

Heat Shock Proteins in Cardiovascular Diseases: From Bench to Bedside



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Abstract Heat shock proteins (HSP) are stress proteins induced in response to a wide variety of physiological and environmental insults. HSP function as molecular chaperones and they are required to maintain the proteome in a folded and functional state, allowing the cells to survive stress conditions. These key proteins, which may be located intracellularly or extracellularly, have multiple functions that range from the regulation of essential cells function to the renaturation of misfolded proteins. In the last decades, the HSP involvement in both normal cell function and disease pathogenesis is widely studied, especially in the context of cardiovascular diseases (CVDs). This chapter covers the current knowledge on the function HSP in the cardiovascular system and particular in the relationship between these proteins and CVDs. Initially, the roles of HSP in cardiovascular health are outlined, followed by an evaluation of the role of HSP in CVDs key processes, such as atherosclerosis, vascular hypertrophy and heart failure. Finally, the therapeutic potential of roles HSP are examined in a CVDs context, considering how the knowledge actually gained may be capitalized in future clinical studies.

Keywords Cardiovascular diseases · Heat shock proteins · Hsp40 · Hsp60 · Hsp70 · Hsp90 · Small heat shock proteins · Therapeutics

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Abbreviations

AIF	apoptosis inducing factor
ApoE ^{-/-}	apolipoproteinE knock out mice
Ca ²⁺	calcium
CEL	celastrol
CVDs	cardiovascular diseases
GGA	geranylgeranylacetone
HSEs	heat shock elements
Hsf1	heat transcription factor 1
HSP	heat shock protein
LDL-C	low-density lipoprotein-cholesterol
RIPC	remote ischemic preconditioning
ROS	reactive oxygen species
sHSP	small heat shock protein
sHSP60	soluble heat shock protein60
SMCs	smooth muscle cells
TLR4s	toll-like receptors 4
VSMCs	vascular smooth muscle cells

Introduction

Nowadays, despite a variety of therapeutic advances, cardiovascular diseases (CVDs) remain a leading cause of mortality worldwide (Kessing et al. 2016). CVDs include various diseases that affect heart and/or blood vessels, such as coronary artery diseases like angina and myocardial infarction. Heat shock proteins (HSP) are stress proteins, also called chaperones that are massively produced by almost all prokaryotic and eukaryotic species when exposed to elevated temperatures. These proteins were discovered in 1962 by Ritossa in the *Drosophila melanogaster* as a set of highly conserved proteins whose expression was induced in salivary gland chromosome in response to transient exposures to elevated temperatures (Kliková et al. 2016; Ritossa 1962). The increased expression of HSP in cells during the heat shock response was demonstrated to inhibit stress-mediated cellular death as shown also by recent experiments indicating a highly versatile role for these proteins (Garrido et al. 2006). These functions are summarized in Fig. 1. HSP play an important role in cell-cycle control, folding, defolding and assembling of protein complexes (Nakai and Ishikawa 2001) and they have been classified into six families on the basis of their approximate molecular weight: small HSP, HSP40, HSP60, HSP70, HSP90 and HSP100 (Khalil et al. 2011). These proteins constitute the 5–10% of the total protein content in cells in physiological conditions and play important roles in cellular homeostasis. They control maturation and turnover of intracellular proteins and play significant roles in the maintenance of cellular integrity.

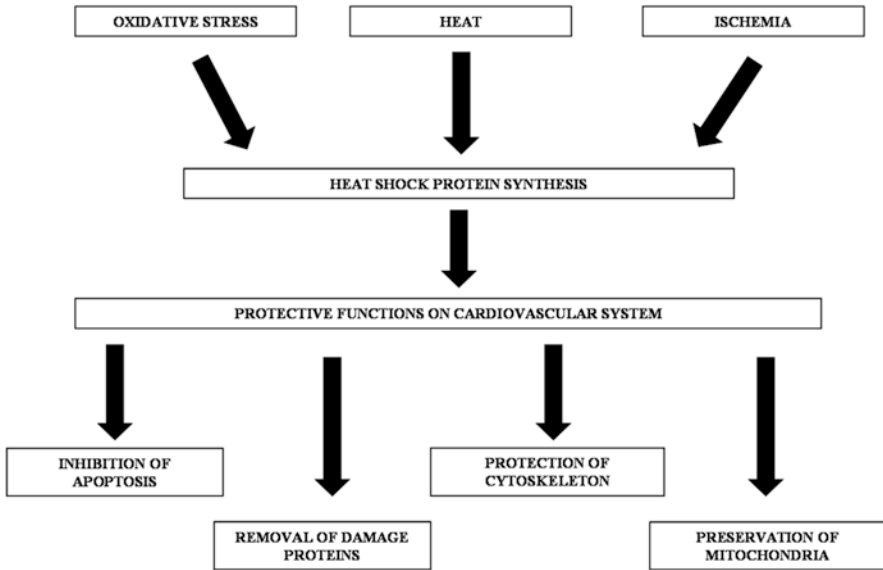


Fig. 1 Main heat shock proteins protective functions at cardiovascular level

Interestingly, the synthesis of these proteins can be markedly induced by various cellular insults ranging from elevation in temperature to exposure to different stimuli which are potentially damaging to the cell (Pockley 2002). These latter conditions include several known risk factors for CVDs onset and development. It is clear, however, from the extensive studies which have been carried out in a variety of cell types that the HSP are induced in response to cellular stress and they confer protection against nonthermal cytotoxic stimuli. A recent *in vitro* experiment demonstrated that induction of the stress response protects endothelial cells against superoxide generation (Chen et al. 2017). Other examples *in vivo* include protection of cardiomyocytes against apoptosis, induced by ischemia/reperfusion injury or by prolonged beta-agonist stimulation (Fan et al. 2004, 2005). Even if new functions continue to be discovered, the stress proteins are generally thought to maintain cellular homeostasis by acting as molecular chaperones, facilitating the proper folding and assembly of nascent polypeptides, as well as assisting in the refolding and stabilization of damaged peptides (Bond and Schlesinger 1985). HSP act as intracellular chaperones, protecting protein structure and folding under stress condition, but are also able to assist with numerous reparative processes including the refolding of denatured proteins and removal of irreparably damaged proteins (Kampinga and Craig 2010). Under particular conditions HSP could be released from cells into the extracellular space, where they exhibit a range of immunoregulatory activities (Chebotareva et al. 2017).

To date numerous studies suggest that the upregulation of HSP could be associated with cardioprotection (Jimenez et al. 2014). In fact, before myocardial infarction, an increase in HSP70 and HSPB1 levels induced through an infection with

Table 1 Heat shock proteins involvement in cardiovascular diseases

Superfamily	Protein names	Alternative names	Low protein expression levels or mutations
shSPs	HSPB1	HSP27,HSP25, HMN2B	Coronary artery disease
	HSPB2	MKBP	Altered mitochondria functions
	HSPB3	HSPL27	Deregulation of myoblast viability
	HSPB5	alphaB-crystallin	Desmin-related myopathy
	HSPB6	HSP20	Decreased contractile function
	HSPB7	CvHSP	No protection against age-related dysfunction
	HSPB8	H11, HSP22	Lack of protection after myocardial ischemia
	HSP40	DNAJA3	Tid-1; Tid11
HSP60	HSP60	Cpn60, Hsp60	Protection from atherosclerosis and proliferation of vascular smooth muscle cells
HSP70	HSPA1A	HSP72; HSPA1	Plaque rupture and thrombotic complications
	HSPA8	HSC70; HSC71	
HSP90	HSPC4	GP96, endoplasmic	Increased in plaque stability
			Cardiomyocyte apoptosis

replication deficient adenovirus encoding for these two HSP has been shown to protect heart (Wei et al. 2006). Moreover, Chen et al. (2005) showed that along with the rise in reactive oxygen species (ROS) levels, there was a significant increase in the heart levels of HSP (HSP70, HSPB1) in response to remote ischemic preconditioning (RIPC). Moreover, treatment with antioxidants reduced the levels of HSP and also attenuated cardioprotection (Chen et al. 2005). These results suggest that HSP production may be induced in condition characterized by an increase in free radical production and this enhancement, in turn, may confer resistance to myocardium against sustained ischemia reperfusion injury. Furthermore, Madamanchi et al. (2001) in an in vitro study showed that upregulation of HSP may be secondary to increase in ROS in vascular smooth muscle cells. In contrast, other studies showed that HSP may stimulate the production of ROS (Baruah et al. 2014; Multhoff et al. 2015). So, more studies are required to better clarify the possible interaction between heat shock proteins HSP in cardioprotection (Singh et al. 2017). Table 1 summarizes the main involvement of H HSP in CVDs. In the following sections, we present an overview of the HSP roles and functions in cardiovascular health conditions and their implication in the onset and development of cardiovascular diseases, firstly we presented the small HSP, characterised by a low molecular weight (15–43 kDa), and then the HSP with a higher molecular weight (HSP40, HSP60, HSP70, HSP90). Finally, we provided a brief summary about potentially preventive and therapeutic approaches that involved HSP modulation.

