

C*amelina sativa*: An Emerging
Biofuel Crop

Shivani Chaturvedi, Amrik Bhattacharya, Sunil Kumar Khare, and Geetanjali Kaushik

Contents

S. Chaturvedi · A. Bhattacharya · S. K. Khare

G. Kaushik (\boxtimes)

Department of Chemistry, Indian Institute of Technology, Delhi, New Delhi, India e-mail: [shivani.d123@gmail.com;](mailto:shivani.d123@gmail.com) [amrik.du@gmail.com;](mailto:amrik.du@gmail.com) skhare@rocketmail.com

MGM's Jawaharlal Nehru Engineering College, Mahatma Gandhi Mission, Aurangabad, Maharashtra, India e-mail: geetanjaliac@gmail.com

[©] Springer Nature Switzerland AG 2019

C. M. Hussain (ed.), Handbook of Environmental Materials Management, https://doi.org/10.1007/978-3-319-73645-7_110

Abstract

In current scenario, the petroleum products should be replaced by renewable plant originated products, but the food crops must be avoided for this. In these lines, Camelina sativa L. Crantz is very promising crop; it needs no special fertilizers, herbicides, or pesticides and produces decent amount of biofuel. C. sativa contains 30–48% oil and 33–47% protein and adequate micronutrients. With such wholesome composition, it displays varied applications, not only as feed but also for multiple industrial uses. It has compounds of bio-significance like glucosinolates, phenolic compounds, tocopherols, polyunsaturated fatty acids, polysaccharides, and lignans.

In biofuel industries it is utilized as biodiesel, jet fuel, glycerol, and biolubricants. In animal and human system, Camelina oil and seeds have applicability in treatment of burns, inflammations, heart disease, neurological abnormalities, cholesterol reduction, ulcers, and cancer. It is also used as healthpromoting supplements. Its application as adhesive, alkyd resins, cosmetic products, soaps, lotions, gum, plastics, and wax manufacturing are many other alternatives to create new dimensions in the pathway of Camelina. In agronomical practices, the deoiled cake of Camelina can be useful as organic fertilizer, pesticide, insecticide, and antifungal agents.

Camelina has high ω-3 fatty acid, which has capacity to be used in therapeutic formulations. It is also rich in antioxidant like α-tocopherol that is responsible to enhance shelf life. With the development of some improved new varieties, Camelina may prove to be a potential crop for future renewable feedstock for biofuel industries.

Keywords

Camelina sativa · Biofuel crop · Antioxidant · Polyphenols · Omega-3-fatty acids

Introduction

Several seed bearing crops like soybean (Glycine max), groundnut (Arachis hypogaea), mustard (Brassica spp.), Camelina (Camelina sativa), etc., are rich source of oil. Some of these varieties are commonly used for edible oil; Camelina is an exception. The "fuel versus food" debate has drawn attention toward the nonfood biodiesel crops which demand less nutrient, water, and land as compare to traditional oil seed crops. Oil from plant origin (generally nonedible type) is being used as an alternative to petroleum products as new renewable industrial products (Carlsson [2009](#page-28-0); Schneider [2006;](#page-34-0) Wittkop et al. [2009](#page-36-0)). C. sativa from Brassicaceae family or mustard family is an example of plant-based renewable oil which not only helps in different industries to produce useful products but is also an environmentally sustainable source of energy. It is known to be economically sustainable, less weather dependent, and requiring less tillage and weed control. It is a short duration biofuel crop having consistent yield (Urbaniak et al. [2008;](#page-35-0) Krohn and Fripp [2012;](#page-31-0) Razeq et al. [2014\)](#page-34-1). Furthermore, it has an ability to grow in very poor quality soil, without any expense on herbicides and pesticides (Iskandarov et al. [2014\)](#page-31-1).

Present review covers various aspects of Camelina research including its origin, morphology, cultivational practices, future potential areas, biologically significant compound, nutritional composition, and multiple applications for medicinal, industrial, and agricultural purposes.

Morphological Features, Cultivation, and Yield

Morphological Features of Camelina

Camelina is self-pollinated and autogamous plant (Mulligan [2002](#page-33-0)). It has shorter root system, a property that makes it suitable for dry land and marginal land cultivation (Putnam et al. [1993\)](#page-33-1). Camelina plant turns woody on maturity and reaches height from 30 to 80 cm (Pavlista et al. [2011](#page-33-2)). It has arrow-shaped, pointed, smooth edges leaves, with 5–9 cm in length (Hulbert et al. [2005](#page-31-2)). Camelina has teardrop-shaped pods having small seeds. Generally, 350,000 seeds per pound are produced, but this depends upon growing conditions and the variety which is selected by the grower (Hulbert et al. [2005](#page-31-2)).

Camelina Plant and Cultivational Practices

Since Camelina has very recently become a crop of significant commercial interest, its agricultural management is still to be developed (Vollmann et al. [2007](#page-36-1)). In normal conditions, the oil content in Camelina is 30–48% on dry weight basis (Pavlista et al. [2011;](#page-33-2) Vollmann et al. [2007](#page-36-1)), which is twice as that of soybean seed oil (18–22%) (Moser [2012](#page-33-3)).

It can be grown in spring as well as winter climate (Gesch and Cermak [2011](#page-30-0)). The seeds of Camelina can emerge at 0° C in controlled environmental conditions (Allen et al. [2014](#page-27-0)). In comparison to other oil seed crops (flax, rapeseed, and mustard) their seedlings can sustain even lower temperatures like -2 °C (Robinson [1987\)](#page-34-2). This quality makes it suitable to be grown in colder climatic conditions. The germination of Camelina seeds is inhibited or delayed of the early growth development because of high temperature (Zanetti et al. [2013](#page-36-2)). Camelina also has frost tolerance and winter hardiness as spatial and temporal crop (Russo and Reggiani [2012](#page-34-3)). During seed development the temperature increase of more than $25 \degree C$ may lead to a significant reduction of unsaturated fatty acid content (Zubr and Matthäus [2002;](#page-36-3)

Gilbertson et al. [2007\)](#page-30-1). If seed bed is prepared with standard agricultural specification, then it displays high germination rate (Schillinger et al. [2012](#page-34-4)). Adapted to different environmental conditions and with improved traits, lot of varieties of seeds are now available in the market (Berti et al. [2011\)](#page-28-1).

The seed yield and plant establishment depends upon soil preparation, seeding rate, method of planting, and seeding depth (Berti et al. [2011](#page-28-1)). Camelina crop needs less fertilization to give good yield and can be grown on marginal land (Putnam et al. [1993\)](#page-33-1). In general, 5–7 kg seeds ha⁻¹ is enough for good dense stand. However, use of excessive crop residue without appropriate tilling practices can reduce seedling emergence (Enjalbert and Johnson [2011\)](#page-29-0).

For Camelina production (growth, yield, and quality of seed) soil nutrient management is necessary. The key factors that affect cultivation are the type of soil, pre-plant soil fertility, and soil moisture. By prior researches it has been demonstrated that with the increase in input of nitrogen fertilizer, the yield of seed is increased but oil content in the seeds is reduced (Jiang et al. [2014\)](#page-31-3). For this, nitrogen 50–120 kg ha⁻¹, phosphorous 15–30 kg ha⁻¹, and 10–25 kg ha⁻¹ sulfur is considered to be adequate (Jiang et al. [2014](#page-31-3); Obour et al. [2015](#page-33-4)). Varying quantities of nitrogen give sufficient yield such as $45-50$ kg ha⁻¹ (Obour et al. [2015](#page-33-4)), 120 kg N ha^{-1} (Wysocki et al. [2013](#page-36-4)), and 185 kg N ha^{-1} (Solis et al. [2013\)](#page-35-1). It has been reported earlier that if nitrogen application increased beyond an optimal dose, it may lead to increased lodging (Berti et al. [2011](#page-28-1); Rathke et al. [2006\)](#page-34-5).

Proper crop spacing is an important factor for high yield and better oil content in seed crops. Row spacing should be maintained for better vegetative and productive phases. Prior researchers have standardized row spacing under central western Himalayas of India (Kumari et al. [2012](#page-32-0)). The harvesting should not be delayed since it can result in pod shattering and can lead to loss of yield (Obour et al. [2015\)](#page-33-4). It is recommended that harvesting be done early as soon as plants achieve physiological maturity or when 50–75% silicles are turned brown in color (Berti et al. [2011\)](#page-28-1).

According to field experiments, the fallow period between two wheat crops in the field may be utilized for Camelina cultivation in semiarid region (Obour et al. [2015\)](#page-33-4). This will help in increasing nutrient cycling with decomposition of soil organic matter (Chen et al. [2015\)](#page-29-1). Being short duration crop of 85–100 days, suitability of Camelina as potential alternative crop to fill abovementioned fallow period is further strengthened (McVay and Lamb [2008\)](#page-32-1).

Crop Pest, Insect, Disease, and Weed Management

Unlike Canola, Camelina is unaffected by flea beetle and bird damage (Pavlista et al. [2011\)](#page-33-2). It demonstrated tolerance against spring freezing and drought conditions in comparison to Canola crop (McVay and Lamb [2008](#page-32-1)). It was also resistant to insect pests (Iskandarov et al. [2014;](#page-31-1) Kirkhus et al. [2013](#page-31-4)). For Camelina crop, quizalofop herbicide is used for weed control (Assure II, Yuma GL) and glyphosate is useful for the preharvest weeds control (Jha and Stougaard [2013](#page-31-5)). Prior researchers had used treflan and bonanza as pre-plant herbicides to control broad leaf and grassy weed (Yang et al. [2016](#page-36-5)). Sethoxydim is the only labeled herbicide available for weed control for this crop and even get control on grass weeds but ineffective on broad leaves (Obour et al. [2015\)](#page-33-4).

A disease Downy Mildew (Peronospora parasitica) was detected in Camelina (Séguin-Swartz et al. [2009\)](#page-34-6). Sclerotinia (Sclerotinia sclerotiorum) is also found in fields of Camelina, which may reduce yield (Yang et al. [2016\)](#page-36-5).

There is limited information available on application of pre-emergence herbicides (PRE) on weed management of Camelina (Schillinger et al. [2012;](#page-34-4) Steiner et al. [2006\)](#page-35-2). Therefore, available alternatives of weed management have to apply a labeled broad-spectrum herbicide before sowing or remove the weeds done mechanically in field which is very time-consuming practice (Froment et al. [2006\)](#page-30-2).

Crop Yield

According to United States Department of Agriculture (USDA) and Department of Energy oil seeds are included in the bioenergy crops (USDA). Oil seed accounts for 36 billion gallons biofuel production of which around 0.5 billion gallons is expected to come from these oil seed crops.

Camelina seed yield is around 106–907 L ha⁻¹, which is significantly higher than sunflower (500–750 L ha⁻¹) and soybean (247–562 L ha⁻¹) with similar kind of conditions (Moser and Vaughn [2010\)](#page-33-5).

Cultivation Aspects in Indian Context

In Indian conditions, Camelina seeds produce around 320–480 g kg⁻¹ and yield is up to 600–1700 kg ha⁻¹ (Kumari et al. [2012\)](#page-32-0). The spacing between the rows significantly impact the yield and oil content of Camelina sativa cv. Calena (Kumari et al. [2012](#page-32-0)). Wider row spacing results in higher flowering (Kumari et al. [2012\)](#page-32-0). It was found that early flowering, pod formation, taller plants, and early physiological maturity were found in wider (35 cm) spacing (Kumari et al. [2012\)](#page-32-0). This may be due to sufficient availability of light (Rana and Pachauri [2001\)](#page-33-6) and soil nutrition (Kumari et al. [2012\)](#page-32-0). Proper row spacing results in efficient translocation, utilization of photosynthetic products, and better availability of nutrients, moisture, light, and space (Kumari et al. [2012\)](#page-32-0).

In Indian tropical climate, period between August to February was suitable for sowing Camelina, while month of October was found the best for optimum seed production. Seed sown in October yields highest $217-218$ g m⁻², that is, $2171-2180$ kg ha⁻¹.

Camelina was grown in field with standardized agronomic factors at Defence Institute of Bio-Energy Research project site biofuel Park at Harsola, Defence Research and Development Organisation (DRDO) India. They intercropped the Camelina with Jatropha plantation. The sowing was carried both in rows and broadcast; on an average 120 plant per $m²$ of plot were sown. It took 5–9 days for germination, 35–45 days for flowering, and 55–60 days for fruiting (Parker [2014](#page-33-7)).

DRDO-DIBER project site Secunderabad, Andhra Pradesh, and Pithoragarh (India) also under took field trials for its agricultural standardization and intercropping with Jatropha crop (Parker [2014](#page-33-7)). The crop rotations of Camelina with leguminous fodder crops were done. Berseem and Lucerne were rotated with main crop of Camelina. The yield of Camelina seed was recorded improved upto 2000–2200 kg ha⁻¹. The possible reason could be presence of "Sinorhizobium" meliloti" in root nodules of Lucern, which has ability to fix nitrogen in the soil. In the same way, Berseem if ploughed into the soil will act as green manure to the soil and improve the nitrogen content in the soil part (Parker [2014\)](#page-33-7). With above experimental demonstration of different topographic cultivational practices, Camelina exhibits favorable agronomic and environmental conditions of three different zones of Indian subcontinent as an alternative biofuel crop.

Bioactive Constituents in Camelina

According to earlier researchers, the variation in place and environment not only affects the yield but also made a difference in composition.

Camelina has vast number of neutraceutically important bioactive compounds. Bioactive components like different classes of phenolics, glucosinolates, tocopherols, poly unsaturated fatty acids (PUFA), mono unsaturated fatty acids (MUFA), polysaccharides, and lignans are reported by researchers in Camelina. They display their significance in defense mechanism, reducing risk of cancer, and other vital activity of organisms. In prior researches using soybean crop, the saponin, isoflavone, phenol, and flaonoid content antioxidant activity were explored for health benefits (Chaturvedi et al. [2012,](#page-28-2) [2014\)](#page-28-3). It was concluded that by utilization of other bioactive compounds present in oil seeds can be helpful in lowering overall bio-fuel cost. The list of some of these bioactive compounds present in Camelina is mentioned below; it can be helpful in lowering overall biofuel cost. These compounds not only improve the health of the consumers but also affect the cost effectiveness of fuel crops. The list of some of these bioactive compounds present in Camelina is mentioned below.

