plant tissues including in the leaves of Ficus plants (Solis et al. 2014). Some might propose that these recent results may be due to a cultivation bias in favor of basidiomycetes. However, this hypothesis was supported by a culture independent approach used for rice leaf yeasts, which also showed that more than half of the reported phylotypes were previously unknown, pointing to the flaws of relying solely on culturing methods for characterizing endophytic yeast communities (Tantirungkij et al. 2015). One point supporting the basidiomycetous-dominated hypothesis is the fact that basidiomycetes have a larger metabolic diversity which could allow them to adapt better to the in planta environment (Ichinose 2013). Regardless, these differing hypotheses support the need for further research using high throughput culture-independent sequencing approaches in order to better understand endophytic yeast diversity and prevalence.

3 Plant Growth-Promoting Characteristics

The majority of our knowledge of plant growth-promoting characteristics of endophytes comes from studying bacterial endophytes and comparisons can be useful for studying endophytic yeasts. This hints at evidence of convergent evolution of habitat use between yeasts and bacteria that occupy the phytosphere. This is understandable from their similar lifestyles in terms of their ecological niches, as well as their physiology, including their single-celled existence adapted to aquatic environments, fast growth rates, and the importance of biofilms in their lifecycle. Additionally, there has been evidence indicating the possibility of mixed biofilm formation containing both bacteria and yeasts (Bandara et al. 2006; Firrincieli et al. 2015). These convergent traits, as well as ecological and physical proximity, seem to indicate the possibility of cross-kingdom horizontal gene transfer events. These events have been hinted at in the past but little conclusive evidence has been found as of yet (Firrincieli et al. 2015; Hall et al. 2005; Marcet-Houben and Gabaldon 2010).

Several growth-promoting traits are commonly found in endophytic yeasts and endophytes in general including phytohormone production, stress alleviation, protection against pathogens and increasing nutrient uptake by the plant. All of these traits have been found in yeasts but three stand out in particular as being common to endophytic yeasts, and seem to be unifying characteristics. These are IAA production, siderophore production and ACC deaminase activity.

Plant hormone production provides a direct method of plant growth promotion by endophytes. Auxins and gibberellins have many growth-promoting properties in plants including promoting root growth and stem elongation, as well as more broadly, cell proliferation and elongation. Specifically, the production of indole-3-acetic acid (IAA) by endophytic yeasts has been widely reported by several groups and extensively reviewed (Doty 2013; Moller et al. 2016; Nassar et al. 2005). Interestingly, this trait was reported to only occur in media supplemented with tryptophan (Hardoim et al. 2008; Xin et al. 2009). This also seems to be true in the case of endophytic bacteria (Kandel et al. 2017). This observation provides a strong basis for the plant endophyte symbiosis. Tryptophan is a complex amino acid which is costly to produce for the yeast but is found inside the plant and in plant exudates (Kamilova et al. 2006). While it may be too costly for the yeast to produce tryptophan to make IAA on its own, when the plant provides it with tryptophan, the yeast can, in return, provide IAA and subsequently promote the growth of its host. This also serves as an indirect way for the plant to divert nitrogen into root and shoot mass, especially in situations where nitrogen is plentiful.

ACC, or 1-aminocyclopropane-1-carboxylate, is a molecule that serves as a precursor to the production of ethylene, a plant stress hormone. ACC deaminases are enzymes that cleave the molecule ACC into α -ketobutyrate and ammonia and have commonly been found in endophytic bacteria, as well as endophytic yeasts (Glick et al. 2007; Nutaratat et al. 2014; Sun et al. 2009). The endophytic yeasts that have this activity may therefore serve as sinks for ACC, therefore decreasing total ethylene concentrations in the plant. Ethylene is a stress hormone that inhibits

growth of the plant. Yeasts may therefore prevent this growth inhibition, and generally increase the plant's tolerance to stress. These enzymes may also serve as a mechanism for ammonia secretion by the yeasts, which have been reported in the past and could serve as a way for the plant to recycle nitrogen using its symbiotic partners (Nutaratat et al. 2014).

