

## Chapter 8

# An Ancestral Immune Surveillance System in the Amphibian *Xenopus* Connecting Certain Heat Shock Proteins with Classical and Nonclassical MHC Class I Molecules



Jacques Robert, Maureen Banach, and Eva-Stina Edholm

**Abstract** Studies in the amphibian *Xenopus*, a vertebrate species that diverged from a common ancestor with mouse and human more than 350 million years ago, provide evolutionary insights into the convergent roles of certain hsps such as gp96 and HSP70 as well as classical and nonclassical MHC class I molecules in cancer immune surveillance. Evidence that in *Xenopus* gp96 and HSP70 can elicit potent antitumor responses dependent on antigen representation by nonclassical MHC class Ib molecules and presumably involving innate T cells suggests the existence of an ancestral immune surveillance system in antigen-presenting cells such as macrophages integrating hsps with classical and nonclassical MHC molecules. The particular connection revealed in *Xenopus* between hsps and nonclassical MHC molecules presenting conserved patterns to innate T cells affords new avenues to develop therapeutic strategies against cancer.

**Keywords** Comparative immunology · Innate T cells · Tumor immunity · Evolution · Unconventional T cells

## 8.1 Introduction

Heat shock proteins (hsps) are evolutionarily ancient and highly conserved molecular chaperones constituting several multigenic families that are produced by all cell types and perform essential biological functions under normal as well as stressful physiological conditions [1]. Some of these hsps including gp96 (a member of the hsp90 family) and the cytosolic 70 kDa hsps or HSP70 (defining indistinctively

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J. Robert (✉) · M. Banach · E.-S. Edholm  
Department of Microbiology and Immunology, University of Rochester Medical Center,  
Rochester, NY, USA  
e-mail: [Jacques\\_Robert@urmc.rochester.edu](mailto:Jacques_Robert@urmc.rochester.edu)

both the inducible hsp72 and the constitutively expressed hsc73) have received a lot of attention because of their potential use in tumor immunotherapy (reviewed in [2–4]). HSP70 and gp96 have been shown to elicit potent CD8 T-cell responses specific against the antigenic peptides they chaperone not only in humans and mice [5–7] but also in frogs [8, 9]. These hsp-mediated CD8 T-cell responses are MHC class I restricted and depend on the internalization of the hsp-antigen complexes by endocytic receptors such as the  $\alpha$ 2-macroglobulin receptor CD91 at the surface of antigen-presenting cells (APCs; [10, 11]). This is followed by the representation of chaperoned antigenic peptides by MHC class Ia molecules on APCs to CD8 T cells [7, 12, 13]. The functional connection between hsp chaperoning and MHC class I antigen presentation may have even further ramifications than previously thought considering that in addition to classical MHC class Ia (class Ia) a growing number of nonclassical MHC class Ib (class Ib) and class I-like gene have been characterized (reviewed in [14, 15]). Some of these class Ib genes encode proteins that are hypothesized to be indicators of intracellular stress and malignancy (reviewed in [16, 17]). The potential role of these class Ib molecules is of particular relevance in immune surveillance and recognition of aggressive class Ia low or negative tumor cells through their interaction with T-cell receptors and/or non-T-cell inhibitory or triggering receptors expressed by NK and unconventional T cells.

Focusing on two of the most conserved hsps, gp96 and hsp70, studies in the amphibian *Xenopus* have provided compelling evidence that the immunological properties of these molecular chaperones, especially their significant antitumor responses, have been conserved during evolution (Reviewed in [18]). Comparably, while nonclassical MHC class Ib genes in *Xenopus* do not share a direct common ancestor with their mammalian counterparts, some of these genes encode molecules with striking analogous functions including class Ib-restricted unconventional T-cell-mediated antitumor immune responses.

We review here recent advances using the amphibian *Xenopus* to explore the potential of an ancestral immune surveillance system composed of hsps such as gp96 and hsp70, endocytic receptors such as CD91 and classical and nonclassical MHC class I molecules.

## 8.2 The *Xenopus* Immune System

The immune system of the South African clawed frog *Xenopus laevis* exhibits all the basic elements of jawed vertebrate immunity. The primary immune organs thymus and spleen and adaptive B- and T-cell effectors expressing a wide Ig and TCR repertoire generated by RAG-mediated somatic diversification as well as innate cell effectors such as neutrophils and macrophages are all conserved in *Xenopus* (reviewed in [19]). In fact, the fully sequenced and annotated genomes of two different *Xenopus* species, *X. tropicalis* and *X. laevis*, have provided compelling

evidence of the remarkably high degree of overall conservation of immune genes between *Xenopus* and human.

One intriguing aspect of anuran amphibians such as *Xenopus* that is not encountered in mammals is that the development of the immune system occurs at two distinct times: first during larval life and then again during the metamorphic transition from tadpole to adult [20, 21]. Specifically, the *Xenopus* thymus is first colonized by embryonic stem cells a few days after fertilization [22]. During metamorphosis, the thymus loses about 90% of its lymphocytes [23]. This loss is followed by a second wave of stem cell immigration [24, 25]. The tadpole is free-swimming and amenable to a variety of surgical (e.g., thymectomy, transplantation) and nonsurgical (e.g., adoptive transfer of leucocytes, injection of hormones, antibodies) interventions. Therefore, studies in *Xenopus* tadpoles can be helpful in collecting valuable information otherwise difficult to gather from in utero studies in mammals (e.g., development of self-tolerance to adult-specific antigens, acquisition of a second T-cell repertoire, and ontogeny of T-cell subsets in a natural setting).

A second aspect of *Xenopus* immunology that makes it attractive as a model is the absence of classical MHC class Ia protein expression in tadpoles until the onset of metamorphosis. Surface class Ia expression is first detected on erythrocytes and on splenic leukocyte populations at pro-metamorphic stages [21, 26, 27]. Although tadpoles are immunocompetent and have CD8 T cells, the larval thymus lacks significant expression of class Ia and LMP7 genes until metamorphosis, which suggests an inefficient class Ia-restricted T-cell education during larval life [21, 28]. Conversely, multiple class Ib genes are expressed by thymocytes at the onset of thymic organogenesis consistent with a role of class Ib molecules in early T-cell development.