Glucosinolates

Glucosinolates (GSL) are natural pesticides (Fahey et al. [2003](#page-29-2); Halkier and Gershenzon [2006](#page-30-3)). In defatted Camelina seed meal, the glucosinolate content vary in the range of $10-25 \mu \text{mol/g}$ due to varying growing conditions, genotype, and losses during processing (Berhow et al. [2013\)](#page-28-4). It has both aliphatic and aromatic forms of glucosinolate, and their biological activity is also different.

Camelina seed contains two aliphatic glucosinolates in a significant level; these are glucoarabin (9-(methylsulfinyl)nonyglucosinolate); GSL9 and glucocamelinin (10-(methylsulfinyl) Decylglucosinolate); GSL10 while the third glucosinolate 11-(ethylsulfinyl) undecylglucosynolate); GSL11 is in very small amount (Vaughn and Berhow [2005](#page-36-6)).

Plants having GSL have myrosinase (this is a group of thiohydrolase enzyme) that can cleave glucose from GSL in the presence of water. After forming an unstable intermediate it converts into thiocynate, isothiocyanate, or nitrile, which helps in plant defense (Spencer and Daxenbichler [1980](#page-35-3)).

The indolylglucosinolates (in aromatic form) have been reported to promote some cancers, even though they protect against others (Androutsopoulos et al. [2009\)](#page-27-1).

The glucosinolate (in aliphatic form) hydrolysis products are inhibitors of cytochrome P450 which could be cancerous (Skupinska et al. [2009\)](#page-34-7). It has methyl sulfinyl aliphatic which are similar to glucoraphanin the parent glucosinolates to sulforaphane. The methyl sulfinyl aliphatic provides similar health benefits as sulforaphane (it is an anticancerous compound).

Phenolic Compounds

Flavonoids, tannins, chalcones along with coumarins, and phenolic acids are the main class of phenolics and all of these display antioxidant properties. Flavonoids are the important constituent of food and most potent antioxidant derived from plant (Hertog et al. [1992;](#page-30-4) Jovanovic et al. [1994](#page-31-6)). Majority of flavonoids are present in Camelina like flavonols, flavanols, flavones, isoflavones, anthrocinins or anthrocynadins, and flavonones.

In the Camelina plants, the presence of phenolics is not uniform at tissue, cellular, and subcellular levels. The phenolic compounds remain attached to cell walls but some of them are in the cytoplasmic vacuoles along with various carbohydrates and organic acids. This makes choosing the extraction method difficult. Methanol mix with water is a good solvent for most of the phenolics. Methanol not only disrupts cell wall but also inhibits the action of enzymes (Obied et al. [2005\)](#page-33-8).

The quantification of phenolics is dependent on its chemical nature, assay method, selection of standards, and the interfering substance present (Naczk and Shahidi [2006](#page-33-9)). To remove nonphenolic compounds few additional steps are required (Oomah et al. [1995\)](#page-33-10). After solvent extraction, solid phase extraction, and purification process, it was found that the residue or cake has most of the phenolic content (Koski et al. [2003](#page-31-7); Terpinc et al. [2012\)](#page-35-4).

Fresh Camelina oil has 123 mg kg^{-1} (expressed as chlorogenic acid) of polar phenolic compounds (Abramovič et al. [2007](#page-27-2)). The total phenolic contents were highest in Camelina cake extract (1666 \pm 40)mg CA 100 g⁻¹ (here CA is chlorogenic acid) followed by Camelina seed extract (1536 \pm 91)mg CA 100 g⁻¹ and very low amount is in crude Camelina oil extract (9.1 ± 0.6) mg CA 100 g⁻¹. This high value of total phenolic was possibly due to high temperature and pressure of water during steaming of seeds which resulted in phenolic acids getting released from its conjugated esters (insoluble forms) attached to cell wall of seeds.

Camelina leaf has quercetin in it (Onyilagha et al. [2003\)](#page-33-11). Camelina cake consists of high quercetin glycosides, which display antioxidant activity (Salminen et al. [2006\)](#page-34-8).

Camelina cake has good amount of catechin, quercetin, quercetin-3-0-glucoside, and rutin. However, they were at low levels in oil part.

Among the phenolic acids, only sinapic acid was transferred to oil in high amount. Salcyclic acid and hydroxyl benzoic acid were in low amount in oil. Protocatechuic acid content was higher in cake as compared to seed probably due to steaming and pressing of seed. Þ-hydroxibenzoic acid and salicylic acid were also very high in cake as compared to seed due to heat treatment (Terpinc et al. [2012\)](#page-35-4).

The ellagic acid content in oil is similar to catechin and salicyclic acid. Moreover, it is higher than in cake because of roasting and processing of the seeds (Peričin et al. [2009;](#page-33-12) Terpinc et al. [2012](#page-35-4)).

Most of the antioxidant properties to the scavenging effectiveness of Camelina extract against DPPH radicals are due to phenolic acids, 4-vinyl derivatives, sinapine, ellagic acid, and flavonoids (Hayes et al. [2009](#page-30-5); Lue et al. [2010](#page-32-2); Terpinc and Abramovič [2010](#page-35-5); Terpinc et al. [2011](#page-35-6)). Camelina cake is found to be the best source of total phenolics and shows highest reducing power and also most effective against DPPH radicals. While Camelina oil has low presence of phenolic compounds, it shows high iron-chelating ability and displays protective action against oxidation of emulsion as demonstrated through β-carotene bleaching test. Camelina cake is not only cheap source for natural polyphenol but also easy and reasonable raw material for food or plasma industries (Terpinc et al. [2012\)](#page-35-4).

Camelina seed has different flavonoids out of which flavonol quercetin is in high amount (Onyilagha et al. [2003](#page-33-11)). As compared to other vegetables of cruciferous family, the size of bioactive molecules were found to be much larger in Camelina ranging between $C_9 - C_{11}$.

Tocopherols

Camelina shows high tocopherols and phenolic compounds responsible for oxidative stability than other highly unsaturated oils like flax (Hrastar et al. [2009](#page-30-6); Peschel et al. [2007](#page-33-13)). Since Camelina oil is unstable in high temperature, it is advisable to use in cold dishes. Camelina oil-based salad dressing has similar kind of oxidative stability as sunflower oil. This suggestion was on the basis of value of peroxide, þ-anicinidine, total oxidation, conjugated diene levels, and conjugated triene levels (Eidhin and O'Beirne [2010\)](#page-29-3).

The comparative oil stability test conducted among Camelina tuna fish and salmon oil showed that oil from Camelina was more stable at 60 \degree C for 20 days (Eidhin and O'Beirne [2010\)](#page-29-3). Few researchers observed the improved quality of meat in those birds that used Camelina as food. They found 1.7-fold increases in γ-tocopherols and antioxidant activity in thigh meat, which may be due to tocopherols and other flavonoids supply to the birds through Camelina feed (Aziza et al. [2010b\)](#page-27-3).

The tocopherol content in Camelina seed is 700 mg kg^{-1} (Waraich et al. [2013\)](#page-36-7), in meal it is 200 μ g g⁻¹ (Aziza et al. [2010a](#page-27-4), [b](#page-27-3); Cherian et al. [2009](#page-29-4)), and in oil part it is reported 806 ppm–800 mg kg⁻¹ (Zubr and Matthäus [2002](#page-36-3)).

The amount of α -tocopherol in seed oil is 28 mg kg⁻¹ (Zubr and Matthäus [2002](#page-36-3)), in meal 5.2 mg kg⁻¹ (Aziza et al. [2010a,](#page-27-4) [b](#page-27-3); Cherian et al. [2009\)](#page-29-4), γ -tocopherol in oil 742 mg kg⁻¹ (Zubr and Matthäus [2002](#page-36-3)), in meal 201.7 mg kg⁻¹ (Aziza et al. [2010a,](#page-27-4) [b;](#page-27-3) Cherian et al. [2009](#page-29-4)), δ-tocopherol 20.47 mg kg⁻¹ (Zubr and Matthäus [2002\)](#page-36-3), plastocharomanonol 14.94 mg kg⁻¹ (Zubr and Matthäus [2002\)](#page-36-3).

World's majority of population depends on the eggs and meat to fulfill their omega-3 FA and tocopherols level. If Camelina will be used as fodder for poultry, then this will definitely enhance the quality and quantity of these essential FA and ultimately humans will be benefited by this (Cherian [2012\)](#page-29-5).

Poly Unsaturated Fatty Acids (PUFA), Mono Unsaturated Fatty Acids (MUFA), and Saturated Fatty Acids

Camelina oil contains fatty acids, which are poly unsaturated fatty acid (PUFA), mono unsaturated fatty acid (MUFA), and saturated fatty acid. The Camelina oil has almost 90% of unsaturated fatty acids, with 5.8 double bonds per triglycerides (Kim et al. [2015\)](#page-31-8), higher amount of unsaturated fatty acid and degree of unsaturation in comparison to soybean oil. This quality as Camelina oil makes it useful for industrial applications (derivatives of higher functionality for higher performance biopolymers).

Being the structural component of mammalian cell membranes, n-3 and n-6 polyunsaturated fatty acids (PUFA) are essential component of human diet (Vella et al. [2013](#page-36-8); Ganesan et al. [2014\)](#page-30-7). Three important n-3 PUFA are eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA), and α-linolenic acid (ALA). Out of these, eicosapentaeoic acid (EPA) and docosahexaenoic acid (DHA) are found in marine sources (fish and algae) (Bermúdez-Aguirre and Barbosa-Cánovas [2011](#page-28-5)). While third α-linolenic acid (ALA) can be obtained from Camelina. In general, n-3 PUFA is called "good fats" (Table [3](#page-17-0)). The ratio is 0.90 to 0.70 between ω -6 FA: ω -3 essential FA. ω -3 FAs major portion 29% is of α-linolenic acid (18:3 n-3), while 23% constitute by linoleic acid (18:2 n-6) (Aziza et al. [2010b](#page-27-3)).

Major MUFA is oleic acid, which is followed by eicosenoic acid (20:1). Palmitoleic (16:1) and erucic acid (22:1, $>2\%$) are also present as MUFAs. The eicosenoic acid is used in making polyamide bioplastic (polyamide-11). Research has revealed that eicosenoic acid (C20:1) is present in Camelina in good amount and has demonstrated unique industrial applications (Zubr and Matthäus [2002](#page-36-3)). In saturated fatty acids, Camelina meal has palmitic acid (16:0, 9%) and stearic acid (18:0, 2.5%) (Aziza et al. [2010b\)](#page-27-3), which are also very useful compounds.

If 10% Camelina meal is included in chicken feed, there is threefold increase in ω-3 FAs in meat of chicken and eightfold increase in egg of chicken, which will enhance the nutritional value of meat (Aziza et al. [2010b](#page-27-3)). It is concluded by earlier research that Camelina meal is rich in bioactive compounds, as it is providing ω-3 FAs and simultaneously reducing the oxidative stress (Aziza et al. [2010b\)](#page-27-3).

Polysaccharides (Hydrocolloids)

Like other members of *Brassica* family, Camelina is rich in nonstarch polysaccharides (Budin et al. [1995](#page-28-6)). Mucilage or hydrocolloids are gums derived from natural resources. It can develop high viscosity in low concentrations. It is used both in food or nonfood industries as thickening agent. In agriculture crops, yellow mustard or flaxseeds also produce gums from coat layers of their seeds (Cui [2000](#page-29-6)). Being a member of the mustard family, Camelina can also be used as gum.

Camelina gum (CG) contains polysaccharides (75.1%) and protein (12.3%). The polysaccharide part is formed by four monosugars, glactose (58.1%), glucose (25.0%), rhamnose (11.6%), and xylose (5.2%) (Li et al. [2016](#page-32-3)).

With experiments it was found that at 1% concentration, the viscosity of Camelina gum was more than two times and four times higher than that of commercially available gums such as k-carrageenan and hydroxyethyl cellulose (HEC) (Li et al. [2016\)](#page-32-3). These properties made Camelina as excellent food and industrial thickening agent or stabilizing agent (Li et al. [2016\)](#page-32-3).

Lignans

Cinnamic acid residues or their biogenetic equivalent unite to form lignan and are found in plants. Lignans have their important enantiomers which are same compounds displaying different biological effects. Usually one of the enantiomeric compound is present in certain species (Ayres and Loike [1990\)](#page-27-5). A particular type of enantiomer displays peculiar biological effects, thus it is also important to know enantiomeric composition of lignans in specific food (Smeds et al. [2012](#page-35-7)).

In earlier studies, the lignans are found in berry and oilseed species (Smeds et al. [2012\)](#page-35-7). In prior researches, it has been proved that lignans are concentrated in hull part of cereal grains, so they investigated the hull portion in oil seeds. The lignin is mainly dependent on genetic and environmental variations. They emphasis the hull part of seeds to be used in diet so that the loss of lignans can be recovered, since oil seed crops are generally used after dehulling practices (Smeds et al. [2007\)](#page-35-8). Therefore, it is recommended to use seed intact along with its hull portion.

In another studies lignin was found to be high in Camelina seeds dominated by (-) lariciresinol (LAR) and (+) Secoisolariciresinol (SECO). In addition, hydroxymatairesinol (HMR) 38 μ g 100 g⁻¹ and (7S,8R,8'R)-(-)-7-hydroxymatairesinol (major isomer, called HMR2) were also detected (Willför et al. [2003;](#page-36-9) Smeds et al. [2012](#page-35-7)). Enantiomeric compositions as analyzed by Smeds et al. [\(2012](#page-35-7)) are $(-)$ SYR, $(-)$ MED, $(+)$ PIN, $(-)$ LAR, and $(-)$ SECO (slightly high).

Lignans have wide range of health promoting qualities like anti-tumorigenic, antioxidant, antiviral (Saarinen et al. [2007\)](#page-34-9), anti-obesity, anti-diabetic (Bhathena and Velasquez [2002](#page-28-7)) effects, and protective in cardiovascular diseases (Vanharanta et al. [1999\)](#page-35-9), and so by utilizing intact seed (without hulling) of Camelina we can enjoy the health benefits of lignans present in seed.

Major Applications of Camelina

In total, 80% of Camelina oil is utilized in health-food products. Approximately, 14% of oil is used in industrial raw materials like cosmetics, detergents, surfactants, emulsifiers, lubricants, fuel, oleo chemicals, plasticizers, and adhesives (Carlsson [2009\)](#page-28-0). Use of Camelina as edible oil is very limited, so it can contribute in fulfilling the oil demand of biofuel (Davis et al. [2011](#page-29-7)). This crop has unique fatty acid profile; therefore, it has multiple applications in medicinal, agricultural, nutritional, and biofuel industries as demonstrated in (Hulbert et al. [2012\)](#page-31-9) Fig. [1](#page-11-0).