Siderophore production is another common trait of endophytic yeasts that has been substantially reviewed for arboreal endophytes and found in rice and sugar cane leaf endophytes (Moller et al. 2016; Nutaratat et al. 2014). Siderophores are iron chelating compounds microorganisms produce when facing iron starvation (Loaces et al. 2011). These compounds are secreted and then recaptured once they have chelated iron. This could serve as a method for plant growth promotion as they could provide the plant with chelated iron that it cannot absorb directly from the soil. Additionally, these compounds could help endophytic yeasts colonize the plant and exclude other microorganisms and could explain antifungal characteristics of some yeasts as described later in this review. This is well known in epiphytic yeasts, which prevent fruit spoilage, for example (Loaces et al. 2011; Nutaratat et al. 2014).

Several other plant growth-promoting characteristics have been reported recently but less commonly, possibly as a result of a lack of in depth characterization of these endophytes. One of these characteristics is catalase activity, an enzyme that decreases the presence of reactive oxygen species and could help promote stress tolerance in the plant (Khalifa et al. 2016). This catalase activity has been reported recently in many of the yeast endophytes found in sugar cane and rice leaves in Thailand (Nutaratat et al. 2014). Nutaratat et al. also reported ammonium secretion independent of ACC deaminase activity, as well as increased phosphate solubilization by several of the yeasts. Additionally, the production of polyamines by these yeasts was also reported. These compounds have a complex role in plant growth and are involved in many cellular processes including the synthesis of macromolecules, as well as growth, survival and stress tolerance of cells. However, they could also generally function in plant growth promotion by yeasts (Takahashi and Kakehi 2010).

4 Industrial and Environmental Applications

Given the growth-promoting characteristics of endophytic yeasts, their potential applications in agriculture are great. Growth promotion by a poplar endophyte *Rhodotorula graminis* has been reported in bell peppers and poplar (Khan et al. 2012; Knoth et al. 2013). Additionally, another *Rhodotorula* strain has been shown to increase germination of cucumber seedlings both through inoculation and treatment by filtered supernatant (Akhtyamova and Sattarova 2013). However, very few of these plant experiments exist as most of the research in the past has been focused on the diversity and ecology of these yeasts rather than their applications.

The use of these yeast endophytes in agriculture could reduce inputs of water and fertilizer and are important to study in the future given our growing population and increasing food demand.

A couple of the studies that showed growth promotion by endophytic yeasts focused on their potential to be used for bioaugmented phytoremediation of heavy metals. Heavy metal contaminated sites pose significant health risks and current technologies for remediation are expensive and insufficient (Deng et al. 2012; Dhankher et al. 2012). Although decontamination using plants as bioaccumulators offers a cheaper alternative that could provide potential for concurrent bioenergy production, this strategy is often inefficient given that heavy metals negatively affect plant health (Deng et al. 2012; Sheng et al. 2008). However, these negative impacts on plant health could be remediated by the addition of pollutant tolerant endophytes, termed bioaugmented phytoremediation. Deng et al. reported that a Cryptococcus sp. from rape roots not only increased the survival rate of Chinese kale seedlings in metal contaminated soil but also promoted their growth. Additionally, Wang et al. showed that Brassica sp. seedlings inoculated with endophytic Rhodotorula sp. showed increased growth and increased extraction of Cd, Pb, Zn, and Cu (Wang et al. 2013). In addition to the previously discussed advantages of using endophytic yeasts over bacteria, yeasts may also be better suited for bioaugmented phytoremediation given that contaminated sites are often contaminated by multiple organic and inorganic pollutants and fungi tend to have a broader range of tolerances to pollutants (D'Annibale et al. 2006; Deng et al. 2012).

Another promising use of endophytic yeasts in agriculture is their use as biocontrol agents against fungal and bacterial pathogens. This use of endophytes for biocontrol has been reported many times for bacterial endophytes (Ait Barka et al. 2002; Brooks et al. 1994; Compant et al. 2005; Kandel et al. 2017; Miotto-Vilanova et al. 2016). Conversely, the use of endophytic yeasts as potential biocontrol agents has not been studied until very recently. One group reported that a *Rhodotorula rubra* strain, isolated from rice, showed strong inhibition of various *Fusarium* species, which are one of the most economically important fungal pathogens (Akhtyamova and Sattarova 2013). Additionally, the yeast also showed strong inhibition of growth of *Xanthomoonas malvacearum* and *Erwinia* species, both important bacterial plant pathogens. This biocontrol ability was also found in the filtered supernatant of the yeast (Akhtyamova and Sattarova 2013). Additionally, Kandel et al. reported the antifungal activity of the poplar endophyte *Rhodotorula graminis* against the common fungal pathogen *Rhizoctonia solani* (Kandel et al. 2017).

