# **Chapter 9 Resistance to Inhibitors of Angiogenesis**



Nili Dahan, Ksenia Magidey, and Yuval Shaked

**Abstract** Angiogenesis, a process that is predominantly driven by the vascular endothelial growth factor (VEGF) signaling pathway, plays an essential role in tumor progression and metastasis. Accordingly, a range of anti-angiogenic agents, most of which block VEGF or its receptor, have been approved for the treatment of various malignant diseases. However, the clinical benefits of anti-angiogenic therapy are relatively modest for several reasons, some of which are related to the development of therapy resistance. Since anti-angiogenic agents target the tumorsupporting vascular system rather than the tumor cells themselves, resistance is dependent on the interplay between the host- and tumor-mediated pathways. In general, the activation of various evasive mechanisms allows for sustained tumor vascularization and growth despite the therapeutic blockade of the drug target. These mechanisms include the upregulation of bypass angiogenic pathways, proangiogenic activity of infiltrating stromal cells and alternative vascularization processes. In addition, off-target effects of anti-angiogenic drugs have implications for tumor aggressiveness. In this chapter, we discuss the molecular and cellular mechanisms contributing to therapy resistance as well as possible strategies to improve the clinical outcome.

**Keywords** Chemokines • Chemokine receptors • Tumor microenvironment • Angiogenesis • Bone marrow-derived cells

# Abbreviations

- BMDC Bone marrow-derived cell
- CAF Cancer-associated fibroblast
- CRC Colorectal cancer

N. Dahan • K. Magidey • Y. Shaked (🖂)

© Springer International Publishing AG, part of Springer Nature 2018 Y. Yarden, M. Elkabets (eds.), *Resistance to Anti-Cancer Therapeutics Targeting* 

Receptor Tyrosine Kinases and Downstream Pathways, Resistance to Targeted Anti-Cancer Therapeutics 15, https://doi.org/10.1007/978-3-319-67932-7\_9

Department of Cell Biology and Cancer Science, Rappaport Faculty of Medicine, Technion—Israel Institute of Technology, 1 Efron St. Bat Galim, Haifa 31096, Israel e-mail: yshaked@tx.technion.ac.il

ECM	Extracellular matrix
EGF	Epidermal growth factor
EMT	Epithelial-mesenchymal transition
FDA	Food and Drug Administration
FGF	Fibroblast growth factor
GBM	Glioblastoma multiforme
G-CSF	Granulocyte colony stimulating factor
GIST	Gastrointestinal stromal tumor
HCC	Hepatocellular carcinoma
HGF	Hepatocyte growth factor
HIF-1	Hypoxia inducible factor-1
MDSC	Myeloid-derived suppressor cell
NSCLC	Non-small-cell lung cancer
OS	Overall survival
PDGF	Platelet-derived growth factor
PFS	Progression-free survival
PIGF	Placental growth factor
PNET	Pancreatic neuroendocrine tumor
RCC	Renal cell carcinoma
SCF	Stem cell factor
SDF-1α	Stromal derived factor-1α
TAM	Tumor-associated macrophage
TEM	Tie2-expressing monocyte
TH17	T helper type 17
TKI	Tyrosine kinase inhibitor
VEGF	Vascular endothelial growth factor
VEGFR	Vascular endothelial growth factor receptor

# 9.1 Introduction

Angiogenesis, the formation of new blood vessels from existing vasculature, plays an essential role in tumor progression and metastasis. The angiogenic process involves the activation, proliferation and migration of endothelial cells toward angiogenic stimuli produced by the tumor and supporting stromal cells within the tumor microenvironment. This ultimately results in the formation of new blood vessels that supply the growing tumor with nutrients and oxygen. This "angiogenic switch" is recognized as a rate-limiting event in tumor progression [1]. The concept of anti-angiogenic therapy was first proposed over four decades ago by Judah Folkman. He postulated that since the growth of all solid tumors is dependent on angiogenesis, inhibiting this process should suppress tumor growth [2]. It is now well-established that one of the most potent factors driving angiogenesis, and especially tumor angiogenesis, is the vascular endothelial growth factor-A (VEGF-A). Based on the key role of this factor in tumor angiogenesis, numerous therapies that block various components of the VEGF signaling pathway have been developed [3]. Several such therapies have been approved for the treatment of a variety of human cancers and there are more in preclinical and clinical trials. However, despite the potent activity of these agents and the high expectations for this therapeutic strategy, the clinical benefits are proving to be relatively mild. In the majority of patients, anti-angiogenic therapy achieves transient tumor control, with only a modest gain in long-term survival [4]. This can be explained by several mechanisms of resistance that allow the tumor to evade the therapeutic inhibition of angiogenesis. Here we discuss the molecular and cellular events underlying resistance in different tumor contexts, distinguishing between tumor- and host-mediated mechanisms.

# 9.2 Inhibitors of Angiogenesis: Mode of Action and Clinical Use

Although angiogenesis is a highly complex process, it is driven by one predominant key player, VEGF-A (hereafter, referred to as VEGF) in both physiological and pathological conditions. VEGF signals through its main receptor expressed on endothelial cells, VEGFR2, thereby coordinating the biological processes necessary for new vessel formation. These processes include: endothelial cell proliferation, migration, invasion and survival; chemotaxis and homing of bone-marrow-derived endothelial precursor cells; vascular permeability; and vasodilation [3, 5]. Whereas autocrine VEGF, released by endothelial cells, maintains vascular homeostasis [6], paracrine VEGF, released by both tumor cells and stromal myeloid cell types, increases vessel branching resulting in abnormal, tortuous vasculature [7]. VEGF is upregulated in most solid tumors. Furthermore, slight increases in tumor VEGF levels are sufficient to promote angiogenesis and tumor growth. Accordingly, it was proposed that neutralizing circulating VEGF would suppress tumor growth, as demonstrated by a number of cancer models in mice [3, 8].

Since 2004, several drugs that target VEGF or its receptor have been approved by the Food and Drug Administration (FDA) for the treatment of various malignant diseases (Table 9.1), and there are more in clinical trials. These drugs include neutralizing antibodies against VEGF and VEGFRs, soluble VEGF receptor hybrids (VEGF traps) and tyrosine kinase inhibitors (TKIs) with selectivity for VEGFRs. It should be noted that, due to their mode of action at the ATP-binding pocket, TKIs designed to target VEGFRs may also significantly inhibit other kinases. Nevertheless, their potent anti-angiogenic activity has been demonstrated in preclinical studies [3]. Bevacizumab, a monoclonal antibody against VEGF, was the first anti-angiogenic drug to be approved by the FDA. It is currently used as first-line therapy in metastatic colorectal cancer (CRC), non-small-cell lung cancer (NSCLC) and renal cell carcinoma (RCC), as second-line therapy in CRC and glioblastoma multiforme (GBM), and as maintenance therapy in advanced ovarian cancer (Table 9.1).

		Ref.	[139]	[140]	[141]	[142]	[143, 144]	[145, 146]	[147, 148]	[149, 150]	[151]	[152]	[153]	[154]	[155]	[156, 157]	[158]	[159, 160]	[161]	[162]	[163, 164]
	Improvement in OS	(months)	4.7	2.1	1.4	2	SN	NS	2	NS/4.8 <sup>a</sup>	NS		2.2	1.4	1.4	SN	2.8	4.6	NA	NA	SN
	Improvement in PFS	(months)	4.4	2.6	1.4	1.7	0.4-0.6	3.3	4.8	NS	3.8	eported	1.5	0.8	2.2	2.7	NS	9	20.9	5.9	5
	Improvement in	RR (%)	10	14.1	0	20	10.3–14	12.4	18	19	NA	Only phase II data re	12	0.8	8.7	8	1	25	NA	9.3	27
0	Treatment	combination	Chemotherapy			Chemotherapy		IFNα		Chemotherapy		Monotherapy	Chemotherapy	Monotherapy	Chemotherapy	Monotherapy	Monotherapy	Monotherapy	Monotherapy	Monotherapy	Monotherapy
C 0		Approved use	Metastatic CRC			Metastatic NSCLC		Metastatic RCC		Advanced ovarian cancer		GBM	Metastatic GEJ		Metastatic CRC	Metastatic RCC	Metastatic HCC	Metastatic RCC	Metastatic GIST	PNET	Metastatic RCC
J.,		Drug class	VEGF-A	antibody									VEGFR2	antibody	VEGF trap	TKI		TKI			TKI
		Drug	Bevacizumab										Ramucirumab		Aflibercept	Sorafenib		Sunitinib			Pazopanib

 Table 9.1
 Clinical benefits of approved anti-angiogenic drugs

			Treatment	Improvement in	Improvement in PFS	Improvement in OS	
Drug	Drug class	Approved use	combination	RR (%)	(months)	(months)	Ref.
Axitinib	TKI	Advanced RCC	Monotherapy	NA	2.6	NS	[165] <sup>b</sup>
Regorafenib		Metastatic CRC	Monotherapy	0.6	0.2	1.4	[166]
Nintedanib	TKI	Advanced NSCLC	Chemotherapy	NA	0.7	1	[167] <sup>c</sup>
Vandetanib	TKI	Advanced medullary thyroid cancer	Monotherapy	43	6.2	NA	[168]
Cabozantinib	TKI	Advanced medullary thyroid cancer	Monotherapy	28	7.2	NS	[169]
		U.1	-	•			

CRC colorectal cancer, GBM glioblastoma multiforme, GEJ gastric and gastroesophageal junction cancer, GIST gastrointestinal stromal cancer, HCC hepatocellular carcinoma, NA not available, NS not significant, NSCLC non-small-cell lung cancer, OS overall survival, PFS progression-free survival, PNET pancreatic neuroendocrine tumors, RCC renal cell carcinoma, RR response rate, TKI tyrosine kinase inhibitor

<sup>a</sup>In poor-prognosis patients

<sup>b</sup>Axitinib vs. sorafenib

<sup>c</sup>Approved only in Europe

Of note, bevacizumab generally failed to provide significant benefits when used as monotherapy. However, with the exception of GBM, it has been approved for use as combination therapy for the treatment of the above-mentioned advanced-stage cancers [9]. The TKIs, sorafenib, sunitinib, pazopanib and axitinib are approved as monotherapies for the treatment of metastatic RCC, a highly vascularized tumor type. In addition, sunitinib is approved for gastrointestinal stromal tumors (GIST) and pancreatic neuroendocrine tumors (PNET), and sorafenib for hepatocellular carcinoma (HCC), for advanced-stage disease in all cases (Table 9.1). Other antiangiogenic therapies approved for late-stage, metastatic disease are described in Table 9.1. These include: ramucirumab, a VEGFR2 monoclonal antibody; aflibercept, a VEGF-trap that binds 3 VEGF family ligands; and other VEGFR TKIs. Antiangiogenic agents have also been evaluated for early-stage disease, specifically in the adjuvant setting, when treatment is administered after surgical removal of the primary tumor. It has been postulated that inhibiting angiogenesis after tumor resection would prevent local relapse or growth of micrometastases [10]. However, two large phase III post-operative adjuvant trials of bevacizumab in combination with chemotherapy in patients with early-stage CRC failed to provide significant benefits when compared to treatment with chemotherapy alone [11-13]. The use of antiangiogenic therapy in the neo-adjuvant setting in order to downsize or downstage a tumor before resection has also been evaluated. However, two large trials testing the efficacy of neoadjuvant bevacizumab in combination with chemotherapy in comparison to neoadjuvant chemotherapy alone in patients with primary breast cancer revealed conflicting findings in terms of long-term benefits [14, 15]. The diverse outcomes of anti-angiogenic therapy in different clinical scenarios highlight the effects of specific parameters, such as disease stage and cancer type, on therapy efficacy. However, our understanding of the underlying mechanisms is still incomplete.