As Feed Stock for Biofuels

In Egyptian times, plant originated castor oil was used for cosmetics and illuminations. The unprocessed pure raw seed oil is commonly used in lubricating chainsaws, concrete mould release oils, and lamp illuminations (Zanetti et al. [2013](#page-36-2)). At present, it has a variety of applications in different industries such as biodiesel, jet fuel, glycerol, and as biolubricants.

Biodiesel

The benefits of using Camelina as biofuel have been explored time to time by many researchers. Camelina proved as a better low cost biodiesel feed stock (Bernardo et al. [2003](#page-28-8)) due to its lower energy life cycle as compared to canola and soyabean. Camelina can be used as biodiesel additive for improving its characteristics (Reaney et al. [2006](#page-34-10); Johnson [2007](#page-31-10)). After cold pressing and filtration, the oil has improved power output and causes less smoke as compared to petroleum fuel (Wu and Leung [2011](#page-36-10)). The oil of Camelina generates methyl ester similar to rapeseed that can be used in industries; it also has high iodine content and protects against rapid deterioration of the lubricating oil (Fröhlich and Rice [2005\)](#page-30-8).

The fatty acid profile of a transestrified biodiesel is similar to the fatty acid of parent oil. The physical properties and chemical composition of transestrified biodiesel has to be up to the customer's satisfaction. The pure form of biodiesel (B100) should meet the American society for testing and material standards or ASTM D6751 in North America and EN14214 in European Union specified parameters, then only it can be applied for use in diesel engines (Table [1](#page-12-0)). The fatty acid composition of parent oil affects the standards of cetane number, kinematic viscosity, distillation temperature, oxidative stability, and cold flow properties (Ciubota-Rosie et al. [2013\)](#page-29-8). The chain length of parent oil fatty acid esters and its degree of unsaturation affects the properties of biodiesel (Knothe [2005\)](#page-31-11). So, it becomes imperative to check the profile of fatty acid.

The low value (from both the standards) of sulfur (S), phosphorus (P), sodium (Na), potassium (K), calcium (Ca), and magnesium (Mg) is good for the fuel quality of biodiesel. Camelina biodiesel's cetane number is 49.7, whichis almost near to both US and European standards but still not up to the mark. Thus, its use as a fuel

Fig. 1 Various applications of *C. Sativa* (Source: compiled by authors)

during combustion of engine may cause knocking of engine due to incomplete combustion and will produce higher NOx (Yang et al. [2016\)](#page-36-5).

The higher unsaturation degree and its cold soak filtration time is not up to the standards of ASTM D6751 and EN 14214, which could be due to the presence of free steryl glycosides and this may result in cold soak filtration failure. Cost effective

Table 1 Physio-chemical properties and composition of Camelina seed oil suitable for biofinal Table 1 Physio-chemical properties and composition of Camelina seed oil suitable for biofuel

Source: Compiled by authors Source: Compiled by authors

and convenient method should be developed for the transesterification (alkalicatalysed) of parent oil at farm level. The high level of polyunsaturated fatty acids (lenoleic acid 19.1 wt $\%$ and lenolenic acid 33.5 wt $(\%)$ will cause poor oxidative stability and lower induction time. For Camelina the induction time is 1.9 h in comparison to 3 h limit by ASTM D6571 standard. Earlier researchers concluded that addition of antioxidants or blend petro-diesel in specific ratio and standardized amount may improve its oxidative stability and induction time (Yang et al. [2016;](#page-36-5) Liang et al. [2006\)](#page-32-4). Similarly, the other group of researchers suggested that genetic engineering or plant breeding can solve this problem by developing a genetically engineered Camelina crop having less unsaturated fatty acids (Kang et al. [2011\)](#page-31-12).

Jet Fuel

The use of Camelina as jet fuel can reduce the carbon emission up to 84% as compared to petroleum-based jet fuel (Lane [2008](#page-32-5)). Camelina possesses good biofuel properties, and fuel produced by Camelina has been used in military and commercial aircraft (Winchester et al. [2013](#page-36-11); Soriano Jr and Narani [2012;](#page-35-10) Wu and Leung [2011;](#page-36-10) Moser and Vaughn [2010](#page-33-5)).

Hydrogenated renewable jet fuel derived from crop oil is approved by American society for testing and materials standards (ASTM) for use in commercial aviation. Commercial aviation industry of USA sets a target of carbon neutral growth by 2020 and reduction of petroleum generated jet fuel consumption 50% by year 2050 (IATA A [2009](#page-31-13)). Industrial oil seed crops can be helpful in achieving this goal (Fishel et al. [2011\)](#page-29-9). The cost of feedstock for biofuel can be reduced by using food oil crops and developing low cost for hydrogenated renewable jet fuel production (Johnson [2007](#page-31-10)).

To develop the cost effective and reliable feedstock for hydrogenated renewable jet fuel (HRJ), proper cropping system along with its suitability with ecoregion, crop yield balance, agricultural input costs, choice of cultivar, and optimum species needs to be considered (Gesch et al. [2015](#page-30-9)).

In USA the Camelina-based jet fuel is produced by many companies like "Accelergy corp. Attair, Inc.," "Biojet corp., and sustainable oils, LLC" (Moser and Vaughn [2010\)](#page-33-5). In commercial, military and private aviation flights of USA, Camelina-based jet oil with blending are being tested [\(http://www.susoils.com/](http://www.susoils.com/flights.php) fl[ights.php](http://www.susoils.com/flights.php)). This testing not only met all aviation fuel specifications but was successful in both categories of flights. Its application also reduced 75–80% of the carbon emission comparison to petroleum products (Shonnard et al. [2010\)](#page-34-11). This not only provides cleaner environment but also reduces pressure on fossil fuel resources.

Glycerol

The glycerol generated from Camelina can be helpful in many ways due to its nontoxicity and biodegradability as compared to other petroleum sources. In polymer industries, it can be added as stabilizer, plasticizer, and cosolvent in emulsion polymerization. It can also be used as alkyd resins (Güner et al. [2006\)](#page-30-10). Its palmitic and stearic fatty acid esters (mono and di) have been used as emulsifiers in processed foods (Soares et al. [2011\)](#page-35-11).

Glycerol is used in medicinal drugs as nitroglycerin form. Glycerol can form glycerol carbonate, which is an intermediate compound in the chemical synthesis process and useful as gelation agent in polyglycerols formation. Polyglycerols are useful in cosmetics, medicinal applications, and controlled drug release (Behr et al. [2008\)](#page-27-6). In biodiesel production, transesterification of triglycerides with methanol was done to get fatty acid methyl esters (biofuel) and glycerol. In an experimental estimation, 100 kg of biodiesel along with 10 kg of crude glycerol was produced through this process (Soares et al. [2011](#page-35-11)).

As per another report, the properties of fatty acid methyl ester content in Camelina was compared with the EN 14214 and ASTM D6751 standards (Yang et al. [2016\)](#page-36-5). The methanol content is in accordance with both the standards. The standard value of flash point from both the standards is in the range of $93-101$ °C; however, the value of Camelina oil is 152 \degree C which makes it much safer for transportation and storage point of view. Other important values like density, kinematic viscosity, acid number, water content free glycerol, total glycerol, mono-, di-, and triglyceride content were in compliance with both the standards (ASTM D6751 and EN 14214).

Biolubricants

Mineral oil based lubricants are being replaced by vegetable oil based lubricants. As far as degradability is concerned, petroleum based lubricant leads to carcinogenic toxicity (Dalbey and Biles [2003\)](#page-29-10), while biolubricants are nontoxic and safe for environment (Luna et al. [2011](#page-32-6)). It also displays the performance benefits like higher flash point, higher detergency, better lubricity, lower volatility, higher viscosity indices, lower compressibility, higher resistance to humidity, higher shear stability, and higher dispersancy (Rani et al. [2015](#page-33-16); Cermak et al. [2013](#page-28-9)). However, they have certain limitations such as feed stock reliability, poor low temperature performance, and low thermal oxidative stability (Soni and Agarwal [2014\)](#page-35-12).

A lot of research has been undertaken to improve these physical properties. The feed stock for developing biolubricants is vegetable oil and mainly triglycerides, esters of glycerol, and three straight chained fatty acids (C12–C24 range fatty acid) (McNutt [2016](#page-32-7)). Monounsaturated fatty acid like oleic and palmitoleic acid found to have good balance of low melting point with good thermo-oxidative stability and viscosity (Nagendramma and Kaul [2012](#page-33-17)). Genetically modified varieties of Camelina were used which can alter its seed oil composition (Liu et al. [2015;](#page-32-8) Gryglewicz et al. [2013\)](#page-30-11). In current researches vegetable oil generated biolubricants were formed by chemical modification with the help of esterification/transesterification, estolide formation, and epoxidation methods (McNutt [2016\)](#page-32-7). The Camelina generated biolubricants are good in quality and safe for the environment.

Animal Feed

The Camelina base feed is given to different terrestrial animals, aquatic animals, and birds to improve their general health. It is also helpful in introducing some peculiar

health enhancer (like ω-3 fatty acids, antioxidants) to human as they consumed their meat. Previous researchers used Camelina seeds and cakes as feed in terrestrial animals, aquatic animals, and birds.

Terrestrial Animals

Camelina meal is full of mineral content that makes it suitable for animal feed. It has 10,214–12,231 ppm phosphorus, 13,204–14,879 ppm potassium, 2,597–2,703 ppm calcium, 4,696 ppm magnesium, 18.7 ppm chloride, 9,122 ppm sulfur, 15.4–17.6 ppm sodium, 151 ppm iron, 25.1 ppm manganese, 61.1 ppm zinc, 9.18 ppm copper, and 5.37 ppm aluminum (Frame et al. [2007;](#page-29-11) Cherian et al. [2009\)](#page-29-4). Even Camelina meal can be used as additive for animal feed because it has good amount of protein and omega-3-FA (Ryhänen et al. [2007;](#page-34-12) Rokka et al. [2002](#page-34-13)) (Table [2](#page-15-0)). It is also rich in protein and poly unsaturated fatty acids (PUFAs) (Hurtaud and Peyraud [2007\)](#page-31-14) and can be used in cattle or pig feed (Cappellozza et al. [2012;](#page-28-10) Ciuca [2013](#page-29-12)). Camelina meal has high amount of protein and energy, which can be a good source for ruminant feeds (Matthäus and Zubr [2000](#page-32-9)).

Being a good source of polyphenolic compounds, Camelina meal could be given in the ratio of 1:23 in the diet of dairy cows (275.67 \pm 15.19 mg GAE l VS⁻¹ 189.37 ± 8.67 mg GAE l VS⁻¹). It was found that polyphenols in plasma of cow fed with Camelina meal was around 12.31 ± 7.17 mg GAE l⁻¹ (Chedea et al. [2014\)](#page-29-13).

In dairy cows, this meal can reduce milk fat, resulting in more spreadable butter (Hurtaud and Peyraud [2007\)](#page-31-14). Addition of 10–20% cake of Camelina in the feed of ewes could change the aroma and the content of volatile compounds in milk (Cais-Sokolińska et al. [2013;](#page-28-11) Szumacher-Strabel et al. [2011\)](#page-35-13).

Few researchers found that Camelina seed is helpful in reducing atherogenic and thrombogenic indexes of rabbit meat (Peiretti et al. [2007\)](#page-33-18). If it is added to rabbit feed (upto 15% addition), it will increase concentration of poly unsaturated fatty acids

				Zubr	Cherian et al.	Aziza et al.	Sampath
References	Bullerwell et al. 2016			2010	2009	$2010a$, b	2009
Composition	Seed	HORM	SECM	Cake	Meal	Meal	Cake
Dry matter $\%$	91.9	89.5	90.8			$\overline{}$	
Gross energy kcal/ kg	6123	4653	4270		4600	4755	
Ash	3.78	5.89	6.29		6.5	6.5	\equiv
Crude Protein%	25.2	35.7	39.0	45	36.2	\equiv	40
Residual oils%	34.6	9.9	2.8	10	\equiv	\equiv	15
Acid detergent fiber $\%$	23.6	18.3	18.0	15	41.8	8.4	12
Glucosinolates (GSLS) and flavonoids%				14	21.2	21.2	

Table 2 Proximate composition of Camelina seed cake and defatted cake

Source: Compiled by authors

– Not available

and decrease the concentration of saturated FA in the body of rabbits. It was also found that addition of more than 5% of Camelina meal in pig feed can cause reduction in growth and meat palatability (Flachowsky et al. [1997\)](#page-29-14).

Aquatic Animals

Prior researchers included Camelina not only in the form of meal but also used oil and other forms in aquatic animals and ended up with encouraging results. Camelina seeds can produce oil, pressed meal, and solvent extracted Camelina meal (SECM). Camelina seed has 35% α-linolenic acid and 15% linoleic acid as essential FA. To get meal from seed, cold pressing technique is used, which produces press cake. The meal has still some residual oil (approximately 10%), 45% crude protein, and 12.8% of crude fiber. This kind of meal is called "high oil residue meal" (HORM). If this meal is again processed to remove oil by solvent extraction technique, it is called "solvent extracted Camelina meal" (SECM), which has high protein content than HORM (Korsrud et al. [1978](#page-31-15)).

In earlier research, the effect of Camelina meal and oil as feed for rainbow trout, Atlantic cod has been tested (Betancor et al. [2015;](#page-28-13) Ye et al. [2016](#page-36-13); Hixson et al. [2014a](#page-30-12), [b;](#page-30-13) Morais et al. [2012\)](#page-32-10). According to experimental data, the use of fish oil can be replaced with Camelina oil in Atlantic salmon diet, without any impact on weight gain (Hixson et al. [2014a](#page-30-12)). In prior research, it was found that in rainbow trout if HORM is given up to 160 g/kg, it has no adverse effect on growth performance (Pan et al. [2011](#page-33-19)). The other group experimented on Atlantic salmon, and they have given SECM at the rate of 50 g kg⁻¹ rate. Lower level (50 g kg⁻¹ SECM) did not affect fish performance while higher doses (100 g kg^{-1}) did affect the fish health (Ye et al. [2016\)](#page-36-13). Some scientists performed 112 days trial on rainbow trout and found that 100 g kg^{-1} Camelina seed, HORM, or SECM in the diet does not affect negatively on the performance of fish, while the obtained fish meat is having better nutrition as compared to fish feed with conventional feed (Bullerwell et al. [2016\)](#page-28-12). Hence, it is clear that Camelina products as diet are suitable. HORM, SECM, and seed has good amount of micronutrient (Bullerwell et al. [2016\)](#page-28-12).

