



# Pharmacology of Inhaled Anesthetics

*Elizabeth Demers Lavelle and Swamy Kurra*

- 10.1 Introduction – 179**
- 10.2 Physical and Chemical Properties of Inhaled Anesthetics – 179**
  - 10.2.1 Nitrous Oxide – 179
  - 10.2.2 Halothane – 180
  - 10.2.3 Enflurane – 180
  - 10.2.4 Isoflurane – 180
  - 10.2.5 Sevoflurane – 180
  - 10.2.6 Desflurane – 180
  - 10.2.7 Xenon – 181
- 10.3 Mechanism of Action – 181**
- 10.4 Systemic Effects of Inhaled Anesthetics – 182**
  - 10.4.1 Effects on Central Nervous System – 183
  - 10.4.2 Effects on Cardiovascular System – 183
  - 10.4.3 Effects on Respiration – 183
  - 10.4.4 Effects on Neuromuscular Junction – 184
  - 10.4.5 Effects on Renal Function – 184
  - 10.4.6 Effects on Hepatic Function – 184
  - 10.4.7 Effects on Hematologic and Immune Systems – 184
- 10.5 Biotransformation and Toxicity of Inhaled Anesthetics – 184**
  - 10.5.1 Nitrous Oxide – 184
  - 10.5.2 Halothane – 184
  - 10.5.3 Methoxyflurane – 185
  - 10.5.4 Isoflurane – 185
  - 10.5.5 Desflurane – 186
  - 10.5.6 Sevoflurane – 186
  - 10.5.7 Possible Neurotoxicity of Inhaled Anesthetics – 186
- 10.6 Minimum Alveolar Concentration and Its Affecting Factors – 186**

---

The original version of this chapter was revised. The abbreviation “MAP mean arterial pressure” was additionally present in the footnote of Table 10.2. This has now been removed. The correction to this chapter can be found at [https://doi.org/10.1007/978-3-319-62067-1\\_39](https://doi.org/10.1007/978-3-319-62067-1_39)

- 10.7 Trace Concentrations, Operating Room Pollution,  
and Personnel Hazards – 187**
  
- 10.8 Comparative Pharmacokinetics of Inhaled Anesthetics – 187**
  - 10.8.1 Uptake and Partial Pressure Equilibrium – 187
  - 10.8.2 Elimination – 189
  
- 10.9 Questions and Answers – 189**
  - References – 191

**Key Points**

1. Inhalational agents are chemical compounds that possess general anesthetic properties and can be administered via inhalation. The contemporary agents available for clinical use include nitrous oxide and the volatile agents: halothane, isoflurane, desflurane, and sevoflurane.
2. Currently, the mechanics of inhaled volatile anesthetics are believed to occur through a combined effect by prolongation of inhibitory effects (GABA<sub>A</sub> and glycine receptors) and inhibition of excitatory effects. This has minimized the belief in the Meyer and Overton theory that proposed the lipid membrane was the primary site of anesthetic action.
3. Volatile anesthetics decrease mean arterial pressure in a dose-dependent manner, which varies with the type of agent used. All volatile agents depress ventilation and blunt responses to changes in PaCO<sub>2</sub>.
4. The minimum alveolar concentration (MAC) of an inhaled anesthetic is the alveolar concentration at which 50% of patients will not show a motor response to a standardized surgical incision.
5. Inhalational anesthetics undergo biotransformation to many different degrees and locations depending primarily on their lipophilicity and clinical stability. The major organs involved in biotransformation, the liver and kidneys, are exposed to the highest metabolite concentrations; and therefore, are the primary sites of toxicity
6. The single most important factor in determining the speed of induction and recovery for inhalational agents is the blood:gas coefficient, which expresses the agent's distribution between the blood and gas at the same partial pressure. The higher the agent's solubility in the blood, the slower its induction rate.

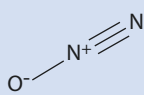
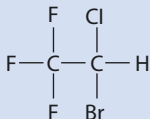
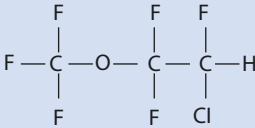
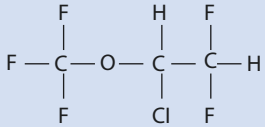
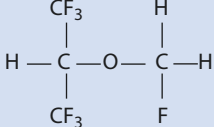
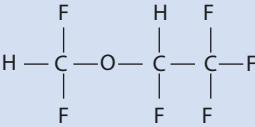
**10.1 Introduction**

Inhalational agents are used in anesthesia primarily to produce a loss of consciousness, but may have other effects such as muscle relaxation and analgesia. Inhalational anesthetics have been in use since the 1840s, when agents such as ether, chloroform, and nitrous oxide were introduced. Due to safety issues, the search for better inhalational agents was begun and fluorinated ethers and hydrocarbons were introduced. Halothane was introduced into clinical practice in 1956 and revolutionized anesthetic practices. However, secondary to its arrhythmogenic effects with epinephrine and possible postoperative liver failure, alternative agents were developed to minimize negative effects. Enflurane, a methyl ether derivative, was not arrhythmogenic or hepatotoxic, but had side effects including lowering the seizure threshold. With further research, the modern inhalational agents of fluorinated ethers including isoflurane, sevoflurane, and desflurane were introduced by the 1980s. These drugs resist metabolism and make organ toxicity unlikely. As research continues, the noble gas, xenon, has a potential for future development. These agents have improved safety and reliability.

**10.2 Physical and Chemical Properties of Inhaled Anesthetics****10.2.1 Nitrous Oxide**

Nitrous oxide (■ Fig. 10.1) is a low-molecular weight inorganic gas that is odorless and colorless. Although it is non-explosive, it does support combustion [1]. Because of its low potency and poor blood solubility (■ Table 10.1), it is commonly administered in conjunction with volatile anesthetics or narcotics to produce general anesthesia. Nitrous oxide has

■ Fig. 10.1 Molecular structure of inhalational anesthetics

| Nitrous Oxide   | Halothane  | Enflurane   |
|---|--|---|
|  |  |  |
| Isoflurane  | Sevoflurane  | Desflurane  |
|  |  |  |

**Table 10.1** Physical and chemical properties of inhaled anesthetics

|                                 | Nitrous Oxide | Halothane | Methoxyflurane | Enflurane | Isoflurane | Desflurane | Sevoflurane |
|---------------------------------|---------------|-----------|----------------|-----------|------------|------------|-------------|
| Molecular weight                | 44            | 197       | 165            | 184       | 184        | 200        | 168         |
| Boiling point (°C)              | −89           | 51        | 105            | 57        | 48         | 23         | 59          |
| Blood:gas coefficient @ 37 °C   | 0.47          | 2.5       | 12             | 1.9       | 1.4        | 0.45       | 0.65        |
| Oil: gas coefficient @ 37 °C    | 1.3           | 197       | 950            | 99        | 97         | 19         | 53          |
| Vapor pressure (mmHG @ 20 °C)   | Gas           | 244       | 22.5           | 172       | 240        | 669        | 170         |
| MAC % (30–55 year old at 1 atm) | 104           | 0.75      | 0.2            | 1.63      | 1.17       | 6.0        | 2.0         |
| Stable in soda lime             | Yes           | No        | Yes            | Yes       | Yes        | Yes        | No          |

Sources: Bovill [2] and Yasuda et al. [3]

the lowest potency of the inhalational agents, with a minimum alveolar concentration (MAC) value of 104%, which is clinically not achievable and thus cannot provide general anesthesia when administered solely. Nitrous oxide is a gas at room temperature, but can be stored as a liquid under pressure as its critical temperature lies above room temperature. Although it does have amnestic and analgesic properties, it does not provide muscle relaxation as other inhalational agents do. Controversy exists over the role of nitrous oxide in postoperative nausea and vomiting (PONV), which may occur through activation of the chemoreceptor trigger zone and vomiting center in the medulla [4].

### 10.2.2 Halothane

Halothane is a halogenated alkane (Fig. 10.1) that remains a clear liquid at room temperature. Carbon-fluoride bonds make it nonflammable and non-explosive. It is well tolerated for inhalational inductions with a notable sweet, non-pungent odor. It has a high potency and intermediate solubility, which allows for an intermediate onset and recovery from anesthesia (Table 10.1). Thymol must be added to halothane which also must be stored in amber bottles to prevent spontaneous oxidative decomposition [5, 6].

### 10.2.3 Enflurane

Enflurane is a halogenated ether (Fig. 10.1) with an ethereal odor that remains a liquid at room temperature. It has an intermediate solubility and high potency allowing for intermediate onset and recovery times from anesthesia (Table 10.1). Enflurane is oxidized in the liver and can produce nephrotoxic fluoride ions [7, 8].

### 10.2.4 Isoflurane

Isoflurane is a fluorinated methyl ethyl ether (Fig. 10.1) that is a nonflammable liquid at room temperature. Although it is an isomer of enflurane, it has different physiochemical properties and different manufacturing methods. Like enflurane and halothane, it has an intermediate solubility and high potency allowing for intermediate onset and recovery times from anesthesia (Table 10.1). It has a pungent ethereal odor [8].

### 10.2.5 Sevoflurane

Sevoflurane is a fluorinated methyl isopropyl ether (Fig. 10.1). It has a potency similar to enflurane. However, it has a significantly lower solubility in blood (Table 10.1). This property allows for a rapid increase in alveolar concentration and a rapid on and offset of anesthesia. Combined with its nonpungent odor, these attributes make sevoflurane an ideal inhalational induction agent. Its vapor pressures allow for the use of a conventional vaporizer. Sevoflurane is susceptible to metabolism, with 3–5% undergoing biodegradation. Unlike other volatile agents, sevoflurane is not metabolized to acyl halide intermediates (as with halothane, enflurane, isoflurane, and desflurane), which can potentially cause hepatotoxicity or cross-sensitivity between drugs.

### 10.2.6 Desflurane

Desflurane is also a fluorinated methyl ethyl ether (Fig. 10.1) that differs from isoflurane only by a substitution of a fluoride for the chlorine atom. The “minor change” of fluorination increases the vapor pressure, enhances molecular stability,

and decreases the potency of the drug [9]. Because of its vapor pressure (Table 10.1), desflurane will boil at room temperature at high altitudes. This requires a vaporizer (Tec 6, GE Healthcare, Chicago, IL) designed specifically to handle this inhalational agent. The vaporizer is heated to 39°C and pressurized to 2 atm. No fresh gas flows through the vaporizer pump; rather pure desflurane vapor joins the fresh gas flows before exiting the vaporizer [10]. Low solubility and potency allow for a rapid on and offset of anesthesia. Its lower blood-gas solubility creates precise control and the ability for more rapid recovery times from anesthesia [8]. Desflurane has a pungent odor, which limits its utility for inhalational inductions.

