# **Neurodevelopmental Disorders of the Cerebellum: Autism Spectrum Disorder**

**Mehnosh Toback, Kambiz Zangeneh, Tabrez J. Siddiqui, and Hassan Marzban**

**Abstract** Autism spectrum disorder (ASD) is a neurodevelopmental disorder with an incidence of 1 in 68 children. Cerebellar abnormalities have been observed in many ASD patients. The cerebellum is an elaborate brain region critically important for motor learning and coordination of movement, and increasing lines of evidence indicate that the cerebellum also contributes to emotion and cognition. In this chapter, we will review the genetic and environmental factors that may cause cerebellar deficits in ASD patients. Structural and functional cerebellar abnormalities based on neuroimaging and histopathological studies and current approaches to management will be discussed.

**Keywords** Cerebellum • Neurodevelopmental disorders • Motor skills • Language • Cognition • Autism spectrum disorder

M. Toback

Foothills Hospital, 1403, 29 Street N.W, Calgary, Alberta T2N 2T9, Canada

K. Zangeneh Sina Laboratory Building, Shakaraby Avenue, Arak, Iran

T.J. Siddiqui Department of Physiology and Pathophysiology, Max Rady College of Medicine, University of Manitoba, Winnipeg, MB, Canada

Neuroscience Research Program, Kleysen Institute for Advanced Medicine, Health Sciences Centre, Winnipeg, MB, Canada

H. Marzban  $(\boxtimes)$ 

© Springer International Publishing AG 2017 369 H. Marzban (ed.), *Development of the Cerebellum from Molecular Aspects to Diseases*, Contemporary Clinical Neuroscience, DOI 10.1007/978-3-319-59749-2\_18

Department of Human Anatomy and Cell Science, The Children's Hospital Research Institute of Manitoba (CHRIM), Max Rady College of Medicine, Rady Faculty of Health Science, University of Manitoba, Winnipeg, MB R3E 0J9, Canada e-mail: [Hassan.Marzban@umanitoba.ca](mailto:Hassan.Marzban@umanitoba.ca)

## **Introduction**

Autism is a commonly occurring complex neurodevelopmental disorder. Leo Kanner, a child psychiatrist (1943), first described patients with "a powerful desire for aloneness" and "an obsessive insistence on persistent sameness" as "early infantile autism" [[1–](#page-10-0)[4\]](#page-10-1). A similar behavioral disorder, "Asperger's syndrome," was reported by Hans Asperger [\[5](#page-10-2)]. To avoid using different terminologies, these disorders were together named "autism disorders" in 1987. Recently, the Diagnostic and Statistical Manual of Mental Disorders (DSM-V) collectively designated all autismlike disorders as "autism spectrum disorder" (ASD) [\[3](#page-10-3)].

ASD is characterized by a triad of symptoms: (1) impairment in social interaction, (2) communication difficulties, and (3) restricted, repetitive, and stereotyped patterns of behavior [[3,](#page-10-3) [6](#page-10-4)[–10\]](#page-10-5). Current methods can diagnose the ASD in children as young as 2 years old, and males are four times more likely to be diagnosed with ASD than females [[11\]](#page-10-6). The prevalence of ASD is estimated at 1 in 68 children in the United States [\[12](#page-10-7)] and 1–2% in Asia, Europe, and North America (see chapter "Epidemiology of Cerebellar Disorders"). The etiology of ASD is complicated: in some patients it is unknown, and in some cases, individuals are affected due to gene mutations and/or environmental factors [[7\]](#page-10-8). However, the interplay of genetic, environmental, and epigenetic factors probably underlies the mechanisms of ASD [[8,](#page-10-9) [13](#page-10-10)].

A subset of ASD patients, about one in five, display increased head circumference and brain volume in early childhood, typically until 5–6 years of age [[14,](#page-10-11) [15\]](#page-10-12). In these patients, the cortical white matter, the thickness of the corpus callosum, and the volume of cerebrospinal fluid (CSF) in the subarachnoid space are increased at the age of 6–9 months [[16–](#page-10-13)[18\]](#page-10-14). The frontal cortex has been reported to be larger, probably due to increased neuronal density in the prefrontal cortex [\[19](#page-11-0)]. Other brain regions that are prominently implicated in ASD include the cerebellum; brainstem and limbic system, including the hippocampus; and basal ganglia [\[20](#page-11-1)]. These areas are most likely responsible for the symptoms of those patients with abnormalities related to social behavior, executive functions, atypical use of language, and difficulties with speech [\[21](#page-11-2)]. Additionally, enlargement of the amygdala and caudate nucleus may cause anxiety and repetitive behavior [[22\]](#page-11-3).

Recent advances in molecular genetics and imaging technologies have shown that the cerebellum is one of the most consistently affected brain regions in ASD patients [\[8](#page-10-9), [23](#page-11-4), [24\]](#page-11-5). The cerebellar neurodevelopmental deficits in ASD include abnormalities in the cerebellar cortex, neurodegeneration, and impaired cerebellar circuits. Together, these deficits affect motor, sensory, language, and cognitive functions [[25–](#page-11-6)[28](#page-11-7)].

#### **Autism Spectrum Disorder Pathogenesis**

Emerging evidence from genetic association studies and postmortem human brain tissue indicates that ASD is either hereditary or caused probably by de novo mutations in a number of genes. Additionally, certain environmental risk factors have

been proposed to be causative in ASD. The set of molecular pathways, neural circuits, and behaviors affected in the different autisms is highly complex, and as a result, it has been difficult to uncover the neurobiological underpinnings of ASD [\[9](#page-10-15), [29\]](#page-11-8). How the cerebellum contributes to the etiology of ASD has been particularly underappreciated.

The cerebellum develops from early embryogenesis to the first year postnatally in human. This long period of pre- and postnatal cerebellar development makes it susceptible to many risk factors  $[30-34]$  $[30-34]$ . In this section, we briefly review the findings regarding currently identified genetic and environmental risk factors in ASD. Epigenetic susceptibility factors have been discussed in chapter "Epigenetics and Cerebellar Neurodevelopmental Disorders."

#### *Genetic Factors*

Several lines of evidence have revealed that ASD is a neurodevelopmental disorder determined largely by genetic factors [\[35\]](#page-11-11). For example, twin studies have higher concordance rates for monozygotic twins than for dizygotic; approximately 80% of monozygotic twins are concordant compared to 10% of dizygotic twins, with a heritability of over 90% [[36\]](#page-11-12). Recently, genome-wide association studies have identified many genes as risk factors for ASD. These genes span several chromosomal loci, and many are highly expressed in, and are involved in, the development of the cerebellum [\[37](#page-11-13), [38\]](#page-11-14). Sadakata et al. [\[38](#page-11-14)] categorized these genes based on their role in development of the nervous system and synapse development and function. Some of these genes, such as CDH9, CDH10, RELN, and PTEN, are involved in developmental process such as neuronal differentiation, migration, and circuit formation. An important category of ASD-associated genes regulates synaptic adhesion and synaptic transmission, including genes encoding for neurexins, neuroligins, leucine-rich repeat transmembrane neuronal proteins (LRRTMs), Shanks, and SynGAP [[39–](#page-12-0)[42\]](#page-12-1). Another category of ASD risk genes encodes for proteins required for transcription and translation such as EN2, TSC1, FMR1, and MECP2 [\[38](#page-11-14), [43](#page-12-2)].

Chromodomain-helicase-DNA-binding protein 8 (CHD8), previously called Duplin, is one of the genes most strongly associated with ASD [\[44](#page-12-3), [45\]](#page-12-4). It was the first identified in the screen for novel interactors within the canonical Wnt/β-catenin pathway [\[46](#page-12-5)]. CHD8 is an ATP-dependent chromatin-remodeling factor [\[47](#page-12-6)] and may serve as a "master regulator" for other ASD risk genes during fetal development [[44,](#page-12-3) [48\]](#page-12-7). Knockdown of CHD8 in human neural stem cells affects the expression of several ASD risk genes [\[44](#page-12-3)], and human patients with mutations in CHD8 display ASD symptoms and have macrocephaly and gastrointestinal difficulties [\[37](#page-11-13)]. Taken together, these data suggested that CHD8 targets a set of genes during brain development and regulates other ASD risk genes [[44\]](#page-12-3). Some of the ASD risk genes regulate developmental processes in the cerebellum [[13\]](#page-10-10). These include genes encoding for Reelin, RORα, EN2, BDNF, neuroligins, and neurexins [[8,](#page-10-9) [49\]](#page-12-8).

RELN dysregulation has been observed in a subset of autistic individuals (reviewed by Ishii et al. [\[36](#page-11-12), [49\]](#page-12-8)). Reelin, encoded by the RELN gene (located on chromosome 7 in human and chromosome 5 in mice), is a 388 kDa extracellular matrix glycoprotein which is essential for proper neuronal migration and positioning during embryonic and perinatal development of the brain/cerebellum [[8,](#page-10-9) [36\]](#page-11-12). Though the precise mechanisms of RELN's role in ASD pathogenesis is uncertain, trinucleotide repeat expansion in the RELN gene has been observed in autistic individuals [\[8](#page-10-9), [49\]](#page-12-8). Persico et al. (2001) first reported that the polymorphic GGC repeats located in the 50 untranslated region (50 UTR) of the RELN are associated with ASD disorder [\[50](#page-12-9)]. The finding was subsequently replicated in three studies [[51–](#page-12-10) [53\]](#page-12-11), but there were no confirmed association between the triplet repeats in the 50 UTR of the RELN and autism. The family-based association analyses revealed that many CGG repeats present in RELN alleles may cause ASD particularly in patients with speech difficulties [\[54](#page-12-12)].

Reelin mutations in mouse models lead to irregular cortex formation and abnormal layering which may be responsible for behavioral and neurological disorders [\[55\]](#page-12-13). Adulthood changes in Reelin protein level caused cognitive impairment and reduced synaptic plasticity [[55](#page-12-13)[–58](#page-13-0)]. Given that the genetic evidence implicates RELN in the etiopathology of ASD, it has been attempted to add biochemical evidence by measuring the Reelin level in brain tissue and blood by using Western blotting. They showed that the levels of Reelin were significantly reduced in patients with ASD [[59\]](#page-13-1).