The metabolic or protective function in plant may be due to presence of antinutritive factors (ANF) but may impair the nutritive value (Blades and Zubr [2010;](#page-28-14) Matthäus and Zubr [2000\)](#page-32-9).

Birds

Camelina meal is rich source of both protein and ω -3 FA, so it is a very good food supplement for birds. Cake or meal is utilized in poultry (Rokka et al. [2002](#page-34-13)). It is also being given to young turkey (Frame et al. [2007](#page-29-11)) and layer birds (Cherian et al. [2009\)](#page-29-4).

Camelina oil mixed with chicken feed increases n-3 (omega-3) content in the egg without any unpleasant flavor (Rokka et al. [2002](#page-34-13)). If meal is added to broiler chicken feed, it may reduce feed intake and growth of chickens, but it improves ω-3-fatty acids which is indirectly beneficial for human nutrition when they consume them (Ryhänen et al. [2007](#page-34-12)). Few researchers found that if more than 5% meal of Camelina is used as feed, then the weight of young turkey will decrease (Frame et al. [2007](#page-29-11)). In another research, the layer birds feeding higher level of energy, protein, essential ω -3

Table 3 Camelina seed oil fatty acid composition reported in different countries Table 3 Camelina seed oil fatty acid composition reported in different countries

Source: Compiled by authors

FA, and n-6 FA was found in the meal of Camelina. But more than 10% can adversely affect the health of birds (Cherian et al. [2009](#page-29-4)).

Human Feed

Using Camelina as animal feed encouraged researchers to use them as human feed. As already proved that Camelina oil is a rich source of essential FA (linoleic and α-linolenic acids) as well as ω-3 FA (α-linolenic) (Hrastar et al. [2009](#page-30-6); Zubr and Matthäus [2002\)](#page-36-3) (Table [3\)](#page-17-0). There are many varied ways to consume this nutrient rich meal and from last few decades, research is going well on this aspect. They utilized it in direct or indirect form of diet.

In Form of Edible Oil

Camelina oil can be promoted in the form of dietary supplement in human and animal nutrition (Obour et al. [2015\)](#page-33-4). The crushed form of Camelina seeds was used as one of the ingredients in forming bread (Zubr and Matthäus [2002](#page-36-3)). The cultivation conditions (genotype, location, fertilizer, and environmental conditions) affect the profile of Camelina fatty acids (Jiang et al. [2014](#page-31-3)). This Camelina oil after purification can be used in different cooking process as cooking medium or as additives, while high erucic acid limits its use (Zubr [1997](#page-36-14)).

Camelina Meal as Patties

Camelina meal is nutritionally very good as it has high ω -3-fatty acid (>35%), vitamin E content, crude protein $($ >45%), and fiber content $(10-11)$ (Meadus et al. [2014\)](#page-32-12). Some researchers used the protein, lipid, and antioxidant high meal to prepare pork meat patties (Salminen et al. [2006](#page-34-8)). This form of preparation can be easily accepted by consumers, and they are benefited with its high nutritional qualities.

Camelina Oil in Fortified Yogurt Form

Our daily diet contains naturally bioactive compounds but fortified food is also available in market. In human diet, n-3 and n-6 polyunsaturated fatty acids are very important component. These "healthful fats" are helpful in maintaining normal health of humans.

The recommended basic requirement of PUFA (ALA, EPA, and DHA) in human body is 250 mg by European council, 2006, while 2 g dose was recommended by European Union 2012 (European council regulation no. 432/2012). Globally, present intake of ω-3 PUFA is not sufficient (Sioen et al. [2009](#page-34-16)).

Marine sources for n-3 PUFA have fishy flavor which limits its use in food, so the vegetable sources were accepted for food fortification (Dal Bello et al. [2015\)](#page-29-15). Vegetable sources as explored by many researchers are flaxseed, soybean, kiwi, rapeseed, echium, raspberry, and Camelina (Botelho et al. [2013](#page-28-15); Waraich et al. [2013;](#page-36-7) Ganesan et al. [2014\)](#page-30-7).

Bello et al. [\(2015](#page-29-15)) used five different vegetable oils and fortified with skim milk before lactic acid fermentation at 10% of daily recommendation 2 g day⁻¹ of α-lenolenic acid according to European council regulations that is around 200 mg

of 125 g per serving size (Dal Bello et al. [2015\)](#page-29-15). To prevent oil from rising to the surface, the modified vegetable originated starch Novation indulge 1720 (Prodotti Gianni S.p.A, Milan, Italy) was mixed with oil before adding it to milk. The milk was slightly heated (5 min at 60 \degree C and cooled to 42 \degree C) and starter is added to it. The starter is cultures of Lactobacillus delbruccki ssp. Bulgaricus and Streptococcus thermophilus (LYOFAST Y450B, Clerici-Sacco, Milan, Italy). The microbiological, chemical, and sensory evaluation was done of fortified yogurt after 0, 7, 14, and 21 days of storage at 4° C.

The oil content addition does not affect the growth of lactic acid bacteria, and all yogurts were accepted by consumers except raspberry and E-plantagineum oil. Many of the yogurts were sensorially appreciated specially when fortification was done with the oil of flaxseeds and Camelina seeds. So by this α-linolenic acid with significantly higher amount is provided with natural fortification in a yogurt form (Dal Bello et al. [2015](#page-29-15)).

Bakery Products and Juices

The ω-3 PUFA benefits (Welch et al. [2010\)](#page-36-15) can be used in infant formula, some dairy meat (Özer and Kirmaci [2010;](#page-33-20) Escobar et al. [2011](#page-29-16)), and bakery products, and juices (Ganesan et al. [2014\)](#page-30-7) have been used as vehicles for EPA and DHA fortification. These fortified backery items and dairy meat products are easily accepted by consumers and simultaneously enjoy the benefits of nutrients.

Medicinal Therapeutic Applications

Health Promoting Supplements

As earlier mentioned, Camelina cake contains a good amount of bioactive compounds such as glucosinolates, vitamins, and antioxidants (Matthäus [2002;](#page-32-13) Eidhin et al. [2003\)](#page-29-17). α-linoleic acid substrate is helpful in human metabolism. By including Camelina oil in diet, the n-6/n-3 fatty acid proportion improves in food. It has high α-linoleic acid, tocopherols, and antioxidants contents (Simopoulos [1999](#page-34-17)). It helps to improve general health (Rokka et al. [2002](#page-34-13); Lu and Kang [2008\)](#page-32-14). Lignans present in Camelina has antioxidant and antiviral properties (Saarinen et al. [2007](#page-34-9)). It also displays antidiabeteic and antiobesity qualities (Bhathena and Velasquez [2002](#page-28-7)).

Camelina extract has natural phenolic compounds and have antioxidant activity (Matthäus [2002;](#page-32-13) Zubr and Matthäus [2002](#page-36-3)). Camelina oil contains 760 mg tocopherols kg^{-1} of oil (Abramovič et al. [2007\)](#page-27-2). Tocopherols not only act as natural antioxidants but also provide oxidative stability to the oil.

Cake of Camelina seed contains many phenolic compounds, which showed antioxidant activity like tocopherols, sinapine, and sinapic acid (Salminen et al. [2006\)](#page-34-8). Flavanols and flavonols both are also responsible for antioxidant activity (Salminen and Heinonen [2008\)](#page-34-18).

Ellagic acid (a polyphenol with four OH groups) is also an important antioxidant showing high radical scavenging activity (Hayes et al. [2009;](#page-30-5) Terpinc et al. [2012](#page-35-4)).

A major phenolic ester sinapine (sinapoylcholine) can be derived from sinapic acid (Matthäs [1997](#page-32-15); Thiyam et al. [2004\)](#page-35-15). Sinapine found in Camelina cake is one of the effective antioxidant (Matthäus and Zubr [2000](#page-32-9); Salminen et al. [2006](#page-34-8)). Camelina has another kind of phenolic compound 4-vinyl derivatives of hydroxycinnamicacid; it is formed by enzymatic or thermal decarboxylation (Vanbeneden et al. [2008\)](#page-35-16). 4-vinyl derivatives contain antioxidant properties; it reduces 2, 2-diphenyl-1 picrylhydrazyl (DPPH) radicals and the superoxide anion radicals (Terpinc et al. [2011\)](#page-35-6). These four vinyl derivatives were lowest in oil, lesser in cake than its defatted seeds. This may be due to the steaming and crushing of seeds where highly volatile nature of these compounds gets lost (Terpinc et al. [2011\)](#page-35-6).

Burns and Inflammations

Camelina oil is also helpful in treatment of burns, wounds, and eye inflammations, and it is applied topically (Rode [2002](#page-34-15)). It has anti-inflammatory properties (Sampath [2009\)](#page-34-14). It can be used as remedy for the treatment of burns, wounds, and eye inflammations. Camelina oil also helps in regeneration of cells, recovery of skin elasticity, and slenderness, and so improves skin condition (Vollmann et al. [1996](#page-36-16)). In Slovenia, Camelina oil is used as traditional home remedy (Rode [2002\)](#page-34-15). In USA, this oil is used in cosmetics and in biofuel industries. Camelina oil has not yet GRAS status, but in France and UK and Denmark it has legal food status (Sampath [2009\)](#page-34-14).

Coronary Heart Diseases

Camelina oil is also very helpful in preventing coronary heart disease, arrhythmias, and thrombosis (Trumbo et al. [2002\)](#page-35-17). In general, n-3 PUFA are called "healthful fats." They are helpful in maintaining normal blood pressure, levels of triglycerides in blood, reduces the risk of cardiovascular diseases, protects in some cancers and tumors, also good for brain, nervous system, and retina of eye (Harris et al. [2008;](#page-30-16) Gogus and Smith [2010\)](#page-30-17). Some lignans may also have protective ability for cardiovascular diseases (Willför et al. [2003](#page-36-9)).

Neurological Abnormalities

Deficiency of α linoleic acid results in neurological abnormalities and poor growth. To cure this deficiency Camelina oil can be added to the diet that will make available α linoleic acid (ALA), eicosapentaenoic acid (EPA), and docosahexanoic acid (DHA) components (Trumbo et al. [2002\)](#page-35-17). By adding Camelina in diet, one can reduce the risk of poor growth and abnormalities.

Cholesterol Reduction

Camelina has cholesterol reducing effect on mildly and moderately hypercholesteromic subjects (Karvonen et al. [2002\)](#page-31-17). It is therefore advisable to add Camelina in any acceptable form to the daily diet of hypercholestromic patients.

Ulcers

In traditional home treatments, Camelina oil is useful in the treatment of stomach and duodenal ulcers (Rode [2002](#page-34-15)). Following previous work it was found that

sulforaphane and other hydrolysis products may have multiple sites of action, upregulation of the phase II detoxification enzyme NAD(P)-H: quinoneoxidoreductase 1 (NQ01) is often used as a biomarker of beneficial bioactivity (Cuendet et al. [2006](#page-29-18)). Oil of Camelina gives soothing effect to the ulcer patients.

Cancers and Tumors

Lignans are antitumorigenic (Willför et al. [2003\)](#page-36-9). Some researchers concluded that the consumption of isothiocyanaes showed chemoprotective activity if consumed by mammals (Jeffery and Araya [2009\)](#page-31-18). It not only acts against tumor initiation but also helps in steroid hormone metabolism modification. Isothiocyanate ingestion induces detoxification enzymes, inhibits enzyme activation, and by these overall helps in improvement of consumers defense system.

The aliphatic glucosinolates hydrolysis products behave as inhibitors of cytochrome P450 activation of carcinogens (Skupinska et al. [2009](#page-34-7)). The form of glucosinolates in Camelina is methyl sulfinyl aliphatic which are similar to glucoraphanin the parent glucosinolates to sulforaphane. This methyl sulfinyl aliphatic provides similar health benefits as sulforaphane (anticarcinogenic).

In same lines, other researchers added aliphatic glucosinolate hydrolysis products along with NQ01 were associated with upregulation of anti-inflammatory, antioxidant, and anticarcinogenic effects (Androutsopoulos et al. [2009](#page-27-1)). Following research evaluated induction of phase I and phase II detoxification enzymes, cytochrome P450 1A1 (CYP 1A1) and NAD(P)-H:quinine oxidoreductase1 (NQ01) as biomarkers of potential bioactivity (Das et al. [2014](#page-29-19)). They also found the impact of glucosinolate 9 and 10, quercetin, and rutin in Camelina seed and defatted meal extract (Das et al. [2014\)](#page-29-19). The role of various bioactive components like rutin, catechin, quercetin, and flavonoids are natural powerful antimicrobial, antimutagenic, anticarcinogenic, and antioxidant (Rice-Evans et al. [1997](#page-34-19)); since Camelina is rich in these bioactive compounds it may help in reducing the number of cancer sufferers.

Industrial Use

There is huge possibility to use Camelina oil in manufacturing of many bio-based products. Since the oil has about 90% of unsaturated FA, it contributes in quick drying, thus useful in making polymers, paints, varnishes, cosmetics, and dermatological products (Kasetaite et al. [2014;](#page-31-19) Hixson et al. [2013](#page-30-18); Merrien et al. [2012;](#page-32-16) Zaleckas et al. [2012](#page-36-17)). It can also be used in many other industries like lubricants, resins, coatings, and adhesives (Obour et al. [2015\)](#page-33-4). Researchers optimized the method of epoxidation by formic acid and hydrogen peroxide for Camelina oil, which have the potential to be used in biopolymer industry to produce pressure sensitive adhesives, resins, and coatings (Obour et al. [2015;](#page-33-4) Kim and Netravali [2012;](#page-31-20) Kim et al. [2015\)](#page-31-8).

Bioadhesive

It is reported that the amino acid profile of Camelina protein is similar with canola protein. This was already reported that canola protein has a potential to be replace the

conventional petroleum based adhesives (Li et al. [2011](#page-32-17), [2014](#page-32-18)). Likewise soybean also shows promising bio-based adhesive source (Qi et al. [2012](#page-33-21); Qi and Sun [2011\)](#page-33-22), but since soybean has been used for human food consumption. Alternatives like Camelina should be explored as bioadhesives (Li et al. [2015](#page-32-19)). It was reported that albumin, globulin, and gluteline are three fractions of protein from meal of Camelina (Li et al. [2014](#page-32-18)). To test the adhesive quality water solubility, protein electrophoresis profile, crystallization, thermal, morphological and rhenological properties were examined. Since, albumin has strong water swallowing properties it has poor ability as adhesive (Li et al. [2015\)](#page-32-19). The adhesive strength of Camelina protein fractions were isolated from defatted Camelina meal were studied and they are globulin and glutelins. The results showed that overall adhesive strength of globulin is better than glutelin fraction. He also found that glutelin had higher protein aggregation than globulin. Due to glutelins compact structure the properties of adhesion is lower than globulin (Li et al. [2015](#page-32-19)). Pressure sensitive adhesives could be useful in many applications like graphics, medical supplies, tapes, and labels (Tse and Jacob [1996;](#page-35-18) Satas [1989](#page-34-20)).

