

# Chapter 32

## Sperm DNA Testing: Where Do We Go from Here?

Ahmad H. Al-Malki and Armand Zini

### 32.1 Introduction

The assessment of male fertility potential traditionally depends on the semen analysis, and the most important parameters of this analysis are sperm concentration, motility, and morphology. Unfortunately, the clinical value of these parameters in the diagnosis of male infertility remains limited [1]. While some authors recognize the importance of semen parameters in the assessment of male fertility potential [2, 3], others question the prognostic value of this test [4–6]. Moreover, with the introduction of intracytoplasmic sperm injection (ICSI), the clinical importance of the semen analysis has declined [7].

The genomic integrity of the spermatozoon is essential for the accurate transmission of genetic information and for the proper development and maturation of the embryo [8, 9]. Animal models of sperm chromatin and DNA damage have clearly shown that sperm DNA fragmentation (e.g., experimentally induced damage) is associated with reduced male fertility potential [10–13]. These experimental studies have shown that sperm DNA damage is associated with adverse reproductive outcomes after ARTs, lower pregnancy rates, chromosomal abnormalities, pregnancy loss, reduced longevity, and birth defects [14–17]. These studies have raised concerns regarding the potential adverse outcomes associated with the use of DNA-damaged sperm in the context of human assisted reproduction.

A large number of tests have been developed to measure sperm chromatin and DNA damage in human spermatozoa [18, 19]. These tests were developed with the hope that they might further our understanding of sperm nuclear architecture, accurately measure sperm chromatin and DNA damage, and be valuable tools in clinical

---

A.H. Al-Malki • A. Zini (✉)

Department of Surgery, Division of Urology, McGill University, Montreal, QC, Canada  
e-mail: [aalmalki14@hamad.qa](mailto:aalmalki14@hamad.qa); [ziniarmand@yahoo.com](mailto:ziniarmand@yahoo.com)

practice. To date, the studies show that sperm DNA tests may be good markers of male fertility potential. Prospective studies of couples with unknown fertility status have shown that sperm DNA damage is associated with a lower probability of conception (odds ratio = ~7) and a prolonged time to pregnancy [20–23]. These studies also reveal that sperm DNA test results may be better predictors of pregnancy than conventional sperm parameters in this context [23].

Several systematic reviews of studies correlating sperm DNA test results and reproductive outcomes after ARTs have shown that sperm DNA damage is associated with lower intrauterine insemination (IUI) (odds ratio = ~9) and conventional in vitro fertilization (IVF) pregnancy rates (odds ratio = ~1.6–1.9) [19, 24–27]. In contrast, systematic reviews have shown that the relationship between sperm DNA damage and intracytoplasmic sperm injection (ICSI) pregnancy rates is weak (OR = ~1.3) [19, 24–27]. Several systematic reviews have also shown that sperm DNA damage is associated with an increased risk of pregnancy loss after an established natural, IVF, or ICSI pregnancy [28, 29].

The widespread clinical application of sperm DNA tests in the evaluation of infertile men and in the management of couples enrolled in IUI and IVF treatment cycles has not been firmly established despite a large number of clinical studies (40–50 relevant studies). One of the important reasons for the poor acceptance of sperm DNA tests in the evaluation of infertile men is the marked heterogeneity of the study characteristics. Studies on sperm DNA damage and reproductive outcomes differ in their design (prospective, retrospective, case-control) and in patient (e.g., female factors) and cycle characteristics (e.g., day of embryo transfer). Moreover, it is difficult to compare studies because they use one of several sperm DNA tests (e.g., SCSA (sperm chromatin structure assay), comet assay (also known as single-cell gel electrophoresis), TUNEL (terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling) assay).

Another reason sperm DNA tests have not been widely utilized in clinical practice is the limited understanding of what the individual assays actually measure [30]. All of the assays require some preparation of the sperm nucleus (variable degree of nuclear decondensation) prior to addition of an enzyme or dye that permits detection of the target sites (e.g., sites of damaged DNA). As such, it remains unclear if a test measures real damage or damage induced by the assay conditions. Ultimately, it is believed that all sperm DNA tests provide an indirect measure of DNA damage (e.g., SCSA, TUNEL) because the assay conditions alter the chromatin state [9, 31, 32]. It is the unique property of the sperm nucleus (i.e., with a tightly packaged chromatin) that limits the accessibility of assay reagents to all areas of the genome and complicates the correct interpretation of assay results [33]. The limited and variable accessibility of reagents to potentially damaged sites in the sperm DNA and chromatin is one reason that the precise nature, location, and clinical significance of sperm DNA damage remain poorly understood.

The lack of consensus on what is considered an acceptable assay and/or assay conditions has been another reason for the limited utilization of these assays in the clinic [30, 34]. Similarly, the lack of standardized protocols for these assays is

another worry voiced by many clinicians. This has led to some concern regarding precision, reproducibility, and repeatability of the various assays. Another important weakness of these studies is the fact that multiple cutoffs or thresholds have been used, even for the same assay (e.g., 15% or 30% for DFI using the SCSA). The variability of DNA damage thresholds has led to some confusion and misinterpretation of test results [35]. Moreover, the thresholds for many of these tests have not been adequately validated (not adequately powered or not using appropriate control populations).