Emerging Roles of Heat Shock Proteins in Cardiovascular Diseases

Small Heat Shock Proteins

Small HSP (sHSP), also called HSP β (HSPBs), have molecular weight which varied from 15 to 43 kDa and are mainly known for their chaperoning function in the process of embryonic development (Jee 2016). This family of HSP consists of ten members (HSPB1-HSPB10), which are characterized by different functions and display different expression profiles. Some members of this family are widely expressed, like HSPB1, HSPB5 and HSPB8, while some other members show exclusively a restricted expression pattern. In particular, HSPB2 and HSPB3 are only found in cardiac and muscle cells, HSPB4 is lens specific, while HSPB9 and HSPB10 are specifically expressed in testis (Clark et al. 2012; Fontaine et al. 2003; Kappe et al. 2003; Sugiyama et al. 2000; Suzuki et al. 1998; Verschuure et al. 2003). Some members of this family, such as HSPB1 and HSPB4, exert mainly refolding activities, while other members display only anti-aggregation function (HSPB6, HSPB7 and HSPB8). Furthermore, other members exert very specialized activities, for example HSPB2 and HSPB3 seem to have an essential role in muscle differentiation, while HSPB1, HSPB5 and HSPB7 show very specific chaperone functions at the levels of nuclear structures forming during stress (Boncoraglio et al. 2012).

sHSP present a highly conserved sequence of 80–100 amino acids called the α -crystallin domain. This structural domain is responsible for many intra- and inter-molecular interactions leading to the formation of dimers, which are considered as sHSP basic unit (Kim et al. 1998; Van Montfort et al. 2001a, b). These dimers can interact with each other forming higher molecular weight oligomers. Besides the α -crystallin domain, both the C-terminal and the N-terminal regions are involved in the stabilization of the oligomers (Kim et al. 1998; Lambert et al. 1999; Van Montfort et al. 2001a, b). However, besides these different aspects, ten members of the HSPB family share the same properties and so alteration of these properties represent a key mechanism that can lead to diseases onset, also at cardiovascular level.

Cardiac and skeletal muscle cells express the largest variety of HSPBs: HSPB1, HSPB2, HSPB3, HSPB5, HSPB6, HSPB7 and HSPB8. These family members are commonly considered the main focus of interest of CVDs researches and new scientific evidences emphasize the functions of these molecules in cardioprotection (Carra et al. 2017; Charmpilas et al. 2017; Edwards et al. 2011; Fan et al. 2005; Martin et al. 2014; Weintraub and Rubinstein 2013).

HSPB1 is a ubiquitously expressed, multifunctional protein chaperone. A decrease in HSPB1 secretion was shown in human atherosclerotic plaques respect to control vessels and also plasmatic HSPB1 levels are reduced in atherosclerotic patients compared to healthy subjects (Martin-Ventura et al. 2004). Moreover,

Seibert et al. (2013) showed that in human low serum HSPB1 levels are associated with coronary artery disease and prognostic of adverse clinical events; while in mouse models of atherosclerosis the increase in HSPB1 level in aorta induce a reduction of atherosclerotic lesion progression and promote plaque stability. It was also shown that HSPB1 overexpression in atherosclerotic-prone mouse models (apolipoproteinE knock out mice – ApoE^{-/-}) modulates plaque formation preventing inflammation, foam cells formation and reducing atherosclerotic plaque area (Rayner et al. 2008). In heart tissue HSPB1 expression was associated with sarcomeres and so it was found to be cardioprotective (Brundel et al. 2006a, b). Moreover, Ghayour-Mobarhan et al. (2012) reported that increased expression of myocardial HSPB1 is a prognostic marker of myocardial ischemia. HSPB1 has been demonstrated to play also an important role in regulating intracellular redox homeostasis and anti-apoptotic pathway (Arrigo et al. 2005; Zhang et al. 2010). Panneerselvam et al. (2017) observed that differential expression of HSP in the rat myocardium could serve as a balance between pro-survival and death signal during acute fluoride-induced heart failure. In detail, the Authors suggested that increased expression of myocardial HSPB1 serves as a prognostic marker for myocardial ischemia and this increase is correlated with myocardial necrosis, impaired contractile function and regulation of intracellular redox homeostasis.

HSPB2 has shown to be highly expressed in heart and it has protective effects against heart diseases such as cardiac hypertrophy and ischemia (Ishiwata et al. 2012; Nakagawa et al. 2001; Sugiyama et al. 2000). Moreover, HSPB2 was associated with the outer membrane of mitochondria and involved in mitochondria permeability transition and calcium uptake. Overexpression of this sHSP was found to conserve ATP synthesis during ischemic/reperfusion injury in mice (Nakagawa et al. 2001). Moreover, mice knockout for HSPB2 with ischemic stress show altered mitochondria respiration rates and reduced ATP production as well as modification in expression of several metabolic and mitochondrial regulators (Ishiwata et al. 2012). These findings suggest that HSPB2 has cardioprotective effects maintaining mitochondrial function and metabolic activity during cardiac stress.

Golenhofen et al. (2006) showed that the cytosolic calcium increase due to the lack of HSPB2 in knockout animals may modify the calcium sensitivity of myofibrils altering the cardiac contractility, suggesting an involvement of HSPB2 in maintaining muscular elasticity during ischemic insult. Interestingly, mice with HSPB2 overexpression in heart revealed lower levels of cardiac injury biomarker troponin I in the blood after ischemia/reperfusion stress, confirming the involvement of HSPB2 in preserving contractile function of the heart (Grose et al. 2015).

HSPB3 was considered not expressed in the heart (Vos et al. 2009), but recently Carra et al. (2017) reported in patients with myopathy two novel mutations in the HSPB3 gene that lead respectively to protein aggregation or truncation and destabilization. Remarkably, these mutations could abolish HSPB2-HSPB3 complex formation, leading to free HSPB2 proteins that tend to mislocalize inside the cells. Moreover, the same authors reported that HSPB2-HSPB3 interaction would lead to deregulation of HSPB2, with potential consequences on myoblast function and viability. These studies suggested that alterations in expression and solubility of

specific HSPBs due to aggregation propensity and deregulated association with other HSPBs could be the basis of complex diseases, such as myopathies.

HSPB5 was found in cardiomyocytes on the I-band and M-line region of sarcomeres (van de Klundert et al. 1998). It is known to bind and stabilize intermediate filaments, actin microfilaments, and sarcomeric proteins, including actin, desmin and titin in physiological conditions (Bullard et al. 2004; Ghosh et al. 2007; Perng et al. 1999). HSPB5 has shown to be involved also in stabilization of the cytoskeleton (Vicart et al. 1998). Interestingly, mutations in HSPB5 are associated with cardiac and muscular disorders. In fact, HSPB5 mutations result in an irregular protein structure and defective chaperone-like function (Bova et al. 1999), which leading to desmin-related myopathy and also early onset of cardiomyopathy (Selcen and Engel 2003; Vicart et al. 1998).

HSPB6 is abundantly expressed in skeletal muscle and heart in two complex formations: 43 kDa dimers and 470 kDa multimers and that this protein is able to bind itself and other HSPBs, like HSPB1, HSPB5 and HSPB8 (Pipkin et al. 2003). Recently, in a more recent study, the overexpression of HSPB6 resulted in enhanced cardiac function by interaction with protein phosphatase 1 and in turn inducing calcium (Ca^{2+}) cycling and sarcoplasmic reticulum Ca^{2+} load (Qian et al. 2011). In addition, this stress protein induces the Ca^{2+} cycling in the sarcoplasmic reticulum and increases the contractile function of the cardio myocyte (Qian et al. 2011). Further, in an experimental study in mice, the phosphorylation of HSPB6 at serine 16 level was shown to be needed for modulating cell injury during ischemia/reperfusion (Qian et al. 2009). So, HSPB6 acts maintaining the heart integrity in mice with ischemia/reperfusion injury (Fan et al. 2005).

HSPB7 is expressed in heart and skeletal muscle. It was shown that HSPB7 is significantly expressed in aged muscle like to HSPB5 (Doran et al. 2007). HSPB7 was also shown to be upregulated in the muscular dystrophy-affected diaphragm, suggesting its induction under stress conditions. Additionally, HSPB7 is involved in protections of cells from protein aggregation, probably by facilitating cargo delivery to autophagosomes (Vos et al. 2010). To note, HSPB4, HSPB6 or HSPB7 could not promote the cellular capacity to chaperone heat-denatured luciferase as shown by HSPB1, indicating different functions and properties of HSPB members (Vos et al. 2010, 2011). Additionally, in cardiomyocytes was observed the co-localization of HSPB7 on myofibrils, suggesting a protective role maintaining the sarcomeric structure (Golenhofen et al. 2004).

HSPB8 is showed in striated and smooth muscles, brain, and keratinocytes level (Vos et al. 2008). This protein can be phosphorylated *in vitro*, but, respect to HSPB1 and HSPB5, HSPB8 phosphorylation influences only marginally its tertiary and quaternary structure. HSPB8 exists, both in its wild type and phosphorylated form, as low molecular mass oligomers. Respect to HSPB1 and HSPB5 that showed reduced oligomeric size and increased chaperone activity after phosphorylation, the phosphorylation of HSPB8 results in larger oligomeric structures and decreased chaperone activity (Basha et al. 2006). In *in vitro* experiments, it has been shown that HSPB8 interacts with several proteins and forms stoichiometric complexes (Carra et al. 2008a). This complexes was found to induce autophagy, which may be benefi-

cial in response to irreparable protein damage (Carra et al. 2008b, 2009). In addition, HSPB8 has been shown to be cardioprotective in experimental models of myocardial ischemia. In detail, Depre et al. 2006 showed that HSPB8 overexpression promotes cardiomyocyte survival after ischemia in mice, while Chen et al. 2011 indicated that this sHSP attenuates the myocardial damage and contractile dysfunction in experimental animals. On the other hand, it was demonstrated that depletion of HSPB8 in mice with pressure overload supports cardiac dysfunction and promotes transition to heart failure (Qiu et al. 2011). Other studies showed that HSPB8 maintains mitochondrial function and energy production that results in attenuation of oxidative stress in infarcted hearts (Marunouchi et al. 2014). In contrast these beneficial HSPB8 effects on cardiomyocyte function, HSPB8 overexpression, both in in vitro and in vivo model systems, was found to induce cardiac hypertrophy (Depre et al. 2002; Hedhli et al. 2008). Therefore, the function of HSPB8 seems to have two faces in heart diseases: HSPB8 acts as a cardioprotective protein during myocardial ischemia by conserving the mitochondrial function and energy production but, at the same time, HSPB8 is a mediator of cardiac hypertrophy and thereby results in heart failure.