The majority of pressure sensitive adhesives, which are useful in forming adhesion joints, are being manufactured by petroleum based acrylated monomers (Li et al. [2014,](#page-32-18) [2015](#page-32-19); Czech [2007\)](#page-29-20). The oleo chemicals and biopolymers are also used for adhesion which was derived from oil of Camelina (Kim et al. [2015](#page-31-8)).

Acryicpolyol has both vinyl group and multihydroxyl functionalities; Camelina oil can synthesize acrylic polyol with some reaction steps like epoxidation, partial acrylation, and dihydroxyletion of remaining epoxy group (Kim et al. [2015](#page-31-8)). In these steps, various intermediates were formed like epoxidized Camelina oil (ECO), acrylated epoxidized Camelina oil (AECO), and dihydroxyl acrylated epoxidized Camelina oil (DAEO), which is an acrylic polyol. Acryl polyol offers good polar attraction to substrate when used as pressure sensitive adhesives. Addition of a moderate amount of rosin ester reduces adhesive plateau modulus and energy dissipation while maintaining high cohesive strength. This will convert PSAs with good peel, tack, and shear performance (Kim et al. [2015](#page-31-8)).

Bio Alkyd Resin

In paint industries, a commercial polymer alkyd resins is used and it is useful in metal, wood or wood based materials, cement, and gypsum plasters. The soybean oil and linseed oils and synthetic pentaerythriol and pthalic anhydride were used as raw material in the preparation of alkyd resins. These synthetic compounds (pentaerythriol and pthalic acids) were toxic for the environment (Gandini [2011\)](#page-30-19). For future production of alkyd resins, the fatty acid raw material should be in large amount and it should not be in direct competition with our food crops. Camelina is appropriate for such type of selection (Nosal et al. [2015\)](#page-33-14). The number and distribution of hydroxyl groups in polyols are important for its drying properties. Therefore, resins produced from pentaerythriol showed better coating than glycerol. Pentaerythritol dry out faster, turnout harder, and more resistant toward moisture (Erhan [2005](#page-29-21)). The glycerol is not only easily renewable raw material but also economical cheap so more commonly used for coating raw matter (Tuck [2000\)](#page-35-19).

In some researches, they have developed one pot synthesis of alkyd resin by Camelina oil (Nosal et al. [2015\)](#page-33-14). In the presence of LiOH (0.1 wt%) at 245 °C temperature, the oligomerization of glycerol was done. This oligomerized product was alcoholized with purified Camelina oil. This alcoholysis product was again gone through polycondensation with pthalic and malic anhydrides at $230-250$ °C temperature and form alkyd resin (Nosal et al. [2015\)](#page-33-14).

Nowadays, we try to reduce the edible oil which is generally consumed in industrial purposes and they have lower unsaturated fatty acid than in normal oils. As comparison to other oil seed crops, Camelina oil has high iodine number and is suitable for drying oils for producing paints.

The key parameters of prepared resins indicate that Camelina oil proves to be very promising raw material. Alkyd resin can be produced from diglycerol and polyglycerols with some properties like flexibility and drying time similar to the product from pentaerythriol. The varnish films formulated from alkyd resin based on penetration by scratching than those based on diglycerol and far better than poly glycerols. The resin formed with pentaerythritol also has better hardness than glycerol and poly glycerols (Nosal et al. [2015\)](#page-33-14).

Cosmetics and Soaps

Oil of Camelina is very useful in the small scale industries like soaps and cosmatics. In USA, small businesses like Manor hall soap company springfield; Siberian Tiger Naturals Inc. Seattle, etc., make soaps for smaller markets. Some cosmetics like lotions and creams are also manufactured with Camelina oil (Obour et al. [2015;](#page-33-4) Bernardo et al. [2003\)](#page-28-8). There is a group of people who prefer vegetable oil in their cosmetics (herbal cosmatics), so for them these Camelina based products is preferable.

Film, Fibers, and Thermoplastics

Camelina oil seed could be used for films, fibers, and thermoplastics when grafted with various acrylates. For this application, optimization to increase its tensile properties should be done (Reddy et al. [2012](#page-34-21)). Researchers have been done to improve its tensile qualities and applications in manufacturing of fibers, films, and other useful products.

Bio-Gum

Hydrocolloids (polysaccharides) are used in industries as thickening agents, swelling agents, emulsify agent, crystal inhibitors, foam stabilizers, gelling agents, and water bonding agents (Walter [1997](#page-36-18)). These are also known as natural gums. Camelina seed is also one of the natural resource for making these gums. Camelina meal showed distinct quality of binding in water (Taasevigen [2010\)](#page-35-20). Soil exposure and erosion can be amended with the Camelina gum during construction works (Grady and Nleya [2010](#page-30-20)).

During their extensive research on Camelina gum, it was found that the physiochemical and rheological properties are mostly similar to hydrocolloids (Li et al. [2016](#page-32-3)). Camelina gum has thinning behavior which may be due to six mono sugars, high viscosity, storage modulus, pH, and temperature tolerance.

Bioplastic

Camelina is rich source of uncommon fatty acid eicosenoic acid used in making poly amide-11 a form of bioplastic (Zubr and Matthäus [2002](#page-36-3); Zanetti et al. [2013\)](#page-36-2). Researchers have isolated protein fractions from Camelina meal (Li and Sun [2014\)](#page-32-20). They tried to develop biodegradable plastics and composites from meal of Camelina (Kim and Netravali [2012](#page-31-20); Reddy et al. [2012](#page-34-21)). Still there is a dearth of techniques to produce high-value chemicals and polymers from oil of Camelina (Balanuca et al. [2014;](#page-27-7) Kasetaite et al. [2014;](#page-31-19) Kim et al. [2015\)](#page-31-8).

Cuticular Waxes and Suberins

Plant waxes have aliphatic and phenolic based barriers which protect them against environmental interfaces like water loss and various stresses. The aerial surfaces of plant are coated with cuticle which is made from cutin and waxes (Samuels et al. [2008\)](#page-34-22). The cutin polymer forms structural backbone of cuticle. Cutin is formed of glycerol and long-chained (C16–C18) fatty acid derivatives and covers the cell wall of plant. The cuticular waxes are mixtures of low polarity compounds, including very long chain (\geq C20) straight chain aliphatics, triterpinoids, and phenolic lipids (von Wettstein-Knowles [2012\)](#page-36-19). Cuticular waxes limit nonstomatal water loss (Vogg et al. [2004](#page-36-20)). Epicuticular waxes are in direct contact with the environment; they work on insect, pathogen interaction, and also help on dust water and spores shedding (Kerstiens [1996](#page-31-21)). The composition of cuticular waxes varies between species, organs of the same plant, development stages, and environmental condition (Jetter et al. [2008](#page-31-22)).

Suberin is also a hydrophobic surface barrier; it is polyester of glycerol, phenolic, and fatty acid derivatives (Franke and Schreiber [2007](#page-29-22)). It is found in cell walls inner surface (endodermis and peridermis) called root waxes (Kosma et al. [2012;](#page-31-23) Li et al. [2007](#page-32-21)).

In Camelina, these surface waxes make tolerance to abiotic and biotic stresses (Razeq et al. [2014\)](#page-34-1). The wax mixture obtained from stem, leaves, flowers, seeds, and root part gives qualitative and quantitative information of Camelina boundary tissues wax. This work also made the understanding about wax biosynthetic pathways in every organ of plant. The knowledge of Camelina wax protective surface, drought tolerance, and pathogen-resistant properties will help to make renewable high value natural products of industrial utility (Razeq et al. [2014](#page-34-1)).

Organic Fertilizer

Oil cakes of different plants have been utilized as compost for years and increased yield is reported for crops like pongamia for tomato crop (Chaturvedi et al. [2013\)](#page-28-16) and

jatropha compost for tomato crop (Chaturvedi et al. [2009,](#page-28-17) [2010](#page-28-18), [2013;](#page-28-16) Chaturvedi [2011;](#page-28-19) Chaturvedi and Kumar [2012](#page-28-20)). Camelina cake having high protein content may be used in animal feed and as fertilizer. With new researches this cake is also utilized in the production of industrial enzymes, antibiotics, biopesticides, vitamins, and other biochemicals (Ramachandran et al. [2007](#page-33-23)).

Pesticide or Insecticide and Antifungal Activities

Camelina can be used as potent antifungal, pesticidal, or insecticide in the field crops. When Camelina meal is applied at 5% and 1% to soil, it suppresses the Phymatotrichopsis omnivore (Duggar) sclerotial germination and hyphal growth of fungi (Hu et al. [2011](#page-30-21)). This fungus causes cotton root rot to the cotton and Alfalfa production; by using Camelina meal one can protect fungal attack (Hu et al. [2011\)](#page-30-21).

GSLs are secondary metabolites and act as natural pesticides and prevent herbivory (Fahey et al. [2003](#page-29-2); Halkier and Gershenzon [2006](#page-30-3)). GSL in the presence of water after an unstable intermediate formation converts into thiocynate, isothiocyanate, or nitrile. These compounds help in defense of plant (Spencer and Daxenbichler [1980\)](#page-35-3).

Shelf Life Enhancer

The oil extracted by mechanically pressed Camelina seed called virgin, crude, or unrefined oil. It has mustard like odor with mild nutty notes and golden yellow in color. This crude oil showed 12–24 months shelf life without change in flavor and aroma under normal environmental conditions of storage (Crowley and Fröhlich [1998\)](#page-29-23). The oil can get degraded by light rather than temperature (Abramovic and Abram [2005](#page-27-8)).

This unrefined Camelina oil has a natural antioxidant (Tocopherols) in it which makes it stable in ambient storage conditions (Sampath [2009](#page-34-14)). Among tocopherol, the γ-tocopherol (vitamin E) is responsible for good shelf life and it does not need special storage conditions (Zubr and Matthäus [2002](#page-36-3)).

Oil having high omega-3-FA shows low shelf life (Eidhin et al. [2003](#page-29-17)). Camelina has very high omega-3-fatty acid so its oxidative stability could be also low. But it was observed that Camelina displayed longer shelf life than similar oils. In conditions of room temperature, it has higher stability than fish oil (Eidhin et al. [2003\)](#page-29-17). This quality will be used both for meal and oil during storage transportation (Brandess [2007](#page-28-21)).

Phytoremediation

Bioenergy crops can be used for phytoremediation activity for polluted lands. Apart from application in cleaning-up process, several end products can be obtained like charcoal, biodiesel, bioethanol, wood, alkaloids, fiber, bioplastics, etc. (Tripathi et al. [2016\)](#page-35-21). Utilizing polluted and abandoned land for the production of bioenergy with phytoremediation is getting attention globally (Abhilash et al. [2013](#page-27-9)). The growth of the crop or production will be hampered due to several pollutants toxicity (like metal, metalloids, persistent organic pollutants, petroleum hydrocarbons, organo phosphorus pesticides, carbamate insecticides, herbicides, radio nuclides, nanoparticles, and other new and emerging pollutants).

To overcome this problem, appropriate crop species and agronomical practices must be optimized (Tripathi et al. [2016](#page-35-21)). The photoproducts generated at the end may have pollutants, so it is a kind of challenge to research on these aspects (Abhilash et al. [2013\)](#page-27-9). Camelina can phytoremediate cadmium (Cd) and atrazine along with biofuel production (Fairley [2011](#page-29-24); Graham-Rowe [2011](#page-30-22)).

Reasons for Low Acceptability by the Growers

Despite its several applications, Camelina has some constraints which affect its production and economic viability. The initial problem is with its cultivation. Farmers' faces lack of information regarding specification of genotype recommendations in particular type of climate (winter or spring). There is less awareness about best agronomic practices in cultivation (Obour et al. [2015\)](#page-33-4). At existing production cost and lack of markets, Camelina is a less profitable crop. Harvesting and postharvesting losses also makes it less beneficial as oil seed crop (Obour et al. [2015\)](#page-33-4). Shattering and uneven maturation causes loss in yield (McVay and Lamb [2008;](#page-32-1) Lenssen et al. [2012](#page-32-22)). Downy mildew infestation causes lesser outcome of seeds (Obour et al. [2015](#page-33-4)). Consequently, farmers face loss by cultivating Camelina in fields leading to less preferable crop.

Another problem arises with its compositional aspect that is antinutrient. Camelina meal contains erucic acid, sinapine, and glucosinolates as antinutrient which may be harmful for human health. In Camelina, the amount of tannins are 2.2 mg g⁻¹ and 2–4 mg kg⁻¹ sinapine (Meadus et al. [2014;](#page-32-12) Matthäs [1997\)](#page-32-15). The presence of phytic acid in Camelina causes poor bioavailability of minerals and proteins $(19 \text{ mg } \text{g}^{-1})$ (Matthäs [1997\)](#page-32-15). The effect of glucosinolates (19–25 mmol kg^{-1}) causes impairment of growth and fertility, irritation of gastrointestinal mucosa, followed by necrosis in livestock (Meadus et al. [2014](#page-32-12); Russo and Reggiani [2012](#page-34-3); Matthäs [1997\)](#page-32-15). The erucic acid 2.4–5% can cause fat deposits in heart muscle and myocardial lesions in experimental animals (Pavlista et al. [2011;](#page-33-2) Zubr and Matthäus [2002](#page-36-3)). According to some research, Camelina has some properties like unusual fatty acid profile, high α -linoleic acid, low erucic acid, unusually high cholesterol content (45 mg 100 g^{-1} of oil) which makes it unpopular as oil (Matthaus [2004\)](#page-32-23).

Camelina based bioindustries face major challenge as its processing is relatively high cost and also due to lack of known effective techniques for the meal and oil conversion into high value biobased products (Kim et al. [2015\)](#page-31-8).

Potential for Future

There are many reasons for this crop to be grown in future as it is exceptionally rich in essential fatty acid, qualitatively high polyunsaturated FA as compared to other vegetable oil (Abramovic and Abram [2005;](#page-27-8) Angelini et al. [1997](#page-27-10); Goffman et al. [1999\)](#page-30-23), and high α -linolenic acid (n-3) that can be used as food additive.

