
Regulation of CTL Infiltration Within the Tumor Microenvironment

3

Sarah E. Church and Jérôme Galon

3.1 The Predictive Capability of Tumor Infiltrating Lymphocytes

3.1.1 T-Lymphocyte Infiltration in the Tumor and Immunoscore

Classical methods for determining malignant disease prognosis are based upon the morphology and location of tumor cells at the primary sites and in lymph node tissues, and the existence of distant metastases. While this analysis provides important information about a patient's disease it fails to capture the biological complexity of the tumor microenvironment and the contribution of the anti-tumor immune response. Immunohistochemical and gene analyses of immune cells, particularly CD3⁺ T lymphocytes

in the primary tumor, provides a prognostic biomarker that is highly statistically accurate for predicting clinical outcome in the vast majority of cancer types including colorectal, lung, melanoma, ovarian, head and neck, breast, urothelial, hepatocellular, gallbladder, and esophageal (reviewed in [1–3]). Furthermore, basic histological quantification of T lymphocyte density, cytotoxicity, and memory by CD3, CD8, and CD45RO, respectively, has demonstrated that increased infiltration of T lymphocytes is associated with statistically significant improvement in patients' disease-free survival (DFS) and overall survival (OS) [2, 4, 5]. In colorectal carcinoma (CRC), further delineating the location of cytotoxic T lymphocytes (CTL--CD3⁺, CD8⁺) into two areas within the primary tumor, the center (CT) and the invading margin (IM), provides a statistically accurate prediction of clinical outcome [4]. Quantification of the density, phenotype, and location (CT or IM) of T lymphocytes has been termed Immunoscore [6–8]. In fact, for the first time it was shown that analysis of a marker, CD3, surpassed the gold standard of diagnostics via tumor-stage, lymph node, and metastatic invasion. Immunoscore defines patients into five categories (I0–I4) based on the distinct location (CT and IM) of CD3⁺ and CD8⁺ T lymphocytes within the primary tumor, where I0 has no CD3⁺ or CD8⁺ cells and I4 has high densities of both

S.E. Church (✉) • J. Galon
Laboratory of Integrative Cancer Immunology,
INSERM, UMRS1138, 15 Rue de l'École de
Médecine, Paris, France

Université Paris Descartes, Paris, France

Cordeliers Research Centre, Université Pierre et
Marie Curie Paris 6, Paris, France
e-mail: church.immunology@gmail.com;
jerome.galon@crc.jussieu.fr

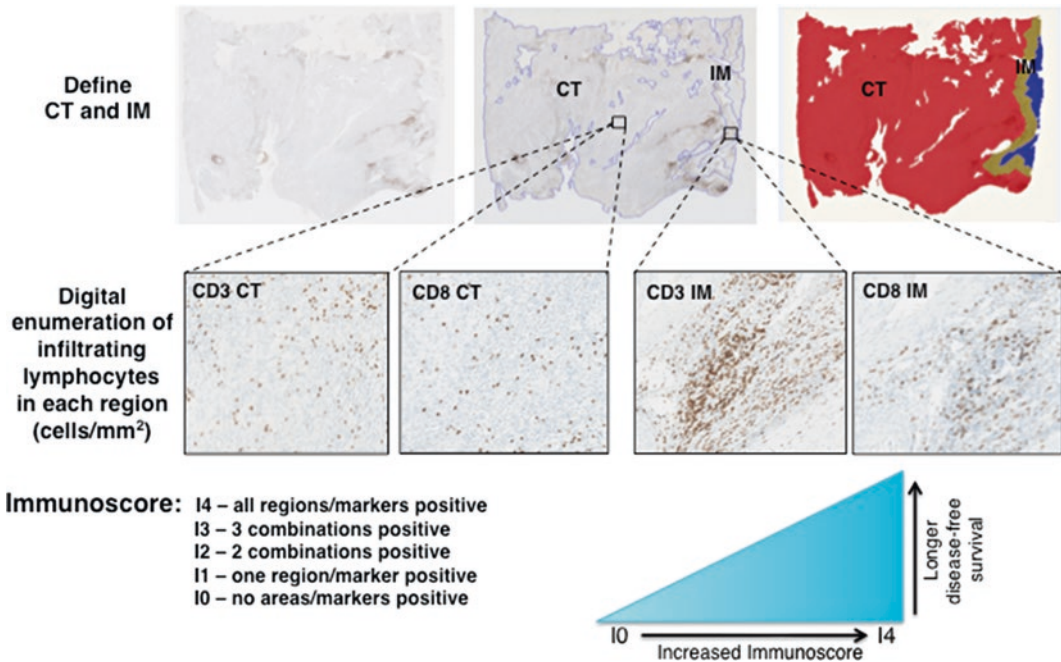


Fig. 3.1 Overview of Immunoscope. Immunoscope classifies tumors by density and location of CD3- and CD8-positive T lymphocytes. Paraffin-embedded tumor tissue is stained for CD3 or CD8. Stained tissue samples are analyzed for tumor and normal epithelium. The tumor center (CT) and invasive margin (IM) are then defined

using digital software. The software then enumerates infiltrating lymphocytes in each region. Immunoscope is calculated based on the density of each marker in both regions. Patients with high Immunoscope (I4) have significantly longer disease-free survival compared to patients with low Immunoscope (I0)

CD3⁺ and CD8⁺ cells in the CT and IM (Fig. 3.1) [9, 10]. Immunoscope utilizes simple analysis of two markers to accurately predict a patient's clinical outcome. By combining Immunoscope with quantification of additional immune components associated with the tumor microenvironment, as a part of the immune contexture, we continue to enrich our understanding of why tumors become resistant, avoid elimination, or fail to generate a tumor-specific cytotoxic response. In particular, identifying characteristics of the tumor microenvironment that lead to low densities of immune infiltrates and consequently low Immunoscope (I0 and I1) could dramatically improve the selection of personalized treatments for cancer. In the cases with high Immunoscope, patients might be more likely to respond to immunotherapies that stimulate an existing immune response, such as checkpoint blockades, while patients with low Immunoscope

would need therapy that primes a *de novo* anti-tumor response or facilitates trafficking of CTL to the tumor site.

3.1.2 Memory and Cytotoxic Lymphocytes Indicate Improved Prognosis

The importance of memory and CTL in the tumor microenvironment is well established. The presence of effector memory T cells in primary colorectal tumors is negatively correlated with signs of early metastatic invasion, as defined by presence of vascular emboli, lymphatic invasion, and perineural invasion [5]. This observation is supported by phenotypic analysis by flow cytometry of effector memory T lymphocytes, where patients with signs of early metastatic invasion have significantly fewer CD3⁺CD8⁺CD45RO⁺

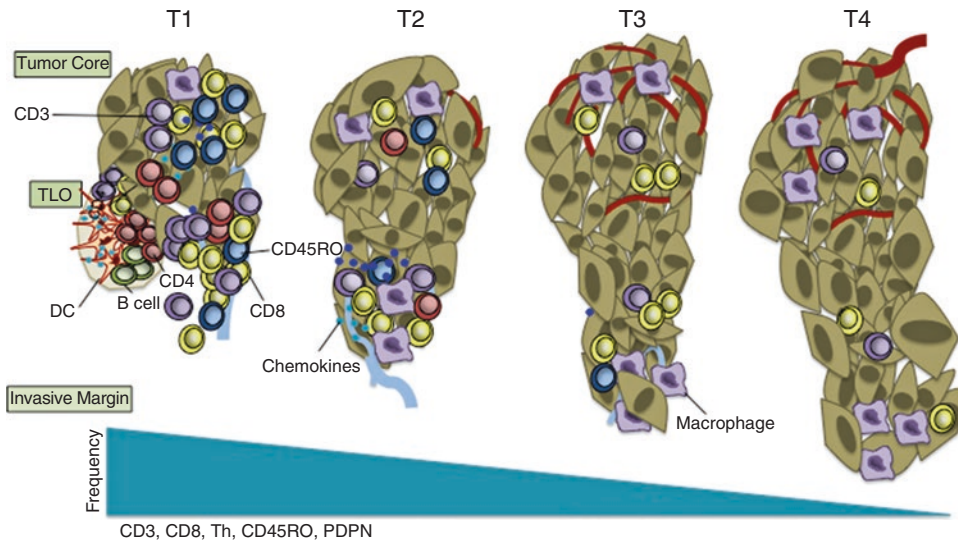


Fig. 3.2 The tumor microenvironment by progressive tumor stage. Depiction of the microenvironment of tumors from stage T1 to T4. Early stage tumors are smaller with many CD3, CD8, CD4, CD45RO, and TLO at the invasive margin and in the tumor center. T2 and T3 tumors are larger and progressively have less T lymphocytes, less

lymphatic vessels (PDPN) and more angiogenesis. T4 tumors have very few T lymphocytes, increased macrophages and increased angiogenesis. The presence of CTL, memory and helper lymphocytes at the invasive margin, tumor center and in TLO predicts better clinical outcomes

and $CD3^+CD8^+CCR7^-$ cells in the tumor, as well as by immunohistochemistry measuring CD45RO, where patients with metastatic progression have significantly fewer CD45RO-positive cells in the primary tumor compared to non-invasive disease [5]. Furthermore, a high density of CD45RO⁺ cells in the primary tumor significantly predicts better overall and disease-free survival compared to patients with low density of CD45RO⁺ expression within their tumors [5]. High expressers had median OS of 36.5 months and DFS of 53.2 months, compared to 11.1 and 20.6 months, respectively, for low density CD45RO. This indicates that not only are CTL an excellent biomarker for determining patient disease-related survival, but also further delineates the importance of the transition of CTL into memory phenotype, which can help refining the predictive capability of intratumoral CTL on patients' clinical outcome, as illustrated in Fig. 3.2. Two important mechanisms of memory T lymphocyte development and maintenance are determined by cytokine stimulation and help by CD4⁺ T lymphocytes [11–14]. The role of

these two mechanisms in maintaining anti-tumor memory and cytotoxic lymphocytes are discussed in the following sections.