Thus, the high degree of functional conservation of the *Xenopus* immune system with human, the natural class Ia-deficient tadpole stages, as well as the amenability of *Xenopus* to in vivo experimentation make it a highly relevant nonmammalian model (reviewed in [19, 29]). In particular, *Xenopus* is well suited to study tumor immune surveillance and as such has proven instrumental to exploring innovative approaches for cancer immunotherapy (reviewed in [19, 30]).

### 8.3 Lymphoid Tumors and Tumor Immunity in *Xenopus*

*X. laevis* is the only amphibian species in which a series of true lymphoid tumor cell lines have been derived and characterized from spontaneously occurring thymic tumors ([31, 32]. Two similar thymic tumors were also reported at the *Xenopus* colony at Tulane University around the same time [33]. More recently, another type of spontaneous leukocytic, possibly monocytic, tumor very different from the thymic tumors originally characterized was described [34].

Importantly, the occurrence of spontaneous thymic tumors in MHC-defined inbred and *X. laevis*/*X. gilli* isogenetic clones has provided a unique opportunity to derive

lymphoid tumor lines growing in in vitro culture as well as in vivo following transplantation in compatible *X. laevis* host [32, 35]. From the partially inbred F strain homozygous of the *f* MHC haplotype, two different tumor lines (B3B7 and ff-2) were derived, whereas from the isogenetic clone LG-15 heterozygous for the MHC haplotype *a/c*, 15/0 and 15/40 lines were obtained. These cell lines are all nonadherent and grow continuously at 27 °C with a generation time of 18–24 h [36]. All four tumor lines share a mixed immature T/B-cell phenotype: they all express several pan T-cell markers such as CD8 and CD5 but have also rearranged their Ig gene loci. All the tumor cell lines also express the cortical thymocyte-specific *Xenopus* cell surface marker (CTX), a marker of immature thymocytes that in the organism is only expressed by cortical thymocytes [37, 38]. Another salient feature exhibited by all these tumor lines is the expression of high level expression of several *Xenopus* non-classical MHC class Ib (*XNC*) genes, including XNC1, 4, 10, and 11 as well as  $\beta$ 2-microglobulin [39]. In contrast, only the ff-2 tumor expresses low levels of classical MHC class Ia at the cell surface, whereas 15/0, 15/40, and B3B7 cell lines are all class Ia-negative [32, 35].

Two of these lymphoid tumor cell lines have remained transplantable in compatible hosts. The ff-2 tumor is transplantable in the MHC homozygous *fff* partially inbred F strain, whereas the 15/0 can grow in the isogenetic LG-15 background. Interestingly, the ff-2 tumor line is tumorigenic when transplanted into F tadpoles but not into F adults. The rejection of ff-2 tumor in F adults is abrogated by  $\gamma$ -irradiation that preferentially depletes thymocytes and is impaired in T-cell-deficient thymectomized animals, which suggests the critical involvement of adult T cells that differentiate just after metamorphosis [35, 40]. Comparably, the 15/0 tumor cells are highly tumorigenic when transplanted into both tadpole and adult LG-15 hosts [32, 35]. In addition, the 15/0 tumor line is transplantable and tumorigenic in another isogenetic clone, LG-6, that shares the same MHC haplotypes (*a/c*) with LG-15 animals but differs at multiple minor histocompatibility (H) loci [41]. This difference in minor H-antigens has been instrumental in exploring antigen-specific antitumor immunity in *Xenopus* as delineated in the next chapter.

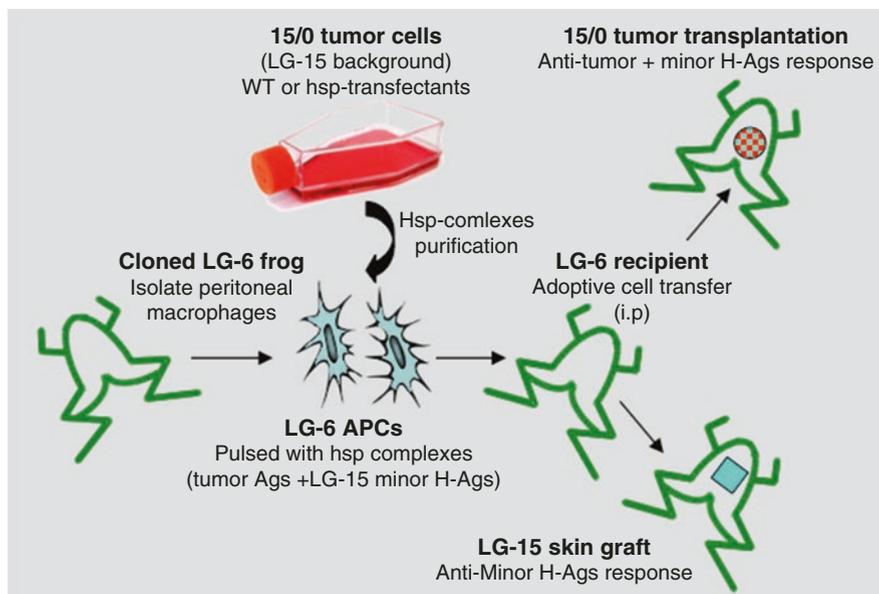
Initial in vivo and in vitro studies have revealed that in *X. laevis* as in mammals NK and CD8 T cells are critical antitumor cell effectors [41]. Briefly, the involvement of NK cells was demonstrated by anti-NK antibody treatment followed by tumor transplantation assays and by an in vitro cytotoxic assay [41–43]. Thymectomy at early developmental stage before cell precursor immigration and sublethal  $\gamma$ -irradiation that mainly affect dividing thymocytes and circulating T cell provided evidence of CD8 T cells requirement to control malignancy [35, 40, 44]. Importantly, taking advantage of the absence of class Ia expression by 15/0 tumor cells has allowed us to shed light on the unappreciated roles of nonclassical MHC class Ib molecules and unconventional class Ib-restricted T cell in *X. laevis* tumor immunity (see Chap. 5).