### 10.2.7 Xenon

Xenon is a noble gas found in the atmosphere; and was recognized as an anesthetic in 1951. It has a MAC value of 71% and can be combined with oxygen to deliver anesthesia. The blood:gas partition coefficient is 0.12, which results in rapid onset and recovery. Xenon depresses post-sympathetic excitatory transmission through N-methyl-D-aspartate (NMDA) receptor blocks. There are minimal cardiovascular side effects, even in the setting of severely limited myocardial reserve. Xenon affects anesthetic-induced preconditioning of the heart and brain against ischemic damage in the same way as volatile agents. Xenon may have neuroprotective action, but it may be offset by an increase in cerebral blood flow. It is a non-irritant to the airway for easy induction. Although a mild respiratory depressant, it decreases respiratory rate and increases tidal volume, in contrast to the volatile agents. Xenon has a high relative density, which causes an increase in pulmonary resistance. Caution is advised in patients who have severe chronic obstructive pulmonary disease (COPD) or in premature infants. It is not metabolized in the liver or kidneys and it does not trigger malignant hyperpyrexia. Xenon is also a potent intraoperative analgesic, attenuating responses to surgical stimuli to a greater extent than sevoflurane.

Xenon anesthesia provides more stable intraoperative blood pressure, lower heart rate, and faster recovery from anesthesia than volatile agents. However, it is associated with higher postoperative nausea and vomiting. The main limitations for wider use are lack of studies, need for hyperbaric conditions, impracticality in surgery, and inefficiency of conventional anesthesia equipment. These limitations make xenon cost prohibitive.

### 10.3 Mechanism of Action

The exact mechanism of action for volatile anesthetics is complex and still unknown. Currently, the mechanics of inhaled volatile anesthetics is believed to occur through a combined effect by prolongation of inhibitory effects (GABA<sub>A</sub> and glycine receptors) and inhibition of excitatory effects

(nicotinic acetylcholine and glutamate receptors). Typical anesthetic agents produce anesthesia, amnesia, analgesia, and immobilization.

Initially, Meyer and Overton proposed a lipid theory and believed the lipid membrane was the primary site of anesthetic action by correlating inhaled anesthetics potency with their solubility in lipids. They observed a strong correlation between the potency of inhalational anesthetics and their solubility in oil, theorizing they had a nonspecific lipid membrane mechanism of action [11]. Later, researchers demonstrated that proteins may also be the site of action for inhaled anesthetics [12, 13]. Additional research on the mechanism of action for inhaled anesthetics explained ligand gated ion channels proteins are mostly likely the targets of inhaled anesthetics [14].

Electrical activity in human cells is generated through influx and efflux of ions (mostly Na<sup>+</sup>, Ca<sup>2+</sup>, Cl<sup>-</sup> and K<sup>+</sup>) through a variety of ion channels. Some receptor-mediated ion channels are targets of inhaled anesthetics at clinical anesthetic concentrations, such as serotonin receptors, GABA<sub>A</sub> receptors, glycine receptors, and NMDA or AMPA (α[alpha]-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid) receptors (glutamate neurotransmitter) [15–18].

GABA<sub>A</sub>-related anesthetic action is common for all volatile anesthetics due to its abundance in the brain. Normal physiologic function of GABA<sub>A</sub> and glycine receptors (Cl<sup>-</sup> ion channels) is to inhibit the excitation of postsynaptic neurons. At effective clinical concentrations, volatile anesthetics enhance the GABA<sub>A</sub> receptor-mediated activity by increasing its sensitivity to gamma-aminobutyric acid (GABA) and the sensitized receptors prolong the inhibition of excitatory neurons. Gaseous anesthetics, such as nitrous oxide, have a minimal effect on GABA-related mechanisms [19–23]. Normally K<sup>+</sup> channels maintain a polarized state of neurons and are targeted sites for isoflurane. Isoflurane activates the K<sup>+</sup> channel and leads to a decrease of neuronal excitation [24].

Inhibition of excitatory neurotransmission can be achieved either by inhibition of a neurotransmitter release from presynaptic nerve endings or a postsynaptic receptor blockade. Halothane and isoflurane at clinical concentrations inhibit the NMDA receptor (Na<sup>+</sup> ion channel) associated excitation by postsynaptic blockades or decreasing presynaptic glutamate release. The volatile anesthetics also can inhibit the presynaptic release of an excitatory neurotransmitter by blocking presynaptic voltage-gated Na<sup>+</sup> channels at clinical concentrations [25]. Excitatory postsynaptic nicotinic acetylcholine receptors and NMDA-sensitive glutamate channels are inhibited by gaseous anesthetics to inhibit the excitation of excitatory neurons [26].

The mechanism of action for immobilization and amnesia occurs at distinct sites. Studies have proven that immobilization to surgical stimulus can be achieved at the spinal cord level without brain involvement. Immobility to surgical stimuli occurs by inhibiting ascending transmission of pain stimuli to the brain from the spinal cord. At the spinal cord level, volatile anesthetics prolong the inhibitory effects of glycine receptors and inhibit the postsynaptic excitatory effects

of NMDA and AMPA receptors [27–29]. Amnesia can be induced without immobilization at lower clinical concentrations. Specific loci in the brain are responsible for this amnesic effect. Inhaled anesthetics act on nicotinic acetylcholine receptors in the brain and impair the memory process leading to amnesia [30–33].

GABA receptors function differently between the growing brain in children and the brain in adults. In the growing brain, GABA receptors function as stimulators and in the adult brain they act as inhibitors; therefore, a neurotoxic effect from anesthetics can be seen in the growing brain. In contrast to its inhibitory action in the adult brain, GABA receptors act as an excitatory neurotransmitter in the growing brain of the child. These GABA receptors generate action potentials directly opening voltage-dependent calcium channels and increase the calcium concentration in the brain. This increase in intracellular calcium can lead to apoptosis. In addition, the mitochondrion appears to be the mediator between anesthesia-induced increased calcium levels and cell apoptosis, leading to mitochondrial damage. Every year,

millions of children are treated with anesthetic agents. There is evidence that suggests that exposure to anesthetics may be neurotoxic to the developing brain and lead to long-term neurological effects.

Lithium protects against anesthesia-induced developmental neuroapoptosis along with melatonin. Coadministration of hydrogen gas acts as part of the carrier gas mixture and may suppress neuronal apoptosis. Therefore, there may not be a safe anesthetic, but only safe anesthetic concentrations and exposure durations.

#### 10.4 Systemic Effects of Inhaled Anesthetics

Inhaled anesthetics have several effects on systemic organs. Anesthetics effects on the central nervous system, cardiovascular system, pulmonary function, neuromuscular junction, renal and liver function, and hematology and immune systems have been described in sub-sections and are summarized in [Table 10.2](#).

**Table 10.2** Systemic effects of inhaled anesthetics

|                          | Nitrous oxide  | Halothane      | Methoxyflurane | Enflurane | Isoflurane | Desflurane     | Sevoflurane |
|--------------------------|----------------|----------------|----------------|-----------|------------|----------------|-------------|
| Cardiovascular           | ↔              | ↓              | ↓              | ↓         | ↓          | ↓              | ↓           |
| HR                       | ↔              | ↓              | ↑              | ↑         | ↑          | ↑ <sup>b</sup> | ↔           |
| SVR                      | ↔              | ↔              | ↔              | ↓         | ↓          | ↓              | ↓           |
| CO                       | ↑ <sup>a</sup> | ↓              | ↓              | ↓         | ↔          | ↔              | ↓           |
| Myocardial contractility | ↔              | ↓              | ↔              | ↓         | ↔          | ↔              | ↔           |
| Respiratory              | ↓              | ↓              | ↓              | ↓         | ↓          | ↓              | ↓           |
| TV                       | ↑              | ↑              | ↑              | ↑         | ↑          | ↑              | ↑           |
| RR                       | ↔              | ↑              | ↑              | ↑         | ↑          | ↑              | ↑           |
| PaCO <sub>2</sub>        | ↑              | ↔              | ↔              | ↔         | ↔          | ↔              | ↔           |
| PVR                      | ↓              | ↓↓             | ↓              | ↓         | ↓          | Irritant       | ↓           |
| Airway resistance        |                |                |                |           |            |                |             |
| Cerebral                 | ↑              | ↑↑             | ↑              | ↑         | ↑          | ↑              | ↑           |
| Blood flow               | ↑              | ↑              | ↑              | ↑         | ↑          | ↑              | ↑           |
| ICP                      | ↑              | ↑ <sup>a</sup> | ↓              | ↓         | ↓          | ↓              | ↓           |
| CMRO <sub>2</sub>        | ↓              | ↓              | ↓              | ↑         | ↓          | ↓              | ↓/↑         |
| Seizures                 |                |                |                |           |            |                |             |
| Nondepolarizing blockade | ↑              | ↑              | ↑              | ↑         | ↑          | ↑              | ↑           |
| Renal                    | ↓              | ↓              | ↓              | ↓         | ↓          | ↓              | ↓           |
| Blood flow               | ↓              | ↓              | ↓              | ↓         | ↓          | Unknown        | Unknown     |
| GFR                      | ↓              | ↓              | ↓              | ↓         | ↓          | Unknown        | Unknown     |
| Urine                    |                |                |                |           |            |                |             |
| Hepatic                  | ↓              | ↓              | ↓              | ↓         | ↓          | ↓              | ↓           |
| Blood flow               |                |                |                |           |            |                |             |

*Abbreviations:* HR heart rate, SVR systemic vascular resistance, CO cardiac output, TV tidal volume, RR respiratory rate, PaCO<sub>2</sub> partial pressure of carbon dioxide, PVR pulmonary vascular resistance, ICP intracranial pressure, CMRO<sub>2</sub> cerebral metabolic rate of oxygen, GFR glomerular filtration rate

<sup>a</sup>Minimal

<sup>b</sup>With rapid change in inhaled concentration

### 10.4.1 Effects on Central Nervous System

The changes in an electroencephalogram (EEG) are noticed after the induction of inhaled anesthetics. At lower clinical concentrations (low MAC), the volatile anesthetics and gaseous anesthetics produce high frequency and low amplitude (Beta waves) waves in the EEG, and are transformed to low frequency and high amplitude waves (Delta waves) at clinical anesthetic concentrations [33]. Some volatile anesthetics, such as isoflurane and desflurane at 1.5–2 MAC anesthetic concentration, cause electrical silence in the EEG [34].