3.1.3 Th1 and Th17 Have Opposing Effects on Disease-Specific Survival

It has been previously shown that incorporating the subtype and location of T helper (Th) lymphocytes, in addition to CTL, improves the accuracy of disease-specific survival prediction [15]. Primary CRC tumors from 125 patients for the expression of 45 immune genes representing four T helper populations, Th1, Th2, Th17 and regulatory T lymphocytes (Treg) were analyzed. Hierarchical clustering revealed eight categories of Th genes as follows: Th1 cytotoxic (IRF1, GZMB, IL27, GNLY, PRF1, CCL5, CD8a, STAT1), Th1 (IL12RB1, CD28, CCR5, HLA-DMB, IL12RB2, CD38, CXCR6, TBX21), Th17 (RORC, IL-17A), Th2 (IL4, IL5, IL13) or (CCR7, CD3E, CD40LG, CCL19, CCR4,

GATA3) or (IFNGR1/2, STAT3, IL10RB, IL4R, STAT6), Th2/Treg (FOXP3, CTLA-4, CCL17, CCL22) or Tregs (IL-10, TGFB). Interestingly, when disease-free survival (DFS) was assessed based on expression of Th1 cytotoxic genes, patients with high expression had significantly increased time to relapse versus patients with low expression (78 *versus* 18 months, $p = 0.01$). Conversely, patients with low expression of Th17 genes had prolonged disease-free survival with 80% of patients not experiencing relapse after 9 years. Since the Th1 cytotoxic and Th17 gene profiles exhibited this extreme contrast in prediction of DFS, the two gene profiles were assessed for complementarity. The patients were separated into 4 groups based on high or low Th1 cytotoxic or Th17 gene expression, Th1-Hi Th17-Hi, Th1-Hi Th17-Lo, and Th1-Lo Th17-Lo. Remarkably, the few patients with Th1-Hi and Th17-Lo had no tumor recurrence at 5 years, while patients with Th1-Hi and Th17-Hi had a DFS of 65% at 5 years and patients with Th1-Lo and Th17-Hi tumors had the worst outcome with DFS of 40% at 5 years. These findings were confirmed at the protein level by immunohistochemistry (IHC) analysis of IL-17 and CD8. Density and location (CT or IM) of IL17- and CD8-positive cells were analyzed where “high” is positive in both the CT and IM, “heterologous” has high density in either the CT or IM, and “low” has low densities of cells in both regions. DFS analysis showed the vigorously augmenting effect of IL17 expression on incidence of relapse, whereas IL17-low CD8-heterologous, IL17-heterologous CD8-heterologous and IL17-high and CD8-heterologous had 8, 40, and 80% observed relapse, respectively ($p < 0.001$). These data demonstrate the benefit of complementary analysis of Th1, Th17, and CTL in the tumor microenvironment.

The finding that high density of IL-17 expressing cells in the primary CRCs is a negative prognostic biomarker is not unprecedented because IL-17 production by T lymphocytes (Th, NK, Tc, $\gamma\delta$), NK, neutrophil, and innate lymphoid cells has been associated with colon tumorigenesis [16]. One mechanism for this is via commensal bacteria that skew Th17-directed inflammation,

leading to hyperplasia of normal colon cells and eventually colon cancer [17]. IL-17 also induces colorectal cell lines and primary cells to secrete pro-angiogenic factors, including vascular endothelial growth factor (VEGF) and can cause resistance to anti-angiogenic therapies [18, 19]. This pro-angiogenic stimulation likely also prevents trafficking of tumor-specific CTL to the tumor site, discussed in further detail below. Additional support for Th17 cells as a negative prognostic factor is that IL-17 expression defines patients with decreased disease-specific survival for pancreatic, breast, and gastric cancer [19–21], as well as increased tumor growth for intraocular lymphoma [22].

Contrary to Th17, Th1 cells are slowly becoming recognized for their important role in anti-tumor immunity for multiple types of cancer. Expression of CD4⁺ cells within the primary tumor correlates with improved prognosis in esophageal squamous carcinoma and non-small cell lung cancer, with statistically significant additive predictability when combined with enumeration of CTL [23, 24]. Helper CD4⁺ T lymphocytes, particularly tumor antigen-specific Th, guide CTL trafficking and maintain their cytolytic function within the tumor [25–27]. Tumor antigen-specific Th1 lymphocytes produce IFN- γ in the tumor microenvironment leading to the expression of CTL chemoattractants including CXCL9, CXCL10, CCL2, CCL3 and CCL5 [25]. Tumor-specific Th cells also produce IL-2, which is critical for CTL survival and can inhibit PD-1 mediated exhaustion of tumor-specific CTL leading to better efficacy of adoptive immunotherapy [27]. Additionally, mesenchymal stromal cells in the tumor microenvironment can directly activate CD4 T lymphocytes to become Th1 cells via IL-12 and consequently make tumor cells more vulnerable to CTL mediated destruction [28].

Regulatory T cells (Tregs) have an important role in the immune system to prevent uncontrolled immune responses to self-antigens. In cancer, this can lead to immunosuppression of the anti-tumor immune response due to self-antigens present on tumor cells. It has been shown that Treg gene signatures were not correlated to patient outcome; however, high densities

of FoxP3 protein expression was associated with increased disease-free survival of patients with colorectal cancer [15]. This is supported by other studies suggesting that in colorectal cancer, Tregs are not correlated with immunosuppression of the anti-tumor immune response and are significantly associated with high densities of CTL and Th1 T cell infiltration in the tumor [15, 29]. The effectiveness of Tregs as a prognostic maker has been variable between cancer types [30–32]. The comparisons of effector to Treg ratios, in hepatocellular and ovarian cancer showed that increased ratio of effector T cells to Tregs has positive prognostic value [33, 34]. Altogether this suggests that Tregs are a complicated biomarker for predicting patient outcome.

Finally, follicular T helper (Tfh) lymphocytes should be mentioned. Tfh cells are specialized to provide help to T and B lymphocytes, maintain memory B lymphocytes and produce IL-21 [35]. It has been previously reported that expression of the Tfh cells markers, CXCL13, CXCR5, and IL21 in the tumor were significantly correlated with prolonged disease-free survival [36]. Furthermore patients with aberration in the CXCL13 gene leading to gene deletion and dysfunction had significantly shorter disease-free survival compared to CRC patients with no aberration. High density of Tfh infiltration in the primary tumor has also been associated with prolonged disease-free survival in breast cancer [37]. The role of CXCL13 and IL-21 on CTL function will be further discussed in the following sections.

3.2 Factors Regulating Tumor Infiltration of Lymphocytes

3.2.1 T-Cell Homing Molecules Mediate Migration of CTL to Tumors

Chemokines have an important role in orchestrating both innate and adaptive immune cells chemotaxis and localization within the tumor. Chemokines can direct development and maintenance of tertiary lymphoid organs (TLO) that

prime tumor-specific CTL at the tumor site, which has been described in multiple cancer types including non-small cell lung cancer, melanoma, and colorectal carcinoma [36, 38–40].

We examined the predictive capability of chemokines using data integration of gene expression in primary tumors from CRC patients [41]. We discovered a significant prolongation of DFS in patients with high expression of the chemokines CX3CL1, CXCL10, and CXCL9. CX3CL1, also known as fractalkine, mediates T lymphocyte and monocytes migration and promotes strong adhesion to endothelial cells [42]. CXCL10, also named IFN- γ protein 10, and CXCL9 are closely related cytokines in the monokine-induced by IFN- γ family. CXCL10 and CXCL9 facilitate migration of CTL, monocytes, NK and dendritic cells, inhibit angiogenesis, and have anti-tumor properties [43, 44]. CRC patients with elevated gene expression of one of these three chemokines had increased percentage and density of CD3⁺CD8⁺ T lymphocytes in the tumor as assessed by flow cytometry and immunohistochemistry [41]. Analysis by location, CT or IM, within the tumor microenvironment showed that: (I) patients with high intra-tumoral CX3CL1 expression also had significantly increased density of effector-activated CTL (GZMB⁺) and Th1 (T-Bet⁺) cells; (II) tumors of patients with high CXCL9 and CXCL10 expression levels contained a significantly increased number of memory T lymphocytes (CD45RO⁺) and macrophage (CD68⁺). TCR repertoire analysis of ten patients randomly selected from the same cohort showed that the TCR repertoire of patients with a high CX3CL1 level was clearly distinguishable from the repertoire of patients with low CX3CL1 expression level. One cluster with CX3CL1, CXCL9, or CXCL10 gene expression levels correlated with a specific CTL repertoire (Vb5.2L08, Vb2L03, Vb2L07), thus suggesting that these chemokines attract clonal CTL with distinct tumor-specificity. Strikingly, when CRC tumors had high expression of any of these three TCRs the patients overall 3-year survival was 100%, as opposed to 28% with low expression of these TCRs. This suggests that CX3CL1, CXCL9, and CXCL10 in the tumor