Future research prospects for Camelina are several. There is need to develop production recommendations for Camelina as fallow replacement crop in nonirrigated cropping systems. Its nitrogen and sulfur fertility requirements are to be standardized (Obour et al. [2015](#page-33-4)). The difficulty of postharvest management, seed shattering, and harvesting should be more explored. Postemergence broadleaf weeds control and downy mildew control should be researched (Obour et al. [2015\)](#page-33-4). High yielding Camelina cultivars, high oil content desired fatty acid composition, improved cetane value and quality with regional environmental adaptation (like less sensitive toward heat stress) will be developed through plant breeding researches (Obour et al. [2015\)](#page-33-4). Recommendations for alternative uses of meal and oil to develop bio-based useful products will create ready markets for growers of Camelina (Obour et al. [2015](#page-33-4)).

References

- Abhilash P, Dubey RK, Tripathi V, Srivastava P, Verma JP, Singh H (2013) Remediation and management of POPs-contaminated soils in a warming climate: challenges and perspectives. Environ Sci Pollut Res 20(8):5879–5885
- Abramovic H, Abram V (2005) Physico-chemical properties, composition and oxidative stability of Camelina sativa oil. Food Technol Biotechnol 43(1):63–70
- Abramovič H, Butinar B, Nikolič V (2007) Changes occurring in phenolic content, tocopherol composition and oxidative stability of Camelina sativa oil during storage. Food Chem 104(3):903–909
- Allen B, Vigil M, Jabro J (2014) Camelina growing degree hour and base temperature requirements. Agron J 106(3):940–944
- Androutsopoulos VP, Tsatsakis AM, Spandidos DA (2009) Cytochrome P450 CYP1A1: wider roles in cancer progression and prevention. BMC Cancer 9(1):187
- Angelini LG, Moscheni E, Colonna G, Belloni P, Bonari E (1997) Variation in agronomic characteristics and seed oil composition of new oilseed crops in central Italy. Ind Crop Prod 6(3):313–323
- Ayres DC, Loike JD (1990) Lignans: chemical, biological and clinical properties. Cambridge University Press, Cambridge
- Aziza A, Quezada N, Cherian G (2010a) Feeding Camelina sativa meal to meat-type chickens: effect on production performance and tissue fatty acid composition. J Appl Poultry Res 19(2): 157–168
- Aziza A, Quezada N, Cherian G (2010b) Antioxidative effect of dietary Camelina meal in fresh, stored, or cooked broiler chicken meat. Poult Sci 89(12):2711–2718
- Balanuca B, Lungu A, Hanganu AM, Stan LR, Vasile E, Iovu H (2014) Hybrid nanocomposites based on POSS and networks of methacrylated Camelina oil and various PEG derivatives. Eur J Lipid Sci Technol 116(4):458–469
- Behr A, Eilting J, Irawadi K, Leschinski J, Lindner F (2008) Improved utilisation of renewable resources: new important derivatives of glycerol. Green Chem 10(1):13–30
- Berhow MA, Polat U, Glinski JA, Glensk M, Vaughn SF, Isbell T, Ayala-Diaz I, Marek L, Gardner C (2013) Optimized analysis and quantification of glucosinolates from *Camelina sativa* seeds by reverse-phase liquid chromatography. Ind Crop Prod 43:119–125
- Bermúdez-Aguirre D, Barbosa-Cánovas GV (2011) Quality of selected cheeses fortified with vegetable and animal sources of omega-3. LWT-Food Sci Technol 44(7):1577–1584
- Bernardo A, Howard-Hildige R, O'Connell A, Nichol R, Ryan J, Rice B, Roche E, Leahy J (2003) Camelina oil as a fuel for diesel transport engines. Ind Crop Prod 17(3):191–197
- Berti M, Wilckens R, Fischer S, Solis A, Johnson B (2011) Seeding date influence on Camelina seed yield, yield components, and oil content in Chile. Ind Crop Prod 34(2):1358–1365
- Betancor M, Sprague M, Usher S, Sayanova O, Campbell P, Napier JA, Tocher DR (2015) A nutritionally-enhanced oil from transgenic *Camelina sativa* effectively replaces fish oil as a source of eicosapentaenoic acid for fish. Sci Rep 5:8104
- Bhathena SJ, Velasquez MT (2002) Beneficial role of dietary phytoestrogens in obesity and diabetes. Am J Clin Nutr 76(6):1191–1201
- Blades M, Zubr J (2010) Carbohydrates, vitamins and minerals of Camelina sativa seed. Nutr Food Sci 40(5):523–531
- Botelho PB, da Rocha Mariano K, Rogero MM, de Castro IA (2013) Effect of Echium oil compared with marine oils on lipid profile and inhibition of hepatic steatosis in LDLr knockout mice. Lipids Health Dis 12(1):1
- Brandess A (2007) Modeling the profitability of *Camelina sativa* as a biofuel feedstock in eastern Colorado, Master's Thesis in Agricultural and Resource Economics, Colorado State University, Fort Collins, US
- Budin JT, Breene WM, Putnam DH (1995) Some compositional properties of Camelina (Camelina sativa L. Crantz) seeds and oils. J Am Oil Chem Soc 72(3):309–315
- Bullerwell CN, Collins SA, Lall SP, Anderson DM (2016) Growth performance, proximate and histological analysis of rainbow trout fed diets containing *Camelina sativa* seeds, meal (high-oil and solvent-extracted) and oil. Aquaculture 452:342–350
- Cais-Sokolińska D, Majcher M, Pikul J, Bielińska S, Czauderna M, Wójtowski J (2013) The effect of Camelina sativa cake diet supplementation on sensory and volatile profiles of ewe's milk. Afr J Biotechnol 10(37):7245–7252
- Cappellozza BI, Cooke R, Bohnert D, Cherian G, Carroll J (2012) Effects of Camelina meal supplementation on ruminal forage degradability, performance, and physiological responses of beef cattle. J Anim Sci 90(11):4042–4054
- Carlsson AS (2009) Plant oils as feedstock alternatives to petroleum–a short survey of potential oil crop platforms. Biochimie 91(6):665–670
- Cermak SC, Biresaw G, Isbell TA, Evangelista RL, Vaughn SF, Murray R (2013) New crop oils – properties as potential lubricants. Ind Crop Prod 44:232–239
- Chaturvedi S (2011) Influence of composted biodiesel cake on growth, yield, and micronutrient composition of tomato. Commun Soil Sci Plant Anal 42(21):2642–2653
- Chaturvedi S, Kumar A (2012) Bio-diesel waste as tailored organic fertilizer for improving yields and nutritive values of Lycopercicum esculatum (tomato) crop. J Soil Sci Plant Nutr 12(4):801–810
- Chaturvedi S, Kumar V, Satya S (2009) Composting effects of Pongamia pinnata on tomato fertilization. Arch Agron Soil Sci 55(5):535–546
- Chaturvedi S, Singh B, Nain L, Khare SK, Pandey AK, Satya S (2010) Evaluation of hydrolytic enzymes in bioaugmented compost of Jatropha cake under aerobic and partial anaerobic conditions. Ann Microbiol 60(4):685–691
- Chaturvedi S, Hemamalini R, Khare SK (2012) Effect of processing conditions on saponin content and antioxidant activity of Indian varieties of soybean (Glycine max Linn.) Cell 91:09350408194
- Chaturvedi S, Kumar A, Singh B, Nain L, Joshi M, Satya S (2013) Bioaugmented composting of Jatropha de-oiled cake and vegetable waste under aerobic and partial anaerobic conditions. J Basic Microbiol 53(4):327–335
- Chaturvedi S, Luqman S, Khare SK (2014) Facet of isoflavone, phenol and flavonoid content in soybean (Glycine max Merrill) varieties under dissimilar processing conditions. Annals of Phytomedicine 3(1):50–55
- Chedea V, Pelmus R, Smaranda T, Taranu I, Grosu H, Dragomir C (2014) Evaluation of Camelina meal as a dietary source of polyphenol for dairy cows. Bull Univ Agric Sci Vet Med Cluj-Napoca Anim Sci Biotechnol 71(2):279–280
- Chen C, Bekkerman A, Afshar RK, Neill K (2015) Intensification of dryland cropping systems for bio-feedstock production: evaluation of agronomic and economic benefits of *Camelina sativa*. Ind Crop Prod 71:114–121
- Cherian G (2012) Camelina sativa in poultry diets: opportunities and challenges. In: Biofuel co-products as livestock feed: opportunities and challenges. FAO, Rome, pp 303–310
- Cherian G, Campbell A, Parker T (2009) Egg quality and lipid composition of eggs from hens fed Camelina sativa. J Appl Poultry Res 18(2):143–150
- Ciubota-Rosie C, Ruiz JR, Ramos MJ, Pérez Á (2013) Biodiesel from Camelina sativa: a comprehensive characterisation. Fuel 105:572–577
- Ciuca N (2013) Effect of the dietary by-product Camelina meal on performances and carcass quality of TOPIGS pigs. Bull Univ Agric Sci Vet Med Cluj-Napoca Anim Sci Biotechnol 70(2):205–213
- Crowley J, Fröhlich A (1998) Factors affecting the composition and use of Camelina. Citeseer, Dublin
- Cuendet M, Oteham CP, Moon RC, Pezzuto JM (2006) Quinone reductase induction as a biomarker for cancer chemoprevention⊥. J Nat Prod 69(3):460–463
- Cui SW (2000) Polysaccharide gums from agricultural products: processing, structures and functionality. CRC Press, Technomic Publishing Company, Inc. Lancaster, USA
- Czech Z (2007) Synthesis and cross-linking of acrylic PSA systems. J Adhes Sci Technol 21(7):625–635
- Dal Bello B, Torri L, Piochi M, Zeppa G (2015) Healthy yogurt fortified with n-3 fatty acids from vegetable sources. J Dairy Sci 98(12):8375–8385
- Dalbey WE, Biles RW (2003) Respiratory toxicology of mineral oils in laboratory animals. Appl Occup Environ Hyg 18(11):921–929
- Das N, Berhow MA, Angelino D, Jeffery EH (2014) Camelina sativa defatted seed meal contains both alkyl sulfinyl glucosinolates and quercetin that synergize bioactivity. J Agric Food Chem 62(33):8385–8391
- Davis PB, Menalled FD, Peterson RK, Maxwell BD (2011) Refinement of weed risk assessments for biofuels using Camelina sativa as a model species. J Appl Ecol 48(4):989–997
- Eidhin DN, O'Beirne D (2010) Oxidative stability of Camelina oil in salad dressings, mayonnaises and during frying. Int J Food Sci Technol 45(3):444–452
- Eidhin DN, Burke J, O'Beirne D (2003) Oxidative stability of ω3-rich Camelina oil and Camelina oil-based spread compared with plant and fish oils and sunflower spread. J Food Sci 68(1):345–353
- Enjalbert J, Johnson J (2011) Guide for producing dryland Camelina in eastern Colorado. Colorado State University Extension, Fort Collins
- Erhan SZ (2005) Industrial uses of vegetable oils. AOCS Press, Champaign
- Escobar D, Clark S, Ganesan V, Repiso L, Waller J, Harte F (2011) High-pressure homogenization of raw and pasteurized milk modifies the yield, composition, and texture of queso fresco cheese. J Dairy Sci 94(3):1201–1210
- Fahey JW, Wade KL, Stephenson KK, Chou FE (2003) Separation and purification of glucosinolates from crude plant homogenates by high-speed counter-current chromatography. J Chromatogr A 996(1):85–93
- Fairley P (2011) Introduction: next generation biofuels. Nature 474(7352):S2–S5
- Fishel CV, Lakeman M, Lawrence M, Tindal C (2011) Roundtable: aviation biofuels: can biotech make a difference? Ind Biotechnol 7(3):172–179
- Flachowsky G, Schaarmann G, Jahreis G, Schone F, Richter G, Bohme H, Schneider A (1997) Influence of feeding of oilseeds and byproducts from oilseeds on vitamin E concentration of animal products. Fett-Lipid 99(2):55–60
- Frame DD, Palmer M, Peterson B (2007) Use of *Camelina sativa* in the diets of young turkeys. J Appl Poultry Res 16(3):381–386
- Franke R, Schreiber L (2007) Suberin – a biopolyester forming apoplastic plant interfaces. Curr Opin Plant Biol 10(3):252–259
- Fröhlich A, Rice B (2005) Evaluation of Camelina sativa oil as a feedstock for biodiesel production. Ind Crop Prod 21(1):25–31
- Froment M, Mastebroek D, Van Gorp K (2006) A growers manual for Calendula officinalis L. <http://www.defra.gov.uk /farm/crops/industrial/ research/reports/ Calendula%. 20Manual.pdf>
- Gandini A (2011) The irruption of polymers from renewable resources on the scene of macromolecular science and technology. Green Chem 13(5):1061–1083
- Ganesan B, Brothersen C, McMahon DJ (2014) Fortification of foods with omega-3 polyunsaturated fatty acids. Crit Rev Food Sci Nutr 54(1):98–114
- Gesch R, Cermak S (2011) Sowing date and tillage effects on fall-seeded Camelina in the northern corn belt. Agron J 103(4):980–987
- Gesch R, Isbell T, Oblath E, Allen B, Archer D, Brown J, Hatfield J, Jabro J, Kiniry J, Long D (2015) Comparison of several Brassica species in the north central US for potential jet fuel feedstock. Ind Crop Prod 75:2–7
- Gilbertson P, Johnson B, Berti M, Halvorson M (2007) Seeding date and performance of specialty oilseeds in North Dakota. Issues in new crops and new uses. ASHS Press, Alexandria, pp 105–110
- Goffman FD, Thies W, Velasco L (1999) Chemotaxonomic value of tocopherols in Brassicaceae. Phytochemistry 50(5):793–798
- Gogus U, Smith C (2010) n-3 Omega fatty acids: a review of current knowledge. Int J Food Sci Technol 45(3):417–436
- Grady K, Nleya TM (2010) Camelina production. South Dakota Cooperative Extension Service, Brookings
- Graham-Rowe D (2011) Agriculture: beyond food versus fuel. Nature 474(7352):S6–S8
- Gryglewicz S, Muszyński M, Nowicki J (2013) Enzymatic synthesis of rapeseed oil-based lubricants. Ind Crop Prod 45:25–29
- Gugel R, Falk K (2006) Agronomic and seed quality evaluation of *Camelina sativa* in western Canada. Can J Plant Sci 86:1047
