Chapter 20 Arbuscular Mycorrhizal Fungi: Green Approach/Technology for Sustainable Agriculture and Environment

Priyanka Srivastava, Bhawna Saxena, and Bhoopander Giri

Abstract To feed the growing population, global food production needs to be doubled by 2050. The fertilizers cost have increased several folds in the last few years, which necessitates agrarian community to be less reliable on chemicals to grow and protect their crops. Moreover, dependency on chemical fertilizers and pesticides has led to the deterioration of human health, disruption of ecosystem functioning and degradation of our environment. To overcome these problems, there is a need to explore and exploit the beneficial plant-soil microbe interactions to meet the food demand without affecting the relationship between the man and his environment. Arbuscular mycorrhizal fungi (AMF) are known to form symbiotic association with the roots of more than 90% of the terrestrial plants. They serve as biofertilizer and enhance the plant growth by accelerating nutrient uptake, particularly of inaccessible nutrients like phosphorus and nitrogen from the soil. Beside mineral nutrition, AMF also maintain the root hydraulic conductivity, increase the plant net photosynthetic capacity, improve stomatal conductance. The multifunctional extraradical hyphae of the fungus provide numerous ecological advantages like maintaining the soil health by influencing the beneficial microbes, aggregating soil particle and preventing soil erosion, conferring resistance to various stresses, enhance ecosystem productivity, bioremediation of degraded land, serving as soil carbon sink. In this chapter we attempt to discuss different role played by AMF, which make them potential tool for sustainable agriculture and environment. It is tempting to state that AMF served for 3E's i.e. eco-friendly, economic and enhanced yield.

P. Srivastava

Department of Botany, Banaras Hindu University, Varanasi 221005, Uttar Pradesh, India

B. Saxena • B. Giri (🖂)

Department of Botany, Swami Shraddhanand College, University of Delhi, Delhi 110036, India

e-mail: bhoopg@yahoo.com

[©] Springer International Publishing AG 2017

A. Varma et al. (eds.), *Mycorrhiza - Nutrient Uptake, Biocontrol, Ecorestoration*, https://doi.org/10.1007/978-3-319-68867-1_20

20.1 Introduction

The world's population is growing rapidly and has been predicted to reach around nine billion by the middle of this century (Rodriguez and Sanders 2015), which will put a tremendous pressure on the global agriculture to meet the growing demand for food. Therefore, the increase in crop production and land productivity in agriculture is necessary. It is beyond the doubt that due to the application of chemical fertilizers and agrochemicals, food production in agriculture has increased significantly. The use of these chemicals has been predicted to be increased significantly in future too (Weber 2014). However, the frequent use of chemical fertilizers along with other chemicals like pesticides and weedicides or herbicides has already generated environmental issues such as deterioration of soil quality, surface and ground water, soil biodiversity and ecosystem functioning. Any increase in the dosage of these chemicals to promote the agriculture. Therefore, in order to safeguard the environment health as well as agriculture productivity, it is importance to reduce dependence of farmers on these agrochemicals to promote plant growth and yield.

Soil is an excellent habitat for a wide array of microorganisms, which could play specific role in maintaining soil productivity, ecological processes and environment health. These microbes could play a key role in sustainable agriculture as they improve the fertility and health of the soil. They safeguard plant from enemies, enhance nutrient cycling and assist host plant to acquire immobile nutrients like N and P from the plant (Aggarwal et al. 2011; Wagg et al. 2014; Bender et al. 2016; Hunter 2016a, b) also established symbiotic associations with a wide range of plant species (Rillig et al. 2016) and thereby benefit their partner plant. The application of beneficial soil microbes could be a potential source to sort-out the issue of intensive use of costly chemical fertilizers for agricultural production as they can increase nutrient availability, plant tolerance against various kinds of stresses and therefore can provide a sustainable way for agricultural practices (Aggarwal et al. 2011). In the soil, these microorganisms are present either in the free-living (Plant Growth Promoting Rhizobacteria, like Azotobactor, Azospirillum, Pseudomonads) state or may develop mutual association with plant roots (Rhizobia, mycorrhizas and mycorrhiza-like organisms like *Piriformospora indica*) (Prasad et al. 2015).

Amongst the diverse groups of soil microorganisms, mycorrhizas are the most ubiquitous soil fungi (Schüßler et al. 2001; Smith and Read 2008; Gianinazzi et al. 2010; Leifheit et al. 2014). It has been predicted that mycorrhizal fungi may have existed even when the first plants appeared on land, which is estimated around more than 400 million years ago (Brundrett 2002). They form a mutual symbiotic relationship with the roots of plant. They are called mycorrhiza originated from the Greek 'mukés', meaning fungus, and 'rhiza' meaning roots. So far, seven different mycorrhizal fungi have been discovered from natural soils, out of these arbuscular mycorrhizal fungi (AMF) are rather common amongst the wide range of plant species. More than 90% of terrestrial plant species are colonized by AMF (Gomes et al. 2017). AMF exist in two environments; in the soil, where they form an extensive extraradical mycelium, which scavenges mineral nutrients, and within the root, where they grow between and within cortical cells developing symbiotic interfaces-the finger-like profusely branched arbuscules or intracellular coils and balloon like structures, the vesicles (Smith and Smith 2011). Arbuscules are the functional sites of nutrient transfer and vesicles act as storage organs. Extraradical hyphae extend beyond the depletion zone and support host plant for acquiring mineral nutrients and water from the soil that are not easily accessible to the normal root system. This AMF-plant relationship encourages plant growth and development of root (Kaur et al. 2014). In return, fungus avail food/carbon from the partner plant. The fungus utilizes carbon/carbohydrate for its own growth and reproduction and also in the synthesis of excretory molecules like glycoprotein (known as glomalin), which release to the soil and helps in improving soil structure with soil organic matter content (Kaur et al. 2014; Sharma et al. 2017). AMF reproduce through asexual mode of reproduction. The sexual mode of reproduction in AMF has not yet well understood. AMF are potential components of sustainable management systems. These fungi are known to exist in a wide range of environment and play a vital role in maintaining plant-water relations, nutrient uptake and ionic balance and improve soil quality and health and productivity of plants (Ruiz-Lozano et al. 2012; Nadeem et al. 2014). Under abiotic stresses, they could improve accumulation of osmo-protectants, maintain membrane integrity and osmotic adjustment and prevent oxidative damages thereby reduce adverse effects of environmental stresses and improve plant growth (Wu and Xia 2006; Evelin et al. 2009). In addition, AMF notably enhance accumulation of active ingredients in several herbal medicinal plants (Mandal et al. 2013). It is estimated that AMF could reduce up to 50% usage of chemical fertilizers for optimal agriculture production depending upon plant species and environmental conditions. Therefore, AMF could be a potential tool for sustainable agricultural practices with improved agronomic strategies which can improve fertility, health and environment of the soil, thereby the plant growth and yield (Aggarwal et al. 2011; Bender et al. 2016; Rillig et al. 2016). In this review, we attempt to highlight the role of AMF as an essential component of sustainable agriculture and environment, due to their involvement in the increased nutrient uptake, biomass production, improved photosynthesis capacity, improving quality and quantity of plant secondary metabolites, reducing dependence on fertilizers and other agro-chemical and improving plant's tolerance to environmental stresses.

20.2 Green Approaches in Relation to Agriculture and Environment

The development of agriculture sector, defined in terms of increased production with decreased average cost along with healthy and safe environment. Sustainable agriculture includes four main objectives; (i) environmental health management (including plant and soil), (ii) economic effectiveness and (iii) enhanced yield, and (iv) social and financial justice (Brodt et al. 2011). The techniques, which make products and processes more sustainable and environmental friendly may be considered as green or clean approaches. Green approaches aim to make the environment sustainable by reducing environmental degradation, conserving natural resources and reducing emissions of green house gases (CO_2 , N_2O , CH_4 etc).

20.3 Potential Solutions for Sustainable Agriculture and Environment

In the previous decades, the consumption of chemical fertilizers and pesticides has been increased massively to increase the crop production to meet the growing demands of rising population for food, which has largely affected the environment and ecosystem of the agricultural fields in the form of deterioration of soil quality and fertility, contamination of surface and ground water, air pollution and decrease in ecosystem functioning and biodiversity (Schultz et al. 1995; Socolow 1999). A potential solution to fulfill current and future generation requirements could be efficient use of the community waste and sewage sludge, which is an inexpensive alternative as manure. However, these sources are having high amount of heavy metal content, which could impose negative impact on the soil microorganisms, and heavy metals may get accumulated in the crop itself and crop field (Giller et al. 1998; Graham 2000).

Soil contains the diverse groups of microorganisms, which could play specific roles in maintaining the soil productivity and ecological processes. To overcome the environmental and ecological issues along with increased crop yield, the application of soil microorganisms could be a cheaper, affordable and eco-friendly solution (Aggarwal et al. 2011). The utilization of beneficial soil microbes for the sustainable agriculture and environment could connect sustainability with enhanced productivity, and agricultural productivity could be efficiently maintained by careful planning of conservation as well as utilization of soils (managing soil fertility/quality). Transferring/minimizing existing agropractices (use of chemical fertilizers and pesticides) with microbial applications is not only economical to grow safe and sufficient food but also for keeping the environment clean and contamination free. Further, sustainable agriculture with organic produce is paying attention of farmers, governments and development agencies. Organic farming is found in coordination with the local environment using land husbandry techniques such as soil-conservation measures, crop rotation, and the application of agronomic, biological and manual methods instead of synthetic inputs like mineral fertilizers and agro-chemicals for maximum outputs. Organic agriculture is a cost-effective approach for economically weaker farmers who cannot pay for expensive techniques of green revolution (Manimozhi and Gayathri 2012). Since arbuscular mycorrhizal fungi colonize roots of a majority of land plant species, provide them with many benefits and play a key role in ecosystem management and functioning, their application could be a potential approach for sustainable agricultural practices, and the improved agronomic strategies with AMF can improve the health of soil and environment (Schüßler et al. 2001; Aggarwal et al. 2011).

20.4 Arbuscular Mycorrhizal Fungi

Arbuscular mycorrhizal fungi (also known as Endomycorrhizae or AMF) are soil fungi, which forms symbiotic association with roots of many vascular plants (including crop plants). AMF are obligate symbionts, belonging to the phylum Glomeromycota, which comprises many genera and species (Schüßler et al. 2001). They colonize all angiosperm families except Betulaceae, Utricaceae, Commelinaceae, Cyperaceae and Polygonaceae. AMF are present as a bridge between roots and soil, where they provide the host plant with mineral nutrients and water from the soil in exchange of carbohydrates from host plant for their own growth and reproduction (Fig. 20.1) (Smith and Read 2008). In this symbiotic



Propagation cycle of arbuscular mycorrhizal fungi

Fig. 20.1 Propagation cycle of arbuscular mycorrhizal fungi. They are considered asexual because sexual reproduction is remaining to be studied. The asexual life cycle includes three main phases, asymbiotic, presymbiotic and symbiotic phases. AM fungal spores present in the soil communicate with plant root (**A**), start germinating (**B**) subsequently develop dense extraradical hyphae, which grow through soil particles and eventually enter plant roots (**C**), forming a bulbous structure called appresorium. Inside plant root, fungal hyphae grow between and within cortical cells, forming intracellular arbuscules and intraradical vesicles (**D**)

association, the fungus develops different structures in soil and in root of host plant. They develop two types of hyphal network, the extraradical hyphae (present in soil) and intraradical hyphae (present in plant root). The extraradical hyphae extract important nutrients and forms dense branched filamentous structures in the soil (Smith and Read 2008). They extend several meters far from the root and are responsible for acquisition of mineral nutrients and water for host plant along with propagation of the fungal spore. AMF fungi produce different types of soil hyphae like thick or runner hyphae and thin or absorptive hyphae (Friese and Allen 1991). The hyphal network is long-lived, and is able to colonize new plant roots as they come in contact with.

Arbuscular mycorrhizal symbiosis initiates as the fungal hyphae start responding to the presence of a plant root, establishing a molecular communication and growing along its surface. Indeed, one or a few hyphae produce appressoria (a kind of swelling like structure), these appressoria penetrate epidermal cells of the root and proceed for entering the root (Brundrett 2008). These hyphae penetrate the hypodermis, start profusely branching in the root cortex region and spread in both directions and form a network. Where, the hyphae develop highly dichotomously branched structure called arbuscules. Arbuscules are formed between the cell wall and the cell membrane of root. They start to form about 2 days after root penetration and never come into the direct contact with the nucleus or other cell organelles of root cell (Brundrett et al. 1985). The exchange in this mutual association occurs in such a way that fungus facilitates plant to uptake nutrients from soil and plant supply carbohydrate to the fungus. This exchange of nutrients takes place at the cell membrane-arbuscule interface (Balestrini and Bonfante 2014). Indeed, AMF hyphae also produce balloon-like swellings in the root cortex known as vesicles, which accumulate the storage products. These arbuscules may be interor intracellular and may function as propagules by developing thick walls in older roots (Biermann and Linderman 1983; Brundrett 2008; Balestrini and Bonfante 2014). Due to the presence of vesicles and arbuscules these fungi were referred to as arbuscular mycorrhizal or AM, however, it has been observed that a few species of AMF do not produce vesicles; therefore, it was suggested to refer as arbuscular mycorrhizal fungi. AMF reproduce asexually through thick-walled spores produced on the extraradical hyphae, and stay in the soil for long periods.

