

Cerebellar Developmental Disorders and Cerebellar Nuclei

Hong-Ting Prekop, Alessio Delogu, and Richard J.T. Wingate

Abstract While significant progress has been made in the last 10 years in understanding the development of cerebellar nuclei, they remain a relatively less well-studied cell group in the brain. In this chapter, we review the anatomical organisation of the cerebellar nuclei and their connections to highlight outstanding developmental questions. We then describe recent progress in dissecting the lineages of cerebellar neurons that may point to new understanding of their involvement in congenital clinical disorders.

Keywords Dentate nucleus • Interposed nucleus • Fastigial nucleus • Inferior olive • Purkinje cell • Rhombic lip • Ventricular zone • Ptf1a • Atoh1 • Pax2 • Nuclear transitory zone

What Are Cerebellar Nuclei?

The cerebellar nuclei (CN) are the final output units for cerebellar processing. For the most part, the CN output is a high-frequency tonic excitation, which is directed towards the midbrain and thalamus. However, a distinct, long-range inhibitory axon tract allows the CN to influence the activity of the inferior olive (IO), which in turn drives Purkinje cell (PC) activity via climbing fibres. CN output is modulated by the patterned firing of inhibitory PCs. They thus form the final common pathway for the integrated activity of a series of nested re-entrant loops via the inferior olive but also via the thalamus, cortex and pons (Fig. 1).

Despite the central position of CN within these major long-range networks, relatively little is known about their component cell types, the synaptic arrangement of their component interneurons or their processing role. Their development has only

H.-T. Prekop • R.J.T. Wingate (✉)
Medical Research Council Centre for Neurodevelopmental Disorders,
King's College London, London, UK
e-mail: RICHARD.WINGATE@KCL.AC.UK

A. Delogu
Wohl Institute, King's College London, London, UK

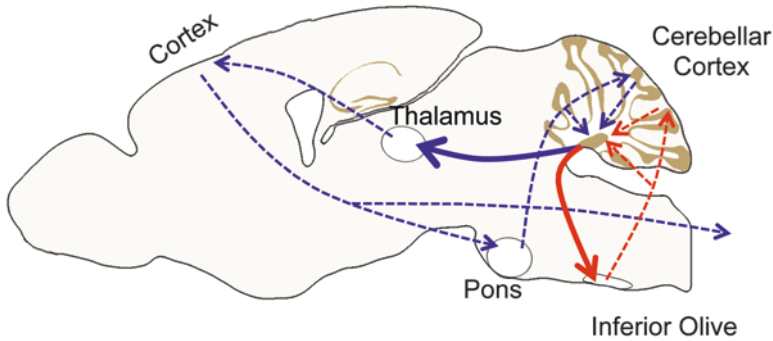


Fig. 1 The cerebellar nuclei are central to cerebellar circuitry. They lie at the centre of two cerebellar loops: the cerebello-thalamo-cerebro-cortical circuit (*blue*) which link the cerebellum back to the cerebral cortex and the olivo-cortico-nucleo-olivary loop (*red*)

recently been described, and, even then, the picture is partial. Major questions remain as to how nuclei achieve their spatial arrangement, integrate cell types of different origins and make connections. For a population of such significance for a wide variety of brain functions, this is a major omission. Similarly, while some nuclear disorders in humans have been described, the lack of anatomical and molecular description has hampered a systematic analysis of clinical disorders.

Cellular Anatomy and Diversity

The earliest descriptions of CN neurons distinguished cells with long axons from those with short axons [1] and identified large and small soma size [2]. The most detailed morphological studies of the rat and primate dentate (lateral) cerebellar nucleus were carried out by Victoria Chan-Palay in the 1970s. Using Golgi, Nissl and Weigert preparations combined with electron microscopy, she mapped out the complex, non-uniform cellular organisation of the nucleus [3–5] and demonstrated the presence of two types of projection neurons with at least three different types of cells with short axons and small soma. These latter neurons were designated as local interneurons on the basis of dendrite and axon morphology and could be distinguished by their multipolarity or bipolarity and fusiform soma.

Immunohistological and molecular techniques have subsequently shown large projection neurons to be glutamatergic (projecting to the red nucleus, thalamus or brainstem), while projection neurons with very small soma that project to the inferior olive are GABAergic [6–8] (Fig. 2). In addition to these latter nucleo-olivary inhibitory projections, glycinergic neurons can project to both the brainstem [9] or to the granule cell layer of the cerebellar cortex [3, 10–12]. Unlike the other CN cell types, these latter nucleo-cortical neurons are not spontaneously active but instead are mostly silent. They most likely target Golgi interneurons, which express glycine receptors, unlike most cells of the granule cell layer [13].