## 8.4 Conservation of Antitumor Properties of Heat Shock Proteins

The *X. laevis* tumor immunity model has provided evolutionary evidence of the ability of certain hsp such as the endoplasmic resident gp96 and the cytosolic HSP70 to elicit potent antitumor protective T-cell responses. In mammals, these molecules can induce pro-inflammatory cytokines, stimulate NK cells, and elicit potent cytotoxic CD8 T-cell responses against the antigenic peptides they chaperone [2–4]. The representation of antigens chaperoned by these hsp in the context of MHC class Ia by APCs critically involves the endocytic receptor CD91 [10, 11] as well as other scavenger receptors [45–47]. The additional interaction of these hsp with various signalling receptors such as TLRs is associated with their ability to stimulate inflammation [48, 49].

Given the high degree of evolutionarily conservation of gp96 and hsp70 across vertebrate and even invertebrate species, it was of interest to determine whether the immunostimulatory properties of these hsp, especially regarding antitumor immunity, were also conserved in amphibians such as *Xenopus*. Using minor H-Ags differences between LG-15 and LG-6 cloned frogs, it was first demonstrated that, as in mouse and human, both gp96 and hsp70 were able to represent chaperoned minor H-Ags and generate efficient CD8 T-cell responses recognizing and killing targets expressing the same minor H-Ags in a MHC-restricted fashion [8]. Immunization by direct subcutaneous injection of hsp70 or gp96 chaperoning minor H-Ags as well as by adoptive transfer of macrophages pulsed with hsp70/gp96-minor H-Ag complexes was shown to generate immunological memory to minor H-Ags leading to accelerated rejection of minor H-Ag-matched skin grafts [8, 50]. As in mammals, *Xenopus* gp96 and HSP70 can interact with the endocytic receptor CD91 at the surface of APCs, which leads to its rapid internalization and the representation of its bound antigens by MHC class Ia [51]. These studies in *Xenopus* strongly suggest that certain hsp (gp96, HSP70) and hsp receptors (CD91) are all integral parts of an ancestral system of immune surveillance. The importance of this system in controlling neoplasia is highlighted by its conservation for more than the 350 million years that separate amphibian and mammals from their common ancestor.

Furthermore, since, in contrast to skin grafts, the 15/0 lymphoid tumor does not express class Ia molecules, our comparative tumor immunity model has permitted investigation of the potential roles of hsp in stimulating MHC class Ia-unrestricted NK and unconventional T cells in the context of antitumor immunity. Both in vivo and in vitro studies demonstrated that immune responses against 15/0 tumor cells in *X. laevis* involve NK cells and unconventional classical class Ia-unrestricted CD8 cytotoxic T cells (CCU-CTLs) that both were shown to kill 15/0 tumor cells but not class Ia expressing non-tumoral lymphoblast targets in vitro [41]. The critical involvement of chaperoned antigens in hsp-mediated anti-15/0 tumor immune responses in the absence of class Ia presentation is supported by several lines of evidence. For both gp96 and hsp70, native forms purified from non-tumoral organs (e.g., liver) or recombinant forms



**Fig. 8.1** Schematic of the antigen representation assay developed in *Xenopus*. Peritoneal macrophages elicited by stimulation with heat-killed *E. coli* are recovered from LG-6 adults by peritoneal lavage and used as APCs. Hsps are purified from 15/0 tumor WT or stable transfectant expressing tagged recombinant *Xenopus* hsps. Since 15/0 tumor is on the LG-15 background, hsp chaperone both minor H and tumor Ags. LG-6 macrophages are pulsed for 1 h on ice with the hsp complexes at a concentration of 0.5–1 mg per  $1 \times 10^5$  cells, extensively washed, and then adoptively transferred into LG-6 recipients ( $5 \times 10^5$  cells per animal). Hsp-mediated immune responses elicited against minor H-Ags can be monitored *in vivo* by monitoring the rejection time of minor H-disparate LG-15 skin graft. Hsp-mediated antitumor immune response can be monitored by determining the time of tumor appearance following injection of 15/0 tumors

produced from bacteria or non-15/0 cells (e.g., B3B7 cells) did not elicit significant anti-15/0 tumor immune response and the removal of ligands from hsp70 by ADP abrogated anti-15/0 immunogenicity [9, 50].

To specifically address MHC class Ia-dependent and class Ia-independent antigen representation, we developed an *in vivo* adoptive cell transfer assay using *X. laevis* peritoneal macrophage (pMac) as APCs that is depicted in Fig. 8.1. First, we demonstrated that adoptive transfer of pMac exposed to either gp96- or hsp70-minor H-Ags complexes generated a CD8 T-cell response specifically against minor H-skin Ags and that this response was dependent on the endocytic receptor CD91 [51]. We then showed that a similar but class Ia-independent representation of hsp chaperoned antigens was involved in the case of the anti-15/0 tumor immune response [50]. Accordingly, LG-6 pMac exposed to tumor-derived gp96 and adoptively transferred into LG-6 hosts markedly impaired the growth of transplanted 15/0 tumor in a CD91-dependent manner.

In the case of hsp70, we went further to distinguish the respective role of the inducible hsp72 and the cognate or constitutively expressed hsc73. Although these two types of cytosolic hsp70 share very similar primary structure, they exhibit significant differences in their peptide- or ligand-binding domains, subcellular localization, and some of their function [52]. To be able to examine the tumor immunogenicity of each hsp70 isoform, we produced *X. laevis* recombinant cognate hsc73 and the inducible hsp72 from stable 15/0 tumor transfectants. Both hsp72 and hsc73-Ag complexes exhibited a similar ability for eliciting class Ia-mediated T-cell responses against minor H-Ag skin grafts. In contrast, our *in vivo* representation assay revealed that hsp72 was more potent than hsc73 in generating protective immune responses against the class Ia-negative 15/0 tumors in an Ag-dependent and putatively class Ib-mediated manner. This study provided the first evidence that although hsc73 is as potent as hsp72 in facilitating class Ia-restricted T-cell responses, it is less efficient than hsp72 in eliciting class Ia-unrestricted antitumor T-cell responses that are class Ib-mediated.