There are a few hypotheses that explain the potential mechanisms of these biocontrol abilities. One involves the production of siderophores described earlier in this review. In addition to their potential role to promote plant growth, side-rophores that scavenge and sequester iron could prevent the establishment of fungal pathogens since other fungi need iron in order to grow and establish. This has been shown in epiphytic yeasts and bacteria (Calvente et al. 2001; Loaces et al. 2011). However, antifungal characteristics have been found in yeasts that do not produce siderophores. These antifungal characteristics seem to be fairly uncommon which points to a more specific mechanism (Nutaratat et al. 2014). A likely hypothesis is

the secretion by endophytic yeasts of cellulases and cell wall degrading enzymes, described as the killer yeast phenomenon (Magliani et al. 1997). Some yeasts produce toxins that have strong antagonistic effects on other fungi and bacteria and likely phytopathogens which could help protect their host (Magliani et al. 1997). Finally, endophytic yeasts might help exclude phytopathogens simply by occupying their niche, thus preventing deleterious fungi and oomycetes from colonizing plant tissues (Prior et al. 2017).

Endophytic yeasts could also be used for the production of a variety of biochemicals as was previously reviewed (Doty 2013). Possessing a wide variety of metabolic options given their adaptations to fluctuations in their environments and their wide host range, endophytic yeasts could provide opportunities for more efficient metabolism of various, difficult to produce, chemicals.

5 Anthropogenic Impacts on Endophytic Yeast Diversity

Recently, there has been increased interest in the research of the anthropogenic impacts on yeast diversity. Notably, Prior et al. studied the impact of fungicide on yeast communities. Fungicides are highly toxic to soil communities and cause significant stress to plants (Jorgensen et al. 2012). However, their direct impact on endophytes has not been studied until recently. This group looked closely at the impacts of two types of fungicides, contact fungicides, such as sulfur and copper, and systemic fungicides such as azoxystrobin. Sulfur and copper had a strong effect on the species richness and colony count of common bean endophytes. The systemic fungicide, azoxystrobin, had an even stronger impact on the endophytic community. Interestingly, once the fungicides leached out of the plant host, the niche inside the plant was eventually recolonized. However, this recolonization also represented a drastic shift in species diversity. This may indicate a recolonization of the plant by potentially pathogenic fungi where endophytes lived before, and could hint at an explanation for the variable success of fungicides in some crops.

Another group has focused on the changes in endophytic communities in plants close to urban areas. This group previously reported a stronger presence of human pathogenic and opportunistic yeast species in pollen in urban environments which may be responsible for increased allergic reactions (Glushakova et al. 2015). More recently, they reported significant changes in the endophytic populations of the fruits of *Malus domestica* and *Pyrus communis* (Glushakova and Kachalkin 2017). Notably, there was a significantly greater abundance of the opportunistic human pathogen *C. parapsilosis.* Overall, they reported that the anthropogenic load has significant impacts on fruit in urban areas. One could imagine potentially significant consequences to endophytic yeast communities within plants living in urban areas and endophytes potentially being outcompeted and replaced by non-native species and human opportunist pathogens. This anthropogenic impact has also been reported by Solis et al. who noticed that plants in greenhouses in Berlin that were moved across various locations had a greater diversity of endophytic yeasts as

compared to ones that were left in soil throughout the year (Solis et al. 2014). These data point to a strong anthropogenic impact on endophytic yeast communities that may have significant consequences on plant health.