Several lines of evidence indicate that genes encoding retinoic acid receptorrelated orphan receptors (RORs) are also associated with ASD. The RORα, RORβ, and RORγ are nuclear receptors that regulate a range of physiological processes during brain development  $[60–62]$  $[60–62]$  $[60–62]$  $[60–62]$ . ROR $\alpha$  and ROR $\gamma$  are broadly expressed in the body, whereas ROR $\beta$  expression is more restricted to the central nervous system [[62,](#page-13-3) [63\]](#page-13-4). RORα protein expression significantly decreases in the brains of ASD patients probably through epigenetic alterations [\[64](#page-13-5)]. Devanna and Vernes demonstrated that miR-137, a microRNA implicated in neuropsychiatric disorders, targets a number of genes associated with ASD including ROR $\alpha$  [[65\]](#page-13-6). ROR $\alpha$  is a transcription factor that is critically important for development of the cerebellum [\[60,](#page-13-2) [61](#page-13-7), [66](#page-13-8)]. The role of the RORα in neural development has been demonstrated in mouse strain *staggerer*, which harbors a spontaneous deletion within ROR $\alpha$  [[67](#page-13-9)]. These mice have small stature and develop ataxia and hypotonia. The major neural deficit was underdevelopment of the cerebellar cortex with a pronounced deficiency in both granule and Purkinje cells [\[67](#page-13-9)]. Furthermore, disruption of RORα in *staggerer* mice shows behavioral phenotypes such as abnormal spatial learning, reduced exploration, limited maze patrolling, and perseverative behavior, which are associated with ASD [\[61](#page-13-7), [62\]](#page-13-3).

Engrailed 2 (EN2), a homeobox transcription factor, has been associated with normal cerebellar development, and mutations or deletions of EN2 result in reduced cerebellum volume and structural abnormalities [[68](#page-13-10), [69](#page-13-11)], which are both associated with susceptibility to ASD [[70](#page-13-12)]. Brain-derived neurotrophic factor (BDNF) plays a key role in the development of the nervous system and modulation of neuronal activity, both of which impact complex human behaviors. Several studies have been performed to measure peripheral blood levels of BDNF in an attempt to

find a biomarker for children with ASD. Peripheral blood levels of BDNF are known to be highly correlated with brain BDNF levels [\[71](#page-13-13)]. Although there is no consistency in the association between BDNF levels in blood and ASD, a recent review by Xiao-Yan et al. (2016) using meta-analysis indicated that there are increased peripheral blood levels of BDNF in ASD patients [[72\]](#page-13-14). Furthermore, Ca2+-dependent activator protein for secretion 2 (CADPS2) contributes to normal cerebellar development by enhancing release of BDNF and neurotrophin-3 (NT-3) [\[73](#page-13-15), [74\]](#page-14-0). The CADPS family is a secretory-related protein family that regulates secretory granule exocytosis, which in vertebrates consists of two genes, CAPS1/ CADPS1 and CAPS2/CADPS2. The expression level of the CAPS2 has been observed to be unusually high in some patients with ASD [\[38](#page-11-14), [75](#page-14-1)].

Mutations in the methyl CpG-binding protein 2 (MECP2) gene are known to cause Rett syndrome, a disorder characterized by language impairments, motor deficiencies, and stereotypical behavior [\[76](#page-14-2)], which is under the umbrella of ASD. Patients with Rett syndrome frequently have cerebellar atrophy that increases with age [\[13](#page-10-10)] (see chapter ["Epigenetics and Cerebellar Neurodevelopmental Disorders](http://dx.doi.org/10.1007/978-3-319-59749-2_10)").

Tuberous sclerosis complex (TSC) is a genetic disease that causes benign tumors in the body, including the brain [\[77\]](#page-14-3). Mutation in the TSC1 and TSC2 genes causes TSC with a neurodevelopmental disorder that involves higher rates of ASD [\[77,](#page-14-3) [78\]](#page-14-4). TSC produces a protein that negatively regulates the target of the rapamycin (mTOR) signaling pathway to control molecular and cellular process. Tsai et al. (2012) designed a mutant mouse model in which the gene for Tsc1 is not expressed in Purkinje cells [[78\]](#page-14-4). These mutant mice displayed ASD-like behaviors, such as abnormal social interaction and ultrasonic vocalization, and inflexibility. In addition, recent discovery has shown that the granule cells/Purkinje cells are important for cognitive processing in the cerebellum [[79\]](#page-14-5). These studies are significant because they demonstrated a clear involvement of the cerebellum in non-motor functions as well [\[78](#page-14-4)].

#### *Environmental Factors*

It has been suggested that the risk of developing ASD increases with exposure to environmental factors such as teratogenic substances (e.g., thalidomide, valproate, and misoprostol), infection with viruses (e.g., influenza, rubella, and cytomegalovirus) during pregnancy, and advanced age of parents (for reviews, see references [\[38](#page-11-14), [80](#page-14-6), [81\]](#page-14-7)).

Some environmental risk factors such as exposure to valproic acid during prenatal development may cause abnormalities in cerebellar development and ASD [[82](#page-14-8)]. In rat, valproic acid exposure reduces the number of Purkinje cells in the cerebellum accompanied by increases in the number of apoptotic cells [[83\]](#page-14-9). Cole et al. (2011) have shown changes in cerebellar gene expression in mice treated with chlorpyrifos [\[84](#page-14-10)]. Dermal exposure of young adult mice to chlorpyrifos causes increased glial fibrillary acidic protein expression of the cerebellum [\[85\]](#page-14-11). Furthermore, Purkinje cell numbers are reduced in rats prenatally exposed to chlorpyrifos [\[86\]](#page-14-12). Other factors

such as organophosphate pesticides and antiepileptic drugs have been shown to affect cerebellar development and potentially cause ASD [[87\]](#page-14-13).

Maternal fever is another environmental risk factor that affects the cerebellum and leads to apoptosis. It also interferes with neuronal maturation and may cause heat shock protein activation during cerebellum development in ASD [[88–](#page-14-14)[90\]](#page-14-15).

Viral infections can affect cerebellar and neocortical development during preand neonatal and cause neuropathy in ASD [[91,](#page-14-16) [92\]](#page-14-17). Influenza virus also has the same impact on cerebellum development such as reduced the number of Purkinje cells and interruption in migration of Purkinje and granule cells during perinatal development, which may cause deficits in working memory and behavioral impairments [[93–](#page-15-0)[96\]](#page-15-1) (see chapter ["Infections of the Cerebellum](http://dx.doi.org/10.1007/978-3-319-59749-2_12)").

Functional gastrointestinal disorders (FGIDs) are disorders independent of organic or physiological conditions that are the most common causes of GI disorders in children with ASD. FGID symptoms include abdominal pain, constipation, irritable bowel syndrome, and functional dyspepsia [\[97](#page-15-2)]. The FGIDs are associated with impaired behaviors and sensory responses, as well as changes in sleep patterns [\[98](#page-15-3)]. It is suggested that inadequate brain-gut interactions may be responsible for these symptoms in ASD patients [[97\]](#page-15-2). Changing the gut microbiome to treat the ASD behaviors such as anxiety and depression is a new line of study that hopes to find alternate treatments for ASD patients [\[99](#page-15-4), [100](#page-15-5)].

The exposure to air pollution, which may cause immune responses, is another likely environmental risk factor for ASD [[101\]](#page-15-6). The immune response results in activation of immune cells and antibody production and increases the leukocyte migration to the brain tissue by increasing diffusion through the blood-brain barrier. It is suggested that maternal immune activation at a critical time points impair cerebellar morphology and a variety of motor and non-motor behaviors [\[102](#page-15-7)]. The abnormal level of blood immunological markers in ASD patients is evidence of interactions between genetic/environmental factors with their immune system in these patients is shown [\[103](#page-15-8), [104\]](#page-15-9) (see chapter "[Neuroimmune Mechanisms of](http://dx.doi.org/10.1007/978-3-319-59749-2_13)  [Cerebellar Development and Its Developmental Disorders: Bidirectional Link](http://dx.doi.org/10.1007/978-3-319-59749-2_13)  [Between the Immune System and Nervous System](http://dx.doi.org/10.1007/978-3-319-59749-2_13)").

#### **Diagnosis of ASD**

Studies on patients with ASD using advance brain imaging, genetic, and behavioral observations improved our knowledge of ASD symptoms. As of yet there are no biomarkers for the diagnosis of ASD, and currently clinical diagnosis of these patients is based on behavioral observations combined with patient history [\[22](#page-11-3), [105\]](#page-15-10). Three ASD diagnosis criteria – social reciprocity, communication, and restricted/repetitive behavior – have been published by *DSM*-IV. However, it recently has been revised by *DSM*-V and International Classification of Diseases, Tenth Edition (ICD-10), into two domains of diagnosis criteria, (1) deficits in social communication/interaction and (2) restricted and repetitive behaviors, with

evidence of persistent symptoms that cause functional impairment [\[105](#page-15-10)]. Murphy et al. (2016) highlighted three key issues, sleep, GI problems, and epilepsy, regarding physical health that may be important in the diagnosis of ASD patient [\[105](#page-15-10)]. The mental health issues are present both in adults and children with ASD, which include mood and anxiety disorders, obsessive-compulsive disorder (OCD), attentiondeficit hyperactivity disorder (ADHD), and psychotic disorders. These persist from childhood to adulthood in both sexes. Additionally, ASD patients have specific cognitive anomalies, including poor planning, decision-making, timing, and motor skills, which impact their daily activities [[105–](#page-15-10)[107\]](#page-15-11).