20.5 Role of Arbuscular Mycorrhizal Fungi in Sustainable Agriculture

20.5.1 Plant Growth and Productivity

AMF supply amount of nutrients and water to crop plants, helping in overcoming stresses therefore lead to boost the productivity of various crops (Lekberg and Koide 2005; Suharno et al. 2017). Karagiannidis and Hadjisavva-Zinoviadi (1998)

observed twofold increase in the biomass production of *Triticum turgitum* var. durum inoculated with *Glomus mosseae*, grow in ten different soils. Improved productivity and improved content of protein, Fe and Zn was recorded in mycorrhizal chick pea (Pellegrino and Bedini 2014). In alkaline soil, the synergistic effect of AMF and *Rhizobium leguminosarum* bv. *viciae* was observed on the growth of *Vicia faba* L. (Abd-Alla et al. 2014). Some of the AMF species, such as *Glomus geosporum* and *Glomus claroideum* accumulate heavy metals of soil into plant roots and prevent their translocation to shoot, acting as a filter (Sambandan et al. 1992; Del Val et al. 1999; Leyval et al. 2002; Meier et al. 2015). They enhance productivity and quality of plant in contaminated soils. Under saline condition, wheat varieties colonized with AMF have shown increased productivity, which may be attributed to the mycorrhiza-assisted reduced uptake of Na and Cl and improved uptake of mineral nutrients by wheat plants (Daei et al. 2009).

20.5.2 Mineral Nutrition

AMF provide host plant with mineral nutrients (Smith and Read 2008). The extraradical mycelium of AMF emerged out from the host root, extends into the soil and increases the surface area to acquire locked nutrients from the soil and alleviate nutrient deficiency by providing adequate supply of mineral nutrients to host plant (Marschner and Dell 1994; Johnson et al. 2010; Nouri et al. 2014). Phosphorus (P) and nitrogen (N) both are important determinants for the AM symbiosis and the colonization of the host plant is controlled by feedback mechanisms between both nutrients (Kytoviita 2005; Fellbaum et al. 2014).

Nitrogen is the most important nutrient for plant development and AMF assist plants to avail more than 50% of required nitrogen from soil and organic compounds by the way of N cycling and producing large amount of external hyphae (Hodge and Fitter 2010; McFarland et al. 2010; Veresoglou et al. 2012a). Studies suggest that AMF can obtain substantial amount of N from decayed organic matter/ residue or accelerate N mineralization from organic matter (Atul-Nayyar et al. 2009; Hodge and Fitter 2010) and also affect the carbon supply through soil microbial communities during decomposition (Herman et al. 2012). AMF improves symbiotic nitrogen fixation, nodulation and increase the activities of pectinase, xylo-glucanase and cellulose which are involved in the nitrogen metabolism and increase the decomposition of soil organic matter (Barea 1991). AMF may exert a priming effect on the soil bacterial communities shifting denitrifying communities by reducing the availability of soluble N and can also reduce denitrification and N₂O emission rates (Ames et al. 1984; Amora-Lazcano et al. 1998; Scheublin et al. 2010; Veresoglou et al. 2012b; Bender et al. 2014).

AMF-inducible NO_3^- or NH_4^+ transporters have been identified in several plants including tomato and soyabean (Hildebrandt et al. 2002; Kobae et al. 2010). The NH_4^+ ammonium transporter has been found to be expressed in arbuscules, suggesting control of AMF on the supply of nutrients to host (Kobae et al. 2010).

Comparing with typical uptake systems of plants, the NH4⁺ uptake system of AMF have a five times higher affinity for NH4⁺, which enables the fungus to obtain NH4⁺ from the soil even under low N supply conditions (Pérez-Tienda et al. 2012). Another important source of N in the soil is free amino acids. AMF influence the uptake of several of these amino acids such as aspartic acid, serine, glycine, glutamic acid, glutamine, cysteine or methionine (Cliquet et al. 1997; Nakano et al. 2001). Monoxenic root-organ culture experiments have shown that translocation of N in plant occurs as arginine but converts to NH4⁺ before transfer to the plant across the symbiotic interface (Smith and Smith 2011). In *Sorghum bicolor*, AMF increase the uptake of phenylalanine, methionine, asparagine, tryptophan, and cysteine, along with increased uptake of the charged amino acids like arginine, lysine, and histidine, which seems to be possible with the help of such transporters only (Cappellazzo et al. 2008; Whiteside et al. 2012).

P is an essential nutrient for plant but difficult to be acquired from the soil (Lambers et al. 2015) due to its extremely low diffusion rate (Shen et al. 2011). Mycorrhizal fungi release phosphatases, which solubilize soil organic P and increase the uptake of P by both responsive and non-responsive plants (Smith et al. 2003; Li et al. 2006). Further, phosphatases act on phytate, a major source of organic P in the soil, and release the H_2PO_4 which is utilized by the plant (Joner et al. 2000). Mycorrhizal fungi increase the exploration of P in soil, increase its translocation to plants through arbuscules, help in efficient transfer of P to plant roots and increased storage of absorbed P (Bhat and Kaveriappa 2007). AMF-plant interaction specifically induces the expression of plant Pi transporters (Harrison et al. 2002; Balestrini et al. 2007; Walder et al. 2015). In sorghum (Sorghum bicolor) and flax (Linum usitatissimum) two genes coding for inorganic phosphate transporters are identified that were induced in roots colonized by AMF but acquisition of inorganic phosphorus (Pi) was strongly affected by the combination of plant and AMF species (Walder et al. 2016). The utilization of AMF along with phosphate solubilising bacteria and other soil microbes could improve plant productivity even with low grade rock phosphates as a source of P (Bagyaraj et al. 2015). However, the effectiveness of acquisition of P varies between fungal species and even between isolates within a species (Smith and Read 2008). Jansa et al. (2005) reported that Glomus mosseae and Glomus intraradices are more efficient in P uptake than Glomus claroideum and Glomus mosseae.

Along with N and P, AMF also help in transfer of sulfur (S) to the plants with the help of sulfate transporters (Allen and Shachar-Hill 2009; Giovannetti et al. 2014). Plants take up S primarily as the sulfate anion, which is often found in low concentrations in the soil (Leustek 1996; Allen and Shachar-Hill 2009). Several researchers have reviewed the uptake, transfer and utilization of S by plants (Allen and Shachar-Hill 2009; Rennenberg et al. 2007). Sulfur is a mobile element. It is commonly lost through soil leaching. Therefore, a large proportion of the soil S remains present in the organically bound forms such as sulfate esters, synthesized by soil microorganisms, which is difficult to utilize by the plant (Fitzgerald 1976; Scherer 2001; Allen and Shachar-Hill 2009). Mycorrhizal fungi have been found to increase the percentage of S in pot-grown onion and maize plants under the

conditions of low S availability (Mohamed et al. 2014; Guo et al. 2007). However, very little is known about the mechanism of S assimilation and its regulation in mycorrhizal fungi. Allen and Shachar-Hill (2009) conducted research experiments to determine the role of AMF in acquisition of S, its metabolism and transfer to host roots and to identify effectors of S transfer to host roots. They found that sulfate was taken up by the AMF and transferred to AMF-colonized roots, increasing root S contents under moderate concentration of sulfate.

20.5.3 Water Uptake/Root Hydraulic Conductivity

Root hydraulic conductivity and permeability are the reliable indicators for water uptake activity in plants (Aubrecht et al. 2006). Mycorrhizal mycelium network affects moisture retention properties of soils and improves water uptake capacity of plant by regulating stomatal conductance (Augé et al. 2015). AMF inoculated plants have shown altered root morphology and hydraulic conductivity, demonstrating that increased root hydraulic conductivity is favorable for plant growth as it credits to improved water uptake by host plant (Augé 2001; Cseresnyés et al. 2013). Cseresnyés et al. (2014) observed that the mycorrhizal bean and cucumber plants exhibit higher rate of daily transpiration and root electrical capacitance than that of the non-mycorrhizal plants. Ruiz-Lozano (2003) revealed that to mitigate the water deficiency, AMF help to improve water uptake by enhancing the absorptive surface area of root (Aggarwal et al. 2011) which ultimately increase the soil water movement into root of plants and delay the decline in water potential (Porcel and Ruiz-Lozano 2004).

20.5.4 Role of AMF in Managing Rate of Photosynthesis

Mycorrhizal symbiosis increases photosynthetic capacity of plants by increasing the activity of hormones like cytokinin, gibberellins and of auxins which could elevate the rate of photosynthesis by influencing stomatal opening (Allen et al. 1982). Increasing colonization of *Boswellia papyrifera* seedlings resulted in greater stomatal conductance (Birhane et al. 2012). The inoculation of *Robinia pseudoacacia* L. (Black Locust) with AMF, *Funneliformis mosseae* and *Rhizophagus intraradices* grow in a lead toxic soil has shown increased photosynthetic pigment content in leaves and higher gas exchange capacity, non-photochemistry efficiency, and photochemistry efficiency (Yang et al. 2015). AMF greatly influence the photosynthesis of black locust seedlings with significantly greater leaf area, higher carboxylation efficiency, chlorophyll content and net photosynthetic rate. They significantly increase the photochemical efficiency of PS II and enhance the carbon content and calorific value of the seedlings (Zhu et al. 2014). Similarly, *Citrus tangerine* (tangerine) colonized by *Glomus versiforme* exhibited higher photosynthetic rates, stomatal conductance, transpiration rate and improve osmotic adjustments, substantiating influence of AMF on plant photosynthesis (Wu and Xia 2006).

20.5.5 Accumulation of Secondary Metabolites/Active Ingredients

Plant metabolites are divided into two broad categories, primary and secondary. The primary metabolites are vital for the basic processes of life including photosynthesis and respiration whereas secondary metabolites are important defense and specialized metabolites noteworthy for the plant interaction with its environment. Secondary metabolites play significant role in defense, and protection of plants against biotic and abiotic stress. AMF studies performed on herbal and aromatic medicinal plants showed that mycorrhizal symbiosis influences plant secondary metabolism in both above and belowground parts of the plants, increasing accumulation of active ingredients (Table 20.1). Farmer et al. (2007) observed accumulation of β -carotene in the tuber of *Ipomoea batatas*. Similarly, Sailo and Bagyaraj (2005) found increasing level of diterpene forskol in the roots of *Coleus forskohlii*. On mycorrhization of Glycyrrhiza uralensis plants, Liu et al. (2007) recorded higher concentration of glycyrrhizin. Mandal et al. (2013) carried out research experiments to examine the influence of AMF, Rhizophagus fasciculatus on the yield of secondary metabolites in Stevia rebaudiana, in relation to mycorrhizainduced physiological changes in addition to improved P uptake. AMF inoculation of Stevia with P-supplementation produced higher concentrations of steviol glycosides in comparison to control plants. The higher content of stevioside and rebaudioside-A (steviol glycosides) in AMF inoculated plants could be attributed to increased nutrients uptake and biomass production, and carbohydrates and jasmonic acid, contributing to more biosynthesis of steviol glycosides. Enhanced nutrient uptake and sugar concentration due to increased photosynthesis in AMF inoculated stevia could control up-regulation of the transcription of steviol glycosides biosynthesis genes (Mandal et al. 2015). Mycorrhizal symbiosis also facilitates the accumulation of essential oils in plants like Anethum graveolens, Trachyspermum ammi and Foeniculum vulgare (Kapoor et al. 2002, 2004; Mandal et al. 2013). Inoculation of Glycyrrhiza uralensis with Glomus mosseae has been found to improve accumulation of flavonoids, liquiritin, isoliquiritin, isoliquiritigenin and glycyrrhizic acid (Chen et al. 2017) under nutrient stress condition. Marjoram and lemon balm inoculation with AMF considerably increased the yield of rosmarinic acid and lithospermic acid isomers (Engel et al. 2016). Similarly, in Lettuce they enhanced the accumulation of vitamins, nutraceuticals and minerals (Baslam et al. 2013). In *Dioscorea* spp., the inoculation of *Glomus* etunicatum increased content of polyphenols, flavonoids and anthocyanin (Lu et al. 2015). These observations substantiate the fact that the quantity and quality of