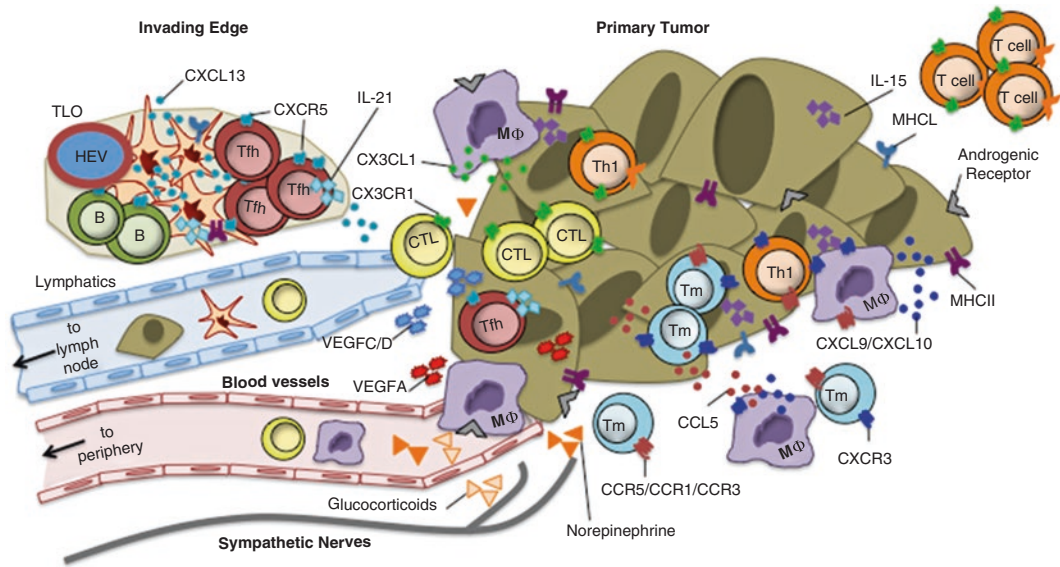


Fig. 3.3 Modulators of T lymphocytes in the tumor microenvironment. Overview of lymphatics, blood vessels, nerves, tertiary lymphoid organs (TLO), immune and tumor cells that produce cytokines, chemokines, hor-

mones, and immunosuppressive factors that regulate function and trafficking of lymphocytes in tumors. Expression of many of these factors can predict prognosis of patients with cancer, described in Table 3.1

microenvironment recruit tumor-specific CTL to eliminate malignant cells, and tumors become resistant to CTL-mediated death when these chemokines are not present. High expression density of CXCL9 and CXCL10 also accurately predicts prolonged disease-specific survival in melanoma patients [39, 45]. Pre-clinical studies with melanoma show that blocking CXCL9 or CXCL10 substantially reduces the ability of CTL to traffic to the primary tumor and distant metastatic lesions [45], which may be due to their role in directing CTL homing to the tumor by CD4 T lymphocyte help.

Another chemokine, CXCL13, was recently found to be associated with follicular helper T (Tfh) lymphocytes and also predicts patients' clinical outcome. CXCL13 is produced by and has been associated with generation of tertiary lymphoid organs (TLO) within the invasive margin of primary tumors [38, 46]. The presence of TLO in primary tumors is positively correlated to prolonged disease-free survival in multiple cancers [38, 39]. It is hypothesized that priming and

activation of tumor-specific CTL is orchestrated by dendritic cells presenting tumor-antigens within these TLO. In conjunction with this observation, CXCL13 as a single biomarker can accurately predict patients' clinical outcome [36]. Earlier it was discussed that low protein expression density or chromosomal aberration of CXCL13 is associated with worse clinical prognosis in CRC [36]. Similarly, in specific subtypes of breast cancer, elevated expression of CXCL13 in the tumor is associated with increased disease-free survival compared to tumors with low expression of CXCL13 [37]. Additionally, there is evidence supporting that high or low density CXCL13 expression can accurately predict patients' response to chemotherapy [47, 48]. It seems that the CXCL13-CXCR5 axis has the highest predictive score in HER2-positive breast cancers as opposed to other breast cancer subtypes [49]. This may be due to potential immunogenicity of HER2 for generating HER2-specific T helper and CTL immune response against the tumor. Figure 3.3 shows an overview of many

Table 3.1 Factors regulating lymphocyte infiltration into tumors

	Tumor immune factor	Prognosis	Ref
<i>Chemokines</i>			
CX3CL1	Mediates T-lymphocyte and monocyte migration to tumors and adhesion to endothelial cells	Good	[35, 36]
CXCL9/10 CCL5	Induced on tumor cells and MΦ by IFN-γ to promote CTL, monocyte, NK, and dendritic cell migration to the tumor and anti-angiogenic properties	Good	[25, 35, 37, 38]
CXCL13	Produced by dendritic cells in the TLO and signals through the CXCR5 receptor on B cells and Tfh cells controlling the organization of TLO	Good	[30, 32, 40]
<i>Cytokines</i>			
IL-17	Associated with tumorigenesis. Induces tumor and primary cells to secrete pro-angiogenic factors	Poor	[15, 18, 19]
IL-15	Regulates memory T lymphocyte maintenance and homing capabilities. Shown to rescue tolerant T cells and augment tumor-reactive CTL function and survival	Good	[11, 48, 50–52]
IL-21	Produced by NKT and Th cells. Activates and prevents exhaustion of tumor-specific CTL	Good	[30, 48, 55]
<i>Angiogenesis/Lymphatics</i>			
VEGFA	Generates leaky vasculature that prevents trafficking of leukocytes to the tumor. Stimulates suppressive Tregs and MDSC and induces immune checkpoints on endothelium (PD-L1, B7-H3, and TIM3)	Poor	[71–74]
VEGFC/D	Generates lymphatic vessels that are dysfunctional in fluid mechanics. Associated with chronic inflammation and induces secretion of immunosuppressive factors	Poor	[75, 77]
TLO/HEV	Facilitates priming, maintenance, and migration of lymphocytes in tumors. Presence in stroma correlates with high density of T and B cells	Good	[32, 78]
<i>Neural</i>			
Glucocorticoids	Induce expression of chemokine, cytokine, complement family members, innate immune-related genes, and TLR and repress adaptive immune-related genes. Reduce adaptive immune gene expression and skew Th1 cells to a Th2 phenotype. Upregulate IL-7Ra, enhance IL-7-mediated signaling and function, and inhibit apoptosis	Both	[82–84, 86]
Norepinephrine AR	Downregulates MHC-I, co-stimulatory molecules and increases production of IDO by tumor cells via beta-AR. AR signaling enhances Treg-mediated suppression, polarizes MΦ to a M2 phenotype and increases infiltration of MDSC	Poor	[87–92]

MΦ macrophage, Th T helper lymphocytes, CTL cytotoxic T lymphocytes, TLO tertiary lymphoid organ, MDSC myeloid-derived suppressor cells, MHC I MHC Class I, AR androgenic receptors

prognostic chemokines involved in T lymphocyte recruitment to the tumor. It is becoming increasingly clear that chemokines have an essential role in trafficking CTL to the tumor site. Furthermore, the addition of chemokine expression to Immunoscore has potential to predict patient response to chemotherapy [48].

3.2.2 Cytokines Contribute to the Distribution of CTL within the Tumor

An immense number of studies have been performed to investigate the components of the cytokine milieu that regulate lymphocytes in the tumor microenvironment. Interferon-gamma (IFN-γ) is well appreciated for its capacity to

prevent tumor growth during cancer immunotherapy [50]. Detection of an IFN- γ signature within the tumor has been associated with prolonged disease-specific survival in melanoma, colorectal, gastrointestinal, and ovarian cancer [51, 53]. To dissect the role of cytokines in tumor progression, a large cohort of CRC primary tumors has been analyzed for copy number variations in cytokines and cytokine receptors [54]. Fifty-nine soluble and membrane-bound proteins and their corresponding receptors from the IFN, IL, transforming growth factor (TGF), and tumor necrosis factor (TNF) families were analyzed. The majority (75%) of CRC patients displayed no difference in genomic alterations in the cytokine gene and receptor families studied. Of the remaining patients, the highest level of gain was in *IL29* and loss was in *IL15*. Furthermore, clinically advanced patients with distant metastases displayed a higher frequency of deletions in the interleukin family members *IL2*, *IL8*, *IL15*, and *IL21*. Interestingly, three of these deletions are in cytokines from the common γ -chain family, which has essential functions for maintenance, proliferation, and migration of memory, CTL, and Th lymphocytes [55]. Most strikingly, only patients with deletions in *IL2*, *IL15*, and *IL21* had significantly higher risk of tumor relapse [54]. On the other hand, gains or deletions of suppressive cytokines genes, *IL8*, *IL10*, and *TGF β* did not correlate to tumor recurrence.

Considering the previously discussed importance of localization and density of memory CD8⁺ T lymphocytes in the CRC tumor microenvironment, and the predominant role of IL-15 in the regulation of memory T lymphocyte homing and maintenance [56, 57], the cellular source of IL-15 within the CRC tumor microenvironment was investigated, both *in silico* and *in vitro*, by ClueGo and CluePedia, and IHC, respectively. It was discovered that tumor and myeloid cells were the source of IL-15, and increased IL-15 expression could significantly predict prolonged DFS [54]. IL-15 has been shown to rescue tolerant T lymphocytes [58] and augment the therapeutic efficacy of tumor-reactive CTL [11]. Moreover, patients with high expression of IL-15 in the tumor microenvironment have increased immune

cell density, immune gene expression, and DFS compared to medium or low expressing CRC tumors [54]. These data show that deletion of the *IL15* gene and reduced production of IL-15 by tumor cells is a substantial mechanism in preventing CTL infiltration and elimination of tumor cells. Furthermore, IL-15 signaling is highly effective in augmenting the anti-tumor CTL response, both as a mechanism to enhance CD4 T cell help and to maintain adoptively-transferred CTL survival [11, 58–60].