#### **The Cerebellum and ASD**

Cerebellar malformations have been associated with a range of developmental impairments and behavior disorders including ASD (see review by Bolduc and Limperopoulos [[31\]](#page-11-15)). Motor impairment and clumsiness has been noted as an important feature in ASD [[108\]](#page-15-12). It is shown that about 80% of children with ASD have motor coordination deficits, which have a positive correlation with the severity of the ASD and intellectual disabilities [\[109](#page-15-13), [110](#page-15-14)]. Cerebellar motor dysfunction in ASD includes eye movement abnormalities, fine and gross motor deficits, gait, balance and coordination impairment, postural instability, and motor learning deficits [\[109](#page-15-13), [111\]](#page-16-0). Motor impairments are among the earliest signs of an autistic phenotype [\[112](#page-16-1)]. It has been shown that motor impairments are predictive of the ASD outcome. During early movement activities, individuals, who are later diagnosed with ASD, have poor fine and gross motor skills that are accompanied by delays of language development [[109,](#page-15-13) [113](#page-16-2)]. Similarly, difficulties in oral and manual motor skills in infancy can label individuals as an ASD patient, and late speech fluency is predictable [[114\]](#page-16-3). In addition, early motor delays are more common in infants at risk for ASD and are related to later communication delays [[115\]](#page-16-4). Therefore, the timing of language acquisition may serve as an indicator for neurodevelopmental and behavior disorders and may be a marker to diagnose people with ASD.

Emotional/behavioral disturbance and communication disorders may be associated with the motor task performance in ASD patients [\[116](#page-16-5)]. It has been suggested that the lack of gesture and imitation in ASD patients might be related to motor dysfunction, providing a mechanism by which cerebellar dysfunction could impact the core social communication symptoms of patient with ASD [[109,](#page-15-13) [117\]](#page-16-6).

### **ASD and Cerebellar Structure Abnormalities**

Cerebellar abnormalities are the most consistently reported brain structural changes in ASD, such as decreased cerebellar cortex, which is a key landmark for diagnosis in ASD brains [\[109](#page-15-13), [118](#page-16-7)]. Cerebellar enlargement has been reported in ASD young children in comparison with the total brain volume and may be associated with the cerebellar white matter [\[26](#page-11-16), [119](#page-16-8), [120\]](#page-16-9). However, the growth rate declines later during development and eventually results in a smaller cerebellar volume by adulthood in ASD patients [[120,](#page-16-9) [121](#page-16-10)].

By MRI, cerebellar lobules VI and VII are hyperplastic in patients with ASD, and it is suggested that these alterations may be responsible for increased stereotype and repetitive movements [[122\]](#page-16-11). The language impairment in ASD may be associated with a decreased volume of the vermis and anterior lobe and abnormal left lateralization in lobule VIIIA [[123,](#page-16-12) [124\]](#page-16-13). A study conducted using voxel-based morphometry suggested that structural differences such as increase and decrease in cerebellar gray and white matters are related to specific abnormalities at the different stages of cerebellar development in ASD patients [[109,](#page-15-13) [125\]](#page-16-14).

Neurohistological studies show changes in the anatomy of the cerebellum in patients with ASD, including a decrease in the number of Purkinje cells [[126,](#page-16-15) [127\]](#page-16-16), immature cerebellar development [[128–](#page-16-17)[130\]](#page-17-0), morphological changes in the size of the cerebellar nuclei which are small and abnormal, and an increase in the number of Bergmann glia cells [[131](#page-17-1), [132](#page-17-2)]. The low density of the Purkinje cells in the cerebellum of ASD patients was observed in the vermis, Crus I–II, lobules IV–VI, and lobule X [\[133](#page-17-3)]. The smaller size of Purkinje cells may indicate the occurrence of an atrophic process [\[134](#page-17-4)]. Because of the large size, and due to numerous synapses with the parallel and climbing fibers, Purkinje cells have a high metabolic demand. Therefore they have extensive amounts of calcium storage that may cause increases in intracellular calcium, which elevates the risk of excitotoxicity and cell death [[135\]](#page-17-5).

It has been reported that the cortical-pontine-cerebellar-thalamic circuit is immature and abnormal both functionally and anatomically in patients with ASD [\[9](#page-10-15)]. It is also shown that the cerebellar input and output pathways in relation to neocortical areas are unusual in ASD patients [[136,](#page-17-6) [137\]](#page-17-7). The corticopontocerebellar pathway carries inputs that originate from the primary sensory and motor cortex, posterior parietal, prefrontal, orbitofrontal, cingulate, temporal, and basal nuclei and projects to the cerebellum [[138,](#page-17-8) [139\]](#page-17-9). Outputs originate from the cerebellar nuclei and through the thalamus project to the neocortex  $[140-142]$  $[140-142]$ . These circuits are specialized for cognitive and behavioral activities such as executive functions, language, and emotions. Thus, the cerebellum may be responsible for cognitive impairment, sensorimotor behavior, and social disconnection in ASD [\[13](#page-10-10)].

Eye gaze abnormalities during social interaction are an early diagnostic indicator in ASD patients. Gaze fixation is naturally used to fix the fovea on an image or object. The oculomotor system maintains fixation, which is supported by the nuclei of the brainstem. Therefore, inputs from the frontal eye fields and superior colliculus actively block the saccades away from the object of interest [\[143\]](#page-17-12). The pontine nuclei stimulate Purkinje cells in lobules VI–VII vermis cerebellum, and inhibitory outputs from the oculomotor vermis help to stop undesired eye movements and keep an image on the fovea [[144\]](#page-17-13) which could potentially be used as an early marker of ASD patients.

Control of upper limb movement is related to the frontoparietal cortex as well as the cerebellar cortex and its output nuclei [\[145](#page-17-14)]. Upper limb and manual motor deficits that are associated with atrophy of the intermediate and lateral cerebellum involve lobules I–V as well as more lateral areas of lobules V–VI extending into Crus I–II in patients with upper limb ataxia. [[146\]](#page-17-15). These loops are responsible for regulating the amplitude, duration, and timing of movements [\[147](#page-17-16), [148\]](#page-17-17). Patients with ASD have difficulties coordinating grasping and reaching activities [[149\]](#page-17-18). These difficulties may be caused by central defects in integrating sensory feedback information and motor output as well as deficits in neocortical-posterior cerebellar circuitry. The compromised motor learning in individuals with ASD could be related to disturbances in the anterior cerebellar lobules IV–VI and their connectivity to frontal as well as parietal regions of the cortex. These effects may damage upper limb and manual motor actions that ultimately impact the patient's ability to control motor behavior and learn new skills. Therefore, the development of more complex social motor skills in these patients is disabled. Medial and intermediate cerebellar circuits affected by insufficiency in both sensory feedback and forward control appear to cause motor impairments and difficulties in posture, gait, and walking in ASD patients [\[150](#page-18-0)]. The motor deficits start from infancy and extend to adolescence and adulthood [[108,](#page-15-12) [151–](#page-18-1)[153\]](#page-18-2).

Cognitive function deficits such as attention and memory impairment, executive function, and cognitive flexibility deficits are common features in ASD [[154](#page-18-3)]. The cerebellum communicates with Brodmann areas 46 and 9 of the prefrontal cortex, which are involved in cognitive functions, memory, planning, decision-making, and cognitive flexibility [[155](#page-18-4)[–157](#page-18-5)]. The cerebellum to prefrontal cortex pathway could affect cognitive functions directly or perhaps indirectly through the ventral tegmental area, which contains dopaminergic neurons that project and terminate in the prefrontal cortex [[158\]](#page-18-6). Notably, the function of the prefrontal cortex dopaminergic pathway is associated with attention selection, cognitive flexibility, and memory  $[157]$ . A maldevelopment of connectivity in connection of the cerebellum to this higher-order circuit may explain the cognitive involvement of the cerebellum in patients with ASD.

#### **Assessment and Treatment**

There is very limited accurate and practical information to assess, diagnose, and manage ASD conditions. Therefore, because the number of ASD patients has rapidly increased during past decade, there is an urgent need to improve knowledge and develop assessment tools and treatment of ASD patients [\[105](#page-15-10)].

ASD diagnosis can be difficult because of a large amount of heterogeneity, varying presentation, and variability in symptoms [[159\]](#page-18-7). There are no biomarkers to diagnose ASD; therefore behavioral presentation of the patients is used for diagnosis [\[160](#page-18-8)]. The gold standard for clinical diagnosis in these patients is based on current diagnostic classification systems and proceeding very careful assessment practices. These assessments include physical examination, hearing test, observation of children's behavior, and a structured parent interview that covers the patient's full developmental history [[160\]](#page-18-8). Currently, the best practice to diagnose ASD patients is a step-by-step strategy that is recommended by the American Psychological Association [[161\]](#page-18-9). This diagnostic strategy starts with child's parent/ caregiver concern and followed by a formal diagnostic assessment conducted by a pediatrician or/and appropriate referrals. The formal diagnostic assessment includes medical and functional evaluation such as everyday verbal and nonverbal skills and level of ability as well as analyze/assess of behaviors based on developmental aspect [\[162](#page-18-10)]. However, because of differences in cognitive function, age, language level, and the source of information, diagnosis of ASD is very difficult [[159\]](#page-18-7).

Children who are diagnosed with ASD need to be reevaluated continuously during preschool years to identify their weakness, inabilities, and difficulties [[159\]](#page-18-7). There are also some diagnostic instruments for ASD such as the Autism Diagnostic Observational Schedule – Generic (ADOS-G) [\[163](#page-18-11)], which assesses communication, play, and creative use of materials and possibilities for children who may have ASD. The best Screening Tool for ASD in Toddlers and Young Children (STAT) [\[164](#page-18-12)] is structured to identify children between 24 and 36 months of age with ASD. One of the measures of early communication in children 8–24 months is the Communication and Symbolic Behavior Scales (CSBS) [\[165](#page-18-13)]. Additionally for the parent interview, there is a clinical diagnostic instrument named the Autism Diagnostic Interview-Revised (ADI-R) that is addressing early development, communication/language, social interactions/interests, and restricted and repetitive behaviors [\[166](#page-18-14)]. The Social Communication Questionnaire (SCQ) is an appropriate method to get information from parents [[167\]](#page-18-15).