In general, protein-based anti-angiogenic drugs, such as bevacizumab and aflibercept, have only shown significant activity when combined with cytotoxic chemotherapy, whereas TKIs are effective when used as monotherapy, without an additive effect when combined with chemotherapy [10]. Conceivably, in cases where single-agent activity is observed, such as in RCC, therapy-induced vessel regression is the major mechanism of action contributing to the efficacy of therapy. In cases where anti-angiogenic therapies only show activity when combined with cytotoxic chemotherapy, such as in CRC, mechanisms other than vessel regression may play a role [3]. A widely-held view is that anti-angiogenic therapy improves the delivery of co-administered chemotherapy through a process known as "vascular normalization". This is based on the principle that the abnormal tumor vasculature, which is known to be dysfunctional, leaky and tortuous, can be "normalized" by suppressing VEGF signaling. The resulting improvement in vessel function and blood flow is presumed to increase delivery of cytotoxic agents [16]. An alternative possibility explaining the benefit of combined therapy is that anti-angiogenic agents block the activity of bone marrow-derived endothelial progenitor cells that have been shown to infiltrate tumors in response to chemotherapy drugs [8, 17–19]. However, given that the clinical relevance of such phenomena is dependent on cancer type and drug class, additional unknown mechanisms likely play a role [4]. A recent study suggests that the vessel phenotype of tumors contributes to the response to different treatment strategies. Using preclinical models and clinical samples, it was shown that cancers that are more responsive to bevacizumab in combination with chemotherapy have a stromal-vessel phenotype, where the vessels are surrounded by a well-developed stroma. In contrast, cancers that are more responsive to TKI monotherapy have a tumor-vessel phenotype, where the vessels are in close proximity to the tumor cells [20]. In addition, tumor-specific differences likely account for why certain anti-angiogenic therapies show efficacy in some cancers, but not in others, although the precise molecular mechanisms are not known [10].

Although anti-angiogenic therapy has been incorporated into the standard protocol for certain cancer types, there are a number of concerns, the foremost being its modest clinical benefits. The gain in progression-free survival (PFS) and overall survival (OS) is generally in the order of months. In addition, initial response rates and gains in PFS do not always translate into significant improvements in OS (Table 9.1). These limited clinical benefits strongly suggest that tumors treated with anti-angiogenic agents develop resistance to therapy. Such resistance can be classified as intrinsic, where tumors are unresponsive from the beginning of treatment, and acquired, where tumors initially respond but then progress during the course of treatment [21]. Thus, there is an urgent need to overcome these limitations and to develop improved strategies for the treatment of cancer at all stages of the disease.

#### 9.3 Mechanisms of Resistance to Inhibitors of Angiogenesis

There is a growing interest in understanding the mechanisms underlying both acquired and intrinsic resistance to anti-angiogenic therapy. Classical drug resistance mechanisms involve the clonal selection of tumor cells harboring genomic mutations that either alter the drug target or affect drug uptake or efflux [22]. However, since anti-angiogenic therapy targets the vascular supply of the tumor mass rather than the tumor cells themselves, resistance in this case is mainly indirect and involves an interplay between tumoral cues and host-mediated pathways. In addition, given that endothelial cells are more genetically stable than tumor cells, they are less likely to acquire mutations after exposure to such drugs [23]. In general, resistance to anti-angiogenic therapy is manifested by the activation of alternative mechanisms that sustain tumor vascularization and growth while the specific target of the drug remains inhibited [21]. These evasive mechanisms are described in detail below. A graphical summary is shown in Fig. 9.1.



**Fig. 9.1** Mechanisms of resistance to anti-angiogenic therapy. Tumors develop resistance to antiangiogenic therapy via a range of tumor- and host-mediated processes. These evasive mechanisms sustain tumor vascularization and/or progression despite the blockade of VEGF signaling imposed by anti-angiogenic agents. Increased tumor hypoxia, which occurs as a direct result of antiangiogenic therapy, drives many of these processes. The processes are not mutually exclusive, and some are interdependent (indicated by thin arrows). BMDC, bone marrow-derived cell; EMT, epithelial-mesenchymal transition

# 9.3.1 Upregulation of Alternative Angiogenic Pathways

Tumor angiogenesis is mainly driven by the VEGF signaling pathway. However, there are also numerous complementary non-VEGF pathways that contribute to blood vessel formation. Tumor hypoxia, which occurs as a direct result of antiangiogenic therapy, modulates the interplay between these various angiogenic pathways via the master regulator, hypoxia inducible factor-1 (HIF-1), a transcription factor that regulates the expression of multiple pro-angiogenic genes [24]. The activation of alternative or compensatory angiogenic pathways allows for persistent neovascularization despite VEGF inhibition and represents the most common means by which tumors evade the blockade of angiogenesis. Preclinical trials in a murine pancreatic cancer model demonstrated an initial response to anti-VEGFR2 therapy (DC101) followed by restoration of tumor growth and vascularization shortly after initiation of therapy. Interestingly, at the time of progression, these tumors expressed higher levels of various pro-angiogenic factors such as fibroblast growth factor (FGF) 1 and 2, ephrin A1 and A2 and angiopoietin 1. Similarly, tumor cells subjected to hypoxic conditions upregulated most of these genes. Blocking both VEGF and FGF signaling attenuated revascularization and slowed tumor growth, suggesting that upregulation of FGF signaling contributes to anti-angiogenic therapy resistance [25]. Several additional pro-angiogenic factors have been implicated in

resistance to anti-angiogenic therapy in various murine tumor models. These include: placental growth factor, PIGF [26]; platelet-derived growth factor, PDGF [27]; hepatocyte growth factor, HGF, and its receptor, c-Met [28, 29]; epidermal growth factor, EGF [30]; interleukin-8, IL-8 [31]; granulocyte colony stimulating factor, G-CSF, and Bv8 [32], among others (recently reviewed in [33]). Similar to the seminal study of Casanovas et al. [25], the above-mentioned studies report elevated levels of the specific factor in resistant tumors and that dual inhibition of the VEGF pathway and the specific factor or its pathway enhances therapeutic outcome. It should be noted that these upregulated pro-angiogenic factors may be derived from tumor cells or host stromal cells residing within the tumor microenvironment. The former case involves a direct response of tumor cells to hypoxia. In the latter case, stromal cells may be responding to cues from the tumor, environmental signals or systemic effects of the drug [10, 34].

There is a wealth of clinical evidence showing that circulating levels of proangiogenic factors are elevated just prior to disease progression or during the relapse phase in cancer patients treated with angiogenesis inhibitors suggesting that these factors contribute to the development of acquired resistance [35–40]. There are also cases in which patients do not respond at all to anti-angiogenic therapy suggestive of intrinsic resistance. In late stage malignancies, pre-existing upregulation of alternative pro-angiogenic pathways may compensate for the inhibition of VEGF signaling [21].

# 9.3.2 Pro-angiogenic Effects of Local and Bone Marrow-Derived Stromal Cells

The release of pro-angiogenic factors in response to anti-angiogenic therapy activates local stromal cells and stimulates the recruitment of bone marrow-derived cells (BMDCs) to the tumor environment. BMDCs include vascular progenitors, which differentiate into cells that make up physical components of the blood vessel walls, and pro-inflammatory monocytes, which produce a diverse assortment of soluble factors that regulate vascular cell survival, proliferation and motility as well as extracellular matrix (ECM) remodeling [21, 41].

The effect of hypoxia on BMDC recruitment was described by Du et al. in an orthotopic model of GBM. They demonstrated that HIF-1 $\alpha$ , the direct effector of hypoxia, promotes angiogenesis and tumor growth by inducing an influx of various pro-angiogenic bone-marrow derived CD45<sup>+</sup> myeloid cells as well as endothelial and pericyte progenitor cells to the tumor [42]. In addition, treating tumor-bearing mice with vascular-disrupting agents, which cause massive tumor hypoxia, triggers an acute mobilization of circulating endothelial progenitor cells that home to the tumor margins in sufficient numbers to facilitate revascularization [17]. Thus, therapy-induced hypoxia represents a major contributing factor to resistance via the action of recruited BMDCs.

The involvement of specific BMDC types and local stromal cells in resistance to anti-angiogenic therapy is described below:

Immature myeloid cells, also known as CD11b<sup>+</sup>Gr1<sup>+</sup>myeloid-derived suppressor cells (MDSCs), produce a variety of factors that influence endothelial cell behavior resulting in new vessel formation [41]. Shojaei et al. demonstrated that tumors resistant to anti-VEGF therapy exhibit higher levels of infiltrating MDSCs in comparison to therapy-sensitive tumors [43]. This is due to an upregulation of G-CSF and Bv8 [32], factors that promote the mobilization of MDSCs from the bone marrow and their infiltration to tumor tissue [44]. In pancreatic tumor models that are resistant to anti-VEGF therapy, increased levels of proinflammatory factors including several CXCR2 ligands, IL-1a, IL-1b and Angptl-2 stimulate the recruitment of CD11b<sup>+</sup> myeloid cells to the tumor environment [45]. In agreement with the above-mentioned studies, blocking chemotherapy-induced infiltration of MDSCs to tumors using Bv8 neutralizing antibodies enhances therapy outcome in mouse models of pancreatic cancer [46]. In another study, it was suggested that tumor-infiltrating T helper type 17 (TH17) cells and IL-17 induce the recruitment of immature myeloid cells to the tumor microenvironment. Blocking TH17 cell function renders resistant tumors sensitive to anti-VEGF therapy [47].

**Tumor-associated macrophages** (TAMs) are recruited to tumors as monocytes from the circulation and, as they extravasate across the tumor vasculature, they differentiate into macrophages. In the tumor environment, TAMs are predominantly polarized towards an M2-like phenotype underlying their ability to promote tumor growth and angiogenesis [48]. TAMs promote angiogenesis mostly through their production of VEGF [41]. However, TAM-derived PIGF can also stimulate angiogenesis in some tumors, representing a possible mechanism for acquired resistance to VEGF/VEGFR-targeted therapies [49]. In HCC xenografts, sorafenib increases CXCL12 levels and TAM infiltration. Furthermore, depletion of TAMs enhances the inhibitory effect of therapy on tumor angiogenesis, growth and metastasis demonstrating the contribution of TAMs to therapy resistance [50].

**Tie2-expressing monocytes** (TEMs) represent a distinct subpopulation of TAMs expressing the angiopoietin receptor, Tie2. They physically associate with vessels and secrete growth factors and matrix-remodeling proteins that stimulate the angiogenic process in a paracrine manner [51]. TEMs are recruited and activated via endothelial cell- and tumor-secreted chemoattractants, Ang2 and CXCL12, respectively [52, 53]. Their recruitment to spontaneously growing tumors promotes angiogenesis [54]. Furthermore, TEMs infiltrate hypoxic tumors treated with a vascular-disrupting agent, and inhibiting such infiltration enhances treatment efficacy [53]. Lastly, dual targeting of VEGF and Ang2 has been shown to delay tumor growth and improve the outcome of anti-angiogenic therapy in preclinical studies [55–57]. These collective findings highlight the possible contribution of TEMs to resistance to anti-angiogenic therapy.