## 8.5 Conserved Roles of Nonclassical MHC and Innate T Cells in Tumor Immunity

As a method of immune evasion, tumors often downregulate their class Ia expression and thus facilitate their escape from conventional T-cell-mediated immune recognition and killing [53]. Importantly, loss of class Ia expression constitutes a loss of “self-signal” and can subsequently render malignant cells more susceptible to NK cell-mediated cytotoxicity. Consequentially, in order to avoid NK-mediated killing, many different types of tumors induce or upregulate the expression of class Ib genes [16]. Accordingly, an increased expression of certain class Ib molecules has been postulated to be an indicator of malignancy and/or intracellular stress [16]. Although the critical implication of classical MHC class Ia in tumor immune surveillance by eliciting effective antitumor CD8 cytotoxic T-cell effectors is well established from *Xenopus* to mammals, the roles of nonclassical MHC class Ib molecules and the effectors interacting with these molecules from NK to unconventional and innate T cells are less well understood.

The functional relevance of class Ib molecules in the cancer field is still unclear and often contradictory. Clinical studies have confirmed class Ib upregulated expression as a hallmark of certain tumors and shown that this typically correlates with unfavorable prognostics. HLA-E and HLA-G, in particular, have been shown to be indicators of poor clinical outcome in several different types of cancer [54–58]. On the other hand, other class Ib proteins, both in human and mouse, have been credited with the ability to mediate protective immunity against a variety of different cancers. In fact, due to their critical regulatory roles in immunity, certain class Ib molecules have emerged as attractive therapeutic targets against malignant neoplastic growths [59, 60]. Among potential class Ib targets, CD1d is perhaps the most

studied. CD1d is critical for the development and function of CD1d-restricted invariant natural killer T-cells (iNKT) cells, which despite their relatively small numbers play critical regulatory roles promoting antitumor responses [59–61]. Several ongoing clinical trials are evaluating the effect of CD1d-mediated stimulation of iNKT cells with  $\alpha$ -galactosylceramide ( $\alpha$ -GalCer) on cancer patients (reviewed in [62]). Even though no clear tumor regression was observed, the iNKT-based therapies increased INF- $\gamma$  blood levels, provided disease stabilization, and prolonged mean survival in patients no longer responding to chemo- or radiotherapies.

However, efficient clinical implementation of CD1 and iNKT cell-based therapies is still far from realization and requires a deeper and comprehensive understanding of the biology of this system.

From an evolutionary perspective, both class Ia and class Ib genes have been found in all jawed vertebrates studied to date (reviewed in [63]). Although relationships between evolutionarily distant class Ib molecules are difficult to establish, functional analogs, such as the primate HLA-E and the mouse Qa-1b, have been identified [64]. Representatives of the CD1 family of genes are found in mammals [65, 66], birds [67, 68], and reptiles [69] but in neither fish nor amphibians. In *X. laevis* there are at least 23 class Ib (*XNCs*) genes that, like other vertebrate class Ibs, are heterogeneous, less polymorphic, and less ubiquitously expressed than class Ia [39, 70–72]. Many of these *XNC* genes have an unusually high degree of conservation between *X. laevis* and *X. tropicalis* species both in primary sequence and genomic organization [70, 72]. The strong gene selection maintained in these two *Xenopus* species that diverged from a common ancestor as long ago as primates and rodents (~65 million years; [73]), is in support of important biological functions of *XNC* genes.

In this context, the high expression levels of several *XNC* genes by tumor lines derived from several independent lymphoid thymic tumors take on particular relevance. The possible involvement of certain *XNC* genes and *XNC*-restricted innate T cells in tumorigenesis and antitumor immunity in connection with hsp90s are all exciting avenues of investigation offered by the *Xenopus* model. To begin elucidating the functions of these *XNCs* in our tumor immunity model, we have chosen a loss-of-function reverse genetic approach based on RNA interference to silence *XNCs* at the level of the tumor. More specifically, the relevance of these *XNCs* for 15/0 tumorigenicity was investigated both indirectly by silencing *b2m*, which is usually required for surface expression of MHC class I molecules including class Ibs, and directly by silencing the expression of multiple *XNC* genes by targeting a consensus sequence shared by most *XNC* transcripts [74]. In fact in the case of *XNC10*, we were able to show the requirement of *b2m* surface expression. Interestingly, both types of silencing resulted in comparable results. 15/0 tumor transfectants deficient in either *b2m* or *XNCs* expression were more susceptible to NK-mediated killing but more resistant to killing by CD8 T cells in vitro. Moreover, 15/0 tumor transfectants were more tumorigenic in vivo upon transplantation in LG-15 adult recipients [74]. The faster tumor development of these *XNC*- or *b2m*-deficient tumor transfectants despite their decreased resistance to NK cell killing in vitro further suggested an

important involvement of unconventional T cells interacting with XNC molecules rather than being restricted by MHC class Ia molecules.

However, further elucidation of the role of distinctive *XNC* gene products in this tumor model has revealed this to be more complex than previously thought. *XNC10* represented an ideal candidate to focus on, since it is among the highest *XNC* expressed in 15/0 tumor and it is conserved, not only in *X. laevis* and *tropicalis* but also across ten different *Xenopus* species. Intriguingly, the specific silencing of *XNC10* in 15/0 tumor resulted in an acute rejection of these tumor transfectants by syngeneic LG-15 adults as well as naturally class Ia-deficient LG-15 tadpoles [75]. In tadpoles, the rejection was more potent toward 15/0 tumor transfectants with stronger *XNC10* knockdown. Furthermore, the rejection of *XNC10*-deficient tumors implicated cell-mediated cytotoxicity that could be enhanced by priming [75]. As such, *XNC10* is necessary for the immune evasion of the thymic-derived 15/0 tumors to escape immune recognition and class Ia-independent cytotoxicity. Taken together these findings suggest that various XNC molecules have different and possibly even opposing roles in immune surveillance, underlining the critical roles of class Ib molecules in tumor immunity. It is possible that different XNCs interact with distinct effector cells resulting in a balance between inhibitory and activating signals leading to either increased or decreased tumorigenicity.