- Güner FS, Yağcı Y, Erciyes AT (2006) Polymers from triglyceride oils. Prog Polym Sci 31(7):633–670
- Halkier BA, Gershenzon J (2006) Biology and biochemistry of glucosinolates. Annu Rev Plant Biol 57:303–333
- Harris WS, Miller M, Tighe AP, Davidson MH, Schaefer EJ (2008) Omega-3 fatty acids and coronary heart disease risk: clinical and mechanistic perspectives. Atherosclerosis 197(1):12–24
- Hayes J, Stepanyan V, Allen P, O'Grady M, O'Brien N, Kerry J (2009) The effect of lutein, sesamol, ellagic acid and olive leaf extract on lipid oxidation and oxymyoglobin oxidation in bovine and porcine muscle model systems. Meat Sci 83(2):201–208
- Hertog MG, Hollman PC, Venema DP (1992) Optimization of a quantitative HPLC determination of potentially anticarcinogenic flavonoids in vegetables and fruits. J Agric Food Chem 40(9): 1591–1598
- Hixson SM, Parrish CC, Anderson DM (2013) Effect of replacement of fish oil with Camelina (Camelina sativa) oil on growth, lipid class and fatty acid composition of farmed juvenile Atlantic cod (Gadus morhua). Fish Physiol Biochem 39(6):1441–1456
- Hixson SM, Parrish CC, Anderson DM (2014a) Use of Camelina oil to replace fish oil in diets for farmed salmonids and Atlantic cod. Aquaculture 431:44–52
- Hixson SM, Parrish CC, Anderson DM (2014b) Full substitution of fish oil with Camelina (Camelina sativa) oil, with partial substitution of fish meal with Camelina meal, in diets for farmed Atlantic salmon (Salmo salar) and its effect on tissue lipids and sensory quality. Food Chem 157:51–61
- Hrastar R, Petrisic MG, Ogrinc N, Košir IJ (2009) Fatty acid and stable carbon isotope characterization of Camelina sativa oil: implications for authentication. J Agric Food Chem 57(2):579–585
- Hrastar R, Petrisic MG, Ogrinc N, Kosir IJ (2011) Fatty acid and stable carbon isotope characterization of Camelina sativa oil: implications for authentication. J Agric food Chem 57:579–585
- Hu P, Wang A, Engledow A, Hollister E, Rothlisberger K, Matocha J, Zuberer D, Provin T, Hons F, Gentry T (2011) Inhibition of the germination and growth of Phymatotrichopsis omnivora (cotton root rot) by oilseed meals and isothiocyanates. Appl Soil Ecol 49:68–75
- Hulbert AJ, Turner N, Storlien L, Else P (2005) Dietary fats and membrane function: implications for metabolism and disease. Biol Rev 80(01):155–169
- Hulbert S, Guy S, Pan W, Paulitz T, Schillinger W, Sowers K, Wysocki D (2012) Camelina production in the dryland Pacific Northwest. Washington State University Extension, Pullman
- Hurtaud C, Peyraud J (2007) Effects of feeding Camelina (seeds or meal) on milk fatty acid composition and butter spreadability. J Dairy Sci 90(11):5134–5145
- IATA A (2009) Global approach to reducing aviation emissions. First Stop: Carbon Neutral Growth from 2020
- Imbrea F, Jurcoane S, Halmajan HV, Duda M, Botos L (2011) Camelina sativa: a new source of vegetable oil. Rom Biotechnol Lett 16:6263–6270
- Iskandarov U, Kim HJ, Cahoon EB (2014) Camelina: an emerging oilseed platform for advanced biofuels and bio-based materials. In: McCann M., Buckeridge M., Carpita N. (eds) Plants and BioEnergy. Advances in Plant Biology, vol 4. Springer, New York, NY
- Jeffery EH, Araya M (2009) Physiological effects of broccoli consumption. Phytochem Rev 8(1): 283–298
- Jetter R, Kunst L, Samuels AL (2008) 4 composition of plant cuticular waxes. Annu Plant Rev Biol Plant Cuticle 23:145
- Jha P, Stougaard RN (2013) Camelina (Camelina sativa) tolerance to selected Preemergence Herbicides. Weed Technol 27(4):712–717
- Jiang Y, Caldwell CD, Falk KC (2014) Camelina seed quality in response to applied nitrogen, genotype and environment. Can J Plant Sci 94(5):971–980
- Johnson D (2007) Camelina: an emerging crop for bioenergy. In: In vitro cellular & developmental biology-Animal. Springer, New York, pp S12–S13
- Jovanovic SV, Steenken S, Tosic M, Marjanovic B, Simic MG (1994) Flavonoids as antioxidants. J Am Chem Soc 116(11):4846–4851
- Kang J, Snapp AR, Lu C (2011) Identification of three genes encoding microsomal oleate desaturases (FAD2) from the oilseed crop Camelina sativa. Plant Physiol Biochem 49(2):223–229
- Karvonen HM, Aro A, Tapola NS, Salminen I, Uusitupa MI, Sarkkinen ES (2002) Effect of [alpha] linolenic acid [ndash] rich *Camelina sativa* oil on serum fatty acid composition and serum lipids in hypercholesterolemic subjects. Metabolism 51(10):1253–1260
- Kasetaite S, Ostrauskaite J, Grazuleviciene V, Svediene J, Bridziuviene D (2014) Camelina oil-and linseed oil-based polymers with bisphosphonate crosslinks. J Appl Polym Sci 131(17):8536
- Kerstiens G (1996) Signalling across the divide: a wider perspective of cuticular structure – function relationships. Trends Plant Sci 1(4):125–129
- Kim JT, Netravali AN (2012) Non-food application of Camelina meal: development of sustainable and green biodegradable paper-Camelina composite sheets and fibers. Polym Compos 33(11): 1969–1976
- Kim N, Li Y, Sun XS (2015) Epoxidation of *Camelina sativa* oil and peel adhesion properties. Ind Crop Prod 64:1–8
- Kirkhus B, Lundon AR, Haugen J-E, Vogt G, Borge GIA, Henriksen BI (2013) Effects of environmental factors on edible oil quality of organically grown *Camelina sativa*. J Agric Food Chem 61(13):3179–3185
- Knothe G (2005) Dependence of biodiesel fuel properties on the structure of fatty acid alkyl esters. Fuel Process Technol 86(10):1059–1070
- Korsrud G, Keith M, Bell J (1978) A comparison of the nutritional value of crambe and Camelina seed meals with egg and casein. Can J Anim Sci 58(3):493–499
- Koski A, Pekkarinen S, Hopia A, Wähälä K, Heinonen M (2003) Processing of rapeseed oil: effects on sinapic acid derivative content and oxidative stability. Eur Food Res Technol 217(2):110–114
- Kosma DK, Molina I, Ohlrogge JB, Pollard M (2012) Identification of an Arabidopsis fatty alcohol: caffeoyl-coenzyme A acyltransferase required for the synthesis of alkyl hydroxycinnamates in root waxes. Plant Physiol 160(1):237–248
- Krohn BJ, Fripp M (2012) A life cycle assessment of biodiesel derived from the "niche filling" energy crop Camelina in the USA. Appl Energy 92:92–98
- Kumari A, Mohsin M, Arya MC, Joshi PK, Ahmed Z (2012) Effect of spacing on Camelina sativa: A new biofuel crop in India. Bioscan 7(4):575–577
- Lane J (2008) India ministerial group approves national bio-Fuel policy, avoids key decisions on bio-diesel, Jatropha Support. BioFuel Digest
- Lenssen AW, Iversen WM, Sainju UM, Caesar-TonThat T, Blodgett SL, Allen BL, Evans RG (2012) Yield, pests, and water use of durum and selected crucifer oilseeds in two-year rotations. Agron J 104(5):1295–1304
- Li Y, Sun XS (2014) Di-hydroxylated soybean oil polyols with varied hydroxyl values and their influence on UV-curable pressure-sensitive adhesives. J Am Oil Chem Soc 91(8):1425–1432
- Li Y, Beisson F, Ohlrogge J, Pollard M (2007) Monoacylglycerols are components of root waxes and can be produced in the aerial cuticle by ectopic expression of a suberin-associated acyltransferase. Plant Physiol 144(3):1267–1277
- Li N, Wang Y, Tilley M, Bean SR, Wu X, Sun XS, Wang D (2011) Adhesive performance of sorghum protein extracted from sorghum DDGS and flour. J Polym Environ 19(3):755–765
- Li N, Qi G, Sun XS, Wang D, Bean S, Blackwell D (2014) Isolation and characterization of protein fractions isolated from Camelina meal. Trans ASABE 57(1):169–178
- Li N, Qi G, Sun XS, Xu F, Wang D (2015) Adhesion properties of Camelina protein fractions isolated with different methods. Ind Crop Prod 69:263–272
- Li N, Qi G, Sun XS, Wang D (2016) Characterization of gum isolated from Camelina seed. Ind Crop Prod 83:268–274
- Liang YC, May CY, Foon CS, Ngan MA, Hock CC, Basiron Y (2006) The effect of natural and synthetic antioxidants on the oxidative stability of palm diesel. Fuel 85(5):867–870
- Liu J, Tjellström H, McGlew K, Shaw V, Rice A, Simpson J, Kosma D, Ma W, Yang W, Strawsine M (2015) Field production, purification and analysis of high-oleic acetyl-triacylglycerols from transgenic Camelina sativa. Ind Crop Prod 65:259–268
- Lu C, Kang J (2008) Generation of transgenic plants of a potential oilseed crop Camelina sativa by agrobacterium-mediated transformation. Plant Cell Rep 27(2):273–278
- Lue B-M, Nielsen NS, Jacobsen C, Hellgren L, Guo Z, Xu X (2010) Antioxidant properties of modified rutin esters by DPPH, reducing power, iron chelation and human low density lipoprotein assays. Food Chem 123(2):221–230
- Luna FMT, Rocha BS, Rola EM, Albuquerque MC, Azevedo DC, Cavalcante CL (2011) Assessment of biodegradability and oxidation stability of mineral, vegetable and synthetic oil samples. Ind Crop Prod 33(3):579–583
- Marquard VR, Kuhlmann H (1986) Investigations of productive capacity and seed quality of linseed dodder (Camelina sativa Crtz.). Fette Seifen Anstrichm 88:245–249
- Matthäs B (1997) Antinutritive compounds in different oilseeds. Lipid/Fett 99(5):170–174
- Matthäus B (2002) Antioxidant activity of extracts obtained from residues of different oilseeds. J Agric Food Chem 50(12):3444–3452
- Matthaus B (2004) Camelina sativa-revival of an old vegetable oil? Ernahrungs-Umschau 51(1):12–15
- Matthäus B, Zubr J (2000) Variability of specific components in *Camelina sativa* oilseed cakes. Ind Crop Prod 12(1):9–18
- McNutt J (2016) Development of biolubricants from vegetable oils via chemical modification. J Ind Eng Chem 36:1
- McVay K, Lamb P (2008) Camelina production in Montana. A self-learning resource from MSU extension, US. www.msuextension.org/publications.asp
- Meadus WJ, Duff P, McDonald T, Caine WR (2014) Pigs fed Camelina meal increase hepatic gene expression of cytochrome 8b1, aldehyde dehydrogenase, and thiosulfate transferase. J Anim Sci Biotechnol 5(1):1
- Merrien A, Carre P, Quinsac A (2012) Des ressources oléagineuses variées potentiellement au service du développement de la chimie verte. Oléagineux, Corps gras, Lipides 19(1):6–9
- Morais S, Edvardsen RB, Tocher DR, Bell JG (2012) Transcriptomic analyses of intestinal gene expression of juvenile Atlantic cod (Gadus morhua) fed diets with Camelina oil as replacement for fish oil. Comp Biochem Physiol B: Biochem Mol Biol 161(3):283–293
- Moser BR (2010) Camelina (Camelina sativa L.) oil as a biofuels feedstock: golden opportunity or false hope? Lipid Technol 22(12):270–273
- Moser BR (2012) Biodiesel from alternative oilseed feedstocks: Camelina and field pennycress. Biofuels 3(2):193–209
- Moser BR, Vaughn SF (2010) Evaluation of alkyl esters from *Camelina sativa* oil as biodiesel and as blend components in ultra low-sulfur diesel fuel. Bioresour Technol 101(2):646–653
- Mulligan GA (2002) Weedy introduced mustards (Brassicaceae) of Canada. Can Field-Naturalist 116(4):623–631
- Naczk M, Shahidi F (2006) Phenolics in cereals, fruits and vegetables: occurrence, extraction and analysis. J Pharm Biomed Anal 41(5):1523–1542
- Nagendramma P, Kaul S (2012) Development of ecofriendly/biodegradable lubricants: an overview. Renew Sust Energ Rev 16(1):764–774
- Nosal H, Nowicki J, Warzała M, Nowakowska-Bogdan E, Zarębska M (2015) Synthesis and characterization of alkyd resins based on Camelina sativa oil and polyglycerol. Prog Org Coat 86:59–70
- Obied HK, Allen MS, Bedgood DR, Prenzler PD, Robards K, Stockmann R (2005) Bioactivity and analysis of biophenols recovered from olive mill waste. J Agric Food Chem 53(4):823–837
- Obour A, Sintim H, Obeng E, Zheljazkov D (2015) Oilseed Camelina (Camelina sativa L Crantz): production systems, prospects and challenges in the USA Great Plains. Adv Plants Agric Res 2:1–10
- Onyilagha J, Bala A, Hallett R, Gruber M, Soroka J, Westcott N (2003) Leaf flavonoids of the cruciferous species, Camelina sativa, Crambe spp., Thlaspi arvense and several other genera of the family Brassicaceae. Biochem Syst Ecol 31(11):1309–1322
- Oomah BD, Kenaschuk EO, Mazza G (1995) Phenolic acids in flaxseed. J Agric Food Chem 43(8):2016–2019
- Özer BH, Kirmaci HA (2010) Functional milks and dairy beverages. Int J Dairy Technol 63(1):1–15
- Pan X, Xie W, Caldwel C, Anderson D (2011) Growth performance and carcass composition of rainbow trout (Oncorhynchus mykiss) fed practical diets containing graded levels of high fat residue Camelina meal. In: Canadian Journal of animal science, vol 3. Agricultural Institute Canada, Ottawa, pp 484–484
- Parker A (2014) Camelina sativa: success of a temperate biofuel crop as intercrop in tropical conditions of Mhow, Madhya Pradesh, India. Curr Sci 107(3):359
- Pavlista A, Isbell T, Baltensperger D, Hergert G (2011) Planting date and development of springseeded irrigated canola, brown mustard and Camelina. Ind Crop Prod 33(2):451–456
- Peiretti P, Mussa P, Prola L, Meineri G (2007) Use of different levels of false flax (Camelina sativa L.) seed in diets for fattening rabbits. Livest Sci 107(2):192–198