Active	AME anapias			
produced	involved	Host plant	Family	Reference
Glycyrrhizin	Glomus mosseae	Glycyrrhiza uralensis	Fabaceae	Liu et al. (2007)
Hypericin and pseudohypericin	Rhizophagus intraradices	Hypericum perforatum	Hypericaceae	Zubek et al. (2012)
Polyphenols, flavonoids, and anthocyanin	G. etunicatum	Dioscorea sp	Dioscoreaceae	Lu et al. (2015)
Camptothecin	G. diaphanum, Acaulospora mellea and Sclerocystis sinuosa	Camptotheca acuminata	Nyssaceae	Zhao et al. (2007)
Flavonoids, lig- nin, DPPH activity and phenolic compounds	Funneliformis mosseae	<i>Cucumis sativus</i> L. seedlings	Cucurbitaceae	Chen et al. (2013)
Phenol	Glomus aggregatum	Catharanthus roseus	Apocynaceae	Srinivasan and Govindasamy (2014)
Chicoric and Caffeic acid derivative	Rhizophagus intraradices	Ocimum basilicum	Lamiaceae	Scagel and Lee (2012)
Stevioside and rebaudioside	Rhizophagus fasciculatus	Stevia rebaudiana	Asteraceae	Mandal et al. (2013)
Linalool and Methyl chavicol	Glomus sp.	Ocimum basilicum	Lamiaceae	Zolfaghari et al. (2012)
2-hydroxy-4- methoxy benzaldehyde	Glomus sp.	Decalepis hamiltonii	Asclepidaceae	Matam and Parvatam (2017)
Cinnamic acid, anthocyanin, Total phenol	Glomus etunicatum and Glomus mosseae	Mentha spicata	Lamiaceae	Bagheri et al. (2014)
Total phenol	Glomus intraradices	Ocimum gratissimum L	Lamiaceae	Hazzoumi et al. (2015)
Phytoalexins	Glomus fasciculatum	Vigna unguiculata	Leguminaceae	Sundaresan et al. (1993)
Rishitin and solavetivone	Glomus etunicatum	Solanum tuberosum	Solanaceae	Yao et al. (2003)
Trigonelline	Gigaspora rosea	Prosopis laevigata	Leguminaceae	Rojas- Andrade et al. (2003)
Cyanidin-3- glucoside	Glomus intraradices	Fragaria x ananassa Duch.	Rosaceae	Castellanos- Morales et al. (2010)

 Table 20.1
 Influence of AMF on accumulation of active ingredients in plants

(continued)

Active ingredients produced	AMF species involved	Host plant	Family	Reference
Linalool, linalyl acetate	Acaulospora morrowiae, Rhizophagus clarus, Scutellospora calospora	Mentha × piperita L. var. citrata (Ehrh.) Briq.	Lamiaceae	Silva et al. (2014)
Artemisinin, Jasmonic acid	Rhizophagus intraradices	Artemisia annua	Asteraceae	Mandal et al. (2015)
Flavonoids, Carotenoids	Glomus intraradices	Bituminaria bituminosa	Fabaceae	Pistelli et al. (2015)
p- hydroxybenzoic acid, Rutin	Rhizophagus irregularis	Viola tricolor L.	Violaceae	Zubek et al. (2015)
Ferulic acid, Caffeic acid, Kaempferol and Luteolin, Quercetin	Piriformospora indica	Brassica campestris ssp. chinensis L.	Brassicaceae	Khalid et al. (2017)
Tanshinones, Salvianolic acid B	Glomus mosseae, Glomus aggregatum, Glo- mus versiforme, Glomus intraradices	Salvia miltiorrhiza	Lamiaceae	Yang et al. (2017)

Table 20.1 (continued)

active ingredients of plants could be improved by the application of optimized inoculum of arbuscular mycorrhizal fungi.

20.5.6 Abiotic Environmental Stress Tolerance

Salinity stress is one of the serious threats, limiting the plant growth and productivity by decreasing water uptake, adversely affecting nutrient absorbance, hydraulic conductivity, stomatal conductance and net photosynthetic rate (Al-Karaki et al. 2001; Koca et al. 2007). Mycorrhizal associations have been found to influence the physiological and morphological properties of plant, thereby help the plant to combat biotic and abiotic stresses (Miransari et al. 2008; Evelin et al. 2009; Saxena et al. 2017; Giri and Saxena 2017). Under environmental stresses like drought and salinity they play an important role in maintaining ionic balance and nutrient supply in plant and soil for proper functioning and productivity of crop plants (Porcel et al. 2012; Evelin et al. 2013; Augé et al. 2015). Availability of water is the major limitation in agricultural fields particularly in arid and semi-arid areas. Several studies have shown that presence of AMF in water-stressed plants increase leaf and root growth and positively maintain root to shoot ratio (Quilambo 2000). The extraradical hyphae increase the absorptive surface area of roots for the better uptake of mineral nutrients and water under stress conditions (Hampp et al. 2000). Moreover, AMF aid plant to overcoming the adverse effects of environmental stresses as they improve balance between K:Na and Ca:Na (Elhindi et al. 2017). AMF facilitate plant for the accumulation of osmo-protectants, osmotic adjustment and maintenance of membrane integrity, therefore preventing plant from oxidative damage during environmental stresses (Evelin et al. 2013). Molecular studies have shown that in AMF colonized plants like *Glycine max* and *Lactuca sativa* changes gene expression of gene encoding plasma membrane aquaporins in response to drought stress (Porcel et al. 2005). Similarly, AM symbiosis could regulate expression of genes involved in oxidative stress, proline synthesis and in dehydrin proteins (Porcel et al. 2005; Fan and Liu 2011). Several nutritional and non-nutritional mycorrhiza-mediated mechanisms, helping plants to overcome salinity stress may be programmed as (1) nutrient uptake (like P, N, Mg and Ca) (Evelin et al. 2009), (2) Biochemical changes like accumulation of proline, betaines, polyamines, carbohydrates and enzymatic and non-enzymatic antioxidants (Evelin et al. 2009), (3) Physiological changes like enhanced photosynthetic efficiency, gas exchange and water use efficiency (Ruiz-Lozano et al. 2012; Elhindi et al. 2017; Saxena et al. 2017).

20.5.7 Weed Control

In agricultural fields, weeds are a major hazard to the crops, as they compete with crops and negatively affect the crop productivity (Oerke 2006; Ampong-Nyarko and Datta 1991). Although the management of weeds is possible using herbicides, weedicides and toxic chemicals, but the drawbacks of using these techniques are (Heap 2015; Shakeel and Yaseen 2016); (i) they are expensive, (ii) not suitable for sensitive crops (iii) several weeds are resistant against the herbicides, (iv) not safe for ecosystem and environment. Arbuscular mycorrhizal fungi have been found effective against several weeds, hence could be considered as the potent microorganisms to control agricultural weeds (Rinaudo et al. 2010; Veiga et al. 2011). Lendzemo (2004) revealed that *Striga hermonthica* (witch weed), which seriously affects the cereal crops can be controlled by applying AMF inoculants, as they directly reduce growth of weed or suppress their growth by increasing the competitiveness of crop through increased nutrient uptake of crop for essential nutrients leaving weeds deprived of nutrition. Rinaudo et al. (2010) recorded about 47% reduction in biomass due to invasion of six weeds, which were grown together with a crop, which could be controlled using AMF (Veiga et al. 2011; Adeyemi et al. 2015). Another mycorrhizal approach in weed control can be inhibiting weed growth by changing the relative abundance of mycorrhiza-infected and nonmycorrhizalinfected weed species in agroecosystem (Jordan et al. 2000) most likely due to secretion of some substances from AMF extraradical mycelium (Veiga et al. 2012).

20.5.8 Disease Resistance

Through several mechanisms like damage compensation, direct competition for colonization sites or food, changes in root morphology and rhizosphere microbial community composition, biochemical changes associated with plant defense mechanisms and the activation of plant defense. AMF can induce control over various plant diseases (Huang et al. 2003; Whipps 2004). Shalaby and Hanna (1998) observed improved growth of AMF colonized Soyabean plant in a soil infested with pathogenic fungi *Fusarium solani* and other pathogens. Similarly, AMF colonization increased resistance in Poinsettia against pathogens like *Pythium ultimumon* (Kaye et al. 1984). Coffee plant infested with mycorrhizal fungi enhanced competitiveness against disease causing *Bidens pilosa* (França et al. 2016) whereas Tea plant inoculated with mycorrhizal fungi and other plant growth promoters provided resistance for rot disease (Chakraborty et al. 2016). *Fusarium* induces wilt disease in tomato worldwide, at large its control and management is difficult. Inoculation of tomato with AMF has shown control in the severity of the disease and enhanced overall growth in tomato seedlings (Mwangi et al. 2011).

20.6 Role in Ecosystem and Environment Management

20.6.1 Soil Health and Fertility

One of the most important functions of the mycorrhizal symbiosis is to enhance acquisition of nutrients from the soil with the help of fungal hyphae. The extraradical hyphae of AM fungi often acquire those nutrients, which are hard to be extracted by plant roots. Although phosphorus is an essential nutrient for plants but remain present in the immobile form in the soil. It can be solubilized with the enzymes released by AMF (Joner and Johansen 2000). Ecosystems instability severely influences the physical, chemical and biological properties of the soil. However, mycorrhizal fungi have been found to improve such properties/processes (Augé 2004). They not only indeed affect the individual plant, even actively engaged in the processes like soil aggregation at the ecosystem level and improve the soil health and quality (Rillig 2004a, b). Nonetheless, soil aggregation helps in improving soil porosity and gaseous exchange, and nicely maintains soil nutrient cycle, protects soil carbon in aggregates, and influences beneficial soil micro-fauna (Diaz-Zorita et al. 2002; Jastrow et al. 1998; Johansson et al. 2004). By acting as a biofertilizer, and biocontrol agent mycorrhiza benefits the overall soil health and productivity (Jeffries and Barea 2012; Abdel Latef et al. 2016). Although plants absorb both macro-and micronutrients from the soil to accomplish their metabolic activities, the excessive accumulation of heavy metals like Zn, Cu, Pb, As, Cd, Cu etc in soil due to various anthropogenic activities and deposition of agro-chemicals rigorously degrade the soil and its fertility and productivity (Liu et al. 1997). Several researchers have observed that AMF could play important role in mitigating such effects by minimizing the negative impact of heavy metal contamination attributed to their enhanced adsorption and precipitation (Nadeem et al. 2014; Turnau et al. 2008; Malekzadeh et al. 2011).

20.6.2 Ecosystem Functioning and Biodiversity

The relationships between mycorrhizal fungi and plant is of great importance as they could play a major role in the ecosystem functioning and maintaining soil biodiversity (Gerz et al. 2016), contributing to biogeochemical cycles, food and timber production, and other benefits (Gianinazzi et al. 2010; van der Heijden et al. 2015; Bender et al. 2016). In fact, AMF are involved in many biogeochemical cycles, the most studied are C, N, and P (Smith and Read 2008; Prieto et al. 2012; Ekblad et al. 2013; Hodge and Storer 2015; Mayor et al. 2015; Lazcano et al. 2014). Moreover, their widespread distribution among a wide array of plant species makes them a powerful tool to manage agricultural and environment sustainability (Azul et al. 2014; Bhardwaj et al. 2014). Mycorrhizal fungi influence plant community structure and play a pivotal role in plant community assemblage and succession (Hartnett and Wilson 1999; Heneghan et al. 2008; Kikvidze et al. 2010; Lin et al. 2015). Further, AMF facilitate the regeneration and emergence of newly developed seedlings (Stanley et al. 1993; Barea et al. 2002), alter species interactions, and change the dynamics of plant communities' thereby increasing plant diversity in terrestrial ecosystems (van der Heijden et al. 1998; Klironomos et al. 2000; Dhillion and Gardsjord 2004; Simard and Austin 2010; Horn et al. 2017). AM association enhances plant's survival rate under unfavourable soil conditions, hence increasing the plant community diversity and ecosystem productivity (van der Heijden et al. 1998; Klironomos et al. 2000; Dhillion and Gardsjord 2004). It has been noticed that ecosystem instability is increasing in the modern agriculture system that might be due to lack of AMF interactions in soil (Helgason et al. 1998; Jeffries and Barea 2012). However, the incorporation of AMF could reverse the threat to biodiversity and ecosystem instability in the present day cultivation (Tilman 1996).