Expression of another common receptor λ -chain cytokine family member, IL-21, also predicts clinical outcome in CRC patients. CRC patients with chromosomal aberration of the *IL21* gene leading to deletion had higher risk of relapse than those without a deletion [54]. Overrepresentation of *IL2*, *IL15*, and *IL21* deletions was seen in patients with metastases, suggesting that these cytokines may be involved in putative anti-tumor immune mechanisms. IL-21 has a broad range of therapeutic anti-cancer properties, including activating and preventing exhaustion of tumor-specific CTL [61]. IL-21 is produced by NKT cells, Th1, Th17, and Tfh cells, again suggesting a role of Tfh as a substantial player in orchestrating the CTL response in the tumor.

3.3 Global Factors that Contribute to the Immune Contexture of Tumors

3.3.1 Mutagenesis and CTL Specificity

In melanoma, historically one of the most immune responsive cancers, it is known that the most potent tumor-specific T lymphocytes are directed toward neoantigens expressed by melanoma cells [62, 63]. Similarly, in other cancer types it has been documented that CTL specific for tumor neoantigens are extremely effective at immunosurveillance, elimination of tumor cells, and predicting clinical outcome [64, 65]. Recently, Alexandrov and colleagues reported extensive somatic mutational analysis describing 30 types of human cancer, where highly immunogenic cancers including melanoma, lung and

colorectal carcinoma had the highest prevalence of somatic mutations in their genome [66], thus integrating the paradigm that increased frequency of tumorigenic mutations provides better tumor-specific CTL targets. From this, it might be hypothesized that somatic mutations could be used as predictive biomarkers of cancer patient survival and response to therapy; however, the data up to this point has been inconsistent. Enumeration of total numbers of somatic mutations does not always predict prolonged disease-free survival, however presence of selected immunogenic mutations can distinguish patients with better clinical outcome [65, 67–69]. On the other hand, low numbers of genomic mutations can predict the presence of immunosuppressive mechanisms within the tumor [70]; furthermore, immunogenic mutational gene signatures have been shown to accurately predict benefit from CTLA-4 and PD-1 blocking immunotherapies [68, 69].

A few studies have reported that cancer-driver mutations are associated with immune gene signatures in the microenvironment, most notably in the RAS and EGFR genes. Interestingly, both of these genes have been linked to immune regulatory pathways. The presence of RAS mutations in colorectal carcinoma has been associated with decreased immunogenicity of tumors [71, 72]. Evidence also suggests that mutations in KRAS correlate with downregulation of MHC Class I molecules on tumor cells [71]. Additionally, it has been found that 20 immune genes encompassing checkpoint (CTLA-4, PD-1/L1/L2, TIM3 and LAG3), MHC class II, and Th1 genes were significantly under expressed in patients with KRAS mutations, independently of microsatellite stable or unstable disease [72]. EGFR mutations in NSCLC have been linked with upregulation of PD-L1 on tumor cells leading to inhibition of T lymphocyte response [73, 74].

This suggests that mutations in cancer driver genes themselves are not the most accurate measurement of patient prognosis, but the enumeration of immune-related gene mutations, particularly those involved in MHC-processing [70]. The predictive capability of expression of genes regulat-

ing immune cells and MHC-processing has been reported in colorectal, lung, ovary, breast, brain, and renal cancers [67, 75, 76].

3.3.2 Intratumoral Blood and Lymphatic Vessels Modulate CTL Trafficking

Tumor-stimulated angiogenesis is a well-established target for anti-cancer therapies because of the necessity for tumors to obtain a sufficient supply of nutrients. However, because blood vessels generated by tumor-induced angiogenesis lack structure causing blood flow to be leaky, leukocytes are unable to traffic properly [77]. Furthermore, many of the angiogenic tumor-derived factors have promiscuous functions in stimulating suppressive immune mechanisms, such as chemotaxis of Tregs and myeloid-derived suppressor cells to the tumor [78]. Angiogenic promoters also reduce endothelial adhesion molecules, preventing CTL from attaching to the vascular walls and migrating into the tumor [79]. The early inflammatory response driven by TNF- α induces adhesion molecules such as VCAM-1 in normal endothelium, however, when pro-angiogenic factors such as basic fibroblast growth factor are present, TNF- α loses the ability to invoke adhesion molecules [79]. Lastly, pro-angiogenic factors can also induce expression of immune checkpoints, including PD-L1/L2, B7-H3, galectin-1, and TIM3, on the endothelium putting the brakes on CTL activation [79, 80]. A few studies have demonstrated that combination therapy using angiogenesis inhibitors with anti-tumor immune stimulation can restore the migratory potential of CTL [81].

Tumors not only stimulate angiogenesis, but also the generation of new lymphatic vasculature by lymphangiogenesis via production of vascular endothelial growth factor C (VEGFC) and VEGFD [82]. VEGFC and VEGFD are often associated with poor clinical prognosis and increased cancer progression. Tumor-induced lymphatic vessels are important factor for dissemination of tumor cells into the lymph node and distant metastases. These tumor-induced

vessels have dysfunctional fluid mechanics that augment chronic inflammation and secrete immunosuppressive factors, such as TGF- β [83].

On the other hand, the presence of well-ordered lymphatic structures, including TLO and high endothelial venules (HEV) have been demonstrated to facilitate priming, maintenance, and migration of lymphocytes into solid tumors in melanoma, breast, ovarian, colon, and lung cancer [38, 84]. The presence of HEVs in the tumor stroma strongly correlates with increased infiltration of T and B lymphocytes. In breast cancer, high density of tumor HEVs is associated with the extravasation of Th1, CTL, and effector memory T lymphocytes into the tumor [84]. Furthermore patients with high density of tumor HEVs have longer metastasis-free survival, DFS, and OS. These observations were independent of the density of blood vessels within the tumor. In conjunction with these reports, the Authors have observed that patients with high density of lymphatic vessels, as measured by podoplanin (PDPN), in the IM of colorectal tumors are less likely to have metastatic invasion [85]. It is possible that high ordered lymphatic vessels facilitate CTL infiltration into the tumor at the edge of the invading tumor where activated CTL function to prevent metastatic dissemination.

3.3.3 Neuromodulators in Tumor Microenvironment

Chronic exposure to hormones, such as norepinephrine, progesterone, glucocorticoids, and androgenic receptor signaling have been linked to tumorigenesis and metastatic invasion of multiple cancer types (reviewed in [86, 87]). Glucocorticoids potential for immunosuppression is harnessed as an anti-inflammatory treatment for uncontrolled immune cells in patients with autoimmune disease [88]. Glucocorticoids are the major immunomodulatory agents used in clinical medicine. However, their actions as anti-inflammatory and immunosuppressive drugs are both beneficial and deleterious. Glucocorticoids induce the expression of chemokine, cytokine, complement family members, and innate immune-

related genes, including scavenger and Toll-like receptors [89]. In contrast, glucocorticoids repress the expression of adaptive immune-related genes [90]. Glucocorticoids modulate T helper lymphocyte differentiation by blocking IL-12-induced Stat4 phosphorylation without altering IL-4-induced Stat6 phosphorylation, therefore leading to suppressive action on the Th1 cellular immune response and a shift toward the Th2 humoral immune response [91]. However, glucocorticoids, in addition to their immunosuppressive function, enhance T-lymphocyte responses [92]. Glucocorticoids up-regulate IL-7RA and enhance IL-7-mediated signaling and function. Moreover, IL-7-mediated inhibition of apoptosis is increased in the presence of glucocorticoids, in a concentration-dependent manner, suggesting enhanced cell sensitivity to IL-7 following glucocorticoid exposure. These observations provide a mechanism by which glucocorticoids may also have a positive influence on T lymphocyte survival and function. Norepinephrine has been shown to downregulate expression of MHC class I molecules and co-stimulatory receptors, as well as increase production of IDO by tumor cells via beta-androgenic receptors [93]. Beta-androgenic receptor signaling has also been implicated in enhanced Treg suppression, polarization of macrophages to the M2 phenotype, and increased infiltration of myeloid-derived suppressor cells [94–98]. These hormones can be produced by cells in the tumor microenvironment or enter through the tumor vasculature. Altogether this suggests tumors that develop in conditions of chronic stress leading to hormone release may be preconditioned to an immunosuppressive immune contexture, which might lead to decreased infiltration of CTL.

3.4 Predicting Patients' Response to Treatment

The ultimate goal of predicting patient survival by immune gene signature and by the presence of immune cells in the tumor microenvironment is to accurately determine which personalized treatment will result in optimal tumor regression.

More and more clinical trials are incorporating immune cell quantification by immunohistochemistry, exome sequencing, gene expression, and flow cytometry to delineate why patients respond to therapy.