Usually an assessment starts with medical evaluation which will be conducted by physicians and, if the ASD suspected, referring the patient for diagnostic assessment as well as considering the pediatrician as an alternative referral. When diagnosis is confirmed, treatment planning should involve the professional health team [[159\]](#page-18-7).

#### **Summary**

Many genetic and environmental factors may cause ASD. The mechanisms are unknown, but presumably genetic and environmental factors affect normal brain development and, consequently, lead to functional disorders in patients with ASD.

There is mounting evidence that developmental abnormalities in the cerebellum may underlie the pathogenetic mechanisms that are associated with the ASD phenotype. Cerebellar developmental disorders that are associated with ASD pathogenesis show deficits in motor coordination, balance, motor memory, and higher-order dysfunctions including speech and attention regulation.

The major goal of management in ASD patients is early diagnosis for behavioral and medical interventions to enhance the functional ability of these children. The new approach involving brain-gut-microbiome interactions may provide a biomarker associated with GI disorders that could be helpful in the early diagnosis of these patients. Because the number of ASD patients is increasing, studies are needed to develop assessment tools and treatment, increase public awareness, and develop strategy for health care of patients with ASD.

## **References**

- <span id="page-10-0"></span>1. Millon T. On the history and future study of personality and its disorders. Annu Rev Clin Psychol. 2012;8:1–19. PubMed PMID: 22035244.
- 2. Tonge BJ, Dissanayake C, Brereton AV. Autism: fifty years on from Kanner. J Paediatr Child Health. 1994;30(2):102–7. PubMed PMID: 8198840.
- <span id="page-10-3"></span>3. Volkmar FR, McPartland JC. From Kanner to DSM-5: autism as an evolving diagnostic concept. Annu Rev Clin Psychol. 2014;10:193–212. PubMed PMID: 24329180.
- <span id="page-10-1"></span>4. Olmsted D, Blaxill M. Leo Kanner's mention of 1938 in his report on autism refers to his first patient. J Autism Dev Disord. 2016;46(1):340–1. PubMed PMID: 26231203.
- <span id="page-10-2"></span>5. Barahona-Correa JB, Filipe CN. A concise history of Asperger syndrome: the short reign of a troublesome diagnosis. Front Psychol. 2015;6:2024. PubMed PMID: 26834663. Pubmed Central PMCID: 4725185.
- <span id="page-10-4"></span>6. Won H, Mah W, Kim E. Autism spectrum disorder causes, mechanisms, and treatments: focus on neuronal synapses. Front Mol Neurosci. 2013;6:19. PubMed PMID: 23935565. Pubmed Central PMCID: 3733014.
- <span id="page-10-8"></span>7. Hampson DR, Blatt GJ. Autism spectrum disorders and neuropathology of the cerebellum. Front Neurosci. 2015;9:420. PubMed PMID: 26594141. Pubmed Central PMCID: 4635214.
- <span id="page-10-9"></span>8. Fatemi SH, Aldinger KA, Ashwood P, Bauman ML, Blaha CD, Blatt GJ, et al. Consensus paper: pathological role of the cerebellum in autism. Cerebellum. 2012;11(3):777–807. PubMed PMID: 22370873. Pubmed Central PMCID: 3677555.
- <span id="page-10-15"></span>9. Mosconi MW, Wang Z, Schmitt LM, Tsai P, Sweeney JA. The role of cerebellar circuitry alterations in the pathophysiology of autism spectrum disorders. Front Neurosci. 2015;9:296. PubMed PMID: 26388713. Pubmed Central PMCID: 4555040.
- <span id="page-10-5"></span>10. Hagmeyer S, Mangus K, Boeckers TM, Grabrucker AM. Effects of trace metal profiles characteristic for autism on synapses in cultured neurons. Neural Plast. 2015;2015:985083. PubMed PMID: 25802764. Pubmed Central PMCID: 4352758.
- <span id="page-10-6"></span>11. Ismail MM, Keynton RS, Mostapha MM, ElTanboly AH, Casanova MF, Gimel'farb GL, et al. Studying autism spectrum disorder with structural and diffusion magnetic resonance imaging: a survey. Front Hum Neurosci. 2016;10:211. PubMed PMID: 27242476. Pubmed Central PMCID: 4862981.
- <span id="page-10-7"></span>12. Vuong HE, Hsiao EY. Emerging roles for the gut microbiome in autism spectrum disorder. Biol Psychiatry. 2016;81:411–23. PubMed PMID:27773355.
- <span id="page-10-10"></span>13. Rogers TD, McKimm E, Dickson PE, Goldowitz D, Blaha CD, Mittleman G. Is autism a disease of the cerebellum? Integr Clin Pre-Clin Res Front Syst Neurosci. 2013;7:15. PubMed PMID: 23717269. Pubmed Central PMCID: 3650713.
- <span id="page-10-11"></span>14. Wallace GL, Dankner N, Kenworthy L, Giedd JN, Martin A. Age-related temporal and parietal cortical thinning in autism spectrum disorders. Brain J Neurol. 2010;133(Pt 12):3745–54. PubMed PMID: 20926367. Pubmed Central PMCID: 2995883.
- <span id="page-10-12"></span>15. Lainhart JE, Piven J, Wzorek M, Landa R, Santangelo SL, Coon H, et al. Macrocephaly in children and adults with autism. J Am Acad Child Adolesc Psychiatry. 1997;36(2):282–90. PubMed PMID: 9031582.
- <span id="page-10-13"></span>16. Herbert MR, Ziegler DA, Deutsch CK, O'Brien LM, Kennedy DN, Filipek PA, et al. Brain asymmetries in autism and developmental language disorder: a nested whole-brain analysis. Brain J Neurol. 2005;128(Pt 1):213–26. PubMed PMID: 15563515.
- 17. Schumann CM, Bloss CS, Barnes CC, Wideman GM, Carper RA, Akshoomoff N, et al. Longitudinal magnetic resonance imaging study of cortical development through early childhood in autism. J Neurosci: Off J Soc Neurosci. 2010;30(12):4419–27. PubMed PMID: 20335478. Pubmed Central PMCID: 2859218.
- <span id="page-10-14"></span>18. Wolff JJ, Gu H, Gerig G, Elison JT, Styner M, Gouttard S, et al. Differences in white matter fiber tract development present from 6 to 24 months in infants with autism. Am J Psychiatry. 2012;169(6):589–600. PubMed PMID: 22362397. Pubmed Central PMCID: 3377782.
- <span id="page-11-0"></span>19. Courchesne E, Mouton PR, Calhoun ME, Semendeferi K, Ahrens-Barbeau C, Hallet MJ, et al. Neuron number and size in prefrontal cortex of children with autism. JAMA. 2011;306(18):2001–10. PubMed PMID: 22068992.
- <span id="page-11-1"></span>20. Sudarov A. Defining the role of cerebellar Purkinje cells in autism spectrum disorders. Cerebellum. 2013;12(6):950–5. PubMed PMID: 23703312. Pubmed Central PMCID: 3795842.
- <span id="page-11-2"></span>21. Taylor MJ, Doesburg SM, Pang EW. Neuromagnetic vistas into typical and atypical development of frontal lobe functions. Front Hum Neurosci. 2014;8:453. PubMed PMID: 24994980. Pubmed Central PMCID: 4061489.
- <span id="page-11-3"></span>22. Ecker C, Bookheimer SY, Murphy DG. Neuroimaging in autism spectrum disorder: brain structure and function across the lifespan. Lancet Neurol. 2015;14(11):1121–34. PubMed PMID: 25891007.
- <span id="page-11-4"></span>23. D'Mello AM, Stoodley CJ. Cerebro-cerebellar circuits in autism spectrum disorder. Front Neurosci. 2015;9:408. PubMed PMID: 26594140. Pubmed Central PMCID: 4633503.
- <span id="page-11-5"></span>24. Basson MA, Wingate RJ. Congenital hypoplasia of the cerebellum: developmental causes and behavioral consequences. Front Neuroanat. 2013;7:29. PubMed PMID: 24027500. Pubmed Central PMCID: 3759752.
- <span id="page-11-6"></span>25. Palmen SJ, van Engeland H, Hof PR, Schmitz C. Neuropathological findings in autism. Brain J Neurol. 2004;127(Pt 12):2572–83. PubMed PMID: 15329353.
- <span id="page-11-16"></span>26. Amaral DG, Schumann CM, Nordahl CW. Neuroanatomy of autism. Trends Neurosci. 2008;31(3):137–45. PubMed PMID: 18258309.
- 27. Whitney ER, Kemper TL, Bauman ML, Rosene DL, Blatt GJ. Cerebellar Purkinje cells are reduced in a subpopulation of autistic brains: a stereological experiment using calbindin-D28k. Cerebellum. 2008;7(3):406–16. PubMed PMID: 18587625.
- <span id="page-11-7"></span>28. Geschwind DH, Levitt P. Autism spectrum disorders: developmental disconnection syndromes. Curr Opin Neurobiol. 2007;17(1):103–11. PubMed PMID: 17275283.
- <span id="page-11-8"></span>29. Dolen G, Sahin M. Editorial: essential pathways and circuits of autism pathogenesis. Front Neurosci. 2016;10:182. PubMed PMID: 27199644. Pubmed Central PMCID: 4844597.
- <span id="page-11-9"></span>30. ten Donkelaar HJ, Lammens M, Wesseling P, Thijssen HO, Renier WO. Development and developmental disorders of the human cerebellum. J Neurol. 2003;250(9):1025–36. PubMed PMID: 14504962.
- <span id="page-11-15"></span>31. Bolduc ME, Limperopoulos C. Neurodevelopmental outcomes in children with cerebellar malformations: a systematic review. Dev Med Child Neurol. 2009;51(4):256–67. PubMed PMID: 19191827.