Enflurane has the tendency to induce convulsions (seizures) to decreased PaCO<sub>2</sub>, MAC >2, and repetitive auditory stimuli [35]. Isoflurane has anti-convulsive properties, and desflurane does not produce seizures [36, 37]. There are case reports that support that sevoflurane can produce seizure activity [38, 39].

Typically, cerebral blood flow is autoregulated and depends on the cerebral oxygen consumption (CMRO<sub>2</sub>), and PaCO<sub>2</sub>. All inhaled anesthetics increase cerebral blood flow in a dose-dependent manner despite a decrease in cerebral oxygen consumption. Inhalation agents also partially preserve the autoregulation of CBF to changes in PaCO<sub>2</sub>. Desflurane and isoflurane preserve the responsiveness of CBF to changes in PaCO<sub>2</sub> [40]. Cerebral metabolic oxygen requirements are dose-dependent and are decreased with volatile anesthetics [41]. Increased intracranial pressure is seen with halothane use due to significant increases in cerebral blood flow compared to other inhaled anesthetics [42].

Preconditioning and postconditioning is a mechanism for inhaled anesthetics for neuroprotective effects. Inhalational anesthetics provide neuroprotective effects against brain ischemia by pre-, pro- and post-conditionings. Preconditioning is a process where a relatively small amount of inhalational agent is administered prior to the ischemic insult. Postconditioning is applied after the cerebral ischemic event has developed. Many studies have confirmed the protection of pre- and post-conditioning of inhalational anesthetics in their neuroprotection against cerebral ischemia. Sevoflurane preconditioning and early postconditioning reduced both cerebral infarct size and neurological defect score at 24 h of reperfusion. Pretreatment with sevoflurane or its early administration at reperfusion provided neuroprotection via mitoKATP in a rat model for focal cerebral ischemia.

Although sevoflurane and isoflurane are similar in their systemic effects, they appear to differ in cerebral circulation. Sevoflurane can maintain cerebral autoregulation up to 1.5 MAC; whereas, isoflurane results in loss of autoregulation. Thus, cerebral autoregulation is better preserved during 1.5 MAC sevoflurane than isoflurane and is a better neuroanesthetic agent.

### 10.4.2 Effects on Cardiovascular System

Volatile anesthetics decrease mean arterial pressure in a dose-dependent manner, which varies with the type of agent used. At clinical anesthetic concentration, halothane decreases the mean arterial pressure by decreasing myocardial contractil-

ity and cardiac output; whereas, isoflurane, sevoflurane, and desflurane decrease systemic vascular resistance. Enflurane decreases both systemic vascular resistance and cardiac output. The change in heart rate is variable with type of agent used and the type of pharmacological agent administered during surgery. The decrease in heart rate is observed with halothane use due to suppression of the carotid sinus to changes in systemic blood pressure and rate of sinus node depolarization. At anesthetic concentrations, the heart rate is increased with desflurane, enflurane, sevoflurane (>1 MAC) and isoflurane use [41, 43]. Nitrous oxide has very little effect in mean arterial pressure and heart rate changes [44, 45]. Significant decrease in cardiac output is noticed with halothane and enflurane, and sevoflurane can decrease cardiac output at MAC between 1 and 1.5. Sevoflurane can prolong the QT interval and should be cautiously used in patients with prolonged QT interval syndrome or patients susceptible to QT interval changes [46]. Volatile agents can induce arrhythmias. Halothane and isoflurane sensitize the heart to epinephrine, compared to desflurane and sevoflurane, and cause cardiac arrhythmias [47, 48]. Coronary steal syndrome, wherein normally responsive coronary anoles are dilated “stealing” blood from vessels supplying ischemic zones, may be associated with isoflurane [49]. Isoflurane is known to be a potent coronary artery vasodilator. Isoflurane-induced coronary artery vasodilatation can lead to redistribution of coronary blood flow away from diseased areas, which have decreased ability to vasodilate. Thereby, blood is redistributed in greater amounts to areas with normally responsive coronary arteries. However, most clinical studies failed to prove a higher incident of myocardial ischemia due to isoflurane. Sevoflurane and desflurane do not cause coronary steal syndrome.

### 10.4.3 Effects on Respiration

All volatile agents depress ventilation and blunt responses to changes in PaCO<sub>2</sub>. Volatile agents cause rapid, shallow breathing. There is a reduction in the tidal volumes and minute ventilation. The increase in respiratory rate does not adequately compensate for the amount of tidal volume decrease and hence causes an increase in PaCO<sub>2</sub>.

Volatile agents reduce minute ventilation by reducing tidal volumes. Reduced tidal volume causes a slight increase in PaCO<sub>2</sub>. The agents minimally suppress the responsiveness to increased PaCO<sub>2</sub> (hypercapnia) from decreased tidal volume at central medullary respiratory centers [50, 51]. Nitrous oxide has very little effect in ventilation depression, bronchial muscle tone, and in hypoxic drive [52]. Increases in respiratory rate are associated with all volatile anesthetics. Halothane, isoflurane and sevoflurane decrease airway resistance in COPD and asthmatic patients [53]. Due to low airway irritant effects, nitrous oxide, halothane and sevoflurane can be used for induction of anesthesia. Isoflurane and desflurane can irritate the airways during induction with MAC greater than 1.5 and 1, respectively, but have little or no effect during the maintenance of anesthesia. Desflurane is a pungent gas that

can cause airway irritability during induction, manifested as breath-holding, salivation, coughing, and possibly laryngospasm. Small doses of opioid administration and humidification help to reduce irritant properties [54–56].

#### 10.4.4 Effects on Neuromuscular Junction

Volatile anesthetics enhance the effects of neuromuscular blocking drugs by inhibiting nicotinic acetylcholine receptors [57]. Volatile anesthetics produce dose-dependent muscle relaxation; whereas, nitrous oxide can cause skeletal muscle rigidity (>1 MAC) [58]. One potential complication of volatile agents is malignant hyperthermia (MH). Succinylcholine administration with a volatile agent potentiates a patient susceptible to MH. Malignant hyperthermia can occur even without succinylcholine administration in genetically susceptible patients [59–61]. Halothane has a higher tendency to produce MH than other volatile agents. MH can appear hours after uneventful anesthesia with desflurane [62] and sevoflurane [63, 64]. Nitrous oxide does not manifest this complication [11].

#### 10.4.5 Effects on Renal Function

Volatile agents have little effect on renal physiology. The decrease in renal blood flow that is clinically observed is a product of their glomerular filtration rate and urine output is systemic vascular effects. There is no direct effect of inhalational agents on renal blood flow. Inorganic fluorides and metabolites, such as compound A, produced from the metabolism of volatile anesthetics can be nephrotoxic; and these effects are further discussed in the next section (Biotransformation and Toxicity of Inhaled Anesthetics).

#### 10.4.6 Effects on Hepatic Function

All inhaled anesthetics reduce the hepatic blood flow. Severe hepatic injury following volatile anesthetics administration is very rare, with a ratio of 1:10,000,000 [65]. Anesthetics agents interfere with hepatic metabolism of other pharmacological agents that are administered during the anesthesia [66, 67]. Hepatotoxicity can occur with inhaled anesthetics due to inadequate hepatic oxygenation from reduced hepatic blood flow. Hepatotoxicity incidences are higher with halothane induction compared with other inhaled anesthetics. These effects are further discussed in the next section (Biotransformation and Toxicity of Inhaled Anesthetics).

#### 10.4.7 Effects on Hematologic and Immune Systems

Prolonged exposure to nitrous oxide can interfere with bone marrow function. Nitrous oxide affects DNA synthesis by inhibiting vitamin B<sub>12</sub> dependent enzymes (methionine synth-

etase) [68, 69]. Megaloblastic changes are noticed in patients who receive nitrous oxide for a duration of 24 h. Agranulocytosis occurs in patients with 4 days or longer exposure to nitrous oxide. Volatile anesthetics have an immunosuppressive effect on both innate immunity (neutrophils, NK cells, and macrophages) and cell-mediated immunity (T-cells and B-cells), and are dose-dependent. Volatile agents impair neutrophil, macrophage, dendritic, and T-cell function. The suppressive action on immunity is from a combined exposure of the patient to surgery and anesthesia. Surgery releases stress hormones (catecholamines and corticosteroids) [70]. Inhaled anesthetics inhibit the actions of polymorphonuclear cells such as chemotaxis and phagocytosis. Sevoflurane and isoflurane can induce dose-dependent apoptosis in lymphocytes. Isoflurane and sevoflurane also reduce the expression of adhesion molecules on lymphocytes and macrophages; and thus, decreases the recruitment and accumulation of immune cells at inflammatory sites [71].

### 10.5 Biotransformation and Toxicity of Inhaled Anesthetics

Inhalational anesthetics undergo biotransformation to many different degrees and locations depending primarily on their lipophilicity and clinical stability. The major organs involved in biotransformation, the liver and kidneys, are exposed to the highest metabolite concentrations, and therefore, are the primary sites of toxicity (see ■ Table 10.3).

#### 10.5.1 Nitrous Oxide

Nitrous oxide undergoes very little biotransformation (0.004%) and is almost solely eliminated by exhalation during emergence [8]. Anaerobic bacteria in the gastrointestinal (GI) tract are responsible for the minimal amount of metabolism. Nitrous oxide irreversibly oxidizes the cobalt atom in vitamin B<sub>12</sub>, including methionine synthetase and thymidylate synthetase. These enzymes are responsible for myelin formation and DNA synthesis; and thus, nitrous oxide has been questioned to cause bone marrow suppression and neurologic deficiencies in prolonged usage.