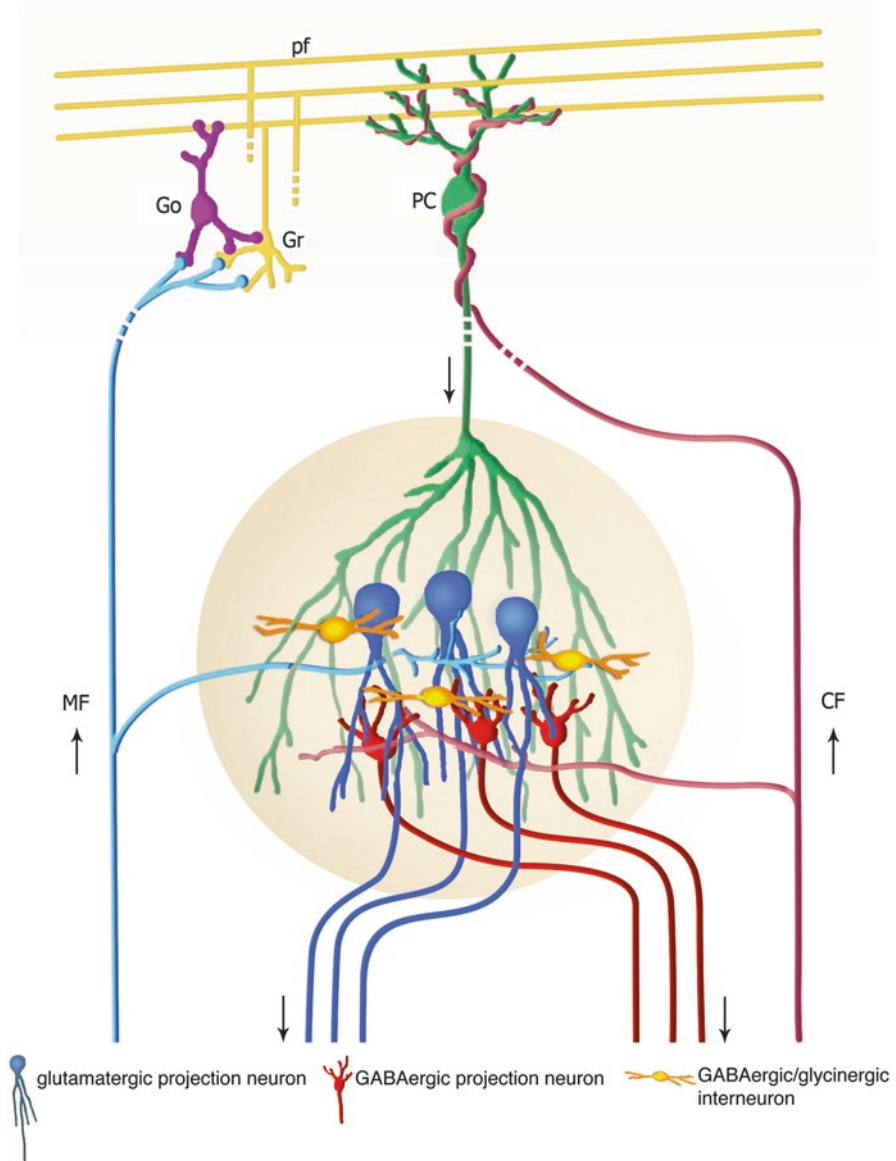


Fig. 2 The cellular composition of the cerebellar nuclei. Nuclei receive inputs from the Purkinje cells in the cerebellar cortex (green), as well as collaterals from the mossy fibres (light blue) and climbing fibres (pink) as they travel to the cortex. Within the nuclei, there are two types of projection neuron: large glutamatergic cells (blue), which are efferent cells in the cerebello-thalamo-cerebro-cortical circuits, and the nucleo-olivary neurons (red), which project to the inferior olive, forming the olivo-cerebellar loop. Interneurons (orange) participate in as yet uncharacterised local circuits

Other larger glycinergic projection neurons are found in the medial nuclei [14] and project ipsilaterally to the vestibular nuclei, the ventral brainstem and the ipsilateral ventromedial medullary reticular formation. These are hence the ipsilaterally projecting counterparts to the large glutamatergic neurons of the same region, which project contralaterally to the same regions. This has raised suggestions that posture and balance rely on a system of cross-midline control, similar system to that of the vestibular control of horizontal eye movements [15].

Relatively little is known about the local interneurons. Chan-Palay [4] noted small GABAergic neurons with fusiform or multipolar somas, limited dendritic trees and short axons, but it is possible that some of the cells observed could be the small nucleo-olivary neurons. A population of glycinergic neurons with small somata have also been found in the interposed and lateral nuclei. Because glycinergic terminals are found mainly on adjacent, presumptive glutamatergic projection neurons, it has been suggested that these are interneurons [14, 15], which colocalise with GABA [16]. GABAergic terminals that did not derive from PCs are also indicative of GABAergic interneurons or possibly local collaterals from the nucleo-olivary neurons. Though it is not possible to differentiate nucleo-olivary neurons from other GABAergic cell types in the CN based on size, there are some electrophysiological differences that aid identification [9].

Despite the fact that cells differ along both rostral-caudal and lateral-medial axes in terms of prevalence and dendritic/axonal trees, models of cerebellar function assume a homogeneous spread of each CN cell type, paralleling the long-assumed homogenous and stereotyped circuitry of the cerebellar cortex, which itself is undergoing re-examination [17]. For example, there is a higher density of nucleo-olivary neurons in the ventral lateral and interposed CN [18]. Accordingly, the PC axon terminals spread in a different manner in these parts when compared to more dorsal and medial regions of the CN [19]. On the whole, the diversity, connectivity and processing function of local interneurons have remained elusive and thus disregarded in circuitry models.

The origins of CN, how their distribution is specified and how local circuits are set up and refined are all important questions that remain to be addressed. PCs can inhibit GABAergic CN neurons, so disinhibiting glutamatergic projection neurons through local networks.

Outputs of the Cerebellar Nuclei

The CN translate cerebellar output to the cerebral cortex via the thalamus, brainstem and spinal cord through two main long-range projection systems: glutamatergic projection neurons send signals to the red nucleus, thalamus, or brainstem, while the GABAergic nucleo-olivary neurons connect the cerebellum to the inferior olive [7]. Meanwhile, other forms of efferent connections have also been found linking the CN to the vestibular nuclei and the cerebellar cortex [10, 15].

Glutamatergic projection neurons form a vital link in the assorted cerebello-thalamo-cerebro-cortical circuits which link the cerebellum back to different parts of the cerebral cortex [20]. The nucleo-olivary neuronal projections are thought to form the olivo-cortico-nucleo-olivary (OCNO) loop, a closed feedback loop between the inferior olive, cerebellar cortex and CN, made up at a fine scale of individual closed loops, or cerebellar modules, of local connections via the CN [21]. While this closed loop model is challenged by the existence of bilaterally extending nucleo-olivary neurons [22, 23], it remains a compelling architecture to describe the functional properties of the cerebellar circuit.

The origins of the diversity and the mechanisms underlying the targeting of their axons are largely unexplored. Each of these characteristics is core to an understanding of how the cerebellum influences other parts of the brain.

Inputs to Cerebellar Nuclei

The inputs to the CN comprise a complex matrix that modulates cerebellar output by influencing the spontaneous baseline firing rate of CN neurons [24, 25]. The most significant of these inputs are PCs from cortical layers directly above the corresponding part of the CN: the medial receiving input from the vermis, interposed from paravermis and the lateral receiving the bulk of its input the hemispheric PCs [26]. Sugihara et al. mapped PC projections to the various CN and found correspondence between aldolase C expression in subsets of PCs and the terminations in specific subdivisions of CN, demonstrating some conservation of topographic organisation [27].

While both PCs and CN neurons are spontaneously active [28, 29], evidence of synaptic plasticity at the CN neurons shows that the CN are involved in modulating cerebellar cortical output and not merely relaying information from the PC population [30–32]. When PC and CN neurons are monitored simultaneously, they do not give the expected reciprocal firing rates that would result from PC inhibition [33–36]. Instead CN neurons are extremely sensitive to the synchronous activity of PC inputs [37] suggesting that the development of a mapping of PC populations into the CN is a critical factor in cerebellum function.

In addition to afferents from the PCs, the CN also receive collaterals from mossy fibres (MFs) and climbing fibres (CFs). These send signals directly to the CN, bypassing cerebellar cortical processing [26]. In the overlying cerebellar cortex, MFs and CFs are topographically mapped onto GCs and PCs, and their collateral projections to CN follow approximately the same topography. MFs from the pontine nuclei, nucleus reticularis tegmenti pontis and lateral reticular nucleus send their cortical terminations such that they divide the cerebellar cortex into zones to process information from particular parts of the body or sensory modes [23, 38, 39]. In contrast, the MF collaterals to the CN are bilateral and show a looser zonal organisation [26, 40]. Likewise, anterograde tracing from the inferior olive has revealed a strict topographic alignment of CFs to the zebrin II-positive PC parasagittal zones in the contralateral cerebellar cortex [19]. The collaterals of these same CFs target the contralateral CN and terminate in specific areas of the CN [27, 41, 42].

Relatively little is known of how inputs to the CN are organised at a cellular level and the intrinsic networks that are built up by interneurons and local collaterals. A natural entry point to these questions is trying to understand the degree of convergence of a relatively orderly PC layer on to the three-dimensional assembly of CN neurons. In terms of numbers, there are around 20 PC to every CN neuron [43, 44] with inputs targeting both glutamatergic [45, 46] and GABAergic projection neurons [8]. However, since the PC axonal target field is wide and conical [47], it is estimated that each PC can encompass tens of CN neurons complicating a simple explanation of convergence. Similarly the proximity of axon terminations to the soma of CN neurons is likely to be of considerable significance in determining synaptic strength [14]. Chan-Palay noted that around 14% of larger neurons in the lateral CN were not innervated directly on their somata by PCs, setting apart a subset of projections neurons [48], which may comprise the glycinergic, nucleo-cortical neurons [11].