- 32. Limperopoulos C. Autism spectrum disorders in survivors of extreme prematurity. Clin Perinatol. 2009;36(4):791–805. vi. PubMed PMID: 19944836.
- 33. Wang SS, Kloth AD, Badura A. The cerebellum, sensitive periods, and autism. Neuron. 2014;83(3):518–32. PubMed PMID: 25102558. Pubmed Central PMCID: 4135479.
- <span id="page-11-10"></span>34. Bolduc ME, Du Plessis AJ, Sullivan N, Khwaja OS, Zhang X, Barnes K, et al. Spectrum of neurodevelopmental disabilities in children with cerebellar malformations. Dev Med Child Neurol. 2011;53(5):409–16. PubMed PMID: 21418200.
- <span id="page-11-11"></span>35. Geschwind DH, State MW. Gene hunting in autism spectrum disorder: on the path to precision medicine. Lancet Neurol. 2015;14(11):1109–20. PubMed PMID: 25891009. Pubmed Central PMCID: 4694565.
- <span id="page-11-12"></span>36. Ishii K, Kubo KI, Nakajima K. Reelin and neuropsychiatric disorders. Front Cell Neurosci. 2016;10:229. PubMed PMID: 27803648. Pubmed Central PMCID: 5067484.
- <span id="page-11-13"></span>37. Barnard RA, Pomaville MB, O'Roak BJ. Mutations and modeling of the chromatin remodeler CHD8 define an emerging autism etiology. Front Neurosci. 2015;9:477. PubMed PMID: 26733790. Pubmed Central PMCID: 4681771.
- <span id="page-11-14"></span>38. Sadakata T, Shinoda Y, Sato A, Iguchi H, Ishii C, Matsuo M, et al. Mouse models of mutations and variations in autism spectrum disorder-associated genes: mice expressing Caps2/ Cadps2 copy number and alternative splicing variants. Int J Environ Res Public Health. 2013;10(12):6335–53. PubMed PMID: 24287856. Pubmed Central PMCID: 3881117.
- <span id="page-12-0"></span>39. Roppongi RT, Karimi B, Siddiqui TJ. Role of LRRTMs in synapse development and plasticity. Neurosci Res. 2016;116:18–28. PubMed PMID: 27810425.
- 40. Sudhof TC. Neuroligins and neurexins link synaptic function to cognitive disease. Nature. 2008;455(7215):903–11. PubMed PMID: 18923512. Pubmed Central PMCID: 2673233.
- 41. Sala C, Vicidomini C, Bigi I, Mossa A, Verpelli C. Shank synaptic scaffold proteins: keys to understanding the pathogenesis of autism and other synaptic disorders. J Neurochem. 2015;135(5):849–58. PubMed PMID: 26338675.
- <span id="page-12-1"></span>42. Baig DN, Yanagawa T, Tabuchi K. Distortion of the normal function of synaptic cell adhesion molecules by genetic variants as a risk for autism spectrum disorders. Brain Res Bull. 2017;129:82–90. PubMed PMID: 27743928.
- <span id="page-12-2"></span>43. Li X, Zou H, Brown WT. Genes associated with autism spectrum disorder. Brain Res Bull. 2012;88(6):543–52. PubMed PMID: 22688012.
- <span id="page-12-3"></span>44. Cotney J, Muhle RA, Sanders SJ, Liu L, Willsey AJ, Niu W, et al. The autism-associated chromatin modifier CHD8 regulates other autism risk genes during human neurodevelopment. Nat Commun. 2015;6:6404. PubMed PMID: 25752243. Pubmed Central PMCID: 4355952.
- <span id="page-12-4"></span>45. Willsey AJ, Sanders SJ, Li M, Dong S, Tebbenkamp AT, Muhle RA, et al. Coexpression networks implicate human midfetal deep cortical projection neurons in the pathogenesis of autism. Cell. 2013;155(5):997–1007. PubMed PMID: 24267886. Pubmed Central PMCID: 3995413.
- <span id="page-12-5"></span>46. Sakamoto I, Kishida S, Fukui A, Kishida M, Yamamoto H, Hino S, et al. A novel betacatenin-binding protein inhibits beta-catenin-dependent Tcf activation and axis formation. J Biol Chem. 2000;275(42):32871–8. PubMed PMID: 10921920.
- <span id="page-12-6"></span>47. O'Roak BJ, Vives L, Girirajan S, Karakoc E, Krumm N, Coe BP, et al. Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations. Nature. 2012;485(7397):246–50. PubMed PMID: 22495309. Pubmed Central PMCID: 3350576.
- <span id="page-12-7"></span>48. Hormozdiari F, Penn O, Borenstein E, Eichler EE. The discovery of integrated gene networks for autism and related disorders. Genome Res. 2015;25(1):142–54. PubMed PMID: 25378250. Pubmed Central PMCID: 4317170.
- <span id="page-12-8"></span>49. Fatemi SH. The role of Reelin in pathology of autism. Mol Psychiatry. 2002;7(9):919–20. PubMed PMID: 12399938.
- <span id="page-12-9"></span>50. Persico AM, D'Agruma L, Maiorano N, Totaro A, Militerni R, Bravaccio C, et al. Reelin gene alleles and haplotypes as a factor predisposing to autistic disorder. Mol Psychiatry. 2001;6(2):150–9. PubMed PMID: 11317216.
- <span id="page-12-10"></span>51. Zhang H, Liu X, Zhang C, Mundo E, Macciardi F, Grayson DR, et al. Reelin gene alleles and susceptibility to autism spectrum disorders. Mol Psychiatry. 2002;7(9):1012–7. PubMed PMID: 12399956.
- 52. Skaar DA, Shao Y, Haines JL, Stenger JE, Jaworski J, Martin ER, et al. Analysis of the RELN gene as a genetic risk factor for autism. Mol Psychiatry. 2005;10(6):563–71. PubMed PMID: 15558079.
- <span id="page-12-11"></span>53. Dutta S, Guhathakurta S, Sinha S, Chatterjee A, Ahmed S, Ghosh S, et al. Reelin gene polymorphisms in the Indian population: a possible paternal 5′UTR-CGG-repeat-allele effect on autism. Am J Med Genet Part B Neuropsychiatr Genet: Off Publ Int Soc Psychiatr Genet. 2007;144B(1):106–12. PubMed PMID: 16941662.
- <span id="page-12-12"></span>54. Chudley AE. Genetic landmarks through philately – autism spectrum disorders: a genetic update. Clin Genet. 2004;65(5):352–7. PubMed PMID: 15099341.
- <span id="page-12-13"></span>55. Fatemi SH, Stary JM, Halt AR, Realmuto GR. Dysregulation of Reelin and Bcl-2 proteins in autistic cerebellum. J Autism Dev Disord. 2001;31(6):529–35. PubMed PMID: 11814262.
- 56. Adams JM, Cory S. The Bcl-2 protein family: arbiters of cell survival. Science. 1998;281(5381):1322–6. PubMed PMID: 9735050.
- 57. de Bergeyck V, Nakajima K, Lambert de Rouvroit C, Naerhuyzen B, Goffinet AM, Miyata T, et al. A truncated Reelin protein is produced but not secreted in the 'Orleans' reeler mutation (Reln[rl-Orl]). Brain Res Mol Brain Res. 1997;50(1–2):85–90. PubMed PMID: 9406921.
- <span id="page-13-0"></span>58. Lacor PN, Grayson DR, Auta J, Sugaya I, Costa E, Guidotti A. Reelin secretion from glutamatergic neurons in culture is independent from neurotransmitter regulation. Proc Natl Acad Sci U S A. 2000;97(7):3556–61. PubMed PMID: 10725375. Pubmed Central PMCID: 16278.
- <span id="page-13-1"></span>59. Fatemi SH, Stary JM, Egan EA. Reduced blood levels of reelin as a vulnerability factor in pathophysiology of autistic disorder. Cell Mol Neurobiol. 2002;22(2):139–52. PubMed PMID: 12363196.
- <span id="page-13-2"></span>60. Boukhtouche F, Brugg B, Wehrle R, Bois-Joyeux B, Danan JL, Dusart I, et al. Induction of early Purkinje cell dendritic differentiation by thyroid hormone requires RORalpha. Neural Dev. 2010;5:18. PubMed PMID: 20663205. Pubmed Central PMCID: 2918593.
- <span id="page-13-7"></span>61. Hamilton BA, Frankel WN, Kerrebrock AW, Hawkins TL, FitzHugh W, Kusumi K, et al. Disruption of the nuclear hormone receptor RORalpha in staggerer mice. Nature. 1996;379(6567):736–9. PubMed PMID: 8602221.
- <span id="page-13-3"></span>62. Wang Y, Billon C, Walker JK, Burris TP. Therapeutic effect of a synthetic RORalpha/gamma agonist in an animal model of autism. ACS Chem Neurosci. 2016;7(2):143–8. PubMed PMID: 26625251. Pubmed Central PMCID: 4759619.
- <span id="page-13-4"></span>63. Huh JR, Leung MW, Huang P, Ryan DA, Krout MR, Malapaka RR, et al. Digoxin and its derivatives suppress TH17 cell differentiation by antagonizing RORgammat activity. Nature. 2011;472(7344):486–90. PubMed PMID: 21441909. Pubmed Central PMCID: 3172133.
- <span id="page-13-5"></span>64. Nguyen A, Rauch TA, Pfeifer GP, Hu VW. Global methylation profiling of lymphoblastoid cell lines reveals epigenetic contributions to autism spectrum disorders and a novel autism candidate gene, RORA, whose protein product is reduced in autistic brain. FASEB J: Off Publ Fed Am Soc Exp Biol. 2010;24(8):3036–51. PubMed PMID: 20375269. Pubmed Central PMCID: 2909294.
- <span id="page-13-6"></span>65. Devanna P, Vernes SC. A direct molecular link between the autism candidate gene RORa and the schizophrenia candidate MIR137. Sci Rep. 2014;4:3994. PubMed PMID: 24500708. Pubmed Central PMCID: 3915307.
- <span id="page-13-8"></span>66. Boukhtouche F, Doulazmi M, Frederic F, Dusart I, Brugg B, Mariani J. RORalpha, a pivotal nuclear receptor for Purkinje neuron survival and differentiation: from development to ageing. Cerebellum. 2006;5(2):97–104. PubMed PMID: 16818384.