**Pericytes**, the periendothelial support cells of the microvasculature, are derived from local or bone marrow-derived mesenchymal stem cells. They provide important support for blood vessel formation, structure and function. Furthermore, tight cross-talk between pericytes and endothelial cells maintains blood vessel integrity [58]. While anti-angiogenic therapy reduces tumor vascularity, the vessels that remain are functional, distinctively thin and tightly covered with pericytes [21, 59, 60]. Owing to their important role in maintaining vessel integrity, the remaining pericytes, along with basement membrane-associated cells, facilitate a rapid regrowth of blood vessels after cessation of treatment with angiogenesis inhibitors [61]. Importantly, pericytes mediate endothelial cell quiescence and survival and therefore their presence presumably reduces responsiveness to anti-angiogenic therapy [21]. The underlying molecular mechanism involves pericyte-induced survival signals that induce an autocrine activation loop of VEGF signaling and antiapoptotic Bcl-w expression in tumor endothelial cells [62]. Additional pericytederived endothelial survival signals, specifically via the Ang1/Tie2 and EGF pathways, may also contribute to anti-angiogenic therapy resistance [30, 63]. Accordingly, it has been suggested that targeting both endothelial cells and pericytes may improve the efficacy of anti-angiogenic therapy. Indeed, such dual targeting improves therapy outcome in a variety of murine tumor models [63–65]. However, severe reduction in pericyte coverage damages the integrity of the vasculature, enabling local intravasation of tumor cells thereby facilitating metastasis [66]. In support of this concept, a recent study demonstrated that TKI-induced pericyte depletion enhances metastasis due to increased vessel leakiness and hypoxiaassociated epithelial-mesenchymal transition (EMT) [67]. Collectively, enhanced as well as reduced pericyte coverage contribute to anti-angiogenic therapy resistance via different mechanisms.

**Cancer-associated fibroblasts** (CAFs) are tumor-localized, activated fibroblasts originating from connective tissue fibroblasts proximal to neoplasms or from local and bone marrow-derived mesenchymal stem/progenitor cells. They promote angiogenesis by producing a variety of pro-angiogenic signaling factors, chemoattractants and ECM-degrading enzymes [41]. Crawford et al. showed that the upregulation of PDGF-C in CAFs from anti-VEGF resistant tumors compensates for the inhibition of VEGF-dependent angiogenesis. Furthermore, CAFs isolated from resistant tumors can stimulate the growth of therapy-sensitive tumors even when VEGF is inhibited. This suggests that, once activated by the tumor environment, CAFs retain their ability to induce angiogenesis independent of tumor cells [27].

### 9.3.3 Alternative Vascularization Mechanisms

Primary tumors and metastases may gain access to a blood supply via mechanisms that are independent of classical sprouting angiogenesis. These alternative vascularization mechanisms are not affected by antiangiogenic drugs and therefore represent another mode of resistance to such therapy [68].

**Vessel co-option** refers to the migration of tumor cells along existing and established blood vessels in the host organ to invade healthy tissue. This process is mostly observed in highly vascularized tissues such as brain, lungs and liver, where tumor cells can co-opt the abundant pre-existing blood vessels [69]. Preclinical and clinical data show that glioblastomas become more infiltrative with the use of antiangiogenic therapy, facilitating vessel co-option [42, 70–74]. In addition, vessel co-option has been implicated in resistance to anti-angiogenic therapy in HCC [75] and metastases in lymph nodes [76], brain [77], liver [78] and lung [79].

**Vasculogenic mimicry** is a mechanism by which highly aggressive tumor cells form vessel-like structures in an angiogenesis-independent manner. These vessellike structures may connect to the endothelial-lined vasculature to provide a perfusion pathway for the transport of fluid, nutrients and oxygen to the core of the malignant mass [68, 80]. Since its first description in uveal melanoma [81], vasculogenic mimicry has been observed in several tumor types and is associated with poor prognosis [82]. By virtue of their plasticity, tumor cells can dedifferentiate and acquire expression of vascular markers thereby "mimicking" endothelial cells during this process [81, 83, 84]. However, despite expression of various vascular markers, such tumor cells are resilient to treatment with angiogenesis inhibitors [84–87]. Furthermore, antiangiogenic treatment has been shown to induce vasculogenic mimicry in preclinical models of various cancers [86, 87]. This may be due to treatment-induced hypoxia that upregulates vasculogenic mimicry pathways in tumor cells [88, 89]. Collectively, anti-angiogenic therapy not only triggers alternative vascularization mechanisms, but may also select for more aggressive tumor cells with an intrinsic ability to evade the blockade of angiogenesis.

# 9.3.4 The Host Response to Inhibitors of Angiogenesis: Implications for Tumor Aggressiveness

Targeting the host-mediated angiogenic process that supports tumor growth has its limitations. As detailed in the previous sections, anti-angiogenic therapies may trigger an array of evasive mechanisms that involve the activity of host cells such as pro-inflammatory myeloid cells and endothelial progenitor cells in the tumor microenvironment. Furthermore, anti-angiogenic therapy has been shown to augment the invasive and metastatic potential of tumors despite overall inhibition of tumor growth [90, 91]. This seemingly paradoxical phenomenon is proposed to arise, at least in part, from a direct response of the host to anti-angiogenic therapy, independent of the tumor. Ebos et al. showed that short-term sunitinib treatment of mice prior to intravenous injection of tumor cells accelerates metastasis and reduces survival. Similarly, adjuvant short-term sunitinib treatment after resection of the primary tumor enhances spontaneous metastatic tumor burden [90]. The mechanisms underlying this effect may involve a drug-induced change in the levels of circulating factors implicated in tumor progression. For example, healthy, tumor-free mice treated with VEGF receptor TKIs exhibit a dose-dependent increase in the levels of circulating G-CSF, SDF-1a, SCF and osteopontin demonstrating a systemic tumor-independent response to therapy [92]. Similarly, cancer patients treated with sunitinib exhibit increased circulating levels of pro-angiogenic factors [93, 94]. In theory, such systemic host-mediated responses could promote the formation of "pre-metastatic niches" in distant organs, thereby facilitating metastasis [95]. The deleterious effect of anti-angiogenic therapy on the host vasculature represents another factor that may explain increased metastasis in response to such therapy. The systemic action of VEGF receptor TKIs may damage the integrity of the vasculature by targeting endothelial cells as well as pericytes. This facilitates local intravasation of invasive tumor cells and creates permissive niches for extravasation of tumor cells in target organs [67, 96, 97].

It should be emphasized that several steps are required for disease progression from a local primary tumor to established metastatic disease. These include loss of cellular adhesion, increased motility, intravasation, survival in the bloodstream, homing, extravasation, seeding of micrometastases, and finally colonization and growth at a distant site [98]. Therefore, it is conceivable that the above-mentioned host-mediated responses act in concert with tumor-derived effects to promote overall tumor aggressiveness in response to anti-angiogenic therapy. Paez-Ribes et al. demonstrated that the anti-VEGFR2 antibody, DC101, and VEGF receptor TKI, sunitinib, promote local primary tumor invasion and metastasis in mouse models of pancreatic neuroendocrine carcinoma and glioblastoma. The researchers suggest that therapy-induced hypoxia in the primary tumor triggers a switch to a hyperinvasive condition in tumor cells [91]. In agreement with this, several preclinical studies demonstrate that VEGF-targeted therapies cause tumor cells to undergo hypoxiaassociated EMT, thereby promoting invasion and metastasis [67, 99, 100]. Collectively, both host- and tumor-dependent responses to anti-angiogenic therapy contribute to the invasive and metastatic potential of treated tumors.

Whether anti-angiogenic therapy causes increased tumor aggressiveness in patients is still a debatable issue. A retrospective analysis found no evidence for accelerated tumor growth in metastatic RCC patients treated with sunitinib [101]. Similarly, a meta-analysis of several randomized phase III trials of bevacizumab found no evidence for accelerated disease progression after discontinuation of therapy in patients with metastatic renal, pancreatic, breast and colorectal cancer [102]. On the other hand, rapid tumor regrowth has been reported after treatment discontinuation in RCC patients receiving sunitinib or sorafenib [103, 104], and in CRC patients receiving bevacizumab and chemotherapy [105]. In addition, several clinical studies describe an increased infiltrative growth pattern of glioblastomas in response to anti-angiogenic therapy [70, 71, 73]. The differences in preclinical and clinical findings may be explained by the animal model used, tumor type, disease stage, drug type, dosage, duration of treatment, or combination with chemotherapy [10].

Other anti-cancer treatment modalities, such as chemotherapy, radiation and surgery, can also produce undesirable pro-angiogenic and pro-metastatic effects that arise from the response of the host to therapy. Accordingly, blunting this host response using combinatorial therapies may improve treatment outcomes [34]. For example, the elevation in circulating endothelial progenitor cell levels following treatment with chemotherapeutic or vascular-disrupting agents can be blocked using anti-VEGF or anti-VEGFR2 neutralizing antibodies. This combinatorial treatment enhances therapy efficacy and delays tumor regrowth in comparison to cytotoxic therapy alone [18]. Recent preclinical studies suggest that the reverse may be true as well; cytotoxic therapy can be used to blunt tumor aggressiveness induced by anti-angiogenic drugs thereby improving treatment efficacy. For example, concurrent paclitaxel chemotherapy was shown to block the increase in primary tumor local invasion and distant metastases induced by anti-VEGFR2 antibody (DC101) therapy in mouse models of breast cancer [106]. In addition, co-administration of chemotherapy counteracted the sunitinib-induced increase in metastasis in mice bearing early stage Lewis lung carcinoma [107]. Thus, add-on therapy that counteracts host- or tumor-dependent responses represents a possible strategy to overcome increased tumor aggressiveness and resistance in response to anti-angiogenic therapy.

#### 9.4 Future Directions

The limited clinical benefits of anti-angiogenic therapy contrast with findings of preclinical studies conducted over the last two decades that demonstrate treatment efficacy. This can be explained by the disparity between preclinical models used to test efficacy and clinical scenarios. Due in part to ethical issues, patients enrolled in clinical trials are generally at an advanced stage of the disease. In contrast, preclinical experimental setups mostly involve localized primary tumors, with suppression of tumor growth after a short-term drug exposure considered a sign of efficacy. Therefore, more relevant preclinical models should be used to study the effects of anti-angiogenic therapy at all stages of disease, including metastatic and adjuvant settings, with clinically-relevant endpoints [108].

In theory, alternative pro-angiogenic pathways upregulated in response to antiangiogenic therapy may be targeted as a strategy to overcome resistance. Multitargeted inhibitors such as brivanib, a dual VEGFR and FGFR TKI, and nintedanib, a triple angiokinase inhibitor for VEGFR, FGFR and PDGFR, are being tested in clinical trials [109, 110]. Importantly, host-mediated evasive mechanisms induced in response to anti-angiogenic therapy may also be targeted in order to improve anti-angiogenic therapy outcomes. The major BMDC recruiting factor, SDF1a (CXCL12), represents a potential target for cancer therapy. Recent preclinical and clinical data support the use of anti-CXCL12 agents to reduce BMDC infiltration as a potential strategy to overcome resistance to anti-angiogenic therapy [18, 111]. Macrophages are key regulators in the tumor microenvironment, and have been implicated in resistance to anti-angiogenic therapy. Therefore, specifically blocking macrophage infiltration is also a potential means for overcoming resistance. Antibodies against the monocyte chemotactic protein, CCL2, and the macrophageexpressed CSF-1 receptor are being tested in clinical trials as monotherapies [112-114]. It will be interesting to test whether such agents synergistically increase efficacy when combined with anti-angiogenic agents in the clinical setting [115].