How the PC axon numbers are developmentally matched to CN targets and the mechanisms that regulate mapping are unknown. Similarly, how the topography of collateral projections from different afferent populations is coordinated within the nucleus is an important question that remains to be addressed. For example, it has been suggested that collaterals of inputs to the cerebellar cortex form a template for topographic refinement of outputs of Purkinje cells to the CN.

Development of Cerebellar Nuclei

The origins of the cerebellum, which sits at the boundary of the midbrain and hind-brain, were an intensely investigated problem at the end of the last century. The advent of molecular techniques revised the concept that the cerebellum received contributions from both the midbrain and hindbrain and identified the cerebellar anlage within the dorsal part of rhombomere (r)1 of the hindbrain [49–51]. Within the anlage, two distinct progenitor zones, which are defined by the mutually exclusive expression of basic helix-loop-helix (bHLH) transcription factors Ptf1a and Atoh1, produce all the cell types of the cerebellum [52]. Ptf1a is expressed in the dorsal ventricular zone of r1 and characterises progenitors of GABAergic cells [53]. The boundary between the ventricular zone and the dorsal roof plate is known as the rhombic lip [54] and expresses Atoh1 [55]. This highly proliferative zone of Atoh1 induction gives rise to glutamatergic cerebellar neurons [56, 57].

Birthdating has shown that some neurons within the CN are among the first-born cell types of the cerebellum [58]. Experiments using either BrdU or a replication-defective adenovirus [59] have shown that PCs are born around the same time as the CN. The time window for the production of glutamatergic and the GABAergic projection neurons in mice lies between E10.75 and E12.5 [60] and appears to be regulated by a common temporal signal [61]. However, the allocation of GABAergic versus glutamatergic fate is strictly a property of progenitor position within either a Ptf1a- or Atoh1-positive pool [53, 56, 57, 61, 62].

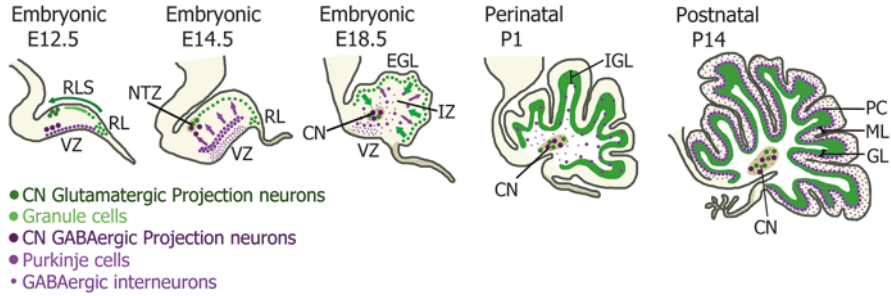


Fig. 3 The developmental timeline of the cerebellum, depicted in sagittal view. GABAergic neurons are derived from the ventricular zone (VZ), while glutamatergic neurons arise at the rhombic lip (RL). The cerebellar nucleus projection neurons are the first born from both progenitor zones, preceding first Purkinje cells (VZ-derived) and then granule cells (RL-derived). Cerebellar nucleus interneurons are believed to be born alongside other cerebellar cortical interneurons, which are generated from E13 from the VZ and later a stem cell population within the future white matter

Origin of Glutamatergic Neurons

One key motif of CN development is the assembly of neurons within an embryonic nuclear transitory zone (NTZ), which appears as almost a “staging post” in the formation of distinct CN (Fig. 3). The derivation of glutamatergic CN neurons initially appeared to be via a radial migration from the ventricular zone [63]. A detailed analysis of postmitotic precursors of CN neurons identified the expression of the transcription factors *Lhx2/Lhx9*, *Meis 1*, *Meis 2* and *Ir3*, as well as genes that are not frequently used as markers in development: *Gja9*, *Mbd2*, *Htr3a* and *Girk4* [64]. Subsequent analysis showed that *Meis 2* co-expresses with *Lhx2/Lhx9* in glutamatergic projection neurons of the lateral CN derived from the rhombic lip [57], while *Ir3* may instead represent a separate population of neurons, likely the GABAergic nucleo-olivary neurons [65].

Glutamatergic projection neurons represent the first cohort in a sequence of neurogenesis from the rhombic lip that ends with the generation of granule cells [49, 56, 57]. A separate domain of *Atoh1* expression at the midbrain-hindbrain boundary gives rise to earlier-born extracerebellar neurons [66]. At the rhombic lip, lateral and then medial CN are produced in discrete temporal waves [67, 68]. CN neurons actively migrate from the rhombic lip in a subpial layer guided by diffusible netrin and slit proteins [69, 70] and sequentially express *Pax6*, *Tbr2*, *Tbr1* and *Lmx1a* [65, 71]. As the postmitotic neurons enter the NTZ, *Tbr1* and *Tbr2* are upregulated and *Pax6* is downregulated [71]. In the absence of *Pax6*, rhombic lip-derived CN neurons are absent from the cerebellum [65]. The differential retention of transcription factors defines different CN populations in mouse. *Tbr1* expression is retained until E14.5 for lateral and interposed CN and into adulthood for the medial CN. In contrast, the lateral and interposed CN projection neurons express *Brn2* at early postnatal stages.

Origin of GABAergic Projection Neurons

The developmental origins of the GABAergic nucleo-olivary neurons are enigmatic. It is assumed that they are born from the ventricular zone like the other GABAergic cell types of the cerebellum, although direct evidence for this is lacking. Like the glutamatergic populations of the CN, GABAergic neurons are likely to arise as part of a discrete temporal window of cell production. It is thought that the GABAergic projection nucleo-olivary neurons are first in a ventricular zone temporal lineage (Kim et al. 2011) that subsequently gives rise to PCs (e10.5–e12.5 in mouse) followed by other GABAergic interneurons [72]. In contrast to these later-born cell types, both PCs and GABAergic CN neurons express Neurog2 [73]. Postmitotic cells expressing Neurog1 appear to be candidate CN nucleo-olivary projection neurons [74]. *Irx3* immunopositive cells are evident in the VZ from E10.25 to E12.5, the NTZ at E13.5 and by E15.5 the cells have migrated into an intermediate zone outside the NTZ [64, 65]. *Irx3* expression persists in the *sey/sey* (“small eye” *pax6* null) cerebellum confirming that the specification of GABAergic and glutamatergic neurons is independent of each other.

Other GABAergic Neurons

VZ progenitors require the expression of *Ptf1a* for GABAergic specification [53, 62]. Within the *Ptf1a* ventricular zone, combinatorial gene expression demarcates discrete germ zones that are thought to give rise to the different types of interneurons [64, 72, 74–79]. Thus, for example, Neurog1 and Neurog2 expression defines subsets of the *Ptf1a*+ VZ population.