- <span id="page-13-9"></span>67. Gold DA, Gent PM, Hamilton BA. ROR alpha in genetic control of cerebellum development: 50 staggering years. Brain Res. 2007;1140:19–25. PubMed PMID: 16427031.
- <span id="page-13-10"></span>68. Liu A, Losos K, Joyner AL. FGF8 can activate Gbx2 and transform regions of the rostral mouse brain into a hindbrain fate. Development. 1999;126(21):4827–38. PubMed PMID: 10518499.
- <span id="page-13-11"></span>69. Kuemerle B, Gulden F, Cherosky N, Williams E, Herrup K. The mouse Engrailed genes: a window into autism. Behav Brain Res. 2007;176(1):121–32. PubMed PMID: 17055592. Pubmed Central PMCID: 2791532.
- <span id="page-13-12"></span>70. Benayed R, Choi J, Matteson PG, Gharani N, Kamdar S, Brzustowicz LM, et al. Autismassociated haplotype affects the regulation of the homeobox gene, ENGRAILED 2. Biol Psychiatry. 2009;66(10):911–7. PubMed PMID: 19615670. Pubmed Central PMCID: 2783416.
- <span id="page-13-13"></span>71. Fernandes BS, Berk M, Turck CW, Steiner J, Goncalves CA. Decreased peripheral brainderived neurotrophic factor levels are a biomarker of disease activity in major psychiatric disorders: a comparative meta-analysis. Mol Psychiatry. 2014;19(7):750–1. PubMed PMID: 24342989.
- <span id="page-13-14"></span>72. Qin XY, Feng JC, Cao C, Wu HT, Loh YP, Cheng Y. Association of peripheral blood levels of brain-derived neurotrophic factor with autism spectrum disorder in children: a systematic review and meta-analysis. JAMA Pediatr. 2016;170(11):1079–86. PubMed PMID: 27654278.
- <span id="page-13-15"></span>73. Sato A, Sekine Y, Saruta C, Nishibe H, Morita N, Sato Y, et al. Cerebellar development transcriptome database (CDT-DB): profiling of spatio-temporal gene expression during the postnatal development of mouse cerebellum. Neural Netw: Off J Int Neural Netw Soc. 2008;21(8):1056–69. PubMed PMID: 18603407.
- <span id="page-14-0"></span>74. Sadakata T, Furuichi T. Developmentally regulated Ca2+−dependent activator protein for secretion 2 (CAPS2) is involved in BDNF secretion and is associated with autism susceptibility. Cerebellum. 2009;8(3):312–22. PubMed PMID: 19238500.
- <span id="page-14-1"></span>75. Sadakata T, Washida M, Iwayama Y, Shoji S, Sato Y, Ohkura T, et al. Autistic-like phenotypes in Cadps2-knockout mice and aberrant CADPS2 splicing in autistic patients. J Clin Invest. 2007;117(4):931–43. PubMed PMID: 17380209. Pubmed Central PMCID: 1821065.
- <span id="page-14-2"></span>76. Amir RE, Van den Veyver IB, Wan M, Tran CQ, Francke U, Zoghbi HY. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. Nat Genet. 1999;23(2):185–8. PubMed PMID: 10508514.
- <span id="page-14-3"></span>77. Ertan G, Arulrajah S, Tekes A, Jordan L, Huisman TA. Cerebellar abnormality in children and young adults with tuberous sclerosis complex: MR and diffusion weighted imaging findings. J Neuroradiol J Neuroradiol. 2010;37(4):231–8. PubMed PMID: 20381146.
- <span id="page-14-4"></span>78. Tsai PT, Hull C, Chu Y, Greene-Colozzi E, Sadowski AR, Leech JM, et al. Autisticlike behaviour and cerebellar dysfunction in Purkinje cell Tsc1 mutant mice. Nature. 2012;488(7413):647–51. PubMed PMID: 22763451. Pubmed Central PMCID: 3615424.
- <span id="page-14-5"></span>79. Wagner MJ, Kim TH, Savall J, Schnitzer MJ, Luo L. Cerebellar granule cells encode the expectation of reward. Nature. 2017;544:96–100. PubMed PMID: 28321129.
- <span id="page-14-6"></span>80. Folstein SE, Rosen-Sheidley B. Genetics of autism: complex aetiology for a heterogeneous disorder. Nat Rev Genet. 2001;2(12):943–55. PubMed PMID: 11733747.
- <span id="page-14-7"></span>81. Grabrucker AM. Environmental factors in autism. Front Psych. 2012;3:118. PubMed PMID: 23346059. Pubmed Central PMCID: 3548163.
- <span id="page-14-8"></span>82. Cheh MA, Millonig JH, Roselli LM, Ming X, Jacobsen E, Kamdar S, et al. En2 knockout mice display neurobehavioral and neurochemical alterations relevant to autism spectrum disorder. Brain Res. 2006;1116(1):166–76. PubMed PMID: 16935268.
- <span id="page-14-9"></span>83. Lee MH, Kim M, Lee BH, Kim JH, Kang KS, Kim HL, et al. Subchronic effects of valproic acid on gene expression profiles for lipid metabolism in mouse liver. Toxicol Appl Pharmacol. 2008;226(3):271–84. PubMed PMID: 17963808.
- <span id="page-14-10"></span>84. Cole TB, Fisher JC, Burbacher TM, Costa LG, Furlong CE. Neurobehavioral assessment of mice following repeated postnatal exposure to chlorpyrifos-oxon. Neurotoxicol Teratol. 2012;34(3):311–22. PubMed PMID: 22425525. Pubmed Central PMCID: 3367041.
- <span id="page-14-11"></span>85. Krishnan K, Mitra NK, Yee LS, Yang HM. A comparison of neurotoxicity in cerebellum produced by dermal application of chlorpyrifos in young and adult mice. J Neural Transm. 2012;119(3):345–52. PubMed PMID: 21922192.
- <span id="page-14-12"></span>86. Abou-Donia MB, Khan WA, Dechkovskaia AM, Goldstein LB, Bullman SL, Abdel-Rahman A. In utero exposure to nicotine and chlorpyrifos alone, and in combination produces persistent sensorimotor deficits and Purkinje neuron loss in the cerebellum of adult offspring rats. Arch Toxicol. 2006;80(9):620–31. PubMed PMID: 16482470.
- <span id="page-14-13"></span>87. Moore SJ, Turnpenny P, Quinn A, Glover S, Lloyd DJ, Montgomery T, et al. A clinical study of 57 children with fetal anticonvulsant syndromes. J Med Genet. 2000;37(7):489–97. PubMed PMID: 10882750. Pubmed Central PMCID: 1734633.
- <span id="page-14-14"></span>88. Khan VR, Brown IR. The effect of hyperthermia on the induction of cell death in brain, testis, and thymus of the adult and developing rat. Cell Stress Chaperones. 2002;7(1):73–90. PubMed PMID: 11892990. Pubmed Central PMCID: 514805.
- 89. Maroni P, Bendinelli P, Tiberio L, Rovetta F, Piccoletti R, Schiaffonati L. In vivo heat-shock response in the brain: signalling pathway and transcription factor activation. Brain Res Mol Brain Res. 2003;119(1):90–9. PubMed PMID: 14597233.
- <span id="page-14-15"></span>90. Dean SL, Wright CL, Hoffman JF, Wang M, Alger BE, McCarthy MM. Prostaglandin E2 stimulates estradiol synthesis in the cerebellum postnatally with associated effects on Purkinje neuron dendritic arbor and electrophysiological properties. Endocrinology. 2012;153(11):5415–27. PubMed PMID: 23054057. Pubmed Central PMCID: 3473195.
- <span id="page-14-16"></span>91. Johnson RT. Effects of viral infection on the developing nervous system. N Engl J Med. 1972;287(12):599–604. PubMed PMID: 4560094.
- <span id="page-14-17"></span>92. Atladottir HO, Thorsen P, Ostergaard L, Schendel DE, Lemcke S, Abdallah M, et al. Maternal infection requiring hospitalization during pregnancy and autism spectrum disorders. J Autism Dev Disord. 2010;40(12):1423–30. PubMed PMID: 20414802.
- <span id="page-15-0"></span>93. Shi L, Fatemi SH, Sidwell RW, Patterson PH. Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring. J Neurosci: Off J Soc Neurosci. 2003;23(1):297–302. PubMed PMID: 12514227.
- 94. Beraki S, Aronsson F, Karlsson H, Ogren SO, Kristensson K. Influenza A virus infection causes alterations in expression of synaptic regulatory genes combined with changes in cognitive and emotional behaviors in mice. Mol Psychiatry. 2005;10(3):299–308. PubMed PMID: 15241434.
- 95. Asp L, Beraki S, Kristensson K, Ogren SO, Karlsson H. Neonatal infection with neurotropic influenza A virus affects working memory and expression of type III Nrg1 in adult mice. Brain Behav Immun. 2009;23(6):733–41. PubMed PMID: 19362585.
- <span id="page-15-1"></span>96. Shi L, Smith SE, Malkova N, Tse D, Su Y, Patterson PH. Activation of the maternal immune system alters cerebellar development in the offspring. Brain Behav Immun. 2009;23(1):116– 23. PubMed PMID: 18755264. Pubmed Central PMCID: 2614890.
- <span id="page-15-2"></span>97. Luna RA, Savidge TC, Williams KC. The brain-gut-microbiome axis: what role does it play in autism spectrum disorder? Curr Dev Disord Rep. 2016;3(1):75–81. PubMed PMID: 27398286. Pubmed Central PMCID: 4933016.
- <span id="page-15-3"></span>98. Mazurek MO, Vasa RA, Kalb LG, Kanne SM, Rosenberg D, Keefer A, et al. Anxiety, sensory over-responsivity, and gastrointestinal problems in children with autism spectrum disorders. J Abnorm Child Psychol. 2013;41(1):165–76. PubMed PMID: 22850932.