6 Colonization

One of the most intriguing aspects of endophytic yeast biology is the ability of the veasts to fully colonize plants. Unlike bacteria, yeasts are not motile and do not have chemotactic capabilities towards, for example, plant root exudates. This means that yeasts that are in the soil most likely cannot colonize plants simply by swimming in the direction of the roots like bacteria do. Additionally, they do not typically form a mycelium and therefore, cannot produce hyphae that are able to grow towards their hosts. Epiphytic yeasts, in addition to being passively diffused in their environment by air and water, are dispersed by insects. A couple of specific examples include the vectoring of *Saccharomyces cerevisiae* by social wasps, as well as the sweet potato epiphyte, Candida kunwiensis, which has been found on the bumblebees that pollinate the sweet potato plants (Hong et al. 2003; Stefanini et al. 2012). Further research showed that yeasts are strongly associated with insects including in the guts of beetles and on fruit flies (Chandler et al. 2012; Suh et al. 2005). This association was extensively reviewed by Ganter, and supports the convincing hypothesis that endophytic yeasts are vectored by insects and are able to colonize their plant hosts in this way (Ganter 2006). Supporting evidence includes the recent study showing that several endophytic yeasts of rice leaves were also found in association with brown plant hoppers and beetles (Tantirungkij et al. 2015). Additionally, there have been recent studies suggesting the importance of endophytic veasts in multipartite associations between tree-associated insects, their host trees, and yeasts which were thoroughly reviewed by Moller et al. (2016). Overall, this idea offers a fairly convincing hypothesis of one of the ways endophytic yeasts are able to move around their environment and colonize different plant hosts.

The other important question is how the yeasts can systemically colonize the host plant. The endophytic yeast, *Rhodotorula graminis*, is able to colonize the entire plant, including leaf and xylem tissue when the plant is inoculated from the roots (Kandel et al. unpublished). One simple explanation is that the yeast can colonize the plant passively, simply by growing inside of the root until the population can gain access to the xylem tissue, and from there, colonize the rest of the plant as water is shuttled through the plant. Although this has not been shown directly, as plant associated fungi, endophytic yeasts likely have the enzymatic capabilities to degrade plant cell walls. This has been directly shown in bacterial and filamentous endophytes (Santoyo et al. 2016; Uzma et al. 2016).

Another interesting hypothesis is the possible role of a dimorphic life cycle of some endophytic yeasts. As discussed previously, many endophytic yeasts are basidiomycetes, a phylum which contains many plant and animal pathogens with dimorphic lifecycles including a saprobic yeast phase as well as a sexual filamentous phase (Morrow and Fraser 2009). In particular, *Cryptococcus* and *Rhodotorula*, two genera that are commonly found inside plant hosts, contain many yeast strains with observed filamentous stages.

To give a specific example, Rhodotorula graminis strain WP1 is a poplar endophyte that was the first endophytic yeast to have its genome fully sequenced (Firrincieli et al. 2015). According to recent phylogenies, this endophyte is very closely related to Rhodotorula glutinis var. glutinis (Biswas et al. 2001). Using a clever experiment with auxotrophic mutants of two different mating types of this species, this yeast was shown to form a mycelium when opposite mating types were mixed together (Banno 1967). Given the evolutionary relationship between the two yeasts, it is extremely likely that *R. graminis* is also able to form a mycelium. This is further supported by recent investigations into the mating locus of R. graminis which has a similar structure to R. glutinis (Joubert, unpublished). Additionally, genomic analysis by Maia et al. showed strong conservation between the mating locus of *R. gramins* strain WP1 and other closely related species (Maia et al. 2015). These preliminary investigations have strongly supported the presence of two separate mating types of R. graminis (Firincelli et al., unpublished). However, the significance of this sexual filamentous stage in yeast colonization needs to be evaluated. These mycelial forms could enable the active spread of the endophyte throughout the plant and allow it to colonize its host from the roots to the leaves. However, this is unlikely to play a significant role in plant colonization given that a large portion of endophytic yeasts are ascomycetes which do not form a filament and the fact that the yeast can seemingly colonize the plant in its haploid yeast stage. One thing is for certain, these filamentous sexual forms are a great source of recombination and diversity, potentially allowing the yeasts to adapt rapidly to different plant hosts and environmental conditions.