HSP60

HSP60 protein is structurally highly conserved and abundantly expressed by prokaryotic and eukaryotic cells under stress conditions. It is considered a constitutively cytosolic protein that translocated to the mitochondria (Gething and Sambrook 1992), but it is also stress inducible, indeed ischemia has been demonstrated to be a potent inducer (Marber et al. 1993). During heart failure this protein is translocated to the plasma membrane and released into the plasma; in fact was shown that HSP60 levels were doubled in end-stage heart failure (Knowlton et al. 1998; Lin et al. 2007). Some studies reported that high titers of anti-HSP60 were correlated with coronary atherosclerosis (Prohászka et al. 2001; Zhu et al. 2001), moreover high levels of anti-HSP60 in children increase the probability to develop CVDs (Cohen and Young 1991). Other studies suggested that plasmatic levels of HSP60 was increased early in heart failure (Brundel et al. 2006a, b). During heart failure, HSP60 localizes to the plasma membrane and the cell surface, in addition to its normal distribution to the mitochondria and cytosol (Wang et al. 2010) and acts as a “danger signal”. This localization of HSP60 could be linked with an increase in apoptosis of the affected cell, because the cell-surface expression of HSP60 may be able to interact with other cells to trigger the innate immune response, resulting in the release of pro-inflammatory cytokines which could induce myocyte loss and contribute to heart disease. Previous studies (Malik et al. 2013; Gupta and Knowlton 2007) demonstrated that HSP60, via exosomal pathway, was released by cardiomyocytes and this stress protein was increased by exposure of the cardiomyocytes to ethanol. Li et al. (2011) reported that, HSP60 induced inflammation in the heart, via activation

of the TLR4- MyD88-IRAK-1 pathway, and led to cardiomyocyte apoptosis, promoting progression of heart failure.

Some studies in human associated HSP60 expression with atherosclerosis and its severity and it was detected on endothelium, smooth muscle cells (SMCs) and mononuclear cells (Kleindienst et al. 1993; Lamb et al. 2002). Other studies reported a positive association between titers of antibody to HSP60 and the extent of atherosclerosis (Burian et al. 2001; Veres et al. 2002; Xu et al. 2000; Zhao et al. 2015; Zhu et al. 2001). Xiao et al. (2005) reported that soluble HSP60 (sHSP60) is probably involved in activating proinflammatory processes associated with early vessel pathology providing the first prospective data confirming an association between elevated levels of sHSP60 and early carotid atherosclerosis. Moreover, studies in cholesterol-fed rabbits showed that plasma titers of anti-HSP60 are consistent with atherosclerotic formation and increased expression of HSP60 on the endothelium during atherogenesis (Ghayour-Mobarhan et al. 2007; Khan et al. 1998; Lamb et al. 2002; Pfister et al. 2005; Zhao et al. 2015). Recently, in vitro experiments have shown that HSP60 can activate the proliferation of vascular SMCs (VSMCs) (de Graaf et al. 2006; Fukuoka et al. 2004; Sasu et al. 2001; Zhao et al. 2015). Moreover, Zhao et al. (2015), in an experimental study, suggested that HSP60 is implicated in the VSMC migration during atherosclerosis. The authors reported that activation of HSP60 could be considered one of the most powerful methods of sending a ‘danger signal’ to the immune system to generate chemokine, such as interleukin-8, which is involved in stimulation of VSMC migration.

HSP70

HSP70 is one major member of HSP family, which is able to bind its substrate through the substrate binding domain, dephosphorylate substrate and in turn maintain its “normal” functions (Connarn et al. 2014). This HSP have some function in the cells including binding to partially synthesized peptide sequences, keeping translocation competent structures of endoplasmic reticulum and mitochondrial precursors in the cytosol and simplify translocation from inside the target compartment (Craig et al. 1994; Mamipour et al. 2017). Previous studies have provided evidences that during myocardial ischemia/reperfusion HSP70 had protective effects and it was able to reduce the deleterious effects characteristics of this pathological condition (Feng et al. 2014; Garrido et al. 2006; Vicencio et al. 2015). In particular, during myocardial ischemia/reperfusion HSP70 accelerates the dephosphorylation of the stress kinase JNK inducing cardiac protection (Gabai et al. 2000). Moreover, in this pathological condition HSP70 is able to prevent the nucleus translocation of apoptosis inducing factor (AIF), acting as a molecule chaperone, so it stabilizes nucleus and antagonizes cardiomyocyte apoptosis (Choudhury et al. 2011). Other data confirming this action showed that the deletion of the domain of AIF involved in interaction with HSP70 enhanced AIF translocation to nucleus inducing apoptosis (Gurbuxani et al. 2003; Zhang et al. 2017). Recently, Vicencio et al. (2015) showed that plasma exosomes,

which carry HSP70 on their surfaces, attenuated myocardial reperfusion injury in rats. In *in vitro* studies, exosomes derived from plasma, containing HSP70, bound to toll-like receptors 4 (TLR4s) on cardiomyocytes and activated various kinases leading to cardioprotection and suggesting the therapeutic use of exosomes for reperfusion injury. Mayr et al. (1999) showed that, in human atherosclerotic aorta, HSP70 is expressed with a heterogeneous pattern respect to control vessels. This HSP70 altered pattern could be due to its release from damaged cells into the plaque. Mazzaferro et al. (2003) hypothesized that insufficient HSP70 in SMC of the aorta leads to their death and favors plaque rupture and thrombotic complications. Moreover atherosclerotic plaque thickness was correlated with the HSP70 expression to the center of the lesion (Pockley et al. 2003). Other studies attributed to HSP70 a protective role in atherosclerosis due to its pro-survival effect on VSMCs. It was demonstrated that HSP70 localization changed during plaque development and progression and was positively related to atherosclerosis severity (Dybdahl et al. 2005; Johnson et al. 1993; Pockley et al. 2003). Furthermore Zhu et al. (2003) have reported that there is an inverse relation between HSP70 expression and severity of coronary disease.

HSP90

The 90 kDa heat shock protein, HSP90, is one of the most abundantly expressed protein in eukaryotic cells, comprising 1–2% of cellular proteins under physiological conditions. It contributes to various cellular processes as signal transduction, protein folding and degradation and morphological evolution (Csermely et al. 1998; Pearl and Prodromou 2001; Picard 2002; Pratt and Toft 2003; Richter and Buchner 2001; Sreedhar et al. 2004). The activity of HSP90 is in cooperation with other co-chaperones, playing a key role in the folding of newly synthesized proteins and stabilization and refolding of denatured proteins during stress conditions (Sreedhar et al. 2004). Some studies indicated that HSP90 plays an important protective role in ischemic pre-conditioning and post-conditioning, acting as a cardioprotective and antiapoptotic protein (Amour et al. 2009; Zhong et al. 2014). Moreover, targeted overexpression of HSP90 protects myocardial reperfusion injury reducing infarct size and myocardial dysfunction (Kupatt et al. 2004). On the other hand, Wang et al. (2009) showed that HSP90 inhibition increases cardiomyocyte apoptosis. It has also been shown that, in isolated hearts, HSP90 attenuates ischemia/reperfusion-induced cardiomyocyte apoptosis and necrosis (Budasz et al. 2010). Moreover, recent reports demonstrated that HSP90 is anti-apoptotic in cardiomyocytes acting as antioxidant and preserving mitochondrial function under stress conditions (Wu et al. 2012).

Previously, a human study demonstrated augmented systemic HSP90-specific cellular and humoral responses in carotid atherosclerosis patients (Mu et al. 2017; Rigano et al. 2007). Furthermore, other authors showed that HSP90 is highly expressed in plaque and serum from patients with atherosclerosis, in which immune responses are stimulated, suggesting that HSP90 is a potential target in the pathogenesis of carotid atherosclerosis (Businaro et al. 2009; Mu et al. 2017). Recently, Mu et al. (2017) showed

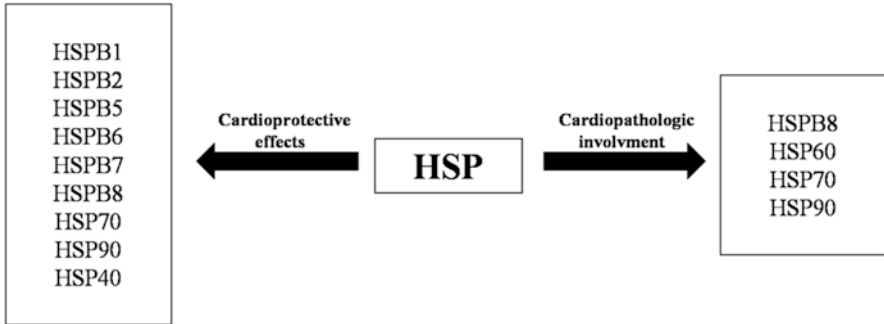


Fig. 2 Schematic representation of heat shock proteins involvement in cardioprotective and/or cardiopathological processes

that in a mouse model of atherosclerosis HSP90 overexpression in vulnerable carotid plaque induced a decrease in plaque stability, along with increased accumulation of lipids, macrophages, inflammation, as well as a decrease in VSMCs and collagen, while HSP90 inhibition exerted opposing effects. So, these authors suggested that regulation of the local HSP90 expression could be a new route for atheroprotection.

HSP40

HSP40 is the co-chaperone of HSP70. This heat shock protein has also been shown to be involved in the pathology of dilated cardiomyopathy that is an important cause of sudden cardiac death and heart failure (Hayashi et al. 2006; Henderson and Pockley 2012). In fact, inactivation of the gene encoding mitochondrial HSP40 in mice resulted in the development of severe cardiomyopathy which induced death probably due to the role played by HSP40 in mitochondrial biogenesis (Hayashi et al. 2006). Hayashi et al. (2006) showed that in cardiomyocytes lacking HSP40 there were progressive respiratory chain deficiency and reduced copy number of mitochondrial DNA, underlining the key role of this protein in CVDs. Figure 2 summarizes the HSP involvement in cardioprotective/cardiopathological processes.