In the previous section we describe the use of immunogenetic mutational analysis of tumors to predict response to CTLA-4 and PD-1 blocking immunotherapy. Recently there have been a number of reports using immunohistochemical analysis to study immune cells, particularly CTL before and after therapy. A study in patients with melanoma investigated the expression of CD8 and PD-1/PD-L1 in the tumor center and invasive margin of tumor biopsies prior to and following treatment with a humanized blocking antibody to PD-1 [99]. Patients exhibiting beneficial response had significantly higher expression of CD8, PD-1, and PD-L1 in their invasive margin before treatment than patients whose tumors progressed following treatment. This data suggests that therapy blocking the PD-1/PD-L1 pathway would be most beneficial to patients that have pre-existing CTL in the tumor microenvironment. Even though one might assume that patients with high density of immune cells would respond better to immunotherapies targeting the immune system, the presence of immune genes and TILs also predicts patients' response to traditional chemotherapies [48, 100–102]. Analysis of colorectal cancer tumors from 1156 stage III patients treated with 5-fluorouracil based chemotherapy found that patients with TILs at the time of treatment had a better survival advantage after treatment than patients lacking TILs [102]. Additionally, two studies in breast cancer compare the predictive value of TILs for three types of therapy, docetaxel, doxorubicin, and trastuzumab [100, 101]. In the first study, patients with HER2-positive breast cancer were compared for high and low density of TILs. Patients with tumors containing high densities of TILs had significantly longer disease-free (5-year 78.6 vs. 47%) and overall (5-year 92.9 vs. 70.7%) survival after treatment with doxorubicin than patients with low densities of TILs. Interestingly, this was not the case when doxorubicin and docetaxel were administered in conjunction. In

another study it was shown that patients with HER2 positive breast cancer and high density TILs had better response to treatment with the anti-HER2 antibody, trastuzumab. These studies portray the importance of characterizing the immune microenvironment of tumors to determine optimal personalized beneficial treatments.

3.5 Conclusions

We ascertain that the evaluation of the CTL densities in primary tumors is a superior method of predicting patient survival for the majority cancer types. Moreover, patients that lack CTL in their primary tumor have the worst clinical prognosis and have tumors that are resistant to CTL killing because CTL are not able to traffic to the tumor site. We propose this is due to the lack of T helper lymphocytes, memory promoting cytokines, and chemoattractants, as well as dysfunctional blood and lymphatic flow preventing CTL from getting into the tumor microenvironment. We also hypothesize that potential immunosuppressive factors from stress-related hormones deters CTL from the tumor. These mechanisms are illustrated in Fig. 3.3. Plainly, if CTL are not present at the tumor site they are not able to eliminate tumor cells.

The difficult question remains, how do we induce CTL trafficking to the tumor for patients with low Immunoscore (I0-I1)? Currently systemic treatment of both recombinant human IL-15 and IL-21 are being tested in clinical trials with favorable results [57, 103]. In addition, IL-15 and IL-21 are being used in combination with adoptive immunotherapy to stimulate CTL *ex vivo* or as supplemental systemic administration. Initial studies using intratumoral injection of membrane-anchored chemokine fusion proteins, including CXCL10, are being used as a method to induce CTL trafficking to the tumor site [104, 105]. Another potential target to improve CTL migration to the tumor is by combination immunotherapy with angiogenesis-inhibitors. Inhibition of angiogenesis improves the organization of the vasculature allowing for better extravasation and migration of CTL into

the tumor [78, 79, 81, 106]. Angiogenesis inhibition has improved the therapeutic efficacy of both adoptive immunotherapy and vaccine-induced anti-tumor immunity. Finally, it will be essential that clinical studies incorporate tumor immune microenvironment analysis, such as Immunoscore, to fully understand the factors managing tumor-specific CTL trafficking to the tumor and quality of response to cancer therapy.

References

1. Angell H, Galon J. From the immune contexture to the Immunoscore: the role of prognostic and predictive immune markers in cancer. *Curr Opin Immunol*. 2013;25:261–7.
2. Fridman W-H, Pagès F, Sautes-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer*. 2012;12:298–306.
3. Galon J, Angell HK, Bedognetti D, Marincola FM. The continuum of cancer immunosurveillance: prognostic, predictive, and mechanistic signatures. *Immunity*. 2013;39:11–26.
4. Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P, Cugnenc P-H, Trajanoski Z, Fridman W-H, Pagès F. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science*. 2006;313:1960–4.
5. Pagès F, Berger A, Camus M, Sanchez-Cabo F, Costes A, Molitor R, Mlecnik B, Kirilovsky A, Nilsson M, Damotte D, Meatchi T, Bruneval P, Cugnenc P-H, Trajanoski Z, Fridman W-H, Galon J. Effector memory T cells, early metastasis, and survival in colorectal cancer. *N Eng J Med*. 2005;353:2654–66.
6. Galon J, Mlecnik B, Bindea G, Angell HK, Berger A, Lagorce C, Lugli A, Zlobec I, Hartmann A, Bifulco C, Nagtegaal ID, Palmqvist R, Masucci GV, Botti G, Tatangelo F, Delrio P, Maio M, Laghi L, Grizzi F, Asslaber M, D'Arrigo C, Vidal-Vanaclocha F, Zavadova E, Chouchane L, Ohashi PS, Hafezi Bakhtiari S, Wouters BG, Roehrl M, Nguyen L, Kawakami Y, Hazama S, Okuno K, Ogino S, Gibbs P, Waring P, Sato N, Torigoe T, Itoh K, Patel PS, Shukla SN, Wang Y, Kopetz S, Sinicrope FA, Scripcariu V, Ascierto PA, Marincola FM, Fox BA, Pagès F. Towards the introduction of the 'Immunoscore' in the classification of malignant tumours. *J Pathol*. 2014;232:199–209.
7. Galon J, Pages F, Marincola FM, Angell HK, Thurin M, Lugli A, Zlobec I, Berger A, Bifulco C, Botti G, Tatangelo F, Britten CM, Kreiter S, Chouchane L, Delrio P, Arndt H, Asslaber M, Maio M, Masucci GV, Mihm M, Vidal-Vanaclocha F, Allison JP, Gnjatic S, Hakansson L, Huber C, Singh-Jasuja H, Ottensmeier C, Zwierzina H, Laghi L, Grizzi F, Ohashi PS, Shaw PA, Clarke BA, Wouters BG, Kawakami Y, Hazama S, Okuno K, Wang E, O'Donnell-Tormey J, Lagorce C, Pawelec G, Nishimura MI, Hawkins R, Lapointe R, Lundqvist A, Khleif SN, Ogino S, Gibbs P, Waring P, Sato N, Torigoe T, Itoh K, Patel PS, Shukla SN, Palmqvist R, Nagtegaal ID, Wang Y, D'Arrigo C, Kopetz S, Sinicrope FA, Trinchieri G, Gajewski TF, Ascierto PA, Fox BA. Cancer classification using the Immunoscore: a worldwide task force. *J Transl Med*. 2012;10:205.
8. Galon J, Pages F, Marincola FM, Thurin M, Trinchieri G, Fox BA, Gajewski TF, Ascierto PA. The immune score as a new possible approach for the classification of cancer. *J Transl Med*. 2012;10:1.
9. Mlecnik B, Tosolini M, Kirilovsky A, Berger A, Bindea G, Meatchi T, Bruneval P, Trajanoski Z, Fridman W-H, Pagès F, Galon J. Histopathologic-based prognostic factors of colorectal cancers are associated with the state of the local immune reaction. *J Clin Oncol Off J Am Soc Clin Oncol*. 2011;29:610–8.
10. Pagès F, Kirilovsky A, Mlecnik B, Asslaber M, Tosolini M, Bindea G, Lagorce C, Wind P, Marliot F, Bruneval P, Zatloukal K, Trajanoski Z, Berger A, Fridman W-H, Galon J. In situ cytotoxic and memory T cells predict outcome in patients with early-stage colorectal cancer. *J Clin Oncol*. 2009;27:5944–51.
11. Klebanoff CA, Finkelstein SE, Surman DR, Lichtman MK, Gattinoni L, Theoret MR, Grewal N, Spiess PJ, Antony PA, Palmer DC, Tagaya Y, Rosenberg SA, Waldmann TA, Restifo NP. IL-15 enhances the in vivo antitumor activity of tumor-reactive CD8+ T cells. *Proc Natl Acad Sci U S A*. 2004;101:1969–74.
12. Sun JC, Bevan MJ. Defective CD8 T cell memory following acute infection without CD4 T cell help. *Science*. 2003;300:339–42.
13. Sprent J, Surh CD. Cytokines and T cell homeostasis. *Immunol Lett*. 2003;85:145–9.
14. Janssen EM, Lemmens EE, Wolfe T, Christen U, von Herrath MG, Schoenberger SP. CD4+ T cells are required for secondary expansion and memory in CD8+ T lymphocytes. *Nature*. 2003;421:852–6.
15. Tosolini M, Kirilovsky A, Mlecnik B, Fredriksen T, Mauger S, Bindea G, Berger A, Bruneval P, Fridman WH, Pages F, Galon J. Clinical impact of different classes of infiltrating T cytotoxic and helper cells (Th1, th2, treg, th17) in patients with colorectal cancer. *Cancer Res*. 2011;71:1263–71.
16. Wu D, Wu P, Huang Q, Liu Y, Ye J, Huang J. Interleukin-17: a promoter in colorectal cancer progression. *Clin Dev Immunol*. 2013;2013:436307.
17. Wu S, Rhee KJ, Albesiano E, Rabizadeh S, Wu X, Yen HR, Huso DL, Brancati FL, Wick E, McAllister F, Housseau F, Pardoll DM, Sears CL. A human colonic commensal promotes colon tumorigenesis