#### 10.5.2 Halothane

The liver is the primary site of biotransformation and metabolism for most drugs, particularly lipophilic drugs such as halothane [74, 75]. Approximately 25% of administered halothane is oxidized by an isoenzyme of P450 (CYP2E1) into its principal metabolite trifluoroacetic acid (TFA), as well as lesser amounts of bromide and chloride [76]. The TFA metabolites react with tissue proteins to form trifluoroacetylated protein adducts. Clinical exposure to halothane results in 2 distinct types of hepatitis [77–79]. Type I hepatotoxicity is benign and self-limiting and occurs in 25–30% of patients



**Table 10.3** Biotransformation and toxicity of inhaled anesthetics

|   | Nitrous oxide  | Halothane                                     | Methoxyflu-<br>rane                         | Enflurane                                   | Isoflurane                                  | Desflurane   | Sevoflurane  |
|---|--|---|---|---|---|--|--|
| Tissue metabolism (%)   | 0.004  | 25  | 70  | 2.5   | 0.2   | 0.02   | 5  |
| Oxidating enzymes   | Anaerobic bacteria in gastrointestinal tract                   | CYP2E1, CYP2A6                                | CYP2E1, CYP1A2, CYP2C9/10, CYP2D6           | CYP2E1                                      | CYP2E1                                      | CYP2E1   | CYP2E1   |
| Principal metabolite  | Inactivation of methionine synthetase                          | TFA, bromine, chloride                        | Oxalic acid, free fluoride                  | TFA, small fluoride level rise              | TFA, small fluoride level rise              | TFA (fluoride levels unchanged from pre-anesthetic levels) | Fluoride   |
| Trifluoroacetylated hepatocellular protein degree of modification | None   | +++++   | None  | ++  | +   | +  | None   |
| CO <sub>2</sub> stability   | Yes  | CO from desiccated carbon dioxide absorbent   | CO from desiccated carbon dioxide absorbent | CO from desiccated carbon dioxide absorbent | CO from desiccated carbon dioxide absorbent | CO from desiccated carbon dioxide absorbent                | Compound A from desiccated carbon dioxide absorbent, heat production |
| Possibly toxicities   | DNA synthesis, Bone marrow suppression, Vitamin B12 deficiency | <b>Hepatic</b> (1:20,000 fulminant hepatitis) | <b>Renal</b> , hepatic                      | Hepatic (1:300,000 fulminant hepatitis)     | Hepatic (rare fulminant hepatitis)          | Hepatic (rare fulminant hepatitis)                         | Hepatic (few case reports fulminant hepatitis)                       |

Sources: Yasuda et al. [72, 73]

Abbreviations: TFA trifluoroacetic acid, CO<sub>2</sub> carbon dioxide, CO carbon monoxide

receiving halothane. Symptoms include transient nausea, fever, and serum transaminase levels. “Halothane hepatitis,” or type II hepatotoxicity, has been reported in 1:5000 to 1:35,000 cases of halothane administration. This immune-mediated reaction is believed to result from the trifluoroacetylated protein adducts in the liver. Clinical symptoms of halothane hepatitis include fever, eosinophilia, and jaundice. Laboratory findings include elevated serum alanine and aspartate transferase and elevated bilirubin. Patients also have a positive IgG against TFA. Severe cases are associated with centrilobular necrosis that may lead to fulminant liver failure with a mortality rate of 50% [80]. Higher rates of halothane hepatitis are found in patients exposed to multiple halothane anesthetics in a short period of time, obese patients, patients >50 years old, female patients, and patients with a history of postanesthetic fever or jaundice.

### 10.5.3 Methoxyflurane

As the most lipophilic inhaled anesthetic, methoxyflurane undergoes the most biotransformation at an estimated 70% of

the drug administered [81]. Only a small amount of the drug, taken into body tissue, is exhaled and respiratory clearance from muscle and fat can extend over a period of several days. Methoxyflurane is metabolized in both the kidneys and the liver, and inorganic fluoride (F<sup>-</sup>) is produced during its metabolism in clinically significant quantities [82, 83]. Many studies have demonstrated direct links between methoxyflurane dosages, metabolism, and fluoride production. Inorganic fluoride likely causes renal injury with a nephrotoxic threshold of 50 μ(mu)mol/L [84]. Methoxyflurane, the first modern halogenated ether anesthetic, is no longer in clinical use because it is now known to produce polyuric renal insufficiency. More recent anesthetics have been cautiously studied for their renal impairment and fluoride production abilities [85].

### 10.5.4 Isoflurane

The minimal metabolism (0.2%) of isoflurane results in extremely low rates of hepatic or renal impairment. TFA is the primary metabolite, but serum fluoride levels have not been shown to cause renal dysfunction [86].

### 10.5.5 Desflurane

Desflurane undergoes extremely low metabolism rates in humans (0.02%); and thus, the serum and urine fluoride levels are essentially unchanged from pre-anesthetic levels. More than the other volatile agents (desflurane > enflurane > isoflurane), desflurane is susceptible to degradation in desiccated carbon dioxide absorbency to carbon monoxide, when water content falls below 1.4% for soda lime and 5% for baralyme. This carbon monoxide can lead to increased levels of blood carboxyhemoglobin [87–89].

### 10.5.6 Sevoflurane

Inorganic fluoride ions in plasma concentrations greater than enflurane and hexafluoroisopropanol are produced during the metabolism of sevoflurane in humans. The overall rate of sevoflurane metabolism is more than 10 times that of isoflurane (5%), clinically producing higher serum fluoride levels. Despite this, clinical studies have demonstrated no clinical nephrotoxicity with sevoflurane administration, even with peak concentrations of 50  $\mu$ (mu)mol/L. Production of fluoride ions of sevoflurane is mainly in the liver and, therefore, has minimal effect on the kidney function. The liver metabolizes 2–5% of the sevoflurane. Typical fluoride levels after 2–3 MAC hours are 20–30  $\mu$ (mu)mol/L [90]. Because of sevoflurane's low blood:gas solubility and rapid elimination, fluoride concentrations fall quickly and renal toxicity is not clinically present. In the presence of a strong alkali, such as those in carbon dioxide absorbents, sevoflurane has been shown to degrade to compounds toxic to animals, particularly compound A (fluoromethyl-1, 1-difluoro-1(trimethyl) vinyl-ether) [91–93]. Larger amounts of compound A are produced with lower gas flows, increased respiratory temperatures, high sevoflurane concentrations, anesthetics of long duration and desiccated soda lime. Amsorb® (Armstrong Ltd., Coleraine, Northern Ireland) is a newer absorbent that does not contain strong base and does not form CO or compound A in vitro. It is clinically recommended to maintain fresh gas flows greater than 2 L/min to limit possible compound A production. Despite proven nephrotoxicity in rats, no postoperative renal impairment or injury has been seen in humans. This difference may be secondary to the lower  $\beta$ (beta)-lyase activity in humans [94]. Degradation of sevoflurane to hydrogen fluoride in the presence of metal and environmental impurities can also occur. Hydrogen fluoride can cause respiratory mucosal burns. Degradation is inhibited through the addition of water in manufacturing and packaging in plastic containers [90]. The US Food and Drug Administration (FDA) recommends the use of sevoflurane with fresh gas flow rates at least 1 L/min for exposure up to 1 h and at least 2 L/min for exposures greater than 1 h.

### 10.5.7 Possible Neurotoxicity of Inhaled Anesthetics

The possibility of neurotoxic effects of inhaled and other general anesthetics does exist in patients of extreme ages [95, 96]. The greatest concern is the use of general anesthetics in the youngest patients, where rapid brain development is occurring. Widespread neuronal apoptosis in 7-day-old rats after exposure to midazolam, isoflurane, and nitrous oxide caused lasting deficits in behavior, learning, and memory centers [97, 98]. Continued research has demonstrated, in nonhuman species including primates, sensitive periods of early brain development when anesthetics can accelerate apoptosis [99]. Clinical studies in humans have demonstrated mixed results. Although a possible association with multiple anesthetic exposure and impaired neurocognitive development was demonstrated in one study, others have found no cognitive outcome differences [100–102]. Ongoing clinical trials will provide more information on this important issue and additional information is available at smarttots.org.

### 10.6 Minimum Alveolar Concentration and Its Affecting Factors

Minimal alveolar concentration of an inhaled anesthetic is defined as the alveolar concentration at which 50% of the patients are immobile to a standard surgical incision at 1 atmospheric pressure [62, 103]. The immobility is achieved in 99% of patients with 1.3 MAC. This is the state where somatic responses are lost. The potency of the anesthetics is measured by MAC. Anesthetics with higher MAC have a lower potency (eg, N<sub>2</sub>O) and vice versa. As the MAC of nitrous oxide exceeds 100%, it cannot be used alone to provide general anesthesia. It is typically combined in a 70% concentration with 30% oxygen and in concert with more potent agents. The MAC values are additive when used in combinations. The MACs of different inhalational anesthetics are summarized in ■ Table 10.1. Numerous factors are involved in affecting MAC values and they are described in ■ Table 10.4 [104, 105].

“MAC-awake” is defined as the concentration at which response to the vertebral commands are lost in 50% of the patients. This is the state where amnesia occurs. Amnesia is observed before the immobility occurs. The MAC-awake values are significantly lower than the MAC values.

Many studies have demonstrated an age-related MAC value for the volatile agents. MAC is highest at 6 months of age, after which it begins to decline. After age 40, MAC declines ~6% per decade such that by 80 years of age, MAC is about 0.75 that of a 40-year-old.

**Table 10.4** Factors affecting minimum alveolar concentrations (MAC)

| Factors decreasing MAC requirements     | Factors increasing MAC requirements   |
|---|---------------------------------------|
| Decreased catecholamine levels in CNS   | Increased catecholamine levels in CNS |
| Hyponatremia                            | Hypernatremia                         |
| Hypothermia                             | Hyperthermia                          |
| Older age (elderly)                     | Infants (greatest MAC at 6 months)    |
| Acute alcohol intoxication              | Chronic alcohol usage                 |
| Anemia (hematocrit <10%)                | Red hair (20% increase)               |
| Hypercarbia ( $P_{aCO_2} >95$ mmHg)     | Hyperthyroidism                       |
| Hypoxia ( $P_{aO_2} <40$ mmHg)          |                                       |
| Pregnancy (decreased by 30% at 8 weeks) |                                       |
| Drugs                                   | Drugs                                 |
| Opioids                                 | Cocaine                               |
| Ketamine                                | Ephedrine                             |
| Benzodiazepines                         | MAO inhibitors                        |
| Clonidine, Dexmetomidine, Methylopa     | Amphetamine (acute)                   |
| Local Anesthetics                       |                                       |
| Lithium                                 |                                       |
| Amphetamines (chronic)                  |                                       |

*CNS* central nervous system, *PaCO<sub>2</sub>* partial pressure of carbon dioxide, *PaO<sub>2</sub>* partial pressure of oxygen, *MAO* monoamine oxidase

## 10.7 Trace Concentrations, Operating Room Pollution, and Personnel Hazards

Health care workers, surgeons, and anesthesiologists in operating theaters or anesthetizing locations are at risk of exposure to trace concentrations of anesthetics. Spontaneous abortions and congenital defects were observed in rats that were exposed to nitrous oxide for longer times [106, 107]. Even in humans, potential occupational associated risks are noticed after prolonged exposure to nitrous oxide. Spontaneous abortions and decreases in fertility occurred in female workers who were exposed to nitrous oxide in the absence of scavenging systems during nitrous oxide administration [108, 109]. Halogenated agents in vitro are embryolethal and produced teratogenic effects in animals [110–112]. In humans, halogenated agents can cause spontaneous abortions and there is some evidence they might produce congenital defects in the offspring of exposed pregnant women [112–114]. Appropriate safety measures, such as scavenging

systems and proper ventilation systems, must be taken in hospital operating rooms and anesthetizing locations to prevent occupational-related hazards, especially in females.