However, this topographic explanation of diversity is complicated by evidence that proliferation continues within a single population of *Pax2*+ precursors from the VZ [80] that persists in the prospective white matter well into postnatal development in mouse. Heterotopic and heterochronic grafting experiments have found that *Pax2* progenitors generate all the remaining inhibitory interneurons [80, 81], including Neurog1 (*Ngn1*)-positive interneurons of the CN, which are born at E17.5 in mouse [82]. Mutation of PC progenitor transcription factors *Olig2* and *Gsx1* disrupts the production of *Pax2* lineages suggesting that the latter is derived from the former in development [83]. The origin and development of the various types of glycinergic neurons in the CN have yet to be characterised.

Nucleogenesis and Cell Migration

The different developmental origins of different types of CN neurons require that cells recognise each other and assemble nuclei distant to their origins. How nucleogenesis – the migration, organisation and synaptogenesis of CN neurons – is organised is

unknown. Clearly, either intrinsic programming or cues in the surrounding environment or a combination of both will be key factors in this developmental process.

For rhombic lip derivatives, unipolar neuroblasts move within a subpial stream towards the NTZ guided by both diffusible netrin and slit [69, 70] (NTZ); however the cues that determine the position of the NTZ itself are unclear. One possible determinant is the underlying axon scaffold of the fasciculus uncinatus, to which first-born CN cells then contribute [67, 69]. Changing the fate of CN neuroblasts blurs the boundaries between distinct populations in the NTZ but does not compress or expand the map of presumptive CN. Thus when either *Lhx9* (lateral CN in mouse) is overexpressed in chick [67] or *Tbr1* knocked down in mouse [71], CN neuron number remains similar but boundaries are less discrete. From the NTZ, cells are then incorporated into the white matter through what might constitute an active radial migration or a passive translocation as a consequence of the overall pattern of cerebellar morphogenesis [60, 63].

Evidence in favour of radial migration being a component of nucleogenesis comes from the analysis of the *reeler* mouse. *Pax6/reelin*-positive neuroblasts migrate from the rhombic lip, and at least some go on to become *Tbr2*-positive CN neurons. The *reeler* mouse has disrupted CN architecture; however, the initial tangential migration of rhombic lip derivatives to the NTZ is normal [71].

Evolution and the Diversification of Cerebellar Nuclei

While some aspects of the cerebellar circuit are among the most evolutionarily conserved across vertebrates, cerebellar nuclei are relatively variable in composition [84]. There is some debate over whether an organism is considered to have cerebelloid structures if they lack CN, since it is these cells that form the dominant output [85]. For example, teleost fish have no white matter or CN. Instead, their PCs project to eurydendroid cells, which then project to other parts of the brain. However, eurydendroid cells also receive inputs from granule cells via parallel fibres and are found within the granule cell layer and so are not homologous to CN projection neurons in terms of inputs [86, 87].

The replacement of CN by eurydendroid cells appears to be a ray-finned fish adaptation as there is evidence for a single cerebellar nucleus in the shark [88]. CN are absent in lampreys, where the cerebellum is reduced or absent. Across fish species the medial and dorsal octavolateral nuclei receive inputs from lateral line systems and are involved in spatial calculations that are analogous to those carried out in the cerebellum. It seems conceivable, though yet to be proved, that these may be considered as ontological homologues of CN [89].

Like sharks, amphibians have a single CN; however the number and diversity of CN increases in amniotes. There are two CN in birds [90] and three sets of CN in rodents: the medial, interpositus and lateral [91, 92]. In cats, rabbits and primates, there are four major CN: the medial, or fastigial, nucleus; the anterior and posterior interposed and the lateral, or dentate, nucleus. Each of these nuclei can be functionally

further subdivided such that complexity of CN organisation is a marked feature of mammalian brains [14]. This systematic variation in organisation suggests that comparative studies may offer an important insight into the significant genetic factors in the development of CN diversity.

Cerebellar Nuclei and Disease

The relatively recent discoveries of the developmental lineages of CN neurons highlight previously unexplored relationships in cerebellar disorders and disease. Glutamatergic projection neurons are formed from *Atoh1* progenitors that not only generate granule cells but also neurons in the pons, vestibular and auditory systems of the hindbrain [57, 93]. GABAergic neurons share a progenitor transcriptional profile with auditory nuclei and, perhaps most prominently, the inferior olive [53].

This is particularly significant in that developmental disorders where cerebellar nucleus exclusively malformed have not been reported. Congenital dysplasia of the dentate and olivary nuclei (DOD), though rarely recorded [94], can sometimes be detected as a minor pathology of more extensive developmental defects (Table 1). Though pathogenesis may differ across different forms of DOD, it is interesting to note that many of the below conditions have pathologies of the inferior olive too. While the correlation in pathologies could be linked by lineage, the possibility of retrograde degeneration of the cerebellar nucleus as a result of inferior olive dysplasia cannot be discounted. Similarly, the possibility that the modularity of the cerebellar-inferior olive closed loop extends to a single cell level [95] means that heavily interconnected microzones might suffer a conductive degeneration when any element of the system is disrupted.

While DOD might represent a failure of *Ptf1a* lineage development, pontocerebellar dysplasia might conversely reflect a dysgenesis of *Atoh1* lineage neurons, affecting both precerebellar and granule cell populations in addition to portions of the dentate CN. In both cases, the spectrum of associated phenotypes raises the possibility of a developmental origin within the specification or maturation of specific populations of derivatives.

Future Perspectives on Cerebellar Nucleus Development

In recent years, significant progress has been made with regard to understanding the development of the glutamatergic CN neurons, while physiologically, models of cerebellar function increasingly recognise how plasticity and modulation within the CN by mossy fibre and climbing fibre collaterals place these cells at the heart of cerebellar networks [43, 115]. However, less is known of other, equally significant, CN neuronal types and key questions about their specification and lineage remain

Table 1 Cerebellar disorders exhibiting nuclear pathology

Disorder	Aetiology	Pathology	Clinical features	Reference
Zellweger (cerebro-hepato-renal) syndrome	Autosomal recessive disease caused by mutations in PEX genes. Migration failure from 14 weeks of gestation in humans	Dysplasia of the dentate and olivary nuclei (DOD), as well as cerebellar hypoplasia and migrational defects of PCs	Developmental delay, seizures and EEG abnormalities, as well as generalised hypotonia, renal cysts and joint calcifications	[96–98]
Dentato-olivary dysplasia with intractable seizures in infancy	Unknown, though suggested to be autosomal recessively inherited	DOD – The dentate nuclei are seen as a solid ovoid or tear-shaped structure rather than the characteristic thin, convoluted band	Hypotonia with frequent seizures from birth and gross developmental delays. Survival is no longer than 3 years	[99, 100]
Joubert syndrome	Autosomal recessive disease approximately 50% of cases are genetically linked to mutations in genes that encode parts of the primary cilia. These may be important in progenitor cells for sensing morphogens like Wnt and Shh during development	Fragmentation of the dentate CN as one of many hindbrain symptoms, along with hypoplasia of vermis (molar tooth sign), dysplasia of the inferior olive and non-decussation of the SCP	Congenital ataxia, hypotonia, episodic breathing dysregulation and mental retardation	[101–103]
Rhombencephalosynapsis	Defective dorsal patterning and proliferation in the rhombic lips during early foetal development	Absence or severe dysgenesis of the cerebellar vermis. This leads to fusing of the two cerebellar hemispheres, peduncles and in the CN so that morphologically, there seems only to be one dentate nucleus that spans the breadth of the white matter	Cerebellar dysfunction, hypotonia, nystagmus, ataxia and mild to severe mental and motor developmental delays	[60, 104–106]