20.6.3 Reclamation of Degraded Land/Bioremediation

Increase in industrialization, urbanization, mining and several other human-induced activities across the globe have led to increased accumulation of toxic elements/ heavy metals in the groundwater and soil (Sharma et al. 2017), which largely affect both soil fertility as well as plant productivity (Whitmore 2006). AMF play an impending role in the restoration of degraded waste lands (Asmelash et al. 2016; Nicolson 1967). The extraradical hyphae of AMF improve the vegetation cover of the degraded lands by significantly enhancing the assimilation of nutrients (Jha

et al. 1994; Khan 2006; Gohre and Paskowski 2006). AMF attribute to the seedlings emergence and their establishment and survival in the polluted areas/lands and maintain the diversity in severely heavy metals contaminated soils (Hassan et al. 2011; Karthikeyan and Krishnakumar 2012; Manaut et al. 2015). AMF alleviate the impact of heavy metals by immobilizing them in the soil, chelation upon absorption, adsorption on the cell wall, altering pH of the rhizosphere soil and by enhancing phytostabilization and phytoextraction ability of the plants (Gaur and Adholeya 2004; Malekzadeh et al. 2011; Abdel Latef et al. 2016). Garg and Singla (2012) observed that inoculation with AMF increased the chlorophyll and relative water content in pea plants growing in the arsenic contaminated soils. Shabani et al. (2016) reported reduced transportation of Ni from the root to shoot along with the enhanced level of ABC transporter and metalothein on inoculation with Glomus mossae and Festuca arundinacea growing in the Ni polluted soil. Several studies have demonstrated that molecular expression of genes like Zn (GintZnT1). As (GiArsA), Cu and Cd (GintABC1, GmarMt1) likely to play a crucial role in plant's heavy metal tolerance (Lanfranco et al. 2002; Gonzalez-Guerrero et al. 2005; Lenoir et al. 2016). AMF predominantly prevail in the stressed habitats and unusable soils, and establish a mutual relationship with the plant species growing on the toxic soil (Cabral et al. 2015; Lenoir et al. 2016). The glomalin produced by AMF adsorbs the heavy metal from the soil and reduce the soil toxicity (Gil-Cardeza et al. 2014; Wu et al. 2014a, b). Experimentation under axenic conditions has shown that AMF exhibit potential for bioremediation of contaminated soils (Mugnier and Mosse 1987; Declerck et al. 2005; Cabral et al. 2010). In the light of occurrence and importance of AMF in the degraded waste land, it may be tempting to state that AMF could prove to be a potential approach to reclaim disturbed/contaminated soils. However, to establish the fact further research, particularly at molecular level is required.

20.6.4 Preventing Soil Erosion

Increasing soil erosion due to anthropogenic activities has become a potential cause of land degradation and desertification. Besides, increasing applications of P fertilizers of which only a small amount is absorbed by the plant and rest of it gets washed-off due to the tillage-induced soil erosion and reduced water retention capacity of soil. Since AM symbiosis plays key role in ecosystem functioning, these fungi could be applied to overcoming the problem of soil erosion (Miller and Jastrow 1990; Perumal and Maun 1999; Enkhtuya et al. 2003; Estaun et al. 2007). Of all the processes involved, production of mucilage, glomalin related soil protein (GRSP) and other extracellular compounds are the most studied one. These compounds provide strength to soil particles to formulate aggregations (Wright and Upadhaya 1996; Rillig et al. 2002; Rillig 2004a, b). A positive correlation has been found between the length of AMF hyphae and water stable aggregate through the production of glomalin related soil protein (GRSP) (Miller and Jastrow 1990; Wright and Anderson 2000; Rillig et al. 2002). Glomalin related soil protein has also been found to maintain stability of soil. Mycorrhiza-induced GRSP fraction has a major role in aggregate stability in citrus rhizosphere (Wu et al. 2014a, b). However, soil aggregate stability may differ among the AMF species. Schreiner et al. (1997) observed a variation in the size of the aggregate formed by *Glomus mossae*, which were larger than those formed by *G. etunicatum* and *Gigaspora rosae*. Miller and Jastrow (1990) revealed a direct correlation between the spore density and water stable aggregate. Beside glomalin production, AMF-mediated dense hyphae production, entanglement of soil particles with hyphae, improved soil microbiota also play important role in soil aggregation (Rillig et al. 2002; Rillig and Mummey 2006; Sharma et al. 2017). Thus, in order to avail maximum benefits of AMF for a soil improvement program more studied are required to better understand, how AMF get involved in the formation of soil aggregation and what are other mechanisms behind it must be examined?

20.6.5 Mitigation of Global Warming and Climate Change

In addition to promoting plant growth, AMF can directly or indirectly contribute to the stabilization of carbon in the soil. AMF extraradical mycelium (AEM) has a carbon-rich component known as glycoprotein or glomalin that can stay in the soil for decades. AMF seem to have a priming impact on the other soil microbes to convert plant waste into stable soil carbon (Cheng et al. 2012), attributing to reduce atmospheric carbon-based green house gases (Giri and Saxena 2017). Research experiments conducted to study climate change impact have assessed that there is already too much GHGs in the atmosphere. To stop the progress of and reverse the effects of human-induced climate change, GHGs production must be reduced and controlled in the atmosphere. Plants have in built capacity, if harnessed appropriately, to fix carbon back into the soil in sufficient amount to make a significant contribution to combat climate change. AMF facilitate supply of the mineral nutrients to the host plant indeed increase the growth and biomass of the plant, harnessing more atmospheric carbon. Further, the sugar produced by the host is a prerequisite of AMF to meet their C requirement for own growth and life-cycle (Giri and Saxena 2017). AMF could play a vital role in the global carbon cycle, because these fungi can exploit a large percentage of the carbon fixed by their host (about 20%) under ambient atmospheric CO₂ conditions (Jakobsen and Rosendahl 1990, Drigo et al. 2010). They could help in depositing slow cycling organic compounds and protect organic matter from microbial attack (Smith and Read 2008; Verbruggen et al. 2013). This exhibits that AMF in fact promote aggregation of soil particles and help in managing soil erosion (Wilson et al. 2009). AMF help in soil C sequestration, particularly under elevated CO_2 concentration (Treseder 2016) as these fungi are the only producers of glomalin (a recalcitrant glue-like glycoprotein), which enters into the soil on the death of fungal mycelium and become a source of soil organic carbon sink (Wright et al. 1996, Wright and Upadhaya 1996).

In addition to atmospheric CO₂, nitrous oxide (N₂O) is another effective greenhouse gas involved in the destruction of the protective ozone layer in the stratosphere. It contributes to global warming with a several times higher global warming potential than atmospheric CO₂ (Forster et al. 2007; Ravishankara et al. 2009). Montzka et al. (2011) demonstrated that after carbon dioxide and methane, the nitrous oxide place highest impact on the greenhouse effect. The importance of N₂O is likely to increase due to its prolonged existence and a predicted increase in future emissions. However, the ecological processes regulating N_2O emissions from soil are not well known. Bender et al. (2014) revealed that the presence of AMF, which have a profound impact on a wide range of ecosystem functions, could shrink N₂O emissions from soil. They manipulated the abundance of AMF in two independent greenhouse experiments and two different soils and found that N₂O emissions increase by 42 and 33% in microcosms with decreased AMF abundance than microcosms with well-established AMF propagules. These results suggested that the N₂O emission from soil is controlled by AMF. They further explained that the reduced N₂O emission in the atmosphere could partially be attributed to the increased N immobilization into microbial or plant biomass and decreased level of soil N as a substrate for N₂O emission and altered water relations. Bender and his colleagues (2014) concluded that the intensification of agricultural practices may further contribute to increased N₂O emissions as it disrupts the development and proliferation of AMF hyphal network. At the high soil moisture levels AMF control N₂O emissions by increasing use of soil water (Lazcano et al. 2014).

20.7 Research Need and Approaches

AMF characteristics make them a potential tool to be utilized for the sustainable management of agriculture and environment (Rodriguez and Sanders 2015; Bhardwaj et al. 2014). Nevertheless, several agronomic practices including crop rotation, soil tillage, use of fertilizers and pesticides largely influence abundance and infectivity of mycorrhizal fungi (Jansa et al. 2002). These practices disturb AMF hyphal networks, destruct their colonization of roots and decrease the absorption of phosphorus from the soil (Douds et al. 1995). Further, conventional tillage significantly decreases mycorrhizal diversity (Alguacil et al. 2008) whereas in zero tillage condition AMF sporulation increased twofold, even in highly fertilized soil (Brito et al. 2011; Verzeaux et al. 2017). Further, land and air pollution, mining, deforestation, and many biotic and abiotic factors largely impacts mycorrhizal survivability, and cause severe loss to the viability of mycorrhizal propagules, resulting in a significant reduction in the mycorrhizal colonization of roots (Gehring and Bennett 2009; Zobel and Öpik 2014; Antoninka et al. 2015; Borriello et al. 2015; Klabi et al. 2015). Therefore, it important to understand that; (1) better understanding of the relative contribution of AMF to any aspect to sustainability by attaining a broad view of all the possible pathways by which they can influences sustainability, including their interactions with other soil microbial biome (Rillig et al. 2016), (2) Focus on the conservation and management of AMF diversity and abundance to accomplish the goal of sustainability. Special attention is required towards the sensitive ecosystems, which are directly affected due to inadequate soil management and climate change such as glaciers (Haeberli and Beninston 1998; Jiang and Zhang 2015), and highly exploited ecosystems like mountains (Kohler and Maselli 2009; Gurung and Bajracharya 2012) should also be prioritised for conservation.

Green technology/approach not only includes the decrease in use of chemical fertilizers but also the enhancement of the sustainable techniques. For the exploitation of AMF as a tool for green technology, we aim to optimize its benefits by increasing its abundance and diversity (Rillig et al. 2016) in the soil through various strategies including (1) Field study Assessment, (2) Analysis, (3) Management and Protection policies, (4) Advanced Improvement techniques, (5) Field trials with AMF (Fig. 20.2). In field study, various soil parameters like physical (structure), chemical (pH, moisture, etc), nutrient status and abundance and diversity of beneficial microorganisms (mycorrhiza and associated microbes) are monitored, as the abundance and diversity of AMF are influenced by the soil chemistry (Casazza et al.



AMF - Green Technology

Fig. 20.2 An overview of various approaches associated with AMF-colonized plant roots, and their possible influencing role in sustainable agriculture and environment

2017). Data from field assessment records show the status of soil of the studied field and about the existing mycorrhizal and other microbial community which further provide site-specific information requirement about management and protection of AMF abundance and functioning (Rillig et al. 2016). Various crop management practices have shown improved yield in relation to mycorrhizal abundance (Monreal et al. 2015). Strategies with zero or less tillage and cover cropping have also shown enhanced root colonization and density of AMF in agriculture fields (Brito et al. 2013; Bowles et al. 2016). Moreover, advanced technologies of microbial community engineering and techniques like plant breeding are required to maintain the abundance and diversity of beneficial mycorrhiza and for manipulation and improvement of AMF with desirable traits respectively (Mueller and Sachs 2015; Hohmann and Messmer 2017). Production and field applications of AMF inoculum is the direct way to enhance the AMF propagules density in the agricultural field (Solaiman et al. 2014; Hijri 2016); however, constrain is the functioning of AMF in collaboration with many other microbes.

20.8 Conclusion and Future Perspectives

To enhance the ecosystem functioning and agricultural productivity without disturbing the balance of ecosystems and environment, utilization of arbuscular mycorrhizal fungi as a biofertilizer could be a potential solution; however, to achieve increased production, a major constraint is the development of AMF culture, abundance and density in the crop fields. Although AMF play a positive role in maintaining soil and plant health, the re-establishment of natural level of AMF richness is a major task, which could substitute the harmful chemical fertilizers and recompense a way to sustainable agriculture and environment. AMF could be utilized as a sustainable tool to improve the concentration of both macro- and micronutrients; hence could be an alternative to agronomic bio-fortification. Since AMF facilitate plants with macro- and micronutrients, it could positively influence the herbage, yield and quality of the crop, which is probably difficult to achieve using agro-chemicals. Therefore, we can recommend that field inoculation with AMF, depending upon the types of plant species and their environmental conditions, could be an effective alternative to agrochemicals. As the population is increasing day-by-day, there is a need of increased food production too, which could be only possible with the help of sustainable techniques or green techniques, like AMF-based bio-fertilizations along with PGPRs. To begin with, field trials with AMF inoculants and farmer's awareness towards cost-effective techniques are important to reducing input of agrochemicals in the agriculture. Once agricultural fields are enriched with microbial inoculants, this approach can be of great significant for managing sustainable agriculture and environment at the large scale.