Nitrous oxide enhances the greenhouse effect just as carbon dioxide does, but is 300 times more potent, accounting for 6% of the heating effect and causing ozone depletion. In addition, inhaled anesthetics also contribute to global climate change. Isoflurane, sevoflurane, and desflurane undergo very little in vivo metabolism in clinical use, and upon exhalation these agents remain in a form that may pollute the environment. Whenever  $N_2O$  or a volatile anesthetic is administered, a continuous flow fresh air ventilation system or scavenger must be used to prevent waste gas accumulation (WGA). Health care facilities are accountable for ensuring that all anesthesia equipment, including the scavenging system, is properly maintained to promote a safe and healthy environment.

## 10.8 Comparative Pharmacokinetics of Inhaled Anesthetics

The pharmacokinetics of inhaled anesthetics describes their uptake from the alveoli into the systemic circulation, distribution in the body, and primary elimination by the lungs or liver metabolism. Although the mechanism of action of these agents remains hypothetical, their therapeutic effect ultimately depends on their tissue concentration in the central nervous system. The goal of delivering the inhaled anesthetic is to obtain an optimal brain partial pressure ( $P_{br}$ ) of the anesthetic. The alveolar partial pressure ( $P_A$ ), in equilibrium, mirrors the  $P_{br}$  and is used as an index of anesthetic depth. The pharmacokinetics can be influenced by aging and increases in body fat [115].

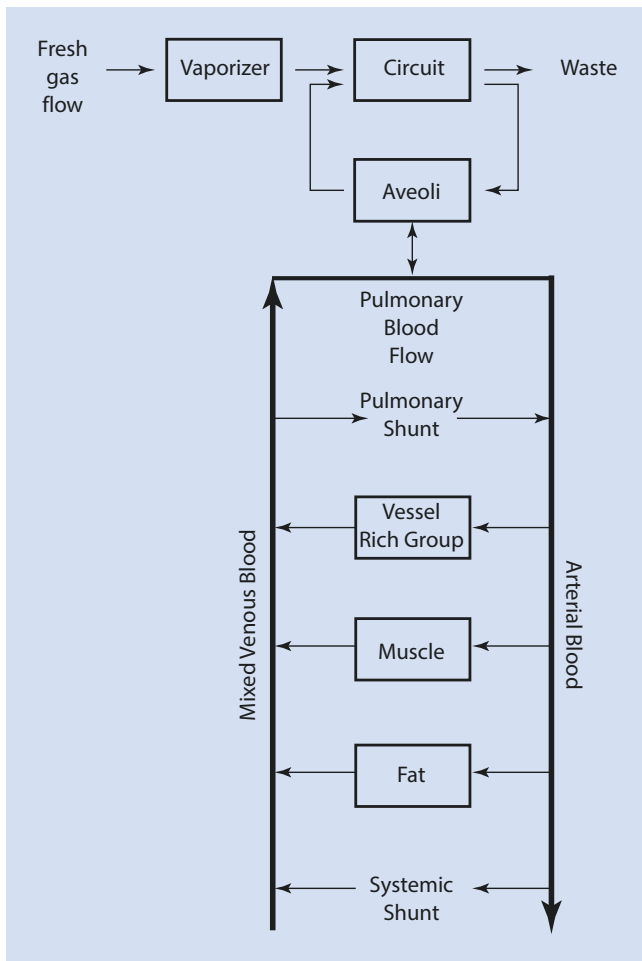
### 10.8.1 Uptake and Partial Pressure Equilibrium

There are many steps involved between the administration of the inhaled anesthetic from a vaporizer and its distribution into the central nervous system (see Fig. 10.2).

A series of partial pressure gradients drives the forward movement and systemic absorption of the gas. The principal objective is to achieve equal partial pressures on both sides of each single barrier in the gas flow.

$P_A$  (alveolar partial pressure)  $\leftrightarrow P_a$  (arterial partial pressure)  $\leftrightarrow P_{br}$  (brain partial pressure):

The alveolar partial pressure is dependent on the inspired pressure, ventilation, and breathing system components. This gradient begins with the inspiratory concentration of the gas leaving the anesthesia machine. This content depends on the concentration set by the vaporizer as well as the fresh gas flow, the volume of the breathing circuit, and the possible absorption of the gas by the circuit. Increasing ventilation promotes the input of anesthetics to offset the tissue uptake. The effect is a more rapid rate of increased in the  $P_A$  [117].



**Fig. 10.2** The uptake and distribution of inhaled anesthetics in the body (Adapted from Miller and Pardo [116])

The second component in the uptake of inhaled anesthetics is the alveolar gas concentration that is achieved in lung tissues during anesthesia. As the agent is taken up in the pulmonary blood stream during induction, the alveolar tissue concentrations remain less than in inspired concentrations. When the blood uptake of the agent is greater, its rate of rise in the alveolar gas is slower. Increasing the inspired concentration of an agent not only increases its alveolar concentration, but also its rate of rise ( $F_A/F_I$ ). This phenomenon is known as the concentration effect. The alveolar partial

pressure of an agent is important because it determines the partial pressure of the anesthetic in the blood and ultimately in the brain. Hence, the partial pressure in the brain is directly proportional to its brain tissue concentration and therefore its clinical effect.

The impact of a right-to-left shunt on the rate of increase in the  $P_a$  depends on the solubility of the anesthetic. A right-to-left shunt slows the rate of increase of the  $P_a$  of a poorly soluble anesthetic more than that of a soluble anesthetic. It appears unlikely that a right-to-left shunt alone will alter the speed of induction of anesthesia significantly. Left-to-right shunts result in delivery to the lungs of blood containing a higher partial pressure of anesthetic than that present in blood that has passed through tissues. As a result, left-to-right shunts offset the dilutional effects of a right-to-left shunt on the  $P_a$ .

$P_A$  (alveolar partial pressure)  $\leftrightarrow$   $P_a$  (arterial partial pressure)  $\leftrightarrow$   $P_{br}$  (brain partial pressure):

The uptake of the inhaled anesthetics from the alveoli into the pulmonary capillary blood depends on its solubility in body tissue (the partition coefficients), the cardiac output, and the alveolar-venous partial pressure difference [3, 118]. A slower rate of induction occurs if there is a greater uptake of the agent and a greater difference between the inspired and alveolar concentrations. The blood:gas partition coefficient is the single most important factor in determining the speed of induction and recovery (Table 10.5). A partition coefficient is a property of a chemical that describes its relative distribution at equilibrium given the same temperature, pressure, and volume. For anesthetics, the blood:gas coefficient is an important measure describing an inhalational agent's distribution between the blood and gas at the same partial pressure. A higher blood:gas coefficient correlates with higher blood solubility and thus a slower induction rate. A lower blood:gas coefficient transiently corresponds with a faster induction rate; for instance, nitrous, desflurane, and sevoflurane have faster induction rates than isoflurane and halothane (see Table 10.5). The second-gas effect states that a high volume of uptake of one gas will accelerate the rate of increase of the  $P_A$  of a simultaneously administered second gas. For instance, a high uptake of nitrous oxide will accelerate the uptake of a second gas, such as a volatile anesthetic [119, 120].

The cardiac output, in the absence of pulmonary shunting, directly affects the uptake of the inhaled agent into the

**Table 10.5** Partition coefficients of inhaled anesthetics

| Partition coefficient @ 37 °C | Nitrous Oxide | Halothane | Methoxyflurane | Enflurane | Isoflurane | Desflurane | Sevoflurane |
|-------------------------------|---------------|-----------|----------------|-----------|------------|------------|-------------|
| Blood: gas                    | 0.47          | 2.5       | 12             | 1.9       | 1.4        | 0.45       | 0.65        |
| Brain: blood                  | 1.1           | 2.9       | 2              | 1.5       | 2.6        | 1.3        | 1.7         |
| Fat: blood                    | 2.3           | 60        | 49             | 36        | 45         | 27         | 48          |

blood stream. As the cardiac output increases, a more rapid uptake will occur, which causes the rate of rise in the  $P_A$  to slow and the induction rate to decrease. Insoluble anesthetics display less effect from the cardiac output since little is taken up in the alveolar blood flow.

The final factor in determining the alveolar blood anesthetic uptake is the alveolar to venous partial pressure difference. A larger gradient slows the rise in  $P_A$ . These factors are determined by the tissue uptake of the anesthetic, primarily in vessel-rich groups that receive 75% of the cardiac output. The vessel rich groups—including the brain, heart, and kidneys—equilibrate rapidly with the  $P_a$ . In approximately 3 time constants, 75% of the returning venous blood has the same partial pressure as the  $P_A$ .

$P_A$  (alveolar partial pressure)  $\leftrightarrow$   $P_a$  (arterial partial pressure)  $\leftrightarrow$   $P_{br}$  (brain partial pressure):

The anesthetic partial pressure in the brain is the final component, and the clinically significant end result of drug administration. It is affected by the blood:brain partition coefficient, the cerebral blood flow and the arterial to venous partial pressure difference (■ Table 10.4).

## 10.8.2 Elimination

Recovery from anesthesia is represented as the lowering of the anesthetic concentration in the brain tissue. A majority of modern anesthetic elimination is through exhalation; however, a small percentage is elimination in biotransformation or transcutaneous loss. The most important route of elimination is through ventilation and the alveolus. As such, many of the same factors that determine induction speed account for the speed of recovery: elimination of rebreathing, high fresh gas flows, low circuit absorption, decreased agent solubility, high cerebral blood flow, and increased ventilation [72, 73]. The main difference in recovery from anesthetics is that, in recovery, different tissues in the body have different partial pressures of the inhaled anesthetic. Therefore, recovery is not as controllable as induction [121]. Because nitrous oxide is eliminated so quickly, it can dilute alveolar oxygen and carbon dioxide, causing diffusion hypoxia. Clinically, this hypoxia is avoided by administering 100% oxygen for 5–10 min after discontinuing nitrous oxide [122].