## 8.6 Conserved Roles of Class Ib-Restricted Innate T Cell in Antitumor Immunity

Among MHC class Ib-restricted effector cells, innate T (iT) cells such as CD1d-restricted iNKT cells have recently emerged as a potentially critical component of tumor immunity as they can orchestrate both innate and adaptive immunity [76–79]. These lymphocytes are T cells with natural killer cell markers and expressing semi-invariant T-cell receptor (TCR) repertoires [14]. Although iT cells generally occur at low frequencies [80], they can control immune responses via rapid and potent release of either pro-inflammatory or anti-inflammatory cytokines [81].

Notably, we have recently demonstrated that iT cells are not only conserved in *Xenopus*, but may constitute a more prominent component of their immune system than in mammals, especially during tadpole life [82]. To date we have been able to characterize the iT cell subset restricted by *XNC10* [15, 82]. Using a reverse genetic approach combining transgenesis with RNA interference, we showed that *XNC10* is required for the development of these iT cells. Furthermore, based on TCR diversity, *XNC10* tetramer binding, and CD8 antibody staining, two subpopulations have been characterized within the *Xenopus* *XNC10*-restricted iT cells, type I *XNC10*-T<sup>+</sup>/CD8<sup>-</sup> and *XNC10*-T<sup>dim+</sup>CD8<sup>dim+</sup>, which are reminiscent of mammalian type I iNKT and type II NKT cells, respectively [82].

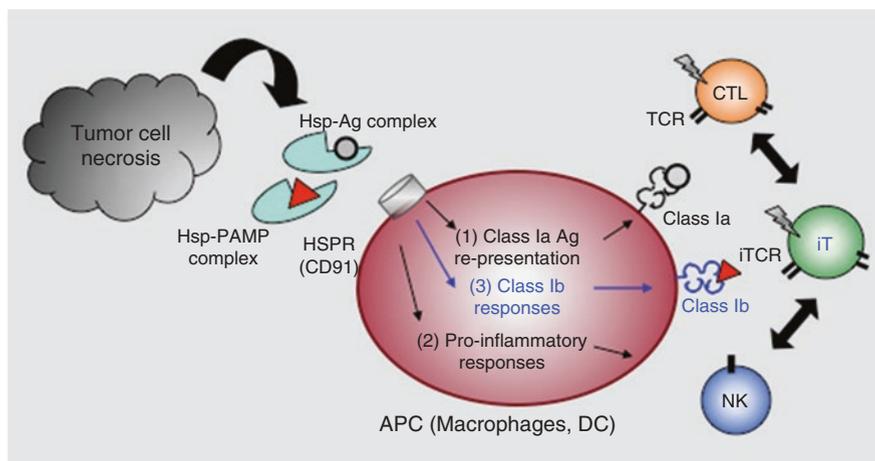
Interestingly, rapid infiltration of *XNC10*-iT cells is observed following intraperitoneal 15/0 tumor transplantation into LG-15 tadpoles [75]. Similar early

infiltration of XNC10-iT cells also occurs when transplanting ff-2 tumor into inbred F tadpoles (Banach and Robert, unpublished observations). Intriguingly, knock-down of XNC10 in 15/0 tumor triggers a substantially increased infiltration of XNC10-iT cells, which is again consistent with the use of XNC10 as an immune evasion strategy by the 15/0 tumors.

## 8.7 Conclusions and Perspective

Antigen presentation by classical MHC class Ia molecules as a way to induce potent antigen-specific CD8 T-cell responses is a pivotal component of the immune surveillance system. More specifically, in the context of tumor immune surveillance, APCs are postulated to acquire tumor antigens generated by deregulated gene expression and/or mutations from the malignant cell and then generate an adaptive T-cell response specific to these antigens. Hsps, such as cytosolic HSP70, and ER-resident gp96 can contribute to elicit this antitumor response by chaperoning tumor antigens thus facilitating efficient cross-presentation as well as by enhancing the co-stimulation responses important for potent activation of T cells.

Here, we propose that hsps, classical MHC class Ia, nonclassical MHC class Ib molecules, and their respective effector cells are integrated in an ancestral immune surveillance system (Fig. 8.2). Indeed, the critical involvement of class Ib molecules



**Fig. 8.2** Proposed ancestral immune surveillance system. Hsp-peptide complexes released in the extracellular compartment from infected or stressed cells (e.g., apoptosis, cell lysis) are internalized by APCs through receptor-mediated endocytosis (e.g., CD91). (1) Antigenic peptides channeled into the class Ia presentation pathway activate CD8 T cells. (2) Hsps internalized by the same receptors or interacting with other receptors (e.g., TLRs) stimulate pro-inflammatory responses. (3) Hsps are proposed to also stimulate class Ib-mediated responses by an as yet unknown mechanism that is likely to be Ag-specific and involve iT cell populations

in amphibian hsp-mediated antitumor responses and the finding that class Ib-restricted antitumor iT cells are present and prominent outside mammals raise the intriguing possibility that this system is ancestral and widespread across jawed vertebrates. Although the role of nonclassical MHC molecules and unconventional T cells, including iT cells in tumor immunity, is still far from fully elucidated, the inherent ability of class Ib molecules to present nonprotein antigens such as lipids and other conserved molecular motifs or patterns offers an extended avenue of detectable antitumor determinants. The limited variation of these class Ib-binding patterns and their conservation during evolution could be exploited as target of choice for future immunotherapy. In addition, the potent and rapid activation of unconventional class Ia-unrestricted T cells such as iT cells may be critical in promoting antitumor versus pro-tumor suppressive microenvironments.

In this context, the ability of hsps to also promote iT cell responses through class Ib molecules is a promising new avenue to investigate. Given that during tumor progression class Ia molecules are often downregulated, cancer immunotherapies that exploit class Ia-restricted T-cell effectors are usually insufficient to maintain potent antitumor responses. Conversely, as some class Ib molecules remain expressed on tumors or in some cases are even upregulated, these molecules and their interacting immune effector cells could serve as additional persisting immunogenic targets. Thus, the elucidation of the roles of class Ib molecules in tumor immunity is of fundamental scientific and clinical interest.