Another interesting hypothesis to explain yeast colonization is the role of the production of IAA. This hormone affects plant roots by inhibiting differentiation of plant root cells and promoting root elongation. This could positively affect plant health overall and could also provide opportunities for colonization by yeast endophytes. The main barrier to microbial colonization of the xylem is the Casparian strip in mature roots. However, if the yeast produces enough IAA in the rhizosphere, as they are capable of doing in vitro, it could prevent root cells from maturing, and therefore, decrease the presence of the Casparian strip (Kandel et al. 2017; Verbon and Liberman 2016). This could provide an avenue for colonization by the yeast. Interestingly, ACC deaminase activity also promotes root elongation and therefore might provide a similar benefit to the yeast in colonization (Glick et al. 2007).

A possible requirement to the establishment of a yeast population is the formation of a biofilm. The yeast lifestyle in general is adapted for aquatic environments, such as the plant xylem, but biofilm formation is likely a requirement for full colonization of the xylem by the yeast (Moller et al. 2016). Genes encoding a polysaccharide capsule were also found in the genome of *R. graminis* strain WP1 (Firrincieli et al. 2015). These endophyte biofilms could allow a form of protection from various environmental conditions, as well as a site of reproduction and the formation of resistance stages as they do in other yeasts (Ramage et al. 2009). This could also be a strong source of expression of plant growth-promoting characteristics of these yeasts, and potentially provide an explanation for the paradox of how yeasts are able to fight off fungal phytopathogens using antifungal molecules without harming themselves. Although this has not been directly studied in endophytic yeasts, this theory seems to be supported by evidence that endophytes have stronger endophytic characteristics in mixed biofilms including both bacteria and filamentous endophytes (Bandara et al. 2006).

7 Conclusion

While the biology of endophytic yeasts remains relatively unexplored, recent research is encouraging as to their potential for industrial and agricultural applications. While more research and especially field trials need to be done to assess this potential, it seems the use of endophytic yeasts could be a viable way of reducing fertilizer and water inputs in agriculture, and potentially increasing yields. These yeasts could also have significant advantages over their bacterial and filamentous neighbors. Their application seems especially promising in the field of remediation of heavy metal pollutants, and as biocontrol agents to protect plants from pathogens without harming the plant's microbiome. Still, many questions remain, especially when it comes to the biology and diversity of these yeasts. The way yeasts are able to colonize plants still remains a mystery, and many experiments on the process of inoculation of the host need to be done in order to elucidate this mystery. The role of insects in the life history of yeasts and their dispersal seem to be a promising avenue for research as well. Finally, many questions on the diversity of yeast species across different plant hosts still need to be answered, and the possibility of significant cultivation biases needs to be thoroughly evaluated.

References

- Ait Barka E, Gognies S, Nowak J, Audran J-C, Belarbi A (2002) Inhibitory effect of endophyte bacteria on Botrytis cinerea and its influence to promote the grapevine growth. Biol Control 24 (2):135–142. https://doi.org/10.1016/S1049-9644(02)00034-8
- Akhtyamova N, Sattarova RK (2013) Endophytic yeast Rhodotorula rubra strain TG-1: antagonistic and plant protection activities. Biochem Physiol: Open Access 02(01). https:// doi.org/10.4172/2168-9652.1000104
- Bandara WM, Seneviratne G, Kulasooriya SA (2006) Interactions among endophytic bacteria and fungi: effects and potentials. J Biosci 31(5):645–650
- Banno I (1967) Studies on the sexuality of Rhodotorula. J Gen Appl Microbiol 13(2):167–196. https://doi.org/10.2323/jgam.13.167
- Biswas SK, Yokoyama K, Nishimura K, Miyaji M (2001) Molecular phylogenetics of the genus Rhodotorula and related basidiomycetous yeasts inferred from the mitochondrial cytochrome b

gene. Int J Syst Evol Microbiol 51(Pt 3):1191–1199. https://doi.org/10.1099/00207713-51-3-1191