## 10.9 Questions and Answers

### ? Questions (Choose the most Appropriate Answer)

- The rate of uptake of an anesthetic gas from the lungs and hence the rate of induction with an inhalational anesthetic:
  - Increases when a premedication has been administered prior to induction
  - Is proportional to the solubility of the inhalational agent in the blood
  - Is increased if tidal volumes are decreased
  - Is dependent only on the MAC of the inhalational agent
  - Correlates with the vapor pressure of the inhalational agent
- A pediatric patient presents for an inhalational induction. The reason desflurane is not the most appropriate agent in this scenario is:
  - Desflurane has a low blood:gas partition coefficient
  - Desflurane has a high vapor pressure
  - Desflurane may produce hepatitis postoperatively
  - Desflurane may produce airway irritability
  - Desflurane cannot attain adequate potency due to its higher MAC value
- The anesthetic agent that should be avoided in patients with a history of seizure activity is:
  - Halothane
  - Isoflurane
  - Desflurane
  - Enflurane
  - All of the above
- While administering only an inhalational agent, you notice that the cardiac output of your patient has decreased. The agent that you are most likely using is:
  - Halothane
  - Isoflurane
  - Desflurane
  - Nitrous Oxide
  - Sevoflurane
- The recommended fresh gas flows when using sevoflurane is 2 L/min because:
  - Sevoflurane biodegrades into peak concentrations of 50  $\mu$ (mu)mol/L of fluoride, which may cause nephrotoxicity.
  - Metal degrades sevoflurane into hydrogen fluoride.
  - Alkali, such as soda lime, can degrade sevoflurane compound A.
  - Nitrous oxide remaining in the circuit can cause degradation of sevoflurane.
  - Sevoflurane is less pungent to airways with higher gas flows.
- A patient presents to the operating room for a knee arthroscopy. She is a 65-year-old woman with obesity. Postoperatively, she develops fever, eosinophilia, jaundice, and elevated serum transaminase levels. Which is the most likely inhalational agent he received during his case?
  - Halothane
  - Isoflurane
  - Desflurane
  - Nitrous Oxide
  - Sevoflurane

7. A patient undergoes a 25-h anesthetic for hand reconstruction after a crush injury with sevoflurane, nitrous oxide, fentanyl, and rocuronium. He is observed on postoperative day one to have megaloblastic anemia. What is the most likely source?:  
 A. Sevoflurane  
 B. Nitrous Oxide  
 C. Fentanyl  
 D. Inadequate Ventilation  
 E. Rocuronium
8. The anesthetic agent that most can produce regional myocardial ischemia during tachycardia due to a preferential dilation of the normal coronary arteries is:  
 A. Halothane  
 B. Isoflurane  
 C. Desflurane  
 D. Nitrous Oxide  
 E. Sevoflurane
9. The resting PaCO<sub>2</sub> is elevated in patients undergoing a general anesthetic with volatile agents primarily because:  
 A. The respiratory rate is decreased.  
 B. Central ventilator depression occurs.  
 C. Bronchodilation causes an elevated PaCO<sub>2</sub>.  
 D. The patient becomes apneic.  
 E. The tidal volumes are decreased.
10. Metabolism plays an important role in the emergence from anesthesia with which of the following agents:  
 A. Halothane  
 B. Methoxyflurane  
 C. Desflurane  
 D. Nitrous Oxide  
 E. None of the above
- seizures. There are case reports that support sevoflurane can produce seizure activity.
4. A. At clinical anesthetic concentration, halothane decreases the mean arterial pressure by decreasing myocardial contractility and cardiac output; whereas, isoflurane, sevoflurane, and desflurane decrease systemic vascular resistance. Nitrous oxide increases cardiac output due to a mild increase in sympathetic tone.
5. C. Alkali, such as soda lime, can degrade sevoflurane into another proven nephritic product in animal models, compound A. Larger amounts of compound A are produced with lower gas flows, increased respiratory temperatures, high sevoflurane concentrations, anesthetics of long duration, and desiccated soda lime. It is clinically recommended to maintain fresh gas flows greater than 2 L/min to limit possible compound A production. Despite proven nephrotoxicity in rats, it has never shown proven nephrotoxicity in humans to indicate injury or toxicity in humans.
6. A. "Halothane hepatitis," or type II hepatotoxicity, has been reported in 1:5000 to 1:35,000 cases of halothane administration. This immune-mediated reaction is believed to result from the trifluoroacetylated protein adducts in the liver. Clinical symptoms of halothane hepatitis include fever, eosinophilia, and jaundice. Severe cases are associated with centrilobular necrosis that may lead to fulminant liver failure with a mortality rate of 50%.
7. B. Nitrous oxide irreversibly oxidizes the cobalt atom in vitamin B<sub>12</sub>, including methionine synthetase and thymidylate synthetase. These enzymes are responsible for myelin formation and DNA synthesis; and thus, nitrous oxide has been questioned to cause bone marrow suppression. Megaloblastic changes are noticed in patients who receive nitrous oxide for duration of over 24 h.
8. B. Coronary steal syndrome may be associated with isoflurane. Sevoflurane and desflurane do not cause coronary steal syndrome. When the perfusion pressure of a coronary artery is reduced, only the vessels that are capable of dilation can effectively compensate. Atherosclerotic coronary vessels cannot effectively dilate and blood is diverted further from these areas to those with the dilation, "stealing" the blood and causing ischemia.
9. E. Volatile agents cause rapid, shallow breathing. There is a reduction in the tidal volumes and minute ventilation. The increase in respiratory rate does not compensate for the amount of tidal volume decrease and hence causes an increase in PaCO<sub>2</sub>.
10. B. As the most lipophilic inhaled anesthetic, methoxyflurane undergoes the most biotransformation at an estimated 70% of the drug administered.

### ✓ Answers

1. B. Inhalational agents with high solubility in the blood are taken up very rapidly from the alveoli. This rapid uptake lowers their partial pressure in the lung and increases the latency for induction of anesthesia. Therefore, the higher the agent's solubility in the blood, the slower its induction rate. A low blood solubility of an agent is desirable as induction and recovery times are faster.
2. D. Desflurane is a pungent gas that can cause airway irritability during induction, manifested as breath-holding, salivation, coughing, and possibly laryngospasm. Although its low blood:gas partition coefficient would allow for a rapid induction, this agent is not well suited for pediatric inductions due to its airway irritability.
3. D. Enflurane has the tendency to induce convulsions (seizures) to decreased PaCO<sub>2</sub>, MAC >2, and repetitive auditory stimuli. Isoflurane has anti-convulsive properties, and desflurane does not produce

Only a small amount of the drug, taken into body tissue, is exhaled and respiratory clearance from muscle and fat can extend over a period of several days. Methoxyflurane is metabolized in both the kidneys and the liver and inorganic fluoride (F<sup>-</sup>) is produced during its metabolism in clinically significant quantities.