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## References

1. Hendrick JP, Hartl FU (1993) Molecular chaperone functions of heat-shock proteins. *Annu Rev Biochem* 62:349–384
2. Calderwood SK, Gong J (2016) Heat shock proteins promote cancer: it's a protection racket. *Trends Biochem Sci* 41:311
3. Srivastava P (2002a) Interaction of heat shock proteins with peptides and antigen presenting cells: chaperoning of the innate and adaptive immune responses. *Annu Rev Immunol* 20:395–425
4. Srivastava P (2002b) Roles of heat-shock proteins in innate and adaptive immunity. *Nat Rev Immunol* 2:185–194
5. Bendz H, Ruhland SC, Pandya MJ, Hainzl O, Riegelsberger S, Brauchle C, Mayer MP, Buchner J, Issels RD, Noessner E (2007) Human heat shock protein 70 enhances tumor antigen presentation through complex formation and intracellular antigen delivery without innate immune signaling. *J Biol Chem* 282:31688–31702

6. Blachere NE, Li Z, Chandawarkar RY, Suto R, Jaikaria NS, Basu S, Udono H, Srivastava PK (1997) Heat shock protein-peptide complexes, reconstituted in vitro, elicit peptide-specific cytotoxic T lymphocyte response and tumor immunity. *J Exp Med* 186:1315–1322
7. Suto R, Srivastava PK (1995) A mechanism for the specific immunogenicity of heat shock protein-chaperoned peptides. *Science* 269:1585–1588
8. Robert J, Gantress J, Rau L, Bell A, Cohen N (2002) Minor histocompatibility antigen-specific MHC-restricted CD8 T cell responses elicited by heat shock proteins. *J Immunol* 168:1697–1703
9. Robert J, Menoret A, Basu S, Cohen N, Srivastava PR (2001) Phylogenetic conservation of the molecular and immunological properties of the chaperones gp96 and hsp70. *Eur J Immunol* 31:186–195
10. Basu S, Binder RJ, Ramalingam T, Srivastava PK (2001) CD91 is a common receptor for heat shock proteins gp96, hsp90, hsp70, and calreticulin. *Immunity* 14:303–313
11. Binder RJ, Han DK, Srivastava PK (2000) CD91: a receptor for heat shock protein gp96. *Nat Immunol* 1:151–155
12. Binder RJ, Srivastava PK (2005) Peptides chaperoned by heat-shock proteins are a necessary and sufficient source of antigen in the cross-priming of CD8+ T cells. *Nat Immunol* 6:593–599
13. Lammert E, Arnold D, Nijenhuis M, Momburg F, Hammerling GJ, Brunner J, Stevanovic S, Rammensee HG, Schild H (1997) The endoplasmic reticulum-resident stress protein gp96 binds peptides translocated by TAP. *Eur J Immunol* 27:923–927
14. Adams EJ, Luoma AM (2013) The adaptable major histocompatibility complex (MHC) fold: structure and function of nonclassical and MHC class I-like molecules. *Annu Rev Immunol* 31:529–561
15. Edholm ES, Grayfer L, Robert J (2014b) Evolution of nonclassical MHC-dependent invariant T cells. *Cell Mol Life Sci* 71:4763–4780
16. Gleimer M, Parham P (2003) Stress management: MHC class I and class I-like molecules as reporters of cellular stress. *Immunity* 19:469–477
17. Gomes AQ, Correia DV, Silva-Santos B (2007) Non-classical major histocompatibility complex proteins as determinants of tumour immunosurveillance. *EMBO Rep* 8:1024–1030
18. Robert J, Goyos A, Nedelkovska H (2009) *Xenopus*, a unique comparative model to explore the role of certain heat shock proteins and non-classical MHC class Ib gene products in immune surveillance. *Immunol Res* 45:114
19. Robert J, Ohta Y (2009) Comparative and developmental study of the immune system in *Xenopus*. *Dev Dyn* 238:1249–1270
20. Flajnik MF, Du Pasquier L (1990) The major histocompatibility complex of frogs. *Immunol Rev* 113:47–63
21. Flajnik MF, Kaufman JF, Hsu E, Manes M, Parisot R, Du Pasquier L (1986) Major histocompatibility complex-encoded class I molecules are absent in immunologically competent *Xenopus* before metamorphosis. *J Immunol* 137:3891–3899
22. Kau CL, Turpen JB (1983) Dual contribution of embryonic ventral blood island and dorsal lateral plate mesoderm during ontogeny of hemopoietic cells in *Xenopus laevis*. *J Immunol* 131:2262–2266
23. Du Pasquier L, Weiss N (1973) The thymus during the ontogeny of the toad *Xenopus laevis*: growth, membrane-bound immunoglobulins and mixed lymphocyte reaction. *Eur J Immunol* 3:773–777
24. Bechtold TE, Smith PB, Turpen JB (1992) Differential stem cell contributions to thymocyte succession during development of *Xenopus laevis*. *J Immunol* 148:2975–2982
25. Turpen JB, Smith PB (1989) Precursor immigration and thymocyte succession during larval development and metamorphosis in *Xenopus*. *J Immunol* 142:41–47
26. Flajnik MF, Du Pasquier L (1988) MHC class I antigens as surface markers of adult erythrocytes during the metamorphosis of *Xenopus*. *Dev Biol* 128:198–206
27. Rollins-Smith LA, Flajnik MF, Blair PJ, Davis AT, Green WF (1997) Involvement of thyroid hormones in the expression of MHC class I antigens during ontogeny in *Xenopus*. *Dev Immunol* 5:133–144