- Brooks DS, Gonzalez CF, Appel DN, Filer TH (1994) Evaluation of endophytic bacteria as potential biological-control agents for Oak Wilt. Biol Control 4(4):373–381. https://doi.org/10. 1006/bcon.1994.1047
- Calvente V, de Orellano ME, Sansone G, Benuzzi D, Sanz de Tosetti MI (2001) A simple agar plate assay for screening siderophore producer yeasts. J Microbiol Methods 47(3):273–279
- Chandler JA, Eisen JA, Kopp A (2012) Yeast communities of diverse Drosophila species: comparison of two symbiont groups in the same hosts. Appl Environ Microbiol 78(20):7327–7336. https://doi.org/10.1128/AEM.01741-12
- Compant S, Duffy B, Nowak J, Clément C, Barka EA (2005) Use of plant growth-promoting bacteria for biocontrol of plant diseases: principles, mechanisms of action, and future prospects. Appl Environ Microbiol 71(9):4951–4959. https://doi.org/10.1128/AEM.71.9.4951-4959.2005
- D'Annibale A, Rosetto F, Leonardi V, Federici F, Petruccioli M (2006) Role of autochthonous filamentous fungi in bioremediation of a soil historically contaminated with aromatic hydrocarbons. Appl Environ Microbiol 72(1):28–36. https://doi.org/10.1128/AEM.72.1.28-36. 2006
- Deng Z, Wang W, Tan H, Cao L (2012) Characterization of heavy metal-resistant endophytic yeast Cryptococcus sp. CBSB78 from rapes (Brassica chinensis) and its potential in promoting the growth of Brassica spp. in metal-contaminated soils. Water Air Soil Pollut 223(8):5321– 5329. https://doi.org/10.1007/s11270-012-1282-6
- Dhankher OP, Pilon-Smits EAH, Meagher RB, Doty S (2012) 20—biotechnological approaches for phytoremediation A2—Altman, Arie. In: Hasegawa PM (ed) Plant biotechnology and agriculture. Academic Press, San Diego, pp 309–328
- Doty SL (2013) Endophytic yeasts: biology and applications. In: Aroca R (ed) Symbiotic endophytes. Springer, Berlin, pp 335–343
- Firrincieli A, Otillar R, Salamov A, Schmutz J, Khan Z, Redman RS, Doty SL et al (2015) Genome sequence of the plant growth promoting endophytic yeast Rhodotorula graminis WP1. Front Microbiol 6:978. https://doi.org/10.3389/fmicb.2015.00978
- Ganter PF (2006) Yeast and invertebrate associations. In: Péter G, Rosa C (eds) Biodiversity and ecophysiology of yeasts. Springer, Berlin, pp 303–370
- Glick BR, Cheng Z, Czarny J, Duan J (2007) Promotion of plant growth by ACC deaminase-producing soil bacteria. Eur J Plant Pathol 119(3):329–339. https://doi.org/10. 1007/s10658-007-9162-4
- Glushakova AM, Kachalkin AV (2017) Endophytic yeasts in Malus domestica and Pyrus communis fruits under anthropogenic impact. Microbiology 86(1):128–135. https://doi.org/10. 1134/s0026261716060102
- Glushakova AM, Kachalkin AV, Zheltikova TM, Chernov IY (2015) Yeasts associated with wind-pollinated plants-leading pollen allergens in Central Russia. Mikrobiologiia 84(5):612–615
- Hall C, Brachat S, Dietrich FS (2005) Contribution of horizontal gene transfer to the evolution of Saccharomyces cerevisiae. Eukaryot Cell 4(6):1102–1115. https://doi.org/10.1128/EC.4.6. 1102-1115.2005
- Hardoim PR, van Overbeek LS, Elsas JD (2008) Properties of bacterial endophytes and their proposed role in plant growth. Trends Microbiol 16(10):463–471. https://doi.org/10.1016/j.tim. 2008.07.008
- Hong SG, Bae KS, Herzberg M, Titze A, Lachance MA (2003) Candida kunwiensis sp. nov., a yeast associated with flowers and bumblebees. Int J Syst Evol Microbiol 53(Pt 1):367–372. https://doi.org/10.1099/ijs.0.02200-0
- Ichinose H (2013) Cytochrome P450 of wood-rotting basidiomycetes and biotechnological applications. Biotechnol Appl Biochem 60(1):71–81. https://doi.org/10.1002/bab.1061