## References

1. Neuman GG, Sidebotham G, Negoianu E, et al. Laparoscopy explosion hazards with nitrous oxide. *Anesthesiology*. 1993;78(5):875–9.
2. Bovill JG. Inhalation anaesthesia from diethyl ether to xenon. *Handb Exp Pharmacol*. 2008;182:121–42.
3. Yasuda N, Targ AG, Eiger EI 2nd. Solubility of I-653, sevoflurane, isoflurane, and halothane in human tissues. *Anesth Analg*. 1989;69(3):370–3.
4. Fernandez-Guisasola J, Gomez-Arnau JI, Cabrera Y, et al. Association between nitrous oxide and the incidence of postoperative nausea and vomiting in adults: a systematic review and meta-analysis. *Anaesthesia*. 2010;65(4):378–87.
5. Gmehling J, Onken U, Schulte HW. Vapor-liquid equilibria for the binary systems diethyl ether-halothane (1, 1, 1-trifluoro-2-bromo-2-chloroethane), halothane-methanol, and diethyl ether-methanol. *J Chem Eng Data*. 1980;25(1):29–32.
6. Stachnik J. Inhaled anesthetic agents. *Am J Health Syst Pharm*. 2006;63(7):623–34.
7. Cousins MJ, Greenstein LR, Hitt BA, Mazze RI. Metabolism and renal effects of enflurane in man. *Anesthesiology*. 1976;44(1):44–53.
8. Eger EI 2nd. Characteristics of anesthetic agents used for induction and maintenance of general anesthesia. *Am J Health Syst Pharm*. 2004;61(Suppl 4):S3–10.
9. Eger EI 2nd. Desflurane animal and human pharmacology: aspects of kinetics, safety, and MAC. *Anesth Analg*. 1992;75(4 Suppl):S3–7. discussion S8–9.
10. Andrews JJ, Johnston RV Jr. The new Tec 6 desflurane vaporizer. *Anesth Analg*. 1993;76(6):1338–41.
11. Eger EI 2nd. The pharmacology of inhaled anesthetics. *J Crit Care*. 2005;24(2):89–100.
12. Franks NP, Lieb WR. Where do general anaesthetics act? *Nature*. 1978;274(5669):339–42.
13. Franks NP, Lieb WR. Do general anaesthetics act by competitive binding to specific receptors? *Nature*. 1984;310(5978):599–601.
14. Sonner JM, Antognini JF, Dutton RC, et al. Inhaled anesthetics and immobility: mechanisms, mysteries, and minimum alveolar anesthetic concentration. *Anesth Analg*. 2003;97(3):718–40.
15. Narahashi T, Aistrup GL, Lindstrom JM, et al. Ion channel modulation as the basis for general anesthesia. *Toxicol Lett*. 1998;100-101:185–91.
16. Frazer MJ, Lynch C 3rd. Halothane and isoflurane effects on Ca<sup>2+</sup> fluxes of isolated myocardial sarcoplasmic reticulum. *Anesthesiology*. 1992;77(2):316–23.
17. Franks NP, Lieb WR. Which molecular targets are most relevant to general anaesthesia? *Toxicol Lett*. 1998;100-101:1–8.
18. Franks NP, Lieb WR. Molecular and cellular mechanisms of general anaesthesia. *Nature*. 1994;367(6464):607–14.
19. Jones MV, Harrison NL. Effects of volatile anesthetics on the kinetics of inhibitory postsynaptic currents in cultured rat hippocampal neurons. *J Neurophysiol*. 1993;70(4):1339–49.
20. Zimmerman SA, Jones MV, Harrison NL. Potentiation of gamma-aminobutyric acid A receptor Cl<sup>-</sup> current correlates with in vivo anesthetic potency. *J Pharmacol Exp Ther*. 1994;270(3):987–91.
21. Yamakura T, Harris RA. Effects of gaseous anesthetics nitrous oxide and xenon on ligand-gated ion channels. Comparison with isoflurane and ethanol. *Anesthesiology*. 2000;93(4):1095–101.
22. Mennerick S, Jetovic-Todorovic V, Todorovic SM, et al. Effect of nitrous oxide on excitatory and inhibitory synaptic transmission in hippocampal cultures. *J Neurosci*. 1998;18(23):9716–26.
23. Gyulai FE, Mintun MA, Firestone LLFE. Dose-dependent enhancement of in vivo GABA(A)-benzodiazepine receptor binding by isoflurane. *Anesthesiology*. 2001;95(3):585–93.
24. Patel AJ, Honore E, Lesage F, et al. Inhalational anesthetics activate two-pore-domain background K<sup>+</sup> channels. *Nat Neurosci*. 1999;2(5):422–6.
25. Hemmings HC Jr. Sodium channels and the synaptic mechanisms of inhaled anaesthetics. *Br J Anaesth*. 2009;103(1):61–9.
26. Nishikawa K, MacIver MB. Excitatory synaptic transmission mediated by NMDA receptors is more sensitive to isoflurane than are non-NMDA receptor-mediated responses. *Anesthesiology*. 2000;92(1):228–36.
27. Kendig JJ. In vitro networks: subcortical mechanisms of anaesthetic action. *Br J Anaesth*. 2002;89(1):91–101.
28. Daniels S, Roberts RJ. Post-synaptic inhibitory mechanisms of anaesthesia; glycine receptors. *Toxicol Lett*. 1998;100-101:71–6.
29. Cheng G, Kendig JJ. Enflurane directly depresses glutamate AMPA and NMDA currents in mouse spinal cord motor neurons independent of actions on GABAA or glycine receptors. *Anesthesiology*. 2000;93(4):1075–84.
30. Flood P, Ramirez-Latorre J, Role L. Alpha 4 beta 2 neuronal nicotinic acetylcholine receptors in the central nervous system are inhibited by isoflurane and propofol, but alpha 7-type nicotinic acetylcholine receptors are unaffected. *Anesthesiology*. 1997;86(4):859–65.
31. Raines DE, Claycomb RJ, Forman SA. Nonhalogenated anesthetic alkanes and perhalogenated nonimmobilizing alkanes inhibit alpha(4)beta(2) neuronal nicotinic acetylcholine receptors. *Anesth Analg*. 2002;95(3):573–7.
32. Cordero-Erausquin M, Marubio LM, Klink R, Changeux JP. Nicotinic receptor function: new perspectives from knockout mice. *Trends Pharmacol Sci*. 2000;21(6):211–7.
33. Campagna JA, Miller KW, Forman SA, Mechanisms JA. of actions of inhaled anesthetics. *N Engl J Med*. 2003;348(21):2110–24.
34. Eger EI 2nd, Stevens WC, Cromwell TH. The electroencephalogram in man anesthetized with forane. *Anesthesiology*. 1971;35(5):504–8.
35. Oshima EE, Urabe N, Shingu K, Mori K. Anticonvulsant actions of enflurane on epilepsy models in cats. *Anesthesiology*. 1985;63(1):29–40.
36. Koblin DD, Eger EI 2nd, Johnson BH, et al. Are Convulsant Gases Also Anesthetics? *Anesth Analg*. 1981;60(7):464–70.
37. Neigh JL, Garman JK, Harp JR. The electroencephalographic pattern during anesthesia with ethrane: effects of depth of anesthesia, PaCO<sub>2</sub>, and nitrous oxide. *Anesthesiology*. 1971;35(5):482–7.
38. Oda Y, Toriyama S, Tanaka K, et al. The effect of dexmedetomidine on electrocorticography in patients with temporal lobe epilepsy under sevoflurane anesthesia. *Anesth Analg*. 2007;105(5):1272–7.
39. Kaisti KK, Jaaskelainen SK, Rinne JO, et al. Epileptiform discharges during 2 MAC sevoflurane anesthesia in two healthy volunteers. *Anesthesiology*. 1999;91(6):1952–5.
40. Mielck F, Stephen H, Buhre W, et al. Effects of 1 MAC desflurane on cerebral metabolism, blood flow and carbon dioxide reactivity in humans. *Br J Anaesth*. 1998;81(2):155–60.
41. Torri G. Inhalation anesthetics: a review. *Minerva Anestesiol*. 2010;76(3):215–28.
42. Adams RW, Gronert GA, Smith TM, Michenfekder JD. Halothane, hypocapnia and cerebrospinal fluid pressure in neurosurgery. In: Brock M, Deitz, editors. *Intracranial pressure*. New York: Springer; 1972. p. 320–5.

43. McKay RE, Sonner J, McKay WR. Inhaled anesthetics. In: Stoelting RK, Miller RD, editors. *Basics in anesthesia*. 5th ed. Philadelphia: Churchill Livingstone Elsevier; 2007. p. 77–96.
44. Hornbein TF, Eger EI 2nd, Winter PM, et al. The minimum alveolar concentration of nitrous oxide in man. *Anesth Analg*. 1982; 61(7):553–6.
45. Eger EI. Isoflurane (Forane®): A compendium and reference. Madison, Wisconsin: Ohio Medical Products; 1981.
46. Khan KS, Hayes I, Buggy DJ. Pharmacology of anaesthetic agents II: inhalation anaesthetic agents. *Contin Educ Anaesth Crit Care Pain*. 2014;14(3):106–11.
47. Moore MA, Weiskopf RB, Eger EI 2nd, et al. Arrhythmogenic doses of epinephrine are similar during desflurane or isoflurane anesthesia in humans. *Anesthesiology*. 1993;79(5):943–7.
48. Navarro R, Weiskopf RB, Moore MA, et al. Humans anesthetized with sevoflurane or isoflurane have similar arrhythmic response to epinephrine. *Anesthesiology*. 1994;80(3):545–9.
49. Sakai EM, Connolly LA, Klauk JA. Inhalation anesthesiology and volatile liquid anesthetics: focus on isoflurane, desflurane, and sevoflurane. *Pharmacotherapy*. 2005;25(12):1773–88.
50. Doi M, Ikeda K. Respiratory effects of sevoflurane. *Anesth Analg*. 1987;66(3):241–4.
51. Lockhart SH, Rampil IJ, Yasuda N, et al. Depression of ventilation by desflurane in humans. *Anesthesiology*. 1991;74(3):484–8.
52. Lam AM, Clement JL, Chung DC, Knill RL. Respiratory effects of nitrous oxide during enflurane anesthesia in humans. *Anesthesiology*. 1982;56(4):298–303.
53. Goff MJ, Shahbaz R, Arain SR, et al. Absence of bronchodilation during desflurane anesthesia: a comparison to sevoflurane and thiopental. *Anesthesiology*. 2000;93(2):404–8.
54. Jones RM, Cashman JN, Mant TG. Clinical impressions and cardiorespiratory effects of a new fluorinated inhalation anaesthetic, desflurane (I-653), in volunteers. *Br J Anaesth*. 1990;64(1):11–5.
55. Kong CF, Chew ST, Ip-Yam PC. Intravenous opioids reduce airway irritation during induction of anaesthesia with desflurane in adults. *Br J Anaesth*. 2000;85(3):364–7.
56. Wilkes AR, Hall JE, Wright E, Grundler S. The effect of humidification and smoking habit on the incidence of adverse airway events during deepening of anaesthesia with desflurane. *Anaesthesia*. 2000;55(7):685–9.
57. Vitez TS, Miller RD, Eger EI 2nd, et al. Comparison in vitro of isoflurane and halothane potentiation of d- tubocurarine and succinylcholine meeting abstracts. *Anesthesiology*. 1974;41(1):53–6.
58. Hornbein TF, Eger EI 2nd, Winter PM, et al. The minimum alveolar concentration of nitrous oxide in man. *Anesth Analg*. 1982; 61(7):553–6.
59. Papadimos TJ, Almasri M, Padgett JC, Rush JEA. Suspected case of delayed onset malignant hyperthermia with desflurane anesthesia. *Anesth Analg*. 2004;98(2):548–9.
60. Ducart A, Adnet P, Renaud B, et al. Malignant hyperthermia during sevoflurane administration. *Anesth Analg*. 1995;80(3):609–11.
61. Ochiai R, Toyoda Y, Takeda J, et al. Possible association of malignant hyperthermia with sevoflurane anesthesia. *Anesth Analg*. 1992;74(4):616–8.
62. Merkel G, Eger EI 2nd. A comparative study of halothane and halopropane anesthesia including method for determining equipotency. *Anesthesiology*. 1963:346–57.
63. Hoenemann CW, Halene-Holtgraave TB, Booke M, et al. Delayed onset of malignant hyperthermia in desflurane anesthesia. *Anesth Analg*. 2003;96(1):165–7.
64. Gillmeister I, Schummer C, Hommann M, Schummer W. Delayed onset of malignant hyperthermia crisis during a living donor liver transplantation caused by sevoflurane. Article in German. *Anaesthesiol Intensivmed Notfallmed Schmerzther*. 2004;39(3):153–6.
65. Eger EI 2nd, Eisenkraft JB, Weiskopf. The pharmacology of inhaled anesthetics. San Francisco (CA). Not in PubMed. 2002
66. Whelan E, Wood AJ, Koshakji R, et al. Halothane inhibition of propranolol metabolism is stereoselective. *Anesthesiology*. 1989;71(4):561–4.
67. Reilly CS, Wood AJ, Koshakji R, Wood M. The effect of halothane on drug disposition: contribution of changes in intrinsic drug metabolizing capacity and hepatic blood flow. *Anesthesiology*. 1985; 63(1):70–6.
68. Eger EI 2nd, Lasiter MJ, Winegar R, et al. Compound A induces sister chromatid exchanges in Chinese hamster ovary cells. *Anesthesiology*. 1997;86(4):918–22.
69. Flippo TS, Holder WD Jr. Neurologic degeneration associated with nitrous oxide anesthesia in patients with vitamin B12 deficiency. *Arch Surg*. 1993;128(12):1391–5.
70. Stevenson G, Hall SC, Rudnick S, et al. The effect of anesthetic agents on the human immune response. *Anesthesiology*. 1990;72(3):542–52.
71. Stollings LM, Jia LJ, Tang P, et al. Immune modulation by volatile anesthetics. *Anesthesiology*. 2016;125(2):399–411.
72. Yasuda N, Lockhart S, Eger EI 2nd, et al. Kinetics of desflurane, isoflurane, and halothane in humans. *Anesthesiology*. 1991;74(3): 489–98.
73. Yasuda N, Lockhart S, Eger EI 2nd, et al. Comparison of kinetic of sevoflurane and isoflurane in humans. *Anesth Analg*. 1991; 72(3):316–24.
74. Krishna DR, Klotz U. Extrahepatic metabolism of drugs in humans. *Clin Pharmacokinet*. 1994;26(2):144–60.
75. Eger EI 2nd. Partition coefficients of I-653 in human blood, saline, and olive oil. *Anesth Analg*. 1987;66(10):971–3.
76. Spracklin DK, Thummel KE, Kharasch ED. Human reductive halothane metabolism in vitro is catalyzed by cytochrome P450 2A6 and 3A 4. *Drug Metab Dispos*. 1996;24(9):976–83.
77. Summary of the national halothane study: possible association between halothane anesthesia and postoperative hepatic necrosis. *JAMA*. 1966;197(10):775–88.
78. Gut J, Christen U, Huwyler J. Mechanisms of halothane toxicity: novel insights. *Pharmacol Ther*. 1993;58(2):133–55.
79. Ray DC, Drummond GB. Halothane hepatitis. *Br J Anesth*. 1991;67(1):84–99.
80. Kenna JG. Immunoallergic drug-induced hepatitis: lessons from halothane. *J Hepatol*. 1997;26(Suppl 1):5–12.
81. Yoshimura N, Holaday DA, Fiserova-Bergerova V. Metabolism of methoxyflurane in man. *Anesthesiology*. 1976;44(5):372–9.
82. Vandam LD. Report on methoxyflurane. *Anesthesiology*. 1966; 27(5):534–5.
83. Crandell WB, Pappas SG, MacDonald A. Nephrotoxicity associated with methoxyflurane anesthesia. *Anesthesiology*. 1966;27(5): 591–607.
84. Cousins MJ, Mazze RI. Methoxyflurane nephrotoxicity: a study of dose response in man. *JAMA*. 1973;225(13):1611–6.
85. Mazze RI. Methoxyflurane revisited: tale of an anesthetic from cradle to grave. *Anesthesiology*. 2006;105(4):843–6.
86. Kharasch ED, Thummel KE. Identification of cytochrome P450 2E1 as the predominant enzyme catalysing human liver microsomal defluorination of sevoflurane, isoflurane, and methoxyflurane. *Anesthesiology*. 1993;79(4):795–807.
87. Koblin DD. Characteristics and implications of desflurane metabolism and toxicity. *Anesth Analg*. 1992;75(4 Suppl):S10–6.
88. Baxter PJ, Garton K, Kharasch ED. Mechanistic aspects of carbon monoxide formation from volatile anesthetics. *Anesthesiology*. 1998;89(4):929–41.
89. Wissing H, Kuhn I, Warnken U, Dudziak R. Carbon monoxide production from desflurane, enflurane, halothane, isoflurane, and desflurane with dry soda lime. *Anesthesiology*. 2001;95(5): 1205–12.
90. Kharasch ED. Biotransformation of sevoflurane. *Anesth Analg*. 1995;81(6 Suppl):S27–38.
91. Anders MW. Formation and toxicity of anesthetic degradation products. *Annu Rev Pharmacol Toxicol*. 2005;45:147–76.
92. Iyer RA, Anders MW. Cysteine conjugate beta-lyase-dependent biotransformation of the cysteine S-conjugates of the sevoflurane degradation product compound A in humans, nonhuman primate,