28. Salter-Cid L, Nonaka M, Flajnik MF (1998) Expression of MHC class Ia and class Ib during ontogeny: high expression in epithelia and coregulation of class Ia and *Imp7* genes. *J Immunol* 160:2853–2861
29. Du Pasquier L, Schwager J, Flajnik MF (1989) The immune system of *Xenopus*. *Annu Rev Immunol* 7:251–275
30. Goyos A, Robert J (2009) Tumorigenesis and anti-tumor immune responses in *Xenopus*. *Front Biosci* 14:167–176
31. Du Pasquier L, Robert J (1992) In vitro growth of thymic tumor cell lines from *Xenopus*. *Dev Immunol* 2:295–307
32. Robert J, Guet C, Du Pasquier L (1994) Lymphoid tumors of *Xenopus laevis* with different capacities for growth in larvae and adults. *Dev Immunol* 3:297–307
33. Earley EM, Reinschmidt DC, Tompkins R, Gebhardt BM (1995) Tissue culture of a mixed cell thymic tumor from *Xenopus laevis*. *In Vitro Cell Dev Biol Anim* 31:255–257
34. Du Pasquier L, Wilson M, Sammut B (2009) The fate of duplicated immunity genes in the dodecaploid *Xenopus ruwenzoriensis*. *Front Biosci* 14:177–191
35. Robert J, Guet C, Du Pasquier L (1995) Ontogeny of the alloimmune response against a transplanted tumor in *Xenopus laevis*. *Differentiation* 59:135–144
36. Du Pasquier L, Courtet M, Robert J (1995) A *Xenopus* lymphoid tumor cell line with complete Ig genes rearrangements and T-cell characteristics. *Mol Immunol* 32:583–593
37. Chretien I, Robert J, Marcuz A, Garcia-Sanz JA, Courtet M, Du Pasquier L (1996) CTX, a novel molecule specifically expressed on the surface of cortical thymocytes in *Xenopus*. *Eur J Immunol* 26:780–791
38. Robert J, Cohen N (1999) In vitro differentiation of a CD4/CD8 double-positive equivalent thymocyte subset in adult *Xenopus*. *Int Immunol* 11:499–508
39. Goyos A, Ohta Y, Guselnikov S, Robert J (2009) Novel nonclassical MHC class Ib genes associated with CD8 T cell development and thymic tumors. *Mol Immunol* 46:1775–1786
40. Robert J, Guet C, Cohen N, Du Pasquier L (1997) Effects of thymectomy and tolerance induction on tumor immunity in adult *Xenopus laevis*. *Int J Cancer* 70:330–334
41. Goyos A, Cohen N, Gantress J, Robert J (2004) Anti-tumor MHC class Ia-unrestricted CD8 T cell cytotoxicity elicited by the heat shock protein gp96. *Eur J Immunol* 34:2449–2458
42. Horton TL, Minter R, Stewart R, Ritchie P, Watson MD, Horton JD (2000) *Xenopus* NK cells identified by novel monoclonal antibodies. *Eur J Immunol* 30:604–613
43. Rau L, Gantress J, Bell A, Stewart R, Horton T, Cohen N, Horton J, Robert J (2002) Identification and characterization of *Xenopus* CD8+ T cells expressing an NK cell-associated molecule. *Eur J Immunol* 32:1574–1583
44. Horton TL, Stewart R, Cohen N, Rau L, Ritchie P, Watson MD, Robert J, Horton JD (2003) Ontogeny of *Xenopus* NK cells in the absence of MHC class I antigens. *Dev Comp Immunol* 27:715–726
45. Delneste Y, Magistrelli G, Gauchat J, Haeuw J, Aubry J, Nakamura K, Kawakami-Honda N, Goetsch L, Sawamura T, Bonnefoy J, Jeannin P (2002) Involvement of LOX-1 in dendritic cell-mediated antigen cross-presentation. *Immunity* 17:353–362
46. Facciponte JG, Wang XY, Subjeck JR (2007) Hsp110 and Grp170, members of the Hsp70 superfamily, bind to scavenger receptor-A and scavenger receptor expressed by endothelial cells-I. *Eur J Immunol* 37:2268–2279
47. Murshid A, Borges TJ, Calderwood SK (2015) Emerging roles for scavenger receptor SREC-I in immunity. *Cytokine* 75:256–260
48. Asea A, Rehli M, Kabingu E, Boch JA, Bare O, Auron PE, Stevenson MA, Calderwood SK (2002) Novel signal transduction pathway utilized by extracellular HSP70: role of toll-like receptor (TLR) 2 and TLR4. *J Biol Chem* 277:15028–15034
49. Warger T, Hilf N, Rechtsteiner G, Haselmayer P, Carrick DM, Jonuleit H, von Landenberg P, Rammensee HG, Nicchitta CV, Radsak MP, Schild H (2006) Interaction of TLR2 and TLR4 ligands with the N-terminal domain of Gp96 amplifies innate and adaptive immune responses. *J Biol Chem* 281:22545–22553