- Jorgensen LF, Kjaer J, Olsen P, Rosenbom AE (2012) Leaching of azoxystrobin and its degradation product R234886 from Danish agricultural field sites. Chemosphere 88(5):554– 562. https://doi.org/10.1016/j.chemosphere.2012.03.027
- Kamilova F, Kravchenko LV, Shaposhnikov AI, Azarova T, Makarova N, Lugtenberg B (2006) Organic acids, sugars, and L-tryptophane in exudates of vegetables growing on stonewool and their effects on activities of rhizosphere bacteria. Mol Plant Microbe Interact 19(3):250–256. https://doi.org/10.1094/MPMI-19-0250
- Kandel SL, Firrincieli A, Joubert PM, Okubara PA, Leston ND, McGeorge KM, Doty SL (2017) An in vitro study of bio-control and plant growth promotion potential of Salicaceae endophytes. Front Microbiol 8:386. https://doi.org/10.3389/fmicb.2017.00386
- Khalifa AY, Alsyeeh AM, Almalki MA, Saleh FA (2016) Characterization of the plant growth promoting bacterium, Enterobacter cloacae MSR1, isolated from roots of non-nodulating Medicago sativa. Saudi J Biol Sci 23(1):79–86. https://doi.org/10.1016/j.sjbs.2015.06.008
- Khan Z, Guelich G, Phan H, Redman R, Doty S (2012) Bacterial and yeast endophytes from poplar and willow promote growth in crop plants and grasses. ISRN Agron 2012:1–11. https://doi.org/10.5402/2012/890280
- Knoth JL, Kim S-H, Ettl GJ, Doty SL (2013) Effects of cross host species inoculation of nitrogen-fixing endophytes on growth and leaf physiology of maize. GCB Bioenergy 5(4):408– 418. https://doi.org/10.1111/gcbb.12006
- Loaces I, Ferrando L, Scavino AF (2011) Dynamics, diversity and function of endophytic siderophore-producing bacteria in rice. Microb Ecol 61(3):606–618. https://doi.org/10.1007/ s00248-010-9780-9
- Magliani W, Conti S, Gerloni M, Bertolotti D, Polonelli L (1997) Yeast killer systems. Clin Microbiol Rev 10(3):369–400
- Maia TM, Lopes ST, Almeida JM, Rosa LH, Sampaio JP, Goncalves P, Coelho MA (2015) Evolution of mating systems in basidiomycetes and the genetic architecture underlying mating-type determination in the yeast Leucosporidium scottii. Genetics 201(1):75–89. https:// doi.org/10.1534/genetics.115.177717
- Marcet-Houben M, Gabaldon T (2010) Acquisition of prokaryotic genes by fungal genomes. Trends Genet 26(1):5–8. https://doi.org/10.1016/j.tig.2009.11.007
- Miotto-Vilanova L, Jacquard C, Courteaux B, Wortham L, Michel J, Clément C, Sanchez L et al (2016) Burkholderia phytofirmans PsJN confers grapevine resistance against Botrytis cinerea via a direct antimicrobial effect combined with a better resource mobilization. Front Plant Sci 7:1236. https://doi.org/10.3389/fpls.2016.01236
- Moller L, Lerm B, Botha A (2016) Interactions of arboreal yeast endophytes: an unexplored discipline. Fungal Ecol 22:73–82. https://doi.org/10.1016/j.funeco.2016.03.003
- Morrow CA, Fraser JA (2009) Sexual reproduction and dimorphism in the pathogenic basidiomycetes. FEMS Yeast Res 9(2):161–177. https://doi.org/10.1111/j.1567-1364.2008. 00475.x
- Nassar AH, El-Tarabily KA, Sivasithamparam K (2005) Promotion of plant growth by an auxin-producing isolate of the yeast Williopsis saturnus endophytic in maize (Zea mays L.) roots. Biol Fertil Soils 42(2):97–108. https://doi.org/10.1007/s00374-005-0008-y
- Nutaratat P, Srisuk N, Arunrattiyakorn P, Limtong S (2014) Plant growth-promoting traits of epiphytic and endophytic yeasts isolated from rice and sugar cane leaves in Thailand. Fungal Biol 118(8):683–694. https://doi.org/10.1016/j.funbio.2014.04.010
- Prior R, Mittelbach M, Begerow D (2017) Impact of three different fungicides on fungal epi- and endophytic communities of common bean (Phaseolus vulgaris) and broad bean (Vicia faba). J Environ Sci Health B 52(6):376–386. https://doi.org/10.1080/03601234.2017.1292093
- Ramage G, Mowat E, Jones B, Williams C, Lopez-Ribot J (2009) Our current understanding of fungal biofilms. Crit Rev Microbiol 35(4):340–355. https://doi.org/10.3109/ 10408410903241436
- Santoyo G, Moreno-Hagelsieb G, Orozco-Mosqueda Mdel C, Glick BR (2016) Plant growth-promoting bacterial endophytes. Microbiol Res 183:92–99. https://doi.org/10.1016/j. micres.2015.11.008