(continued)

Table 1 (continued)

Disorder	Aetiology	Pathology	Clinical features	Reference
Thanatophoric dysplasia	Due to gain of function mutations of FGF receptor 3 (FGFR3), which is involved in various parts of brain development, so pathological features are widespread across many brain regions as well as bones	Primarily a skeletal dysplasia with macrocephaly. Within the cerebellum, there are abnormalities of the cerebellar cortex, and CN are enlarged and hyperconvoluted and dysplastic. There is also dysplasia of the inferior olive	Generally is a lethal condition where foetuses are usually stillborn or die as neonates due to respiratory failure. For the very few survivors, clinical symptoms include seizures, dependence on ventilator and mental and motor impairments	[107–109]
Pontocerebellar hypoplasias	A group of neurodegenerative autosomal recessive disorders. Some variants are caused by tRNA splicing endonuclease mutations	Common feature is cerebellar hypoplasia and cerebellar and pons atrophy. In the cerebellum, there is scattered loss of PCs and segmental loss of dentate CN neurons, while specific regions of CN are preserved	Severe mental and motor impairments as well as swallowing problems and seizures	[110, 111]
Autism spectrum disorder	Heterogeneous: it may be caused by genetic, epigenetic or environmental factors during neurodevelopment. There is some consensus in that brain connectivity is affected. In the cerebellum, lower levels of GABA synthesis have been found in CN and PCs	Cerebellar vermal hypoplasia, reduction of superior cerebellar peduncle, decreased connectivity between the DN and cerebral regions (dentatorubrothalamic tract)	Heterogeneous spectrum of clinical features affecting social interaction, communication and behaviour	[112–114]

unanswered. A defining feature of development is that cells transit through the NTZ, yet nothing is known of the factors that regulate nucleogenesis.

Similarly, there are relatively few reports that highlight differences in cell types across the different CN. For example, Bagnall et al. [15] identified projections that are restricted to the fastigial CN, while molecular and cellular analyses point to underlying temporal cues that may explain how different nuclei are formed [67, 71]. Given that different densities of CN cell types are found across the already diversely shaped CN, and that the various CN have been found to be involved with wide ranges of motor control, from eye blinks to posture, it may be that connectivity and plasticity differ across similar cells to bring about an assortment of functions.

Finally, the diversity of different CN cells types, their origins and how they develop a network of intranuclear connectivity are key developmental questions whose answers will be of huge significance for functional models of the cerebellar network. The answer to these questions may also point towards new landmarks for the identification of disease processes in the cerebellum. This somewhat neglected population of brain cells is poised at a threshold of new understanding that offers the promise of new perspectives on the both how the cerebellum works and its clinical vulnerabilities.

References

1. Saccozzi A. Sul nucleo dentato del cervelletto. *Riv Sper Fren Med Legale*. 1887;13:93–9.
2. Lugaro E. Sulla struttura del nucleo dentato del cervelletto nell'uomo. *Monit Zool Ital*. 1895;6:5–12.
3. Chan-Palay V. *Cerebellar dentate nucleus: organization, cytology and transmitters*. Berlin: Springer; 1977. 548 p.
4. Chan-Palay V. A light microscope study of the cytology and organization of neurons in the simple mammalian nucleus lateralis: columns and swirls. *Z Anat Entwicklungsgeschichte*. 1973;141(2):125–50. PubMed PMID: 4769549.
5. Chan-Palay V. Cytology and organization in the nucleus lateralis of the cerebellum: the projections of neurons and their processes into afferent axon bundles. *Z Anat Entwicklungsgeschichte*. 1973;141(2):151–9. PubMed PMID: 4769550.
6. De Zeeuw C, Van Alphen A, Hawkins R, Ruigrok T. Climbing fibre collaterals contact neurons in the cerebellar nuclei that provide a GABAergic feedback to the inferior olive. *Neuroscience*. 1997;80(4):981–6.
7. Fredette BJ, Mugnaini E. The GABAergic cerebello-olivary projection in the rat. *Anat Embryol*. 1991;184(3):225–43.
8. Teune TM, van der Burg J, de Zeeuw CI, Voogd J, Ruigrok TJH. Single Purkinje cell can innervate multiple classes of projection neurons in the cerebellar nuclei of the rat: a light microscopic and ultrastructural triple-tracer study in the rat. *J Comp Neurol*. 1998;392(2):164–78.
9. Uusisaari MY, Knöpfel T. Diversity of neuronal elements and circuitry in the cerebellar nuclei. *Cerebellum*. 2012;11(2):420–1.
10. Houck BD, Person AL. Cerebellar loops: a review of the nucleocortical pathway. *Cerebellum*. 2014;13(3):378–85.
11. Uusisaari M, Knöpfel T. GlyT2 neurons in the lateral cerebellar nucleus. *Cerebellum*. 2010;9(1):42–55.