Heat Shock Proteins Modulation as a Potential Therapeutic Target

The broad function on cardiovascular system discussed above strongly sustains the notion that strategies aimed at modulating HSP expression may be important for preserving CVDs onset and related symptoms. In fact, some experimental studies have tested the hypothesis that HSP modulation improves the outcome of CVDs,

particularly in the context of ischemia/reperfusion injury. The modulations of HSP can be performed in different ways such as by the use of small molecules able to modulate HSP expression or through their activation by phosphorylation. The induction of chaperones in response to different stress conditions are mediated at the transcriptional level by the heat transcription factor 1 (Hsf1) that binds to its target sites (heat shock response elements) in the promoter regions of stress-induced genes and thus, following Hsf1 phosphorylation, induces HSP gene expression (Westerheide et al. 2012). In physiological conditions, without stress, Hsf1 is retained in an inactive state in the cytoplasm binded to HSP90. Stress conditions induce dissociation of HSP90 from Hsf1 that, in turn translocate to the nucleus where it up-regulates transcription of chaperone genes. Although the HSP were identified on the basis of their induction by stressful procedures, from a clinical standpoint there are several drugs and natural compounds that modulates HSP and could be beneficial in the treatment of CVDs.

Geranylgeranylacetone (GGA) is a cyclic polyisoprenoid used as a gastromucoprotective drug. In experimental conditions, it has been showed that this pharmacological compound can induce transcriptional activation of HSP70 (Chang et al. 2013; Hirakawa et al. 1996) and that ischemia/reperfusion-induced damage of myocardial cells was prevented in GGA-treated myocytes (Chang et al. 2013; Ooie et al. 2001). Moreover, GGA upregulates HSP expression in heart and attenuates ischemia/reperfusion injury, degradation of myofibrils, and atrial fibrillation genesis caused by rapid atrial pacing or atrial ischemia (Brundel et al. 2006a, b; Chang et al. 2013; Ooie et al. 2001). Recently, Chang et al. (2013) showed that HSP induced by GGA can regulate the atrial arrhythmogenesis in heart failure by modulation of sodium and potassium channels, as well as calcium homeostasis (Chang et al. 2013).

HSP co-inducer arimoclomol has been tested in a number of clinical trials (Cudkowicz et al. 2008; Lanka et al. 2009; Kirkegaard et al. 2016). Arimoclomol belongs to a group of HSP-modulating drugs that act as inducers of HSP70, whose mechanism of action involves stabilization of the interaction of HSF1 with heat shock elements (HSEs), the transcriptional elements controlling HSP production (Anckar and Sistonen 2011; Crul et al. 2013; Kieran et al. 2004; Kirkegaard et al. 2016; Neef et al. 2011; Parfitt et al. 2014; Vigh et al. 1997).

Celastrol (CEL), a quinone methide triterpene, is a derived from the Celastraceae family of plants, that is used in traditional Chinese medicine. This compound has been shown to induce HSF1 and HSP70 expression in skeletal muscle cells (Gwag et al. 2013; Trott et al. 2008; Westerheide et al. 2004). Furthermore, this compound has been demonstrated to exhibit a broad range of functions like antioxidant, anti-inflammatory (Gwag et al. 2013; Lee et al. 2006; Sassa et al. 1990; Trott et al. 2008) and neuroprotective activity (Franklin et al. 2005; Gwag et al. 2013). All of these activities could be related to its action s HSP inducer.

Moreover, Deane and Brown (2016) in a recent in vitro research on neuronal cells showed that co-application of celastrol and arimoclomol induced higher HSP levels compared to heat shock paired with arimoclomol. This co-application targets multiple alterations including protein misfolding, protein aggregation, inflammation and oxidative stress. Several studies demonstrated the beneficial effects of

Table 2 Pharmacological compounds that modulate heat shock proteins and relative beneficial effects

Compounds	Heat shock proteins modulation	Beneficial effects
GGA	↑HSP70	Prevents ischemia/reperfusion-induced damage at myocardial level
		Regulate the atrial arrhythmogenesis in heart failure
ARIMOCLOMOL	↑HSP70	Neuroprotective activity
CELASTROL	↑HSP70	Antioxidant and anti-inflammatory activities
		Neuroprotective activity
STATIN	↑HSP70	Contributes to plaque stability
	↑HSPB1	Reduces both mortality and the incidence of acute coronary syndrome
	↑HSP90	

statins in reducing both mortality and incidence of acute coronary syndrome (Noguchi et al. 2015; Smith et al. 2011; Weintraub et al. 2011). In addition, to reducing levels of serum low-density lipoprotein-cholesterol (LDL-C), statins also may contribute to plaque stability by reducing inflammation (Noguchi et al. 2015; Puato et al. 2010), improving endothelial function (Noguchi et al. 2015; Schönbeck and Libby 2004) and reinforcing the fibrous cap (Hattori et al. 2012; Komukai et al. 2014; Noguchi et al. 2015); these effects play crucial roles in the protection from CVDs (Fuster et al. 1992a, b; Noguchi et al. 2015). Remarkably, Frostegård et al. (2016) showed that statin, in particular atorvastatin, restores the proatherogenic HSP profile, characterized by an increase of pro-atherogenic HSP60 and HSP70 and a decrease of HSPB1, suggesting that statins could modulate immune reactions pivotal to atherosclerosis and CVDs pathogenesis. Simvastatin induces also vascular endothelial cells HSF1 translocate in nucleus and so cause the transcription of HSP70 and HSP90 (Uchiyama et al. 2007; Willis and Patterson 2010). All the above reported studies suggest that several drugs, have the potential to be cardioprotective because of their ability to modulate HSP in cardiovascular system. The effects of pharmacological compounds are summarized in Table 2.

In addition to the pharmacological induction of HSP, physical exercise was also discovered to induce HSPB levels and consequently has important cardioprotective effects. In fact, various studies demonstrated that gene and protein levels of HSPB1 and HSPB6 in heart are increased after physical exercise in animal models. In these studies, HSPB1 and HSPB6 were phosphorylated inducing myofilaments' stabilization, contractile damaged proteins restoration that resulted in improved contractile function of the heart (Boluylt et al. 2006; Burniston 2009; Campos et al. 2012; de Moraes et al. 2015; Rinaldi et al. 2006; Sakamoto et al. 2006; Hu et al. 2017). Moreover, it has been showed that physical exercise induced elevation of myocardial HSP expression (such as HSP70, HSP90 and HSPB5) and that regular endurance exercise protected the heart against ischemia/reperfusion and infarction (Harris and Starnes 2001; Powers et al. 2002). Therefore, physical exercise may represent a promising preventive therapeutic therapy against CVDs by also its activity in HSP

modulation. Furthermore, gene therapy can also be considered as a possible option to modulate HSP levels and function. In particular, adeno-associated viral vectors, which have been successfully utilized in clinical trials for the treatment of cardiomyopathy with SERCA2 overexpression, are currently the most effective in vivo delivery system (Jessup et al. 2011).

Conclusions

During the development of CVDs there is a modulation of chaperones and co-chaperones. Molecular chaperones are members of a large family of proteins that provide physiological control of proteostasis. Lack of regulation of the physiological balance between protein synthesis, folding, and degradation results in accumulation of misfolded proteins, so inducing several CVDs. In particular, we reported in this chapter that different cellular stress proteins have opposed role in promotion or inhibition of CVDs or related symptoms. The data discussed in this manuscript suggest that modulation of molecular chaperone expression could have a protective function in CVDs. Indeed, molecules capable of regulating chaperone expression are currently used in clinical trials, principally for the treatment of neurodegenerative diseases. Therefore, the discovery and the development of novel strategies to modulate chaperone expression and functions could be considered a crucial step for expanding drugs against CVDs.

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References

- Amour, J., Brzezinska, A. K., Weihsrauch, D., et al. (2009). Role of heat shock protein 90 and endothelial nitric oxide synthase during early anaesthetic and ischemic preconditioning. *Anesthesiology*, *110*, 317–325.
- Anckar, J., & Sistonen, L. (2011). Regulation of HSF1 function in the heat stress response: Implications in aging and disease. *Annual Review of Biochemistry*, *80*, 1089–1115.
- Arrigo, A. P., Virost, S., Chaurfour, S., et al. (2005). Hsp27 consolidates intracellular redox homeostasis by upholding glutathione in its reduced form and by decreasing iron intracellular levels. *Antioxidants & Redox Signaling*, *7*, 414–422.
- Baruah, K., Norouzitallab, P., Linayati, L., et al. (2014). Reactive oxygen species generated by a heat shock protein (Hsp) inducing product contributes to Hsp70 production and Hsp70-mediated protective immunity in *Artemia franciscana* against pathogenic vibrios. *Developmental and Comparative Immunology*, *46*, 470–474.
- Basha, E., Friedrich, K. L., & Vierling, E. (2006). The N-terminal arm of small heat shock proteins is important for both chaperone activity and substrate specificity. *The Journal of Biological Chemistry*, *281*, 39943–39952.