The biological variability of sperm DNA tests is also an important point to remember when interpreting sperm DNA test results and using these results in clinical decision-making. It has been shown that tests of sperm DNA damage exhibit a small to moderate degree of biologic variability (coefficient of variation (CV) in the range of 10–30%) such that one may need to repeat the assay to confirm the result [36–40]. Several studies have shown that sperm DNA test results can be influenced by sexual abstinence, with longer abstinence periods being associated with higher levels of sperm DNA damage [41, 42]. Finally, external factors (e.g., fever, infections, medications) can also affect sperm DNA integrity [43–45].

Given the important clinical and biological uncertainties of sperm DNA testing, additional work in this area is much needed. In the future, basic studies should be aimed at improving our understanding of the nature of sperm chromatin and DNA damage and what it is that the various sperm DNA tests truly measure. We should also establish standardized sperm DNA assay protocols that provide reproducible results across different laboratories. Future clinical studies evaluating the relationship between sperm DNA damage and reproductive outcomes should be designed as prospective, controlled trials with well-defined populations. These studies should help establish validated and clinically relevant sperm DNA damage thresholds. Ultimately, such studies will help establish the clinical value of sperm DNA tests as markers of male fertility potential.

## 32.2 Conclusions

A large number of clinical studies (over 50 relevant studies to date) have shown that sperm DNA damage is associated with reproductive outcomes. However, the widespread clinical application of sperm DNA tests in the evaluation of the infertile man has not been firmly established due to a number of limiting clinical and biological factors. The factors responsible for the limited acceptance of sperm DNA tests in the evaluation of infertile men include the marked heterogeneity of clinical studies, the incomplete understanding of sperm chromatin and DNA damage, the lack of standardized sperm DNA test protocols, and the biological variability of these assays. Future studies should be aimed at improving our understanding of the nature of sperm chromatin and DNA damage and, ultimately, help establish the clinical value of sperm DNA tests in the evaluation of infertile men.

## References

1. Guzick DS, et al. Sperm morphology, motility, and concentration in fertile and infertile men. *N Engl J Med*. 2001;345(19):1388–93.
2. Wichmann L, Isola J, Tuohimaa P. Prognostic variables in predicting pregnancy. A prospective follow up study of 907 couples with an infertility problem. *Hum Reprod*. 1994;9(6):1102–8.
3. Tomlinson MJ, Kessopoulou E, Barratt CL. The diagnostic and prognostic value of traditional semen parameters. *J Androl*. 1999;20(5):588–93.
4. Bonde JP, et al. Relation between semen quality and fertility: a population-based study of 430 first-pregnancy planners. *Lancet*. 1998;352(9135):1172–7.
5. Lefevre L, et al. Counting sperm does not add up any more: time for a new equation? *Reproduction*. 2007;133(4):675–84.
6. Chen X, et al. Predictive value of semen parameters in in vitro fertilisation pregnancy outcome. *Andrologia*. 2009;41(2):111–7.
7. Lewis SE. Is sperm evaluation useful in predicting human fertility? *Reproduction*. 2007;134(1):31–40.
8. Gatewood JM, et al. Isolation of four core histones from human sperm chromatin representing a minor subset of somatic histones. *J Biol Chem*. 1990;265(33):20662–6.
9. Gawecka JE, et al. A model for the control of DNA integrity by the sperm nuclear matrix. *Asian J Androl*. 2015;17(4):610–5.
10. Evenson DP, Darzynkiewicz Z, Melamed MR. Comparison of human and mouse sperm chromatin structure by flow cytometry. *Chromosoma*. 1980;78(2):225–38.
11. Doerksen T, Trasler JM. Developmental exposure of male germ cells to 5-azacytidine results in abnormal preimplantation development in rats. *Biol Reprod*. 1996;55(5):1155–62.
12. Cho C, et al. Haploinsufficiency of protamine-1 or -2 causes infertility in mice. *Nat Genet*. 2001;28(1):82–6.
13. Delbes G, Hales BF, Robaire B. Effects of the chemotherapy cocktail used to treat testicular cancer on sperm chromatin integrity. *J Androl*. 2007;28(2):241–9. discussion 250–1
14. Ahmadi A, Ng SC. Fertilizing ability of DNA-damaged spermatozoa. *J Exp Zool*. 1999;284(6):696–704.
15. Fernandez-Gonzalez R, et al. Long-term effects of mouse intracytoplasmic sperm injection with DNA-fragmented sperm on health and behavior of adult offspring. *Biol Reprod*. 2008;78(4):761–72.
16. Perez-Crespo M, et al. Factors from damaged sperm affect its DNA integrity and its ability to promote embryo implantation in mice. *J Androl*. 2008;29(1):47–54.
17. Perez-Crespo M, Pintado B, Gutierrez-Adan A. Scrotal heat stress effects on sperm viability, sperm DNA integrity, and the offspring sex ratio in mice. *Mol Reprod Dev*. 2008;75(1):40–7.
18. Delbes G, Hales BF, Robaire B. Toxicants and human sperm chromatin integrity. *Mol Hum Reprod*. 2010;16(1):14–22.
19. Zini A. Are sperm chromatin and DNA defects relevant in the clinic? *Syst Biol Reprod Med*. 2011;57(1–2):78–85.
20. Evenson DP, et al. Utility of the sperm chromatin structure assay as a diagnostic and prognostic tool in the human fertility clinic. *Hum Reprod*. 1999;14(4):1039–49.
21. Spano M, et al. Sperm chromatin damage impairs human fertility. The Danish First Pregnancy Planner Study Team. *Fertil Steril*. 2000;73(1):43–50.
22. Loft S, et al. Oxidative DNA damage in human sperm influences time to pregnancy. *Hum Reprod*. 2003;18(6):1265–72.
23. Giwercman A, et al. Sperm chromatin structure assay as an independent predictor of fertility in vivo: a case-control study. *Int J Androl*. 2010;33(1):e221–7.
24. Bungum M, et al. Sperm DNA integrity assessment in prediction of assisted reproduction technology outcome. *Hum Reprod*. 2007;22(1):174–9.
25. Collins JA, Barnhart KT, Schlegel PN. Do sperm DNA integrity tests predict pregnancy with in vitro fertilization? *Fertil Steril*. 2008;89(4):823–31.