12. Uusisaari M, Obata K, Knöpfel T. Morphological and electrophysiological properties of GABAergic and non-GABAergic cells in the deep cerebellar nuclei. *J Neurophysiol.* 2007;97(1):901–11.
13. Uusisaari M, Knöpfel T. Functional classification of neurons in the mouse lateral cerebellar nuclei. *Cerebellum.* 2011;10(4):637–46.
14. De Zeeuw CI, Berrebi AS. Postsynaptic targets of Purkinje cell terminals in the cerebellar and vestibular nuclei of the rat. *Eur J Neurosci.* 1995;7(11):2322–33.
15. Bagnall MW, Zingg B, Sakatos A, Moghadam SH, Zeilhofer HU, du Lac S. Glycinergic projection neurons of the cerebellum. *J Neurosci.* 2009;29(32):10104–10.
16. Chen S, Hillman DE. Colocalization of neurotransmitters in the deep cerebellar nuclei. *J Neurocytol.* 1993;22(2):81–91.
17. Cerminara NL, Lang EJ, Sillitoe RV, Apps R. Redefining the cerebellar cortex as an assembly of non-uniform Purkinje cell microcircuits. *Nat Rev Neurosci.* 2015;16(2):79–93.
18. Giaquinta G, Casabona A, Smecca G, Bosco G, Perciavalle V. Cortical control of cerebellar dentato-rubral and dentato-olivary neurons. *Neuroreport.* 1999;10(14):3009–13.
19. Sugihara I, Fujita H, Na J, Quy PN, Li BY, Ikeda D. Projection of reconstructed single Purkinje cell axons in relation to the cortical and nuclear aldolase C compartments of the rat cerebellum. *J Comp Neurol.* 2009;512(2):282–304.
20. D'Angelo E, Casali S. Seeking a unified framework for cerebellar function and dysfunction: from circuit operations to cognition. *Front Neural Circ.* 2013;6:116.
21. Ruigrok TJH. Ins and outs of cerebellar modules. *Cerebellum.* 2011;10(3):464–74.
22. Teune TM, van der Burg J, van der Moer J, Voogd J, Ruigrok TJ. Topography of cerebellar nuclear projections to the brain stem in the rat. *Prog Brain Res.* 2000;124:141–72.
23. Uusisaari M, De Schutter E. The mysterious microcircuitry of the cerebellar nuclei. *J Physiol.* 2011;589(Pt 14):3441–57.
24. Person AL, Raman IM. Purkinje neuron synchrony elicits time-locked spiking in the cerebellar nuclei. *Nature.* 2011;481(7382):502–5. PubMed PMID: 22198670. Pubmed Central PMCID: 3268051.
25. Heck DH, De Zeeuw CI, Jaeger D, Khodakhah K, Person AL. The neuronal code(s) of the cerebellum. *J Neurosci: Off J Soc Neurosci.* 2013;33(45):17603–9. PubMed PMID: 24198351. Pubmed Central PMCID: 3818542.
26. Voogd J, Glickstein M. The anatomy of the cerebellum. *Trends Cogn Sci.* 1998;2(9):307–13.
27. Sugihara I, Shinoda Y. Molecular, topographic, and functional organization of the cerebellar nuclei: analysis by three-dimensional mapping of the olivonuclear projection and aldolase C labeling. *J Neurosci: Off J Soc Neurosci.* 2007;27(36):9696–710.
28. Raman IM, Gustafson AE, Padgett D. Ionic currents and spontaneous firing in neurons isolated from the cerebellar nuclei. *J Neurosci.* 2000;20(24):9004–16.
29. Thach W. Discharge of Purkinje and cerebellar nuclear neurons during rapidly alternating arm movements in the monkey. *J Neurophysiol.* 1968;31(5):785–97.
30. Morishita W, Sastry BR. Postsynaptic mechanisms underlying long-term depression of GABAergic transmission in neurons of the deep cerebellar nuclei. *J Neurophysiol.* 1996;76(1):59–68.
31. Ohyama T, Nores WL, Medina JF, Riusech FA, Mauk MD. Learning-induced plasticity in deep cerebellar nucleus. *J Neurosci.* 2006;26(49):12656–63.
32. Zheng N, Raman IM. Synaptic inhibition, excitation, and plasticity in neurons of the cerebellar nuclei. *Cerebellum.* 2010;9(1):56–66.
33. Armstrong D, Edgley S. Discharges of nucleus interpositus neurones during locomotion in the cat. *J Physiol.* 1984;351:411.
34. Armstrong D, Edgley S. Discharges of Purkinje cells in the paravermal part of the cerebellar anterior lobe during locomotion in the cat. *J Physiol.* 1984;352:403.
35. McDevitt CJ, Ebner TJ, Bloedel JR. Changes in the responses of cerebellar nuclear neurons associated with the climbing fiber response of Purkinje cells. *Brain Res.* 1987;425(1):14–24.
36. McDevitt CJ, Ebner TJ, Bloedel JR. Relationships between simultaneously recorded Purkinje cells and nuclear neurons. *Brain Res.* 1987;425(1):1–13.

37. Person AL, Raman IM. Purkinje neuron synchrony elicits time-locked spiking in the cerebellar nuclei. *Nature*. 2012;481(7382):502–5.
38. Apps R, Hawkes R. Cerebellar cortical organization: a one-map hypothesis. *Nat Rev Neurosci*. 2009;10(9):670–81.
39. Shinoda Y, Sugihara I, Wu H, Sugiuchi Y. The entire trajectory of single climbing and mossy fibers in the cerebellar nuclei and cortex. *Prog Brain Res*. 1999;124:173–86.
40. Wu H, Sugihara I, Shinoda Y. Projection patterns of single mossy fibers originating from the lateral reticular nucleus in the rat cerebellar cortex and nuclei. *J Comp Neurol*. 1999;411(1):97–118.
41. Blenkinsop TA, Lang EJ. Synaptic action of the olivocerebellar system on cerebellar nuclear spike activity. *J Neurosci*. 2011;31(41):14708–20.
42. Sugihara I, Wu H, Shinoda Y. Morphology of single olivocerebellar axons labeled with biotinylated dextran amine in the rat. *J Comp Neurol*. 1999;414(2):131–48.
43. Person AL, Raman IM. Synchrony and neural coding in cerebellar circuits. *Front Neural Circ*. 2012;6:97.
44. Sultan F, König T, Möck M, Thier P. Quantitative organization of neurotransmitters in the deep cerebellar nuclei of the Lurcher mutant. *J Comp Neurol*. 2002;452(4):311–23.
45. Aizenman CD, Huang EJ, Linden DJ. Morphological correlates of intrinsic electrical excitability in neurons of the deep cerebellar nuclei. *J Neurophysiol*. 2003;89(4):1738–47.
46. Matsuno H, Kudoh M, Watakabe A, Yamamori T, Shigemoto R, Nagao S. Distribution and structure of synapses on medial vestibular nuclear neurons targeted by cerebellar Flocculus Purkinje cells and vestibular nerve in mice: light and electron microscopy studies. *PLoS One*. 2016;11(10):e0164037.
47. Chan-Palay V. Afferent axons and their relations with neurons in the nucleus lateralis of the cerebellum: a light microscopic study. *Z Anat Entwicklungsgeschichte*. 1973;142(1):1–21. PubMed.
48. Chan-Palay V. On the identification of the afferent axon terminals in the nucleus lateralis of the cerebellum. An electron microscope study. *Z Anat Entwicklungsgeschichte*. 1973;142(2):149–86. PubMed.
49. Wingate RJ, Hatten ME. The role of the rhombic lip in avian cerebellum development. *Development*. 1999;126(20):4395–404. PubMed PMID: 10498676. Epub 1999/09/28. eng.
50. Millet S, Bloch-Gallego E, Simeone A, Alvarado-Mallart RM. The caudal limit of *Otx2* gene expression as a marker of the midbrain/hindbrain boundary: a study using *in situ* hybridisation and chick/quail homotopic grafts. *Development*. 1996;122(12):3785–97.
51. Zervas M, Millet S, Ahn S, Joyner AL. Cell behaviors and genetic lineages of the mesencephalon and rhombomere 1. *Neuron*. 2004;43(3):345–57. PubMed.
52. Wingate R. Math-Map(ic)s. *Neuron*. 2005;48(1):1–4. PubMed.
53. Hoshino M, Nakamura S, Mori K, Kawauchi T, Terao M, Nishimura YV, et al. *Ptf1a*, a bHLH transcriptional gene, defines GABAergic neuronal fates in cerebellum. *Neuron*. 2005;47(2):201–13. PubMed.
54. His W. Die entwicklung des menschlichen rautenhirns vom ende des ersten bis zum beginn des dritten monats. I. Verlängertes Mark. *Abh Kön Sächs Ges d Wiss Mat Phys Kl*. 1890;29:1–74.
55. Ben-Arie N, Bellen HJ, Armstrong DL, McCall AE, Gordadze PR, Guo Q, et al. *Math1* is essential for genesis of cerebellar granule neurons. *Nature*. 1997;390(6656):169–72.
56. Machold R, Fishell G. *Math1* is expressed in temporally discrete pools of cerebellar rhombic-lip neural progenitors. *Neuron*. 2005;48(1):17–24. PubMed.
57. Wang VY, Rose MF, Zoghbi HY. *Math1* expression redefines the rhombic lip derivatives and reveals novel lineages within the brainstem and cerebellum. *Neuron*. 2005;48(1):31–43. PubMed.
58. Altman J, Bayer SA. Prenatal development of the cerebellar system in the rat. I. Cytogenesis and histogenesis of the deep nuclei and the cortex of the cerebellum. *J Comp Neurol*. 1978;179(1):23–48.