As detailed throughout this review, hypoxia resulting from anti-angiogenic therapy drives tumor aggressiveness and therapy resistance via tumor- and hostmediated mechanisms. Therefore, alleviating hypoxia or targeting HIF-1 represent avenues for future investigation [116]. The former case would involve optimizing the dosage and scheduling of anti-angiogenic agents with the aim of normalizing the abnormal tumor vasculature as opposed to inducing rapid and excessive vessel pruning [16]. Indeed, tumor perfusion and oxygenation correlates with clinical benefit in GBM patients treated with anti-angiogenic therapy [117–120]. Alleviating hypoxia would reduce processes such as EMT, vasculogenic mimicry and the selection of more aggressive tumor cells as well as affect immune and stromal cells within the tumor microenvironment. It is well-established that a hypoxic tumor environment induces BMDC recruitment and reprograms TAMs towards a protumorigenic phenotype. Therefore, alleviating hypoxia through vascular normalization could potentially reprogram the entire tumor microenvironment [16]. Histone deacetylase inhibitors have been shown to strongly repress HIF-1 expression and their use as anti-cancer drugs is currently being explored [121]. A recent phase I clinical trial evaluating the use of a histone deacetylation inhibitor in combination with the anti-angiogenic agent, pazopanib, demonstrated durable tumor regression in 70% of patients with pazopanib-refractory disease [122]. Thus, epigenetic targeting represents a potential strategy to reverse resistance to anti-angiogenic therapy, possibly by targeting HIF-1. The precise molecular mechanisms and clinical benefits should be further characterized.

The combination of anti-angiogenic drugs with immunotherapy represents an emerging strategy for cancer treatment. The rationale for using this combination is based on the systemic influence of VEGF on immune cell function. Specifically, several studies have demonstrated that an elevated level of circulating VEGF in tumor-bearing hosts impedes immune surveillance and destruction of tumor cells [123–125]. Accordingly, anti-angiogenic drugs may be used to neutralize the immunosuppressive activity of VEGF. Moreover, the combination of anti-angiogenic therapy with immunotherapy could potentially offer a synergistic anti-cancer effect. In addition, it has been proposed that alleviating tumor hypoxia via vascular normalization would reprogram the phenotype of the tumor microenvironment from immunosuppressive to immunosupportive, thereby improving the efficacy of anti-cancer immunotherapies [16, 126]. A number of preclinical and clinical studies have demonstrated the benefits of this combination strategy [127–133].

Lastly, a major challenge is to identify robust biomarkers predictive of clinical efficacy of anti-angiogenic therapy. Currently, no validated biomarkers exist to select patients who will benefit from such therapy. Biomarkers under consideration in various cancers include circulating VEGF-A, VEGF-D, Ang2, HGF, osteopontin, IL6 and IL8, among others [4, 134]. With respect to VEGF as a predictive biomarker for bevacizumab-based treatment benefit, phase III trials have reported a correlation between high circulating levels of VEGF and survival benefit in metastatic breast and gastric cancer patients [135, 136], but not in CRC, RCC and lung cancer patients [137]. Other emerging areas for biomarker identification include tumor vessel imaging with dynamic contrast-enhanced MRI, measurement of circulating endothelial

cells, expression arrays, single nucleotide polymorphisms and early pharmacodynamic response to treatment, such as hypertension [138]. The incorporation of predictive biomarkers into routine clinical practice would maximize clinical benefit, reduce unnecessary toxicity and improve costs of cancer care.

## 9.5 Conclusions

The development of anti-angiogenic agents is an important milestone in the field of cancer research. However, their clinical use is proving to be more complex than originally anticipated with major ongoing challenges. A prominent issue in the clinic is resistance to therapy resulting in only modest gains in long-term survival in the majority of patients. Given that anti-angiogenic agents target the tumor-supporting vascular system comprised of a variety of host cells, and that tumor progression is regulated by tumor-host cell cross-talk, resistance is dependent on both tumor- and host-mediated mechanisms (Fig. 9.1). Understanding these mechanisms is key to developing strategies to overcome therapy resistance and improve clinical outcome.

**Acknowledgments** This book chapter is primarily supported by the European Research Council (#260633) and Rappaport Institute funds given to YS. KM is supported by a student fellowship from the Lyon Sachs University of Toronto— Technion Collaboration Fund.

Conflict of Interest The authors declare that they have no conflict of interest.

#### References

- 1. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144:646-74.
- 2. Folkman J. Tumor angiogenesis: therapeutic implications. N Engl J Med. 1971;285:1182-6.
- Ellis LM, Hicklin DJ. VEGF-targeted therapy: mechanisms of anti-tumour activity. Nat Rev Cancer. 2008;8:579–91.
- 4. Vasudev NS, Reynolds AR. Anti-angiogenic therapy for cancer: current progress, unresolved questions and future directions. Angiogenesis. 2014;17:471–94.
- Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. Nature. 2011;473:298–307.
- Lee S, Chen TT, Barber CL, Jordan MC, Murdock J, Desai S, Ferrara N, Nagy A, Roos KP, Iruela-Arispe ML. Autocrine VEGF signaling is required for vascular homeostasis. Cell. 2007;130:691–703.
- Stockmann C, Doedens A, Weidemann A, Zhang N, Takeda N, Greenberg JI, Cheresh DA, Johnson RS. Deletion of vascular endothelial growth factor in myeloid cells accelerates tumorigenesis. Nature. 2008;456:814–8.
- 8. Kerbel RS. Tumor angiogenesis. N Engl J Med. 2008;358:2039-49.
- Boere IA, Hamberg P, Sleijfer S. It takes two to Tango: combinations of conventional cytotoxics with compounds targeting the vascular endothelial growth factor-vascular endothelial growth factor receptor pathway in patients with solid malignancies. Cancer Sci. 2010;101:7–15.

- 9 Resistance to Inhibitors of Angiogenesis
  - 10. Ebos JM, Kerbel RS. Antiangiogenic therapy: impact on invasion, disease progression, and metastasis. Nat Rev Clin Oncol. 2011;8:210–21.
  - Allegra CJ, Yothers G, O'Connell MJ, Sharif S, Petrelli NJ, Colangelo LH, Atkins JN, Seay TE, Fehrenbacher L, Goldberg RM, O'Reilly S, Chu L, Azar CA, Lopa S, Wolmark N. Phase III Trial assessing bevacizumab in stages II and III carcinoma of the colon: results of Nsabp protocol C-08. J Clin Oncol. 2011;29:11–6.
  - Allegra CJ, Yothers G, O'Connell MJ, Sharif S, Petrelli NJ, Lopa SH, Wolmark N. Bevacizumab in stage II–III colon cancer: 5-year update of the national surgical adjuvant breast and bowel project C-08 trial. J Clin Oncol. 2013;31:359–64.
  - 13. de Gramont A, Van Cutsem E, Schmoll HJ, Tabernero J, Clarke S, Moore MJ, Cunningham D, Cartwright TH, Hecht JR, Rivera F, Im SA, Bodoky G, Salazar R, Maindrault-Goebel F, Shacham-Shmueli E, Bajetta E, Makrutzki M, Shang A, Andre T, Hoff PM. Bevacizumab Plus oxaliplatin-based chemotherapy as adjuvant treatment for colon cancer (Avant): A Phase 3 randomised controlled trial. Lancet Oncol. 2012;13:1225–33.
  - 14. von Minckwitz G, Loibl S, Untch M, Eidtmann H, Rezai M, Fasching PA, Tesch H, Eggemann H, Schrader I, Kittel K, Hanusch C, Huober J, Solbach C, Jackisch C, Kunz G, Blohmer JU, Hauschild M, Fehm T, Nekljudova V, Gerber B. Survival after neoadjuvant chemotherapy with or without bevacizumab or everolimus for Her2-negative primary breast cancer (Gbg 44-Geparquinto) dagger. Ann Oncol. 2014;25:2363–72.
  - 15. Bear HD, Tang G, Rastogi P, Geyer CE Jr, Liu Q, Robidoux A, Baez-Diaz L, Brufsky AM, Mehta RS, Fehrenbacher L, Young JA, Senecal FM, Gaur R, Margolese RG, Adams PT, Gross HM, Costantino JP, Paik S, Swain SM, Mamounas EP, Wolmark N. Neoadjuvant plus adjuvant bevacizumab in early breast cancer (Nsabp B-40 [Nrg Oncology]): secondary outcomes of a phase 3. Randomised Controlled Trial Lancet Oncol. 2015;16:1037–48.
  - Jain RK. Antiangiogenesis strategies revisited: from starving tumors to alleviating hypoxia. Cancer Cell. 2014;26:605–22.
  - Shaked Y, Ciarrocchi A, Franco M, Lee CR, Man S, Cheung AM, Hicklin DJ, Chaplin D, Foster FS, Benezra R, Kerbel RS. Therapy-induced acute recruitment of circulating endothelial progenitor cells to tumors. Science. 2006;313:1785–7.
  - 18. Shaked Y, Henke E, Roodhart JM, Mancuso P, Langenberg MH, Colleoni M, Daenen LG, Man S, Xu P, Emmenegger U, Tang T, Zhu Z, Witte L, Strieter RM, Bertolini F, Voest EE, Benezra R, Kerbel RS. Rapid chemotherapy-induced acute endothelial progenitor cell mobilization: implications for antiangiogenic drugs as chemosensitizing agents. Cancer Cell. 2008;14:263–73.
  - 19. Shaked Y, Kerbel RS. Antiangiogenic strategies on defense: on the possibility of blocking rebounds by the tumor vasculature after chemotherapy. Cancer Res. 2007;67:7055–8.
  - Smith NR, Baker D, Farren M, Pommier A, Swann R, Wang X, Mistry S, McDaid K, Kendrew J, Womack C, Wedge SR, Barry ST. Tumor stromal architecture can define the intrinsic tumor response to VEGF-targeted therapy. Clin Cancer Res. 2013;19:6943–56.
  - 21. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. Nat Rev Cancer. 2008;8:592–603.
  - 22. Cree IA, Charlton P. Molecular chess? Hallmarks of anti-cancer drug resistance. BMC Cancer. 2017;17:10.
  - Boehm T, Folkman J, Browder T, O'Reilly MS. Antiangiogenic therapy of experimental cancer does not induce acquired drug resistance. Nature. 1997;390:404–7.
  - Yang Y, Sun M, Wang L, Jiao B. Hifs, angiogenesis, and cancer. J Cell Biochem. 2013;114:967–74.
  - Casanovas O, Hicklin DJ, Bergers G, Hanahan D. Drug resistance by evasion of antiangiogenic targeting of VEGF signaling in late-stage pancreatic islet tumors. Cancer Cell. 2005;8:299–309.
  - 26. Fischer C, Jonckx B, Mazzone M, Zacchigna S, Loges S, Pattarini L, Chorianopoulos E, Liesenborghs L, Koch M, De Mol M, Autiero M, Wyns S, Plaisance S, Moons L, van Rooijen N, Giacca M, Stassen JM, Dewerchin M, Collen D, Carmeliet P. Anti-Plgf inhibits

growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. Cell. 2007;131:463–75.