- via activation of T helper type 17 T cell responses. *Nat Med.* 2009;15:1016–22.
18. Chung AS, Wu X, Zhuang G, Ngu H, Kasman I, Zhang J, Vernes JM, Jiang Z, Meng YG, Peale FV, Ouyang W, Ferrara N. An interleukin-17-mediated paracrine network promotes tumor resistance to anti-angiogenic therapy. *Nat Med.* 2013;19:1114–23.
 19. Liu J, Duan Y, Cheng X, Chen X, Xie W, Long H, Lin Z, Zhu B. IL-17 is associated with poor prognosis and promotes angiogenesis via stimulating VEGF production of cancer cells in colorectal carcinoma. *Biochem Biophys Res Commun.* 2011;407:348–54.
 20. Liu T, Peng L, Yu P, Zhao Y, Shi Y, Mao X, Chen W, Cheng P, Wang T, Chen N, Zhang J, Liu X, Li N, Guo G, Tong W, Zhuang Y, Zou Q. Increased circulating Th22 and Th17 cells are associated with tumor progression and patient survival in human gastric cancer. *J Clin Immunol.* 2012;32:1332–9.
 21. He S, Fei M, Wu Y, Zheng D, Wan D, Wang L, Li D. Distribution and clinical significance of Th17 cells in the tumor microenvironment and peripheral blood of pancreatic cancer patients. *Int J Mol Sci.* 2011;12:7424–37.
 22. Galand C, Donnou S, Crozet L, Brunet S, Touitou V, Ouakrim H, Fridman WH, Sautes-Fridman C, Fisson S. Th17 cells are involved in the local control of tumor progression in primary intraocular lymphoma. *PLoS One.* 2011;6:e24622.
 23. Hiraoka K, Miyamoto M, Cho Y, Suzuoki M, Oshikiri T, Nakakubo Y, Itoh T, Ohbuchi T, Kondo S, Katoh H. Concurrent infiltration by CD8+ T cells and CD4+ T cells is a favourable prognostic factor in non-small-cell lung carcinoma. *Br J Cancer.* 2006;94:275–80.
 24. Cho Y, Miyamoto M, Kato K, Fukunaga A, Shichinohe T, Kawarada Y, Hida Y, Oshikiri T, Kurokawa T, Suzuoki M, Nakakubo Y, Hiraoka K, Murakami S, Shinohara T, Itoh T, Okushiba S, Kondo S, Katoh H. CD4+ and CD8+ T cells cooperate to improve prognosis of patients with esophageal squamous cell carcinoma. *Cancer Res.* 2003;63:1555–9.
 25. Bos R, Sherman LA. CD4+ T-cell help in the tumor milieu is required for recruitment and cytolytic function of CD8+ T lymphocytes. *Cancer Res.* 2010;70:8368–77.
 26. Wong SB, Bos R, Sherman LA. Tumor-specific CD4+ T cells render the tumor environment permissive for infiltration by low-avidity CD8+ T cells. *J Immunol.* 2008;180:3122–31.
 27. Church SE, Jensen SM, Antony PA, Restifo NP, Fox BA. Tumor-specific CD4+ T cells maintain effector and memory tumor-specific CD8+ T cells. *Eur J Immunol.* 2014;44:69–79.
 28. Jin P, Civini S, Zhao Y, De Giorgi V, Ren J, Sabatino M, Jin J, Wang H, Bedognetti D, Marincola F, Stroncek D. Direct T cell-tumour interaction triggers TH1 phenotype activation through the modification of the mesenchymal stromal cells transcriptional programme. *Br J Cancer.* 2014;110:2955–64.
 29. Salama P, Phillips M, Grieu F, Morris M, Zeps N, Joseph D, Platell C, Iacopetta B. Tumor-infiltrating FOXP3+ T regulatory cells show strong prognostic significance in colorectal cancer. *J Clin Oncol.* 2009;27:186–92.
 30. Adams SF, Levine DA, Cadungog MG, Hammond R, Facciabene A, Olvera N, Rubin SC, Boyd J, Gimotty PA, Coukos G. Intraepithelial T cells and tumor proliferation: impact on the benefit from surgical cytoreduction in advanced serous ovarian cancer. *Cancer.* 2009;115:2891–902.
 31. Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, Evdemon-Hogan M, Conejo-Garcia JR, Zhang L, Burow M, Zhu Y, Wei S, Kryczek I, Daniel B, Gordon A, Myers L, Lackner A, Disis ML, Knutson KL, Chen L, Zou W. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med.* 2004;10:942–9.
 32. Merlo A, Casalini P, Carcangiu ML, Malventano C, Triulzi T, Menard S, Tagliabue E, Balsari A. FOXP3 expression and overall survival in breast cancer. *J Clin Oncol.* 2009;27:1746–52.
 33. Gao Q, Qiu SJ, Fan J, Zhou J, Wang XY, Xiao YS, Xu Y, Li YW, Tang ZY. Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection. *J Clin Oncol.* 2007;25:2586–93.
 34. Sato E, Olson SH, Ahn J, Bundy B, Nishikawa H, Qian F, Jungbluth AA, Frosina D, Gnjjatic S, Ambrosone C, Kepner J, Odunsi T, Ritter G, Lele S, Chen YT, Ohtani H, Old LJ, Odunsi K. Intraepithelial CD8+ tumor-infiltrating lymphocytes and a high CD8+/regulatory T cell ratio are associated with favorable prognosis in ovarian cancer. *Proc Natl Acad Sci U S A.* 2005;102:18538–43.
 35. Schmitt N, Ueno H. Human T follicular helper cells: development and subsets. *Adv Exp Med Biol.* 2013;785:87–94.
 36. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC, Angell H, Fredriksen T, Lafontaine L, Berger A, Bruneval P, Fridman W-H, Becker C, Pagès F, Speicher MR, Trajanoski Z, Galon J. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. *Immunity.* 2013;39:782–95.
 37. Gu-Trantien C, Loi S, Garaud S, Equeter C, Libin M, de Wind A, Ravoet M, Le Buanec H, Sibille C, Manfouo-Foutsop G, Veys I, Haibe-Kains B, Singhal SK, Michiels S, Rothe F, Salgado R, Duvillier H, Ignatiadis M, Desmedt C, Bron D, Larsimont D, Piccart M, Sotiriou C, Willard-Gallo K. CD4(+) follicular helper T cell infiltration predicts breast cancer survival. *J Clin Invest.* 2013;123:2873–92.
 38. de Chaisemartin L, Goc J, Damotte D, Validire P, Magdeleinat P, Alifano M, Cremer I, Fridman WH, Sautes-Fridman C, Dieu-Nosjean MC. Characterization of chemokines and adhesion molecules associated with T cell presence in tertiary