59. Hashimoto M, Mikoshiba K. Mediolateral compartmentalization of the cerebellum is determined on the “birth date” of Purkinje cells. *J Neurosci*. 2003;23(36):11342–51.
60. Elsen G, Juric-Sekhar G, Daza R, Hevner RF. Development of cerebellar nuclei. In: Manto M, Gruol D, Schmammann J, Koibuchi N, Rossi F, editors. *Handbook of cerebellum and cerebellum disorders*. Heidelberg: Springer; 2013. p. 179–205.
61. Yamada M, Seto Y, Taya S, Owa T, Inoue YU, Inoue T, et al. Specification of spatial identities of cerebellar neuron progenitors by *ptf1a* and *ato1* for proper production of GABAergic and glutamatergic neurons. *J Neurosci: Off J Soc Neurosci*. 2014;34(14):4786–800. PubMed.
62. Pascual M, Abasolo I, Mingorance-Le Meur A, Martinez A, Del Rio JA, Wright CV, et al. Cerebellar GABAergic progenitors adopt an external granule cell-like phenotype in the absence of *Ptf1a* transcription factor expression. *Proc Natl Acad Sci U S A*. 2007;104(12):5193–8. PubMed PubMed Central PMCID: 1829285. Epub 2007/03/16. eng.
63. Altman J, Bayer SA. Embryonic development of the rat cerebellum. II. Translocation and regional distribution of the deep neurons. *J Comp Neurol*. 1985;231(1):27–41. PubMed PMID: 3968227. Epub 1985/01/01. eng.
64. Morales D, Hatten ME. Molecular markers of neuronal progenitors in the embryonic cerebellar anlage. *J Neurosci: Off J Soc Neurosci*. 2006;26(47):12226–36. PubMed PMID: 17122047.
65. Yeung J, Ha TJ, Swanson DJ, Goldowitz D. A novel and multivalent role of *Pax6* in cerebellar development. *J Neurosci: Off J Soc Neurosci*. 2016;36(35):9057–69. PubMed PMID: 27581449. PubMed Central PMCID: 5005719.
66. Green MJ, Myat AM, Emmenegger BA, Wechsler-Reya RJ, Wilson LJ, Wingate RJ. Independently specified *Atoh1* domains define novel developmental compartments in rhombomere 1. *Development*. 2014;141(2):389–98. PubMed PMID: 24381197. Epub 2014/01/02. eng.
67. Green MJ, Wingate RJ. Developmental origins of diversity in cerebellar output nuclei. *Neural Dev*. 2014;9(1):1. PubMed PMID: 24405572. PubMed Central PMCID: 3929244.
68. Wilson LJ, Wingate RJ. Temporal identity transition in the avian cerebellar rhombic lip. *Dev Biol*. 2006;297(2):508–21. PubMed PMID: 16806151. Epub 2006/06/30. eng.
69. Gilthorpe JD, Papantoniou EK, Chedotal A, Lumsden A, Wingate RJ. The migration of cerebellar rhombic lip derivatives. *Development*. 2002;129(20):4719–28. PubMed PMID: 12361964. Epub 2002/10/04. eng.
70. Alcantara S, Ruiz M, De Castro F, Soriano E, Sotelo C. Netrin 1 acts as an attractive or as a repulsive cue for distinct migrating neurons during the development of the cerebellar system. *Development*. 2000;127(7):1359–72. PubMed PMID: 10704383. Epub 2000/03/08. eng.
71. Fink AJ, Englund C, Daza RA, Pham D, Lau C, Nivison M, et al. Development of the deep cerebellar nuclei: transcription factors and cell migration from the rhombic lip. *J Neurosci: Off J Soc Neurosci*. 2006;26(11):3066–76. PubMed PMID: 16540585. Epub 2006/03/17. eng.
72. Sudarov A, Turnbull RK, Kim EJ, Lebel-Potter M, Guillemot F, Joyner AL. *Ascl1* genetics reveals insights into cerebellum local circuit assembly. *J Neurosci: Off J Soc Neurosci*. 2011;31(30):11055–69.
73. Florio M, Leto K, Muzio L, Tinterri A, Badaloni A, Croci L, et al. Neurogenin 2 regulates progenitor cell-cycle progression and Purkinje cell dendritogenesis in cerebellar development. *Development*. 2012;139(13):2308–20.
74. Zordan P, Croci L, Hawkes R, Consalez GG. Comparative analysis of proneural gene expression in the embryonic cerebellum. *Dev Dyn*. 2008;237(6):1726–35.
75. Chizhikov VV, Lindgren AG, Currie DS, Rose MF, Monuki ES, Millen KJ. The roof plate regulates cerebellar cell-type specification and proliferation. *Development*. 2006;133(15):2793–804.
76. Mizuhara E, Minaki Y, Nakatani T, Kumai M, Inoue T, Muguruma K, et al. Purkinje cells originate from cerebellar ventricular zone progenitors positive for *Neph3* and *E-cadherin*. *Dev Biol*. 2010;338(2):202–14. PubMed PMID: 20004188. Epub 2009/12/17. eng.