References

- Abd-Alla MH, El Enany AWE, Nafady NA, Khalof DM, Morsy FM (2014) Symbiotic interaction of *Rhizobium leguminosarum* bv viciae and Arbuscular mycorrhizal fungi as plant growth promoting biofertilizers for faba bean (*Vicia faba* L.) in alkaline soils. Microbiol Res 169:49–58
- Abdel Latef AA, Hashem A, Rasool A, Abd_Allah EF, Alqarawi AA, Dilfuza E, Sumira Jan, Naser AA, Parvaiz A (2016) Arbuscular mycorrhizal symbiosis and abiotic stress in plants: a review. J Plant Physiol 59:407–426
- Adeyemi OR, Atayese MO, Dare MO, Sakariyawo SO, Adigbo SO, Bakare TO (2015) Weed control efficacy and arbuscular mycorrhizal (AM) colonization of upland rice varieties as affected by population densities. J Biol Agric Healthcare 5:178–185
- Aggarwal A, Kadian N, Tanwar A, Yadav A, Gupta KK (2011) Role of arbuscular mycorrhizal fungi (AMF) in global sustainable development. J Appl Nat Sci 3:340–351
- Alguacil MM, Lumini E, Roldan A, Salinas-García JR, Bonfante P, Bianciotto V (2008) The impact of tillage practices on arbuscular mycorrhizal fungal diversity in subtropical crops. Ecol Appl 18:527–536
- Al-Karaki GN, Hammad R, Rusan M (2001) Response of two tomato cultivars differing in salt tolerance to inoculation with mycorrhizal fungi under salt stress. Mycorrhiza 11:41–47
- Allen JW, Shachar-Hill Y (2009) Sulfur transfer through an arbuscular mycorrhiza. Plant Physiol 149:549–560
- Allen MF, Moore TS, Christensen M (1982) Phytohormone changes in *Bouteloua gracilis* infected by vesicular–arbuscular mycorrhizae. II. Altered levels of gibberellin-like substances and abscisic acid in the host plant. Can J Bot 60:468–471
- Ames RN, Reid CPP, Ingham ER (1984) Rhizosphere bacterial population responses to root colonization by a vesicular arbuscular mycorrhizal fungus. New Phytol 96:555–563
- Amora-Lazcano E, Vazquez MM, Azcon R (1998) Response of nitrogen-transforming microorganisms to arbuscular mycorrhizal fungi. Biol Fertil Soils 27:65–70
- Ampong-Nyarko K, Datta SK (1991) A handbook for weed control in rice. International Rice Research Institute, Manila, Philippines, p 113. ISBN-13:9789712200205
- Antoninka AJ, Ritchie ME, Johnson NC (2015) The hidden Serengeti-Mycorrhizal fungi respond to environmental gradients. Pedobiologia (Jena) 58:165–176
- Asmelash F, Bekele T, Birhane E (2016) The potential role of arbuscular mycorrhizal fungi in the restoration of degraded lands. Front Microbiol 7:1095. https://doi.org/10.3389/fmicb.2016. 01095
- Atul-Nayyar A, Hamel C, Hanson K, Germida J (2009) The arbuscular mycorrhizal symbiosis links N mineralization to plant demand. Mycorrhiza 19:239–246
- Aubrecht L, Staněk Z, Koller J (2006) Electrical measurement of the absorption surfaces of tree roots by the earth impedance methods: 1. Theory. Tree Physiol 26:1105–1112
- Augé RM (2001) Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. Mycorrhiza 11:3–42
- Augé RM (2004) Arbuscular mycorrhizae and soil/plant water relations. Can J Soil Sci 84:373-381
- Augé RM, Toler HD, Saxton AM (2015) Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: a meta-analysis. Mycorrhiza 25:13–24
- Azul AM, Nunes J, Ferreira I, Coelho AS, Veríssimo P, Trovão J, Campos A, Castro P, Freitas H (2014) Valuing native ectomycorrhizal fungi as a Mediterranean forestry component for sustainable and innovative solutions 1. Botany 92:161–171
- Bagheri S, Ebrahimi MA, Davazdahemami S, Moghadam JM (2014) Terpenoids and phenolic compounds production of mint genotypes in response to mycorrhizal bioelicitors. TJEAS J 4:339–348

- Bagyaraj DJ, Sharma MP, Maiti D (2015) Phosphorus nutrition of crops through arbuscular mycorrhizal fungi. Curr Sci 108:1288–1293
- Balestrini R, Bonfante P (2014) Cell wall remodeling in mycorrhizal symbiosis: a way towards biotrophism. Front Plant Sci 5:237. https://doi.org/10.3389/fpls.2014.00237
- Balestrini R, Goimez-Ariza J, Lanfranco L, Bonfante P (2007) Laser microdissection reveals that transcripts for five plants and one fungal phosphate transporter genes are contemporaneously present in arbusculated cells. Mol Plant Microbe Interact 20:1055–1062
- Barea JM (1991) Vesicular-arbuscular mycorrhizae as modifiers of soil fertility. Adv Soil Sci 15:1–40
- Barea JM, Gryndler M, Lemananceau P, Schuepp H, Azcon R (2002) The rhizosphere of mycorrhizal plants. In: Gianinazzi S, Schuepp H, Barea JM, Haselwandter K (eds) Mycorrhizal technology in agriculture: from genes to bioproducts. Birkhauser, Basel
- Baslam M, Garmendia I, Goicoechea N (2013) Enhanced accumulation of vitamins, nutraceuticals and minerals in lettuces associated with arbuscular mycorrhizal fungi (AMF): a question of interest for both vegetables and humans. Agriculture 3:188–209
- Bender SF, Plantenga F, Neftel A, Jocher M, Oberholzer H-R, Koehl L, Giles M, Daniell TJ, van der Heijden MGA (2014) Symbiotic relationships between soil fungi and plants reduce N₂O emissions from soil. ISME J 8:1336–1345
- Bender SF, Wagg C, van der Heijden MGA (2016) An underground revolution: biodiversity and soil ecological engineering for agricultural sustainability. Trends Ecol Evol 31:440–452
- Bhardwaj D, Ansari M, Sahoo R, Tuteja N (2014) Biofertilizers function as key player in sustainable agriculture by improving soil fertility, plant tolerance and crop productivity. Microb Cell Fact 13:66. https://doi.org/10.1186/1475-2859-13-66
- Bhat PR, Kaveriappa KM (2007) Effect of AM fungi on the growth and nutrition uptake in some endemic Myristicaceae members of the Western ghats, India. In: Tiwari M, Sati SC (eds) The mycorrhizae: diversity, ecology and application. Daya Pub. House, Delhi, pp 295–309
- Biermann B, Linderman RG (1983) Increased geranium growth using pre-transplant inoculation with a mycorrhizal fungus. J Am Soc Hortic Sci 108:972–976
- Birhane E, Sterck FJ, Fetene M, Bongers F, Kuyper TW (2012) Arbuscular mycorrhizal fungi enhance photosynthesis, water use efficiency, and growth of frankincense seedlings under pulsed water availability conditions. Oecologia 169:895–904
- Borriello R, Berruti A, Lumini E, Beffa MTD, Scariot V, Bianciotto V (2015) Edaphic factors trigger diverse AM fungal communities associated to exotic camellias in closely located Lake Maggiore (Italy) sites. Mycorrhiza 25:253–265
- Bowles TM, Jackson LE, Loeher M, Cavagnaro TR (2016) Ecological intensification and arbuscular mycorrhizas: a meta-analysis of tillage and cover crop effects. J Appl Ecol. https://doi.org/10.1111/1365-2664.12815
- Brito I, Carvalho M, Goss MJ (2011) The importance of no-till in the development of cropping systems to maximize benefits of arbuscular mycorrhiza symbiosis. In: Elizabeth Stockdale E, Watson C (eds) Proceedings of the Association of applied biologist "Making crop rotations fit for the future". Aspects of applied biology, pp 137–141
- Brito I, Carvalho M, Goss MJ (2013) Soil and weed management for enhancing arbuscular mycorrhiza colonization of wheat. Soil Use Manag 29:540–546
- Brodt S, Six J, Feenstra G, Ingels C, Campbell D (2011) Sustainable agriculture. Nat Edu Know 3:1
- Brundrett MC (2002) Coevolution of roots and mycorrhizas of land plants. New Phytol 154:275–304
- Brundrett MC (2008) Mycorrhizal associations: the web resource. Date accessed
- Brundrett MC, Piche Y, Peterson RL (1985) A developmental study of the early stages in vesicular arbuscular mycorrhiza formation. Can J Bot 63:184–194
- Cabral L, Siqueira J, Soares C et al (2010) Retention of heavy metals by arbuscular mycorrhizal fungi mycelium. Quím Nova 33:25–29

- Cabral L, Soares CR, Giachini AJ, Siqueira JO (2015) Arbuscular mycorrhizal fungi in phytoremediation of contaminated areas by trace elements: mechanisms and major benefits of their applications. World J Microbiol Biotechnol 31:1655–1664
- Cappellazzo G, Lanfranco L, Fitz M, Wipf D, Bonfante P (2008) Characterization of an amino acid permease from the endomycorrhizal fungus *Glomus mosseae*. Plant Physiol 147:429–437
- Casazza G, Lumini E, Ercole E, Dovana F, Guerrina M, Arnulfo A, Minuto L, Fusconi A, Mucciarelli M (2017) The abundance and diversity of arbuscular mycorrhizal fungi are linked to the soil chemistry of screes and to slope in the Alpic paleo-endemic *Berardia subacaulis*. PLoS One 12(2):e0171866. https://doi.org/10.1371/journal.pone.0171866
- Castellanos-Morales V, Villegas J, Wendelin S, Vierheilig H, Eder R, Cardenas-Navarro R (2010) Root colonization by the arbuscular mycorrhizal fungus *Glomus intraradices* alters the quality of strawberry fruits (*Fragaria x ananassa* Duch.) at different nitrogen levels. J Sci Food Agric 90:1774–1782
- Chakraborty K, Bose J, Shabala L, Shabala S (2016) Difference in root K⁺ retention ability and reduced sensitivity of K⁺ permeable channels to reactive oxygen species confer differential salt tolerance in three *Brassica* species. J Exp Bot 67:4611–4625
- Chen SC, Jin WJ, Liu AR, Zhang SJ, Liu DL, He CX (2013) Arbuscular mycorrhizal fungi (AMF) increase growth and secondary metabolism in cucumber subjected to low temperature stress. Sci Hortic (Amst) 160:222–229
- Chen M, Yang G, Sheng Y, Li P, Qui H, Zhou X, Huang L, Chao Z (2017) Glomus mosseae inoculation improves the root system architecture, photosynthetic efficiency and flavonoids accumulation of Liquorice under nutrient stress. Front Plant Sci 8:931. https://doi.org/10.3389/ fpls.2017.00931
- Cheng L, Booker FL, Tu C, Burkey KO, Zhou L, Shew HD, Ruffy TW, Hu S (2012) Arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO₂. Science 337:1084–1087
- Cliquet JB, Murray PJ, Boucaud J (1997) Effect of the arbuscular mycorrhizal fungus *Glomus* fasciculatum on the uptake of amino nitrogen by *Lolium perenne*. New Phytol 137:345–349
- Cseresnyés I, Takács T, Végh RK, Anton A, Rajkai K (2013) Electrical impedance and capacitance method: a new approach for detection of functional aspects of arbuscular mycorrhizal colonization in maize. Eur J Soil Biol 54:25–31
- Cseresnyés I, Takács T, Füzy A, Rajkai K (2014) Simultaneous monitoring of electrical capacitance and water uptake activity of plant root system. Int Agrophys 28:537–541
- Daei G, Ardakani M, Rejali F, Teimuri S, Miransari M (2009) Alleviation of salinity stress on wheat yield, yield components, and nutrient uptake using arbuscular mycorrhizal fungi under field conditions. J Plant Physiol 166:617–625
- Declerck S, Strullu D, Fortin J (2005) In vitro culture of mycorrhizas. Springer, New York
- Del Val C, Barea JM, Azcón-Aguilar C (1999) Assessing the tolerance to heavy metals of arbuscular mycorrhizal fungi isolated from sewage sludge-contaminated soils. Appl Soil Ecol 11:261–269
- Dhillion SS, Gardsjord TL (2004) Arbuscular mycorrhizas influence plant diversity, productivity, and nutrients in boreal grasslands. Can J Bot 82:104–114
- Diaz-Zorita M, Perfect E, Grove JH (2002) Disruptive methods for assessing soil structure. Soil Tillage Res 64:3–22
- Douds DD Jr, Galvez L, Janke RR, Wagoner P (1995) Effect of tillage and farming system upon populations and distribution of vesicular-arbuscular mycorrhizal fungi. Agric Ecosyst Environ 52:111–118
- Drigo B, Pijl AS, Duyts H, Kielak AM, Gamper HA, Houtekamer MJ (2010) Shifting carbon flow from roots into associated microbial communities in response to elevated atmospheric CO₂. Proc Natl Acad Sci USA 107:10939–10942
- Ekblad A, Wallander H, Godbold DL, Cruz C, Johnson D, Baldrian P, Björk RG, Epron D, Kieliszewska-Rokicka B, Kjøller R et al (2013) The production and turnover of extramatrical mycelium of ectomycorrhizal fungi in forest soils: role in carbon cycling. Plant Soil 366:1–27