50. Nedelkovska H, Robert J (2013) Hsp72 mediates stronger antigen-dependent non-classical MHC class Ib anti-tumor responses than hsc73 in *Xenopus laevis*. *Cancer Immunol* 13:4
51. Robert J, Ramanayake T, Maniero GD, Morales H, Chida AS (2008) Phylogenetic conservation of glycoprotein 96 ability to interact with CD91 and facilitate antigen cross-presentation. *J Immunol* 180:3176–3182
52. Callahan MK, Chaillot D, Jacquin C, Clark PR, Menoret A (2002) Differential acquisition of antigenic peptides by Hsp70 and Hsc70 under oxidative conditions. *J Biol Chem* 277:33604–33609
53. Zitvogel L, Tesniere A, Kroemer G (2006) Cancer despite immunosurveillance: immunoselection and immunosubversion. *Nat Rev Immunol* 6:715–727
54. Benevolo M, Mottolise M, Tremante E, Rollo F, Diodoro MG, Ercolani C, Sperduti I, Lo Monaco E, Cosimelli M, Giacomini P (2011) High expression of HLA-E in colorectal carcinoma is associated with a favorable prognosis. *J Transl Med* 9:184
55. de Kruijf EM, Sajet A, van Nes JG, Natanov R, Putter H, Smit VT, Liefers GJ, van den Elsen PJ, van de Velde CJ, Kuppen PJ (2010) HLA-E and HLA-G expression in classical HLA class I-negative tumors is of prognostic value for clinical outcome of early breast cancer patients. *J Immunol* 185:7452–7459
56. He X, Dong DD, Yie SM, Yang H, Cao M, Ye SR, Li K, Liu J, Chen J (2010) HLA-G expression in human breast cancer: implications for diagnosis and prognosis, and effect on allocytotoxic lymphocyte response after hormone treatment in vitro. *Ann Surg Oncol* 17:1459–1469
57. Ye SR, Yang H, Li K, Dong DD, Lin XM, Yie SM (2007) Human leukocyte antigen G expression: as a significant prognostic indicator for patients with colorectal cancer. *Mod Pathol* 20:375–383
58. Yie SM, Yang H, Ye SR, Li K, Dong DD, Lin XM (2007) Expression of HLA-G is associated with prognosis in esophageal squamous cell carcinoma. *Am J Clin Pathol* 128:1002–1009
59. McEwen-Smith RM, Salio M, Cerundolo V (2015) The regulatory role of invariant NKT cells in tumor immunity. *Cancer Immunol Res* 3:425–435
60. Robertson FC, Berzofsky JA, Terabe M (2014) NKT cell networks in the regulation of tumor immunity. *Front Immunol* 5:543
61. Nagato K, Motohashi S, Ishibashi F, Okita K, Yamasaki K, Moriya Y, Hoshino H, Yoshida S, Hanaoka H, Fujii S, Taniguchi M, Yoshino I, Nakayama T (2012) Accumulation of activated invariant natural killer T cells in the tumor microenvironment after alpha-galactosylceramide-pulsed antigen presenting cells. *J Clin Immunol* 32:1071–1081
62. Altman JB, Benavides AD, Das R, Bassiri H (2015) Antitumor responses of invariant natural killer T cells. *J Immunol Res* 2015:652875
63. Flajnik MF, Kasahara M (2001) Comparative genomics of the MHC: glimpses into the evolution of the adaptive immune system. *Immunity* 15:351–362
64. Yeager M, Kumar S, Hughes AL (1997) Sequence convergence in the peptide-binding region of primate and rodent MHC class Ib molecules. *Mol Biol Evol* 14:1035–1041
65. Brossay L, Chioda M, Burdin N, Koezuka Y, Casorati G, Dellabona P, Kronenberg M (1998) CD1d-mediated recognition of an alpha-galactosylceramide by natural killer T cells is highly conserved through mammalian evolution. *J Exp Med* 188:1521–1528
66. Dascher CC (2007) Evolutionary biology of CD1. *Curr Top Microbiol Immunol* 314:3–26
67. Miller MM, Wang C, Parisini E, Coletta RD, Goto RM, Lee SY, Barral DC, Townes M, Rouramir C, Ford HL, Brenner MB, Dascher CC (2005) Characterization of two avian MHC-like genes reveals an ancient origin of the CD1 family. *Proc Natl Acad Sci U S A* 102:8674–8679
68. Salomonsen J, Sorensen MR, Marston DA, Rogers SL, Collen T, van Hateren A, Smith AL, Beal RK, Skjoldt K, Kaufman J (2005) Two CD1 genes map to the chicken MHC, indicating that CD1 genes are ancient and likely to have been present in the primordial MHC. *Proc Natl Acad Sci U S A* 102:8668–8673
69. Yang Z, Wang C, Wang T, Bai J, Zhao Y, Liu X, Ma Q, Wu X, Guo Y, Zhao Y, Ren L (2015) Analysis of the reptile CD1 genes: evolutionary implications. *Immunogenetics* 67:337–346

70. Edholm ES, Goyos A, Taran J, De Jesus Andino F, Ohta Y, Robert J (2014a) Unusual evolutionary conservation and further species-specific adaptations of a large family of nonclassical MHC class Ib genes across different degrees of genome ploidy in the amphibian subfamily Xenopodinae. *Immunogenetics* 66:411–426
71. Flajnik MF, Kasahara M, Shum BP, Salter-Cid L, Taylor E, Du Pasquier L (1993) A novel type of class I gene organization in vertebrates: a large family of non-MHC-linked class I genes is expressed at the RNA level in the amphibian *Xenopus*. *EMBO J* 12:4385–4396
72. Goyos A, Sowa J, Ohta Y, Robert J (2011) Remarkable conservation of distinct nonclassical MHC class I lineages in divergent amphibian species. *J Immunol* 186:372–381
73. Evans BJ (2008) Genome evolution and speciation genetics of clawed frogs (*Xenopus* and *Silurana*). *Front Biosci* 13:4687–4706
74. Goyos A, Gusel'nikov S, Chida AS, Sniderhan LF, Maggirwar SB, Nedelkovska H, Robert J (2007) Involvement of nonclassical MHC class Ib molecules in heat shock protein-mediated anti-tumor responses. *Eur J Immunol* 37:1494–1501
75. Haynes-Gilmore N, Banach M, Edholm ES, Lord E, Robert J (2014) A critical role of non-classical MHC in tumor immune evasion in the amphibian *Xenopus* model. *Carcinogenesis* 35:1807–1813
76. Bassiri H, Das R, Guan P, Barrett DM, Brennan PJ, Banerjee PP, Wiener SJ, Orange JS, Brenner MB, Grupp SA, Nichols KE (2014) iNKT cell cytotoxic responses control T-lymphoma growth in vitro and in vivo. *Cancer Immunol Res* 2:59–69
77. Brennan PJ, Brigl M, Brenner MB (2013) Invariant natural killer T cells: an innate activation scheme linked to diverse effector functions. *Nat Rev Immunol* 13:101–117
78. Pilonis KA, Aryankalayil J, Babb JS, Demaria S (2014) Invariant natural killer T cells regulate anti-tumor immunity by controlling the population of dendritic cells in tumor and draining lymph nodes. *J Immunother Cancer* 2:37
79. Pilonis KA, Aryankalayil J, Demaria S (2012) Invariant NKT cells as novel targets for immunotherapy in solid tumors. *Clin Dev Immunol* 2012:720803
80. Kronenberg M (2005) Toward an understanding of NKT cell biology: progress and paradoxes. *Annu Rev Immunol* 23:877–900
81. Matsuda JL, Mallevaey T, Scott-Browne J, Gapin L (2008) CD1d-restricted iNKT cells, the 'Swiss-Army knife' of the immune system. *Curr Opin Immunol* 20(3):358–368. <https://doi.org/10.1016/j.coi.2008.03.018>. Epub 2008 May 22
82. Edholm ES, Albertorio Saez LM, Gill AL, Gill SR, Grayfer L, Haynes N, Myers JR, Robert J (2013) Nonclassical MHC class I-dependent invariant T cells are evolutionarily conserved and prominent from early development in amphibians. *Proc Natl Acad Sci U S A* 110:14342–14347