- <span id="page-15-4"></span>99. Messaoudi M, Lalonde R, Violle N, Javelot H, Desor D, Nejdi A, et al. Assessment of psychotropic-like properties of a probiotic formulation (Lactobacillus helveticus R0052 and Bifidobacterium longum R0175) in rats and human subjects. Br J Nutr. 2011;105(5):755–64. PubMed PMID: 20974015.
- <span id="page-15-5"></span>100. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, et al. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proc Natl Acad Sci U S A. 2011;108(38):16050–5. PubMed PMID: 21876150. Pubmed Central PMCID: 3179073.
- <span id="page-15-6"></span>101. Flores-Pajot MC, Ofner M, Do MT, Lavigne E, Villeneuve PJ. Childhood autism spectrum disorders and exposure to nitrogen dioxide, and particulate matter air pollution: a review and meta-analysis. Environ Res. 2016;151:763–76. PubMed PMID: 27609410.
- <span id="page-15-7"></span>102. Aavani T, Rana SA, Hawkes R, Pittman QJ. Maternal immune activation produces cerebellar hyperplasia and alterations in motor and social behaviors in male and female mice. Cerebellum. 2015;14(5):491–505. PubMed PMID: 25863812.
- <span id="page-15-8"></span>103. Gottfried C, Bambini-Junior V, Francis F, Riesgo R, Savino W. The impact of neuroimmune alterations in autism spectrum disorder. Front Psych. 2015;6:121. PubMed PMID: 26441683. Pubmed Central PMCID: 4563148.
- <span id="page-15-9"></span>104. Verkhratsky A, Rodriguez JJ, Parpura V. Neuroglia in ageing and disease. Cell Tissue Res. 2014;357(2):493–503. PubMed PMID: 24652503.
- <span id="page-15-10"></span>105. Murphy CM, Wilson CE, Robertson DM, Ecker C, Daly EM, Hammond N, et al. Autism spectrum disorder in adults: diagnosis, management, and health services development. Neuropsychiatr Dis Treat. 2016;12:1669–86. PubMed PMID: 27462160. Pubmed Central PMCID: 4940003.
- 106. Johnston K, Dittner A, Bramham J, Murphy C, Knight A, Russell A. Attention deficit hyperactivity disorder symptoms in adults with autism spectrum disorders. Autism Res: Off J Int Soc Autism Res. 2013;6(4):225–36. PubMed PMID: 23788522.
- <span id="page-15-11"></span>107. Russell AJ, Murphy CM, Wilson E, Gillan N, Brown C, Robertson DM, et al. The mental health of individuals referred for assessment of autism spectrum disorder in adulthood: a clinic report. Autism Int J Res Pract. 2016;20(5):623–7. PubMed PMID: 26471427.
- <span id="page-15-12"></span>108. Fournier KA, Hass CJ, Naik SK, Lodha N, Cauraugh JH. Motor coordination in autism spectrum disorders: a synthesis and meta-analysis. J Autism Dev Disord. 2010;40(10):1227–40. PubMed PMID: 20195737.
- <span id="page-15-13"></span>109. Becker EB, Stoodley CJ. Autism spectrum disorder and the cerebellum. Int Rev Neurobiol. 2013;113:1–34. PubMed PMID: 24290381.
- <span id="page-15-14"></span>110. Hilton CL, Zhang Y, Whilte MR, Klohr CL, Constantino J. Motor impairment in sibling pairs concordant and discordant for autism spectrum disorders. Autism Int J Res Pract. 2012;16(4):430–41. PubMed PMID: 22013131. Pubmed Central PMCID: 4222044.
- <span id="page-16-0"></span>111. Gowen E, Hamilton A. Motor abilities in autism: a review using a computational context. J Autism Dev Disord. 2013;43(2):323–44. PubMed PMID: 22723127.
- <span id="page-16-1"></span>112. Zwaigenbaum L, Bryson S, Garon N. Early identification of autism spectrum disorders. Behav Brain Res. 2013;251:133–46. PubMed PMID: 23588272.
- <span id="page-16-2"></span>113. Landa R, Garrett-Mayer E. Development in infants with autism spectrum disorders: a prospective study. J Child Psychol Psychiatry. 2006;47(6):629–38. PubMed PMID: 16712640.
- <span id="page-16-3"></span>114. Gernsbacher MA, Sauer EA, Geye HM, Schweigert EK, Hill GH. Infant and toddler oraland manual-motor skills predict later speech fluency in autism. J Child Psychol Psychiatry. 2008;49(1):43–50. PubMed PMID: 17979963. Pubmed Central PMCID: 4123528.
- <span id="page-16-4"></span>115. Bhat AN, Galloway JC, Landa RJ. Relation between early motor delay and later communication delay in infants at risk for autism. Infant Behav Dev. 2012;35(4):838–46. PubMed PMID: 22982285. Pubmed Central PMCID: 3538350.
- <span id="page-16-5"></span>116. Papadopoulos N, McGinley J, Tonge B, Bradshaw J, Saunders K, Murphy A, et al. Motor proficiency and emotional/behavioural disturbance in autism and Asperger's disorder: another piece of the neurological puzzle? Autism Int J Res Pract. 2012;16(6):627–40. PubMed PMID: 21949004.
- <span id="page-16-6"></span>117. Jones V, Prior M. Motor imitation abilities and neurological signs in autistic children. J Autism Dev Disord. 1985;15(1):37–46. PubMed PMID: 3980428.
- <span id="page-16-7"></span>118. Ecker C, Rocha-Rego V, Johnston P, Mourao-Miranda J, Marquand A, Daly EM, et al. Investigating the predictive value of whole-brain structural MR scans in autism: a pattern classification approach. NeuroImage. 2010;49(1):44–56. PubMed PMID: 19683584.
- <span id="page-16-8"></span>119. Stanfield AC, McIntosh AM, Spencer MD, Philip R, Gaur S, Lawrie SM. Towards a neuroanatomy of autism: a systematic review and meta-analysis of structural magnetic resonance imaging studies. Eur Psychiatry J Assoc Eur Psychiatr. 2008;23(4):289–99. PubMed PMID: 17765485.
- <span id="page-16-9"></span>120. Courchesne E, Karns CM, Davis HR, Ziccardi R, Carper RA, Tigue ZD, et al. Unusual brain growth patterns in early life in patients with autistic disorder: an MRI study. Neurology. 2001;57(2):245–54. PubMed PMID: 11468308.
- <span id="page-16-10"></span>121. Hallahan B, Daly EM, McAlonan G, Loth E, Toal F, O'Brien F, et al. Brain morphometry volume in autistic spectrum disorder: a magnetic resonance imaging study of adults. Psychol Med. 2009;39(2):337–46. PubMed PMID: 18775096.
- <span id="page-16-11"></span>122. Courchesne E, Campbell K, Solso S. Brain growth across the life span in autism: age-specific changes in anatomical pathology. Brain Res. 2011;1380:138–45. PubMed PMID: 20920490. Pubmed Central PMCID: 4500507.
- <span id="page-16-12"></span>123. Murakami JW, Courchesne E, Press GA, Yeung-Courchesne R, Hesselink JR. Reduced cerebellar hemisphere size and its relationship to vermal hypoplasia in autism. Arch Neurol. 1989;46(6):689–94. PubMed PMID: 2730382.
- <span id="page-16-13"></span>124. Hodge SM, Makris N, Kennedy DN, Caviness VS Jr, Howard J, McGrath L, et al. Cerebellum, language, and cognition in autism and specific language impairment. J Autism Dev Disord. 2010;40(3):300–16. PubMed PMID: 19924522. Pubmed Central PMCID: 3771698.
- <span id="page-16-14"></span>125. Cheung C, Chua SE, Cheung V, Khong PL, Tai KS, Wong TK, et al. White matter fractional anisotrophy differences and correlates of diagnostic symptoms in autism. J Child Psychol Psychiatry. 2009;50(9):1102–12. PubMed PMID: 19490309.
- <span id="page-16-15"></span>126. Bauman M, Kemper TL. Histoanatomic observations of the brain in early infantile autism. Neurology. 1985;35(6):866–74. PubMed PMID: 4000488.
- <span id="page-16-16"></span>127. Whitney ER, Kemper TL, Rosene DL, Bauman ML, Blatt GJ. Density of cerebellar basket and stellate cells in autism: evidence for a late developmental loss of Purkinje cells. J Neurosci Res. 2009;87(10):2245–54. PubMed PMID: 19301429. Pubmed Central PMCID: 2760265.
- <span id="page-16-17"></span>128. Courchesne E. Brainstem, cerebellar and limbic neuroanatomical abnormalities in autism. Curr Opin Neurobiol. 1997;7(2):269–78. PubMed PMID: 9142760.
- 129. Nicot A, Lelievre V, Tam J, Waschek JA, DiCicco-Bloom E. Pituitary adenylate cyclaseactivating polypeptide and sonic hedgehog interact to control cerebellar granule precursor cell proliferation. J Neurosci: Off J Soc Neurosci. 2002;22(21):9244–54. PubMed PMID: 12417650.
- <span id="page-17-0"></span>130. DiCicco-Bloom E, Lord C, Zwaigenbaum L, Courchesne E, Dager SR, Schmitz C, et al. The developmental neurobiology of autism spectrum disorder. J Neurosci: Off J Soc Neurosci. 2006;26(26):6897–906. PubMed PMID: 16807320.
- <span id="page-17-1"></span>131. Vargas DL, Nascimbene C, Krishnan C, Zimmerman AW, Pardo CA. Neuroglial activation and neuroinflammation in the brain of patients with autism. Ann Neurol. 2005;57(1):67–81. PubMed PMID: 15546155.
- <span id="page-17-2"></span>132. Bauman ML, Kemper TL. Neuroanatomic observations of the brain in autism: a review and future directions. Int J Dev Neurosci: Off J Int Soc Dev Neurosci. 2005;23(2–3):183–7. PubMed PMID: 15749244.