- Elhindi KM, El-Din AS, Elgorban AM (2017) The impact of arbuscular mycorrhizal fungi in mitigating salt-induced adverse effects in sweet basil (*Ocimum basilicum* L.). Saudi J Biol Sci 24(1):170–179
- Engel R, Szabó K, Abrankó L, Rendes K, Füzy A, Takács T (2016) Effect of arbuscular mycorrhizal fungi on the growth and polyphenol profile of marjoram, lemon balm, and marigold. J Agric Food Chem 64:3733–3742
- Enkhtuya B, Oskarsson U, Dodd JC, Vosatka M (2003) Inoculation of grass and tree seedlings used for reclaiming eroded areas in Iceland with mycorrhizal fungi. Folia Geobot 38:209–222
- Estaun V, Vicente S, Calvet C, Camprubi A, Busquets M (2007) Integration of arbuscularmycorrhiza inoculation in hydroseeding technology. Effects on plant growth and interspecies competition. Land Degrad Dev 18:621–630
- Evelin H, Kapoor R, Giri B (2009) Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. Ann Bot 104:1263–1280
- Evelin H, Giri B, Kapoor R (2013) Ultrastructural evidence for AMF mediated salt stress mitigation in *Trigonella foenum–graecum*. Mycorrhiza 23:71–86
- Fan Q, Liu J (2011) Colonization with arbuscular mycorrhizal fungus affects growth, drought tolerance and expression of stress-responsive genes in *Poncirus trifoliata*. Acta Physiol Plant 33:1533–1542
- Farmer MJ, Li X, Feng G, Zhao B, Chatagnier O, Gianinazzi S, Gianinazzi-Pearson V, Van Tuinen D (2007) Molecular monitoring of field-inoculated AMF to evaluate persistence in sweet potato crops in China. Appl Soil Ecol 35:599–609
- Fellbaum CR, Mensah J, Cloos A, Pfeffer P, Strahan G, Kiers ET, Bücking H (2014) Fungal nutrient allocation in common mycorrhizal networks is regulated by the carbon source strength of individual host plants. New Phytol 2:646–656
- Fitzgerald JW (1976) Sulfate ester formation and hydrolysis: potentially important yet often ignored aspect of sulfur cycle of aerobic soils. Bacteriol Rev 40:698–721
- Forster P, Ramaswamy V, Artaxo P, Berntsen T, Betts R, Fahey DW (2007) Changes in atmospheric constituents and in radiative forcing. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB et al (eds) Climate change: the physical science basis. Contribution of working group I to the fourth assessment report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge and New York, NY
- França AC, de Freitas AF, dos Santos EA, Grazziotti PH, de Andrade Júnior VC (2016) Mycorrhizal fungi increase coffee plants competitiveness against *Bidens pilosa* interference. Pesqui Agropecu Trop Goiânia 46:132–139
- Friese CF, Allen MF (1991) The spread of VA mycorrhizal fungal hyphae in the soil: inoculum types and external hyphal architecture. Mycologia 83:409–418
- Garg N, Singla P (2012) The role of *Glomus mosseae* on key physiological and biochemical parameters of pea plants grown in arsenic contaminated soil. Sci Hortic 143:92–101
- Gaur A, Adholeya A (2004) Prospects of arbuscular mycorrhizal fungi in phytoremediation of heavy metal contaminated soils. Curr Sci 86:528–534
- Gehring C, Bennett A (2009) Mycorrhizal fungal-plant-insect interactions: the importance of community approach. Environ Entomol 38:93–102
- Gerz M, Bueno CG, Zobel M, Moora M (2016) Plant community mycorrhization in temperate forests and grasslands: relations with edaphic properties and plant diversity. J Veg Sci 27:89–99
- Gianinazzi S, Gollotte A, Binet MN, van Tuinen D, Redecker D, Wipf D (2010) Agroecology: the key role of arbuscular mycorrhizas in ecosystem services. Mycorrhiza 20:519–530
- Gil-Cardeza ML, Ferri A, Cornejo P et al (2014) Distribution of chromium species in a Cr-polluted soil: presence of Cr (III) in glomalin related protein fraction. Sci Total Environ 493:828–833
- Giller KE, Witter E, McGrath SP (1998) Toxicity of heavy metals to microorganisms and microbial processes in agriculture soils: a review. Soil Biol Biochem 30:1389–1414

- Giovannetti M, Tolosano M, Volpe V, Kopriva S, Bonfante P (2014) Identification and functional characterization of a sulfate transporter induced by both sulfur starvation and mycorrhiza formation in *Lotus japonicus*. New Phytol 204:609–619
- Giri B, Saxena B (2017) Response of arbuscular mycorrhizal fungi to global climate change and their role in terrestrial ecosystem C and N cycling. In: Varma A, Prasad R, Tuteja N (eds) Mycorrhiza-functions, diversity and state of the art. Springer, Cham, pp 305–327
- Gohre V, Paskowski U (2006) Contribution of the arbuscular mycorrhizal symbiosis to heavy metal phytoremediation. Planta 223:1115–1122
- Gomes SIF, Merckx VSFT, Saavedra S (2017) Fungal-host diversity among mycoheterotrophic plants increases proportionally to their fungal-host overlap. Ecol Evol 10:3623–3630
- Gonzalez-Guerrero M, Azcon-Aguilar C, Mooney M (2005) Characterization of a *Glomus intraradices* gene encoding a putative Zn transporter of the cation diffusion facilitator family. Fungal Genet Biol 42:130–140
- Graham JH (2000) Assessing cost of arbuscular mycorrhizal symbiosis in agrosystems. In: Podila GK, Donds DD (eds) Current advances in mycorrhizae research. APS Press, St Paul, pp 127–140
- Guo T, Zhang JL, Christie P, Li XL (2007) Pungency of spring onion as affected by inoculation with arbuscular mycorrhizal fungi and sulfur supply. J Plant Nutr 30:1023–1034
- Gurung J, Bajracharya RM (2012) Climate change and glacial retreat in the Himalaya: implications for soil and plant development. Kathm Univ J Sci Engin Tech 8:153–163
- Haeberli W, Beninston M (1998) Climate change and its impacts on glaciers and permafrost in the Alps. AMBIO J Hum Environ 27:258–265
- Hampp R, Nehls U, Wallenda T (2000) Physiology of mycorrhiza. In: Esser K, Kadereit JW, Lüttge U, Runge M (eds) Progress in botany. Genetics, physiology, systemates, ecology. Springer, Berlin, pp 223–254
- Harrison MJ, Dewbre GR, Liu JY (2002) A phosphate transporter from *Medicago truncatula* involved in the acquisition of phosphate released by arbuscular mycorrhizal fungi. Plant Cell 14:2413–2429
- Hartnett DC, Wilson WT (1999) Mycorrhizae influence plant community structure and diversity in tall grass prairie. Ecology 80:1187–1195
- Hassan SE, Boon E, St-Arnaud M, Hijri M (2011) Molecular biodiversity of arbuscular mycorrhizal fungi in trace metal-polluted soils. Mol Ecol 20:3469–3483
- Hazzoumi Z, Moustakime Y, Elharchli EH, Khalid AJ (2015) Effect of arbuscular mycorrhizal fungi (AMF) and water stress on growth, phenolic compounds, glandular hairs, and yield of essential oil in basil (*Ocimum gratissimum* L). Chem Biol Technol Agric 2:10. https://doi.org/ 10.1186/s40538-015-0035-3
- Heap I (2015) The international survey of herbicide resistant weeds. Retrieved from www. weedscience.org
- Helgason T, Daniell TJ, Husband R, Fitter AH, Young JPW (1998) Ploughing up the wood-wide web? Nature 394:431
- Heneghan L, Miller SP, Baer S, Callaham MA, Montgomery J, Pavao-Zuckerman M (2008) Integrating soil ecological knowledge into restoration management. Restor Ecol 16:608–617
- Herman DJ, Firestone MK, Nuccio E, Hodge A (2012) Interactions between an arbuscular mycorrhizal fungus and a soil microbial community mediating litter decomposition. FEMS Microbiol Ecol 80:236–247
- Hijri M (2016) Analysis of a large dataset form field mycorrhizal inoculation trials on potato showed highly significant increase in yield. Mycorrhiza 26:209–214
- Hildebrandt U, Janetta K, Bothe H (2002) Towards growth of arbuscular mycorrhizal fungi independent of a plant host. Appl Environ Microbiol 68:1919–1924
- Hodge A, Fitter AH (2010) Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N cycling. Proc Natl Acad Sci USA 107:13754–13759
- Hodge A, Storer K (2015) Arbuscular mycorrhiza and nitrogen: Implications for individual plants through to ecosystems. Plant Soil 386:1–19

- Hohmann P, Messmer MM (2017) Breeding for mycorrhizal symbiosis: focus on disease resistance. Euphytica 213:113
- Horn S, Hempel S, Verbruggen E, Rillig MC, Caruso T (2017) Linking the community structure of arbuscular mycorrhizal fungi and plants: a story of interdependence? ISME J 11:1400–1411
- Huang Z, Krishnamurthy S, Panda A, Samal SK (2003) Newcastle disease virus V protein is associated with viral pathogenesis and functions as an alpha interferon antagonist. J Virol 77:8676–8685
- Hunter P (2016a) Plant microbiomes and sustainable agriculture. EMBO Rep 17:1696-1699
- Hunter P (2016b) Deciphering the plant microbiome and its role in nutrient supply and plant immunity has great potential to reduce the use of fertilizers and biocides in agriculture. Sci Soc 17:1696–1699
- Jakobsen I, Rosendahl L (1990) Carbon flow into soil and external hyphae from roots of mycorrhizal cucumber plants. New Phytol 115:77–83
- Jansa J, Mozafar A, Anken T, Ruh R, Sanders IR, Frossard E (2002) Diversity and structure of AMF communities as affected by tillage in a temperate soil. Mycorrhiza 12:225–234
- Jansa J, Mozafar A, Frossard E (2005) Phosphorus acquisition strategies within arbuscular mycorrhizal fungal community of a single field site. Plant Soil 276:163–176
- Jastrow JD, Miller RM, Lussenhop J (1998) Contributions of interacting biological mechanisms to soil aggregate stabilization in restored prairie. Soil Biol Biochem 30:905–916
- Jeffries P, Barea JM (2012) Arbuscularmycorrhiza a key component of sustainable plant-soil ecosystems. In: Hock B (ed) The mycota, Fungal associations, vol IX, 2nd edn. Springer, Berlin, Heidelberg, pp 51–75
- Jha DK, Sharma GD, Mishra RR (1994) Ecology of vesicular-arbuscular mycorrhiza. In: Prasad AB, Bilgrami RS (eds) Microbes and environments. Narendra Publishing House, Delhi, pp 199–208
- Jiang C, Zhang L (2015) Climate change and its impact on the eco-environment of the three-rivers headwater region on the Tibetan Plateau, China. Int J Environ Res Public Health 12:12057–12081
- Johansson RC, Gowda PH, Mulla DJ, Dalzell BJ (2004) Metamodelling phosphorus best management practices for policy use: a frontier approach. Agric Econ 30:63–74
- Johnson NC, Wilson GWT, Bowker MA, Wilson JA, Miller RA (2010) Resource limitation is a driver of local adaptation in mycorrhizal symbioses. Proc Natl Acad Sci USA 107:2093–2098
- Joner EJ, Johansen A (2000) Phosphatase activity of external hyphae of two arbuscular mycorrhizal fungi. Mycol Res 104:81–86
- Joner EJ, Briones R, Leyval C (2000) Metal-binding capacity of arbuscular mycorrhizal mycelium. Plant Soil 226:227–234
- Jordan NR, Zhang J, Huerd S (2000) Arbuscular-mycorrhizal fungi: potential roles in weed management. Weed Res 40:397–410
- Kapoor R, Giri B, Mukerji KG (2002) Glomus macrocarpum: a potential bioinoculant to improve essential oil quality and concentration in dill (Anethum graveolens L.) and carum (Trachyspermum ammi (Linn.) Sprague). World J Microbiol Biotechnol 18:459–463
- Kapoor R, Giri B, Mukerji KG (2004) Improved growth and essential oil yield and quality in *Foeniculum vulgare* mill on mycorrhizal inoculation supplemented with P-fertilizer. Bioresour Technol 93:3007–3011
- Karagiannidis N, Hadjisavva-Zinoviadi S (1998) The mycorrhizal fungus *Glomus mosseae* enhances growth, yield and chemical composition of a durum wheat variety in 10 different soils. Nutr Cycl Agroecosyst 52:1–7
- Karthikeyan A, Krishnakumar N (2012) Reforestation of bauxite mine spoils with *Eucalyptus* tereticornis Sm. seedlings inoculated with arbuscular mycorrhizal fungi. Ann For Res 55:207–216
- Kaur R, Singh A, Kang JS (2014) Influence of different types mycorrhizal fungi on crop productivity. Curr Agric Res 2:51–54