- lymphoid structures in human lung cancer. *Cancer Res.* 2011;71:6391–9.
39. Messina JL, Fenstermacher DA, Eschrich S, Qu X, Berglund AE, Lloyd MC, Schell MJ, Sondak VK, Weber JS, Mule JJ. 12-chemokine gene signature identifies lymph node-like structures in melanoma: potential for patient selection for immunotherapy? *Sci Rep.* 2012;2:765.
 40. Coppola D, Nebozhyn M, Khalil F, Dai H, Yeatman T, Loboda A, Mule JJ. Unique ectopic lymph node-like structures present in human primary colorectal carcinoma are identified by immune gene array profiling. *Am J Pathol.* 2011;179:37–45.
 41. Mlecnik B, Tosolini M, Charoentong P, Kirilovsky A, Bindea G, Berger A, Camus M, Gillard M, Bruneval P, Fridman W-H, Pagès F, Trajanoski Z, Galon J. Biomolecular network reconstruction identifies T-cell homing factors associated with survival in colorectal cancer. *Gastroenterology.* 2010;138:1429–40.
 42. Umehara H, Bloom E, Okazaki T, Domae N, Imai T. Fractalkine and vascular injury. *Trends Immunol.* 2001;22:602–7.
 43. Dufour JH, Dziejman M, Liu MT, Leung JH, Lane TE, Luster AD. IFN-gamma-inducible protein 10 (IP-10; CXCL10)-deficient mice reveal a role for IP-10 in effector T cell generation and trafficking. *J Immunol.* 2002;168:3195–204.
 44. Romagnani P, Annunziato F, Lazzeri E, Cosmi L, Beltrame C, Lasagni L, Galli G, Francalanci M, Manetti R, Marra F, Vanini V, Maggi E, Romagnani S. Interferon-inducible protein 10, monokine induced by interferon gamma, and interferon-inducible T-cell alpha chemoattractant are produced by thymic epithelial cells and attract T-cell receptor (TCR) alphabeta+ CD8+ single-positive T cells, TCRgammadelta+ T cells, and natural killer-type cells in human thymus. *Blood.* 2001;97:601–7.
 45. Harlin H, Meng Y, Peterson AC, Zha Y, Tretiakova M, Slingluff C, McKee M, Gajewski TF. Chemokine expression in melanoma metastases associated with CD8+ T-cell recruitment. *Cancer Res.* 2009;69:3077–85.
 46. Crotty S. T follicular helper cell differentiation, function, and roles in disease. *Immunity.* 2014;41:529–42.
 47. Denkert C, von Minckwitz G, Brase JC, Sinn BV, Gade S, Kronenwett R, Pfitzner BM, Salat C, Loi S, Schmitt WD, Schem C, Fisch K, Darb-Esfahani S, Mehta K, Sotiriou C, Wienert S, Klare P, Andre F, Klauschen F, Blohmer JU, Krappmann K, Schmidt M, Tesch H, Kummel S, Sinn P, Jackisch C, Diel M, Reimer T, Untch M, Loibl S. Tumor-infiltrating lymphocytes and response to neoadjuvant chemotherapy with or without carboplatin in human epidermal growth factor receptor 2-positive and triple-negative primary breast cancers. *J Clin Oncol.* 2015;33:983–91.
 48. Stoll G, Enot D, Mlecnik B, Galon J, Zitvogel L, Kroemer G. Immune-related gene signatures predict the outcome of neoadjuvant chemotherapy. *Oncoimmunology.* 2014;3:e27884.
 49. Razis E, Kalogeras KT, Kotoula V, Eleftheraki AG, Nikitas N, Kronenwett R, Timotheadou E, Christodoulou C, Pectasides D, Gogas H, Wirtz RM, Makatsoris T, Bafaloukos D, Aravantinos G, Televantou D, Pavlidis N, Fountzilias G. Improved outcome of high-risk early HER2 positive breast cancer with high CXCL13-CXCR5 messenger RNA expression. *Clin Breast Cancer.* 2012;12:183–93.
 50. Kaplan DH, Shankaran V, Dighe AS, Stockert E, Aguet M, Old LJ, Schreiber RD. Demonstration of an interferon gamma-dependent tumor surveillance system in immunocompetent mice. *Proc Natl Acad Sci U S A.* 1998;95:7556–61.
 51. Delahaye NF, Rusakiewicz S, Martins I, Menard C, Roux S, Lyonnet L, Paul P, Sarabi M, Chaput N, Semeraro M, Minard-Colin V, Poirier-Colame V, Chaba K, Flament C, Baud V, Authier H, Kerdine-Romer S, Pallardy M, Cremer I, Peaudecerf L, Rocha B, Valteau-Couanet D, Gutierrez JC, Nunes JA, Commo F, Bonvalot S, Ibrahim N, Terrier P, Opolon P, Bottino C, Moretta A, Tavernier J, Rihet P, Coindre JM, Blay JY, Isambert N, Emile JF, Vivier E, Lecesne A, Kroemer G, Zitvogel L. Alternatively spliced NKp30 isoforms affect the prognosis of gastrointestinal stromal tumors. *Nat Med.* 2011;17:700–7.
 52. Duncan TJ, Rolland P, Deen S, Scott IV, Liu DT, Spendlove I, Durrant LG. Loss of IFN gamma receptor is an independent prognostic factor in ovarian cancer. *Clin Cancer Res.* 2007;13:4139–45.
 53. Lee JE, Abdalla J, Porter GA, Bradford L, Grimm EA, Reveille JD, Mansfield PF, Gershenwald JE, Ross MI. Presence of the human leukocyte antigen class II gene DRB1*1101 predicts interferon gamma levels and disease recurrence in melanoma patients. *Ann Surg Oncol.* 2002;9:587–93.
 54. Mlecnik B, Bindea G, Angell HK, Sasso MS, Obenauf AC, Fredriksen T, Lafontaine L, Bilocq AM, Kirilovsky A, Tosolini M, Waldner M, Berger A, Fridman W-H, Raffi A, Valge-Archer V, Pagès F, Speicher MR, Galon J. Functional network pipeline reveals genetic determinants associated with in situ lymphocyte proliferation and survival of cancer patients. *Sci Transl Med.* 2014;6:228ra237.
 55. Rochman Y, Spolski R, Leonard WJ. New insights into the regulation of T cells by gamma(c) family cytokines. *Nat Rev Immunol.* 2009;9:480–90.
 56. Wilkinson PC, Liew FY. Chemoattraction of human blood T lymphocytes by interleukin-15. *J Exp Med.* 1995;181:1255–9.
 57. Jakobisiak M, Golab J, Lasek W. Interleukin 15 as a promising candidate for tumor immunotherapy. *Cytokine Growth Factor Rev.* 2011;22:99–108.
 58. Teague RM, Sather BD, Sacks JA, Huang MZ, Dossett ML, Morimoto J, Tan X, Sutton SE, Cooke MP, Ohlén C, Greenberg PD. Interleukin-15 rescues tolerant CD8+ T cells for use in adoptive immunotherapy of established tumors. *Nat Med.* 2006;12:335–41.

59. Oh S, Perera LP, Terabe M, Ni L, Waldmann TA, Berzofsky JA. IL-15 as a mediator of CD4+ help for CD8+ T cell longevity and avoidance of TRAIL-mediated apoptosis. *Proc Natl Acad Sci U S A*. 2008;105:5201–6.
60. Yang T, Wall EM, Milne K, Theiss P, Watson P, Nelson BH. CD8+ T cells induce complete regression of advanced ovarian cancers by an interleukin (IL)-2/IL-15 dependent mechanism. *Clin Cancer Res*. 2007;13:7172–80.
61. Petrella TM, Tozer R, Belanger K, Savage KJ, Wong R, Smylie M, Kamel-Reid S, Tron V, Chen BE, Hunder NN, Hagerman L, Walsh W, Eisenhauer EA. Interleukin-21 has activity in patients with metastatic melanoma: a phase II study. *J Clin Oncol*. 2012;30:3396–401.
62. Lennerz V, Fatho M, Gentilini C, Frye RA, Lifke A, Ferel D, Wolfel C, Huber C, Wolfel T. The response of autologous T cells to a human melanoma is dominated by mutated neoantigens. *Proc Natl Acad Sci U S A*. 2005;102:16013–8.
63. Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, Ivanova Y, Hundal J, Arthur CD, Krebber WJ, Mulder GE, Toebes M, Vesely MD, Lam SS, Korman AJ, Allison JP, Freeman GJ, Sharpe AH, Pearce EL, Schumacher TN, Aebersold R, Rammensee HG, Melief CJ, Mardis ER, Gillanders WE, Artyomov MN, Schreiber RD. Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature*. 2014;515:577–81.
64. Matsushita H, Vesely MD, Koboldt DC, Rickert CG, Uppaluri R, Magrini VJ, Arthur CD, White JM, Chen YS, Shea LK, Hundal J, Wendl MC, Demeter R, Wylie T, Allison JP, Smyth MJ, Old LJ, Mardis ER, Schreiber RD. Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoeediting. *Nature*. 2012;482:400–4.
65. Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. *Science*. 2015;348:69–74.
66. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, Bignell GR, Bolli N, Borg A, Borresen-Dale AL, Boyault S, Burkhardt B, Butler AP, Caldas C, Davies HR, Desmedt C, Eils R, Eyfjord JE, Foekens JA, Greaves M, Hosoda F, Hutter B, Ilicic T, Imbeaud S, Imielinski M, Jager N, Jones DT, Jones D, Knappskog S, Kool M, Lakhani SR, Lopez-Otin C, Martin S, Munshi NC, Nakamura H, Northcott PA, Pajic M, Papaemmanuil E, Paradiso A, Pearson JV, Puente XS, Raine K, Ramakrishna M, Richardson AL, Richter J, Rosenstiel P, Schlesner M, Schumacher TN, Span PN, Teague JW, Totoki Y, Tutt AN, Valdes-Mas R, van Buuren MM, van 't Veer L, Vincent-Salomon A, Waddell N, Yates LR, Zucman-Rossi J, Futreal PA, McDermott U, Lichter P, Meyerson M, Grimmond SM, Siebert R, Campo E, Shibata T, Pfister SM, Campbell PJ, Stratton MR. Signatures of mutational processes in human cancer. *Nature*. 2013;500:415–21.
67. Brown SD, Warren RL, Gibb EA, Martin SD, Spinelli JJ, Nelson BH, Holt RA. Neo-antigens predicted by tumor genome meta-analysis correlate with increased patient survival. *Genome Res*. 2014;24:743–50.
68. Snyder A, Wolchok JD, Chan TA. Genetic basis for clinical response to CTLA-4 blockade. *N Engl J Med*. 2015;372:783.
69. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, Lee W, Yuan J, Wong P, Ho TS, Miller ML, Rehkman N, Moreira AL, Ibrahim F, Bruggeman C, Gasmfi B, Zappasodi R, Maeda Y, Sander C, Garon EB, Merghoub T, Wolchok JD, Schumacher TN, Chan TA. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science*. 2015;348:124–8.
70. Angelova M, Charoentong P, Hackl H, Fischer ML, Snajder R, Krogsdam AM, Waldner MJ, Bindea G, Mlecnik B, Galon J, Trajanoski Z. Characterization of the immunophenotypes and antigenomes of colorectal cancers reveals distinct tumor escape mechanisms and novel targets for immunotherapy. *Genome Biol*. 2015;16:64.
71. Atkins D, Breuckmann A, Schmahl GE, Binner P, Ferrone S, Krummenauer F, Storkel S, Seliger B. MHC class I antigen processing pathway defects, ras mutations and disease stage in colorectal carcinoma. *Int J Cancer*. 2004;109:265–73.
72. Lal N, Beggs AD, Willcox BE, Middleton GW. An immunogenomic stratification of colorectal cancer: implications for development of targeted immunotherapy. *Oncoimmunology*. 2015;4:e976052.
73. Akbay EA, Koyama S, Carretero J, Altabef A, Tchaicha JH, Christensen CL, Mikse OR, Cherniack AD, Beauchamp EM, Pugh TJ, Wilkerson MD, Fecci PE, Butaney M, Reibel JB, Soucheray M, Cohoon TJ, Janne PA, Meyerson M, Hayes DN, Shapiro GI, Shimamura T, Sholl LM, Rodig SJ, Freeman GJ, Hammerman PS, Dranoff G, Wong KK. Activation of the PD-1 pathway contributes to immune escape in EGFR-driven lung tumors. *Cancer Discov*. 2013;3:1355–63.
74. Azuma K, Ota K, Kawahara A, Hattori S, Iwama E, Harada T, Matsumoto K, Takayama K, Takamori S, Kage M, Hoshino T, Nakanishi Y, Okamoto I. Association of PD-L1 overexpression with activating EGFR mutations in surgically resected non-small-cell lung cancer. *Ann Oncol*. 2014;25:1935–40.
75. Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N. Molecular and genetic properties of tumors associated with local immune cytolytic activity. *Cell*. 2015;160:48–61.
76. Bindea G, Mlecnik B, Angell HK, Galon J. The immune landscape of human tumors: implications for cancer immunotherapy. *Oncoimmunology*. 2014;3:e27456.
77. Folkman J. Angiogenesis. *Annu Rev Med*. 2006;57:1–18.
78. Motz GT, Coukos G. The parallel lives of angiogenesis and immunosuppression: cancer and other tales. *Nat Rev Immunol*. 2011;11:702–11.