- Crawford Y, Kasman I, Yu L, Zhong C, Wu X, Modrusan Z, Kaminker J, Ferrara N. Pdgf-C mediates the angiogenic and tumorigenic properties of fibroblasts associated with tumors refractory to anti-VEGF treatment. Cancer Cell. 2009;15:21–34.
- Shojaei F, Lee JH, Simmons BH, Wong A, Esparza CO, Plumlee PA, Feng J, Stewart AE, Hu-Lowe DD, Christensen JG. Hgf/C-Met acts as an alternative angiogenic pathway in sunitinib-resistant tumors. Cancer Res. 2010;70:10090–100.
- Shojaei F, Simmons BH, Lee JH, Lappin PB, Christensen JG. Hgf/C-Met pathway is one of the mediators of sunitinib-induced tumor cell type-dependent metastasis. Cancer Lett. 2012;320:48–55.
- 30. Cascone T, Herynk MH, Xu L, Du Z, Kadara H, Nilsson MB, Oborn CJ, Park YY, Erez B, Jacoby JJ, Lee JS, Lin HY, Ciardiello F, Herbst RS, Langley RR, Heymach JV. Upregulated stromal Egfr and vascular remodeling in mouse xenograft models of angiogenesis inhibitor-resistant human lung adenocarcinoma. J Clin Invest. 2011;121:1313–28.
- Huang D, Ding Y, Zhou M, Rini BI, Petillo D, Qian CN, Kahnoski R, Futreal PA, Furge KA, Teh BT. Interleukin-8 mediates resistance to antiangiogenic agent sunitinib in renal cell carcinoma. Cancer Res. 2010;70:1063–71.
- 32. Shojaei F, Wu X, Qu X, Kowanetz M, Yu L, Tan M, Meng YG, Ferrara N. G-Csf-initiated myeloid cell mobilization and angiogenesis mediate tumor refractoriness to anti-VEGF therapy in mouse models. Proc Natl Acad Sci U S A. 2009;106:6742–7.
- van Beijnum JR, Nowak-Sliwinska P, Huijbers EJ, Thijssen VL, Griffioen AW. The great escape; the hallmarks of resistance to antiangiogenic therapy. Pharmacol Rev. 2015;67:441–61.
- 34. Shaked Y. Balancing efficacy of and host immune responses to cancer therapy: the Yin and Yang effects. Nat Rev Clin Oncol. 2016;13:611–26.
- 35. Batchelor TT, Sorensen AG, di Tomaso E, Zhang WT, Duda DG, Cohen KS, Kozak KR, Cahill DP, Chen PJ, Zhu M, Ancukiewicz M, Mrugala MM, Plotkin S, Drappatz J, Louis DN, Ivy P, Scadden DT, Benner T, Loeffler JS, Wen PY, Jain RK. Azd2171, a Pan-VEGF receptor tyrosine kinase inhibitor, normalizes tumor vasculature and alleviates edema in glioblastoma patients. Cancer Cell. 2007;11:83–95.
- 36. Kopetz S, Hoff PM, Morris JS, Wolff RA, Eng C, Glover KY, Adinin R, Overman MJ, Valero V, Wen S, Lieu C, Yan S, Tran HT, Ellis LM, Abbruzzese JL, Heymach JV. Phase II trial of infusional fluorouracil, irinotecan, and bevacizumab for metastatic colorectal cancer: efficacy and circulating angiogenic biomarkers associated with therapeutic resistance. J Clin Oncol. 2010;28:453–9.
- 37. Motzer RJ, Michaelson MD, Redman BG, Hudes GR, Wilding G, Figlin RA, Ginsberg MS, Kim ST, Baum CM, DePrimo SE, Li JZ, Bello CL, Theuer CP, George DJ, Rini BI. Activity of Su11248, a multitargeted inhibitor of vascular endothelial growth factor receptor and platelet-derived growth factor receptor, in patients with metastatic renal cell carcinoma. J Clin Oncol. 2006;24:16–24.
- Porta C, Paglino C, Imarisio I, Ganini C, Sacchi L, Quaglini S, Giunta V, De Amici M. Changes in circulating pro-angiogenic cytokines, other than VEGF, before progression to sunitinib therapy in advanced renal cell carcinoma patients. Oncology. 2013;84:115–22.
- 39. Rosen LS, Kurzrock R, Mulay M, Van Vugt A, Purdom M, Ng C, Silverman J, Koutsoukos A, Sun YN, Bass MB, Xu RY, Polverino A, Wiezorek JS, Chang DD, Benjamin R, Herbst RS. Safety, pharmacokinetics, and efficacy of Amg 706, an oral multikinase inhibitor, in patients with advanced solid tumors. J Clin Oncol. 2007;25:2369–76.
- 40. Willett CG, Boucher Y, Duda DG, di Tomaso E, Munn LL, Tong RT, Kozin SV, Petit L, Jain RK, Chung DC, Sahani DV, Kalva SP, Cohen KS, Scadden DT, Fischman AJ, Clark JW, Ryan DP, Zhu AX, Blaszkowsky LS, Shellito PC, Mino-Kenudson M, Lauwers GY. Surrogate markers for antiangiogenic therapy and dose-limiting toxicities for bevacizumab with radiation and chemotherapy: continued experience of a Phase I Trial in rectal cancer patients. J Clin Oncol. 2005;23:8136–9.

- 9 Resistance to Inhibitors of Angiogenesis
  - Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. Cancer Cell. 2012;21:309–22.
  - 42. Du R, Lu KV, Petritsch C, Liu P, Ganss R, Passegue E, Song H, Vandenberg S, Johnson RS, Werb Z, Bergers G. Hif1alpha induces the recruitment of bone marrow-derived vascular modulatory cells to regulate tumor angiogenesis and invasion. Cancer Cell. 2008;13:206–20.
  - 43. Shojaei F, Wu X, Malik AK, Zhong C, Baldwin ME, Schanz S, Fuh G, Gerber HP, Ferrara N. Tumor refractoriness to anti-VEGF treatment is mediated by Cd11b+Gr1+ Myeloid Cells. Nat Biotechnol. 2007;25:911–20.
  - 44. Shojaei F, Wu X, Zhong C, Yu L, Liang XH, Yao J, Blanchard D, Bais C, Peale FV, van Bruggen N, Ho C, Ross J, Tan M, Carano RA, Meng YG, Ferrara N. Bv8 regulates myeloidcell-dependent tumour angiogenesis. Nature. 2007;450:825–31.
  - 45. Carbone C, Moccia T, Zhu C, Paradiso G, Budillon A, Chiao PJ, Abbruzzese JL, Melisi D. Anti-VEGF treatment-resistant pancreatic cancers secrete proinflammatory factors that contribute to malignant progression by inducing an Emt cell phenotype. Clin Cancer Res. 2011;17:5822–32.
  - 46. Hasnis E, Alishekevitz D, Gingis-Veltski S, Bril R, Fremder E, Voloshin T, Raviv Z, Karban A, Shaked Y. Anti-Bv8 antibody and metronomic gemcitabine improve pancreatic adenocarcinoma treatment outcome following weekly gemcitabine therapy. Neoplasia. 2014;16:501–10.
  - 47. Chung AS, Wu X, Zhuang G, Ngu H, Kasman I, Zhang J, Vernes JM, Jiang Z, Meng YG, Peale FV, Ouyang W, Ferrara N. An Interleukin-17-mediated paracrine network promotes tumor resistance to anti-angiogenic therapy. Nat Med. 2013;19:1114–23.
  - De Palma M, Lewis CE. Macrophage regulation of tumor responses to anticancer therapies. Cancer Cell. 2013;23:277–86.
  - 49. Rolny C, Mazzone M, Tugues S, Laoui D, Johansson I, Coulon C, Squadrito ML, Segura I, Li X, Knevels E, Costa S, Vinckier S, Dresselaer T, Akerud P, De Mol M, Salomaki H, Phillipson M, Wyns S, Larsson E, Buysschaert I, Botling J, Himmelreich U, Van Ginderachter JA, De Palma M, Dewerchin M, Claesson-Welsh L, Carmeliet P. Hrg inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through down-regulation of Plgf. Cancer Cell. 2011;19:31–44.
  - 50. Zhang W, Zhu XD, Sun HC, Xiong YQ, Zhuang PY, Xu HX, Kong LQ, Wang L, Wu WZ, Tang ZY. Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. Clin Cancer Res. 2010;16:3420–30.
  - 51. Lewis CE, De Palma M, Naldini L. Tie2-expressing monocytes and tumor angiogenesis: regulation by hypoxia and angiopoietin-2. Cancer Res. 2007;67:8429–32.
  - 52. Coffelt SB, Tal AO, Scholz A, De Palma M, Patel S, Urbich C, Biswas SK, Murdoch C, Plate KH, Reiss Y, Lewis CE. Angiopoietin-2 regulates gene expression in Tie2-expressing monocytes and augments their inherent proangiogenic functions. Cancer Res. 2010;70:5270–80.
  - 53. Welford AF, Biziato D, Coffelt SB, Nucera S, Fisher M, Pucci F, Di Serio C, Naldini L, De Palma M, Tozer GM, Lewis CE. Tie2-expressing macrophages limit the therapeutic efficacy of the vascular-disrupting agent combretastatin A4 phosphate in mice. J Clin Invest. 2011;121:1969–73.
  - 54. De Palma M, Venneri MA, Galli R, Sergi Sergi L, Politi LS, Sampaolesi M, Naldini L. Tie2 identifies a hematopoietic lineage of proangiogenic monocytes required for tumor vessel formation and a mesenchymal population of pericyte progenitors. Cancer Cell. 2005;8:211–26.
  - 55. Kienast Y, Klein C, Scheuer W, Raemsch R, Lorenzon E, Bernicke D, Herting F, Yu S, The HH, Martarello L, Gassner C, Stubenrauch KG, Munro K, Augustin HG, Thomas M. Ang-2-VEGF-A Crossmab, a novel bispecific human IgG1 antibody blocking VEGF-A and Ang-2 functions simultaneously, mediates potent antitumor, antiangiogenic, and antimetastatic efficacy. Clin Cancer Res. 2013;19:6730–40.
  - 56. Peterson TE, Kirkpatrick ND, Huang Y, Farrar CT, Marijt KA, Kloepper J, Datta M, Amoozgar Z, Seano G, Jung K, Kamoun WS, Vardam T, Snuderl M, Goveia J, Chatterjee S, Batista A, Muzikansky A, Leow CC, Xu L, Batchelor TT, Duda DG, Fukumura D, Jain RK. Dual

inhibition of Ang-2 and VEGF receptors normalizes tumor vasculature and prolongs survival in glioblastoma by altering macrophages. Proc Natl Acad Sci U S A. 2016;113:4470–5.