- Sheng XF, Xia JJ, Jiang CY, He LY, Qian M (2008) Characterization of heavy metal-resistant endophytic bacteria from rape (Brassica napus) roots and their potential in promoting the growth and lead accumulation of rape. Environ Pollut 156(3):1164–1170. https://doi.org/10. 1016/j.envpol.2008.04.007
- Solis MJL, Yurkov A, dela Cruz TE, Unterseher M (2014) Leaf-inhabiting endophytic yeasts are abundant but unevenly distributed in three Ficus species from botanical garden greenhouses in Germany. Mycol Prog 14(1):1019. https://doi.org/10.1007/s11557-014-1019-6
- Stefanini I, Dapporto L, Legras JL, Calabretta A, Di Paola M, De Filippo C, Cavalieri D et al (2012) Role of social wasps in Saccharomyces cerevisiae ecology and evolution. Proc Natl Acad Sci U S A 109(33):13398–13403. https://doi.org/10.1073/pnas.1208362109
- Suh SO, McHugh JV, Pollock DD, Blackwell M (2005) The beetle gut: a hyperdiverse source of novel yeasts. Mycol Res 109(Pt 3):261–265
- Sun Y, Cheng Z, Glick BR (2009) The presence of a 1-aminocyclopropane-1-carboxylate (ACC) deaminase deletion mutation alters the physiology of the endophytic plant growth-promoting bacterium Burkholderia phytofirmans PsJN. FEMS Microbiol Lett 296 (1):131–136. https://doi.org/10.1111/j.1574-6968.2009.01625.x
- Takahashi T, Kakehi J (2010) Polyamines: ubiquitous polycations with unique roles in growth and stress responses. Ann Bot 105(1):1–6. https://doi.org/10.1093/aob/mcp259
- Tantirungkij M, Nasanit R, Limtong S (2015) Assessment of endophytic yeast diversity in rice leaves by a culture-independent approach. Antonie Van Leeuwenhoek 108(3):633–647. https:// doi.org/10.1007/s10482-015-0519-y
- Uzma F, Konappa NM, Chowdappa S (2016) Diversity and extracellular enzyme activities of fungal endophytes isolated from medicinal plants of Western Ghats, Karnataka. Egypt J Basic Appl Sci 3(4):335–342. https://doi.org/10.1016/j.ejbas.2016.08.007
- Verbon EH, Liberman LM (2016) Beneficial microbes affect endogenous mechanisms controlling root development. Trends Plant Sci 21(3):218–229. https://doi.org/10.1016/j.tplants.2016.01. 013
- Wang W, Deng Z, Tan H, Cao L (2013) Effects of Cd, Pb, Zn, Cu-resistant endophytic Enterobacter sr CBSB1 and Rhodotorula sp. CBSB79 on the growth and phytoextraction of Brassica plants in multimetal contaminated soils. Int J Phytorem 15(5):488–497. https://doi. org/10.1080/15226514.2012.716101
- Xin G, Glawe D, Doty SL (2009) Characterization of three endophytic, indole-3-acetic acid-producing yeasts occurring in Populus trees. Mycol Res 113(Pt 9):973–980. https://doi. org/10.1016/j.mycres.2009.06.001