77. Leto K, Rolando C, Rossi F. The genesis of cerebellar GABAergic neurons: fate potential and specification mechanisms. *Front Neuroanat.* 2012;6:6.
78. Lundell T, Zhou Q, Doughty M. Neurogenin1 expression in cell lineages of the cerebellar cortex in embryonic and postnatal mice. *Dev Dyn.* 2009;238(12):3310–25.
79. Sillitoe RV, Joyner AL. Morphology, molecular codes, and circuitry produce the three-dimensional complexity of the cerebellum. *Annu Rev Cell Dev Biol.* 2007;23:549–77.
80. Maricich SM, Herrup K. Pax-2 expression defines a subset of GABAergic interneurons and their precursors in the developing murine cerebellum. *J Neurobiol.* 1999;41(2):281–94.
81. Leto K, Carletti B, Williams IM, Magrassi L, Rossi F. Different types of cerebellar GABAergic interneurons originate from a common pool of multipotent progenitor cells. *J Neurosci: Off J Soc Neurosci.* 2006;26(45):11682–94.
82. Obana EA, Lundell TG, Kevin JY, Radomski KL, Zhou Q, Doughty ML. Neurog1 genetic inducible fate mapping (GIFM) reveals the existence of complex spatiotemporal cyto-architectures in the developing cerebellum. *Cerebellum.* 2015;14(3):247–63.
83. Seto Y, Nakatani T, Masuyama N, Taya S, Kumai M, Minaki Y, et al. Temporal identity transition from Purkinje cell progenitors to GABAergic interneuron progenitors in the cerebellum. *Nat Commun.* 2014;5:3337. PubMed PMID: 24535035.
84. Butts T, Chaplin N, Wingate RJ. Can clues from evolution unlock the molecular development of the cerebellum? *Mol Neurobiol.* 2011;43(1):67–76. PubMed PMID: 21174175. Epub 2010/12/22. eng.
85. Marzban H, Del Bigio MR, Alizadeh J, Ghavami S, Zachariah RM, Rastegar M. Cellular commitment in the developing cerebellum. *Front Cell Neurosci.* 2015;8:450.
86. Hashimoto M, Hibi M. Development and evolution of cerebellar neural circuits. *Develop Growth Differ.* 2012;54(3):373–89. PubMed PMID: 22524607. Epub 2012/04/25. eng.
87. Murakami T, Morita Y. Morphology and distribution of the projection neurons in the cerebellum in a teleost, *Sebastiscus marmoratus*. *J Comp Neurol.* 1987;256(4):607–23.
88. Ebbesson SO, Campbell CB. On the organization of cerebellar efferent pathways in the nurse shark (*Ginglymostoma cirratum*). *J Comp Neurol.* 1973;152(3):233–54. PubMed PMID: 4130103.
89. Butler A, Hodos W. Comparative vertebrate neuroanatomy: evolution and adaptation. New York: Wiley-Liss; 1996. 514 p.
90. Arends JJ, Zeigler HP. Organization of the cerebellum in the pigeon (*Columba livia*): II. Projections of the cerebellar nuclei. *J Comp Neurol.* 1991;306(2):245–72. PubMed PMID: 1711054. Epub 1991/04/08. eng.
91. Goodman DC, Hallett RE, Welch RB. Patterns of localization in the cerebellar corticonuclear projections of albino rat. *J Comp Neurol.* 1963;121:51–67. PubMed PMID: 14051845.
92. Korneliussen HK. On the morphology and subdivision of the cerebellar nuclei of the rat. *J Hirnforsch.* 1968;10(2):109–22. PubMed PMID: 4181301.
93. Wingate RJT. The rhombic lip and early cerebellar development. *Curr Opin Neurobiol.* 2001;11(1):82–8.
94. Golden J, Harding B. Pathology and genetics. Developmental neuropathology. Basel: ISN Neuropath Press; 2004.
95. Lu H, Yang B, Jaeger D. Cerebellar nuclei neurons show only small excitatory responses to optogenetic olivary stimulation in transgenic mice: in vivo and in vitro studies. *Front Neural Circ.* 2016;10:21.
96. Müller CC, Nguyen TH, Ahlemeyer B, Meshram M, Santrampurwala N, Cao S, et al. PEX13 deficiency in mouse brain as a model of Zellweger syndrome: abnormal cerebellum formation, reactive gliosis and oxidative stress. *Dis Model Mech.* 2011;4(1):104–19.
97. Powers JM, Moser HW, Moser AB, Upshur JK, Bradford BF, Pai SG, et al. Fetal cerebropontorenal (Zellweger) syndrome: dysmorphic, radiologic, biochemical, and pathologic findings in four affected fetuses. *Hum Pathol.* 1985;16(6):610–20.
98. Volpe JJ, Adams RD. Cerebro-hepato-renal syndrome of Zellweger: an inherited disorder of neuronal migration. *Acta Neuropathol.* 1972;20(3):175–98.

99. Harding B, Boyd S. Intractable seizures from infancy can be associated with dentato-olivary dysplasia. *J Neurol Sci.* 1991;104(2):157–65.
100. Martland T, Harding BN, Morton RE, Young I. Dentato-olivary dysplasia in sibs: an autosomal recessive disorder? *J Med Genet.* 1997;34(12):1021–3.
101. Joubert M, Eisenring J-J, Robb JP, Andermann F. Familial agenesis of the cerebellar vermis: a syndrome of episodic hyperpnea, abnormal eye movements, ataxia and retardation. American Academy of Neurology meeting, 1968, Chicago, US; Read in part at the aforementioned conference; 1968 1999: BC Decker.
102. Millen KJ, Gleeson JG. Cerebellar development and disease. *Curr Opin Neurobiol.* 2008;18(1):12–9. PubMed PMID: 18513948. Pubmed Central PMCID: 2474776. Epub 2008/06/03. eng.
103. Yachnis AT, Rorke LB. Cerebellar and brainstem development: an overview in relation to Joubert syndrome. *J Child Neurol.* 1999;14(9):570–3. PubMed PMID: 10488901. Epub 1999/09/17. eng.
104. Pasquier L, Marcocelles P, Loget P, Pelluard F, Carles D, Perez M-J, et al. Rhombencephalosynapsis and related anomalies: a neuropathological study of 40 fetal cases. *Acta Neuropathol.* 2009;117(2):185–200.
105. Utsunomiya H, Takano K, Ogasawara T, Hashimoto T, Fukushima T, Okazaki M. Rhombencephalosynapsis: cerebellar embryogenesis. *Am J Neuroradiol.* 1998;19(3):547–9.
106. Yachnis AT. Rhombencephalosynapsis with massive hydrocephalus: case report and pathogenetic considerations. *Acta Neuropathol.* 2002;103(3):301–4.
107. Coulter CL, Leech RW, Brumback RA, Schaefer GB. Cerebral abnormalities in thanatophoric dysplasia. *Childs Nerv Syst.* 1991;7(1):21–6.
108. Hevner RF. The cerebral cortex malformation in thanatophoric dysplasia: neuropathology and pathogenesis. *Acta Neuropathol.* 2005;110(3):208–21.
109. Miller E, Blaser S, Shannon P, Widjaja E. Brain and bone abnormalities of thanatophoric dwarfism. *Am J Roentgenol.* 2009;192(1):48–51.
110. Namavar Y, Barth PG, Baas F. Classification, diagnosis and potential mechanisms in ponto-cerebellar hypoplasia. *Orphanet J Rare Dis.* 2011;6(1):1.
111. Rudnik-Schöneborn S, Barth PG, Zerres K. Pontocerebellar hypoplasia. *Am J Med Genet C: Semin Med Genet.* Wiley Online Library; 2014.
112. Jeong J-W, Chugani DC, Behen ME, Tiwari VN, Chugani HT. Altered white matter structure of the dentatorubrothalamic pathway in children with autistic spectrum disorders. *Cerebellum.* 2012;11(4):957–71.
113. Olivito G, Clausi S, Laghi F, Tedesco AM, Baiocco R, Mastropasqua C, et al. Resting-state functional connectivity changes between dentate nucleus and cortical social brain regions in autism spectrum disorders. *Cerebellum.* 2017;16:283.
114. Yip J, Soghomonian JJ, Blatt GJ. Decreased GAD65 mRNA levels in select subpopulations of neurons in the cerebellar dentate nuclei in autism: an in situ hybridization study. *Autism Res.* 2009;2(1):50–9.
115. Pugh JR, Raman IM. Mechanisms of potentiation of mossy fiber EPSCs in the cerebellar nuclei by coincident synaptic excitation and inhibition. *J Neurosci.* 2008;28(42):10549–60.