79. Griffioen AW, Damen CA, Blijham GH, Groenewegen G. Tumor angiogenesis is accompanied by a decreased inflammatory response of tumor-associated endothelium. *Blood*. 1996;88:667–73.
80. Melero I, Rouzaut A, Motz GT, Coukos G. T-cell and NK-cell infiltration into solid tumors: a key limiting factor for efficacious cancer immunotherapy. *Cancer Discov*. 2014;4:522–6.
81. Griffioen AW. Anti-angiogenesis: making the tumor vulnerable to the immune system. *Cancer Immunol Immunother*. 2008;57:1553–8.
82. Tammela T, Alitalo K. Lymphangiogenesis: molecular mechanisms and future promise. *Cell*. 2010;140:460–76.
83. Swartz MA, Lund AW. Lymphatic and interstitial flow in the tumour microenvironment: linking mechanobiology with immunity. *Nat Rev Cancer*. 2012;12:210–9.
84. Martinet L, Garrido I, Filleron T, Le Guellec S, Bellard E, Fournie JJ, Rochoix P, Girard JP. Human solid tumors contain high endothelial venules: association with T- and B-lymphocyte infiltration and favorable prognosis in breast cancer. *Cancer Res*. 2011;71:5678–87.
85. Mlecnik B, Bindea G, Kirilovsky A, Angell HK, Obenauf AC, Tosolini M, Church SE, Maby P, Vasaturo A, Angelova M, Fredriksen T, Mauger S, Waldner M, Berger A, Speicher MR, Pages F, Valge-Archer V, Galon J. The tumor microenvironment and Immunoscore are critical determinants of dissemination to distant metastasis. *Sci Transl Med*. 2016;8(327):327ra26.
86. Eng JW, Kokolus KM, Reed CB, Hylander BL, Ma WW, Repasky EA. A nervous tumor microenvironment: the impact of adrenergic stress on cancer cells, immunosuppression, and immunotherapeutic response. *Cancer Immunol Immunother*. 2014;63:1115–28.
87. Garcia SB, Stopper H, Kannen V. The contribution of neuronal-glia-endothelial-epithelial interactions to colon carcinogenesis. *Cell Mol Life Sci*. 2014;71:3191–7.
88. Coutinho AE, Chapman KE. The anti-inflammatory and immunosuppressive effects of glucocorticoids, recent developments and mechanistic insights. *Mol Cell Endocrinol*. 2011;335:2–13.
89. Sternberg EM. Neural regulation of innate immunity: a coordinated nonspecific host response to pathogens. *Nat Rev Immunol*. 2006;6:318–28.
90. Galon J, Franchimont D, Hiroi N, Frey G, Boettner A, Ehrhart-Bornstein M, O'Shea JJ, Chrousos GP, Bornstein SR. Gene profiling reveals unknown enhancing and suppressive actions of glucocorticoids on immune cells. *FASEB J*. 2002;16:61–71.
91. Franchimont D, Galon J, Gadina M, Visconti R, Zhou Y, Aringer M, Frucht DM, Chrousos GP, O'Shea JJ. Inhibition of Th1 immune response by glucocorticoids: dexamethasone selectively inhibits IL-12-induced Stat4 phosphorylation in T lymphocytes. *J Immunol*. 2000;164:1768–74.
92. Franchimont D, Galon J, Vacchio MS, Fan S, Visconti R, Frucht DM, Geenen V, Chrousos GP, Ashwell JD, O'Shea JJ. Positive effects of glucocorticoids on T cell function by up-regulation of IL-7 receptor alpha. *J Immunol*. 2002;168:2212–8.
93. Wang L, Liu H, Chen X, Zhang M, Xie K, Ma Q. Immune sculpting of norepinephrine on MHC-I, B7-1, IDO and B7-H1 expression and regulation of proliferation and invasion in pancreatic carcinoma cells. *PLoS One*. 2012;7:e45491.
94. Bernard AC, Fitzpatrick EA, Maley ME, Gellin GL, Tsuei BJ, Arden WA, Boulanger BR, Kearney PA, Ochoa JB. Beta adrenoceptor regulation of macrophage arginase activity. *Surgery*. 2000;127:412–8.
95. Cosentino M, Fietta AM, Ferrari M, Rasini E, Bombelli R, Carcano E, Saporiti F, Meloni F, Marino F, Lecchini S. Human CD4+CD25+ regulatory T cells selectively express tyrosine hydroxylase and contain endogenous catecholamines subserving an autocrine/paracrine inhibitory functional loop. *Blood*. 2007;109:632–42.
96. Flierl MA, Rittirsch D, Nadeau BA, Sarma JV, Day DE, Lentsch AB, Huber-Lang MS, Ward PA. Upregulation of phagocyte-derived catecholamines augments the acute inflammatory response. *PLoS One*. 2009;4:e4414.
97. Guereschi MG, Araujo LP, Maricato JT, Takenaka MC, Nascimento VM, Vivanco BC, Reis VO, Keller AC, Brum PC, Basso AS. Beta2-adrenergic receptor signaling in CD4+ Foxp3+ regulatory T cells enhances their suppressive function in a PKA-dependent manner. *Eur J Immunol*. 2013;43:1001–12.
98. Qin JF, Jin FJ, Li N, Guan HT, Lan L, Ni H, Want Y. Adrenergic receptor B2 activation by stress promotes breast cancer progression through macrophages M2 polarization in tumor microenvironment. *BMB Rep*. 2015;48(5):295–300.
99. Tumeh PC, Harvieu CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, Chmielowski B, Spasic M, Henry G, Ciobanu V, West AN, Carmona M, Kivork C, Seja E, Cherry G, Gutierrez AJ, Grogan TR, Mateus C, Tomicic G, Glaspy JA, Emerson RO, Robins H, Pierce RH, Elashoff DA, Robert C, Ribas A. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature*. 2014;515:568–71.
100. Loi S, Michiels S, Salgado R, Sirtaine N, Jose V, Fumagalli D, Kellokumpu-Lehtinen PL, Bono P, Kataja V, Desmedt C, Piccart MJ, Loibl S, Denkert C, Smyth MJ, Joensuu H, Sotiriou C. Tumor infiltrating lymphocytes are prognostic in triple negative breast cancer and predictive for trastuzumab benefit in early breast cancer: results from the FinHER trial. *Ann Oncol*. 2014;25:1544–50.
101. Loi S, Sirtaine N, Piette F, Salgado R, Viale G, Van Eenoo F, Rouas G, Francis P, Crown JP, Hitre E, de Azambuja E, Quinaux E, Di Leo A, Michiels S, Piccart MJ, Sotiriou C. Prognostic and predic-

- tive value of tumor-infiltrating lymphocytes in a phase III randomized adjuvant breast cancer trial in node-positive breast cancer comparing the addition of docetaxel to doxorubicin with doxorubicin-based chemotherapy: BIG 02-98. *J Clin Oncol*. 2013;31:860–7.
102. Morris M, Platell C, Iacopetta B. Tumor-infiltrating lymphocytes and perforation in colon cancer predict positive response to 5-fluorouracil chemotherapy. *Clin Cancer Res*. 2008;14:1413–7.
 103. Santegoets SJ, Turksma AW, Powell DJ Jr, Hooijberg E, de Gruijl TD. IL-21 in cancer immunotherapy: at the right place at the right time. *Oncoimmunology*. 2013;2:e24522.
 104. Nelson PJ, Muenchmeier N. Membrane-anchored chemokine fusion proteins: a novel class of adjuvants for immunotherapy. *Oncoimmunology*. 2013;2:e26619.
 105. Muenchmeier N, Boecker S, Bankel L, Hinz L, Rieth N, Lapa C, Mendler AN, Noessner E, Mocikat R, Nelson PJ. A novel CXCL10-based GPI-anchored fusion protein as adjuvant in NK-based tumor therapy. *PLoS One*. 2013;8:e72749.
 106. Shrimali RK, Yu Z, Theoret MR, Chinnasamy D, Restifo NP, Rosenberg SA. Antiangiogenic agents can increase lymphocyte infiltration into tumor and enhance the effectiveness of adoptive immunotherapy of cancer. *Cancer Res*. 2010;70:6171–80.