- 57. Wu FT, Man S, Xu P, Chow A, Paez-Ribes M, Lee CR, Pirie-Shepherd SR, Emmenegger U, Kerbel RS. Efficacy of cotargeting Angiopoietin-2 and the VEGF pathway in the adjuvant postsurgical setting for early breast, colorectal, and renal cancers. Cancer Res. 2016;76:6988–7000.
- Armulik A, Abramsson A, Betsholtz C. Endothelial/pericyte interactions. Circ Res. 2005;97:512–23.
- 59. Thomas M, Kienast Y, Scheuer W, Bahner M, Kaluza K, Gassner C, Herting F, Brinkmann U, Seeber S, Kavlie A, Welschof M, Ries S, Weidner KM, Regula JT, Klein C. A novel Angiopoietin-2 selective fully human antibody with potent anti-tumoral and anti-angiogenic efficacy and superior side effect profile compared to pan-angiopoietin-1/–2 inhibitors. PLoS One. 2013;8:e54923.
- Kumar S, Mokhtari RB, Oliveira ID, Islam S, Toledo SR, Yeger H, Baruchel S. Tumor dynamics in response to antiangiogenic therapy with oral metronomic topotecan and pazopanib in neuroblastoma xenografts. Transl Oncol. 2013;6:493–503.
- 61. Mancuso MR, Davis R, Norberg SM, O'Brien S, Sennino B, Nakahara T, Yao VJ, Inai T, Brooks P, Freimark B, Shalinsky DR, Hu-Lowe DD, McDonald DM. Rapid vascular regrowth in tumors after reversal of VEGF inhibition. J Clin Invest. 2006;116:2610–21.
- Franco M, Roswall P, Cortez E, Hanahan D, Pietras K. Pericytes promote endothelial cell survival through induction of autocrine VEGF-a signaling and Bcl-W Expression. Blood. 2011;118:2906–17.
- 63. Erber R, Thurnher A, Katsen AD, Groth G, Kerger H, Hammes HP, Menger MD, Ullrich A, Vajkoczy P. Combined inhibition of VEGF and PDGF signaling enforces tumor vessel regression by interfering with pericyte-mediated endothelial cell survival mechanisms. FASEB J. 2004;18:338–40.
- 64. Bergers G, Song S, Meyer-Morse N, Bergsland E, Hanahan D. Benefits of targeting both pericytes and endothelial cells in the tumor vasculature with kinase inhibitors. J Clin Invest. 2003;111:1287–95.
- 65. Pietras K, Hanahan D. A multitargeted, metronomic, and maximum-tolerated dose "Chemo-Switch" regimen is antiangiogenic, producing objective responses and survival benefit in a mouse model of cancer. J Clin Oncol. 2005;23:939–52.
- Xian X, Hakansson J, Stahlberg A, Lindblom P, Betsholtz C, Gerhardt H, Semb H. Pericytes limit tumor cell metastasis. J Clin Invest. 2006;116:642–51.
- 67. Cooke VG, LeBleu VS, Keskin D, Khan Z, O'Connell JT, Teng Y, Duncan MB, Xie L, Maeda G, Vong S, Sugimoto H, Rocha RM, Damascena A, Brentani RR, Kalluri R. Pericyte depletion results in hypoxia-associated epithelial-to-mesenchymal transition and metastasis mediated by met signaling pathway. Cancer Cell. 2012;21:66–81.
- Pinto MP, Sotomayor P, Carrasco-Avino G, Corvalan AH, Owen GI. Escaping antiangiogenic therapy: strategies employed by cancer cells. Int J Mol Sci. 2016;17
- 69. Donnem T, Hu J, Ferguson M, Adighibe O, Snell C, Harris AL, Gatter KC, Pezzella F. Vessel co-option in primary human tumors and metastases: an obstacle to effective anti-angiogenic treatment? Cancer Med. 2013;2:427–36.
- de Groot JF, Fuller G, Kumar AJ, Piao Y, Eterovic K, Ji Y, Conrad CA. Tumor invasion after treatment of glioblastoma with bevacizumab: radiographic and pathologic correlation in humans and mice. Neuro-Oncology. 2010;12:233–42.
- 71. di Tomaso E, Snuderl M, Kamoun WS, Duda DG, Auluck PK, Fazlollahi L, Andronesi OC, Frosch MP, Wen PY, Plotkin SR, Hedley-Whyte ET, Sorensen AG, Batchelor TT, Jain RK. Glioblastoma recurrence after cediranib therapy in patients: lack of "rebound" revascularization as mode of escape. Cancer Res. 2011;71:19–28.
- 72. Narayana A, Kelly P, Golfinos J, Parker E, Johnson G, Knopp E, Zagzag D, Fischer I, Raza S, Medabalmi P, Eagan P, Gruber ML. Antiangiogenic therapy using bevacizumab in recurrent high-grade glioma: impact on local control and patient survival. J Neurosurg. 2009;110:173–80.

- 73. Norden AD, Young GS, Setayesh K, Muzikansky A, Klufas R, Ross GL, Ciampa AS, Ebbeling LG, Levy B, Drappatz J, Kesari S, Wen PY. Bevacizumab for recurrent malignant gliomas: efficacy, toxicity, and patterns of recurrence. Neurology. 2008;70:779–87.
- Rubenstein JL, Kim J, Ozawa T, Zhang M, Westphal M, Deen DF, Shuman MA. Anti-VEGF antibody treatment of glioblastoma prolongs survival but results in increased vascular cooption. Neoplasia. 2000;2:306–14.
- Kuczynski EA, Yin M, Bar-Zion A, Lee CR, Butz H, Man S, Daley F, Vermeulen PB, Yousef GM, Foster FS, Reynolds AR, Kerbel RS. Co-option of liver vessels and not sprouting angiogenesis drives acquired sorafenib resistance in hepatocellular carcinoma. J Natl Cancer Inst. 2016;108:djw030. https://doi.org/10.1093/jnci/djw030.
- 76. Jeong HS, Jones D, Liao S, Wattson DA, Cui CH, Duda DG, Willett CG, Jain RK, Padera TP. Investigation of the lack of angiogenesis in the formation of lymph node metastases. J Natl Cancer Inst. 2015;107:699. https://doi.org/10.1111/cup.12571.
- 77. Leenders WP, Kusters B, Verrijp K, Maass C, Wesseling P, Heerschap A, Ruiter D, Ryan A, de Waal R. Antiangiogenic therapy of cerebral melanoma metastases results in sustained tumor progression via vessel co-option. Clin Cancer Res. 2004;10:6222–30.
- 78. Frentzas S, Simoneau E, Bridgeman VL, Vermeulen PB, Foo S, Kostaras E, Nathan MR, Wotherspoon A, Gao ZH, Shi Y, Van den Eynden G, Daley F, Peckitt C, Tan X, Salman A, Lazaris A, Gazinska P, Berg TJ, Eltahir Z, Ritsma L, van Rheenen J, Khashper A, Brown G, Nystrom H, Sund M, Van Laere S, Loyer E, Dirix L, Cunningham D, Metrakos P, Reynolds AR. Vessel co-option mediates resistance to anti-angiogenic therapy in liver metastases. Nat Med. 2016;22:1294–302.
- 79. Bridgeman VL, Vermeulen PB, Foo S, Bilecz A, Daley F, Kostaras E, Nathan MR, Wan E, Frentzas S, Schweiger T, Hegedus B, Hoetzenecker K, Renyi-Vamos F, Kuczynski EA, Vasudev NS, Larkin J, Gore M, Dvorak HF, Paku S, Kerbel RS, Dome B, Reynolds AR. Vessel co-option is common in human lung metastases and mediates resistance to anti-angiogenic therapy in preclinical lung metastasis models. J Pathol. 2016;241:362.
- Kirschmann DA, Seftor EA, Hardy KM, Seftor RE, Hendrix MJ. Molecular pathways: vasculogenic mimicry in tumor cells: diagnostic and therapeutic implications. Clin Cancer Res. 2012;18:2726–32.
- Maniotis AJ, Folberg R, Hess A, Seftor EA, Gardner LM, Pe'er J, Trent JM, Meltzer PS, Hendrix MJ. Vascular channel formation by human melanoma cells in vivo and in vitro: vasculogenic mimicry. Am J Pathol. 1999;155:739–52.
- Paulis YW, Soetekouw PM, Verheul HM, Tjan-Heijnen VC, Griffioen AW. Signalling pathways in vasculogenic mimicry. Biochim Biophys Acta. 2010;1806:18–28.
- Ricci-Vitiani L, Pallini R, Biffoni M, Todaro M, Invernici G, Cenci T, Maira G, Parati EA, Stassi G, Larocca LM, De Maria R. Tumour vascularization via endothelial differentiation of glioblastoma stem-like cells. Nature. 2010;468:824–8.
- Wang R, Chadalavada K, Wilshire J, Kowalik U, Hovinga KE, Geber A, Fligelman B, Leversha M, Brennan C, Tabar V. Glioblastoma stem-like cells give rise to tumour endothelium. Nature. 2010;468:829–33.
- van der Schaft DW, Seftor RE, Seftor EA, Hess AR, Gruman LM, Kirschmann DA, Yokoyama Y, Griffioen AW, Hendrix MJ. Effects of angiogenesis inhibitors on vascular network formation by human endothelial and melanoma cells. J Natl Cancer Inst. 2004;96:1473–7.
- Soda Y, Marumoto T, Friedmann-Morvinski D, Soda M, Liu F, Michiue H, Pastorino S, Yang M, Hoffman RM, Kesari S, Verma IM. Transdifferentiation of glioblastoma cells into vascular endothelial cells. Proc Natl Acad Sci U S A. 2011;108:4274–80.
- Xu Y, Li Q, Li XY, Yang QY, Xu WW, Liu GL. Short-term anti-vascular endothelial growth factor treatment elicits vasculogenic mimicry formation of tumors to accelerate metastasis. J Exp Clin Cancer Res. 2012;31:16.
- Schnegg CI, Yang MH, Ghosh SK, Hsu MY. Induction of vasculogenic mimicry overrides VEGF-a silencing and enriches stem-like cancer cells in melanoma. Cancer Res. 2015;75:1682–90.