26. Practice Committee of the American Society for Reproductive, M. The clinical utility of sperm DNA integrity testing: a guideline. *Fertil Steril*. 2013;99(3):673–7.
27. Simon L, et al. A systematic review and meta-analysis to determine the effect of sperm DNA damage on in vitro fertilization and intracytoplasmic sperm injection outcome. *Asian J Androl*. 2016;19:80.
28. Zini A, et al. Sperm DNA damage is associated with an increased risk of pregnancy loss after IVF and ICSI: systematic review and meta-analysis. *Hum Reprod*. 2008;23(12):2663–8.
29. Robinson L, et al. The effect of sperm DNA fragmentation on miscarriage rates: a systematic review and meta-analysis. *Hum Reprod*. 2012;27(10):2908–17.
30. Barratt CL, et al. Sperm DNA: organization, protection and vulnerability: from basic science to clinical applications—a position report. *Hum Reprod*. 2010;25(4):824–38.
31. Henkel R, et al. TUNEL assay and SCSA determine different aspects of sperm DNA damage. *Andrologia*. 2010;42(5):305–13.
32. Evenson DP. The Sperm Chromatin Structure Assay (SCSA((R))) and other sperm DNA fragmentation tests for evaluation of sperm nuclear DNA integrity as related to fertility. *Anim Reprod Sci*. 2016;169:56–75.
33. Brown R, Harper J. The clinical benefit and safety of current and future assisted reproductive technology. *Reprod BioMed Online*. 2012;25(2):108–17.
34. Zini A, Albert O, Robaire B. Assessing sperm chromatin and DNA damage: clinical importance and development of standards. *Andrology*. 2014;2(3):322–5.
35. Simon L, et al. Comparative analysis of three sperm DNA damage assays and sperm nuclear protein content in couples undergoing assisted reproduction treatment. *Hum Reprod*. 2014;29(5):904–17.
36. Evenson DP, et al. Individuality of DNA denaturation patterns in human sperm as measured by the sperm chromatin structure assay. *Reprod Toxicol*. 1991;5(2):115–25.
37. Erenpreiss J, et al. Intra-individual variation in sperm chromatin structure assay parameters in men from infertile couples: clinical implications. *Hum Reprod*. 2006;21(8):2061–4.
38. Keel BA. Within- and between-subject variation in semen parameters in infertile men and normal semen donors. *Fertil Steril*. 2006;85(1):128–34.
39. Smit M, et al. Clinical correlates of the biological variation of sperm DNA fragmentation in infertile men attending an andrology outpatient clinic. *Int J Androl*. 2007;30(1):48–55.
40. Oleszczuk K, Giwercman A, Bungum M. Intra-individual variation of the sperm chromatin structure assay DNA fragmentation index in men from infertile couples. *Hum Reprod*. 2011;26(12):3244–8.
41. Gosalvez J, et al. Shorter abstinence decreases sperm deoxyribonucleic acid fragmentation in ejaculate. *Fertil Steril*. 2011;96(5):1083–6.
42. Pons I, et al. One abstinence day decreases sperm DNA fragmentation in 90 % of selected patients. *J Assist Reprod Genet*. 2013;30(9):1211–8.
43. Evenson DP, et al. Characteristics of human sperm chromatin structure following an episode of influenza and high fever: a case study. *J Androl*. 2000;21(5):739–46.
44. Moskovtsev SI, et al. Frequency and severity of sperm DNA damage in patients with confirmed cases of male infertility of different aetiologies. *Reprod BioMed Online*. 2010;20(6):759–63.
45. Tanrikut C, et al. Adverse effect of paroxetine on sperm. *Fertil Steril*. 2010;94(3):1021–6.