- Boluyt, M. O., Brevick, J. L., Rogers, D. S., et al. (2006). Changes in the rat heart proteome induced by exercise training: Increased abundance of heat shock protein hsp20. *Proteomics*, 6, 3154–3169.
- Boncoraglio, A., Minoia, M., & Carra, S. (2012). The family of mammalian small heat shock proteins (HSPBs): Implications in protein deposit diseases and motor neuropathies. *The International Journal of Biochemistry & Cell Biology*, 44, 1657–1669.
- Bond, U., & Schlesinger, M. J. (1985). Ubiquitin is a heat shock protein in chicken embryo fibroblasts. *Molecular and Cellular Biology*, 5, 949–956.
- Bova, M. P., Yaron, O., Huang, Q., et al. (1999). Mutation R120G in alphaB-crystallin, which is linked to a desmin-related myopathy, results in an irregular structure and defective chaperone-like function. *Proceedings of the National Academy of Sciences of the United States of America*, 96, 6137–6142.
- Brundel, B. J., Henning, R. H., Ke, L., et al. (2006a). Heat shock protein upregulation protects against pacing-induced myolysis in HL-1 atrial myocytes and in human atrial fibrillation. *Journal of Molecular and Cellular Cardiology*, 41, 555–562.
- Brundel, B. J., Shiroshita-Takeshita, A., Qi, X., et al. (2006b). Induction of heat shock response protects the heart against atrial fibrillation. *Circulation Research*, 99, 1394–1402.
- Budas, G. R., Churchill, E. N., Disatnik, M. H., et al. (2010). Mitochondrial import of PKCepsilon is mediated by HSP90: a role in cardioprotection from ischaemia and reperfusion injury. *Cardiovascular Research*, 88, 83–92.
- Bullard, B., Ferguson, C., Minajeva, A., et al. (2004). Association of the chaperone alphaB-crystallin with titin in heart muscle. *The Journal of Biological Chemistry*, 279, 7917–7924.
- Burian, K., Kis, Z., Virok, D., et al. (2001). Independent and joint effects of antibodies to human heat-shock protein 60 and Chlamydia pneumoniae infection in the development of coronary atherosclerosis. *Circulation*, 103, 1503–1508.
- Burniston, J. G. (2009). Adaptation of the rat cardiac proteome in response to intensity-controlled endurance exercise. *Proteomics*, 9, 106–115.
- Businaro, R., Profumo, E., Tagliani, A., et al. (2009). Heat-shock protein 90: A novel autoantigen in human carotid atherosclerosis. *Atherosclerosis*, 207, 74–83.
- Campos, J. C., Queliconi, B. B., Dourado, P. M., et al. (2012). Exercise training restores cardiac protein quality control in heart failure. *PLoS One*, 7, e52764.
- Carra, S., Seguin, S. J., Lambert, H., et al. (2008a). HspB8 chaperone activity toward poly (Q)-containing proteins depends on its association with Bag3, a stimulator of macroautophagy. *The Journal of Biological Chemistry*, 283, 1437–1444.
- Carra, S., Seguin, S. J., & Landry, J. (2008b). HspB8 and Bag3: A new chaperone complex targeting misfolded proteins to macroautophagy. *Autophagy*, 4, 237–239.
- Carra, S., Brunsting, J. F., Lambert, H., et al. (2009). HspB8 participates in protein quality control by a non-chaperone-like mechanism that requires eIF2 {alpha} phosphorylation. *The Journal of Biological Chemistry*, 284, 5523–5532.
- Carra, S., Alberti, S., Arrigo, P. A., et al. (2017). The growing world of small heat shock proteins: From structure to functions. *Cell Stress & Chaperones*, 22, 601–611.
- Chang, S. L., Chen, Y. C., Hsu, C. P., et al. (2013). Heat shock protein inducer modifies arrhythmogenic substrate and inhibits atrial fibrillation in the failing heart. *International Journal of Cardiology*, 168, 4019–4026.
- Champilas, N., Kyriakakis, E., & Tavernarakis, N. (2017). Small heat shock proteins in ageing and age-related diseases. *Cell Stress & Chaperones*, 22, 481–492.
- Chebotaeva, N., Bobkova, I., & Shilov, E. (2017). Heat shock proteins and kidney disease: Perspectives of HSP therapy. *Cell Stress & Chaperones*, 22, 319–343.
- Chen, Y. S., Chien, C. T., Ma, M. C., et al. (2005). Protection “outside the box” (skeletal remote preconditioning) in rat model is triggered by free radical pathway. *The Journal of Surgical Research*, 126, 92–101.

- Chen, L., Lizano, P., Zhao, X., et al. (2011). Preemptive conditioning of the swine heart by H11 kinase/Hsp22 provides cardiac protection through inducible nitric oxide synthase. *American Journal of Physiology. Heart and Circulatory Physiology*, 300, H1303–H1310.
- Chen, Y., Jiang, B., Zhuang, Y., et al. (2017). Differential effects of heat shock protein 90 and serine 1179 phosphorylation on endothelial nitric oxide synthase activity and on its cofactors. *PLoS One*, 12, e0179978.
- Choudhury, S., Bae, S., Ke, Q., et al. (2011). Mitochondria to nucleus translocation of AIF in mice lacking Hsp70 during ischemia/reperfusion. *Basic Research in Cardiology*, 106, 397–407.
- Clark, A. R., Lubsen, N. H., & Slingsby, C. (2012). sHSP in the eye lens: Crystallin mutations, cataract and proteostasis. *The International Journal of Biochemistry & Cell Biology*, 44, 1687–1697.
- Cohen, I. R., & Young, D. B. (1991). Autoimmunity, microbial immunity and the immunological homunculus. *Immunology Today*, 12, 105–110.
- Connarn, J. N., Assimon, V. A., Reed, R. A., et al. (2014). The molecular chaperone Hsp70 activates protein phosphatase 5 (PP5) by binding the tetratricopeptide repeat (TPR) domain. *The Journal of Biological Chemistry*, 289, 2908–2917.
- Craig, E. A., Weissman, J. S., & Horwich, A. L. (1994). Heat shock proteins and molecular chaperones: Mediators of protein conformation and turnover in the cell. *Cell*, 78, 365–372.
- Crul, T., Toth, N., Piotto, S., et al. (2013). Hydroximic acid derivatives: Pleiotropic HSP co-inducers restoring homeostasis and robustness. *Current Pharmaceutical Design*, 19, 309–346.
- Csermely, P., Schnaider, T., Soti, C., et al. (1998). The 90-kDa molecular chaperone family: Structure, function, and clinical applications. A comprehensive review. *Pharmacology & Therapeutics*, 79, 129–168.
- Cudkowicz, M. E., Shefner, J. M., Simpson, E., et al. (2008). Arimoclolomol at dosages up to 300 mg/day is well tolerated and safe in amyotrophic lateral sclerosis. *Muscle & Nerve*, 38, 837–844.
- de Graaf, R., Kloppenburg, G., Kitslaar, P. J., et al. (2006). Human heat shock protein 60 stimulates vascular smooth muscle cell proliferation through Toll-like receptors 2 and 4. *Microbes and Infection*, 8, 1859–1865.
- de Moraes, W. M., Melara, T. P., de Souza, P. R., et al. (2015). Impact of leucine supplementation on exercise training induced anti-cardiac remodeling effect in heart failure mice. *Nutrients*, 7, 3751–3766.
- Deane, C. A., & Brown, I. R. (2016). Induction of heat shock proteins in differentiated human neuronal cells following co-application of celastrol and arimoclolomol. *Cell Stress & Chaperones*, 21, 837–848.
- Depre, C., Hase, M., Gaussin, V., et al. (2002). H11 kinase is a novel mediator of myocardial hypertrophy in vivo. *Circulation Research*, 91, 1007–1014.
- Depre, C., Wang, L., Sui, X., et al. (2006). H11 kinase prevents myocardial infarction by preemptive preconditioning of the heart. *Circulation Research*, 98, 280–288.
- Doran, P., Gannon, J., O'Connell, K., et al. (2007). Aging skeletal muscle shows a drastic increase in the small heat shock proteins alphaB-crystallin/HspB5 and cvHsp/HspB7. *European Journal of Cell Biology*, 86, 629–640.
- Dybdahl, B., Slørdahl, S. A., Waage, A., et al. (2005). Myocardial ischaemia and the inflammatory response: Release of heat shock protein 70 after myocardial infarction. *Heart*, 91, 299–304.
- Edwards, H. V., Cameron, R. T., & Baillie, G. S. (2011). The emerging role of HSP20 as a multi-functional protective agent. *Cellular Signalling*, 23, 1447–1454.
- Fan, G. C., Chu, G., Mitton, B., et al. (2004). Small heat-shock protein Hsp20 phosphorylation inhibits beta-agonist-induced cardiac apoptosis. *Circulation Research*, 94, 1474–1482.
- Fan, G. C., Ren, X., Qian, J., et al. (2005). Novel cardioprotective role of a small heat-shock protein, Hsp20, against ischemia/reperfusion injury. *Circulation*, 111, 1792–1799.
- Feng, Y., Huang, W., Meng, W., et al. (2014). Heat shock improves Sca-1 + stem cell survival and directs ischemic cardiomyocytes toward a pro survival phenotype via exosomal transfer: A critical role for HSF1/miR-34a/HSP70 pathway. *Stem Cells*, 32, 462–472.