- Sun B, Zhang D, Zhang S, Zhang W, Guo H, Zhao X. Hypoxia influences vasculogenic mimicry channel formation and tumor invasion-related protein expression in melanoma. Cancer Lett. 2007;249:188–97.
- Ebos JM, Lee CR, Cruz-Munoz W, Bjarnason GA, Christensen JG, Kerbel RS. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. Cancer Cell. 2009;15:232–9.
- Paez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, Vinals F, Inoue M, Bergers G, Hanahan D, Casanovas O. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. Cancer Cell. 2009;15:220–31.
- Ebos JM, Lee CR, Christensen JG, Mutsaers AJ, Kerbel RS. Multiple circulating proangiogenic factors induced by sunitinib malate are tumor-independent and correlate with antitumor efficacy. Proc Natl Acad Sci U S A. 2007;104:17069–74.
- 93. Norden-Zfoni A, Desai J, Manola J, Beaudry P, Force J, Maki R, Folkman J, Bello C, Baum C, DePrimo SE, Shalinsky DR, Demetri GD, Heymach JV. Blood-based biomarkers of Su11248 activity and clinical outcome in patients with metastatic imatinib-resistant gastrointestinal stromal tumor. Clin Cancer Res. 2007;13:2643–50.
- 94. Zurita AJ, Khajavi M, Wu HK, Tye L, Huang X, Kulke MH, Lenz HJ, Meropol NJ, Carley W, DePrimo SE, Lin E, Wang X, Harmon CS, Heymach JV. Circulating cytokines and monocyte subpopulations as biomarkers of outcome and biological activity in sunitinib-treated patients with advanced neuroendocrine tumours. Br J Cancer. 2015;112:1199–205.
- 95. Ebos JM, Lee CR, Kerbel RS. Tumor and host-mediated pathways of resistance and disease progression in response to antiangiogenic therapy. Clin Cancer Res. 2009;15:5020–5.
- 96. Chung AS, Kowanetz M, Wu X, Zhuang G, Ngu H, Finkle D, Komuves L, Peale F, Ferrara N. Differential drug class-specific metastatic effects following treatment with a panel of angiogenesis inhibitors. J Pathol. 2012;227:404–16.
- 97. Welti JC, Powles T, Foo S, Gourlaouen M, Preece N, Foster J, Frentzas S, Bird D, Sharpe K, van Weverwijk A, Robertson D, Soffe J, Erler JT, Pili R, Springer CJ, Mather SJ, Reynolds AR. Contrasting effects of sunitinib within in vivo models of metastasis. Angiogenesis. 2012;15:623–41.
- Nguyen DX, Bos PD, Massague J. Metastasis: from dissemination to organ-specific colonization. Nat Rev Cancer. 2009;9:274–84.
- 99. Maione F, Capano S, Regano D, Zentilin L, Giacca M, Casanovas O, Bussolino F, Serini G, Giraudo E. Semaphorin 3a overcomes cancer hypoxia and metastatic dissemination induced by antiangiogenic treatment in mice. J Clin Invest. 2012;122:1832–48.
- 100. Sennino B, Ishiguro-Oonuma T, Wei Y, Naylor RM, Williamson CW, Bhagwandin V, Tabruyn SP, You WK, Chapman HA, Christensen JG, Aftab DT, McDonald DM. Suppression of tumor invasion and metastasis by concurrent inhibition of C-Met and VEGF signaling in pancreatic neuroendocrine tumors. Cancer Discov. 2012;2:270–87.
- Blagoev KB, Wilkerson J, Stein WD, Motzer RJ, Bates SE, Fojo AT. Sunitinib does not accelerate tumor growth in patients with metastatic renal cell carcinoma. Cell Rep. 2013;3:277–81.
- 102. Miles D, Harbeck N, Escudier B, Hurwitz H, Saltz L, Van Cutsem E, Cassidy J, Mueller B, Sirzen F. Disease course patterns after discontinuation of bevacizumab: pooled analysis of randomized phase III trials. J Clin Oncol. 2011;29:83–8.
- 103. Desar IM, Mulder SF, Stillebroer AB, van Spronsen DJ, van der Graaf WT, Mulders PF, van Herpen CM. The reverse side of the victory: flare up of symptoms after discontinuation of sunitinib or sorafenib in renal cell cancer patients. A report of three cases. Acta Oncol. 2009;48:927–31.
- 104. Wolter P, Beuselinck B, Pans S, Schoffski P. Flare-up: an often unreported phenomenon nevertheless familiar to oncologists prescribing tyrosine kinase inhibitors. Acta Oncol. 2009;48:621–4.
- 105. Cacheux W, Boisserie T, Staudacher L, Vignaux O, Dousset B, Soubrane O, Terris B, Mateus C, Chaussade S, Goldwasser F. Reversible tumor growth acceleration following bevaci-

zumab interruption in metastatic colorectal cancer patients scheduled for surgery. Ann Oncol. 2008;19:1659–61.

- 106. Paez-Ribes M, Man S, Xu P, Kerbel RS. Potential proinvasive or metastatic effects of preclinical antiangiogenic therapy are prevented by concurrent chemotherapy. Clin Cancer Res. 2015;21:5488–98.
- 107. Rovida A, Castiglioni V, Decio A, Scarlato V, Scanziani E, Giavazzi R, Cesca M. Chemotherapy counteracts metastatic dissemination induced by antiangiogenic treatment in mice. Mol Cancer Ther. 2013;12:2237–47.
- 108. Francia G, Cruz-Munoz W, Man S, Xu P, Kerbel RS. Mouse models of advanced spontaneous metastasis for experimental therapeutics. Nat Rev Cancer. 2011;11:135–41.
- 109. Ledermann JA, Hackshaw A, Kaye S, Jayson G, Gabra H, McNeish I, Earl H, Perren T, Gore M, Persic M, Adams M, James L, Temple G, Merger M, Rustin G. randomized phase ii placebo-controlled trial of maintenance therapy using the oral triple angiokinase inhibitor BIBF 1120 after chemotherapy for relapsed ovarian cancer. J Clin Oncol. 2011;29:3798–804.
- 110. Llovet JM, Decaens T, Raoul JL, Boucher E, Kudo M, Chang C, Kang YK, Assenat E, Lim HY, Boige V, Mathurin P, Fartoux L, Lin DY, Bruix J, Poon RT, Sherman M, Blanc JF, Finn RS, Tak WY, Chao Y, Ezzeddine R, Liu D, Walters I, Park JW. Brivanib in patients with advanced hepatocellular carcinoma who were intolerant to sorafenib or for whom sorafenib failed: results from the randomized phase III BRISK-PS study. J Clin Oncol. 2013;31:3509–16.
- 111. Duda DG, Kozin SV, Kirkpatrick ND, Xu L, Fukumura D, Jain RK. Cxcl12 (Sdf1alpha)-Cxcr4/Cxcr7 pathway inhibition: an emerging sensitizer for anticancer therapies? Clin Cancer Res. 2011;17:2074–80.
- 112. Ries CH, Cannarile MA, Hoves S, Benz J, Wartha K, Runza V, Rey-Giraud F, Pradel LP, Feuerhake F, Klaman I, Jones T, Jucknischke U, Scheiblich S, Kaluza K, Gorr IH, Walz A, Abiraj K, Cassier PA, Sica A, Gomez-Roca C, de Visser KE, Italiano A, Le Tourneau C, Delord JP, Levitsky H, Blay JY, Ruttinger D. Targeting tumor-associated macrophages with anti-Csf-1r antibody reveals a strategy for cancer therapy. Cancer Cell. 2014;25:846–59.
- 113. Ries CH, Hoves S, Cannarile MA, Ruttinger D. Csf-1/Csf-1r Targeting agents in clinical development for cancer therapy. Curr Opin Pharmacol. 2015;23:45–51.
- 114. Sandhu SK, Papadopoulos K, Fong PC, Patnaik A, Messiou C, Olmos D, Wang G, Tromp BJ, Puchalski TA, Balkwill F, Berns B, Seetharam S, de Bono JS, Tolcher AW. A first-in-human, first-in-class, phase i study of carlumab (Cnto 888), a human monoclonal antibody against CC-chemokine ligand 2 in patients with solid tumors. Cancer Chemother Pharmacol. 2013;71:1041–50.
- 115. Ruffell B, Coussens LM. Macrophages and therapeutic resistance in cancer. Cancer Cell. 2015;27:462–72.
- 116. Rapisarda A, Melillo G. Role of the hypoxic tumor microenvironment in the resistance to anti-angiogenic therapies. Drug Resist Updat. 2009;12:74–80.
- 117. Batchelor TT, Gerstner ER, Emblem KE, Duda DG, Kalpathy-Cramer J, Snuderl M, Ancukiewicz M, Polaskova P, Pinho MC, Jennings D, Plotkin SR, Chi AS, Eichler AF, Dietrich J, Hochberg FH, Lu-Emerson C, Iafrate AJ, Ivy SP, Rosen BR, Loeffler JS, Wen PY, Sorensen AG, Jain RK. Improved tumor oxygenation and survival in glioblastoma patients who show increased blood perfusion after cediranib and chemoradiation. Proc Natl Acad Sci U S A. 2013;110:19059–64.
- 118. Emblem KE, Mouridsen K, Bjornerud A, Farrar CT, Jennings D, Borra RJ, Wen PY, Ivy P, Batchelor TT, Rosen BR, Jain RK, Sorensen AG. Vessel architectural imaging identifies cancer patient responders to anti-angiogenic therapy. Nat Med. 2013;19:1178–83.
- 119. Sorensen AG, Batchelor TT, Zhang WT, Chen PJ, Yeo P, Wang M, Jennings D, Wen PY, Lahdenranta J, Ancukiewicz M, di Tomaso E, Duda DG, Jain RK. A "vascular normalization index" as potential mechanistic biomarker to predict survival after a single dose of cediranib in recurrent glioblastoma patients. Cancer Res. 2009;69:5296–300.
- 120. Sorensen AG, Emblem KE, Polaskova P, Jennings D, Kim H, Ancukiewicz M, Wang M, Wen PY, Ivy P, Batchelor TT, Jain RK. Increased survival of glioblastoma patients who respond to antiangiogenic therapy with elevated blood perfusion. Cancer Res. 2012;72:402–7.

- 121. Hutt DM, Roth DM, Vignaud H, Cullin C, Bouchecareilh M. The histone deacetylase inhibitor, vorinostat, represses hypoxia inducible factor 1 alpha expression through translational inhibition. PLoS One. 2014;9:e106224.
- 122. Aggarwal R, Thomas S, Pawlowska N, Bartelink I, Grabowsky J, Jahan T, Cripps A, Harb A, Leng J, Reinert A, Mastroserio I, Truong TG, Ryan CJ, Munster PN. Inhibiting histone deacetylase as a means to reverse resistance to angiogenesis inhibitors: phase I study of abexinostat plus pazopanib in advanced solid tumor malignancies. J Clin Oncol. 2017.:JCO2016705350;35:1231.
- 123. Gabrilovich DI, Chen HL, Girgis KR, Cunningham HT, Meny GM, Nadaf S, Kavanaugh D, Carbone DP. Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. Nat Med. 1996;2:1096–103.
- 124. Huang Y, Chen X, Dikov MM, Novitskiy SV, Mosse CA, Yang L, Carbone DP. Distinct roles of VEGFR-1 and VEGFR-2 in the aberrant hematopoiesis associated with elevated levels of VEGF. Blood. 2007;110:624–31.
- 125. Terme M, Pernot S, Marcheteau E, Sandoval F, Benhamouda N, Colussi O, Dubreuil O, Carpentier AF, Tartour E, Taieb J. VEGFA-VEGFR pathway blockade inhibits tumor-induced regulatory t-cell proliferation in colorectal cancer. Cancer Res. 2013;73:539–49.
- 126. Huang Y, Goel S, Duda DG, Fukumura D, Jain RK. Vascular normalization as an emerging strategy to enhance cancer immunotherapy. Cancer Res. 2013;73:2943–8.
- 127. Gabrilovich DI, Ishida T, Nadaf S, Ohm JE, Carbone DP. Antibodies to vascular endothelial growth factor enhance the efficacy of cancer immunotherapy by improving endogenous dendritic cell function. Clin Cancer Res. 1999;5:2963–70.
- 128. Hodi FS, Lawrence D, Lezcano C, Wu X, Zhou J, Sasada T, Zeng W, Giobbie-Hurder A, Atkins MB, Ibrahim N, Friedlander P, Flaherty KT, Murphy GF, Rodig S, Velazquez EF, Mihm MC Jr, Russell S, DiPiro PJ, Yap JT, Ramaiya N, Van den Abbeele AD, Gargano M, McDermott D. Bevacizumab plus ipilimumab in patients with metastatic melanoma. Cancer Immunol Res. 2014;2:632–42.
- 129. Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, Santosuosso M, Martin JD, Martin MR, Vianello F, Leblanc P, Munn LL, Huang P, Duda DG, Fukumura D, Jain RK, Poznansky MC. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. Proc Natl Acad Sci U S A. 2012;109:17561–6.
- 130. Ko JS, Zea AH, Rini BI, Ireland JL, Elson P, Cohen P, Golshayan A, Rayman PA, Wood L, Garcia J, Dreicer R, Bukowski R, Finke JH. Sunitinib mediates reversal of myeloid-derived suppressor cell accumulation in renal cell carcinoma patients. Clin Cancer Res. 2009;15:2148–57.
- 131. Manegold C, Dingemans AC, Gray JE, Nakagawa K, Nicolson M, Peters S, Reck M, Wu YL, Brustugun OT, Crino L, Felip E, Fennell D, Garrido P, Huber RM, Marabelle A, Moniuszko M, Mornex F, Novello S, Papotti M, Perol M, Smit EF, Syrigos K, van Meerbeeck JP, van Zandwijk N, Chih-Hsin Yang J, Zhou C, Vokes E. The potential of combined immunotherapy and antiangiogenesis for the synergistic treatment of advanced Nsclc. J Thorac Oncol. 2017;12:194–207.
- 132. Manning EA, Ullman JG, Leatherman JM, Asquith JM, Hansen TR, Armstrong TD, Hicklin DJ, Jaffee EM, Emens LA. A vascular endothelial growth factor receptor-2 inhibitor enhances antitumor immunity through an immune-based mechanism. Clin Cancer Res. 2007;13:3951–9.
- 133. Shrimali RK, Yu Z, Theoret MR, Chinnasamy D, Restifo NP, Rosenberg SA. Antiangiogenic agents can increase lymphocyte infiltration into tumor and enhance the effectiveness of adoptive immunotherapy of cancer. Cancer Res. 2010;70:6171–80.
- Clarke JM, Hurwitz HI. Understanding and targeting resistance to anti-angiogenic therapies. J Gastrointest Oncol. 2013;4:253–63.
- 135. Miles DW, de Haas SL, Dirix LY, Romieu G, Chan A, Pivot X, Tomczak P, Provencher L, Cortes J, Delmar PR, Scherer SJ. Biomarker results from the avado phase 3 trial of first-