- Kaye JW, Pfleger FL, Stewart EL (1984) Interaction of *Glomus fasciculatum* and-*Pythium ultimum* on greenhouse-grown poinsettia. Can J Bot 62:1575–1579
- Khalid M, Hassani D, Bilal M, Liao J, Huang D (2017) Elevation of secondary metabolites synthesis in *Brassica campestris* ssp. *chinensis* L. via exogenous inoculation of *Piriformospora indica* with appropriate fertilizer. PLoS One 12(5):e0177185
- Khan AG (2006) Mycorrhiza remediation-an enhanced form of phytoremediation. J Zhejiang Univ Sci B 7:503–514
- Kikvidze Z, Armas C, Fukuda K, Martínez-García LB, Miyata M, Oda-Tanaka A (2010) The role of arbuscular mycorrhizae in primary succession: differences and similarities across habitats. Web Ecol 10:50–57
- Klabi R, Bell TH, Hamel C, Iwaasa A, Schellenberg M, Raies A, St-Arnaud M (2015) Plant assemblage composition and soil P concentration differentially affect communities of AM and total fungi in a semi-arid grassland. FEMS Microbiol Ecol 91:1–13
- Klironomos JN, McCune J, Hart M, Neville J (2000) The influence of arbuscular mycorrhizae on the relationship between plant diversity and productivity. Ecol Lett 3:137–141
- Kobae Y, Tamura Y, Takai S, Banba M, Hata S (2010) Localized expression of arbuscular mycorrhiza-inducible ammonium transporters in soybean. Plant Cell Physiol 51:1411–1415
- Koca H, Bor M, Özdemir F, Türkan İ (2007) The effect of salt stress on lipid peroxidation, antioxidative enzymes and proline content of sesame cultivars. Environ Exp Bot 60:344–351
- Kohler T, Maselli D (2009) Mountains and climate change from understanding to action. Published by Geographica Bernensia with the support of the Swiss Agency for Development and Cooperation (SDC), and an International Team of Contributors, Bern
- Kytoviita MM (2005) Role of nutrient level and defoliation on symbiotic function: experimental evidence by tracing C-14/N-15 exchange in mycorrhizal birch seedlings. Mycorrhiza 15:65–70
- Lambers H, Martinoia E, Renton M (2015) Plant adaptations to severely phosphorus-impoverished soils. Curr Opin Plant Biol 25:23–31
- Lanfranco L, Bolchi A, Ros EC, Ottonello S, Bonfante P (2002) Differential expression of a metallothionein gene during the presymbiotic versus the symbiotic phase of an arbuscular mycorrhizal fungus. Plant Physiol 130:58–67
- Lazcano C, Barrios-Masias FH, Jackson LE (2014) Arbuscular mycorrhizal effects on plant water relations and soil greenhouse gas emissions under changing moisture regimes. Soil Biol Biochem 74:184–192
- Leifheit EF, Veresoglou SD, Lehmann A, Morris EK, Rillig MC (2014) Multiple factors influence the role of arbuscular mycorrhizal fungi in soil aggregation – a meta-analysis. Plant Soil 374:523–537
- Lekberg Y, Koide RT (2005) Is plant performance limited by abundance of arbuscular mycorrhizal fungi? A meta-analysis of studies published between 1988 and 2003. New Phytol 168:189–204
- Lendzemo VW (2004) The tripartite interaction between sorghum, *Striga hermonthica*, and arbuscular mycorrhizal fungi. PhD thesis, Wageningen University, Wageningen, The Netherlands
- Lenoir I, Fontaine J, Lounès-Hadj A (2016) Arbuscular mycorrhizal fungal responses to abiotic stresses: a review. Phytochemistry 123:4–15
- Leustek T (1996) Molecular genetics of sulfate assimilation in plants. Physiol Plant 97:411-419
- Leyval C, Joner EJ, del Val C, Haselwandter K (2002) Potential of arbuscular mycorrhizal fungi for bioremediation. In: Gianinazzi S, Schüepp H, Barea JM, Haselwandter K (eds) Mycorrhizal technology in agriculture. Birkhäuser, Basel, pp 175–186
- Li HY, Smith SE, Holloway RE, Zhu YG, Smith FA (2006) Arbuscular mycorrhizal fungi contribute to phosphorus uptake by wheat grown in a phosphorus-fixing soil even in the absence of positive growth responses. New Phytol 172:536–543
- Lin G, McCormack ML, Guo D (2015) Arbuscular mycorrhizal fungal effects on plant competition and community structure. J Ecol 103:1224–1232
- Liu C, Muchhal US, Raghothama KG (1997) Differential expression of TPSI1, a phosphate starvation-inducible gene in tomato. Plant Mol Biol 33:867–874

- Liu JN, Wu LJ, Wei SG, Xiao X, Su CX, Jiang P, Song ZB, Wang T, Yu ZL (2007) Effects of arbuscular mycorrhizal fungi on the growth, nutrient uptake and glycyrrhizin production of licorice (*Glycyrrhiza uralensis* Fisch). Plant Growth Regul 52:29–39
- Lu F, Lee C, Wang C (2015) The influence of arbuscular mycorrhizal fungi inoculation on yam (*Dioscorea* spp.) tuber weights and secondary metabolite content. Peer J 3:e1266
- Malekzadeh E, Alikhani AH, Savaghebi-Fioozabadi RG, Zarei M (2011) Influence of arbuscular mycorrhizal fungi and an improving growth bacterium on Cd uptake and maize growth in Cd-polluted soils. Spanish J Agric Res 9:1213–1223
- Manaut N, Sanguin H, Ouahmane L, Bressan M, Thioulouse J, Baudoin E (2015) Potentialities of ecological engineering strategy based on native arbuscular mycorrhizal community for improving afforestation programs with carob trees in degraded environments. Ecol Eng 79:113–119
- Mandal S, Evelin H, Giri B, Singh VP, Kapoor R (2013) Arbuscular mycorrhiza enhances the production of stevioside and rebaudioside-A in *Stevia rebaudiana* via nutritional and non-nutritional mechanisms. Appl Soil Ecol 72:187–194
- Mandal S, Upadhyay S, Wajid S, Ram M, Jain DC, Singh VP, Kapoor R (2015) Arbuscular mycorrhiza increase artemisinin accumulation in *Artemisia annua* by higher expression of key biosynthesis genes via enhanced jasmonic acid levels. Mycorrhiza 25:345–357
- Manimozhi K, Gayathri D (2012) Eco friendly approaches for sustainable agriculture. J Environ Res Dev 7:166–173
- Marschner H, Dell B (1994) Nutrient uptake in mycorrhizal symbiosis. Plant Soil 159:89-102
- Matam P, Parvatam G (2017) Arbuscular mycorrhizal fungi promote enhanced growth, tuberous roots yield and root specific flavor 2-hydroxy-4-methoxy benzaldehyde content of *Decalepis hamiltonii* Wight and Arn. Acta Sci Pol Hortorum Cultus 16:3–10
- Mayor J, Bahram M, Henkel T, Buegger F, Pritsch K, Tedersoo L (2015) Ectomycorrhizal impacts on plant nitrogen nutrition: emerging isotopic patterns, latitudinal variation and hidden mechanisms. Ecol Lett 18:96–107
- McFarland J, Ruess R, Keilland K, Pregitzer K, Hendrick R, Allen M (2010) Cross-ecosystem comparisons of in situ plant uptake of amino acid-N and NH4+. Ecosystems 13:177–193
- Meier S, Cornejo P, Cartes P, Borie F, Medina J, Azcón R (2015) Interactive effect between Cu-adapted arbuscular mycorrhizal fungi and biotreated agrowaste residue to improve the nutritional status of *Oenothera picensis* growing in Cu-polluted soils. J Plant Nutr Soil Sci 178:126–135
- Miller RM, Jastrow JD (1990) Hierarchy of root and mycorrhizal fungal interactions with soil aggregation. Soil Biol Biochem 22:579–584
- Miransari M, Bahrami HA, Rejali F, Malakouti MJ (2008) Using arbuscular mycorrhiza to alleviate the stress of soil compaction on wheat (*Triticum aestivum* L.) growth. Soil Biol Biochem 40:1197–1206
- Mohamed AA, Wedad EEE, Heggo AM, Hassan EA (2014) Effect of dual inoculation with arbuscular mycorrhizal fungi and sulphur-oxidising bacteria on onion (*Allium cepa* L.) and maize (*Zea mays* L.) grown in sandy soil under greenhouse conditions. Ann Agric Sci 59:109–118
- Monreal CM, DeRosa M, Mallubhotla SC, Bindraban PS, Dimkpa C (2015) The application of nanotechnology for micronutrients in soil-plant systems. VFRC report 2015/3. Virtual Fertilizer Research Center, Washington, DC, p 44
- Montzka SA, Dlugokencky EJ, Butler JH (2011) Non-CO2 greenhouse gases and climate change. Nature 476:43–50
- Mueller UG, Sachs JL (2015) Engineering microbiomes to improve plant and animal health. Trends Microbiol 23:606–617
- Mugnier J, Mosse B (1987) Vesicular-arbuscular mycorrhizal infection in transformed rootinducing T-DNA roots grown axenically. Phytopathology 77:1045–1050
- Mwangi MW, Monda EO, Okoth SA, Jefwa JM (2011) Inoculation of tomato seedlings with *Trichoderma Harzianum* and arbuscular mycorrhizal fungi and their effect on growth and control of wilt in tomato seedlings. Braz J Microbiol 42:508–513

- Nadeem SM, Ahmad M, Zahir ZA (2014) The role of mycorrhizae and plant growth promoting rhizobacteria (PGPR) in improving crop productivity under stressful environment. Biotechnol Adv 32:429–448
- Nakano A, Takahashi K, Koide RT, Kimura M (2001) Determination of the nitrogen source for arbuscular mycorrhizal fungi by ¹⁵N application to soil and plants. Mycorrhiza 10:267–273
- Nicolson TH (1967) Vesicular-arbuscular mycorrhiza-a universal plant symbiosis. Sci Prog Oxf 55:561–581
- Nouri E, Breuillin-Sessoms F, Feller U, Reinhardt D (2014) Phosphorus and nitrogen regulate arbuscular mycorrhizal symbiosis in *Petunia hybrida*. PLoS One 9:e90841
- Oerke EC (2006) Crop losses to pests. J Agric Sci 144:31-43
- Pellegrino E, Bedini S (2014) Enhancing ecosystem services in sustainable agriculture: biofertilization and biofortification of chickpea (*Cicer arietinum* L.) by arbuscular mycorrhizal fungi. Soil Biol Biochem 68:429–439
- Pérez-Tienda J, Valderas A, Camañes G, García-Agustín P, Ferrol N (2012) Kinetics of NH₄+ uptake by the arbuscular mycorrhizal fungus *Rhizophagus irregularis*. Mycorrhiza 22:485–491
- Perumal JV, Maun MA (1999) The role of mycorrhizal fungi in growth enhancement of dune plants following burial in sand. Funct Ecol 13:560–566
- Pistelli LA, Ulivieri V, D'Angiolillo F, Giovannelli S, Pistelli LU, Giovannetti M (2015) Influence of arbuscular mycorrhizal fungi (AMF) in the production of secondary metabolites of *Bituminaria bituminosa* L. In: Proceedings of the joint congress SIBV-SIGA, Milano, Italy
- Porcel R, Ruiz-Lozano JM (2004) Arbuscular mycorrhizal influence on leaf water potential, solute accumulation and oxidative stress in soybean plants subjected to drought stress. J Exp Bot 55:1743–1750
- Porcel R, Gómez M, Kaldenhoff R, Ruiz-Lozano JM (2005) Impairment of NtAQP1 gene expression in tobacco plants does not affect root colonization pattern by arbuscular mycorrhizal fungi but decreases their symbiotic efficiency under drought. Mycorrhiza 15:417–423
- Porcel R, Aroca R, Ruiz-Lozano JM (2012) Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. Agron Sustain Dev 32:181–200
- Prasad R, Kumar M, Varma A (2015) Role of PGPR in soil fertility and plant health. In: Egamberdieva D, Shrivastava S, Varma A (eds) Plant Growth-Promoting Rhizobacteria (PGPR) and Medicinal Plants. Springer, Cham, pp 247–260
- Prieto I, Armas C, Pugnaire FI (2012) Water release through plant roots: new insights into its consequences at the plant and ecosystem level. New Phytol 193:830–841
- Quilambo OA (2000). Functioning of peanut (*Arachis hypogaea* L.) under nutrient deficiency and drought stress in relation to symbiotic associations. PhD thesis, University of Groningen, The Netherlands, Van Denderen B.V., Groningen
- Ravishankara AR, Daniel JS, Portmann RW (2009) Nitrous oxide (N2O): the dominant ozonedepleting substance emitted in the 21st century. Science 326:123–125
- Rennenberg H, Herschbach C, Haberer K, Kopriva S (2007) Sulfur metabolism in plants: are trees different? Plant Biol 9:620–637
- Rillig MC (2004a) Arbuscular mycorrhizae and terrestrial ecosystem processes. Ecol Lett 7:740–754
- Rillig MC (2004b) Arbuscular mycorrhizae, glomalin, and soil aggregation. Can J Soil Sci 84:355–363
- Rillig MC, Mummey DL (2006) Mycorrhizas and soil structure. New Phytol 171:41-53
- Rillig MC, Wright SF, Eviner V (2002) The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: comparing effects of five plant species. Plant Soil 238:325–333
- Rillig MC, Sosa-Hernandez MA, Roy J, Aguilar-Trigueros CA, Valyi K, Lehmann A (2016) Towards an integrated mycorrhizal technology: harnessing mycorrhiza for sustainable intensification in agriculture. Front Plant Sci 7:1625
- Rinaudo V, Bàrberi P, Giovannetti M, van der Heijden MGA (2010) Mutualistic fungi suppress aggressive agricultural weeds. Plant Soil 333:7–20
- Rodriguez A, Sanders IR (2015) The role of community and population ecology in applying mycorrhizal fungi for improved food security. ISME J 9:1053–1061