- Fontaine, J. M., Rest, J. S., Welsh, M. J., et al. (2003). The sperm outer dense fiber protein is the 10th member of the superfamily of mammalian small stress proteins. *Cell Stress & Chaperones*, 8, 62–69.
- Franklin, T. B., Krueger-Naug, A. M., Clarke, D. B., et al. (2005). The role of heat shock proteins Hsp70 and Hsp27 in cellular protection of the central nervous system. *International Journal of Hyperthermia*, 21, 379–392.
- Frostegård, J., Zhang, Y., Sun, J., et al. (2016). Oxidized low-density lipoprotein (OxLDL)-treated dendritic cells promote activation of T cells in human atherosclerotic plaque and blood, which is repressed by statins: MicroRNA let-7c is integral to the effect. *J Am Heart Assoc*, 5, e003976.
- Fukuoka, K., Sawabe, A., & Sugimoto, T. (2004). Inhibitory actions of several natural products on proliferation of rat vascular smooth muscle cells induced by Hsp60 from *Chlamydia pneumoniae* J138. *Journal of Agricultural and Food Chemistry*, 52, 6326–6329.
- Fuster, V., Badimon, L., Badimon, J. J., et al. (1992a). The pathogenesis of coronary artery disease and the acute coronary syndromes (1). *The New England Journal of Medicine*, 326, 242–250.
- Fuster, V., Badimon, L., Badimon, J. J., et al. (1992b). The pathogenesis of coronary artery disease and the acute coronary syndromes (2). *The New England Journal of Medicine*, 326, 310–318.
- Gabai, V. L., Meriin, A. B., Yaglom, J. A., et al. (2000). Suppression of stress kinase JNK is involved in HSP72-mediated protection of myogenic cells from transient energy deprivation. HSP72 alleviates the stress-induced inhibition of JNK dephosphorylation. *The Journal of Biological Chemistry*, 275, 38088–38094.
- Garrido, C., Brunet, M., Didelot, C., et al. (2006). Heat shock proteins 27 and 70: Anti-apoptotic proteins with tumorigenic properties. *Cell Cycle*, 5, 2592–2601.
- Gething, M. J., & Sambrook, J. (1992). Protein folding in the cell. *Nature*, 355, 33–45.
- Ghayour-Mobarhan, M., Lamb, D. J., Tavallaie, S., et al. (2007). Relationship between plasma cholesterol, von Willebrand factor concentrations, extent of atherosclerosis and antibody titers to heat shock proteins-60, -65 and -70 in cholesterol-fed rabbits. *International Journal of Experimental Pathology*, 88, 249–255.
- Ghayour-Mobarhan, M., Saber, H., & Ferns, G. A. (2012). The potential role of heat shock protein 27 in cardiovascular disease. *Clinica Chimica Acta*, 413, 15–24.
- Ghosh, J. G., Houck, S. A., & Clark, J. I. (2007). Interactive domains in the molecular chaperone human alphaB crystallin modulate microtubule assembly and disassembly. *PLoS One*, 2, e498.
- Golenhofen, N., Perng, M. D., Quinlan, R. A., et al. (2004). Comparison of the small heat shock proteins alphaB-crystallin, MKBP, HSP25, HSP20, and cvHSP in heart and skeletal muscle. *Histochemistry and Cell Biology*, 122, 415–425.
- Golenhofen, N., Redel, A., Wawrousek, E. F., et al. (2006). Ischemia-induced increase of stiffness of alphaB-crystallin/HSPB2-deficient myocardium. *Pflügers Archiv*, 451, 518–525.
- Grose, J. H., Langston, K., Wang, X., et al. (2015). Characterization of the cardiac overexpression of HSPB2 reveals mitochondrial and myogenic roles supported by a cardiac HspB2 interaction. *PLoS One*, 10, e0133994.
- Gupta, S., & Knowlton, A. A. (2007). HSP60 trafficking in adult cardiac myocytes: role of the exosomal pathway. *American Journal of Physiology. Heart and Circulatory Physiology*, 292, H3052–H3056.
- Gurbuxani, S., Schmitt, E., & Cande, C. (2003). Heat shock protein 70 binding inhibits the nuclear import of apoptosis-inducing factor. *Oncogene*, 22, 6669–6678.
- Gwag, T., Park, K., Kim, E., et al. (2013). Inhibition of C2C12 myotube atrophy by a novel HSP70 inducer, celastrol, via activation of Akt1 and ERK1/2 pathways. *Archives of Biochemistry and Biophysics*, 537, 21–30.
- Harris, M. B., & Starnes, J. W. (2001). Effects of body temperature during exercise training on myocardial adaptations. *American Journal of Physiology. Heart and Circulatory Physiology*, 280, H2271–H2280.
- Hattori, K., Ozaki, Y., Ismail, T. F., et al. (2012). Impact of statin therapy on plaque characteristics as assessed by serial OCT, grayscale and integrated backscatter-IVUS. *JACC Cardiovascular Imaging*, 5, 169–177.

- Hayashi, M., Imanaka-Yoshida, K., et al. (2006). A crucial role of mitochondrial Hsp40 in preventing dilated cardiomyopathy. *Nature Medicine*, *12*, 128–132.
- Hedhli, N., Lizano, P., Hong, C., et al. (2008). Proteasome inhibition decreases cardiac remodeling after initiation of pressure overload. *American Journal of Physiology. Heart and Circulatory Physiology*, *295*, H1385–H1393.
- Henderson, B., & Pockley, A. G. (2012). Proteotoxic stress and circulating cell stress proteins in the cardiovascular diseases. *Cell Stress & Chaperones*, *17*, 303–311.
- Hirakawa, T., Rokutan, K., Nikawa, T., et al. (1996). Geranylgeranylacetone induces heat shock proteins in cultured guinea pig gastric mucosal cells and rat gastric mucosa. *Gastroenterology*, *111*, 345–357.
- Hu, X., Van Marion, D. M. S., Wiersma, M., et al. (2017). The protective role of small heat shock proteins in cardiac diseases: Key role in atrial fibrillation. *Cell Stress & Chaperones*, *22*, 665–674.
- Ishiwata, T., Orosz, A., Wang, X., et al. (2012). HSPB2 is dispensable for the cardiac hypertrophic response but reduces mitochondrial energetics following pressure overload in mice. *PLoS One*, *7*, e42118.
- Jee, H. (2016). Size dependent classification of heat shock proteins: A mini-review. *Journal Exercise Rehabilitation*, *12*, 255–259.
- Jessup, M., Greenberg, B., Mancini, D., et al. (2011). Calcium upregulation by percutaneous administration of gene therapy in cardiac disease (CUPID) investigators. Calcium upregulation by percutaneous administration of gene therapy in cardiac disease (CUPID): A phase 2 trial of intracoronary gene therapy of sarcoplasmic reticulum Ca²⁺-ATPase in patients with advanced heart failure. *Circulation*, *124*, 304–313.
- Jimenez, S. K., Small, B. A., Hsu, A. K., et al. (2014). Heat shock proteins HSP90 and HSP70 mediate opioid- and GSK3 β -induced cardioprotection. *Circulation Research*, *115*, 340.
- Johnson, A. D., Berberian, P. A., Tytell, M., et al. (1993). Atherosclerosis alters the localization of HSP70 in human and macaque aortas. *Experimental and Molecular Pathology*, *58*, 155–168.
- Kampinga, H. H., & Craig, E. A. (2010). The HSP70 chaperone machinery: J proteins as drivers of functional specificity. *Nature Reviews Molecular Cell Biology*, *11*, 579–592.
- Kappe, G., Franck, E., Verschuere, P., et al. (2003). The human genome encodes 10 alpha-crystallin-related small heat shock proteins: HspB1-10. *Cell Stress & Chaperones*, *8*, 53–61.
- Kessing, D., Denollet, J., Widdershoven, J., et al. (2016). Self-care and all-cause mortality in patients with chronic heart failure. *JACC Heart Failure*, *4*, 176–183.
- Khalil, A. A., Kababy, N. F., Deraz, S. F., et al. (2011). Heat shock proteins in oncology: Diagnostic biomarkers or therapeutic targets? *Biochimica et Biophysica Acta*, *1816*, 89–104.
- Khan, I. U., Wallin, R., Gupta, R. S., et al. (1998). Protein kinase A-catalyzed phosphorylation of heat shock protein 60 chaperone regulates its attachment to histone 2B in the T lymphocyte plasma membrane. *Proceedings of the National Academy of Sciences of the United States of America*, *95*, 10425–10430.
- Kieran, D., Kalmar, B., Dick, J. R., et al. (2004). Treatment with arimoclomol, a coinducer of heat shock proteins, delays disease progression in ALS mice. *Nature Medicine*, *10*, 402–405.
- Kim, K. K., Kim, R., & Kim, S. H. (1998). Crystal structure of a small heat-shock protein. *Nature*, *394*, 595–599.
- Kirkegaard, T., Gray, J., Priestman, D. A., et al. (2016). Heat shock protein-based therapy as a potential candidate for treating the sphingolipidoses. *Science Translational Medicine*, *8*, 355ra118.
- Kleindienst, R., Xu, Q., Willeit, J., et al. (1993). Immunology of atherosclerosis. Demonstration of heat shock protein 60 expression and T lymphocytes bearing alpha/beta or gamma/delta receptor in human atherosclerotic lesions. *The American Journal of Pathology*, *142*, 1927–1937.
- Klíková, K., Pilchová, I., Stefaníková, A., et al. (2016). The role of heat shock proteins in leukemia. *Klinická Onkologie*, *29*, 29–38.