- <span id="page-17-3"></span>133. Skefos J, Cummings C, Enzer K, Holiday J, Weed K, Levy E, et al. Regional alterations in Purkinje cell density in patients with autism. PLoS One. 2014;9(2):e81255. PubMed PMID: 24586223. Pubmed Central PMCID: 3933333.
- <span id="page-17-4"></span>134. Fatemi SH, Halt AR, Realmuto G, Earle J, Kist DA, Thuras P, et al. Purkinje cell size is reduced in cerebellum of patients with autism. Cell Mol Neurobiol. 2002;22(2):171–5. PubMed PMID: 12363198.
- <span id="page-17-5"></span>135. Vajda S, Vakser IA, Sternberg MJ, Janin J. Modeling of protein interactions in genomes. Proteins. 2002;47(4):444–6. PubMed PMID: 12001222.
- <span id="page-17-6"></span>136. Catani M, Jones DK, Daly E, Embiricos N, Deeley Q, Pugliese L, et al. Altered cerebellar feedback projections in Asperger syndrome. NeuroImage. 2008;41(4):1184–91. PubMed PMID: 18495494.
- <span id="page-17-7"></span>137. Shukla DK, Keehn B, Lincoln AJ, Muller RA. White matter compromise of callosal and subcortical fiber tracts in children with autism spectrum disorder: a diffusion tensor imaging study. J Am Acad Child Adolesc Psychiatry. 2010;49(12):1269–78. 78 e1-2. PubMed PMID: 21093776. Pubmed Central PMCID: 3346956.
- <span id="page-17-8"></span>138. Dum RP, Strick PL. An unfolded map of the cerebellar dentate nucleus and its projections to the cerebral cortex. J Neurophysiol. 2003;89(1):634–9. PubMed PMID: 12522208.
- <span id="page-17-9"></span>139. Eccles JC, Sasaki K, Strata P. Interpretation of the potential fields generated in the cerebellar cortex by a mossy fibre volley. Exp Brain Res. 1967;3(1):58–80. PubMed PMID: 6031000.
- <span id="page-17-10"></span>140. Percheron G, Francois C, Talbi B, Yelnik J, Fenelon G. The primate motor thalamus. Brain Res Brain Res Rev. 1996;22(2):93–181. PubMed PMID: 8883918.
- 141. Leiner HC, Leiner AL, Dow RS. The human cerebro-cerebellar system: its computing, cognitive, and language skills. Behav Brain Res. 1991;44(2):113–28. PubMed PMID: 1751002.
- <span id="page-17-11"></span>142. Leiner HC, Leiner AL, Dow RS. Cognitive and language functions of the human cerebellum. Trends Neurosci. 1993;16(11):444–7. PubMed PMID: 7507614.
- <span id="page-17-12"></span>143. Zuber BL, Stark L, Cook G. Microsaccades and the velocity-amplitude relationship for saccadic eye movements. Science. 1965;150(3702):1459–60. PubMed PMID: 5855207.
- <span id="page-17-13"></span>144. Kase M, Miller DC, Noda H. Discharges of Purkinje cells and mossy fibres in the cerebellar vermis of the monkey during saccadic eye movements and fixation. J Physiol. 1980;300:539– 55. PubMed PMID: 6770085. Pubmed Central PMCID: 1279371.
- <span id="page-17-14"></span>145. Grodd W, Hulsmann E, Lotze M, Wildgruber D, Erb M. Sensorimotor mapping of the human cerebellum: fMRI evidence of somatotopic organization. Hum Brain Mapp. 2001;13(2):55– 73. PubMed PMID: 11346886.
- <span id="page-17-15"></span>146. Vaillancourt DE, Mayka MA, Corcos DM. Intermittent visuomotor processing in the human cerebellum, parietal cortex, and premotor cortex. J Neurophysiol. 2006;95(2):922–31. PubMed PMID: 16267114. Pubmed Central PMCID: 2366036.
- <span id="page-17-16"></span>147. Spraker MB, Corcos DM, Kurani AS, Prodoehl J, Swinnen SP, Vaillancourt DE. Specific cerebellar regions are related to force amplitude and rate of force development. NeuroImage. 2012;59(2):1647–56. PubMed PMID: 21963915. Pubmed Central PMCID: 3230677.
- <span id="page-17-17"></span>148. Neely KA, Coombes SA, Planetta PJ, Vaillancourt DE. Segregated and overlapping neural circuits exist for the production of static and dynamic precision grip force. Hum Brain Mapp. 2013;34(3):698–712. PubMed PMID: 22109998. Pubmed Central PMCID: 3292669.
- <span id="page-17-18"></span>149. Brisson J, Warreyn P, Serres J, Foussier S, Adrien-Louis J. Motor anticipation failure in infants with autism: a retrospective analysis of feeding situations. Autism Int J Res Pract. 2012;16(4):420–9. PubMed PMID: 22250193.
- <span id="page-18-0"></span>150. Cheng Y, Chou KH, Chen IY, Fan YT, Decety J, Lin CP. Atypical development of white matter microstructure in adolescents with autism spectrum disorders. NeuroImage. 2010;50(3):873– 82. PubMed PMID: 20074650.
- <span id="page-18-1"></span>151. Provost B, Lopez BR, Heimerl S. A comparison of motor delays in young children: autism spectrum disorder, developmental delay, and developmental concerns. J Autism Dev Disord. 2007;37(2):321–8. PubMed PMID: 16868847.
- 152. Brian J, Bryson SE, Garon N, Roberts W, Smith IM, Szatmari P, et al. Clinical assessment of autism in high-risk 18-month-olds. Autism Int J Res Pract. 2008;12(5):433–56. PubMed PMID: 18805941.
- <span id="page-18-2"></span>153. Van Waelvelde H, Oostra A, Dewitte G, Van Den Broeck C, Jongmans MJ. Stability of motor problems in young children with or at risk of autism spectrum disorders, ADHD, and or developmental coordination disorder. Dev Med Child Neurol. 2010;52(8):e174–8. PubMed PMID: 20132135.
- <span id="page-18-3"></span>154. Solomon M, Ozonoff SJ, Cummings N, Carter CS. Cognitive control in autism spectrum disorders. Int J Dev Neurosci: Off J Int Soc Dev Neurosci. 2008;26(2):239–47. PubMed PMID: 18093787. Pubmed Central PMCID: 2695998.
- <span id="page-18-4"></span>155. Middleton FA, Strick PL. Basal ganglia output and cognition: evidence from anatomical, behavioral, and clinical studies. Brain Cogn. 2000;42(2):183–200. PubMed PMID: 10744919.
- 156. Schmahmann JD. From movement to thought: anatomic substrates of the cerebellar contribution to cognitive processing. Hum Brain Mapp. 1996;4(3):174–98. PubMed PMID: 20408197.
- <span id="page-18-5"></span>157. Robbins TW, Roberts AC. Differential regulation of fronto-executive function by the monoamines and acetylcholine. Cereb Cortex. 2007;17(Suppl 1):i151–60. PubMed PMID: 17725997.
- <span id="page-18-6"></span>158. Fallon JH, Riley JN, Moore RY. Substantia nigra dopamine neurons: separate populations project to neostriatum and allocortex. Neurosci Lett. 1978;7(2–3):157–62. PubMed PMID: 19605105.
- <span id="page-18-7"></span>159. Huerta M, Lord C. Diagnostic evaluation of autism spectrum disorders. Pediatr Clin N Am. 2012;59(1):103–11. xi. PubMed PMID: 22284796. Pubmed Central PMCID: 3269006.
- <span id="page-18-8"></span>160. Taylor LJ, Eapen V, Maybery MT, Midford S, Paynter J, Quarmby L, et al. Diagnostic evaluation for autism spectrum disorder: a survey of health professionals in Australia. BMJ Open. 2016;6(9):e012517. PubMed PMID: 27601502. Pubmed Central PMCID: 5020660.
- <span id="page-18-9"></span>161. Filipek PA, Accardo PJ, Ashwal S, Baranek GT, Cook EH Jr, Dawson G, et al. Practice parameter: screening and diagnosis of autism: report of the Quality Standards Subcommittee of the American Academy of Neurology and the Child Neurology Society. Neurology. 2000;55(4):468–79. PubMed PMID: 10953176.
- <span id="page-18-10"></span>162. Zwaigenbaum L, Bauman ML, Choueiri R, Kasari C, Carter A, Granpeesheh D, et al. Early intervention for children with autism spectrum disorder under 3 years of age: recommendations for practice and research. Pediatrics. 2015;136(Suppl 1):S60–81. PubMed PMID: 26430170.
- <span id="page-18-11"></span>163. Lord C, Risi S, Lambrecht L, Cook EH Jr, Leventhal BL, DiLavore PC, et al. The autism diagnostic observation schedule-generic: a standard measure of social and communication deficits associated with the spectrum of autism. J Autism Dev Disord. 2000;30(3):205–23. PubMed PMID: 11055457.
- <span id="page-18-12"></span>164. Stone WL, Coonrod EE, Ousley OY. Brief report: screening tool for autism in two-yearolds (STAT): development and preliminary data. J Autism Dev Disord. 2000;30(6):607–12. PubMed PMID: 11261472.
- <span id="page-18-13"></span>165. Wetherby AM, Allen L, Cleary J, Kublin K, Goldstein H. Validity and reliability of the communication and symbolic behavior scales developmental profile with very young children. J Speech Lang Hear Res: JSLHR. 2002;45(6):1202–18. PubMed PMID: 12546488.
- <span id="page-18-14"></span>166. Kim SH, Lord C. Restricted and repetitive behaviors in toddlers and preschoolers with autism spectrum disorders based on the Autism Diagnostic Observation Schedule (ADOS). Autism Res: Off J Int Soc Autism Res. 2010;3(4):162–73. PubMed PMID: 20589716. Pubmed Central PMCID: 3005305.
- <span id="page-18-15"></span>167. Listhaus AD, Freeman WR. Fluorescein angiography in patients with posterior uveitis. Int Ophthalmol Clin. 1990;30(4):297–308. PubMed PMID: 2228479.