- Rojas-Andrade R, Cerda-Garcia-Rojas CM, Frias-Hernandez JT, Dendooven L, Olalde-Portugal-V, Ramos-Valdivia AC (2003) Changes in the concentration of trigonelline in a semi-arid leguminous plant (*Prosopis laevigata*) induced by an arbuscular mycorrhizal fungus during the presymbiotic phase. Mycorrhiza 13:49–52
- Ruiz-Lozano JM (2003) Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress: new perspectives for molecular studies. Mycorrhiza 13:309–317
- Ruiz-Lozano JM, Porcel R, Azcón C, Aroca R (2012) Regulation by arbuscular mycorrhizae of the integrated physiological response to salinity in plants: new challenges in physiological and molecular studies. J Exp Bot 63(11):4033–4044
- Sailo GL, Bagyaraj DJ (2005) Influence of different AM fungi on the growth, nutrition and forskolin content of *Coleus forskohlii*. Mycol Res 109:795–798
- Sambandan K, Kannan K, Raman N (1992) Distribution of vesicular-arbuscular mycorrhizal fungi in heavy metal polluted soils of Tamil-Nadu, India. J Environ Biol 13:159–167
- Saxena B, Shukla K, Giri B (2017) Arbuscular mycorrhizal fungi and tolerance of salt stress in plants. In: Wu QS (ed) Arbuscular mycorrhizas and stress tolerance of plants. Springer, Singapore, pp 67–97
- Scagel CF, Lee J (2012) Phenolic composition of basil plants is differentially altered by plant nutrient status and inoculation with mycorrhizal fungi. Hortic Sci 47:660–671
- Scherer HW (2001) Sulphur in crop production. Eur J Agron 14:81-111
- Scheublin TR, Sanders IR, Keel C, van der Meer JR (2010) Characterisation of microbial communities colonising the hyphal surfaces of arbuscular mycorrhizal fungi. ISME J 4:752–763
- Schreiner RP, Mihara KL, McDaniel H, Bethlenfalvay GJ (1997) Mycorrhizal fungi influence plant and soil functions and interactions. Plant Soil 188:199–209
- Schultz RC, Colletti JP, Isenhart TM, Simkins WW, Mize CW, Thompson ML (1995) Design and placement of a multi-species riparian buffer strip system. Agrofor Syst 29:1–16
- Schüßler A, Schwarzott D, Walker C (2001) A new fungal phylum, the Glomeromycota: phylogeny and evolution. Mycol Res 105:1413–1421
- Shabani L, Sabzalian MR, Mostafavi S (2016) Arbuscular mycorrhiza affects nickel translocation and expression of ABC transporter and metallothionein genes in *Festuca arundinacea*. Mycorrhiza 26:67–76
- Shakeel M, Yaseen T (2016) A review on exploring the weed suppressing characteristics of arbuscular mycorrhizal fungi for enhanced plant yield and productivity. Sci Technol Dev 35:54–62
- Shalaby AM, Hanna MM (1998) Preliminary studies on interactions between VA mycorrhizal fungus Glomus mosseae, Bradyrhizobium japonicum and Pseudomonas syringae in soybean plants. Acta Microbiol Pol 47:385–391
- Sharma S, Anand G, Singh N, Kapoor R (2017) Arbuscular mycorrhiza augments arsenic tolerance in wheat (*Triticum aestivum* L.) by strengthening antioxidant defense system and thiol metabolism. Front Plant Sci 8:906. https://doi.org/10.3389/fpls.2017.00906
- Sharma S, Prasad R, Varma A, Sharma AK (2017) Glycoprotein associated with *Funneliformis coronatum*, *Gigaspora margarita* and *Acaulospora scrobiculata* suppress the plant pathogens in vitro. Asian J Plant Pathol. https://doi.org/10.3923/ajppaj.2017
- Shen J, Yuan L, Zhang J, Li H, Bai Z, Chen X (2011) Phosphorus dynamics: from soil to plant. Plant Physiol 156:997–1005
- Silva VC, Alves PAC, de Oliveira RA, de Jesus RM, do Bomfim Costa LC, Gross E (2014) Influence of arbuscular mycorrhizal fungi on growth, mineral composition and production of essential oil in *Mentha* × *piperita* L. var. *citrata* (Ehrh.) Briq. under two phosphorus levels. J Med Plants Res 8:1321–1332
- Simard SW, Austin ME (2010) The role of mycorrhizas in forest soil stability with climate change. In: Simard S (ed) Climate change and variability. In Tech, Rijeka, Croatia, pp 275–302
- Smith SE, Read DJ (2008) Mycorrhizal symbiosis. Academic Press, New York
- Smith SE, Smith FA (2011) Roles of arbuscular mycorrhizas in plant nutrition and growth: new paradigms from cellular to ecosystem scales. Annu Rev Plant Biol 62:227–250

- Smith SE, Smith FA, Jakobsen I (2003) Mycorrhizal fungi can dominate phosphate supply to plants irrespective of growth responses. Plant Physiol 133:16–20
- Socolow RH (1999) Nitrogen management and the future of food: lessons from the management of energy and carbon. Proc Natl Acad Sci USA 96:6001–6008
- Solaiman ZM, Abbott LK, Varma A (eds) (2014) Mycorrhizal fungi: use in sustainable agriculture and land restoration. Springer, Berlin
- Srinivasan R, Govindasamy C (2014) Influence of native arbuscular mycorrhizal fungi on growth, nutrition and phytochemical constituents of *Catharanthus roseus* (L.) G. Don. J Coast Life Med 2:31–37
- Stanley MR, Koide RT, Shumway DL (1993) Mycorrhizal symbiosis increases growth, reproduction and recruitment of *Abutilon theophrasti* Medic. in the field. Oecologia 94:30–35
- Suharno, Soetarto ES, Sancayaningsih RP, Kasiamdari RS (2017) Association of arbuscular mycorrhizal fungi (AMF) with *Brachiaria precumbens* (Poaceae) in tailing and its potential to increase the growth of maize (*Zea mays*). Biodiversitas 18:433–441
- Sundaresan P, Raja NU, Gunasekaran P (1993) Induction and accumulation of phytoalexins in cowpea roots infected with the mycorrhizal fungus *Glomus fasciculatum* and their resistance to *Fusarium* wilt disease. J Biosci 18:291–301
- Tilman D (1996) Biodiversity: population versus ecosystem stability. Ecology 77:350-363
- Treseder KK (2016) Model behavior of arbuscular mycorrhizal fungi: predicting soil carbon dynamics under climate change. Botany 94:417–423
- Turnau K, Anielska T, Ryszka P, Gawronski S, Ostachowicz B, Jurkiewicz A (2008) Establishment of arbuscular mycorrhizal plants originating from xerothermic grasslands on heavy metal rich industrial wastes – new solution for waste revegetation. Plant Soil 305:267–280
- van der Heijden MGA, Klironomos JN, Ursic M, Moutoglis P, Streitwolf Engel R, Boller T, Sanders IR (1998) Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. Nature 396:69–72
- van der Heijden MGA, Martin FM, Selosse MA, Sanders IR (2015) Mycorrhizal ecology and evolution: the past, the present, and the future. New Phytol 205:1406–1423
- Veiga RSL, Jansa J, Frossard E, van der Heijden MGA (2011) Can arbuscular mycorrhizal fungi reduce the growth of agricultural weeds? PLoS One 6:e27825
- Veiga RSL, Howard K, van der Heijden MGA (2012) No evidence for allelopathic effects of arbuscular mycorrhizal fungi on the non-host plant Stellaria media. Plant Soil 360:319–331
- Verbruggen E, Veresoglou SD, Anderson IC, Caruso T, Hammer EC, Kohler J (2013) Arbuscular mycorrhizal fungi – short-term liability but long-term benefits for soil carbon storage? New Phytol 197:366–368
- Veresoglou SD, Chen BD, Rillig MC (2012a) Arbuscular mycorrhiza and soil nitrogen cycling. Soil Biol Biochem 46:53–62
- Veresoglou SD, Shaw LJ, Hooker JE, Sen R (2012b) Arbuscular mycorrhizal modulation of diazotrophic and denitrifying microbial communities in the (mycor)rhizosphere of *Plantago lanceolata*. Soil Biol Biochem 53:78–81
- Verzeaux J, Nivelle E, Roger D, Hirel B, Dubois F, Tetu T (2017) Spore density of arbuscular mycorrhizal fungi is fostered by six years of a no-till system and is correlated with environmental parameters in a silty loam soil. Agronomy 7(2):38
- Wagg C, Bender SF, Widmer F, van der Heijden MGA (2014) Soil biodiversity and soil community composition determine ecosystem multifunctionality. Proc Natl Acad Sci USA 111:5266–5270
- Walder F, Brulé D, Koegel S, Wiemken A, Boller T, Courty PE (2015) Plant phosphorus acquisition in a common mycorrhizal network: regulation of phosphate transporter genes of the Pht1 family in sorghum and flax. New Phytol 205:1632–1645
- Walder F, Boller T, Wiemken A, Courty PE (2016) Regulation of plants' phosphate uptake in common mycorrhizal networks: role of intraradical fungal phosphate transporters. Plant Signal Behav 11:e1131372
- Weber JG (2014) A decade of natural gas development: the makings of a resource curse? Resour Energy Econ 37:168–183

- Whipps JM (2004) Prospects and limitations for mycorrhizas in biocontrol of root pathogens. Can J Bot 82:1198–1227
- Whiteside MD, Garcia MO, Treseder KK (2012) Amino acid uptake in arbuscular mycorrhizal plants. PLoS One 7:e47643
- Whitmore A (2006) The emperors new clothes: sustainable mining? J Clean Prod 14:309-314
- Wilson GWT, Rice CW, Rillig MC, Springer A, Hartnett DC (2009) Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: results from long-term field experiments. Ecol Lett 12:452–461
- Wright SF, Anderson RL (2000) Aggregate stability and glomalin in alternative crop rotations for the central great plains. Biol Fertil Soils 31:249–253
- Wright SF, Upadhaya A (1996) Extraction of an abundant and unusual protein from soil and comparison with hyphal protein of arbuscular mycorrhizal fungi. Plant Soil 198:97–107
- Wright SF, Franke-Snyder M, Morton JB, Upadhyaya A (1996) Time-course study and partial characterization of a protein on hyphae of arbuscular mycorrhizal fungi during active colonization of roots. Plant Soil 181:193–203
- Wu QS, Xia RX (2006) Arbuscular mycorrhizal fungi influence growth, osmotic adjustment and photosynthesis of citrus under well-watered and water stress conditions. J Plant Physiol 163:417–425
- Wu Q-S, Cao M-Q, Zou YH, He XH (2014a) Direct and indirect effects of glomalin, mycorrhizal hyphae, and roots on aggregate stability in rhizosphere of trifoliate orange. Sci Rep 4:5823. https://doi.org/10.1038/srep05823
- Wu Z, McGrouther K, Huang J, Wu P, Wu W, Wang H (2014b) Decomposition and the contribution of glomalin-related soil protein (GRSP) in heavy metal sequestration: field experiment. Soil Biol Biochem 68:283–290
- Yang YR, Han XZ, Liang Y, Amit G, Chen J, Tang M (2015) The combined effects of arbuscular mycorrhizal fungi (AMF) and lead (Pb) stress on Pb accumulation, plant growth parameters, photosynthesis, and antioxidant enzymes in *Robinia pseudoacacia* L. PLoS One 10(12): e0145726. https://doi.org/10.1371/journal.pone.0145726
- Yang Y, Ou X, Yang G, Xia Y, Chen M, Guo L, Liu D (2017) Arbuscular mycorrhizal fungi regulate the growth and phyto-active compound of *Salvia miltiorrhiza* seedlings. Appl Sci 7:68
- Yao MK, Desilets H, Charles MT, Boulanger R, Tweddell RJ (2003) Effect of mycorrhization on the accumulation of rishitin and solavetivone in potato plantlets challenged with *Rhizoctonia solani*. Mycorrhiza 13:333–336
- Zhao X, Wang Y, Yan XF (2007) Effect of arbuscular mycorrhiza fungi and phosphorus on camptothecin content in *Camptotheca acuminata* seedlings. Allelopath J 20:51–60
- Zhu XQ, Wang CY, Chen H (2014) Effects of arbuscular mycorrhizal fungi on photosynthesis, carbon content, and calorific value of black locust seedlings. Photosynthetica 52:247–252
- Zobel M, Öpik M (2014) Plant and arbuscular mycorrhizal fungal (AMF) communities-which drives which? J Veg Sci 25:1133–1140
- Zolfaghari M, Nazeri V, Sefidkon F, Rejali F (2012) Effect of arbuscular mycorrhizal fungi on plant growth and essential oil content and composition of *Ocimum basilicum* L. Iranian. J Plant Physiol 3:643–650
- Zubek S, Mielcarek S, Turnau K (2012) Hypericin and pseudohypericin concentrations of a valuable medicinal plant *Hypericum perforatum* L. are enhanced by arbuscular mycorrhizal fungi. Mycorrhiza 22:149–156
- Zubek S, Rola K, Szewczyk A, Majewska ML, Turnau K (2015) Enhanced concentrations of elements and secondary metabolites in *Viola tricolor* L. induced by arbuscular mycorrhizal fungi. Plant Soil 390:129–142