- Knowlton, A. A., Kapadia, S., & Torre-Amione, G. (1998). Differential expression of heat shock proteins in normal and failing human hearts. *Journal of Molecular and Cellular Cardiology*, 30, 811–818.
- Komukai, K., Kubo, T., Kitabata, H., et al. (2014). Effect of atorvastatin therapy on fibrous cap thickness in coronary atherosclerotic plaque as assessed by optical coherence tomography: The EASY-FIT study. *Journal of the American College of Cardiology*, 64, 2207–2217.
- Kupatt, C., Dessy, C., Hinkel, R., et al. (2004). Heat shock protein 90 transfection reduces ischemia-reperfusion-induced myocardial dysfunction via reciprocal endothelial NO synthase serine 1177 phosphorylation and threonine 495 dephosphorylation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 24, 1435–1441.
- Lamb, D. J., El-Sankary, W., & Ferns, G. A. (2002). Molecular mimicry in atherosclerosis: A role for heat shock proteins in immunization. *Atherosclerosis*, 167, 177–185.
- Lambert, H., Charette, S. J., Bernier, A. F., et al. (1999). HSP27 multimerization mediated by phosphorylation-sensitive intermolecular interactions at the amino terminus. *The Journal of Biological Chemistry*, 274, 9378–9385.
- Lanka, V., Wieland, S., Barber, J., et al. (2009). Arimoclomol: A potential therapy under development for ALS. *Expert Opinion on Investigational Drugs*, 18, 1907–1918.
- Lee, J. H., Koo, T. H., Yoon, H., et al. (2006). Inhibition of NF-kappa B activation through targeting I kappa B kinase by celastrol, a quinone methide triterpenoid. *Biochemical Pharmacology*, 72, 1311–1321.
- Li, Y., Si, R., Feng, Y., et al. (2011). Myocardial ischemia activates an injurious innate immune signaling via cardiac heat shock protein 60 and Toll-like receptor 4. *The Journal of Biological Chemistry*, 286, 31308–31319.
- Lin, L., Kim, S. C., Wang, Y., et al. (2007). HSP60 in heart failure: Abnormal distribution and role in cardiac myocyte apoptosis. *American Journal of Physiology. Heart and Circulatory Physiology*, 293, H2238–H2247.
- Madamanchi, N. R., Patterson, C., Li, S., & Runge, S. M. (2001). Reactive oxygen species regulate heat-shock protein 70 via the JAK/STAT pathway. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 21, 321–326.
- Malik, Z. A., Kott, K. S., Poe, A. J., et al. (2013). Cardiac myocyte exosomes: Stability, HSP60, and proteomics. *American Journal of Physiology Heart and Circulatory Physiology*, 304, H954–H965.
- Mamipour, M., Yousefi, M., & Hasanzadeh, M. (2017). An overview on molecular chaperones enhancing solubility of expressed recombinant proteins with correct folding. *International Journal of Biological Macromolecules*, 102, 367–375.
- Marber, M. S., Latchman, D. S., Walker, J. M., et al. (1993). Cardiac stress protein elevation 24 hours after brief ischemia or heat stress is associated with resistance to myocardial infarction. *Circulation*, 88, 1264–1272.
- Martin, T. P., Currie, S., & Baillie, G. S. (2014). The cardioprotective role of small heat-shock protein 20. *Biochemical Society Transactions*, 42, 270–273.
- Martin-Ventura, J. L., Duran, M. C., Blanco-Colio, L. M., et al. (2004). Identification by a differential proteomic approach of heat shock protein 27 as a potential marker of atherosclerosis. *Circulation*, 110, 2216–2219.
- Marunouchi, T., Inomata, S., Sanbe, A., et al. (2014). Protective effect of geranylgeranylacetone via enhanced induction of HSPB1 and HSPB8 in mitochondria of the failing heart following myocardial infarction in rats. *European Journal of Pharmacology*, 730, 140–147.
- Mayr, M., Metzler, B., Kiechl, S., et al. (1999). Endothelial cytotoxicity mediated by serum antibodies to heat shock proteins of *Escherichia coli* and *Chlamydia pneumoniae*: Immune reactions to heat shock proteins as a possible link between infection and atherosclerosis. *Circulation*, 99, 1560–1566.
- Mazzaferro, V., Coppa, J., Carrabba, M. G., et al. (2003). Vaccination with autologous tumor-derived heat-shock protein gp96 after liver resection for metastatic colorectal cancer. *Clinical Cancer Research*, 9, 3235–3245.

- Mu, H., Wang, L., & Zhao, L. (2017). HSP90 inhibition suppresses inflammatory response and reduces carotid atherosclerotic plaque formation in ApoE mice. *Cardiovascular Therapy*, 35(2), 1–9.
- Multhoff, G., Pockley, A. G., Schmida, T. E., et al. (2015). The role of heat shock protein 70 (Hsp70) in radiation-induced immunomodulation. *Cancer Letters*, 368, 179–184.
- Nakagawa, M., Tsujimoto, N., Nakagawa, H., et al. (2001). Association of HSPB2, a member of the small heat shock protein family, with mitochondria. *Experimental Cell Research*, 271, 161–168.
- Nakai, A., & Ishikawa, T. (2001). Cell cycle transition under stress conditions controlled by vertebrate heat shock factors. *The EMBO Journal*, 20, 2885–2895.
- Neef, D. W., Jaeger, A. M., & Thiele, D. J. (2011). Heat shock transcription factor 1 as a therapeutic target in neurodegenerative diseases. *Nature Reviews Drug Discovery*, 10, 930–944.
- Noguchi, T., Tanaka, A., Kawasaki, T., et al. (2015). Effect of intensive statin therapy on coronary high-intensity plaques detected by noncontrast T1-weighted imaging: The AQUAMARINE pilot study. *Journal of the American College of Cardiology*, 66, 245–256.
- Ooie, T., Takahashi, N., Saikawa, T., et al. (2001). Single oral dose of geranylgeranylacetone induces heat-shock protein 72 and renders protection against ischemia/reperfusion injury in rat heart. *Circulation*, 104, 1837–1843.
- Panneerselvam, L., Raghunath, A., & Perumal, E. (2017). Differential expression of myocardial heat shock proteins in rats acutely exposed to fluoride. *Cell Stress Chaperones*, 22, 743–750. [Epub ahead of print].
- Parfitt, D. A., Aguila, M., McCulley, C. H., et al. (2014). The heat-shock response co-inducer arimoclomol protects against retinal degeneration in rhodopsin retinitis pigmentosa. *Cell Death & Disease*, 5, e1236.
- Pearl, L. H., & Prodromou, C. (2001). Structure, function, and mechanism of the Hsp90 molecular chaperone. *Advances in Protein Chemistry*, 59, 157–186.
- Perng, M. D., Cairns, L., van den Ijssel, P., et al. (1999). Intermediate filament interactions can be altered by HSP27 and alphaB-crystallin. *Journal of Cell Science*, 112, 2099–2112.
- Pfister, G., Stroth, C. M., & Perschinka, H. (2005). Detection of HSP60 on the membrane surface of stressed human endothelial cells by atomic force and confocal microscopy. *Journal of Cell Science*, 118, 1587–1594.
- Picard, D. (2002). Heat-shock protein 90, a chaperone for folding and regulation. *Cellular and Molecular Life Sciences*, 59, 1640–1648.
- Pipkin, W., Johnson, J. A., Creazzo, T. L., et al. (2003). Localization, macromolecular associations, and function of the small heat shock-related protein HSP20 in rat heart. *Circulation*, 107, 469–476.
- Pockley, A. G. (2002). Heat shock proteins, inflammation, and cardiovascular disease. *Circulation*, 105, 1012–1017.
- Pockley, A. G., Georgiades, A., Thulin, T., et al. (2003). Serum heat shock protein 70 levels predict the development of atherosclerosis in subjects with established hypertension. *Hypertension*, 42, 235–238.
- Powers, S. K., Lennon, S. L., Quindry, J., et al. (2002). Exercise and cardioprotection. *Current Opinion in Cardiology*, 17, 495–502.
- Pratt, W. B., & Toft, D. O. (2003). Regulation of signaling protein function and trafficking by the hsp90/hsp70-based chaperone machinery. *Experimental Biology and Medicine (Maywood, N.J.)*, 228, 111–133.
- Prohászka, Z., Doba, J., Horváth, L., et al. (2001). Comparative study on antibodies to human and bacterial 60 kDa heat shock proteins in a large cohort of patients with coronary heart disease and healthy subjects. *European Journal of Clinical Investigation*, 31, 285–292.
- Puato, M., Faggini, E., Rattazzi, M., et al. (2010). Atorvastatin reduces macrophage accumulation in atherosclerotic plaques: A comparison of a nonstatin-based regimen in patients undergoing carotid endarterectomy. *Stroke*, 41, 1163–1168.