line bevacizumab plus docetaxel for Her2-negative metastatic breast cancer. Br J Cancer. 2013;108:1052–60.

- 136. Van Cutsem E, de Haas S, Kang YK, Ohtsu A, Tebbutt NC, Ming Xu J, Peng Yong W, Langer B, Delmar P, Scherer SJ, Shah MA. Bevacizumab in combination with chemotherapy as first-line therapy in advanced gastric cancer: a biomarker evaluation from the avagast randomized phase iii trial. J Clin Oncol. 2012;30:2119–27.
- 137. Hegde PS, Jubb AM, Chen D, Li NF, Meng YG, Bernaards C, Elliott R, Scherer SJ, Chen DS. Predictive impact of circulating vascular endothelial growth factor in four phase III trials evaluating bevacizumab. Clin Cancer Res. 2013;19:929–37.
- 138. Jayson GC, Hicklin DJ, Ellis LM. Antiangiogenic therapy--evolving view based on clinical trial results. Nat Rev Clin Oncol. 2012;9:297–303.
- 139. Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med. 2004;350:2335–42.
- 140. Giantonio BJ, Catalano PJ, Meropol NJ, O'Dwyer PJ, Mitchell EP, Alberts SR, et al. Bevacizumab in combination with oxaliplatin, fluorouracil, and leucovorin (FOLFOX4) for previously treated metastatic colorectal cancer: results from the Eastern Cooperative Oncology Group Study E3200. J Clin Oncol. 2007;25:1539–44.
- 141. Saltz LB, Clarke S, Diaz-Rubio E, Scheithauer W, Figer A, Wong R, et al. Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: a randomized phase III study. J Clin Oncol. 2008;26:2013–9.
- 142. Sandler A, Gray R, Perry MC, Brahmer J, Schiller JH, Dowlati A, et al. Paclitaxel-carboplatin alone or with bevacizumab for non-small-cell lung cancer. N Engl J Med. 2006;355:2542–50.
- 143. Reck M, von Pawel J, Zatloukal P, Ramlau R, Gorbounova V, Hirsh V, et al. Phase III trial of cisplatin plus gemcitabine with either placebo or bevacizumab as first-line therapy for nonsquamous non-small-cell lung cancer: AVAil. J Clin Oncol. 2009;27:1227–34.
- 144. Reck M, von Pawel J, Zatloukal P, Ramlau R, Gorbounova V, Hirsh V, et al. Overall survival with cisplatin-gemcitabine and bevacizumab or placebo as first-line therapy for nonsquamous non-small-cell lung cancer: results from a randomised phase III trial (AVAiL). Ann Oncol. 2010;21(9):1804.
- 145. Rini BI, Halabi S, Rosenberg JE, Stadler WM, Vaena DA, Archer L, et al. Phase III trial of bevacizumab plus interferon alfa versus interferon alfa monotherapy in patients with metastatic renal cell carcinoma: final results of CALGB 90206. J Clin Oncol. 2010;28:2137–43.
- 146. Rini BI, Halabi S, Rosenberg JE, Stadler WM, Vaena DA, Ou SS, et al. Bevacizumab plus interferon alfa compared with interferon alfa monotherapy in patients with metastatic renal cell carcinoma: CALGB 90206. J Clin Oncol. 2008;26:5422–8.
- 147. Escudier B, Bellmunt J, Negrier S, Bajetta E, Melichar B, Bracarda S, et al. Phase III trial of bevacizumab plus interferon alfa-2a in patients with metastatic renal cell carcinoma (AVOREN): final analysis of overall survival. J Clin Oncol. 2010;28:2144–50.
- 148. Escudier B, Pluzanska A, Koralewski P, Ravaud A, Bracarda S, Szczylik C, et al. Bevacizumab plus interferon alfa-2a for treatment of metastatic renal cell carcinoma: a randomised, doubleblind phase III trial. Lancet. 2007;370:2103–11.
- 149. Perren TJ, Swart AM, Pfisterer J, Ledermann JA, Pujade-Lauraine E, Kristensen G, et al. A phase 3 trial of bevacizumab in ovarian cancer. N Engl J Med. 2011;365:2484–96.
- 150. Oza AM, Cook AD, Pfisterer J, Embleton A, Ledermann JA, Pujade-Lauraine E, et al. Standard chemotherapy with or without bevacizumab for women with newly diagnosed ovarian cancer (ICON7): overall survival results of a phase 3 randomised trial. Lancet Oncol. 2015;16:928–36.
- Burger RA, Brady MF, Bookman MA, Fleming GF, Monk BJ, Huang H, et al. Incorporation of bevacizumab in the primary treatment of ovarian cancer. N Engl J Med. 2011;365:2473–83.
- 152. Cohen MH, Shen YL, Keegan P, Pazdur R. FDA drug approval summary: bevacizumab (Avastin) as treatment of recurrent glioblastoma multiforme. Oncologist. 2009;14:1131–8.

- 153. Wilke H, Muro K, Van Cutsem E, Oh SC, Bodoky G, Shimada Y, et al. Ramucirumab plus paclitaxel versus placebo plus paclitaxel in patients with previously treated advanced gastric or gastro-oesophageal junction adenocarcinoma (RAINBOW): a double-blind, randomised phase 3 trial. Lancet Oncol. 2014;15:1224–35.
- 154. Fuchs CS, Tomasek J, Yong CJ, Dumitru F, Passalacqua R, Goswami C, et al. Ramucirumab monotherapy for previously treated advanced gastric or gastro-oesophageal junction adenocarcinoma (REGARD): an international, randomised, multicentre, placebo-controlled, phase 3 trial. Lancet. 2014;383:31–9.
- 155. Van Cutsem E, Tabernero J, Lakomy R, Prenen H, Prausova J, Macarulla T, et al. Addition of aflibercept to fluorouracil, leucovorin, and irinotecan improves survival in a phase III randomized trial in patients with metastatic colorectal cancer previously treated with an oxaliplatin-based regimen. J Clin Oncol. 2012;30:3499–506.
- 156. Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Siebels M, et al. Sorafenib in advanced clear-cell renal-cell carcinoma. N Engl J Med. 2007;356:125–34.
- 157. Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Staehler M, et al. Sorafenib for treatment of renal cell carcinoma: Final efficacy and safety results of the phase III treatment approaches in renal cancer global evaluation trial. J Clin Oncol. 2009;27:3312–8.
- 158. Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, et al. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med. 2008;359:378–90.
- 159. Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Oudard S, et al. Overall survival and updated results for sunitinib compared with interferon alfa in patients with metastatic renal cell carcinoma. J Clin Oncol. 2009;27:3584–90.
- Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Rixe O, et al. Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. N Engl J Med. 2007;356:115–24.
- 161. Demetri GD, van Oosterom AT, Garrett CR, Blackstein ME, Shah MH, Verweij J, et al. Efficacy and safety of sunitinib in patients with advanced gastrointestinal stromal tumour after failure of imatinib: a randomised controlled trial. Lancet. 2006;368:1329–38.
- 162. Raymond E, Dahan L, Raoul JL, Bang YJ, Borbath I, Lombard-Bohas C, et al. Sunitinib malate for the treatment of pancreatic neuroendocrine tumors. N Engl J Med. 2011;364:501–13.
- 163. Sternberg CN, Davis ID, Mardiak J, Szczylik C, Lee E, Wagstaff J, et al. Pazopanib in locally advanced or metastatic renal cell carcinoma: results of a randomized phase III trial. J Clin Oncol. 2010;28:1061–8.
- 164. Sternberg CN, Hawkins RE, Wagstaff J, Salman P, Mardiak J, Barrios CH, et al. A randomised, double-blind phase III study of pazopanib in patients with advanced and/or metastatic renal cell carcinoma: final overall survival results and safety update. Eur J Cancer. 2013;49:1287–96.
- 165. Motzer RJ, Escudier B, Tomczak P, Hutson TE, Michaelson MD, Negrier S, et al. Axitinib versus sorafenib as second-line treatment for advanced renal cell carcinoma: overall survival analysis and updated results from a randomised phase 3 trial. Lancet Oncol. 2013;14:552–62.
- 166. Grothey A, Van Cutsem E, Sobrero A, Siena S, Falcone A, Ychou M, et al. Regorafenib monotherapy for previously treated metastatic colorectal cancer (CORRECT): an international, multicentre, randomised, placebo-controlled, phase 3 trial. Lancet. 2013;381:303–12.
- 167. Reck M, Kaiser R, Mellemgaard A, Douillard JY, Orlov S, Krzakowski M, et al. Docetaxel plus nintedanib versus docetaxel plus placebo in patients with previously treated non-smallcell lung cancer (LUME-Lung 1): a phase 3, double-blind, randomised controlled trial. Lancet Oncol. 2014;15:143–55.
- 168. Wells SA Jr, Robinson BG, Gagel RF, Dralle H, Fagin JA, Santoro M, et al. Vandetanib in patients with locally advanced or metastatic medullary thyroid cancer: a randomized, doubleblind phase III trial. J Clin Oncol. 2012;30:134–41.
- 169. Elisei R, Schlumberger MJ, Muller SP, Schoffski P, Brose MS, Shah MH, et al. Cabozantinib in progressive medullary thyroid cancer. J Clin Oncol. 2013;31:3639–46.