- Peričin D, Krimer V, Trivić S, Radulović L (2009) The distribution of phenolic acids in pumpkin's hull-less seed, skin, oil cake meal, dehulled kernel and hull. Food Chem 113(2):450–456
- Peschel W, Dieckmann W, Sonnenschein M, Plescher A (2007) High antioxidant potential of pressing residues from evening primrose in comparison to other oilseed cakes and plant antioxidants. Ind Crop Prod 25(1):44–54
- Putnam D, Budin J, Field L, Breene W (1993) Camelina: a promising low-input oilseed. In: New crops. Wiley, New York, p 314
- Qi G, Sun XS (2011) Soy protein adhesive blends with synthetic latex on wood veneer. J Am Oil Chem Soc 88(2):271–281
- Qi G, Li N, Wang D, Sun XS (2012) Physicochemical properties of soy protein adhesives obtained by in situ sodium bisulfite modification during acid precipitation. J Am Oil Chem Soc 89(2):301–312
- Ramachandran S, Singh SK, Larroche C, Soccol CR, Pandey A (2007) Oil cakes and their biotechnological applications–a review. Bioresour Technol 98(10):2000–2009
- Rana D, Pachauri D (2001) Sensitivity of zero erucic acid genotypes of oleiferous brassicas to plant population and planting geometry. Indian J Agronomy 46(4):736–740
- Rani S, Joy M, Nair KP (2015) Evaluation of physiochemical and tribological properties of rice bran oil–biodegradable and potential base stoke for industrial lubricants. Ind Crop Prod 65:328–333
- Rathke G-W, Behrens T, Diepenbrock W (2006) Integrated nitrogen management strategies to improve seed yield, oil content and nitrogen efficiency of winter oilseed rape (*Brassica napus* L.): a review. Agric Ecosyst Environ 117(2):80–108
- Razeq FM, Kosma DK, Rowland O, Molina I (2014) Extracellular lipids of Camelina sativa: characterization of chloroform-extractable waxes from aerial and subterranean surfaces. Phytochemistry 106:188–196
- Reaney M, Hartley Furtan W, Loutas P (2006) A critical cost benefit analysis of oilseed biodiesel in Canada: a BIOCAP research integration program synthesis paper, Canada. www.biocap.ca
- Reddy N, Jin E, Chen L, Jiang X, Yang Y (2012) Extraction, characterization of components, and potential thermoplastic applications of Camelina meal grafted with vinyl monomers. J Agric Food Chem 60(19):4872–4879
- Rice-Evans C, Miller N, Paganga G (1997) Antioxidant properties of phenolic compounds. Trends Plant Sci 2(4):152–159
- Robinson RG (1987) Camelina: a useful research crop and a potential oilseed crop. Minnesota Agricultural Experiment Station, University of Minnesota, St. Paul
- Rode J (2002) Study of autochthon *Camelina sativa* (L.) Crantz in Slovenia. J Herbs Spices Med Plants 9(4):313–318
- Rokka T, Alén K, Valaja J, Ryhänen E-L (2002) The effect of a Camelina sativa enriched diet on the composition and sensory quality of hen eggs. Food Res Int 35(2):253–256
- Russo R, Reggiani R (2012) Antinutritive compounds in twelve *Camelina sativa* genotypes
- Ryhänen EL, Perttilä S, Tupasela T, Valaja J, Eriksson C, Larkka K (2007) Effect of Camelina sativa expeller cake on performance and meat quality of broilers. J Sci Food Agric 87(8):1489–1494
- Saarinen NM, Wärri A, Airio M, Smeds A, Mäkelä S (2007) Role of dietary lignans in the reduction of breast cancer risk. Mol Nutr Food Res 51(7):857–866
- Salminen H, Heinonen M (2008) Plant phenolics affect oxidation of tryptophan. J Agric Food Chem 56(16):7472–7481
- Salminen H, Estévez M, Kivikari R, Heinonen M (2006) Inhibition of protein and lipid oxidation by rapeseed, Camelina and soy meal in cooked pork meat patties. Eur Food Res Technol 223(4): 461–468
- Sampath A (2009) Chemical characterization of Camelina seed oil. Rutgers University-Graduate School, New Brunswick
- Samuels L, Kunst L, Jetter R (2008) Sealing plant surfaces: cuticular wax formation by epidermal cells. Plant Biol 59(1):683
- Satas D (1989) Characterization and Evaluation of Materials. Handbook of pressure sensitive adhesive technology. Van Nostrand Reinhold company incorporated, Springer Science and Business Media New York
- Schillinger WF, Wysocki DJ, Chastain TG, Guy SO, Karow RS (2012) Camelina: planting date and method effects on stand establishment and seed yield. Field Crop Res 130:138–144
- Schneider MP (2006) Plant-oil-based lubricants and hydraulic fluids. J Sci Food Agric 86(12): 1769–1780
- Séguin-Swartz G, Eynck C, Gugel R, Strelkov S, Olivier C, Li J, Klein-Gebbinck H, Borhan H, Caldwell C, Falk K (2009) Diseases of Camelina sativa (false flax). Can J Plant Pathol 31(4):375–386
- Shonnard DR, Williams L, Kalnes TN (2010) Camelina-derived jet fuel and diesel: sustainable advanced biofuels. Environ Prog Sustain Energy 29(3):382–392
- Simopoulos AP (1999) New products from the agri-food industry: the return of n-3 fatty acids into the food supply. Lipids 34(1):S297–S301
- Sioen I, De Henauw S, Van Camp J, Volatier J-L, Leblanc J-C (2009) Comparison of the nutritional–toxicological conflict related to seafood consumption in different regions worldwide. Regul Toxicol Pharmacol 55(2):219–228
- Skupinska K, Misiewicz-Krzeminska I, Stypulkowski R, Lubelska K, Kasprzycka-Guttman T (2009) Sulforaphane and its analogues inhibit CYP1A1 and CYP1A2 activity induced by benzo [a] pyrene. J Biochem Mol Toxicol 23(1):18–28
- Smeds AI, Eklund PC, Sjöholm RE, Willför SM, Nishibe S, Deyama T, Holmbom BR (2007) Quantification of a broad spectrum of lignans in cereals, oilseeds, and nuts. J Agric Food Chem 55(4):1337–1346
- Smeds AI, Eklund PC, Willför SM (2012) Content, composition, and stereochemical characterisation of lignans in berries and seeds. Food Chem 134(4):1991–1998
- Soares VL, Lachter ER, Rodrigues JdA Jr, Batista LN, Nascimento RS (2011) New applications for soybean biodiesel glycerol. Soyabean- Application and technology, Agricultural and biological sciences. INTECH Open Access Publisher, SE19 SG, London, UK
- Solis A, Vidal I, Paulino L, Johnson BL, Berti MT (2013) Camelina seed yield response to nitrogen, sulfur, and phosphorus fertilizer in South Central Chile. Ind Crop Prod 44:132–138
- Soni S, Agarwal M (2014) Lubricants from renewable energy sources–a review. Green Chem Letters Rev 7(4):359–382
- Soriano NU Jr, Narani A (2012) Evaluation of biodiesel derived from Camelina sativa oil. J Am Oil Chem Soc 89(5):917–923
- Spencer GF, Daxenbichler ME (1980) Gas chromatography-mass spectrometry of nitriles, isothiocyanates and oxazolidinethiones derived from cruciferous glucosinolates. J Sci Food Agric 31(4):359–367
- Steiner J, Griffith S, Mueller-Warrant G, Whittaker G, Banowetz G, Elliott L (2006) Conservation practices in western Oregon perennial grass seed systems. Agron J 98(1):177–186
- Szumacher-Strabel M, Cieślak A, Zmora P, Pers-Kamczyc E, Bielińska S, Stanisz M, Wójtowski J (2011) Camelina sativa cake improved unsaturated fatty acids in ewe's milk. J Sci Food Agric 91(11):2031–2037
- Taasevigen DJ (2010) Camelina composite pellet fuels feasibility for residential and commercial applications. Montana State University-Bozeman, College of Engineering
- Terpinc P, Abramovič H (2010) A kinetic approach for evaluation of the antioxidant activity of selected phenolic acids. Food Chem 121(2):366–371
- Terpinc P, Polak T, Šegatin N, Hanzlowsky A, Ulrih NP, Abramovič H (2011) Antioxidant properties of 4-vinyl derivatives of hydroxycinnamic acids. Food Chem 128(1):62–69
- Terpinc P, Polak T, Makuc D, Ulrih NP, Abramovič H (2012) The occurrence and characterisation of phenolic compounds in Camelina sativa seed, cake and oil. Food Chem 131(2):580–589
- Thiyam U, Kuhlmann A, Stöckmann H, Schwarz K (2004) Prospects of rapeseed oil by-products with respect to antioxidative potential. C R Chim 7(6):611–616
- Tripathi V, Edrisi SA, Abhilash P (2016) Towards the coupling of phytoremediation with bioenergy production. Renew Sust Energ Rev 57:1386–1389
- Trumbo P, Schlicker S, Yates AA, Poos M (2002) Dietary reference intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein and amino acids. J Am Diet Assoc 102(11): 1621–1630
- Tse MF, Jacob L (1996) Pressure sensitive adhesives based on vector sis polymers i. rheological model and adhesive design pathways. J Adhes 56(1–4):79–95
- Tuck N (2000) Waterborne and solvent based alkyds and their end user applications. Wiley, Chichester
- Urbaniak SD, Caldwell CD, Zheljazkov VD, Lada R, Luan L (2008a) The effect of cultivar and applied nitrogen on the performance of *Camelina sativa* L. in the maritime provinces of Canada. Can J Plant Sci 88:111–119
- Urbaniak SD, Caldwell CD, Zheljazkov VD, Lada R, Luan L (2008b) The effect of seeding rate, seeding date and seeder type on the performance of *Camelina sativa* L. in the maritime provinces of Canada. Can J Plant Sci 88:501–508
- Vanbeneden N, Gils F, Delvaux F, Delvaux FR (2008) Formation of 4-vinyl and 4-ethyl derivatives from hydroxycinnamic acids: occurrence of volatile phenolic flavour compounds in beer and distribution of Pad1-activity among brewing yeasts. Food Chem 107(1):221–230
- Vanharanta M, Voutilainen S, Lakka TA, van der Lee M, Adlercreutz H, Salonen JT (1999) Risk of acute coronary events according to serum concentrations of enterolactone: a prospective population-based case-control study. Lancet 354(9196):2112–2115
- Vaughn SF, Berhow MA (2005) Glucosinolate hydrolysis products from various plant sources: pH effects, isolation, and purification. Ind Crop Prod 21(2):193–202
- Vella MN, Stratton LM, Sheeshka J, Duncan AM (2013) Exploration of functional food consumption in older adults in relation to food matrices, bioactive ingredients, and health. J Nutr Gerontol Geriatr 32(2):122–144
- Vogg G, Fischer S, Leide J, Emmanuel E, Jetter R, Levy AA, Riederer M (2004) Tomato fruit cuticular waxes and their effects on transpiration barrier properties: functional characterization of a mutant deficient in a very-long-chain fatty acid β-ketoacyl-CoA synthase. J Exp Bot 55(401):1401–1410
- Vollmann J, Damboeck A, Eckl A, Schrems H, Ruckenbauer P (1996) Improvement of Camelina sativa, an underexploited oilseed. In: Progress in new crops, vol 1. ASHS Press, Alexandria, pp 357–362
- Vollmann J, Moritz T, Kargl C, Baumgartner S, Wagentristl H (2007) Agronomic evaluation of Camelina genotypes selected for seed quality characteristics. Ind Crop Prod 26(3):270–277
- von Wettstein-Knowles P (2012) Plant waxes. eLS-Wiley, Chichester. [https://doi.org/10.1002/](https://doi.org/10.1002/9780470015902:a0001919) [9780470015902:a0001919](https://doi.org/10.1002/9780470015902:a0001919)
- Walter RH (1997) Polysaccharide dispersions: chemistry and technology in food. Academic press, Elsevier, Cambridge, Massachusetts, United States
- Waraich EA, Ahmed Z, Ahmad R, Saifullah MYA, Naeem MS, Rengel Z (2013) Camelina sativa, a climate proof crop, has high nutritive value and multiple-uses: a review. Aust J Crop Sci 7(10):1551
- Welch AS, Shakya-Shrestha MAH, Lentjes NJW, Khaw K (2010) Dietary intake and status of n-3 polyunsaturated fatty acids in a population of fish eating and non-fish eating meat-eaters, vegetarians and vegans and the precursor product ratio of α linolenic acid to long chain n-3 polyunsaturated fatty acids: results from the EPIC-Norflik cohort. Am J Clin Nutr 92:1040– 1051
- Willför S, Hemming J, Reunanen M, Eckerman C, Holmbom B (2003) Lignans and lipophilic extractives in Norway spruce knots and stemwood. Holzforschung 57(1):27–36
- Winchester N, McConnachie D, Wollersheim C, Waitz IA (2013) Market cost of renewable jet fuel adoption in the United States. MIT Joint Program on the Science and Policy of Global Change, Cambridge, MA
- Wittkop B, Snowdon R, Friedt W (2009) Status and perspectives of breeding for enhanced yield and quality of oilseed crops for Europe. Euphytica 170(1–2):131–140
- Wu X, Leung DY (2011) Optimization of biodiesel production from Camelina oil using orthogonal experiment. Appl Energy 88(11):3615–3624
- Wysocki DJ, Chastain TG, Schillinger WF, Guy SO, Karow RS (2013) Camelina: seed yield response to applied nitrogen and sulfur. Field Crop Res 145:60–66
- Yang J, Caldwell C, Corscadden K, He QS, Li J (2016) Industr Crops Prod 81:162–168
- Ye CL, Anderson DM, Lall SP (2016) The effects of Camelina oil and solvent extracted Camelina meal on the growth, carcass composition and hindgut histology of Atlantic salmon (Salmo salar) parr in freshwater. Aquaculture 450:397–404
- Zaleckas E, Makarevičiene V, Sendžikiene E (2012) Possibilities of using *Camelina sativa* oil for producing biodiesel fuel. Transport 27(1):60–66
- Zanetti F, Monti A, Berti MT (2013) Challenges and opportunities for new industrial oilseed crops in EU-27: a review. Ind Crop Prod 50:580–595
- Zubr J (1997) Oil-seed crop: Camelina sativa. Ind Crop Prod 6(2):113-119
- Zubr J, Matthäus B (2002) Effects of growth conditions on fatty acids and tocopherols in *Camelina* sativa oil. Ind Crop Prod 15(2):155–162
- Zubr J (2010) Carbohydrates, vitamins and minerals of Camelina sativa seed. Nutr food sci 40:523–531