- Qian, J., Ren, X., Wang, X., et al. (2009). Blockade of Hsp20 phosphorylation exacerbates cardiac ischemia/reperfusion injury by suppressed autophagy and increased cell death. *Circulation Research*, *105*, 1223–1231.
- Qian, J., Vafiadaki, E., Florea, S. M., et al. (2011). Small heat shock protein 20 interacts with protein phosphatase-1 and enhances sarcoplasmic reticulum calcium cycling. *Circulation Research*, *108*, 1429–1438.
- Qiu, H., Lizano, P., Laure, L., et al. (2011). H11 kinase/heat shock protein 22 deletion impairs both nuclear and mitochondrial functions of STAT3 and accelerates the transition into heart failure on cardiac overload. *Circulation*, *124*, 406–415.
- Rayner, K., Chen, Y. X., McNulty, M., et al. (2008). Extracellular release of the atheroprotective heat shock protein 27 is mediated by estrogen and competitively inhibits acLDL binding to scavenger receptor-A. *Circulation Research*, *103*, 133–141.
- Richter, K., & Buchner, J. (2001). Hsp90: Chaperoning signal transduction. *Journal of Cellular Physiology*, *188*, 281–290.
- Rigano, R., Profumo, E., Buttari, B., et al. (2007). Heat shock proteins and autoimmunity in patients with carotid atherosclerosis. *Annals of the New York Academy of Sciences*, *1107*, 1–10.
- Rinaldi, B., Corbi, G., Boccuti, S., et al. (2006). Exercise training affects age-induced changes in SOD and heat shock protein expression in rat heart. *Experimental Gerontology*, *41*, 764–770.
- Ritossa, F. (1962). A new puffing pattern induced by heat shock and DNP in drosophila. *Experientia*, *18*, 571–573.
- Sakamoto, M., Minamino, T., Toko, H., et al. (2006). Upregulation of heat shock transcription factor 1 plays a critical role in adaptive cardiac hypertrophy. *Circulation Research*, *99*, 1411–1418.
- Sassa, H., Takaishi, Y., & Terada, H. (1990). The triterpene celastrol as a very potent inhibitor of lipid peroxidation in mitochondria. *Biochemical Biophysical Research Communications*, *172*, 890–897.
- Sasu, S., LaVerda, D., Qureshi, N., et al. (2001). Chlamydia pneumoniae and chlamydial heat shock protein 60 stimulate proliferation of human vascular smooth muscle cells via toll-like receptor 4 and p44/p42 mitogen-activated protein kinase activation. *Circulation Research*, *89*, 244–250.
- Schönbeck, U., & Libby, P. (2004). Inflammation, immunity, and HMG-CoA reductase inhibitors: Statins as antiinflammatory agents? *Circulation*, *109*, II18–II26.
- Seibert, T. A., Hibbert, B., Chen, Y. X., et al. (2013). Serum heat shock protein 27 levels represent a potential therapeutic target for atherosclerosis: Observations from a human cohort and treatment of female mice. *Journal of the American College of Cardiology*, *62*, 1446–1454.
- Selcen, D., & Engel, A. G. (2003). Myofibrillar myopathy caused by novel dominant negative alpha B-crystallin mutations. *Annals of Neurology*, *54*, 804–810.
- Singh, L., Randhawa, P. K., Singh, N., et al. (2017). Redox signaling in remote ischemic preconditioning-induced cardioprotection: Evidences and mechanisms. *European Journal of Pharmacology*, *809*, 151–155.
- Smith, S. C., Jr., Benjamin, E. J., Bonow, R. O., et al. (2011). AHA/ACCF secondary prevention and risk reduction therapy for patients with coronary and other atherosclerotic vascular disease: 2011 update: a guideline from the American Heart Association and American College of Cardiology Foundation endorsed by the World Heart Federation and the Preventive Cardiovascular Nurses Association. *Journal of the American College of Cardiology*, *58*, 2432–2446.
- Sreedhar, A. S., Kalmár, E., Csermely, P., et al. (2004). Hsp90 isoforms: Functions, expression and clinical importance. *FEBS Letters*, *562*, 11–15.
- Sugiyama, Y., Suzuki, A., Kishikawa, M., et al. (2000). Muscle develops a specific form of small heat shock protein complex composed of MKBP/HSPB2 and HSPB3 during myogenic differentiation. *The Journal of Biological Chemistry*, *275*, 1095–1104.
- Suzuki, A., Sugiyama, Y., Hayashi, Y., et al. (1998). MKBP, a novel member of the small heat shock protein family, binds and activates the myotonic dystrophy protein kinase. *The Journal of Cell Biology*, *140*, 1113–1124.

- Trott, A., West, J. D., Klaić, L., et al. (2008). Activation of heat shock and antioxidant responses by the natural product celastrol: Transcriptional signatures of a thiol-targeted molecule. *Molecular Biology of the Cell*, *19*, 1104–1112.
- Uchiyama, T., Atsuta, H., Utsugi, T., et al. (2007). HSF1 and constitutively active HSF1 improve vascular endothelial function (heat shock proteins improve vascular endothelial function). *Atherosclerosis*, *190*, 321–329.
- van de Klundert, F. A., Gijzen, M. L., van den Ijssel, P. R., et al. (1998). alpha B-crystallin and hsp25 in neonatal cardiac cells-differences in cellular localization under stress conditions. *European Journal of Cell Biology*, *75*, 38–45.
- Van Montfort, R. L., Basha, E., Friedrich, K. L., et al. (2001a). Crystal structure and assembly of a eukaryotic small heat shock protein. *Nature Structural Biology*, *8*, 1025–1030.
- Van Montfort, R., Slingsby, C., & Vierling, E. (2001b). Structure and function of the small heat shock protein/alpha-crystallin family of molecular chaperones. *Advances in Protein Chemistry*, *59*, 105–156.
- Veres, A., Füst, G., Smieja, M., et al. (2002). Heart Outcomes Prevention Evaluation (HOPE) study investigators. Relationship of anti-60 kDa heat shock protein and anti-cholesterol antibodies to cardiovascular events. *Circulation*, *106*, 2775–2780.
- Verschuure, P., Tatard, C., Boelens, W. C., et al. (2003). Expression of small heat shock proteins HspB2, HspB8, Hsp20 and cvHsp in different tissues of the perinatal developing pig. *European Journal of Cell Biology*, *82*, 523–530.
- Vicart, P., Caron, A., Guicheney, P., et al. (1998). A missense mutation in the alphaB-crystallin chaperone gene causes a desmin-related myopathy. *Nature Genetics*, *20*, 92–95.
- Vicencio, J. M., Yellon, D. M., Sivaraman, V., et al. (2015). Plasma exosomes protect the myocardium from ischemia-reperfusion injury. *Journal of the American College of Cardiology*, *65*, 1525–1536.
- Vígh, L., Literáti, P. N., Horváth, I., et al. (1997). Bimoclomol: A nontoxic, hydroxylamine derivative with stress protein-inducing activity and cytoprotective effects. *Nature Medicine*, *3*, 1150–1154.
- Vos, M. J., Hageman, J., Carra, S., et al. (2008). Structural and functional diversities between members of the human HSPB, HSPH, HSPA, and DNAJ chaperone families. *Biochemistry*, *47*, 7001–7011.
- Vos, M. J., Kanon, B., & Kampinga, H. H. (2009). HSPB7 is a SC35 speckle resident small heat shock protein. *Biochimica et Biophysica Acta*, *1793*, 1343–1353.
- Vos, M. J., Zijlstra, M. P., & Kanon, B. (2010). HSPB7 is the most potent polyQ aggregation suppressor within the HSPB family of molecular chaperones. *Human Molecular Genetics*, *19*, 4677–4693.
- Vos, M. J., Zijlstra, M. P., Carra, S., et al. (2011). Small heat shock proteins, protein degradation and protein aggregation diseases. *Autophagy*, *7*, 101–103.
- Wang, W., Peng, Y., Wang, Y., et al. (2009). Anti-apoptotic effect of heat shock protein 90 on hypoxia-mediated cardiomyocyte damage is mediated via the phosphatidylinositol 3-kinase/AKT pathway. *Clinical and Experimental Pharmacology & Physiology*, *36*, 899–903.
- Wang, Y., Chen, L., Hagiwara, N., et al. (2010). Regulation of heat shock protein 60 and 72 expression in the failing heart. *Journal of Molecular and Cellular Cardiology*, *48*, 360–366.
- Wei, H., Campbell, W., & Vander Heide, R. S. (2006). Heat shock-induced cardioprotection activates cytoskeletal-based cell survival pathways. *American Journal of Physiology. Heart and Circulatory Physiology*, *291*, H638–H647.
- Weintraub, N. L., & Rubinstein, J. (2013). Cooling the fire of atherosclerosis with heat shock protein 27. *Journal of the American College of Cardiology*, *62*, 1455–1456.
- Weintraub, W. S., Daniels, S. R., Burke, L. E., et al. (2011). Value of primordial and primary prevention for cardiovascular disease: A policy statement from the American Heart Association. *Circulation*, *124*, 967–990.
- Westerheide, S. D., Bosman, J. D., Mbadugha, B. N., et al. (2004). Celastrols as inducers of the heat shock response and cytoprotection. *The Journal of Biological Chemistry*, *279*, 56053–56060.

- Westerheide, S. D., Raynes, R., Powell, C., et al. (2012). HSF transcription factor family, heat shock response, and protein intrinsic disorder. *Current Protein & Peptide Science*, *13*, 86–103.
- Willis, M. S., & Patterson, C. (2010). Hold me tight: Role of the heat shock protein family of chaperones in cardiac disease. *Circulation*, *122*, 1740–1751.
- Wu, K., Xu, W., You, Q., et al. (2012). Increased expression of heat shock protein 90 under chemical hypoxic conditions protects cardiomyocytes against injury induced by serum and glucose deprivation. *International Journal of Molecular Medicine*, *30*, 1138–1144.
- Xiao, Q., Mandal, K., Schett, G., et al. (2005). Association of serum-soluble heat shock protein 60 with carotid atherosclerosis: Clinical significance determined in a follow-up study. *Stroke*, *36*, 2571–2576.
- Xu, Q., Schett, G., Perschinka, H., et al. (2000). Serum soluble heat shock protein 60 is elevated in subjects with atherosclerosis in a general population. *Circulation*, *102*, 14–20.
- Zhang, X., Min, X., Li, C., et al. (2010). Involvement of reductive stress in the cardiomyopathy in transgenic mice with cardiac-specific overexpression of heat shock protein 27. *Hypertension*, *55*, 1412–1417.
- Zhang, C., Liu, X., Miao, J., et al. (2017). Heat shock protein 70 protects cardiomyocytes through suppressing SUMOylation and nucleus translocation of phosphorylated eukaryotic elongation factor 2 during myocardial ischemia and reperfusion. *Apoptosis*, *22*, 608–625.
- Zhao, Y., Zhang, C., Wei, X., et al. (2015). Heat shock protein 60 stimulates the migration of vascular smooth muscle cells via Toll-like receptor 4 and ERK MAPK activation. *Scientific Reports*, *5*, 15352.
- Zhong, G. Q., Tu, R. H., Zeng, Z. Y., et al. (2014). Novel functional role of heat shock protein 90 in protein kinase C-mediated ischemic post-conditioning. *The Journal of Surgical Research*, *189*, 198–206.
- Zhu, J., Quyyumi, A. A., Rott, D., et al. (2001). Antibodies to human heat-shock protein 60 are associated with the presence and severity of coronary artery disease: Evidence for an autoimmune component of atherogenesis. *Circulation*, *103*, 1071–1075.
- Zhu, J., Quyyumi, A. A., Wu, H., et al. (2003). Increased serum levels of heat shock protein 70 are associated with low risk of coronary artery disease. *Arteriosclerosis, Thrombosis, and Vascular Biology*, *23*, 1055–1059.