- and rat kidney cytosol and mitochondria. *Anesthesiology*. 1996;85(6):1454–61.
93. Kharasch ED. Sevoflurane and the kidney: a current perspective. *Anesth Clin North Am Annu Anesthetic Pharmacol*. 1996;1:205–22.
  94. Kharasch ED, Schroeder JI, Sheffels P, Liggitt HD. Influence of sevoflurane on the metabolism and renal effects of compound A in rats. *Anesthesiology*. 2005;103(6):1183–8.
  95. Rappaport B, Mellon RD, Simone A, Woodcock J. Defining safe use of anesthesia in children. *N Engl J Med*. 2011;364(15):1387–90.
  96. Hudson AE, Hemmings HC Jr. Are anaesthetics toxic to the brain? *Br J Anaesth*. 2011;107(1):30–7.
  97. Jevtovic-Todorovic V, Hartman RE, Izumi Y, et al. Early exposure to common anesthetic agents causes widespread neurodegeneration in the developing rat brain and persistent learning deficits. *J Neurosci*. 2003;23(3):876–82.
  98. Loepke AW, Soriano SG. An assessment of the effects of general anesthetics on developing brain structure and neurocognitive function. *Anesth Analg*. 2008;106(6):1681–707.
  99. Zou X, Liu F, Zhang X, et al. Inhalation anesthetic-induced neuronal damage in the developing rhesus monkey. *Neurotoxicol Teratol*. 2011;33(5):592–7.
  100. Wilder RT, Flick RP, Sprung J, et al. Early exposure to anesthesia and learning disabilities in a population-based cohort. *Anesthesiology*. 2009;110(4):796–804.
  101. Hansen TG, Pedersen JK, Henneberg SW, et al. Academic performance in adolescence after inguinal hernia repair in infancy: a nationwide cohort study. *Anesthesiology*. 2011;114(5):1076–85.
  102. Sun LS, Li G, DiMaggio CJ, et al. Feasibility and pilot study of the Pediatric Anesthesia NeuroDevelopment Assessment (PANDA) project. *J Neurosurg Anesthesiol*. 2012;24(4):382–8.
  103. Eger EI 2nd. Age, minimum alveolar anesthetic concentration, and minimum alveolar anesthetic concentration-awake. *Anesth Analg*. 2001;93(4):947–53.
  104. Hall RI, Sullivan JA. Does cardiopulmonary bypass alter enflurane requirements for anesthesia? *Anesthesiology*. 1990;73(2):249–55.
  105. Al Q, Eger EI 2nd, Tinker JH. Determination and applications of MAC. *Anesthesiology*. 1980;53(4):315–34.
  106. Vieira E, Cleaton-Jones P, Auston JC, et al. Effects of low concentrations of nitrous oxide on rat fetuses. *Anesth Analg*. 1980;59(3):175–7.
  107. Fink B, Shepard T, Blandau R. Teratogenic activity of nitrous oxide. *Nature*. 1967;214(5084):146–8.
  108. Rowland AS, Baird DD, Weinberg CR, et al. Reduced fertility among women employed as dental assistants exposed to high levels of nitrous oxide. *N Engl J Med*. 1992;327(14):993–7.
  109. Rowland AS, Baird DD, Shore DL, et al. Nitrous oxide and spontaneous abortion in female dental assistants. *Am J Epidemiol*. 1995;141(6):531–8.
  110. Basford AB, Fink BR. The teratogenicity of halothane in the rat. *Anesthesiology*. 1968;29(6):1167–73.
  111. Wharton RS, Wilson AI, Mazze RI, et al. Fetal morphology in mice exposed to halothane. *Anesthesiology*. 1979;51(6):532–7.
  112. Corbett TH, Cornell RG, Endres JL, Lieding K. Birth defects among children of nurse-anesthetists. *Anesthesiology*. 1974;41(4):341–4.
  113. Occupational disease among operating room personnel: a national study. Report of an Ad Hoc Committee on the Effect of Trace Anesthetics on the Health of Operating Room Personnel, American Society of Anesthesiologists. *Anesthesiology*. 1974;41(4):321–40.
  114. Pharoah PO, Aberman E, Doyle P, Chamberlain G. Outcome of pregnancy among women in anaesthetic practice. *Lancet*. 1977;309(8001):34–6.
  115. Strum DP, Eger EI 2nd, Unadkat JD, et al. Age affects the pharmacokinetics of inhaled anesthetics in humans. *Anesth Analg*. 1991;73(3):310–8.
  116. Miller RD, Pardo MC Jr, editors. *Basics of anesthesia*. 8th ed. Philadelphia: Saunders; 2015.
  117. Eger EI 2nd. The effect of inspired concentration on the rate of rise of alveolar concentration. *Anesthesiology*. 1963;24:153–7.
  118. Eger EI 2nd. *Desflurane (Suprane): A compendium and reference*. Anaquest: Nutley, NJ; 1993.
  119. Stoelting RK, Eger EI 2nd. An additional explanation for the second gas effect: a concentrating effect. *Anesthesiology*. 1969;30(3):273–7.
  120. Epstein RM, Rackow H, Salanitro E, Wolf GL. Influence of the concentration effect on the uptake of anesthetic mixtures: the second gas effect. *Anesthesiology*. 1964;25:364–71.
  121. Carpenter RI, Eger EI 2nd, Johnson BH, et al. Pharmacokinetics of inhaled anesthetics in humans: measurements during and after the simultaneous administration of enflurane, halothane, isoflurane, methoxyflurane, and nitrous oxide. *Anesth Analg*. 1986;65(6):575–82.
  122. Fink BR. Diffusion anoxia. *Anesthesiology*. 1955;16(4):511–9.

#### Selected Readings

- Stachnik J. Inhaled anesthetic agents. *Am J Health Syst Pharm*. 2006;63(7):623–34.
- Eger EI 2nd. Characteristics of anesthetic agents used for induction and maintenance of general anesthesia. *Am J Health Syst Pharm*. 2004;61(Suppl 4):S3–10.
- Eger EI 2nd. The pharmacology of inhaled anesthetics. *J Crit Care*. 2005;24(2):89–100.
- Sonner JM, Antognini JF, Dutton RC, et al. Inhaled anesthetics and immobility: mechanisms, mysteries, and minimum alveolar anesthetic concentration. *Anesth Analg*. 2003;97(3):718–40.
- Campagna JA, Miller KW, Forman SA. Mechanisms of actions of inhaled anesthetics. *N Engl J Med*. 2003;348(21):2110–24.
- Torri G. Inhalation anesthetics: a review. *Minerva Anesthesiol*. 2010;76(3):215–28.
- McKay RE, Sonner J, McKay WR. Inhaled anesthetics. In: Stoelting RK, Miller RD, editors. *Basics in anesthesia*. 5th ed. Philadelphia: Churchill Livingstone Elsevier; 2007. p. 77–96.
- Khan KS, Hayes I, Buggy DJ. Pharmacology of anaesthetic agents II: inhalation anaesthetic agents. *Contin Educ Anaesth Crit Care Pain*. 2014;14(3):106–11.
- Rappaport B, Mellon RD, Simone A, Woodcock J. Defining safe use of anesthesia in children. *N Engl J Med*. 2011;364(15):1387–90.
- Ebert TJ. Inhalation anesthesia. In: Barash PG, Cullen BF, Stoelting RK, editors. *Clinical anesthesia*. 5th ed. Philadelphia: Lippincott Williams & Wilkins; 2006.
- Miller RD, Pardo MC Jr, editors. *Basics of anesthesia*. 8th ed. Philadelphia: Saunders; 2015.